

# SCIENTIFIC REPORTS

OPEN

## Re-exploration of U's Triangle *Brassica* Species Based on Chloroplast Genomes and 45S nrDNA Sequences

Chang-Kug Kim<sup>1</sup>, Young-Joo Seol<sup>2</sup>, Sampath Perumal<sup>3,4</sup>, Jonghoon Lee<sup>3,5</sup>, Nomar Espinosa Waminal<sup>3</sup>, Murukarthick Jayakodi<sup>3</sup>, Sang-Choon Lee<sup>3</sup>, Seungwoo Jin<sup>3</sup>, Beom-Soon Choi<sup>6</sup>, Yeisoo Yu<sup>6</sup>, Ho-Cheol Ko<sup>7</sup>, Ji-Weon Choi<sup>8</sup>, Kyoung-Yul Ryu<sup>2</sup>, Seong-Han Sohn<sup>1</sup>, Isobel Parkin<sup>4</sup> & Tae-Jin Yang<sup>3,9</sup>

The concept of U's triangle, which revealed the importance of polyploidization in plant genome evolution, described natural allopolyploidization events in *Brassica* using three diploids [*B. rapa* (A genome), *B. nigra* (B), and *B. oleracea* (C)] and derived allotetraploids [*B. juncea* (AB genome), *B. napus* (AC), and *B. carinata* (BC)]. However, comprehensive understanding of *Brassica* genome evolution has not been fully achieved. Here, we performed low-coverage (2–6 ×) whole-genome sequencing of 28 accessions of *Brassica* as well as of *Raphanus sativus* [R genome] to explore the evolution of six *Brassica* species based on chloroplast genome and ribosomal DNA variations. Our phylogenomic analyses led to two main conclusions. (1) Intra-species-level chloroplast genome variations are low in the three allotetraploids (2–7 SNPs), but rich and variable in each diploid species (7–193 SNPs). (2) Three allotetraploids maintain two 45SnrDNA types derived from both ancestral species with maternal dominance. Furthermore, this study sheds light on the maternal origin of the AC chloroplast genome. Overall, this study clarifies the genetic relationships of U's triangle species based on a comprehensive genomics approach and provides important genomic resources for correlative and evolutionary studies.

Brassicaceae is one of the largest eudicot families; it contains more than 330 genera and 3,800 species. The genomes of species in the tribe Brassiceae share a common whole-genome triplication, which is considered to be a crucial event that drove diversification of the species and intra-species morphotypes<sup>1,2</sup>. Brassiceae includes several economically important crops that are used for vegetables, oils, and fodders. The basic foundation for the systematic relationship of the six major *Brassica* species was classically explained as U's triangle<sup>3</sup>. U's triangle proposed that the three tetraploid species *B. juncea* (AABB genome,  $2n = 4x = 36$ ), *B. napus* (AACC,  $2n = 4x = 38$ ), and *B. carinata* (BBCC,  $2n = 4x = 34$ ) are the derived allotetraploids of the diploid species *B. rapa* (AA,  $2n = 2x = 20$ ), *B. nigra* (BB,  $2n = 2x = 16$ ), and *B. oleracea* (CC,  $2n = 2x = 18$ ), respectively, which arose by natural hybridization and chromosome doubling.

Whole-genome sequencing (WGS) analyses of the A, C, AB, and AC genomes has increased our understanding of *Brassica* genome evolution<sup>4–8</sup>. It has been suggested that the *Brassica* genome diverged from *Arabidopsis thaliana* around 17 million years ago (mya)<sup>9</sup>, and there is evidence that the B genome first diverged from the

<sup>1</sup>Genomics Division, National Institute of Agricultural Sciences, Jeonju, 54874, Korea. <sup>2</sup>International Technology Cooperation Center, Rural Development Administration, Jeonju, 54875, Korea. <sup>3</sup>Department of Plant Science, Plant Genomics and Breeding Institute, Research Institute of Agriculture and Life Sciences, College of Agriculture and Life Sciences, Seoul National University, Seoul, 08826, Republic of Korea. <sup>4</sup>Agriculture and Agri-Food Canada, 107 Science Place, Saskatoon, SK, S7N 0X2, Canada. <sup>5</sup>Joeun Seed, Goesan-Gun, Chungcheongbuk-Do, 28051, Republic of Korea. <sup>6</sup>Phyzen Genomics Institute, Seongnam, 13558, Republic of Korea. <sup>7</sup>National Agrobiodiversity Center, National Institute of Agricultural Sciences, RDA, Jeonju, Jeollabukdo, 54874, Republic of Korea. <sup>8</sup>Postharvest Technology Division, National Institute of Horticultural and Herbal Science, Wanju, Republic of Korea. <sup>9</sup>Crop Biotechnology Institute/GreenBio Science and Technology, Seoul National University, Pyeongchang, 232-916, Republic of Korea. Chang-Kug Kim, Young-Joo Seol and Sampath Perumal contributed equally to this work. Correspondence and requests for materials should be addressed to T.-J.Y. (email: [tjyang@snu.ac.kr](mailto:tjyang@snu.ac.kr))

*Brassica* lineage around 9 mya, followed by divergence of the A and C genomes around 4.5 mya<sup>10,11</sup>. Recent genome sequencing of the two AC and AB genome allotetraploids suggested that they derive from allotetraploidization events that happened approximately 8,000–51,000 years ago<sup>6,8</sup>.

Cells contain three different genomes (nuclear, mitochondrial, and chloroplast) that follow different evolutionary pathways<sup>12</sup>. Chloroplast, mitochondrial, and nuclear ribosomal DNA sequences are crucial resources to understand plant genomic diversity due to their highly conserved nature and strong phylogenetic signals. The chloroplast genome is circular, relatively simple, and inherited uniparentally with a highly conserved gene structure and gene order<sup>13,14</sup>. The chloroplast genome has sufficiently informative nucleotide divergence that it can be utilized to understand genetic diversity, genomic origin, and genetic relationships, as well as for barcode marker development<sup>15–19</sup>. A few systematic studies have explored the *Brassica* chloroplast genome; however, these results have yielded a partial and unresolved understanding<sup>20–23</sup>.

Nuclear ribosomal DNA (nrDNA) sequences are highly homozygous, tandemly repeated transcriptional units that encode important housekeeping functions in nuclear assembly and nuclear function<sup>24,25</sup>. Two nuclear ribosomal DNA blocks, 5SnrDNA and 45SnrDNA, are generally localized on different chromosomes in plants. The 45SnrDNA units contain a highly conserved multicistronic gene with 18S, 5.8S, and 28S RNA sequences and relatively polymorphic internal transcribed spacer (ITS) regions, which makes 45SnrDNA a preferred target for both phylogenetic and barcoding analyses<sup>26–28</sup>.

Advances in next-generation sequencing (NGS) technology and bioinformatics algorithms are facilitating the discovery of extensive natural variations in large populations. Most research has focused on identification of intra-species natural variations in the nuclear genome to explore diversity, adaptation, domestication, and evolution, as well as to mine for new alleles<sup>29</sup>. Our group recently established a method based on ‘genome skimming’ approach called dnaLCW for high-throughput simultaneous *de novo* assembly of chloroplast and 45SnrDNA transcription unit sequences using low-coverage whole-genome NGS to reveal inter-species and intra-species diversity<sup>30–32</sup>.

The objective of the current study is to elucidate the genetic diversity and evolution of *Brassica* species belonging to U’s triangle by performing whole-genome sequencing (WGS). We report the complete sequences of chloroplast genomes and 45SnrDNA transcription units for 28 genotypes. We also investigate genome-wide variation and phylogenomic analysis for chloroplast genomes and 45SnrDNA sequences to revisit the evolution of the six *Brassica* species in U’s triangle compared with the related species *Raphanus sativus*.

