

## Refined solution structure and backbone dynamics of HIV-1 Nef

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### Abstract

The tendency of HIV-1 Nef to form aggregates in solution, particularly at pH values below 8, together with its large fraction of highly mobile residues seriously complicated determination of its three-dimensional structure, both for heteronuclear solution NMR (Grzesiek et al., 1996a, *Nat Struct Biol* 3:340–345) and for X-ray crystallography (Lee et al., 1996, *Cell* 85:931–942). Methods used to determine the Nef structure by NMR at pH 8 and 0.6 mM concentration are presented, together with a detailed description of Nef's secondary and tertiary structure. The described techniques have general applicability for the NMR structure determination of proteins that are aggregating and/or have limited stability at low pH values. Extensive chemical shift assignments are reported for backbone and side chain <sup>1</sup>H, <sup>13</sup>C, and <sup>15</sup>N resonances of the HIV-1 Nef deletion mutants NEF<sup>Δ2–39</sup>, NEF<sup>Δ2–39, Δ159–173</sup>, and of NEF<sup>Δ2–39, Δ159–173</sup> in complex with the SH3 domain of the Hck tyrosine protein kinase. Besides a type II polyproline helix, Nef's structure consists of three  $\alpha$ -helices, a <sub>310</sub> helix, and a five-stranded anti-parallel  $\beta$ -sheet. The analysis of <sup>15</sup>N relaxation parameters of the backbone amide sites reveals that all the secondary structure elements are non-mobile on the picosecond to nanosecond and on the millisecond time scale. A large number of slowly exchanging amide protons provides evidence for the stability of the Nef core even on the time scale of hours. Significant internal motions on the ps to ns time scale are detected for residues 60 to 71 and for residues 149 to 180, which form solvent-exposed loops. The residues of the HIV-1 protease cleavage site (W57/L58) do not exhibit large amplitude motions on the sub-nanosecond time scale, and their side chains insert themselves into a hydrophobic crevice formed between the C-terminus of helix 1 and the N-terminus of helix 2. A refined structure has been determined based on additional constraints for side-chain and backbone dihedral angles derived from a large number of three-bond J-couplings and ROE data.

**Keywords:** heteronuclear NMR; HIV-1; multi-dimensional NMR; Nef; protein dynamics; protein structure

HIV-1 Nef is a 206 amino acid, N-terminal myristoylated and membrane-associated protein that is expressed at high levels in the early stages of HIV infection (Cullen, 1994). Although early reports suggested a negative role on viral replication (Luciw et al., 1987; Ahmad & Venkatesan, 1988; Cheng-Mayer et al., 1989; Niederman et al., 1989), the following evidence suggests a positive role of Nef on viral replication and disease pathogenesis: first, macaques infected with the simian immuno-deficiency virus strain SIV<sub>mac239Δnef</sub> bearing a deletion in the Nef gene show only poor

viral replication, do not develop disease, and are protected from infection by pathogenic SIV (Kestler et al., 1991; Daniel et al., 1992). Moreover, in animals infected with an SIV strain containing a premature stop codon (Kestler et al., 1991) or a 12 bp deletion (Whatmore et al., 1995) in the Nef gene, restoration of the intact and complete Nef reading frame is observed and correlates with disease progression. Second, sequencing of the HIV genome of several groups of long-term (>10 years) survivors of HIV infection, with no clinical signs of disease progression and normal cluster determinant 4 (CD4) counts, revealed the presence of discrete and stable deletions in the Nef gene in at least nine individuals (Deacon et al., 1995; Kirchhoff et al., 1995; Mariani et al., 1996; Prekumar et al., 1996). Full-length sequencing, which was carried out for one of these individuals, showed no other abnormalities in the entire HIV genome (Deacon et al., 1995). Third, Nef was shown to increase HIV-1 replication in primary human

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peripheral blood lymphocytes and in certain cell lines in vitro (deRonde et al., 1992; Skowronski et al., 1993; Chowers et al., 1994; Miller et al., 1994; Spina et al., 1994).

The role of Nef at a molecular level remains unclear. Nef down-regulates cell-surface CD4 even in the absence of other viral gene products by post-translational endocytosis (Guy et al., 1987; Garcia & Miller, 1991; Benson et al., 1993; Garcia et al., 1993; Skowronski et al., 1993; Foster et al., 1994). However, this function seems to be dissociated from the viral infectivity enhancement (Goldsmith et al., 1995; Chowers et al., 1995; Saksela et al., 1995). This latter activity of Nef is linked to a conserved polyproline repeat from residues 69–78 (Goldsmith et al., 1995; Saksela et al., 1995). The polyproline motif is reminiscent of an SH3 target site (Shugars et al., 1993; Saksela et al., 1995), and Nef has been shown to bind specifically and with sub-micromolar affinity to the SH3 domains of a subset of Src family kinases (Lee et al., 1995, 1996; Saksela et al., 1995).

Recently, we have determined the Nef (NEF $\Delta^{2-39, \Delta 159-173}$ ) structure by heteronuclear NMR (Grzesiek et al., 1996a). In this construct, residues 2–39 of the unstructured N-terminus (Freund et al., 1994a) and residues 159–173 of a long solvent exposed loop had been removed without altering the structure of the remaining Nef core domain. Chemical shift changes induced by the binding of SH3 Hck to Nef allowed us to model the Nef–SH3 complex (Grzesiek et al., 1996a). Independently, the structure of Nef (residues 54–205) in complex with a mutated SH3 domain of Fyn tyrosine kinase has been solved by X-ray crystallography (Lee et al., 1996). The two structures of the Nef core domain are identical within the experimental resolution of the two studies, and the position of the SH3 domain relative to Nef in the X-ray structure agrees with our model of the complex. In addition to the Nef–SH3 interaction, chemical shift changes induced by binding to a peptide from the C-terminal tail of CD4 provide evidence for a direct Nef–CD4 interaction and delineate a well-defined binding surface for this peptide (Grzesiek et al., 1996b). This binding site is located in the vicinity of the W57/L58 HIV-1 protease cleavage site and is distinct from the SH3 binding site.

Efforts to determine the structure of Nef by either NMR or crystallography have long been hampered by its large fraction of unstructured residues and Nef's tendency to form aggregates at millimolar concentrations, especially at pH values below 8. Because of sensitivity considerations, NMR structures are usually determined at concentrations  $\geq 1$  mM. Furthermore, the fast ex-

change of amide protons with water can be a serious source of line broadening and information loss at basic pH values. Here, we give a detailed description of the methods that were used to determine the structure of Nef by NMR at 0.6 mM concentration and at pH 8. NMR evidence is presented that defines the various structural elements and  $^{15}\text{N}$  relaxation data provide a measure of the degree of local backbone mobility on the pico- to nanosecond time scale. Based on a substantial number of additional distance and dihedral restraints, a refined structure of Nef has been calculated and its coordinates have been deposited in the Brookhaven data base (accession code 2NEF).

