

## Recommended Nomenclature for Five Mammalian Carboxylesterase Gene Families: Human, Mouse and Rat Genes and Proteins

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### **Abstract**

Mammalian carboxylesterase (*CES* or *Ces*) genes encode enzymes that participate in xenobiotic, drug and lipid metabolism in the body and are members of at least 5 gene families. Tandem duplications have added more genes for some families, particularly for mouse and rat genomes, which has caused confusion in naming rodent *Ces* genes. A new nomenclature system for human, mouse and rat carboxylesterase genes is described which identifies homolog gene families and allocates a unique name for each gene. The guidelines of human, mouse and rat gene nomenclature committees were followed and “*CES*” (human) or “*Ces*” (mouse and rat) root symbols were used followed by the family number (eg human *CES1*). Where multiple genes were identified for a family or where a clash occurred with an existing gene name, a letter was added (eg. human *CES4A*; mouse and rat *Ces1a*) which reflected gene relatedness among rodent species (eg. mouse and rat *Ces1a*). Pseudogenes were named by adding “*P*” and a number to the human gene name (eg. human *CES1P1*) or by using a new letter followed by *ps* for mouse and rat *Ces* pseudogenes (eg. *Ces2d-ps*). Gene transcript isoforms were named by adding the Genbank accession ID to the gene symbol (eg human *CES1\_AB119995* or mouse *Ces1e\_BC019208*). This nomenclature improves our understanding of human, mouse and rat *CES/Ces* gene families and facilitates research into the structure, function and evolution of these gene families. It also serves as a model for naming *CES* genes from other mammalian species.

## **INTRODUCTION**

Five families of mammalian carboxylesterases (CES; E.C.3.1.1.1) have been described including CES1, the major liver enzyme (Munger et al., 1991; Shibata et al. 1993; Ghosh 2000; Holmes et al., 2009a); CES2, the major intestinal enzyme (Langmann et al. 1997; Schewer et al. 1997; Holmes et al., 2009a); CES3, expressed in brain, liver and colon (Sanghani et al. 2004; Holmes et al., 2010); CES5 (also called CES7 or cauxin), a major urinary protein of the domestic cat also present in human tissues (Miyazaki *et al.* 2003; 2006; Holmes et al., 2008a; Zhang et al., 2009); and CES6, a predicted CES-like enzyme in brain (Clark et al., 2003; Holmes et al., 2009a) (reviewed by Williams et al., 2010). These enzymes catalyse hydrolytic and transesterification reactions with xenobiotics, anticancer pro-drugs, and narcotics (Satoh & Hosokawa 1998; 2006; Satoh *et al.* 2002; Ohtsuka *et al.* 2003; Redinbo & Potter 2005), the conversion of lung alveolar surfactant (Ruppert et al. 2006) and several lipid metabolic reactions

(Tsujita & Okuda 1993; Becker et al. 1994; Ghosh 2000; Hosokawa et al. 2007; Diczfalusy et al. 2001), and may assist with the assembly of low density lipoprotein particles in liver (Wang et al. 2007).

Structures for human and animal *CES* genes have been reported, including rodent *CES1* and *CES2* ‘like’ genes (Ghosh et al. 1995; Dolinsky et al. 2001; Hosokawa et al. 2007) and human *CES1* and *CES2* genes (Becker et al. 1994; Langmann et al. 1997; Ghosh 2000; Marsh et al. 2004). Predicted gene structures have been also described for the human *CES3*, *CES5* and *CES6* genes, which are localized with *CES1* and *CES2* in two contiguous *CES* gene clusters on human chromosome 16 (Holmes et al. 2008a; 2009a,b; 2010). In addition, a *CES1*-like pseudogene (currently designated as *CES4*) is located with the *CES1-CES5* gene cluster (Yan et al., 1999). Mammalian *CES* genes usually contain 12 to 14 exons of DNA encoding CES enzyme sequences which may be shuffled during mRNA synthesis, generating several *CES* transcripts and enzymes encoded by each of the *CES* genes (see Thierry-Mieg and Thierry-Mieg, 2006). There are significant sequence similarities for the five *CES* families, especially for key regions previously identified for human liver CES1 (Bencharit et al. 2003; 2006; Fleming et al. 2005). Three-dimensional structural analyses of human CES1 have identified three major ligand binding sites, including the broad specificity active site, the ‘side door’ and ‘Z-site’, where substrates, fatty acids and cholesterol analogues respectively, are bound; and an active site ‘gate’, which may facilitate product release following catalysis (Bencharit et al. 2003; 2006; Fleming et al. 2005).

Because of the confusion associated with the current nomenclature for mammalian *CES* genes, particularly for mouse and rat *CES* genes where significant gene duplication events have generated a large number of *Ces1*-like and *Ces2*-like genes (see Berning et al., 1985; Ghosh et al., 1995; Satoh & Hosokawa, 1995; Dolinsky et al., 2001; Hosokawa et al., 2007), this paper proposes a new nomenclature system which enables easy identification of *CES* family members for this enzyme. The nomenclature follows the guidelines of the human, mouse and rat gene nomenclature committees and allocates a new name for each human (*CES*) or mouse and rat (*Ces*) gene. It also names and identifies the gene family origin for identified *CES* pseudogenes and provides a system for naming transcript isoforms derived from each of the *CES* genes. The nomenclature has the flexibility to accommodate new human, mouse and rat *CES* genes and will assist further research into the structure, function and evolution of these gene families as well as serving as a model for naming *CES* genes from other mammalian species.

## **GUIDING PRINCIPLES FOR THE NEW *CES* NOMENCLATURE**

The new nomenclature system for human, mouse and rat *CES* genes and enzymes is based on the identification of homolog gene families and a subsequent allocation of a unique gene name for each of the genes observed from genome data bases or reported from previous studies. It follows the guidelines of the human, mouse and rat gene nomenclature committees and recommends the naming of homolog *CES* or *Ces* genes among species. The italicized root symbol “*CES*” for human and “*Ces*” for mouse and rat genes were used, followed by an number describing the gene family (examples include *CES1* for human *CES* family 1 or *Ces1* for mouse and rat *Ces* family 1 genes) (Tables 1-3). For mammalian genomes where multiple genes were identified or where a gene required a name which clashed with an existing name, a capital letter (for human genes) (eg. *CES4A*) or a lower case letter (for mouse and rat genes) (eg. *Ces1a*, *Ces1b* for multiple mouse *Ces1*-like genes) was added after the number. The letter used for multiple genes reflected the relatedness of the genes across species (eg. reflecting higher degrees of identity for mouse and rat *Ces1a* genes). When a human *CES* pseudogene was identified, a capital “P” and a number were added to the gene name (eg. *CES1P1*), whereas for mouse and rat *Ces* pseudogenes, a unique lower case letter was used followed by “-ps” (eg. *Ces2d-ps*). Transcript isoforms of human (*CES*), mouse and rat (*Ces*) gene transcripts were designated by following the gene name with the Genbank transcript ID, such as human *CES1\_ABI19997* and *CES1\_ABI187225* which differs from the current nomenclature used for human *CES1* isoforms (*CES1A1* and *CES1A2*, respectively) (see Table 1).

## **HUMAN *CES* GENES**

Table 1 summarizes the locations and exonic structures for human *CES* genes based upon previous reports for human *CES1* and *CES2* (Becker et al. 1994; Langmann et al. 1997; Ghosh 2000; Marsh et al. 2004) and predictions for human *CES3* (Holmes et al., 2010), *CES4A* (Holmes et al., 2009a) and *CES5A* (Holmes et al., 2008a) (the February 2009 human reference sequence (GRCh37) was used in this study) (Rhead et al., 2010). Human *CES1P1* (a *CES1*-like pseudogene), *CES1* and *CES5A* were located in a cluster (cluster 1) on chromosome 16 while *CES2*, *CES3* and *CES4A* were in a separate cluster (cluster 2) on the same chromosome. Cluster 1 *CES* genes (*CES1* and *CES5A*) were transcribed on the negative strand, whereas cluster 2 genes (*CES2*, *CES3* and *CES4A*) were transcribed on the positive strand. Figure 1 summarizes the predicted exonic start sites for human *CES* genes, with *CES1* and *CES4A* containing 14 exons; *CES3* and *CES5A* 13 exons; and *CES2* with 12 exons. These exon start sites were in identical or similar positions to those reported for *CES1* (Ghosh, 2000; March et al., 2004). Figure 2 shows the comparative structures for human *CES* reference sequences and transcripts described