## Results

**Characterization of 28 complete chloroplast genomes.** The complete chloroplast genomes were obtained for 28 genotypes using the dnaLCW approach (Table 1). Annotation of chloroplast genomes revealed conserved quadripartite structures with coherent gene number and gene order among the 28 genotypes (Fig. 1). The chloroplast genome is highly conserved, with 99–100% sequence similarity within each species, although meaningful variations were observed between species with 98.1–99.5% sequence similarity (Fig. 2; Table S1). Chloroplast genome lengths varied by 607 bp among the 28 genotypes, ranging from 153,037 bp (accession A4) to 153,642 bp (accession B4). Chloroplast genome copy numbers were estimated based on read depth for the haploid genome size, ranging from 453 (accession AB2) to 1,279 (accession BC1) copies per cell (Table 1).

The chloroplast genomes showed different levels of intra-species polymorphism (Tables 2, S2, S3). The chloroplast genomes from C genome species had very low intra-species diversity with seven SNPs and four InDels, whereas other chloroplast genomes had relatively high intra-species diversity with 88 SNPs and 16 InDels in the A species genomes, 99 and 24 in the B species genomes, and 193 and 112 in the R genome, respectively (Tables S2, S3). Polymorphism was richer in genic regions than in intergenic regions (Table 3). Abundant polymorphisms were detected on the inter-species level (Tables 2, S4). The highest number of inter-species variations was 2,502 SNPs and 294 InDels between the B and C chloroplast genomes, and the lowest was 257 SNPs and 65 InDels between the A and C chloroplast genomes (Table S4). The three tetraploids showed fewer variations in the chloroplast genome compared to the diploid species.

**Characterization of 45SnrDNA sequences.** The complete 45SnrDNA sequences of the four diploid species ranged from 5,816 to 5,831 bp (Table 1). Only one representative 45S was identified for each of the 16 diploid accessions of the A, B, C, and R genomes. By contrast, two different 45S sequences were identified for each of the tetraploid accessions. Therefore, 24 different 45SnrDNA sequences were identified for all 12 genotypes of the three allotetraploids (AB, AC, and BC genomes) (Table 1). Comparative analysis of the 40 types of 45SnrDNA sequences revealed 39 bp length variations in the 5,818 bp sequence (Figs 3, 4). Compared with the chloroplast genome, 45SnrDNA sequences were less diverse, with 22 SNPs and one InDel among 40 types of 45SnrDNA sequences from 28 genotypes. These variations were distributed among genic and intergenic regions.

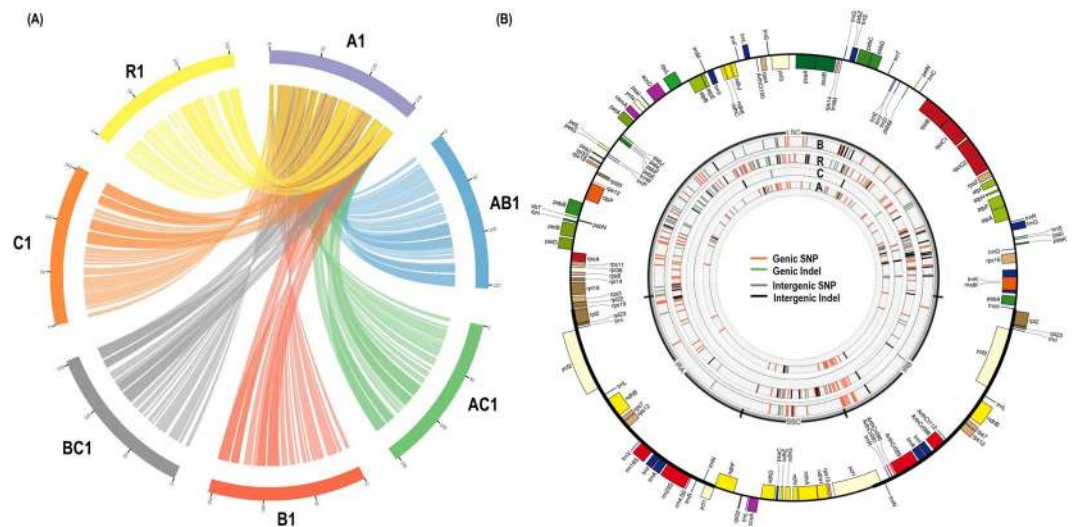
Our analysis identified two types of 45SnrDNA (both parental) in three allotetraploids (AB, AC, and BC genomes). Each 45SnrDNA type in the three allotetraploids showed 100% sequence similarity with their corresponding parental diploid genome. For example, the A and B types of 45SnrDNA in the AB genome were 100% identical with those in the A and B genomes, respectively (Figs 3, 4). Read depth approach was used to estimate the copies of each 45SnrDNA type. Copy numbers differed among the allotetraploid sub-genomes, with 3,000–6,500 copies in the AB genome, 6,000–11,000 copies in the AC genome, and 2,200–2,900 copies in the BC genome. Copy numbers of each 45SnrDNA type in sub-genomes displayed a biased proportion up to 1.5–3-fold, with a higher proportion always occurring in the maternal ancestors of the AB, AC, and BC genomes (Table S5).

**Validation and utilization of species-specific variations.** Although chloroplast and 45SnrDNA sequences are highly conserved, our comparative analyses revealed a considerable number of variations (Fig. 5). There were more SNPs in chloroplast sequences than in 45SnrDNA sequences, with an average of 15 SNPs

