

NIH Public Access

Author Manuscript

Published in final edited form as: J Am Chem Soc. 2006 June 28; 128(25): 8112-8113.

Reciprocity of Steric and Stereoelectronic Effects in the Collagen Triple Helix

Matthew D. Shoulders[†], Jonathan A. Hodges^{†,‡}, and Ronald T. Raines^{†,§}

[†]Department of Chemistry, University of Wisconsin, Madison, Wisconsin 53706

Spepartment of Biochemistry, University of Wisconsin, Madison, Wisconsin 53706

Steric and stereoelectronic effects play a defining role in molecular conformation and reactivity. In small molecules, steric and stereoelectronic effects often have dichotomous consequences. For example, the anomeric effect in glycosides yields axial substituents that are disfavored by sterics.¹ Similarly, replacing the steric effect of a methyl group with the stereoelectronic effect of a fluoro group enables a β -peptide to fold.²

Stereoelectronic effects contribute markedly to the conformational stability of an abundant protein: collagen.³⁴⁵ Collagen is a fibrous protein comprising a bundle of three parallel strands folded into polyproline type II helices.⁶ Each strand consists of ~300 repeats of the unit: Xaa-Yaa-Gly, where Xaa is often (2S)-proline (Pro) and Yaa is often (2S,4R)-4-hydroxyproline (Hyp). The pyrrolidine ring in the Xaa and Yaa positions have C^{γ} -endo and C^{γ} -exo ring puckers, respectively.⁷ These puckers can be preordained by a stereoelectronic effect. Specifically, the gauche effect from a 4S fluoro group stabilizes the C^{γ}-endo pucker; that from a 4R fluoro group stabilizes the C^y-exo pucker (Figure 1).³ These stereoelectronic effects can marked ly enhance the conformational stability of a collagen triple helix. We reasoned that pyrrolidine ring pucker could instead be fixed and hence collagen stability enhanced by steric rather than stereoelectronic effects. Herein, we report on the bestowal of conformational stability to collagen by steric effects that reiterate stereoelectronic effects.

Density functional theory indicated that the pyrrolidine ring of (2S,4R)-4-methylproline (mep) has a strong preference (1.4 kcal/mol) for the C^{γ}-endo pucker and that of (2S,4S)-4methylproline (Mep) has a strong preference (1.7 kcal/mol) for the C^{γ}-exo pucker (Figure 1). These conformational preferences are observed in crystalline Ac-mep-NHMe and Ac-Mep-NHMe,⁸ and follow the trend observed in 4-*tert*-butylprolines.⁹ In the preferred conformations, the methyl group of mep and Mep adopts a pseudo-equatorial conformation. 10 A methyl group in this conformation should protrude radially from a collagen triple helix and thus not instill any deleterious interstrand steric interactions. Accordingly, we synthesized mepOH and MepOH¹¹ and incorporated these nonnatural amino acids into collagen strands to yield: (mep-Pro-Gly)7, (Pro-Mep-Gly)7, and (mep-Mep-Gly)7. We incubated solutions of each strand at <4 °C, and then used circular dichroism (CD) spectroscopy to detect formation of triple helices and assess their conformational stability.

(mep-Pro-Gly)7, (Pro-Mep-Gly)7, and (mep-Mep-Gly)7 formed triple helices at 4 °C, as indicated by an ellipiticity maximum near 225 nm (Figure 2A). The self-association of (Pro-Mep-Gly)₇, (mep-Mep-Gly)₇, and, to a lesser extent, (mep-Pro-Gly)₇ at 4 °C was confirmed by sedimentation equilibrium experiments.¹² (mep-Pro-Gly)₇, (Pro-Mep-Gly)₇, and (mep-

E-mail: raines@biochem.wisc.edu. [‡]Present address: Affinergy, Inc., 21 Davis Dr., Research Triangle Park, NC 27709.

Mep-Gly)₇ triple helices had $T_{\rm m}$ values of 13, 29, and 36 °C, respectively (Table 1), which are much greater than that of (Pro-Pro-Gly)₇. Thus, we conclude that steric effects can indeed stabilize the collagen triple helix.¹³

Mep in the Yaa position confers more stability to a triple helix than does mep in the Xaa position (Table 1). Likewise, (2S,4R)-4-fluoroproline (Flp) in the Yaa position increases triple-helical propensity more than does (2S,4S)-4-fluoroproline (flp) in the Xaa position.^{3,14} We suspected that this dichotomy could arise from the effect of the steric and stereoelectronic effects on the peptide bond itself. The trans:cis ratio of Ac-Pro-OMe in D₂O is only $K_{\text{trans/cis}} = 4.6^{3a}$. Yet, all peptide bonds in the collagen triple helix are in the trans conformation ($\omega = 180^\circ$).⁶