## Results and discussion

### Backbone resonance assignments

A summary of the Nef constructs and samples used for this NMR study is given in Table 1. Preliminary NMR measurements on full-length Nef indicated a large unstructured region of  $\sim 40$  residues (Freund et al., 1994a; A. Bax & P. Wingfield, unpubl. obs.). Limited digestion of Nef with trypsin resulted in the removal of many of the unstructured residues in the NMR spectrum. Trypsin cleaves Nef rapidly and completely between residue 39 and 40 (Freund et al., 1994a; A. Bax & P. Wingfield, unpubl. obs.), and a Nef deletion mutant was constructed where residues 2–39 are removed (NEF $\Delta^{2-39}$ ). This mutant expresses as a soluble product at high levels in *E. coli* (Grzesiek et al., 1996a).

The backbone assignment was first carried out on this NEF $\Delta^{2-39}$  construct. For the assignment of the backbone and  $\text{H}^\beta$  and  $\text{C}^\beta$  resonances, data from the HNCO, HNCA, HNCACB, CBCA-(CO)NH, HBHA(CO)NH,  $^{15}\text{N}$ -edited HOHAHA,  $^{15}\text{N}$ -edited NOESY for samples of NEF $\Delta^{2-39}$  dissolved in 95%  $\text{H}_2\text{O}/5\%$   $\text{D}_2\text{O}$  were used according to Table 2. As Nef forms aggregates at concentrations of  $\sim 1$  mM and is predominantly monomeric only at  $\sim 0.1$  mM (see below), for all the experiments in Table 2 a compromise value of 0.6 mM was used. This set of experiments and the associated assignment process are very similar to a previously applied strategy (Anglister et al., 1994). However, special care was taken to minimize the effects of resonance overlap and fast amide hydrogen exchange caused by the high pH value.

In brief, the assignment strategy consists of classifying all the HN, N-frequency pairs according to cross-peaks in the HNCO spectrum, which is the most well resolved and most sensitive of the

**Table 1.** Sample preparations<sup>a</sup>

No.	Nef construct	Labeling	Solvent
1	NEF $\Delta^{2-39}$	U- $^{15}\text{N}$	95% $\text{H}_2\text{O}/5\%$ $\text{D}_2\text{O}$
2	NEF $\Delta^{2-39}$	U- $^{15}\text{N}$ , U- $^{13}\text{C}$	95% $\text{H}_2\text{O}/5\%$ $\text{D}_2\text{O}$
3	NEF $\Delta^{2-39}$	U- $^{15}\text{N}$ , U- $^2\text{H}$	95% $\text{H}_2\text{O}/5\%$ $\text{D}_2\text{O}$
4	NEF $\Delta^{2-39}$	U- $^{15}\text{N}$ , U- $^{13}\text{C}$ , Phe- $^{12}\text{C}$	$\text{D}_2\text{O}^b$
5	NEF $\Delta^{2-39, \Delta 159-173}$	U- $^{15}\text{N}$	95% $\text{H}_2\text{O}/5\%$ $\text{D}_2\text{O}$
6	NEF $\Delta^{2-39, \Delta 159-173}$	U- $^{15}\text{N}$ , U- $^{13}\text{C}$	95% $\text{H}_2\text{O}/5\%$ $\text{D}_2\text{O}$
7	NEF $\Delta^{2-39, \Delta 159-173}$	U- $^{15}\text{N}$ , U- $^{13}\text{C}$	$\text{D}_2\text{O}^b$
8	NEF $\Delta^{2-39, \Delta 159-173}$ in 1:1 complex with Hck SH3	U- $^{15}\text{N}$ , U- $^{13}\text{C}$	95% $\text{H}_2\text{O}/5\%$ $\text{D}_2\text{O}$

<sup>a</sup>All samples were prepared as 0.6 mM solutions in 5 mM Tris-HCl, 5 mM dithiothreitol at pH 8.0 (uncorrected meter reading).

<sup>b</sup>Lyophilized and redissolved in  $\text{D}_2\text{O}$ .

**Table 2a.** Assignment experiments<sup>a</sup>

Experiment	Sample <sup>b</sup>	Time (h)	Acquisition times (ms)			No. of constraints <sup>c</sup>
HOHAHA15N <sup>d</sup>	1	35	15.4 (H)	39.6 (N)	55.3 (HN)	
HOHAHA12C <sup>e</sup>	4	2	50.4 (H1)	145.1 (H2)		
NOESY12C <sup>f</sup>	4	15	51.0 (H1)	141.3 (H2)		
HNCA <sup>g</sup>	2, 8	33	11.0 (CA)	26.8 (N)	55.3 (HN)	
HNCACB <sup>h</sup>	2, 6	83	7.2 (CACB)	26.8 (N)	55.3 (HN)	
HNCO <sup>i</sup>	2	14	49.6 (CO)	26.3 (N)	55.3 (HN)	
HBHACO <sup>j</sup>	2	63	13.0 (HAHB)	19.8 (N)	55.3 (HN)	
CBCACO <sup>k</sup>	2,6,8	62	6.2 (CACB)	19.8 (N)	55.3 (HN)	133
CH3COSY <sup>l</sup>	4	12	12.8 (C1)	25.8 (C-MET)	52.6 (H-MET)	
CH3DIPSI <sup>m</sup>	4	46	6.2 (C)	25.6 (C-MET)	79.8 (H-MET)	

<sup>a</sup>These experiments were used to derive assignments based solely on J-coupling connectivities (with the exception of NOESY12C).

<sup>b</sup>The experiments were carried out on all of the listed samples. Numbers correspond to Table 1.

<sup>c</sup>Number of constraints derived from the experiment and used in structure calculation.

<sup>d</sup><sup>15</sup>N-separated HOHAHA (Marion et al., 1989), mixing time 32 ms.

<sup>e</sup>2D HOHAHA selective for <sup>12</sup>C-attached protons (Bax et al., 1994), mixing time 32 ms.

<sup>f</sup>2D NOESY selective for <sup>12</sup>C-attached protons (Vuister et al., 1994), mixing time 85 ms.

<sup>g</sup>Constant-time HNCA (Grzesiek & Bax, 1992a) with water flip-back modifications (Grzesiek & Bax, 1993b; Kay et al., 1994).

<sup>h</sup>Constant-time HNCACB (Wittekind & Mueller, 1993) with water flip-back modifications (Grzesiek & Bax, 1993b; Kay et al., 1994).

<sup>i</sup>Constant-time HNCO (Grzesiek & Bax, 1992a) with water flip-back modifications (Grzesiek & Bax, 1993b; Kay et al., 1994).

<sup>j</sup>HBHA(CO)NH (Grzesiek & Bax, 1993a) with WATERGATE (Piotto et al., 1992).