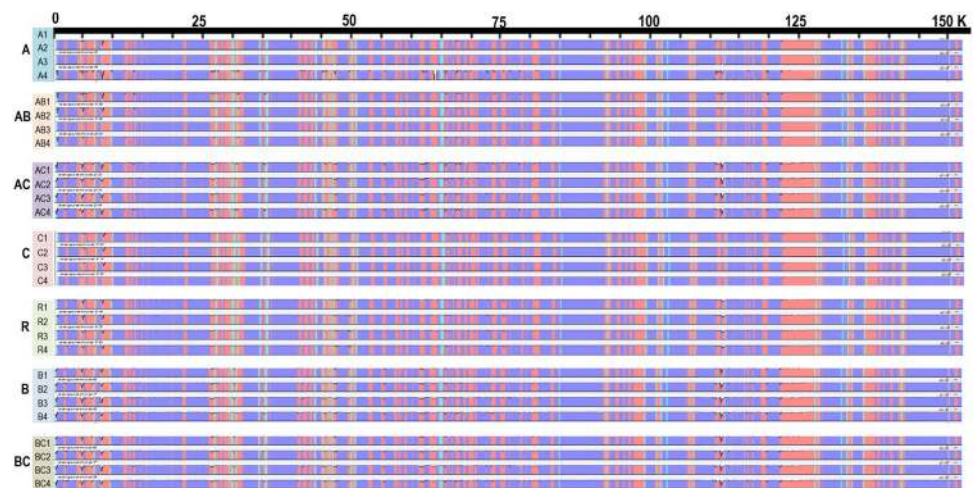
Organism and genome	Genotype ID <sup>a</sup>	Genome size (Mb)	Total reads (Mb)	Chloroplast genome			45SnrDNA		
				Length (bp)	Copy number (x) <sup>b</sup>	Accession number	Length (bp)	Copy number (x) <sup>b</sup>	Accession number
<i>B. rapa</i> (A)	A1	529	1,557	153,483	378	KX681647	5,818	3,216	KX709342
	A2	529	1,214	153,482	305	KX681648	5,818	3,770	KX709343
	A3	529	1,352	153,482	363	KX681649	5,818	3,872	KX709344
	A4	529	1,293	153,037	496	KX681650	5,818	4,183	KX709345
<i>B. nigra</i> (B)	B1	632	1,532	153,633	378	KT878383	5,831	1,819	KX709346
	B2	632	1,632	153,641	221	KX681651	5,831	1,667	KX709347
	B3	632	1,489	153,623	323	KX681652	5,831	1,324	KX709348
	B4	632	1,631	153,642	244	KX681653	5,831	1,571	KX709349
<i>B. oleracea</i> (C)	C1	630	1,489	153,364	278	KX681654	5,811	2,873	KX709350
	C2	630	1,312	153,364	510	KX681655	5,848	1,384	KX709351
	C3	630	1,611	153,364	285	KX681656	5,818	2,768	KX709352
	C4	630	2,115	153,363	347	KX681657	5,819	1,957	KX709353
<i>R. sativus</i> (R)	R1	530	1,467	153,372	264	KX681658	5,816	3,812	KX709354
	R2	530	1,487	153,444	412	KX681659	5,816	2,042	KX709355
	R3	530	1,440	153,376	393	KX681660	5,819	4,174	KX709356
	R4	530	1,470	153,370	343	KX681661	5,823	4,614	KX709357
<i>B. juncea</i> (AB)	AB1-A	1,068	1,469	153,483	779	KX681662	5,818	2,412	KX709358
	AB1-B						5,831	1,589	KX709359
	AB2-A	1,068	1,352	153,483	358	KX681663	5,818	1,883	KX709360
	AB2-B						5,831	690	KX709361
	AB3-A	1,068	1,528	153,490	495	KX681664	5,818	2,192	KX709362
	AB3-B						5,831	1,041	KX709363
	AB4-A	1,068	1,549	153,483	338	KX681665	5,818	3,449	KX709364
	AB4-B						5,831	1,190	KX709365
<i>B. napus</i> (AC)	AC1-A	1,130	1,534	153,452	630	KX681666	5,831	1,445	KX709366
	AC1-C						5,818	689	KX709367
	AC2-A	1,130	1,401	153,429	890	KX681667	5,831	1,169	KX709368
	AC2-C						5,819	879	KX709369
	AC3-A	1,130	1,401	153,429	925	KX681668	5,817	1,009	KX709370
	AC3-C						5,832	865	KX709371
	AC4-A	1,130	1,579	153,453	366	KX681669	5,831	982	KX709372
	AC4-C						5,818	741	KX709373
<i>B. carinata</i> (BC)	BC1-B	1,284	2,156	153,636	762	KX681670	5,818	4,223	KX709374
	BC1-C						5,818	2,409	KX709375
	BC2-B	1,284	1,457	153,636	919	KX681671	5,818	5,865	KX709376
	BC2-C						5,818	3,453	KX709377
	BC3-B	1,284	1,710	153,641	913	KX681672	5,818	2,813	KX709378
	BC3-C						5,817	1,836	KX709379
	BC4-B	1,284	1,511	153,636	540	KX681673	5,818	4,791	KX709380
	BC4-C						5,818	2,551	KX709381

**Table 1.** Summary of chloroplast and 45SnrDNA assemblies from 28 *Brassica* and *Raphanus* genotypes. <sup>a</sup>rDNA from tetraploids was designated as A, B, or C based on the parental genome or sub-genome type. The complete details and list of organisms can be found in Table S6. <sup>b</sup>Copy numbers of chloroplast and 45SnrDNA were estimated based on average read depth mapping and converted into the corresponding haploid genome size.

identified for every 1 kb of chloroplast genome, but only 3 for every 1 kb of 45SnrDNA. We tested the utility of this information on the diversity in chloroplast and 45SnrDNA sequences for identification and authentication of species or cultivars. We began with comprehensive analysis of the SNP and InDel variations in the chloroplast genome and 45SnrDNA to facilitate the development of barcode markers that enable the discrimination of each species. A total of 2,796 chloroplast variations were identified in 28 genotypes, and many of them were potential candidates for species-specific marker development (Tables S2, S3). We performed PCR analysis to validate the sequence polymorphism against several diversity-containing regions, and identified two InDel variations based on the chloroplast genome that could differentiate each diploid genome (A, B, C, and R genomes) (Figure S1). By contrast, only 23 variations (including 22 SNPs and one InDel) were identified based on 40 different 45SnrDNA sequences from 28 genotypes (Fig. 3). The 18S and ITS regions had relatively rich diversity and provide potential targets to differentiate the A, B, C, and R genomes by PCR analysis (Figure S2).



**Figure 1.** Chloroplast genome variations and comparative analysis in seven Brassicaceae species. **(A)** Syntenic comparisons of chloroplast genomes in *Brassica*. Circos-based syntenic comparative map developed for *B. rapa* (A1) against *B. juncea* (AB1), *B. napus* (AC1), *B. nigra* (B1), *B. carinata* (BC1), *B. oleracea* (C1), and *Raphanus sativus* (R1). Syntenic blocks with minimum length of 1 kb were used for the syntenic analysis. **(B)** Distribution of intra-species variations in *B. nigra* (B), *R. sativus* (R), *B. oleracea* (C), and *B. rapa* (R) chloroplast genomes. Outermost chloroplast circular map was developed from the *B. rapa* chloroplast genome (A1) using OGDRAW. Genes are represented in different colors. Positive and negative gene orientations are shown as outer and inner circles, respectively. Inner circles represent variations in the B, R, C, and A genomes, respectively.



**Figure 2.** Comparative analysis based on complete chloroplast genomes identify similar and variable regions among the 28 *Brassica* and *Raphanus* genotypes.

SNP/Indel	A	B	C	R
A	88/16 <sup>a</sup>	280 <sup>b</sup>	65	167
B	2,402 <sup>b</sup>	99/24	294	245
C	257	2,502	7/4	183
R	1,203	2,259	1,293	193/112

**Table 2.** Summary of inter-species and intra-species variations based on chloroplast genomes. <sup>a</sup>18/16 denotes the number of SNP/Indel variations in the A genome. <sup>b</sup>B genome has 2,402 and 280 SNP and InDel variations, respectively, compared with the A genome.



Genome	SNP			Indel		
	Genic	Intergenic	Total	Genic	Intergenic	Total
A	58	30	88	8	8	16
B	68	31	99	13	11	24
C	2	5	7	3	1	4
R	137	56	193	57	55	112

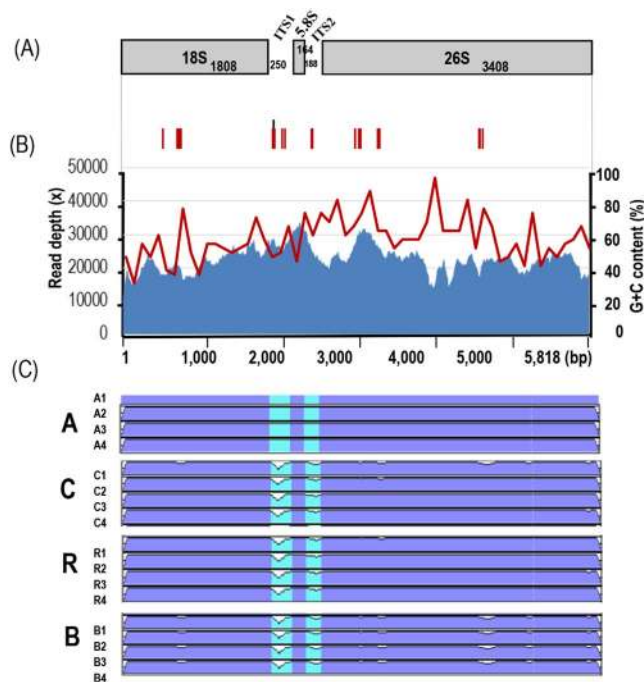
**Table 3.** Summary and distribution of intraspecies SNP and Indel variations based on chloroplast genomes.