on the AceView website (<http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/>) (Thierry-Mieg & Thierry-Mieg, 2006). The *CES* gene and transcript sequences varied in size from 11kb for *CES2* to 79kb for *CES5A*, and exhibited distinct structures in each case. Moreover, several isoforms were generated *in vivo* for each of the human *CES* genes which have different structures, as a result of transcriptional events including truncation of the 5' ends, differential presence or absence of exons, alternative splicing or retention of introns or overlapping exons with different boundaries. In addition, the isoforms are differentially expressed in tissues of the body and may perform distinctive metabolic roles. *CES* isoforms were named by using the gene name followed by the Genbank ID for the specific transcript. Recent studies of human *CES1* have described at least two major isoform transcripts, designated as *CES1A1* (*AB119997*) and *CES1A2* (*AB119996*) (Tanimoto et al., 2007). These isoforms have been redesignated as *CES1\_ABI19997* and *CES1\_ABI19997*, respectively (see Table 1) and encode sequences which differ by only 4 amino acid residues within the N-terminal region (exon 1) (Tanimoto et al., 2007). Distinct 5'-untranslated consensus sequences for binding transcription factors were reported which suggested differences in transcriptional regulation and functional roles in contributing to CPT-11 chemosensitivity for these isoforms (Tanimoto et al., 2007; Hosokawa et al., 2008; Yoshimura et al., 2008). Fukami and coworkers (2008) have also examined human *CES* isoform structure and proposed that *CES1P1*, a *CES1*-like pseudogene on chromosome 16 (designated as *CES1A3*), was derived from the *CES1\_ABI19997* isoform.

## **HUMAN CES AMINO ACID SEQUENCES AND STRUCTURES**

An alignment of the amino acid sequences for human CES-like protein subunits are shown in Figure 1, together with a description of several features for these enzymes. The sequences have been derived from previously reported sequences for *CES1* (Munger et al., 1991; Shibata et al., 1993); *CES2* (Langmann et al., 1997; Schewer et al., 1997); *CES3* (Sanghani et al., 2004); *CES4A* (previously *CES6* or *CES8*) (Holmes et al., 2009a); and *CES5A* (previously *CES7*) (Holmes et al., 2008a) (Table 1). Alignments of the human *CES* subunits showed between 39-46% sequence identities, which suggests that these are products of separate but related gene families, whereas sequence alignments of human *CES1* and *CES2* with mouse *CES1*-like and *CES2*-like subunits exhibited higher levels of sequence identities with the *CES* family homolog in each case (66-78% identities for human and mouse *CES1*-like subunits and 64-72% for human and mouse *CES2*-like subunits, respectively) (data not shown), suggesting that these are members of the same mammalian *CES* families, in each case. Similar results were observed for comparisons of human *CES3*, *CES4A* (previously *CES6* or *CES8*) and *CES5A* (previously

CES7) with the corresponding mouse CES homolog sequences, with 65%, 72% and 69% identities being observed, respectively. This supports the designation of these *CES* genes being members of the same family, in each case.

The amino acid sequences for the human CES subunits examined contained 567 (CES1), 559 (CES2), 571 (CES3), 561 (CES4A) and 575 (CES5A) residues (Figure 1). Previous studies on human CES1 have identified key residues which contribute to the catalytic, oligomeric, subcellular localization and regulatory functions for this enzyme (sequence numbers refer to human CES1). These included the catalytic triad for the active site (Ser221; Glu354; His468) (Cygler et al. 1993); disulfide bond forming residues (Cys87/Cys116 and Cys274/Cys285) (Lockridge et al. 1987); microsomal targeting sequences, including the hydrophobic N-terminus signal peptide (von Heijne 1983; Zhen et al. 1995; Potter et al. 1998) and the C-terminal endoplasmic reticulum (ER) retention sequence (His-Ile-Glu-Leu) (Robbi & Beaufay 1983); as well as ligand binding sites, including the 'Z-site' (Gly356), the 'side door' (Val424-Met425-Phe426) and the 'gate' (Phe550) residues (Bencharit et al. 2003; 2006; Fleming et al. 2005). Identical residues were observed for each of the human CES subunit families for the active site triad and disulfide bond forming residues although changes were observed for some key residues for CES1 subunits, including the 'side-door' and 'gate' of the active site, with family specific sequences or residues in each case; the 'Z-site' (Gly356 for human CES1) has been retained for human CES2 and CES5A sequences, but substituted for CES3 (Ser) and CES4A (Asn); the hydrophobic N-terminal sequence for human CES sequences has undergone major changes although this region retains a predicted signal peptide property; the human CES C-terminal tetrapeptide sequences have also changed, although CES2 (HTEL) and CES3 (QEDL) are similar in sequence with human CES1 (HIEL), which plays a role in the localization of human CES1 within endoplasmic reticulum membranes (Robbi & Beaufay, 1983).

Other key human CES1 sequences included two charge clamps which are responsible for subunit-subunit interaction, namely residues Lys78/Glu183 and Glu72/ which Arg186, contribute to the trimeric and hexameric structures for this enzyme (Bencharit et al. 2003; 2006; Fleming et al. 2005). Other human CES subunit sequences for these charge clamp sites included substitutions with neutral amino acids for the human CES2 and CES5A sequences, while the CES3 and CES4A sequences retained one potential clamp site (Figure 1). Pindel et al. (1997) and Holmes et al (2009b) have reported monomeric subunit structures for human and baboon CES2, which is consistent with the absence of charge clamps for this enzyme. This could have a major influence on the kinetics and biochemical roles for human CES isozymes since three dimensional studies have

indicated that ligand binding to the human CES1 'Z-site' shifts the trimer-hexamer equilibrium towards the trimer facilitating substrate binding and enzyme catalysis (Redinbo & Potter 2005). The N-glycosylation site for human CES1 at Asn79-Ala80-Thr81 (Kroetz et al. 1993; Bencharit et al. 2003; 2006; Fleming et al. 2005) was not retained for any of the other human CES sequences, although potential N-glycosylation sites were observed at other positions, including CES2 (site 3), CES3 (site 2), CES4A (sites 4, 5 and 7) and CES5A (sites 6, 8 and 9) (Table 4). Given the reported role of the N-glycosylated carbohydrate group contributing to CES1 stability and maintaining catalytic efficiency (Kroetz et al. 1993), the N-glycosylation sites predicted for other human CES subunits may perform similar functions or indeed may serve new functions specific to a particular CES family.

Predicted secondary structures for human CES2 (Holmes et al., 2009b), CES3 (Holmes et al., 2010), CES4A (Holmes et al., 2009a) and CES5A (Holmes et al., 2008a) sequences were compared with those reported for human CES1 and similar  $\alpha$ -helix  $\beta$ -sheet structures were observed for all of the CES subunits examined (Bencharit et al., 2003; 2006) (Figure 1). This was especially apparent near key residues or functional domains such as the  $\alpha$ -helix within the N-terminal signal peptide; the  $\beta$ -sheet and  $\alpha$ -helix structures near the active site Ser221 (human CES1) and 'Z-site' (Glu354/Gly356 respectively); the  $\alpha$ -helices bordering the 'side door' site; and the  $\alpha$ -helix containing the 'gate' residue (Phe550 for human CES1). The human CES5A sequence, however, contained a predicted helix at the hydrophobic C-terminus not observed for other CES subunits which may perform a family specific function. Predicted 3-D structures have been previously described for each of the human CES subunits (Holmes et al., 2008a; 2009a, b; 2010) which were similar to the human CES1 structure (Bencharit et al., 2003; 2006).

## **MOUSE *Ces* GENES AND ENZYMES**

Table 2 summarizes the proposed names, locations and overall structures for the *Ces* genes observed for the mouse genome (July 2007 mouse [*Mus musculus*] genome data obtained from the Build 37 assembly by NCBI and the Mouse Genome Sequencing Consortium) (<http://www.ncbi.nlm.nih.gov> was used in this study). The italicized gene name, '*Ces*', is consistent with other mouse gene nomenclature and is preferred to the '*CES*' stem used for human genes. At least 20 mouse *Ces* genes are recognized on the Mouse Genome Database (<http://www.informatics.jax.org/>) (MGI) and further described in terms of their locations on mouse chromosome 8, the number of predicted exons for each gene, predicted strand for transcription, number of amino acid residues and subunit MWs for the encoded CES subunits, and identification symbols from MGI (eg.



MGI3648919 for *Ces1a*), NCBI (Reference Sequences were identified from the National Center for Biotechnology Information database) (<http://www.ncbi.nlm.nih.gov/>), Vega (the Vertebrate Genome Annotation (VEGA) database) (<http://vega.sanger.ac.uk/index.html>), UNIPROT (Universal Protein Resource) (<http://www.ebi.ac.uk/uniprot/>) and Ensembl (Genome Database) (<http://www.ensembl.org/>) database sources.