<sup>k</sup>CBCA(CO)NH (Grzesiek & Bax, 1992b) with WATERGATE (Piotto et al., 1992).

<sup>l</sup>HCCH-COSY (Bax et al., 1990a) optimized for methyl group detection by using constant-time HSQC-type <sup>13</sup>C evolution (Vuister & Bax, 1992).

<sup>m</sup>HCCH-TOCSY (Bax et al., 1990b) optimized for methyl group detection by using constant-time HSQC-type <sup>13</sup>C evolution.

**Table 2b.** J-coupling experiments<sup>a</sup>

Experiment	Sample	Time (h)	Acquisition times (ms)			No. of constraints
HNHA <sup>b</sup>	5	87	15.4 (H)	38.2 (N)	55.3 (HN)	91
HNCOCO <sup>c</sup>	6	69	26.9 (CO)	26.6 (N)	48.3 (HN)	78
HAHB <sup>d</sup>	7	24	11.0 (HAHB)	24.2 (CA)	54.7 (HA)	37
CH3CO/CN <sup>e</sup>	7	36	25.9 (C-MET)	79.5 (H-MET)		9
HN(CG) <sup>f</sup>	6	18	88.8 (N)	47.1 (HN)		
HN(CO)(CG) <sup>g</sup>	6	19	50.0 (N)	47.4 (HN)		19

<sup>a</sup>Experiments that were used to derive dihedral angle restraints from <sup>3</sup>J-couplings.

<sup>b</sup>Water flip-back HNHA (Kuboniwa et al., 1994).

<sup>c</sup>HNCOCO quantitative <sup>3</sup>J<sub>C'C'</sub> correlation (Hu & Bax, 1996; Grzesiek & Bax, 1997).

<sup>d</sup>HACAHB-COSY (Grzesiek et al., 1995b).

<sup>e</sup>2D constant-time spin-echo difference <sup>13</sup>C-HSQC for measurement of <sup>13</sup>C-<sup>15</sup>N (Vuister et al., 1993) and <sup>13</sup>C-<sup>13</sup>C' (Grzesiek et al., 1993) J-couplings.

<sup>f</sup>2D constant-time spin-echo difference <sup>15</sup>N-HSQC for measurement of aromatic <sup>13</sup>C-<sup>15</sup>N J-couplings (Hu et al., 1997).

<sup>g</sup>2D constant-time spin-echo difference experiment for measurement of aromatic <sup>13</sup>C-<sup>15</sup>C' J-couplings (Hu et al., 1997). Constraints are derived from a combined analysis with the HN(CG) experiment.

above set of experiments. Corresponding  $C_i^\alpha$ ,  $C_{i-1}^\alpha$ ,  $C_i^\beta$ ,  $C_{i-1}^\beta$ ,  $H_i^\alpha$ ,  $H_{i-1}^\alpha$ ,  $H_{i-1}^\beta$ ,  $HN_{i-1}$ ,  $HN_{i+1}$  frequencies are extracted from the HNCA, HNCACB, CBCA(CO)NH, HBHA(CO)NH, <sup>15</sup>N-edited HOHAHA, <sup>15</sup>N-edited NOESY. These frequencies are associated with one HNCO cross-peak, assuming that it represents a unique amide bond within the protein backbone. The spin systems asso-

ciated with each HNCO peak are then matched against all others for possible interresidue correlations to generate sequential stretches of amino acid spin systems. These stretches are, in turn, assigned to locations in the primary sequence of the protein based on the amino acid type probability of the  $C^\alpha$  and  $C^\beta$  shifts (Grzesiek & Bax, 1993a).

**Table 2c.** NOESY/ROESY experiments<sup>a</sup>

Experiment	Sample	Time (h)	Mix (ms)	Acquisition times (ms)				No. of constraints
NOE15N15N <sup>b</sup>	3	108	200	20.8 (N1)	20.0 (HN1)	20.8 (N2)	55.3 (HN2)	
NOE15N <sup>c</sup>	1,5	73	70	16.9 (H)	39.6 (N)	55.3 (HN)		355
ROE15N <sup>d</sup>	5	58	20	15.0 (H)	60.0 (N)	47.1 (HN)		128
NOE13C <sup>e</sup>	7	67	74	27.4 (H)	7.0 (C)	49.1 (HC)		93
NOE13C <sup>f</sup>	7	76	80	12.0 (H)	26.8 (Cmet)	70.3 (Hmet)		73
NOE13C <sup>g</sup>	7	86	74	15.0 (H)	7.1 (Caro)	48.9 (Haro)		128
NOE13C12C <sup>h</sup>	4	75	74	15.2 (HC)	6.3 (C)	70.0 (Haro)		

<sup>a</sup>Experiments that were used to derive distance restraints from the nuclear Overhauser effect.

<sup>b</sup>4D <sup>15</sup>N, <sup>15</sup>N-edited NOESY (Grzesiek et al., 1995a).

<sup>c</sup><sup>15</sup>N-edited 3D NOESY with water-flip back (Grzesiek & Bax, 1993b) and radiation damping during mixing period (Lippens et al., 1995).

<sup>d</sup><sup>15</sup>N-edited 3D ROESY with water-flip back (Grzesiek & Bax, 1993b).

<sup>e</sup><sup>13</sup>C-edited 3D NOESY (Ikura et al., 1990; Zuiderweg et al., 1990) optimized for detection of aliphatic <sup>1</sup>H-<sup>13</sup>C groups.

<sup>f</sup><sup>13</sup>C-edited 3D NOESY optimized for methyl group detection with constant-time HSQC (Santoro & King, 1992; Van de Ven & Philippens, 1992; Vuister & Bax, 1992).

<sup>g</sup><sup>13</sup>C-edited 3D NOESY optimized for detection of aromatic <sup>1</sup>H-<sup>13</sup>C groups.

<sup>h</sup><sup>13</sup>C-edited 3D NOESY (Vuister et al., 1994) with detection of <sup>12</sup>C-attached protons in U-<sup>15</sup>N, U-<sup>13</sup>C, Phe-<sup>12</sup>C sample.

**Table 2d.** <sup>15</sup>N relaxation experiments<sup>a</sup>

Experiment	Sample	Time t (h)	Acquisition times (ms)	
<sup>15</sup> N T <sub>1</sub> <sup>b</sup>	5	22	96.3 (N)	55.1 (HN)
<sup>15</sup> N T <sub>2</sub> <sup>b</sup>	5	18	82.2 (N)	55.1 (HN)
{ <sup>1</sup> H}- <sup>15</sup> N NOE <sup>c</sup>	5	33	82.2 (N)	55.1 (HN)

<sup>a</sup>Experiments for the definition of protein dynamics on the ps- to ns-time scale.

<sup>b</sup>Kay et al., 1992.