To determine the effect of a 4-methyl group on the value of $K_{\text{trans/cis}}$, we synthesized [¹³CH₃] Ac-mep-OMe and [¹³CH₃]Ac-Mep-OMe and evaluated $K_{\text{trans/cis}}$ with ¹³C NMR spectroscopy. The trans:cis ratio was twofold greater for Ac-Mep-OMe ($K_{\text{trans/cis}} = 7.4$) than for Ac-mep-OMe ($K_{\text{trans/cis}} = 3.6$). These data provide an explanation for triple helices formed by (Pro-Mep-Gly)₇ being more stable than those formed by (mep-Pro-Gly)₇¹⁵. Apparent ly, a balance exists between preorganization of the proper ring pucker and stabilization of a trans peptide bond.¹⁶

Our findings have numerous implications. Only recently were stereoelectronic effects found to contribute to the conformational stability of a protein.³ Herein, steric effects are shown to reiterate those same stereoelectronic effects. The stability of a nonnatural (mep-Mep-Gly)7 triple helix is indistinguishable from that of the "natural" (Pro-Hyp-Gly)₇ triple helix (Table 1), indicating that side-chain heteroatoms (and hence side-chain solvation) are not necessary for the formation of a stable triple helix. The stereoelectronic effects induced by heteroatoms are not additive in collagen. A (flp-Flp-Gly)₇ triple helix is less stable than is a (flp-Pro-Gly)₇ or (Pro-Flp-Gly)₇ triple helix (Table 1), presumably because of an unfavorable steric interaction between fluoro groups on adjacent strands.¹⁷ In contrast, the steric effects are additive, as a (mep-Mep-Gly)7 triple helix is more stable than is a (mep-Pro-Gly)7 or (Pro-Mep-Gly)7 triple helix (Table 1). The methyl groups of mep and Mep in synthetic collagen can likely be elaborated to larger functionalities without undesirable encumbrance. We imagine the creation of a new class of hyperstable collagen mimetics by the judicious integration of stereoelectronic and steric effects. The application of these venerable principles coupled with recent advances in the self-assembly of collagen fragments¹⁸ provides the means to create sturdy synthetic collagens for applications in biomedicine and biotechnology.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgment

We are grateful to F. W. Kotch, J. -C. Horng, D. R. McCaslin, F. Weinhold, and M. C. Pirrung for helpful discussion. M.D.S. was supported by a Department of Homeland Security (DHS) graduate fellowship. J.A.H. was supported by postdoctoral fellowship AR48057 (NIH). This work was supported by Grant AR44276 (NIH). CD and sedimentation equilibrium experiments were performed at the University of Wisconsin-Madison Biophysics Instrumentation Facility, which was established by Grants BIR-9512577 (NSF) and RR13790 (NIH).

References

- Thatcher, GRJ., editor. The Anomeric Effect and Associated Stereoelectronic Effects. American Chemical Society; Washington, DC: 1993.
- (2). Mathad RI, Gessier F, Seebach D, Jaun B. Helv. Chim. Acta 2005;88:266-280.
- (3)(a). Bretscher LE, Jenkins CL, Taylor KM, DeRider ML, Raines RT. J. Am. Chem. Soc 2001;123:777– 778. [PubMed: 11456609] (b) DeRider ML, Wilkens SJ, Waddell MJ, Bretscher LE, Weinhold F,

JAm Chem Soc. Author manuscript; available in PMC 2008 August 17.

Raines RT, Markley JL. J. Am. Chem. Soc 2002;124:2497–2505. [PubMed: 11890798] (c) Hodges JA, Raines RT. J. Am. Chem. Soc 2003;125:9262–9263. [PubMed: 12889933] (d) Doi M, Nishi Y, Uchiyama S, Nishiuchi Y, Nakazawa T, Ohkubo T, Kobayashi Y. J. Am. Chem. Soc 2003;125:9922–9923. [PubMed: 12914445]