Eight *Ces1*-like genes are located in tandem within a 360 kilobase segment of mouse chromosome 8, with an average gene size of 28 kilobases. The names for these genes (*Ces1a*, *Ces1b*...*Ces1h*) are allocated in the same order as their locations on the mouse genome (Table 3). The *Ces1*-like gene cluster is also located near to the mouse *Ces5a* gene, which is comparable to the *CES1P1-CES1-CES5A* cluster observed for human chromosome 16. Each of these genes contained 13 or 14 exons predicted for transcription on the negative strand, and with encoded CES subunits exhibiting distinct but similar amino acid sequences (554-567 residues). The subunits were 63-85% identical with each other and with the human CES1 sequence, which is consistent with these being members of the mouse *Ces1* gene family. Mouse *Ces1*-like genes included several that have been previously investigated, including *Ces1c* (previously called *Es1*), encoding a major mouse plasma esterase with 554 amino acid residues and also exhibiting lung surfactant convertase activity (Genetta et al., 1988; Krishnasamy et al., 1998); *Ces1d* (previously *Ces3*), encoding a mouse liver enzyme with 565 residues exhibiting triacylglycerol hydrolase activity (Dolinsky et al., 2001); *Ces1e* (previously called *Es22* or *egasyn*) encoding a liver CES with 562 residues exhibiting  $\beta$ -glucuronidase-binding properties (Ovnic et al., 1991); and *Ces1g* (previously *Ces1*) encoding a liver CES which exhibits lipid metabolising activity and has 565 amino acid residues (Table 5) (Ellingham et al., 1998).

Eight *Ces2*-like genes were also observed in a second 286 kb gene cluster on mouse chromosome 8 with an average gene size of ~8 kilobases (Table 2). These genes were named according to their sequence of position on the mouse genome (*Ces2a*, *Ces2b*...*Ces2h*), and included a pseudogene designated as *Ces2d-ps*. Three of these mouse *Ces2*-like genes have been previously described including *Ces2c* (previously *Ces2*), which encodes an inducible liver acyl-carnitine hydrolase enzyme with 561 residues (Furihata et al., 2003); *Ces2e* (previously *Ces5*) encoding a liver and intestinal enzyme with 560 amino acid residues (The MGC Project Team, 2004); and *Ces2a* (previously *Ces6*), encoding a liver and colon enzyme with 558 residues (The MGC Project Team, 2004). The *Ces2*-like cluster was located alongside two *Ces3*-like mouse genes (*Ces3a* and *Ces3b*) and a *Ces4a* gene (Table 3), which is comparable with the *CES2-CES3-CES4A* gene cluster on human chromosome 16 (Table 1). The *Ces3a* gene (previously mouse *esterase 31*

or *Est31*) is expressed strongly in male mouse livers and encodes a 554 residue CES3-like subunit (Aida *et al.*, 1993), whereas the *Ces3b* gene (previously *Es31L* or *EG13909*) is also expressed in liver encoding a 568 residue subunit (The MGC Project Team, 2004). The *Ces4a* gene (previously called *EST8* or *Ces8*) encodes an enzyme predicted for secretion in epidermal cells with 563 amino acid residues showing 72% identity with human CES4A (The MGC Project Team, 2004).

## **RAT *Ces* GENES AND ENZYMES**

Table 3 summarizes the proposed names, locations and structures for *Ces* genes observed for the rat genome (the November 2004 rat (*Rattus norvegicus*) genome assembly based on version 3.4 produced by the Baylor Human Genome Sequencing Center) (Gibbs *et al.*, 2004 was used in this study). Fifteen rat *Ces* genes were identified on the Rat Genome Database (RGD) (<http://rgd.mcw.edu/>) and further characterized by their locations on rat chromosomes 1 and 19, the number of predicted exons for each gene, the predicted strand for transcription, current gene symbols, the number of amino acid residues and subunit MWs for the encoded CES subunits, and the identification symbols from RGD (eg. RGD1583671 for *Ces1a*), NCBI Reference Sequences (<http://www.ncbi.nlm.nih.gov/>), Vega (<http://vega.sanger.ac.uk/index.html>), UNIPROT (<http://www.ebi.ac.uk/uniprot/>) and Ensembl (<http://www.ensembl.org/>) database sources.

Five *Ces1*-like genes were located in tandem within a 201 kilobase segment of rat chromosome 19, with an average gene size of 33 kilobases (Table 3). The names for these genes (*Ces1a*, *Ces1c*...*Ces1f*) were allocated according to their degree of identity with the corresponding mouse *Ces1*-like genes (Table 3). The genes were located in tandem in the same order as the mouse *Ces1*-like genes and were near the rat *Ces5a* gene. This is comparable to the *CES1P1-CES1A-CES5A* gene cluster observed for human chromosome 16. The rat *Ces1*-like genes contained 14 exons, and were predicted for transcription on the positive strand with encoded CES subunits exhibiting similar amino acid sequences (550-565 residues). The subunits were 65-73% identical with each other and with the human CES1 sequence, which is consistent with membership of the rat *Ces1* gene family. The encoded rat *Ces1*-like subunit sequences showed higher levels of identity with the corresponding mouse *Ces1*-like sequences (81-92% for rat and mouse CES1a, CES1c, CES1d, CES1e and CES1f amino acid sequences). At least 3 rat *Ces1*-like genes have been previously described, including *Ces1c* (previously called *Es1*) encoding a rat plasma esterase (Vanlith *et al.*, 1993; Sanghani *et al.*, 2002); *Ces1d* (previously *Ces3*) encoding a rat liver enzyme with 565 residues and exhibiting cholesteryl ester hydrolase

activity (Robbi et al., 1990; Ghosh et al., 1995); and *Ces1e* (previously called *ES-3* or *egasyn*), encoding a rat liver Ces with 561 residues and having  $\beta$ -glucuronidase-binding properties (Robbi & Beaufay, 1994).

Seven rat *Ces2*-like genes were observed on the rat genome which were localized on two chromosomes: chromosome 1 (*Ces2c* and *Ces2i*) and chromosome 19 in 3 locations: *Ces2a* and *Ces2e*; *Ces2j*; and *Ces2g* and *Ces2h* (Table 3). The genes were named according to the degree of sequence identity with the corresponding mouse *Ces2*-like genes. Rat *Ces2*-like genes have been previously investigated, including *Ces2c* (previously *Ces2*), encoding an inducible liver acyl-carnitine hydrolase enzyme containing 561 residues (Furihata et al., 2003); *Ces2e* (previously *Ces5*) encoding a liver and intestinal enzyme with 560 amino acid residues (The MGC Project Team, 2004); and *Ces2a* (previously *Ces6*), encoding a liver and colon enzyme with 558 residues (The MGC Project Team, 2004). The rat *Ces2*-like cluster was located alongside a *Ces3*-like gene (*Ces3a* and *Ces3b*) and a *Ces4a* gene (Table 3), which is comparable with the *CES2A-CES3A-CES4A* gene cluster on human chromosome 16 (Table 1).

## **FUNCTIONS OF MAMMALIAN CES FAMILIES**

Mammalian CES families exhibit broad substrate specificities and specific roles for these enzymes have been difficult to establish because of the promiscuity of the CES active site towards a wide range of substrates and the existence of multiple forms with overlapping specificities (see Leinweber, 1987; Satoh & Hosokawa, 1998; 2006; Redinbo & Potter, 2005; Fleming et al., 2005; Imai, 2006). Table 4 summarizes current knowledge concerning substrates and functions reported for human, mouse and rat *CES* gene family members.

Studies on human CES1 have examined its role in the metabolism of various drugs, including narcotics such as heroin and cocaine (Pindel et al, 1997; Bencharit et al, 2003), warfare nerve agents (Hemmert et al., 2010), psychostimulants (Sun et al, 2004), analgesics (Takai et al., 1997) and chemotherapy drugs (Sanghani et al., 2004). Mammalian liver is predominantly responsible for drug clearance from the body with CES1 and CES2 (with CES1 > CES2) playing major roles, following absorption of drugs into the circulation (Pindel et al. 1997; Imai 2006). Mammalian intestine (with CES2 > CES1) plays a major role in first pass clearance of several drugs, predominantly via CES2 in the ileum and jejunum (Imai et al. 2003). CES1 and CES2 also have different roles in prodrug activation, as shown for the anti-cancer drug irinotecan (CPT-11) which is converted to its active form SN-38 predominantly by CES2 (Humerickhouse et al. 2000). Recent modeling studies have shown that the human CES2 active site cavity is lined

with negatively charged residues which may explain the preference of this enzyme for neutral substrates (Vistoli et al., 2010). The role for human CES3 has not been extensively studied although the enzyme is capable of activating prodrugs such as irinotecan (Sanghani et al., 2004). There are no reports concerning the metabolic role(s) for human CES4A and functional studies on mammalian CES5 function are limited to feline species, where the enzyme is secreted into cat urine and apparently regulates the production of a cat specific amino acid 'felinine', a putative pheromone precursor (Miyazaki et al. 2006).