Locus	18S					ITS1		IT S2		26S														
	4	6	6	6	7	1	1	1	1	1	2	2	2	2	2	2	2	3	3	3	3	4	4	4
Position	9	7	9	9	1	5	7	8	8	9	0	3	3	8	9	9	1	1	1	1	4	4	4	4
	8	1	1	9	6	9	3	3	2	6	5	0	7	4	8	3	7	8	8	9	0	0	3	7
AA	A1	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
	A2	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
	A3	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
	A4	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
BB	B1	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
	B2	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
	B3	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
	B4	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
CC	C1	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
	C2	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
	C3	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
	C4	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
RR	R1	T	C	T	T	T	G	CCGG AAT	C	A	C	A	T	T	A	T	G	T	A	T	A	G	A	C
	R2	T	C	T	T	T	G	CCGG AAT	C	A	C	A	T	T	A	T	G	T	A	T	A	G	A	C
	R3	T	C	T	T	T	G	CCGG AAT	C	A	C	A	T	T	A	T	G	T	A	T	A	G	A	C
	R4	T	C	T	T	T	G	CCGG AAT	C	A	C	A	T	T	A	T	G	T	A	T	A	G	A	C
AA	AB1-A	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
	AB2-A	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
	AB3-A	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
	AB4-A	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
BB	AB1-B	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
	AB2-B	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
	AB3-B	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
	AB4-B	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
AA	AC1-A	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
	AC2-A	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
	AC3-A	T	C	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
	AC4-A	T	T	T	C	C	A	---	T	A	A	A	T	C	G	T	A	C	T	A	G	A	G	T
CC	AC1-C	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
	AC2-C	C	C	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
	AC3-C	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
	AC4-C	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
BB	BC1-B	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
	BC2-B	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
	BC3-B	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
	BC4-B	T	C	C	C	T	G	GCCGATT	C	T	C	G	C	C	G	C	G	T	A	T	A	A	G	T
CC	BC1-C	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
	BC2-C	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
	BC3-C	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T
	BC4-C	C	T	T	C	C	G	GCI GATT	C	A	C	A	T	C	G	T	A	C	T	A	G	A	G	T

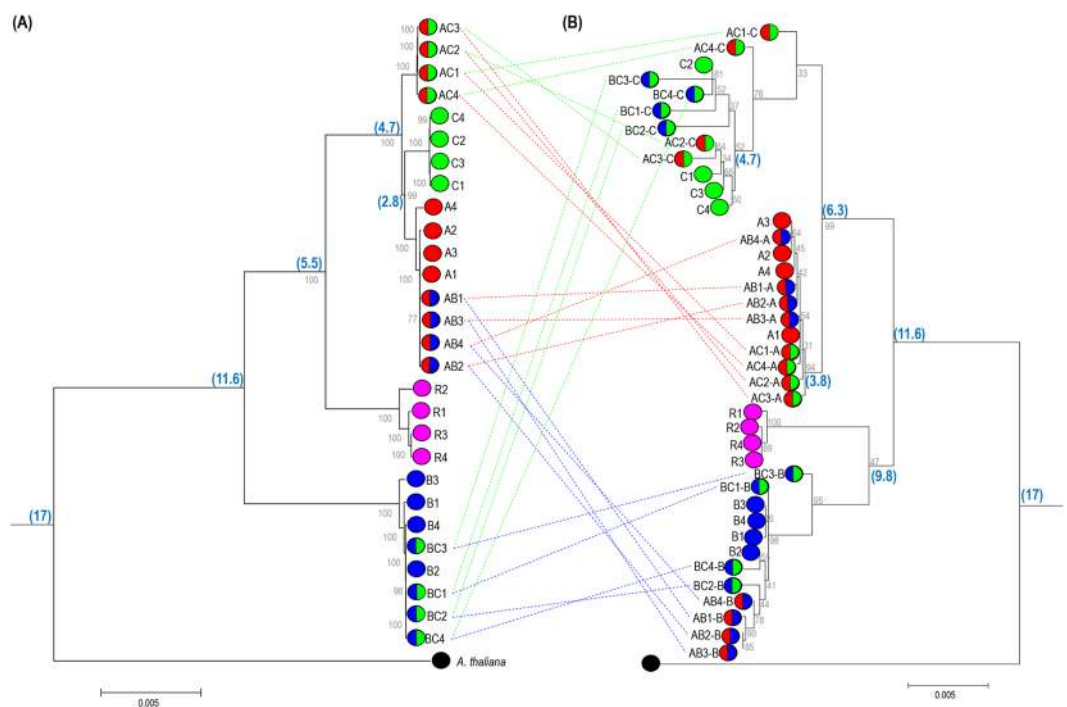
**Figure 3.** Summary of nucleotide variations based on 45SnrDNA sequences from 28 genotypes.

**Phylogenomic exploration of U's triangle.** Separate phylogenetic analyses based on chloroplast and 45SnrDNA sequences identified conserved genetic relationships and displayed essentially identical topologies among the *Brassica* species in U's triangle. The high bootstrap values on the nodes support the reliability of the phylogenies produced based on both chloroplast and 45SnrDNA sequences (Fig. 5).

The chloroplast phylogenetic tree displayed five different clades, with clear discrimination between the four diploid genomes but an ambiguous clade in the AC genome (Figs 5A; S3). The four AC genomes were clustered with each other, but did not group with the parental A or C genomes. The other two allotetraploids (AB and BC) were clustered with one of their parental genomes; AB clustered with the A genome, and BC clustered with the B genome indicating that the A and B genomes were the maternal ancestors for the AB and BC genomes, respectively. However, the AC genome followed neither the A nor C genome as a source of maternal origin and showed an enigmatic relationship with its diploid parental genomes. Furthermore, we did not observe any reciprocal hybridization patterns in any of the three tetraploids.



**Figure 4.** Structure and similarity analyses of 45SrDNA sequences from 28 *Brassica* and *Raphanus* genotypes. (A) Complete structure and gene annotation of 45SrDNA sequences from the A1 genome. (B) Red and black arrowheads indicate the SNP and InDel variations, respectively. (C) Coverage of 45SrDNA-based read mapping. Red lines indicate the proportion of G + C in the 4S 45SrDNA. (D) Comparative analysis of similar and variable regions using mVISTA. Red arrowheads indicate inter-species variations.



**Figure 5.** Phylogenetic relationships of the genus *Brassica* inferred from complete chloroplast (A) and 45SrDNA (B) sequences of 28 *Brassica* and *Raphanus* genotypes. Tree was developed using MEGA7 with 1,000 bootstrap replications. The bootstrap values for clades are shown in corresponding branches of the tree. Taxon markers with single and double circles represent diploid and tetraploid genomes, respectively. The circled legend for 45SrDNA and chloroplast corresponds to each species. Dotted line connects the corresponding allotetraploid genomes of chloroplast and 45SrDNA. Species divergence times were inferred from Bayesian analysis, and are shown at the side of the node in million years (my). *A. thaliana* was used as an outgroup.

The 45SnrDNA phylogenetic tree displayed clear classifications (Figs 5B, S4). The four diploid species diverged into four distinct clades, and each of the tetraploid genomes contained two independent 45SnrDNAs according to their corresponding ancestral maternal/paternal genomes. The AB genome harbored both A-type and B-type 45SnrDNA (AB-A and AB-B, respectively) with AB-A from the maternal genome and AB-B from the paternal. The BC and AC genomes also harbored two original subgenomic 45SnrDNA types; the B and C type in the BC genome (BC-B and BC-C, respectively), and the A and B type in the AB genome (AB-A and AB-B, respectively). Overall, the 45SnrDNA phylogenetic analysis of three allotetraploids revealed the expected parental association with the three diploids.

A phylogenetic tree generated by BEAST analysis showed similar topology as that generated by MEGA. The molecular dating based on chloroplast and 45SnrDNA sequences generated essentially similar divergence times (Figures S3, S4). Tree topologies with inferred speciation dates clearly identified three major divergence periods in both analyses (chloroplast and 45SnrDNA) of *Brassica*. The trees indicated that divergence and speciation in the B genome occurred 11 mya, followed by R genome divergence at 9 mya, and speciation of A and C genomes about 4.5 mya. The allotetraploids appear to have arisen from their diploid ancestors from 0.001 to 0.03 mya (Figure S5).