- (4)(a). For reviews, see:Jenkins CL, Raines RT. Nat. Prod. Rep 2002;19:49–59. [PubMed: 11902439] (b) Raines RT. Protein Sci 2006;15:1219–1225. [PubMed: 16641494]
- (5). Analogous stereoelectronic effects have been observed in elastin-like proteins. See:Kim W, McMillan RA, Snyder JP, Conticello VP. J. Am. Chem. Soc 2005;127:18121–18132. [PubMed: 16366565]
- (6)(a). Ramachandran GN, Kartha G. Nature 1954;174:269–270. [PubMed: 13185286] (b)
 Ramachandran GN, Kartha G. Nature 1955;176:593–595. [PubMed: 13265783] (c) Rich A, Crick FHC. Nature 1955;176:915–916. [PubMed: 13272717] (d) Rich A, Crick FHC. J. Mol. Biol 1961;3:483–506. [PubMed: 14491907] (e) Bella J, Eaton M, Brodsky B, Berman HM. Science 1994;266:75–81. [PubMed: 7695699]
- (7). Vitagliano L, Berisio R, Mazzarella L, Zagari A. Biopolymers 2001;58:459–464. [PubMed: 11241217]
- (8). Flippen-Anderson JL, Gilardi R, Karle IL, Frey MH, Opella SJ, Gierasch LM, Goodman M, Madison V, Delaney NG. J. Am. Chem. Soc 1983;105:6609–6614.
- (9). Koskinen AMP, Helaja J, Kumpulainen ETT, Koivisto J, Mansikkamäkí H, Rissanen K. J. Org. Chem 2005;70:6447–6453. [PubMed: 16050708]
- (10). It is noteworthy that a pseudo-axial C^γ-H allows for greater σ→σ* hyperconjugative interactions with C^δ-N (a stereoelectronic effect) than does a pseudo-axial C^γ-CH₃.
 See:WeinholdFLandisCRValency and Bonding: A Natural Bond Orbital Donor-Acceptor Perspective2005Cambridge University PressCambridge, UK
- (11). Del Valle JR, Goodman M. J. Org. Chem 2003;68:3923-3931. [PubMed: 12737573]
- (12). See: Supporting information.
- (13). CD experiments in solutions containing the osmolyte trimethylamine-N-oxide confirm that triple helices of (mep-Pro-Gly)₇ have a T_m value near 13 °C, but the low molar ellipticity at 227 nm (Figure 2A) and the results of sedimentation equilibrium experiments suggest that (mep-Pro-Gly)₇ is only partially assembled at 4 °C (ref 12).
- (14)(a). Holmgren SK, Taylor KM, Bretscher LE, Raines RT. Nature 1998;392:666–667. [PubMed: 9565027] (b) Holmgren SK, Bretscher LE, Taylor KM, Raines RT. Chem. Biol 1999;6:63–70. [PubMed: 10021421]
- (15). The value of $K_{\text{trans/cis}}$ for a 4-substituted proline residue correlates with its ring pucker, an effect that is attributable to the stabilization of the trans isomer by an $n \rightarrow \pi^*$ interaction in the C[?]-exo conformation. See: ref ^{3b} andHinderaker MP, Raines RT. Protein Sci 2003;12:1188–1194. [PubMed: 12761389]
- (16). Mizuno K, Hayashi T, Peyton DH, Bächinger HP. J. Biol. Chem 2004;279:38072–38078. [PubMed: 15231845]
- (17)(a). Masamitsu D, Nishi Y, Uchiyama S, Nishiuchi Y, Nishio H, Nakazawa T, Ohkubo T, Kobayashi Y. J. Pept. Sci 2005;11:609–616. [PubMed: 15880478] (b) Hodges JA, Raines RT. J. Am. Chem. Soc 2005;127:15923–15932. [PubMed: 16277536]
- (18)(a). Paramonon SE, Gauba V, Hartgerink JD. Macromolecules 2005;38:7555–7561. (b) Kishimoto T, Morihara Y, Osanai M, Ogata S, Kamitakahara M, Ohtsuki C, Tanihara M. Biopolymers 2005;79:163–172. [PubMed: 16094625] (c) Koide T, Homma DL, Asada S, Kitagawa K. Bioorg. Med. Chem. Lett 2005;15:5230–5233. [PubMed: 16185864] (d) Kotch FW, Raines RT. Proc. Natl. Acad. Sci. U.S.A 2006;103:3028–3033. [PubMed: 16488977]

Shoulders et al.





Figure 1.

Ring conformations of 4-substituted proline residues. The C^{γ} -endo conformation is favored strongly by steric effects when $R_1 = Me$, $R_2 = H$ (mep) or stereoelectronic effects when $R_1 = H$ and $R_2 = F$ (flp). Similarly, the C^{γ} -exo conformation is favored strongly by steric effects when $R_1 = H$, $R_2 = CH_3$ (Mep) or stereoelectronic effects when $R_1 = OH$, $R_2 = H$ (Hyp) or $R_1 = F$, $R_2 = H$ (Flp).

Shoulders et al.





Figure 2.

Conformational analysis of (mep-Pro-Gly)₇, (Pro-Mep-Gly)₇, and (mep-Mep-Gly)₇ by CD spectroscopy. (A) Spectra of peptide solutions (0.2 mM in 50 mM acetic acid) incubated at \leq 4 °C for \geq 24 h. (B) Effect of temperature on the molar ellipticity at 225 nm ((Pro-Mep-Gly)₇ and (mep-Mep-Gly)₇) or 227 nm ((mep-Pro-Gly)₇). Data were recorded at intervals of 1 or 3 °C after equilibration for \geq 5 min.

J Am Chem Soc. Author manuscript; available in PMC 2008 August 17.

Table 1

Effect of 4-Methylproline and 4-Fluoroproline Diastereomers on the Conformational Stability of the Collagen Triple Helix

peptide	$T_{\rm m}$ (± 1°C)	ref
(Pro-Flp-Gly)7	45	3a
(mep-Mep-Gly) ₇	36	this work
(Pro-Hyp-Gly) ₇	36	3a 3c
(flp-Pro-Gly) ₇	33	
(Pro-Mep-Gly) ₇	29	this work
(mep-Pro-Gly) ₇	13	this work
(flp-Flp-Gly) ₇	8^a	17b
(Pro-Pro-Gly) ₇	-6^a	17b

 $^a\mathrm{Based}$ on the extrapolation of data from solutions containing TMAO.