## **EVOLUTION OF MAMMALIAN *CES* GENE FAMILIES**

Recent comparative and evolutionary studies (Holmes et al., 2008b; Williams et al., 2010) have concluded that there are at least five major mammalian *CES* gene families. In addition, the gene duplication events which generated the ancestral mammalian *CES1*, *CES2*, *CES3*, *CES4* and *CES5* genes have apparently predated the common ancestor for marsupial and eutherian mammals (Holmes et al., 2008b) which has been estimated at ~ 173-193 million years ago (Woodburne *et al*, 2003) and may coincide with the early diversification of tetrapods approximately 350-360 million years ago (Donoghue & Benton, 2007). The mammalian *CES* gene families are ancient in their genetic origins and were established prior to the appearance of mammals during evolution. Further *CES/Ces* gene duplication events have subsequently occurred during mammalian evolution however, especially for rodent species, for which the mouse and rat *Ces1*-like and *Ces2*-like genes have apparently undergone successive duplication events. At least three of these are likely to have occurred in the common ancestor for rat and mouse during rodent evolution since several homolog genes and proteins were recognized, including *Ces1c* (previously *Es1*), *Ces1d* (*Ces3*), *Ces1e* (*Es22*), *Ces2a* (*Ces6*), *Ces2c* (*Ces2*) and *Ces2e* (*Ces5*) (Tables 3 and 4). With the exception of the rat *Ces2*-like genes, which were located in multiple clusters on chromosomes 1 and 19, human, mouse and rat *CES* genes were localized within 2 clusters of genes on the same chromosome, namely *Ces1-Ces5A* (with multiple *Ces1*-like genes) and *Ces2-Ces3-Ces4A* (with multiple *Ces2*-like genes in mouse and rat). The presence of two *Ces3*-like genes in the mouse suggests that a further duplication event also took place in this species.

## **CONCLUSIONS**

This paper has examined human, mouse and rat carboxylesterase genes and encoded subunits and has proposed a new nomenclature system identifying each of five gene families (designated as *CES1*, *CES2*...*CES5* for human genes or *Ces1*, *Ces2*...*Ces5* for mouse and rat genes) and allocating a unique gene name for each of the genes. The italicized root symbol “*CES*” for human and “*Ces*” for mouse and rat genes followed by a number for the family were used which is consistent with current practice. When multiple genes were identified for a gene family or where a gene required a name which clashed with an existing name, a capital letter (for human genes) (eg. *CES4A*) or a lower case letter (for mouse and rat genes) (eg. *Ces1a*, *Ces1b*) was added after the number. A human *CES* pseudogene was named using a capital “P” and a number (eg. *CES1P1*), whereas mouse and rat *Ces* pseudogenes were named with a unique lower case letter followed by “-ps” (eg. *Ces2d-ps*). This new nomenclature will also assist in naming multiple *CES* genes and proteins from other mammalian species. As an example, Holmes and co-workers (2009c) and Williams et al (2010) have reported multiple *CES1*-like genes on the horse genome which may be designated in accordance with the recommended nomenclature as *CES1A*, *CES1B*, *CES1C* etc in order of the tandem locations of these genes on chromosome 3. Transcript isoforms of *CES* gene transcripts were named by following the gene name with the Genbank ID for the specific transcript. This nomenclature will assist our understanding of the genetic relatedness and the *CES* family origins for individual human, mouse and rat *CES* genes and proteins and facilitate future research into the structure, function and evolution of these genes. It will also serve as a model for naming *CES* genes from other mammalian species.

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## **REFERENCES**

- Aida K, Moore R, Negishi M (1993) Cloning and nucleotide sequence of a novel, male-predominant carboxylesterase in mouse liver. *Biochim Biophys Acta* 1174, 72-74
- Barthel BL, Torres RC, Hyatt JL, Edwards CC, Hatfield MJ, Potter PM, Koch TH (2008) Identification of human intestinal carboxylesterase as the primary enzyme for activation of a doxazoline carbamate prodrug. *J Med Chem* 51, 298-304

- Becker A, Bottcher A, Lackner KJ, Fehring P, Notka F, Aslandis C, Schmithz C (1994) Purification, cloning and expression of a human enzyme with acyl coenzyme A: cholesterol acyltransferase activity, which is identical to liver carboxylesterase. *Arterioscler Thromb* 14, 1346-1355
- Bencharit S, Morton C.L, Xue Y, Potter PM, Redinbo MR (2003) Structural basis of heroin and cocaine metabolism by a promiscuous human drug-processing enzyme. *Nature Struct Biol* 10, 349-356
- Bencharit S, Edwards CC, Morton CL, Howard-Williams EL, Kuhn P, Potter PM, Redinbo MR (2006) Multisite promiscuity in the processing of endogenous substrates by human carboxylesterase 1. *J Mol Biol* 363, 201-214
- Berning W, De Looze SM, von Deimling O (1985) Identification and development of a genetically closely linked carboxylesterase gene family of the mouse liver. *Comp Biochem Physiol*, 80, 859-865
- Cyglar M, Schrag JD, Sussman JL, Harel M, Silman I, Gentry MK, Dostor BP (1993) Relationship between sequence conservation and three-dimensional structure in a large family of esterases, lipases and related proteins. *Protein Science* 2, 366-382
- Diczfalusy MA, Bjorkkem I, Einarsson C, Hillebrant CG, Alexson SE (2001) Characterization of enzymes involved in formation of ethyl esters of long-chain fatty acids. *J Lipid Res* 42, 1025-1032
- Dolinsky VW, Sipione S, Lehner R, Vance DE (2001) The cloning and expression of murine triacylglycerol hydrolase cDNA and the structure of the corresponding gene. *Biochim Biophys Acta* 1532, 162-172
- Donoghue PCJ, Benton MJ (2007) Rocks and clocks: calibrating the tree of life using fossils and molecules. *Trends Genet* 22, 424– 630
- Ecroyd H, Belghazi M, Dacheux J-L, Miyazaki M, Yamashita T, Gatti JL (2006) An epididymal form of cauxin, a carboxylesterase-like enzyme, is present and active in mammalian male reproductive fluids. *Biol Reprod* 74, 439-447.
- Ellingham P, Seedorf U, Assmann G (1998) Cloning and sequencing of a novel murine liver carboxylesterase cDNA. *Biochim. Biophys. Acta* 1397, 175-179
- Fleming C.D, Bencharit S, Edwards CC, Hyatt JL, Tsurkan L, Bai F, Fraga C, Morton CL, Howard-Williams EL, Potter PM, Redinbo MR (2005) Structural insights into drug processing by human carboxylesterase 1: tamoxifen, Mevastatin, and inhibition by Benzil. *J Mol Biol* 352, 165-177
- Fukami T, Nakajima M, Maruichi T, Takahashi S, Takamiya M, Aoki Y, McLeod HL, Yokoi T (2008) Structure and characterization of human carboxylesterase 1A1, 1A2 and 1A3 genes. *Pharm Genomics* 18, 911-920
- Furihata T, Hosokawa M, Nakata F, Satoh T, Chiba K (2003) Purification, molecular cloning, and functional expression of inducible liver acylcarnitine hydrolase in C57BL/6 mouse, belonging to the carboxylesterase multigene family. *Arch Biochem Biophys* 416, 101-109