## Discussion

**Inter-species nucleotide diversity of chloroplast and 45SnrDNA.** Chloroplast genomes are highly stable, have low mutation rates, and produce highly reliable phylogenetic trees that help elucidate plant evolutionary history<sup>15,18,30,33</sup>. Nuclear ribosomal DNAs can remain highly homozygous, whereas nuclear genomes are subject to cross-hybridization and cross-over during meiosis<sup>32</sup>. We produced a comprehensive diversity map for *Brassica* based on 28 complete chloroplast genomes and 40 types of complete 45SnrDNA sequences of the major species listed in the classical U's triangle. The chloroplast and 45SnrDNA sequences exhibited highly conserved gene structures and gene orders at the inter-species and intra-species levels. However, considerable numbers of nucleotide variations were observed in both chloroplast and 45SnrDNA, which represent genus- and species-specific variations that can be developed for barcode markers and molecular breeding analysis. The B genome was highly diverged from the other genomes, suggesting prolonged, independent evolution. The estimated speciation time of the B genome is consistent with this result (Table 2). Phylogenetic analyses based on chloroplast genomes and 45SnrDNA sequences showed general agreement, with the B genome as a sister group to the A and C genomes and the derived allotetraploids following their corresponding progenitor genomes<sup>6,34</sup>. The chloroplast and 45SnrDNA sequences indicate that the R genome was closer to the A and C genomes than the B genome.

### Chloroplast genomes of seven species have different intra-species nucleotide diversity levels.

Four genotypes for each species all showed different levels of divergent intra-species chloroplast genome polymorphism, although there were fewer intra-species polymorphisms than inter-species polymorphisms. The three allotetraploids rarely showed intra-species diversity (as expected) because those species were generated by allotetraploidization less than 0.05 mya<sup>6,8</sup>. Although four genotypes cannot represent the full diversity of each species, our results indicate that the C genome is less diverse than the maternal ancestor genome, the A and B genomes are moderately diverse, and the R genome is more diverse than the maternal ancestor genome. These results are consistent with a recent report of very low diversity in the C genome and relatively rich diversity in the A and B chloroplast genomes<sup>23,35</sup>. Similarly, rich variations were identified in the mitochondrial genome of *R. sativus*. Our previous work showed that dynamic mitochondrial genome rearrangements caused cytoplasmic male sterility and large variations among radish lines<sup>36</sup> compared with the relatively conserved mitochondrial genome structures in *Brassica*<sup>30,37</sup>.

**Two types of 45SnrDNA are derived from two diploid ancestors in allotetraploids.** Some polyploid plants maintain both parental ribosomal DNA genomes (5SnrDNA and 45SnrDNA) after allopolyploidization<sup>38–41</sup>. However, many allopolyploids express nucleolar dominance (ND), in which rRNA from one parent is transcriptionally silenced or recessively expressed<sup>42,43</sup>. ND is anticipated to have a significant role in chromatin modification and genome evolution<sup>44,45</sup>. Homogenization into one of two rDNA types also occurs via concerted evolution, mediated by rearrangements such as repeat loss, replacement, and recombination<sup>46–49</sup>.

There are few reports of the complete 45SnrDNA sequence in plant genomes. Here, we obtained the complete 45S rDNA transcription sequences for 28 accessions. We found only one highly homologous 45SnrDNA sequence in each accession of four diploids, but detected two types of 45SnrDNAs derived from the parental diploid ancestors in all three allotetraploids. Copy number analysis revealed that 45SnrDNA sequence bias toward the maternal genome occurred in the order of A > B > C genomes (Table S5), suggesting that there was genome-specific expansion of 45SnrDNA, which might be caused by sub-genome dominance<sup>6</sup>. However, further studies are required to address the consequences of rDNA copy number variation in allotetraploid *Brassica* (Table 1).

**Origin of the chloroplast genome in the AC genome accessions.** Phylogenetic analyses conducted with only one or a few loci can misrepresent the derived phylogenetic history, and complete information on genetic diversity is required for accurate analysis<sup>50</sup>. Unlike 45SnrDNA, chloroplast-based phylogenetic analysis indicates that the AC genome chloroplasts did not follow either of the parental nuclear genomes (A or C genome). Studies have been performed to clarify the genetic relationships of the major diploid and tetraploid *Brassica* species, but the origin of the chloroplast in the AC genome species is still unclear<sup>15,20,21,51</sup>. Initially, maternal parent of the AC genome was thought to be derived from the C genome due to their similarities in their chloroplast DNA restriction digest patterns; however, analysis with a wider range of accessions suggested that A genome was the maternal source<sup>20</sup>. Moreover, analysis with both chloroplast and nuclear markers suggested that the AC genome arose from several independent hybridization events including artificial introgression of A-genome<sup>51</sup>. A survey of the *rho*



locus revealed that >90% of 488 AC accessions displayed different genotypes than the parental accessions (A and C), but they were classified as an independent group with different origin<sup>20</sup>. Comparison with the recent findings of the *Brassica* chloroplast genome shows overall agreement, such as grouping based on species and the maternal and paternal origin of the allotetraploids<sup>23</sup>. Though there were two different A genome sources for the AC and AB genomes, we did not observe any divergence based on 45SnrDNA, suggesting that 45SnrDNA has been conserved in the *Brassica* genome<sup>8</sup>. Furthermore, chloroplast genomes from nine and seven different A and C genome morphotypes, respectively, formed a single cluster to confirm that the chloroplast genome and 45SnrDNA are stable even upon divergence of different sub-species and morphotypes<sup>23</sup>. In addition, sub-genome parallel selection played a crucial role in evolution of different morphotypes<sup>52</sup>.

Furthermore, A recent chloroplast genome survey of more diverse A genotypes revealed two different types of chloroplast genomes. The rapa-type1 chloroplast genome is generally found in all *B. rapa*, whereas the rapa-type2 is unique for some Italian Broccoletto genotypes of *B. rapa*<sup>23</sup>. Phylogenetic analysis indicated that the rapa-type2 clustered with the chloroplast of the AC genome, which explains why the Italian Broccoletto genotype is the donor for the most abundant AC chloroplast genome. The rapa-type2 chloroplast genome diverged 4.7 mya, which coincides with the currently known A and C genome divergence around 5.4 to 2.7 mya (Fig. 5). By contrast, analysis of the AC genome indicated that allotetraploidization occurred 7,500 years ago<sup>6</sup>. Both of these results indicate that the rapa-type2 chloroplast genome was maintained in the Italian Broccoletto genotype by geographical isolation or maternal dominance since 4.7 mya, and the Italian Broccoletto genome was utilized as the maternal parent to generate the AC genome 7,500 years ago. However, there are still questions about the evolution of the maternal genomes for the A and AC genomes. It is still not known how the rapa-type2 chloroplast genome became associated with the common maternal parent for most AC genomes, although the Italian Broccoletto genotype is not widespread in the A genome.

## Conclusion

This study analyzed the genetic relationships and diversity among *Brassica* species using chloroplast genome and 45SnrDNA sequences. Phylogenetic analysis revealed that the B genome diverged first in the *Brassica* clade, followed by R, A, and C, and with three allotetraploids forming during last 0.1 to 0.01 mya. We cataloged the complete variants in chloroplast and 45SnrDNA sequences, which will serve as excellent resources for the development of barcode markers and species identification. Comparative genome analyses of species-specific variations would facilitate the study of genome evolution and morphological divergence of *Brassica*. The combined results of this study reveal comprehensive genetic relationships of U's triangle species and provide insights into genome evolution in *Brassica*. The results of this study will be extensively applicable for species identification and evolutionary studies.