<sup>c</sup>Grzesiek and Bax, 1993b.

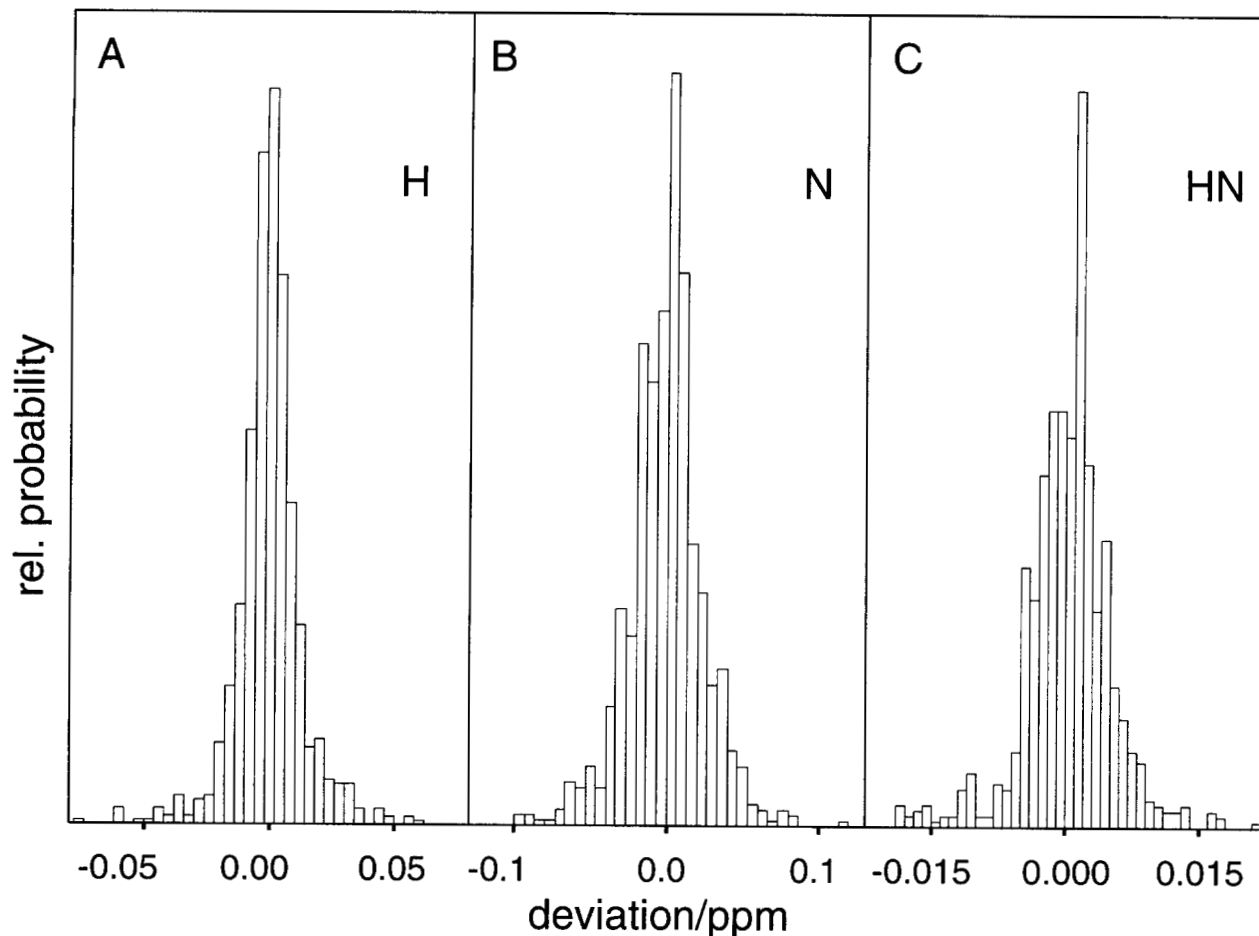
#### Determination of experimental variances of the chemical shifts

A large fraction of the cross-peaks in the 3D spectra listed in Table 2 are well resolved, even in the case of broad linewidths. During the assignment process cross-peaks from different 3D spectra are usually linked together by one or two common frequencies. However, in this one- or two-dimensional frequency sub-space of peak connections, crowding and peak overlap often is a serious problem. To decide whether two frequencies should be considered identical, it is critical to have an accurate estimate of the experimental variance of the peak positions. With frequency identity, the peaks can originate from the same nuclei or from nuclei with degenerate chemical shifts. If two frequencies differ by more than the sum of their experimental uncertainties, the respective cross-peaks very likely originate from different nuclei. If a spectrum contains multiple cross-peaks for a given nucleus, the reproducibility of frequencies derived from the spectrum can be assessed a posteriori once the spectrum has been assigned. In practice, it is not necessary to assign the whole spectrum, but a good estimate can be obtained from a small number of representative cross-peaks. Figure 1 shows a histogram of deviations from the mean frequency of the assigned peaks for the <sup>15</sup>N-edited NOESY spectrum of NEF<sup>Δ2-39,Δ159-173</sup>. The root-mean-square deviations (RMSD)

of the peak positions are 8.4, 1.6, and 3.0 Hz for the indirect proton frequency (H), the nitrogen frequency (N), and the detected amide proton (H<sup>N</sup>) frequency, respectively. For sufficient zero-filling, to a first approximation these frequency variances are inversely proportional to the mean signal duration in the time domain. Besides the transverse relaxation time, T<sub>2</sub>, this mean lifetime of the resonances is determined by the acquisition time and the digital filter function used. For all data sets listed in Table 1, we empirically determined that the variance in the peak positions is ~20–30 times smaller than the inverse of this mean lifetime.

The practical value of a correct estimate of the frequency variations is demonstrated for the amide proton and nitrogen frequency pairs of D123 and R134 in NEF<sup>Δ2-39</sup>. Figure 2 shows orthogonal (vide infra) strip plots for the amide frequencies of D123 for the backbone assignment experiments of Table 2. At first glance, resonances belonging to the two amides might be classified as overlapping. However, the respective chemical shifts as measured from the interresidue H<sup>N</sup>-H<sub>i-1</sub><sup>α</sup> cross-peak in the <sup>15</sup>N-edited NOESY spectrum (Fig. 2, right) are: (N,H<sup>N</sup>) = (124.46,9.008) ppm for D123 and (124.56,9.021) ppm for R134. At the 14 T magnetic field strength used, these differences in chemical shift correspond to 5.9 Hz (N) and 7.8 Hz (H<sup>N</sup>), respectively. This is 3.7 (N) or 2.6 (H<sup>N</sup>) times larger than the observed variance of chemical shifts in the <sup>15</sup>N-edited NOESY spectrum. Therefore, the two sets of cross-peaks clearly can be identified as belonging to different amides.