- Genetta T.L, D'Eustachio P, Kadner SS, Finlay TH (1988) cDNA cloning of esterase 1, the major esterase activity in mouse plasma. *Biochem Biophys Res Commun* 151, 1364-1370
- Ghosh S (2000) Cholesteryl ester hydrolase in human monocyte/macrophage: cloning, sequencing and expression of full-length cDNA. *Physiol Genomics* 2, 1-8
- Ghosh S, Mallonee DH, Grogan W.M (1995) Molecular cloning and expression of rat hepatic neutral cholesteryl ester hydrolase. *Biochim Biophys Acta* 1259, 305-312
- Gibbs RA, Weinstock GM, Metzker ML, Muzny DM, Sodergren EJ, Scherer S, Scott G, Steffen D, Worley KC. (2004) Genome sequence of the Brown Norway rat yields insights into mammalian evolution. *Nature* 428:493-521
- Gilham D, Alam, M, Gao W, Vance DE, Lehner R (2005) Triacylglycerol hydrolase is localized to the endoplasmic reticulum by an unusual retrieval sequence where it participates in VLDL assembly without utilizing VLDL lipids as substrates. *Mol Biol Cell* 16, 984-996
- Hemmert AC, Otto TC, Wierdl M, Edwards CC, Fleming CD, MacDonald M, Cashman JR, Potter PM, Cerasoli DM, Redinbo MR (2010) Human carboxylesterase 1 stereoselectively binds the nerve agent cyclosarin and spontaneously hydrolyzes the nerve agent sarin. *Mol Pharmacol* 77, 508-516
- Holmes RS, Cox LA, VandeBerg JL (2008a) Mammalian carboxylesterase 5: comparative biochemistry and genomics. *Comp Biochem Physiol Part D* 3, 195-204
- Holmes RS, Chan J, Cox LA, Murphy WJ, VandeBerg JL (2008b) Opossum carboxylesterases: sequences, phylogeny and evidence for CES duplication events predating the marsupial-eutherian common ancestor. *BMC Evol Biol* 8, 54
- Holmes RS, VandeBerg JL, Cox LA (2009a) A new class of mammalian carboxylesterase *CES6*. *Comp Biochem Physiol Part D* 4, 209-217
- Holmes RS, Glenn JP, VandeBerg JL, Cox LA (2009b) Baboon carboxylesterases 1 and 2: sequences, structures and phylogenetic relationships with human and other primate carboxylesterases. *J Med Primatol* 38, 27-38
- Holmes RS, Cox LA, VandeBerg JL (2009c) Horse carboxylesterases: evidence for six CES1 and four families of CES genes on chromosome 3. *Comp Biochem Physiol* 4: 54-65
- Holmes RS, Cox LA, VandeBerg JL (2010) Mammalian carboxylesterase 3: comparative genomics and proteomics. *Genetica* DOI 10.1007/s10709-101-9438-z (in press)
- Hosokawa M, Furihata T, Yaginuma Y, Yamamoto N, Kayano N, Fujii A, Nagahara Y, Satoh T, Chiba K (2007) Genomic structure and transcriptional regulation of the rat, mouse and human carboxylesterase genes. *Drug Metab Revs* 39, 1-15
- Hosokawa M (2008) Structure and catalytic properties of carboxylesterase isozymes involved in metabolic activation of prodrugs. *Molecules* 13, 412-431

- Hosokawa M, Furihata T, Yaginuma Y, Yamamoto N, Watanabe N, Tsukada E, Ohhata Y, Kobayashi K, Satoh T, Chiba K (2008) Structure organization and characterization of the regulatory element of the human carboxylesterase (CES1A1 and CES1A2) genes. *Drug Metab Pharmacogenet* 23, 73-84
- Humerickhouse R, Lohrbach K, Li L, Bosron WF, Dolan ME (2000) Characterization of CPT-11 hydrolysis by human liver carboxylesterase isoforms h-CE1 and hCE-2. *Cancer Res* 60, 1189-1192
- Imai T (2006) Human carboxylesterase isozymes: catalytic properties and rational drug design. *Drug Metab Pharmacogen* 21, 173-185
- Imai T, Yoshigae Y, Hosokawa M, Chiba K, Otagiri M (2003) Evidence for the involvement of a pulmonary first-pass effect via carboxylesterase in the disposition of a propanolol ester derivative after intravenous administration. *J Pharmacol Exp Therapeut* 307, 1234-1242
- Ko KW, Erickson B, Lehner R (2009) Es-x/Ces1 prevents triacylglycerol accumulation in McArdle-RH7777 hepatocytes. *Biochim Biophys Acta* 1791, 1133-1143
- Krishnasamy R, Teng AL, Dhand R, Schultz RM, Gross NJ (1998) Molecular cloning, characterization and differential expression pattern of mouse lung surfactant convertase. *Am J Physiol Lung Mol Cell Biol* 275, L969-L975
- Kroetz DL, McBride OW, Gonzalez FJ (1993) Glycosylation-dependent activity of Baculovirus-expressed human liver carboxylesterases: cDNA cloning and characterization of two highly similar enzyme forms. *Biochem* 32, 11606-11617
- Langmann T, Becker A, Aslanidis C, Notka F, Ulrich H, Schwer H, Schmitz G (1997) Structural organization and characterization of the promoter region of a human carboxylesterase gene. *Biochim Biophys Acta* 1350, 65-74
- Lehner R, Vance DE (1999) Cloning and expression of a cDNA encoding a hepatic microsomal lipase that mobilizes stored triacylglycerol. *Biochem J* 343, 1-10.
- Leinweber FJ (1987) Possible physiological roles of carboxyl ester hydrolases. *Drug Metab Revs* 18, 379-439.
- Linke T, Dawson H, Harrison EH (2005) Isolation and characterization of a microsomal retinyl ester hydrolase. *J Biol Chem* 280, 23287-23294
- Lockridge O, Adkins S, La Due BN (1987) Location of disulfide bonds within the sequence of human serum cholinesterase. *J Biol Chem* 262, 12945-12952
- Marsh S, Xiao M, Yu J, Ahluwalia R, Minton M, Freimuth RR, Kwok P-Y, McLeod HL (2004) Pharmacogenomic assessment of carboxylesterases 1 and 2. *Genomics* 84, 661-668
- Masaki K, Hashimoto M, Imai (2007) Intestinal first-pass metabolism via carboxylesterase in rat jejunum and intestine. *Drug Metab Dispos* 35, 1089-1095



Miyazaki M, Yamashita T, Suzuki Y, Saito Y, Soeta S, Taira H, Suzuki A (2006) A major urinary protein of the domestic cat regulates the production of felinine, a putative pheromone precursor. *Chem Biol* 13, 1070-1079

Morton CL, Iacono L, Hyatt JL, Taylor KR, Cheshire PJ, Houghton PJ, Danks MK, Stewart CF, Potter PM (2005) Activation and antitumor activity of CPT-11 in plasma esterase-deficient mice. *Cancer Chemother Pharmacol.* 56, 629-36

Miyazaki M, Kamiie K, Soeta S, Taira H, Yamashita T (2003) Molecular cloning and characterization of a novel carboxylesterase-like protein that is physiologically present at high concentrations in the urine of domestic cats (*Felis Catus*). *Biochem J* 370, 101-110

Miyazaki K, Yamashita T, Suzuki Y, Saito Y, Soeta S, Taira H, Suzuki A (2006) A major urinary protein of the domestic cat regulates the production of felinine, a putative pheromone precursor. *Chem Biol* 13, 10171-10179

Munger JS, Shi GP, Mark EA, Chin DT, Gerard C, Chapman HA (1991) A serine esterase released by human alveolar macrophages is closely related to liver microsomal carboxylesterases. *J Biol Chem* 266, 18832-18838

Mutch E, Nave R, McCracken N, Zech K, Williams FM (2007) The role of esterases in the metabolism of ciclesinide to deisobutyl-ciclesonide in human tissue. *Biochem Pharmacol* 73, 1657-1664

Ohtsuka H, Inoue S, Kameyama M (2003) Intracellular conversion of irinotecan to its active form, SN-38, by native carboxylesterase in human non-small cell lung cancer. *Lung Cancer* 41, 87-198

Okazaki H, Igarashi M, Nishi M, Tajima M, Sekiya M, Okazaki S, Yahagi N, Ohashi K, Tsukamoto K, Michiyo A-K, Yamada N, Aoki J, Moriwawa R, Takanezawa Y, Arai H, Nagai R, Kadowaki T, Osuga J, Ishibashi S (2006) Identification of a Novel Member of the Carboxylesterase Family That Hydrolyzes Triacylglycerol. A Potential Role in Adipocyte Lipolysis *Diabetes* 55, 2091-2097

Okazaki H, Igarashi M, Nishi M, Sekiya M, Tajima M, Takase S, Takanashi M, Ohta K, Tamura Y, Okazaki S, Yahagi N, Ohashi K (2008) Identification of neutral cholesterol hydrolase, a key enzyme removing cholesterol from macrophages. *J Biol Chem* 283, 33357-33364

Ovnic M., Swank R.T., Fletcher C., Zhen L., Novak E.K., Baumann H., Heintz N., Ganschow R.E. (1991) Characterization and functional expression of a cDNA encoding egasyn (esterase-22): the endoplasmic reticulum-targeting protein of beta-glucuronidase. *Genomics* 11, 956-967