## Materials and Methods

**Plant materials and DNA sequencing.** Seeds of four genotypes representing each A, B, C, R, AB, AC, and BC genome were obtained from the RDA Genebank Center, Suwon, South Korea. All plants were grown at 22 °C (day)/18 °C (night) with a 16 h light/8 h dark photoperiod at the RDA experimental farm, Suwon, South Korea, during the spring of 2014. High-quality total genomic DNA was isolated from young leaves using a modified CTAB method<sup>53</sup>. Whole-genome shotgun libraries were generated using the TrueSeq DNA PCR-Free Library Preparation kit (Illumina) according to the manufacturer's instructions. Briefly, 5 ng of high-quality DNA from each accession was fragmented via sonication. Then, the fragments were end-repaired and A-tailed. Adapters were ligated, including the barcoding and multiplex identifier adapters, and the fragments were amplified with 10 PCR cycles. Finally, a paired-end (PE) library with inserts of 400–500 bp was generated. The library was sequenced with the MiSeq System (Illumina) at LabGenomics ([www.labgenomics.co.kr](http://www.labgenomics.co.kr), South Korea). Multiplex adapters were used to separate the 28 genotypes from the bulked raw reads, and the sequence reads were trimmed for adapters and low quality and utilized for further analysis. All trimmed high quality sequences (NN3658-NN3685) for the 28 accessions were deposited into the National Agricultural Biotechnology Information Center (<http://nabic.rda.go.kr>) public database<sup>54</sup> (Table S6).

**Assembly and annotation of chloroplast genome and 45SnrDNA sequences.** Complete chloroplast genome and 45SnrDNA sequences were simultaneously assembled for all 28 *Brassica* and *Raphanus* genotypes using the dnaLCW method<sup>32</sup>. The dnaLCW method is a fast and comparatively easy method that does not require a PCR based gap filling to assemble the chloroplast and rDNA sequences. With slight modification, dnaLCW also allows the characterization of the major repeats in the *Brassica* genome<sup>55</sup>. Briefly, high-quality Illumina paired-end reads were *denovo* assembled using the CLC genome assembler (ver. 4.06 beta, CLC Inc., Aarhus, Denmark) with autonomously controlled overlap size (200–500 bp). After gap closing, the resulting contigs were homology searched against the *Arabidopsis thaliana* chloroplast reference genome (GenBank accession: NC\_000932) using mummer. Contigs related to the chloroplast genome were ordered according to the reference genome. Gaps and other errors such as false SNPs, copies of tandem repeats and homopolymer errors were corrected according to the dnaLCW approach<sup>32</sup>. Likewise, *Arabidopsis thaliana* 45SnrDNA sequence (GenBank accession: X52322.1) was used as a reference to assemble the 45SnrDNA sequences of 28 genotypes. Due to the number of variations in intergenic spacer sequences (up to six types in *B. oleracea*), only the unique 45SnrDNA transcription units were assembled. We also identified both parental types of 45SnrDNA in an allotetraploid genome [i.e., *B. napus* (AC) genome] containing parental or sub-genomes of *B. rapa* (A) and *B. oleracea* (C), which were represented as AC-A and AC-C, respectively.

The chloroplast genomes of the 28 genotypes were annotated for protein-coding genes, transfer RNA (tRNA), and ribosomal RNA (rRNA) using DOGMA (<https://dogma.cccb.utexas.edu/>)<sup>56</sup>. The accuracy of the start and stop codons and intron–exon boundaries were manually annotated based on previously annotated information



from the close relative *A. thaliana*. The complete structure of tRNA genes was validated using tRNAscan-SE v1.2.1<sup>57</sup>. The systematic circular view of the chloroplast genome was created using OGDRAW and in-house customized perl script<sup>58</sup>. Comparative syntenic maps were generated using circos following the BlastZ annotation. A chloroplast-based browser was developed for systematic analysis of the chloroplast genomes of the 28 *Brassica* and *Raphanus* genotypes, which can be accessed at [www.phyzen.co.kr/cpbrowser](http://www.phyzen.co.kr/cpbrowser). The chloroplast browser also contains sequence and gene annotation information for all 28 genotypes. Similarly, 45SnrDNA genes (18S, 5.8S, and 26S) were annotated based on Blast analyses and reported reference units. The mvista tool was used to visualize comparative syntenic relationships with other genotypes<sup>59</sup>. Complete chloroplast genomes and 40 complete 45SnrDNA sequences from 28 genotypes were deposited in GenBank (Table 1).

**Structural variations and PCR analysis of chloroplast and 45SnrDNA.** Extensive manual curation of chloroplast and 45SnrDNA revealed different kinds of non-redundant sequence variations (SV) such as SNPs, InDels, and copy number variations. Inter-species and intra-species structural variations were analyzed for chloroplast and 45SnrDNA sequences from 28 *Brassica* and *Raphanus* genotypes (Tables S2–S5). Putative SNPs and InDels were manually analyzed using the file aligned with MEGA7. Tandem repeats were identified using the Tandem repeats finder (TRF) tool. To detect highly reliable variations, all predicted variations were manually curated for both chloroplast and 45SnrDNA. Some of the randomly selected and highly informative variations were validated by PCR analysis.

To validate the polymorphic regions of chloroplast and 45SnrDNA sequences, specific primers were developed for high-quality structural variations such as SNPs and InDels (Table S7). DNA templates from 28 genotypes were used for target analysis. Each PCR reaction contained 10 ng template DNA, 10 pM primers, 0.5 μM dNTPs, 2 units of Taq polymerase (TAKARA, Japan), and the final volume brought to 20 μl with sterile distilled water. The PCR reactions were 10 min at 95 °C; followed by 36 cycles of 30S at 94 °C, 30S at 55–62 °C, and 30S at 72 °C; with a final extension at 72 °C for 5 min. Amplified fragments were checked with 2% agarose gel electrophoresis to estimate the product size.

**Phylogenetic analysis and divergence estimation based on chloroplast genomes and 45SnrDNA sequences.** Complete chloroplast genomes and 45SnrDNA sequences were independently explored for phylogenetic and divergence analysis. Chloroplast sequences of 28 *Brassica* and *Raphanus* genotypes were aligned with a previously reported *Brassica* chloroplast sequence using MAFFT (<http://mafft.cbrc.jp/alignment/software/>). Phylogenetic trees were constructed in MEGA7 using the neighbor-joining iterative model with 1,000 bootstrap replications<sup>60</sup>. Phylogenetic analysis was performed for 40 types of 45SnrDNA sequences based on 28 genotypes. *A. thaliana* chloroplast and 45SnrDNA sequences were used as an outgroup for the phylogenetic analysis. The reference chloroplast sequence with its annotation of *A. thaliana*, *B. rapa*, *B. oleracea*, *B. nigra*, *B. juncea*, *B. carinata*, and *Raphanus sativus* was obtained from GenBank.

Chloroplast and 45SnrDNA sequences from 28 genotypes were subjected to tree topology analysis and divergence time estimation using Bayesian methods implemented in BEAST (<http://beast.bio.ed.ac.uk/>)<sup>61</sup>. The BEAST program assumes auto-correlation, and is widely used to estimate the uncertainty of divergence dates and branch lengths, to estimate divergence using known speciation dates, and to accommodate the branching rate. The GTR + I + G substitution model was used to construct the tree topology and divergence time. We used an uncorrelated lognormal relaxed clock model to perform 10,000,000 generations of Markov chain Monte Carlo (MCMC) analysis with sampling every 1,000 generations. A Yule tree prior were used to generate the random starting tree. Tracer v. 1.6 was used to obtain the BEAST run after discarding 10% of the generations as burn-in. The remaining BEAST runs were used for the posterior possibilities. The divergence time was estimated using Tree annotator. *A. thaliana* was constrained as the outgroup, and the age of divergence between *A. thaliana* and *Brassica* lineages was constrained by a normal distribution with a mean of 17 million years (my) and standard deviation of 2 my<sup>9</sup>.