In order to visualize these very small chemical shift differences between cross-peaks in two dimensions, it is particularly useful to generate "orthogonal strip plots" for the amides (Fig. 2). For this, two small cross-sections (strips) through the three-dimensional spectrum, taken perpendicular to the F2 (<sup>15</sup>N) and F3 (<sup>1</sup>H<sup>N</sup>) axis of the 3D spectrum, are displayed next to each other. These strips are centered around the F2 and F3 frequencies of interest and therefore correspond to narrow regions of the (F1,F3) and (F1,F2) planes. The common F1-dimension is plotted for both regions with the same scale in the vertical direction. In Figure 2, the width of the strip in the <sup>1</sup>H<sup>N</sup>-dimension is 0.15 ppm (horizontal axis, left side of strips), whereas the width of the strips in the <sup>15</sup>N-dimension is 1.2 ppm (horizontal axis, right side of strips). As the strips in



**Fig. 1.** Histogram of the relative probability of deviations from the mean frequency of assigned peaks, corresponding to individual nuclei for the  $^{15}\text{N}$ -edited NOESY spectrum of NEF $^{\Delta 2-39, \Delta 159-173}$ . **A:** Deviations of the assigned  $^1\text{H}$  frequencies in the indirect dimension. **B:** Same as **A**, for  $^{15}\text{N}$  dimension. **C:** Same as **A** for directly detected  $^1\text{H}^{\text{N}}$  dimension.

Figure 2 are centered at the amide frequencies of D123, it is immediately recognized that the centers for the HNCO cross-peak and for the  $\text{H}_{i-1}^{\alpha}$  peak in the  $^{15}\text{N}$ -edited NOESY for R134 are shifted downfield in both the  $^{15}\text{N}$  and  $^1\text{H}^{\text{N}}$  frequencies. The display of these chemical shift deviation patterns for several spectra in the same graphic makes it easy to classify the individual cross-peaks as belonging either to the amide of D123 or the amide of R134.

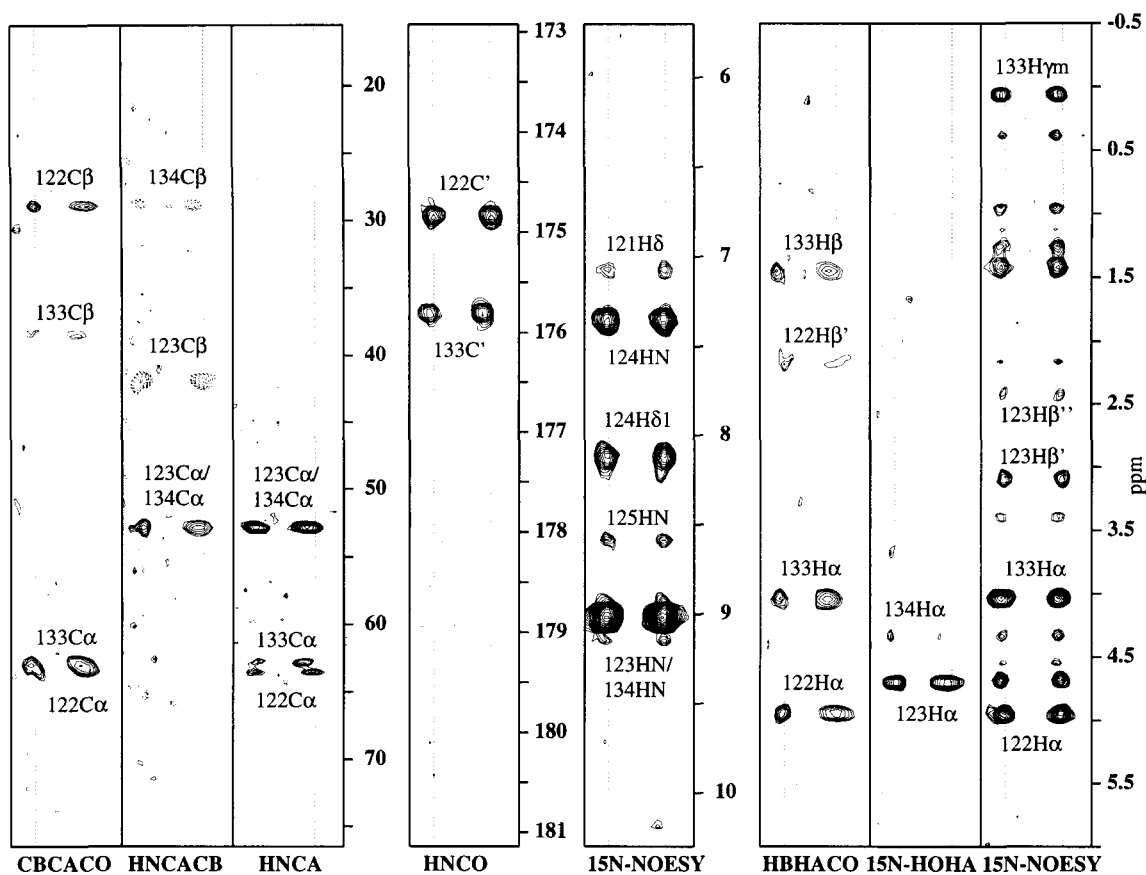
#### *Problems associated with fast amide proton exchange*

As all experiments are carried out at pH 8.0 and 35 °C, the intrinsic rates at which amide protons exchange with  $\text{H}_2\text{O}$  are very fast (Bai et al., 1993). The  $\text{H}_2\text{O}$  magnetization has a long  $T_1$  relaxation time and, unless special care is taken, its net magnetization will be greatly reduced when the NMR experiment is repeated at a rate faster than  $1/T_1$ . Rapid exchange of amide and hydroxyl protons with this largely saturated water resonance, followed by NOE magnetization transfer within the protein, will then attenuate the protein NMR signals. This problem can be solved by the incorporation of water flip-back pulses (Grzesiek & Bax, 1993b) into the amide proton out-and-back experiments (Kay et al., 1994), such as HNCO, HNCA, HNCACB, and by the use of radiation damping during the

NOESY mixing period (Lippens et al., 1995). Using such procedures, all protons except those that exchange faster than ca.  $30\text{ s}^{-1}$  can be detected. Of the 155 backbone amide protons in NEF $^{\Delta 2-39}$ , only 18 could not be detected as a result of fast solvent exchange, and only three of these, Q104, R105, and H193, lie in the structured core region of the protein. This core region is defined as the 102 residues (54–56, 58, 72, 73, 75–95, 97–148, 181–203), which have a heteronuclear  $^{15}\text{N}\{-^1\text{H}\}$  NOE larger than 0.6 or, if the NOE could not be measured, are adjacent to these residues.