Pindel EV, Kedishvili NY, Abraham TL, Brezinski MR, Zhang A, Dean RA, Bosron WF (1997) Purification and cloning of a broad substrate specificity human liver carboxylesterase that catalyzes the hydrolysis of cocaine and heroin. *J Biol Chem* 272, 14769-14775

Potter P.M, Wolverton JS, Morton CL, Wierdl M, Danks MK (1998) Cellular localization domains of a rabbit and human carboxylesterase: influence on irinotecan (CPT-11) metabolism by the rabbit enzyme. *Cancer Res* 58, 3627-32

- Redinbo MR, Potter PN (2005) Mammalian carboxylesterases: from drug targets to protein therapeutics. *Drug Discovery Today* 10, 313-20
- Rhead B, Karolchik D, Kuhn RM, Hinrichs AS, Zweig AS, Fujita P, Diekhans M, Smith KE, Rosenbloom KR, Raney BJ, Pohl A, Pheasant M, Meyer L, Hsu F, Hillman-Jackson J, Harte RA, Giardine B, Dreszer T, Clawson H, Barber GP, Haussler D, Kent WJ (2010) The UCSC Genome Browser database: update 2010. *Nucleic Acids Res* 38: D613-9
- Robbi M., Beaufay H. (1994) Cloning and sequencing of rat liver carboxylesterase ES-3 (egasyn). *Biochem Biophys Res Commun* 203, 1404-1411
- Robbi M, Beaufay H, Octave J-N (1990) Nucleotide sequence of cDNA coding for rat liver pI 6.1 esterase (ES-10), a carboxylesterase located in the lumen of the endoplasmic reticulum. *Biochem J* 269, 451-458
- Ruppert C, Bagheri A, Markart P, Schmidt R, Seegar W, Gunther A (2006) Liver carboxylesterase cleaves surfactant protein (SP-B) and promotes surfactant subtype conversion. *Biochem Biophys Res Coms* 348, 1449-1454
- Sanghani SP, Davis WI, Dumaual NG, Mahrenholz A, Bosron WF (2002) Identification of microsomal rat liver carboxylesterases and their activity with retinyl palmitate (2002) *Eur J Biochem* 269, 4387-4398
- Sanghani SP, Quinney SK, Fredenberg TB, Davis WI, Murray DJ, Bosron WF (2004) Hydrolysis of irinotecan and its oxidative metabolites, 7-ethyl-10-[4-N(5-aminopentanoic acid)-1-piperidino] carbonyloxycamptothecin and 7-ethyl-10-[4-(1-piperidino)-1 amino]-carbonyloxycamptothecin, by human carboxylesterases CES1A1, CES2, and a newly expressed carboxylesterase isoenzyme, CES3. *Drug Metab Dispos* 32, 505-511
- Satoh T, Hosokawa M (1998) The mammalian carboxylesterases: from molecules to functions. *Ann Revs Pharmacol Toxicol* 38, 257-288
- Satoh T, Hosokawa M (2006) Structure, function and regulation of carboxylesterases. *Chem-Biol Interactions* 162, 195-211
- Satoh T, Taylor P, Bosron WF, Sanghani P, Hosokawa M, Du PB (2002) Current progress on esterases: from molecular structure to function. *Drug Metab Dispos* 30, 488-493
- Schewer H, Langmann T, Daig R, Becker A, Aslandis C, Schmidt G (1997) Molecular cloning and characterization of a novel putative carboxylesterase, present in human intestine and liver. *Biochem Biophys Res Commun* 233, 117-120
- Schreiber R, Taschler U, Wolinski H, Seper A, Tamegger SN, Graf M, Kohlwein SD, Haemmerle G, Zimmermann R, Zechner R, Lass A (2009) Esterase 22 and beta-glucuronidase hydrolyze retinoids in mouse liver. *J Lipid Res* 50, 2514-2523
- Shibita F, Takagi Y, Kitajima M, Kuroda T, Omura T (1993) Molecular cloning and characterization of a human carboxylesterase gene. *Genomics* 17, 76-82

Sun Z., Murry DJ, Sanghani SP, Davis WI, Kedishvilli NY, Zou Q, Hurley TD, Bosron WF (2004) Methylphenadate is stereoselectively hydrolyzed by human carboxylesterase CES1A1. *J Pharmacol Exp Ther* 310, 469-476

Takai S, Matsuda A, Usami Y, Adachi T, Sugiyama T, Katagiri Y, Tatematsu M, Hirano (1997) Hydrolytic profile for ester- or amide-linkage by carboxylesterases pI 5.3 and 4.5 from human liver. *Biol Pharm Bull* 20, 869-873

Taketani M, Shii M, Ohura K, Ninomiya S, Imai T (2007) Carboxylesterase in the liver and small intestine of experimental animals and human. *Life Sci* 81: 924-932

The MGC Project Team. (2004) The status, quality, and expansion of the NIH full-length cDNA project: the Mammalian Gene Collection (MGC). *Genome Res* 14, 2121-2127

Thierry-Mieg D, Thierry-Mieg J (2006) AceView: A comprehensive cDNA-supported gene and transcripts annotation. *Genome Biology* 7, S12

<http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/index.html?human>

Tanimoto K, Kaneyasu M, Shimokuni T, Hiyama K, Nishiyama M. (2007) Human carboxylesterase 1A2 expressed from carboxylesterase 1A1 and 1A2 genes is a potent predictor of CPT-11 cytotoxicity *in vitro*. *Pharm Genomics* 17, 1-10

Tsujita T, Okuda H (1993) Palmitoyl-coenzyme A hydrolyzing activity in rat kidney and its relationship with carboxylesterase. *J Lipid Res* 34, 1773-1781

Vanlith H A, Haller M, Vanhoof IJM, Vanderwouw MJA, Vanzutphen BFM, Beynen AC (1993) Characterization of rat plasma esterase ES-1A concerning its molecular and catalytic properties. *Arch Biochem Biophys* 301, 265-274

Vistoli G, Padretti A, Mazzolari A, Testa B (2010) Homology modelling and metabolism prediction of human carboxylesterase-2 using docking analyses by GriDock: a parallelized tool based on AutoDock 4.0. *J Comput Aided Mol Des* PMID: 20623318

von Heijne G (1983) Patterns of amino acids near signal-sequence cleavage sites. *Eur J Biochem* 133, 17-21

Wang H, Gilham D, Lehner R (2007) Proteomic and lipid characterization of apo-lipoprotein B-free luminal lipid droplets from mouse liver microsomes: implications for very low density lipoprotein assembly. *J Biol Chem* 282, 33218-33226

Williams ET, Wang H, Wrighton SA, Qian Y-W, Perkins EJ (2010) Genomic analysis of the carboxylesterases: identification and classification of novel forms. *Mol Phylogenet Evol* [doi:10.1016/j.ympev.2010.05.018](https://doi.org/10.1016/j.ympev.2010.05.018)

Woodburne MO, Rich TH, Springer MS (2003) The evolution of tribospheny and the antiquity of mammalian clades. *Mol Phylogenet Evol* 28,360–385

Xu G, Zhang W, Ma, MK, MacLeod HL (2002) Human carboxylesterase 2 is commonly expressed in tumor tissue and is correlated with the activation of irinotecan. Clin Cancer Res 8, 2605-2611

Yan B, Matoney L, Yang D (1999) Human carboxylesterases in term placenta: enzymatic characterization, molecular cloning and evidence for the existence of multiple forms. Placenta 20, 517-525

Yoshimura M, Kimura T, Ishii M, Ishii K, Matsuura T, Geshi E, Hosokawa M, Muramatsu M (2008) Functional polymorphisms in carboxylesterase1A2 (CES1A2) gene involves specific protein 1 (Sp1) binding sites. Biochem Biophys Res Commun 369, 939-942

Zhang L, Hu Z, ZHU C, Liu Q, Zhou Y, Zhang Y (2009) Identification and characterization of an epididymis-specific gene, *Ces7*. Acta Biochim Biophys Sin 41:809-815

Zhen L, Rusiniak ME, Swank RT (1995) The beta-glucuronidase propeptide contains a serpin-related octamer necessary for complex formation with egasyn esterase and for retention within the endoplasmic reticulum. J Biol Chem 270, 11912-11920