**Availability of data and materials.** All data generated or analysed during this study were obtained from the accession numbers provide at Tables 1 and S6.

## References

- Cheng, F., Wu, J. & Wang, X. Genome triplication drove the diversification of Brassica plants. *Horticulture Research* **1**, 14024 (2014).
- Cheng, F. *et al.* Genome sequencing supports a multi-vertex model for Brassicaceae species. *Current opinion in plant biology* **36**, 79–87, <https://doi.org/10.1016/j.cpb.2017.01.006> (2017).
- U, N. Genome analysis in Brassica with special reference to the experimental formation of *B. napus* and peculiar mode of fertilization. *Jap J Bot* **7**, 389–452 (1935).
- Wang, X. *et al.* The genome of the mesopolyploid crop species *Brassica rapa*. *Nature genetics* **43**, 1035–1039, <https://doi.org/10.1038/ng.919> (2011).
- Liu, S. *et al.* The Brassica oleracea genome reveals the asymmetrical evolution of polyploid genomes. *Nature communications* **5**, 3930, <https://doi.org/10.1038/ncomms4930> (2014).
- Chalhoub, B. *et al.* Early allopolyploid evolution in the post-Neolithic Brassica napus oilseed genome. *Science* **345**, 950–953 (2014).
- Parkin, I. A. *et al.* Transcriptome and methylome profiling reveals relics of genome dominance in the mesopolyploid Brassica oleracea. *Genome biology* **15**, R77, <https://doi.org/10.1186/gb-2014-15-6-r77> (2014).
- Yang, J. *et al.* The genome sequence of allopolyploid Brassica juncea and analysis of differential homoeolog gene expression influencing selection. *Nature genetics* **48**, 1225–1232, <https://doi.org/10.1038/ng.3657> (2016).
- Yang, T.-J. *et al.* Sequence-level analysis of the diploidization process in the triplicated FLOWERING LOCUS C region of Brassica rapa. *The Plant cell* **18**, 1339–1347 (2006).
- Koenig, D. & Weigel, D. Beyond the thale: comparative genomics and genetics of Arabidopsis relatives. *Nature reviews. Genetics* **16**, 285–298, <https://doi.org/10.1038/nrg3883> (2015).
- Mun, J. H. *et al.* Genome-wide comparative analysis of the Brassica rapa gene space reveals genome shrinkage and differential loss of duplicated genes after whole genome triplication. *Genome biology* **10**, R111, <https://doi.org/10.1186/gb-2009-10-10-r111> (2009).

12. Palmer, J. D. In *Cell organelles* 99–133 (Springer, 1992).
13. Birky, C. W. Uniparental inheritance of mitochondrial and chloroplast genes: mechanisms and evolution. *Proceedings of the National Academy of Sciences* **92**, 11331–11338 (1995).
14. Reboud, X. & Zeyl, C. Organelle inheritance in plants. *Heredity* **72**, 132–140 (1994).
15. Palmer, J. D., Shields, C., Cohen, D. & Orton, T. Chloroplast DNA evolution and the origin of amphidiploid Brassica species. *Theoretical and Applied Genetics* **65**, 181–189 (1983).
16. Moore, M. J., Soltis, P. S., Bell, C. D., Burleigh, J. G. & Soltis, D. E. Phylogenetic analysis of 83 plastid genes further resolves the early diversification of eudicots. *Proceedings of the National Academy of Sciences* **107**, 4623–4628, <https://doi.org/10.1073/pnas.0907801107> (2010).
17. Li, X. *et al.* Plant DNA barcoding: from gene to genome. *Biological Reviews* **90**, 157–166 (2015).
18. Nikiforova, S. V., Cavalieri, D., Velasco, R. & Goremykin, V. Phylogenetic analysis of 47 chloroplast genomes clarifies the contribution of wild species to the domesticated apple maternal line. *Molecular biology and evolution* **30**, 1751–1760 (2013).
19. Zhang, Y. *et al.* The complete chloroplast genome sequences of five *Epimedium* species: lights into phylogenetic and taxonomic analyses. *Frontiers in plant science* **7** (2016).
20. Qiao, J. *et al.* High-throughput multiplex cpDNA resequencing clarifies the genetic diversity and genetic relationships among *Brassica napus*, *Brassica rapa* and *Brassica oleracea*. *Plant biotechnology journal* **14**, 409–418 (2016).
21. Sharma, S. *et al.* Two plastid DNA lineages—*Rapa/Oleracea* and *Nigra*—within the tribe Brassiceae can be best explained by reciprocal crosses at hexaploidy: evidence from divergence times of the plastid genomes and R-block genes of the A and B genomes of *Brassica juncea*. *PLoS one* **9**, e93260 (2014).
22. Franzke, A., Lysak, M. A., Al-Shehbaz, I. A., Koch, M. A. & Mummenhoff, K. Cabbage family affairs: the evolutionary history of Brassicaceae. *Trends in plant science* **16**, 108–116 (2011).
23. Li, P. *et al.* A Phylogenetic Analysis of Chloroplast Genomes Elucidates the Relationships of the Six Economically Important Brassica Species Comprising the Triangle of U. *Frontiers in plant science* **8**, 111 (2017).
24. Koo, D. H. *et al.* Rapid divergence of repetitive DNAs in Brassica relatives. *Genomics* **97**, 173–185, <https://doi.org/10.1016/j.ygeno.2010.12.002> (2011).
25. Lim, K. B. *et al.* Characterization of rDNAs and tandem repeats in the heterochromatin of *Brassica rapa*. *Molecules and cells* **19**, 436–444 (2005).
26. Waminal, N. E. *et al.* In *The Brassica rapa Genome* 83–96 (Springer, 2015).
27. Hasterok, R., Jenkins, G., Langdon, T., Jones, R. N. & Maluszynska, J. Ribosomal DNA is an effective marker of Brassica chromosomes. *Theoretical and Applied Genetics* **103**, 486–490 (2001).
28. Warwick, S. I., Mummenhoff, K., Sauder, C. A., Koch, M. A. & Al-Shehbaz, I. A. Closing the gaps: phylogenetic relationships in the Brassicaceae based on DNA sequence data of nuclear ribosomal ITS region. *Plant Systematics and Evolution* **285**, 209–232 (2010).
29. Atwell, S. *et al.* Genome-wide association study of 107 phenotypes in *Arabidopsis thaliana* inbred lines. *Nature* **465**, 627–631 (2010).
30. Yang, J. *et al.* Comparative mitochondrial genome analysis reveals the evolutionary rearrangement mechanism in Brassica. *Plant biology* **18**, 527–536, <https://doi.org/10.1111/plb.12414> (2015).
31. Kim, K. *et al.* Comprehensive Survey of Genetic Diversity in Chloroplast Genomes and 45S nrDNAs within italic *Panax ginseng* / italic Species. *PLoS one* **10**, e0117159, <https://doi.org/10.1371/journal.pone.0117159> (2015).
32. Kim, K. *et al.* Complete chloroplast and ribosomal sequences for 30 accessions elucidate evolution of *Oryza AA* genome species. *Scientific reports* **5**, 15655 (2015).
33. Mandáková, T. & Lysak, M. A. Chromosomal phylogeny and karyotype evolution in  $x = 7$  crucifer species (Brassicaceae). *The Plant cell* **20**, 2559–2570 (2008).
34. Mitsui, Y. *et al.* The radish genome and comprehensive gene expression profile of tuberous root formation and development. *Scientific reports* **5**, 10835, <https://doi.org/10.1038/srep10835> (2015).
35. Seol, Y.-J. *et al.* The complete chloroplast genome of two Brassica species, *Brassica nigra* and *B. oleracea*. *Mitochondrial DNA*, 1–2, <https://doi.org/10.3109/19401736.2015.1115493> (2015).
36. Park, J. Y. *et al.* Complete mitochondrial genome sequence and identification of a candidate gene responsible for cytoplasmic male sterility in radish (*Raphanus sativus* L.) containing DCGMS cytoplasm. *TAG. Theoretical and applied genetics. Theoretische und angewandte. Genetik* **126**, 1763–1774, <https://doi.org/10.1007/s00122-013-2090-0> (2013).
37. Chang, S. *et al.* Mitochondrial genome sequencing helps show the evolutionary mechanism of mitochondrial genome formation in Brassica. *BMC genomics* **12**, 497 (2011).
38. Bennett, R. I. & Smith, A. G. Use of a genomic clone for ribosomal RNA from *Brassica oleracea* in RFLP analysis of Brassica species. *Plant molecular biology* **16**, 685–688 (1991).
39. Sang, T., Crawford, D. J. & Stuessy, T. F. Documentation of reticulate evolution in peonies (*Paeonia*) using internal transcribed spacer sequences of nuclear ribosomal DNA: implications for biogeography and concerted evolution. *Proceedings of the National Academy of Sciences of the United States of America* **92**, 6813–6817 (1995).
40. O’Kane Jr, S. L., Schaal, B. A. & Al-Shehbaz, I. A. The origins of *Arabidopsis suecica* (Brassicaceae) as indicated by nuclear rDNA sequences. *Systematic Botany*, 559–566 (1996).
41. Poczai, P. & Hyvonen, J. Nuclear ribosomal spacer regions in plant phylogenetics: problems and prospects. *Molecular biology reports* **37**, 1897–1912, <https://doi.org/10.1007/s11033-009-9630-3> (2010).
42. Idziak, D. & Hasterok, R. Cytogenetic evidence of nucleolar dominance in allotetraploid species of *Brachypodium*. *Genome* **51**, 387–391, <https://doi.org/10.1139/G08-017> (2008).
43. Tucker, S., Vitins, A. & Pikaard, C. S. Nucleolar dominance and ribosomal RNA gene silencing. *Current opinion in cell biology* **22**, 351–356, <https://doi.org/10.1016/j.ceb.2010.03.009> (2010).
44. Earley, K. *et al.* Erasure of histone acetylation by *Arabidopsis* HDA6 mediates large-scale gene silencing in nucleolar dominance. *Genes & development* **20**, 1283–1293, <https://doi.org/10.1101/gad.1417706> (2006).
45. Preuss, S. B. *et al.* Multimegabase silencing in nucleolar dominance involves siRNA-directed DNA methylation and specific methylcytosine-binding proteins. *Mol Cell* **32**, 673–684, <https://doi.org/10.1016/j.molcel.2008.11.009> (2008).
46. Fuertes Aguilar, J., Rosselló, J. & Nieto Feliner, G. Nuclear ribosomal DNA (nrDNA) concerted evolution in natural and artificial hybrids of *Armeria* (Plumbaginaceae). *Molecular ecology* **8**, 1341–1346 (1999).
47. Bao, Y., Wendel, J. F. & Ge, S. Multiple patterns of rDNA evolution following polyploidy in *Oryza*. *Molecular phylogenetics and evolution* **55**, 136–142, <https://doi.org/10.1016/j.ympev.2009.10.023> (2010).
48. Kotseruba, V. *et al.* The evolution of the hexaploid grass *Zingieriakochii* (Mez) Tzvel. ( $2n = 12$ ) was accompanied by complex hybridization and uniparental loss of ribosomal DNA. *Molecular phylogenetics and evolution* **56**, 146–155, <https://doi.org/10.1016/j.ympev.2010.01.003> (2010).
49. Dobesova, E. *et al.* Silenced rRNA genes are activated and substitute for partially eliminated active homeologs in the recently formed allotetraploid, *Tragopogon mirus* (Asteraceae). *Heredity (Edinb)* **114**, 356–365, <https://doi.org/10.1038/hdy.2014.111> (2015).
50. Bailey, C. D. *et al.* Toward a global phylogeny of the Brassicaceae. *Molecular biology and evolution* **23**, 2142–2160, <https://doi.org/10.1093/molbev/msl087> (2006).
51. Allender, C. J. & King, G. J. Origins of the amphidiploid species *Brassica napus* L. investigated by chloroplast and nuclear molecular markers. *BMC plant biology* **10**, 54 (2010).