#### *Deletion of residues 159 to 173*

Figure 3 shows the  $^1\text{H}\text{-}^{15}\text{N}$  HSQC spectrum of NEF $^{\Delta 2-39}$  together with assignments for the most well-resolved residues. Clearly, a large number of residues are located in the random coil region around 120 ppm ( $^{15}\text{N}$ ) and 8.2 ppm ( $^1\text{H}$ ). A list of resonance assignments for NEF $^{\Delta 2-39}$  is given in the Electronic Appendix. For residues K152 to D174 of NEF $^{\Delta 2-39}$ , the deviations from random coil values for the  $\text{C}^{\alpha}$  and  $\text{C}^{\beta}$  shifts are considerably smaller than 1 ppm, indicating that this region does not adopt a defined secondary structure. Furthermore, all of the assigned amide protons in this region show strong cross-peaks with the water resonance in the  $^{15}\text{N}$ -edited NOESY spectrum (Fig. 4). The intensity of these



**Fig. 2.** Orthogonal strip plots (see text) for the backbone assignment experiments extracted from the three-dimensional data cubes at the  $^1\text{H}$ - $^{15}\text{N}$  amide frequency pair of D123 of NEF $^{\Delta 2-39}$ . Individual experiments are marked at the bottom of each strip. The strip plots have a width of 0.15 ppm in the  $^1\text{H}^{\text{N}}$  dimension (left side of strips), and of 1.2 ppm in the  $^{15}\text{N}$  dimension (right side of strips). Vertical axes correspond to  $^{13}\text{C}^{\alpha}/^{13}\text{C}^{\beta}$  frequencies for CBCA(CO)NH, HNCACB, and HNCA, to  $^{13}\text{C}'$  frequencies for HNCO, and to  $^1\text{H}$  frequencies for the  $^{15}\text{N}$ -edited NOESY, HBHA(CO)NH and  $^{15}\text{N}$ -edited HOHAHA experiments, respectively.

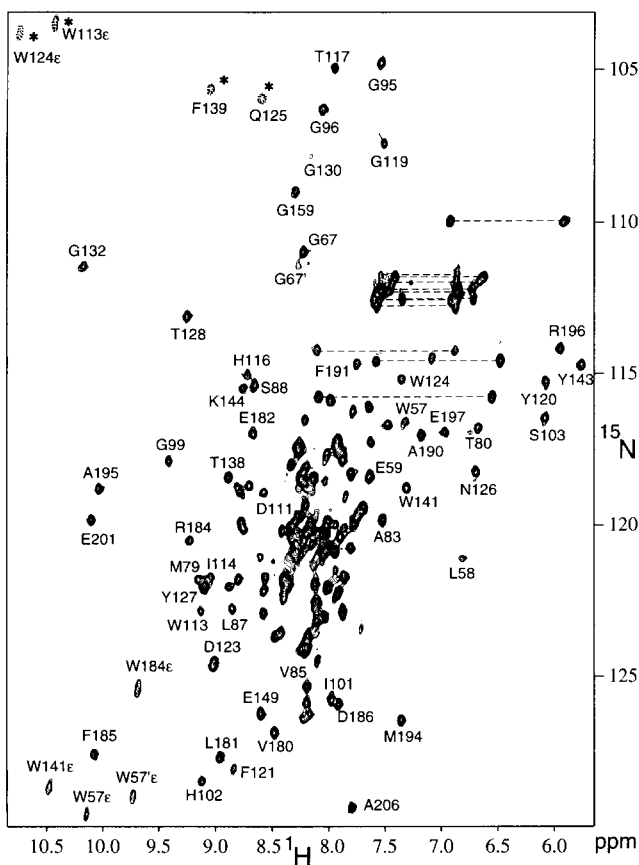
cross-peaks indicates that the amide protons exchange with the solvent at rates  $\geq 10 \text{ s}^{-1}$ . Residues K152 to D174, therefore, are not involved in stable hydrogen bonds and must be solvent exposed. Moreover, the NOE patterns of a four-dimensional  $^{15}\text{N}$ - $^{15}\text{N}$ -edited NOESY carried out on a  $^{15}\text{N}/^2\text{H}$ -labeled sample of NEF $^{\Delta 2-39}$  in  $\text{H}_2\text{O}$  (Grzesiek et al., 1995a) revealed that these residues are bordered on both sides by the two anti-parallel  $\beta$ -strands B4 and B5 (Fig. 8), thus forming a loop-like structure.

Based on the above findings, we deleted 15 residues (159–173) within this loop with the aim of improving the spectral properties of Nef. Unchanged NOE patterns and minimal differences in chemical shifts between NEF $^{\Delta 2-39, \Delta 159-173}$  and NEF $^{\Delta 2-39}$  indicated that the residues of the loop-deletion mutant NEF $^{\Delta 2-39, \Delta 159-173}$  retained the same structure as the corresponding residues in NEF $^{\Delta 2-39}$ . Figure 5 shows the weighted average of amide  $^1\text{H}$  and  $^{15}\text{N}$  chemical shift differences between the two mutants. For most of the residues, this average is considerably smaller than 0.1 ppm, with the exception of a small number of residues adjacent to the deletion site, i.e., K152, E154, D174, D175, E179, and also G132, which is part of the hairpin connecting strands B2 and B3 (Fig. 8). As B2 and B3 belong to the anti-parallel  $\beta$ -sheet consisting of strands B1 to B5, G132 is located close to the site of the deletion in the loop between strands B4 and B5.

Measurement of the concentration dependence of amide proton  $T_2$  values revealed that the deletion of residues 159 to 173 did not

affect Nef's tendency of self-association. However, at 0.6 mM Nef and  $35^\circ\text{C}$ , this deletion increased the average  $T_2$  for non-flexible residues from  $\sim 13$  ms to  $\sim 14$  ms for  $^1\text{H}^{\text{N}}$  and from 63 ms to 69 ms for  $^{15}\text{N}$ . This increase in  $T_2$  of  $\sim 10\%$  is attributed to the  $\sim 9\%$  reduction in Nef's molecular weight resulting from the deletion of 15 amino acids. Although the increase in  $T_2$  seems moderate, the improvement affects all evolution periods of a multi-dimensional experiment as well as the efficiencies of all the magnetization transfer steps. Therefore, the observed gain in sensitivity for the 3D experiments listed in Table 2 is considerably larger than 30%. Consequently, the determination of the three-dimensional structure was carried out for the NEF $^{\Delta 2-39, \Delta 159-173}$  mutant, rather than on NEF $^{\Delta 2-39}$ .