## Figure Legends:

### Figure 1: Amino Acid Sequence Alignments for Human CES1, CES2, CES3, CES4A and CES5A Subunits

See Table 1 for CES isoform sequences aligned; \* shows identical residues for CES subunits; : similar alternate residues; . dissimilar alternate residues; signal peptide sequences for CES1 (1-17), CES2 (1-25), CES3 (1-27), CES4A (1-19) and CES5A (1-24) and C-termini (MTS) microsomal targeting sequences for CES1 (564-567), CES2 (556-569) and CES3 (568-571) are shown in **red**; active site (AS) triad residues (human CES1) Ser221, Glu354 and His468 are highlighted in **green**; 'Side door' (Val424-Met425-Phe426), 'Gate' (Phe550) and cholesterol binding residue (Z site) (Gly356) for human CES1 (Fleming et al, 2005) are highlighted in **khaki**; disulfide bond Cys residues for human CES1 (\*) are shown in **blue**; charge clamp residues identified for human CES1 (Glu72...Arg186; Lys78...Glu183) (Fleming et al, 2005) are highlighted in **purple**; confirmed (CES1) (Asn79-Ala80-Thr81) [site 1] or predicted N-glycosylation sites for human CES2 (Asn111-Met112-Thr113) [site 3], CES3 (Asn105-Ser106-Ser107) [site 2], CES4A (Asn213-Val214-Thr215 [site 4]; Asn276-Ser-277-Thr278) [site 5]; and Asn388-Ile389-Thr390) [site 7] and CES5A (Asn363-Lys364-Ser365 [site 6]; Asn513-Leu514-Thr515 [site 8]; and Asn524-Met525-Ser526 [site 9]) are highlighted in **blue**;  $\alpha$ -helix (human CES1 or predicted) and  $\beta$ -sheet (human CES1 or predicted) regions were highlighted in **yellow** and **grey**, respectively;  $\alpha$ -helices and  $\beta$ -sheets are numbered according to the reported human CES1 3D structure (Fleming et al, 2005); **bold underlined** font shows known or predicted exon start sites; exon numbers refer to the human *CES1* gene (see Langmann et al., 1997).

### Figure 2: Gene Structures and Major Isoforms for Human *CES1*, *CES2*, *CES3*, *CES4A* and *CES5A* Genes.

Derived from AceView website <http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/> (Thierry-Mieg and Thierry-Mieg, 2006) Mature isoform variants (a) are shown with capped 5'- and 3'- ends for the predicted mRNA sequences; exons are in solid color; 5'- and 3'- untranslated regions of the genes are shown as open boxes; introns are shown as a line; the 5'  $\rightarrow$  3' transcription directions are shown; **a** refers to the major transcript isoform for each human *CES* gene. Note that each *CES* gene structure is drawn to a different scale and that the respective gene sizes are shown: *CES1* [34.8kb]; *CES2* [10.9kb]; *CES3* [13.9kb]; *CES4A* [22.3kb]; and *CES5A* [79.3kb].

## Table Legends

### Table 1: Human *CES1*, *CES1P1*, *CES2*, *CES3*, *CES4A* and *CES5A* Genes and Subunits.

RefSeq, GenBank and UNIPROT IDs provide the sources for the gene and protein sequences; the relative gene expression level for human CES genes in comparison with the expression of an average human gene is given in [brackets] <http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/>; <sup>2</sup>refers to GenBank ID number; <sup>3</sup>no current AceView isoform name available; ps refers to a pseudogene (*CES1P1*); gene sizes are given as base pairs of nucleotides; +ve and -ve refer to the transcription strand direction; CES isoform sequences aligned in Figure 1 are shown in **bold**; <sup>4</sup>the human CES2\_BC032095 isoform transcript contains multiple transcription start sites with the shorter CES2 sequence (559 residues) previously reported (see Schwer et al., 1997; Pindel et al., 1997).

### Table 2: Mouse *Ces* Genes and Subunits.

RefSeq, GenBank, UNIPROT, MGI, Vega and Ensembl IDs provide the sources for the gene and protein sequences; ps refers to a pseudogene (*Ces2d-ps*); <sup>^</sup>the relative gene expression level for mouse *Ces* genes in comparison with the expression of an average mouse gene is given in [brackets] <http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/>; gene sizes are given as base pairs of nucleotides; +ve and -ve refer to the transcription strand.

### Table 3: Rat *Ces* Genes and Subunits.

RefSeq, GenBank, UNIPROT, RGD, Vega and Ensembl IDs provide the sources for the gene and protein sequences; gene sizes are given as base pairs of nucleotides; the relative gene expression level for rat *Ces* genes in comparison with the expression of an average rat gene is given in [brackets] <http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/>; +ve and -ve refer to the transcription strand direction.

### Table 4: Functions and Substrates for Human *CES*, and Mouse and Rat *Ces* Genes and Enzymes

<sup>1</sup>Pindel et al., 1997; <sup>2</sup>Bencharit et al., 2003; <sup>3</sup>Satoh & Hosokawa, 2006; <sup>4</sup>Sun et al., 2004; <sup>5</sup>Takai et al., 1997; <sup>6</sup>Humerickhouse et al., 2000; Xu et al., 2002; Ohtsuka et al., 2003; Morton et al., 2005; <sup>7</sup>flurbiprofen derivatives serve as substrates; Imai, 2006; Taketani et al., 2007; Hosokawa, 2008; <sup>8</sup>Diczfalusy et al., 2001; <sup>9</sup>Hemmert et al., 2010; <sup>10</sup>Mutch et al., 2007; <sup>11</sup>Becker et al., 1994; <sup>12</sup>Barthel et al., 2008; <sup>13</sup>Krishnasamy et al., 1998; Ruppert et al., 2006; <sup>14</sup>Morton et al., 2005; <sup>15</sup>Dolinsky et al., 2005; <sup>16</sup>Ovnic et al., 1991; <sup>17</sup>Ellingham et al., 1998; Koh et al., 2009; <sup>18</sup>Furihata et al., 2003; <sup>19</sup>Sanghani et al., 2002; <sup>20</sup>Ghosh et al., 1995; Okazaki et al., 2008; <sup>21</sup>Robbi & Beaufay, 1994; <sup>22</sup>Masaki et al., 2007; <sup>23</sup>Miyazaki et al., 2006; <sup>24</sup>Ecroyd et al., 2006; Zhang et al., 2009 <sup>25</sup>Gilham et al., 2005; <sup>26</sup>Schreiber et al., 2009; <sup>27</sup>Lehner & Vance, 1999; <sup>28</sup>Okazaki et al., 2006; <sup>29</sup>Linke et al., 2005.