52. Cheng, F. *et al.* Subgenome parallel selection is associated with morphotype diversification and convergent crop domestication in *Brassica rapa* and *Brassica oleracea*. *Nature genetics* **48**, 1218–1224 (2016).
53. Allen, G., Flores-Vergara, M., Krasynanski, S., Kumar, S. & Thompson, W. A modified protocol for rapid DNA isolation from plant tissues using cetyltrimethylammonium bromide. *Nature protocols* **1**, 2320–2325 (2006).
54. Seol, Y.-J., Lee, T.-H., Park, D.-S. & Kim, C.-K. NABIC: A New Access Portal to Search, Visualize, and Share Agricultural Genomics Data. *Evolutionary bioinformatics online* **12**, 51 (2016).
55. Perumal, S. *et al.* Elucidating the major hidden genomic components of the A, C, and AC genomes and their influence on Brassica evolution. *Scientific reports* **7**, 17986 (2017).
56. Wyman, S. K., Jansen, R. K. & Boore, J. L. Automatic annotation of organellar genomes with DOGMA. *Bioinformatics* **20**, 3252–3255 (2004).
57. Schattner, P., Brooks, A. N. & Lowe, T. M. The tRNAscan-SE, snoscan and snoGPS web servers for the detection of tRNAs and snoRNAs. *Nucleic acids research* **33**, W686–W689 (2005).
58. Lohse, M., Drechsel, O. & Bock, R. OrganellarGenomeDRAW (OGDRAW): a tool for the easy generation of high-quality custom graphical maps of plastid and mitochondrial genomes. *Current genetics* **52**, 267–274 (2007).
59. Frazer, K. A., Pachter, L., Poliakov, A., Rubin, E. M. & Dubchak, I. VISTA: computational tools for comparative genomics. *Nucleic acids research* **32**, W273–W279 (2004).
60. Kumar, S., Stecher, G. & Tamura, K. MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *Molecular biology and evolution*, <https://doi.org/10.1093/molbev/msw054> (2016).
61. Drummond, A. J., Suchard, M. A., Xie, D. & Rambaut, A. Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Molecular biology and evolution* **29**, 1969–1973, <https://doi.org/10.1093/molbev/mss075> (2012).

## Acknowledgements

This study was conducted with support from the Research Program for Agricultural Science and Technology Development (Project no. PJ010112) of the NAAS, and from the Cooperative Research Program for Agriculture Science & Technology Development (SSAC, Grant no. PJ011650), RDA, Republic of Korea. This work was also partially supported by a Golden Seed Project (Center for Horticultural Seed Development, No. 213003-04-4-SB430), Ministry of Agriculture, Food and Rural Affairs (MAFRA), Cooperative Research Program for Agriculture Science & Technology Development (Project No. PJ01100801), Rural Development Administration, Republic of Korea. The funding body has no role in the design of the study, analysis, and writing the manuscript.

## Author Contributions

C.K., Y.S., S.P. and T.J.Y. planned and designed the research. Y.S., C.K., S.S., J.L., N.E.W., M.J., S.C.L., S.J. Y.Y., H.K., J.C., K.R., and B.C. contributed materials and chloroplast genome assembly and comparative analysis. Y.S. and C.K. conducted genome sequencing. S.P. and T.J.Y. analyzed data and interpreted the results. S.P., C.K., Y.S. and T.J.Y. wrote the manuscript and I.P. and S.S. edited the manuscript.

## Additional Information

**Supplementary information** accompanies this paper at <https://doi.org/10.1038/s41598-018-25585-4>.

**Competing Interests:** The authors declare no competing interests.

**Publisher's note:** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2018