The deletion of residues 159 to 173 did not affect the binding of Nef to either the SH3 domain of Hck (Grzesiek et al., 1996a) or binding to a C-terminal peptide of CD4 (Grzesiek et al., 1996b). Therefore, the V148–V180 loop region, connecting strands B4 and B5, is not involved in the interaction of Nef with these molecules and remains flexible in complexes with Hck SH3 and the CD4 peptide. Similarly, this loop region was found to be disordered in the crystal structure of Nef complexed with Fyn SH3 (Lee et al., 1996). It was previously shown (Grzesiek et al., 1996a) that the regions of defined secondary structure in Nef correspond to highly conserved contiguous sets of amino acids in sequences derived from HIV-1, HIV-2, and primate lentiviruses isolates (Shugars



**Fig. 3.**  $^1\text{H}$ - $^{15}\text{N}$  HSQC spectrum of NEF $^{\Delta 2-39}$  together with assignments for the most well-resolved residues. Resonances marked by asterisks are aliased in the  $^{15}\text{N}$  dimension. Dashed lines mark  $\text{NH}_2$  groups of Asn and Gln residues.

et al., 1993). In contrast, only a relatively small subset of non-contiguous residues in the V148–V180 loop region is highly ( $\geq 95\%$  identity) conserved, i.e., V148, E154, A156, E160, L164, L165, H166, P167, H171, G172, and D175 (Shugars et al., 1993). However, the length of this loop ( $\sim 30$  amino acids) is remarkably well preserved (Shugars et al., 1993). Although this loop is not structured in the previously described complexes of Nef, it might be involved in the interaction with another as yet unidentified ligand. Interestingly, a recent mutational analysis of Nef showed that a D174A,D175A mutation gave rise to fully stable Nef, which is completely inactive in its down-regulation of CD4 (Aiken et al., 1996).

#### Assignment of side chain resonances

Methyl groups were assigned using modified versions of the HCCH-COSY and HCCH-TOCSY experiments (Table 2a). For larger proteins, spectral overlap is usually severe in both HCCH-COSY and HCCH-TOCSY. Therefore, the  $^{13}\text{C}$  evolution period, following the  $^{13}\text{C}$  isotropic mixing, was replaced by a constant-time HSQC sequence (Santoro & King, 1992; Van de Ven & Philippens, 1992; Vuister & Bax, 1992). In this way, the more favorable relaxation properties of the methyl carbon and proton nuclei could be exploited to achieve sufficient resolution in the second and third frequency dimensions.

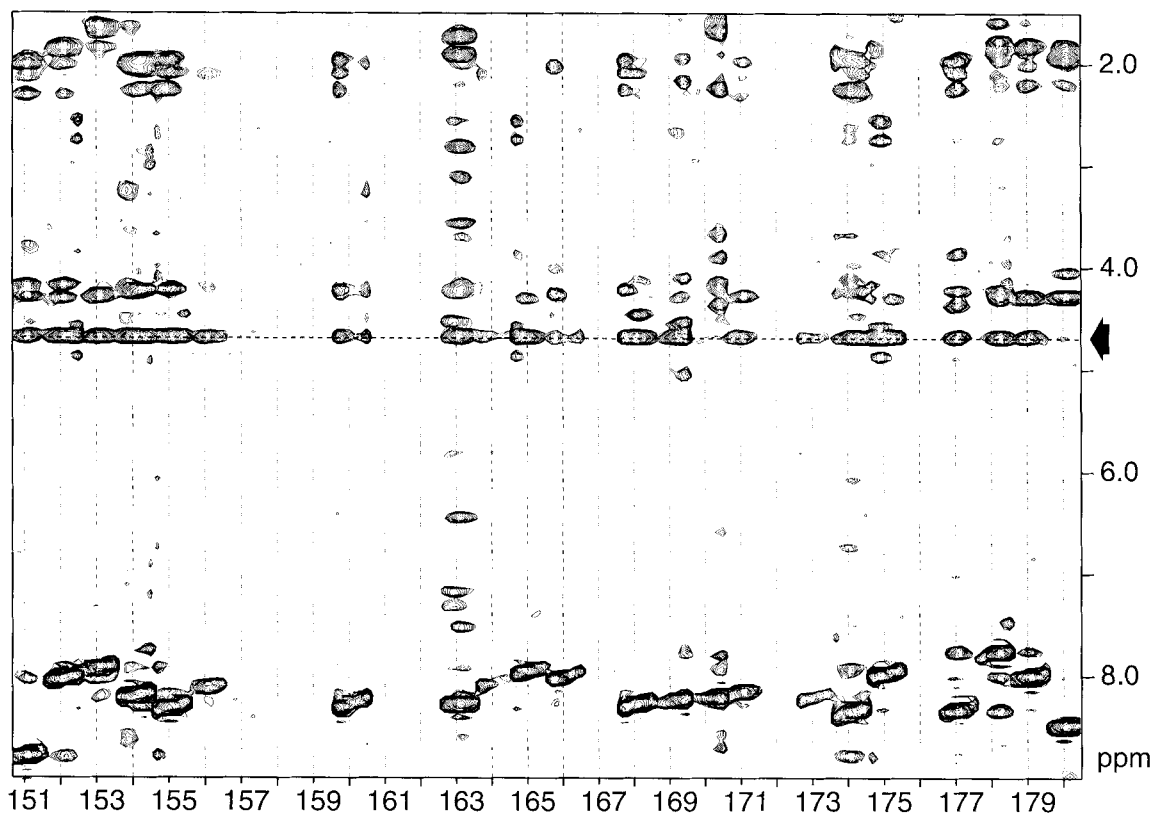
Resonance assignments for the aromatic protons of phenylalanine residues were carried out on a sample of NEF $^{\Delta 2-39}$  labeled uniformly with  $^{15}\text{N}$  and  $^{13}\text{C}$ , but with  $^{12}\text{C}$  inverse labeling for phenylalanine (Vuister et al., 1994). The assignments were derived from a combination of two-dimensional homonuclear NOESY and HOHAHA experiments, which were selective for  $^{12}\text{C}$ -attached protons in both dimensions (Bax et al., 1994; Vuister et al., 1994).

Assignments of NEF $^{\Delta 2-39}$  were transferred to NEF $^{\Delta 2-39,\Delta 159-173}$  in a two-step process. First, assignments of  $^1\text{HN}$ ,  $^{15}\text{N}$ ,  $^{13}\text{C}^\alpha$ , and  $^{13}\text{C}^\beta$  resonances were established by information from the CBCA-(CO)NH and HNCACB experiments carried out on NEF $^{\Delta 2-39,\Delta 159-173}$  and by comparison to the NEF $^{\Delta 2-39}$  data (Table 2a, Fig. 5). Second, assignments for the remaining resonances were transferred to NEF $^{\Delta 2-39,\Delta 159-173}$  by comparing data of the three-dimensional  $^{13}\text{C}$ - or  $^{15}\text{N}$ -edited NOESY experiments for both proteins. Further assignments, in particular for aromatic groups, were derived from a  $^{13}\text{C}$ -edited NOESY experiment with optimized detection of aromatic  $^{13}\text{C}$  and  $^1\text{H}$  resonances (Table 2c). This separate experiment was necessary because the  $^{13}\text{C}$  radio-frequency field strength on the 8-mm probe was not sufficient to cover both aliphatic and aromatic  $^{13}\text{C}$  resonances at the same time. Similar to the HCCH-COSY and HCCH-TOCSY experiments, a  $^{13}\text{C}$ -edited NOESY spectrum with a constant-time  $^1\text{H}$ - $^{13}\text{C}$  HSQC period was also collected, in order to achieve better resolution for methyl groups (Table 2c). A total of 2429 cross-peaks from the various three-dimensional NOESY spectra in Table 2c were assigned for NEF $^{\Delta 2-39,\Delta 159-173}$ , which served not only to derive NOE distance information, but also to verify and extend previous assignments of side chain resonances.