Figure 2

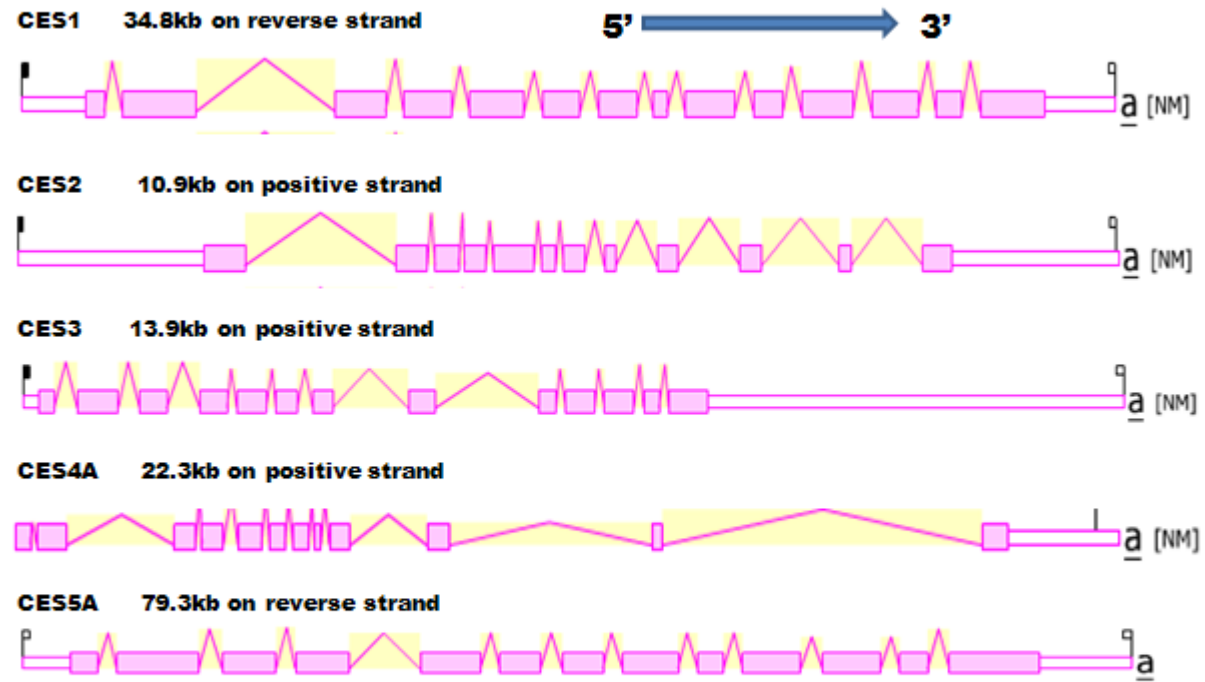




Table 1



Table 2

Mouse CES	Chromosome 8	Gene	Exons	Subunit	Amino	GenBank ID	MGI ID_YZ	Current MGI	Current	NCBI	Vega ID	Ensembl ID
Gene	Coordinates	Size	strand	MW	Acids			Symbol_YZ	Gene	Transcript		
(Proposed)		bps							Symbols			
Ces1a	95,544,116-95,572,091	27,979	14 -ve	61,744	563	BC089371	MGI:3648919	Gm4976	<i>EG244595</i>	NM_001013764	none	ENSMUSG00000071047
Ces1b	95,580,789-95,603,815	23,027	13 -ve	62,197	567	*NM_001081372	MGI:3779470	Gm5158	<i>CesN</i>	NM_001081372	none	ENSMUSG00000078964
Ces1c	95,622,914-95,655,182	32,268	13 -ve	61,172	554	BC028907	MGI:95420	Es1	<i>Es1, Ces-N</i>	NM_007954	ENSMUSG00000024453	ENSMUSG00000057400
Ces1d	95,690,157-95,721,618	31,462	14 -ve	61,788	565	BC019198	MGI:2148202	Ces3	<i>Ces3</i>	NM_053200	ENSMUSG00000024539	ENSMUSG00000056973
Ces1e	95,725,306-95,753,320	28,015	14 -ve	61,582	562	BC019208	MGI:95432	Es22	<i>Es22</i>	NM_133660	ENSMUSG00000024532	ENSMUSG00000061959
Ces1f	95,780,331-95,803,599	23,269	14 -ve	61,698	561	BC013479	MGI:234564	AU018778	<i>CesML1, TGH-2</i>	NM_144930	ENSMUSG00000024519	ENSMUSG00000031725
Ces1g	95,826,807-95,861,053	34,247	14 -ve	62,680	565	BC021150	MGI:88378	Ces1	<i>Ces1</i>	NM_021456	ENSMUSG00000024535	ENSMUSG00000057074
Ces1h	95,875,926-95,903,624	27,699	14 -ve	62,087	562	AK009689	MGI:75704	2310039D24Rik	<i>AK009689</i>	XM_134476	ENSMUSG00000033579	ENSMUSG00000074156
Ces2a	107,257,972-107,265,313	7,342	12 +ve	61,940	558	BC024491	MGI:2142491	Ces6	<i>Ces6</i>	NM_133960	OTTMUSG00000027410	ENSMUSG00000055730
Ces2b	107,355,572-107,362,353	6,782	12 +ve	61,927	556	BC015286	MGI:2448547	BC015286	<i>BC015286</i>	NM_198172	OTTMUSG00000027467	ENSMUSG00000050097
Ces2c	107,371,033-107,378,161	7,129	12 +ve	62,470	561	BC031170	MGI:2389505	Ces2	<i>Ces2</i>	NM_145603	OTTMUSG00000027466	ENSMUSG00000061825
Ces2d-ps	107,391,388-107,397,764	3,762	6 +ve			BC034182	MGI:3704319	Gm9756		XR_002069	none	ENSMUSG00000031884
Ces2e	107,450,221-107,457,611	7,391	12 +ve	62,735	560	BC055062	MGI:2443170	Ces5	<i>Ces5</i>	NM_172759	none	ENSMUSG00000031886
Ces2f	107,471,256-107,479,862	7,335	12 +ve	62,707	561	BC117742	MGI:1919153	2310038E17Rik		NM_001079865	none	ENSMUSG00000062826
Ces2g	107,485,688-107,492,328	6,771	10 +ve	52,731	478	BC027185	MGI:1919611	2210023G06Rik		NM_197999	none	ENSMUSG00000031877
Ces2h	107,524,753-107,544,307	19,554					MGI:3648740	Gm5744		XM_488149	none	none
Ces3a	107,572,572-107,582,000	21,512	13 +ve	61,510	554	AK138932	MGI:102773	Es31	<i>Es31</i>	NM_198672	none	ENSMUSG00000069922
Ces3b	107,607,670-107,617,468	9,799	14 +ve	63,007	568	BC019047	Gm4738	Es31L	<i>Es31L</i>	NM_144511	none	ENSMUSG00000062181
Ces4a	107,655,852-107,673,417	17,566	14 +ve	62,123	563	BC026374	BC026374	Ces8	<i>Ces8</i>	NM_146213	OTTMUSG00000027469	ENSMUSG00000060560
Ces5a	96,038,095-96,059,607	21,512	13 +ve	64,167	575	AB186393	MGI:1915185	Ces7	<i>Ces7</i>	NM_001003951	none	ENSMUSG00000058019

Table 3



Table 4

Mammal	<i>CES</i> ( <i>Ces</i> ) Gene	Current Gene Symbol(s)	Substrates and Function (Hydrolysis or detoxification)
Human	<i>CES1</i>	<i>CES1, hCE-1, CES1A1, HU1</i>	heroin, cocaine <sup>1-3</sup> , methyl phenidate <sup>4</sup> , temocapril <sup>5</sup> , CPT-11 <sup>6</sup> , flurbiprofen <sup>7</sup>
		<i>CES1</i>	fatty acid ethyl ester synthase <sup>8</sup> , sarin <sup>9</sup> , ciclesonide <sup>10</sup> , cholesteryl ester hydrolase <sup>11</sup> , triacylglycerol hydrolase <sup>11</sup>
	<i>CES2</i>	<i>CES2, hCE-2, HU2</i>	procaine <sup>5</sup> , heroin, cocaine <sup>1-3</sup> , temacapril <sup>5</sup> , CPT-11 <sup>6</sup> , flurbiprofen <sup>7</sup> , doxazolidine <sup>12</sup>
	<i>CES3</i>	<i>CES3</i>	CPT-11 <sup>6</sup>
Mouse	<i>Ces1c</i>	<i>Es1, Ces-N</i>	lung surfactant convertase <sup>13</sup> , CPT-11 <sup>14</sup>
	<i>Ces1d</i>	<i>Ces3</i>	triacylglycerol hydrolase <sup>15</sup>
	<i>Ces1e</i>	<i>Es22, egasyn</i>	$\beta$ -glucuronidase binding in the liver endoplasmic reticulum <sup>16</sup> , retinyl ester hydrolase <sup>26</sup>
	<i>Ces1f</i>	<i>CesML1, TGH-2</i>	triacylglycerol hydrolase <sup>27</sup> , monoacylglycerol hydrolase <sup>27</sup> , cholesteryl ester hydrolase <sup>27</sup> , phospholipase <sup>27</sup>
	<i>Ces1g</i>	<i>Ces1</i>	lipid metabolism <sup>17</sup>
	<i>Ces2c</i>	<i>Ces2</i>	inducible liver acylcarnitine hydrolase <sup>18</sup>
Rat	<i>Ces1c</i>	<i>Es1</i>	retinyl palmitate <sup>19</sup>
	<i>Ces1d</i>	<i>Ces3</i>	cholesterol ester hydrolase <sup>20</sup> , triacylglycerol hydrolase <sup>27</sup> , retinyl ester hydrolase <sup>28</sup>
	<i>Ces1e</i>	<i>ES-3</i>	$\beta$ -glucuronidase binding in the liver endoplasmic reticulum <sup>21</sup>
	<i>Ces2a</i>	<i>Ces6</i>	intestinal first pass metabolism <sup>22</sup>
	<i>Ces2c</i>	<i>Ces2</i>	inducible liver acylcarnitine hydrolase <sup>18</sup> , intestinal first pass metabolism <sup>22</sup>
	<i>Ces2e</i>	<i>Ces5</i>	intestinal first pass metabolism <sup>22</sup>
Cat	<i>CES5A</i>	<i>CES7, cauxin</i>	3-methylbutanol-cysteinylglycine hydrolysis in urine releasing pheromone <sup>23</sup>
Rat, sheep	<i>CES5A</i>	<i>CES7, cauxin</i>	lipid transfer reactions in epididymis <sup>24</sup>