#### Stereoassignments and $\chi_1$ -angle restraints

Stereo-assignments and  $\chi_1$ -restraints for valine and threonine side chains were derived from measurements of the 3-bond J-couplings between side chain  $^{13}\text{C}^\gamma$  and backbone  $^{13}\text{C}'$  and  $^{15}\text{N}$  nuclei as described earlier (Grzesiek et al., 1993; Vuister et al., 1993). Stereo-assignments for leucine methyl groups were derived from a two-dimensional  $^1\text{H}$ - $^{13}\text{C}$  constant-time HSQC (Vuister & Bax, 1992; Hiroaki et al., 1996) carried out on NEF $^{\Delta 2-39,\Delta 159-173}$  biosynthetically labeled at 10% in  $^{13}\text{C}$  (Neri et al., 1989).

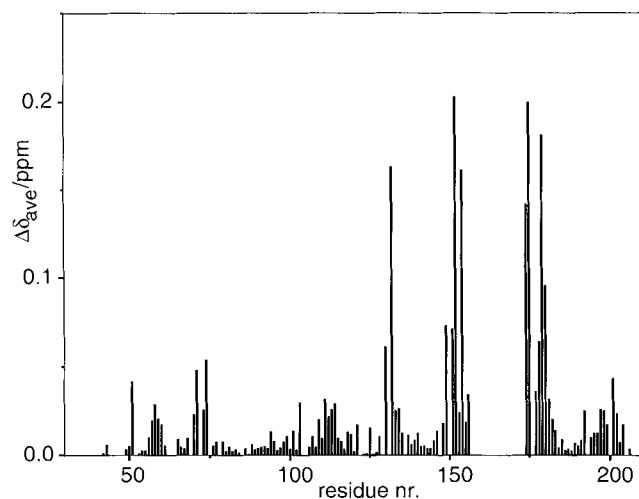
Additional  $\chi_1$ -restraints and stereo-specific assignments for  $\text{H}^\beta$ -protons were derived for many residues by a combined analysis of data from  $^{15}\text{N}$ -edited NOESY and ROESY experiments, together with information from HNHA and HAHB quantitative J-correlation experiments and  $^{13}\text{C}$ -edited NOESY. This is illustrated in Figure 6, which shows strip plots extracted from the corresponding 3D spectra at the  $^1\text{H}^\text{N}, ^{15}\text{N}$ , and  $^1\text{H}^\alpha, ^{13}\text{C}^\alpha$  frequency pairs of residue W183. The ratio of the  $\text{H}^\alpha$  cross-peak to the  $\text{H}^\text{N}$  diagonal peak intensity in the HNHA J-correlation experiment indicates that the  $\text{H}^\text{N}$ - $\text{H}^\alpha$  3-bond J-coupling is larger than 6 Hz. This restrains the  $\phi$ -angle for W183 to values smaller than  $\sim -75^\circ$ , as expected for a  $\beta$ -strand conformation. The ratio of the  $\text{H}^\beta$  cross-peak to the  $\text{H}^\alpha$  diagonal peak intensity in the HAHB quantitative J-correlation spectrum indicates a value of  $>8$  Hz for the  $\text{H}^\alpha$ - $\text{H}^\beta$  3-bond J-coupling. As the  $\text{H}^\beta$  cross-peak is absent in the HAHB experiment, the  $\text{H}^\alpha$ - $\text{H}^\beta$  J-coupling must be smaller than  $\sim 4$  Hz. Therefore,  $\text{H}^\beta$  is *trans* with respect to  $\text{H}^\alpha$  and the  $\chi_1$ -angle is either  $\sim -60^\circ$  or  $\sim 180^\circ$ . On the other hand, the  $^{15}\text{N}$ -edited ROESY shows a cross-peak from the amide proton of W183 to the intraregional  $\text{H}^{\epsilon 3}$  proton. In contrast to NOE, ROE intensities are decreased by indirect magnetization transfer during the mixing period and present a much more



**Fig. 4.** Strip plot of the  $^{15}\text{N}$ -edited NOESY spectrum of NEF $^{\Delta 2-39}$  (mixing time 70 ms), extracted at the  $^1\text{H}$ - $^{15}\text{N}$  amide frequency pairs of residues 151 to 179. The vertical axis shows the indirectly detected  $^1\text{H}$  dimension. The arrow and horizontal dashed line mark the position of the water resonance.

reliable measure for upper distance limits (Bothner-By et al., 1984; Kuboniwa et al., 1995). For W183, the  $\text{H}^{\text{N}}$ - $\text{H}^{\epsilon 3}$  ROE peak is of similar intensity as the intraresidue  $\text{H}^{\text{N}}$ - $\text{H}^{\alpha}$  ROE cross-peak. As the  $\text{H}^{\text{N}}$ - $\text{H}^{\alpha}$  distance is always less than 3.1 Å (Wüthrich, 1986), the

$\text{H}^{\text{N}}$ - $\text{H}^{\epsilon 3}$  distance must also be smaller than this value. This immediately rules out a  $\chi_1$ -angle of  $180^\circ$  and restrains even the  $\chi_2$ -angle to  $\sim -90^\circ$ . The relatively strong  $\text{H}^{\alpha}$ - $\text{H}^{\delta 1}$  NOE cross-peak in the  $^{13}\text{C}$ -edited NOESY spectrum confirms this side-chain conformation. The above information identifies  $\text{H}^{\beta'}$  as  $\text{H}^{\beta 2}$  and  $\text{H}^{\beta''}$  as  $\text{H}^{\beta 3}$ . Results of a recently developed pair of two-dimensional experiments, HN(CG) and HN(CO)(CG), which measure 3-bond J-couplings from aromatic  $^{13}\text{C}^{\gamma}$  to backbone  $^{15}\text{N}$  and  $^{13}\text{C}'$  nuclei, yielded additional  $\chi_1$  information for 19 aromatic residues and confirmed a  $\chi_1$ -angle of  $-60^\circ$  for W183 (Table 2b; Hu et al., 1997).

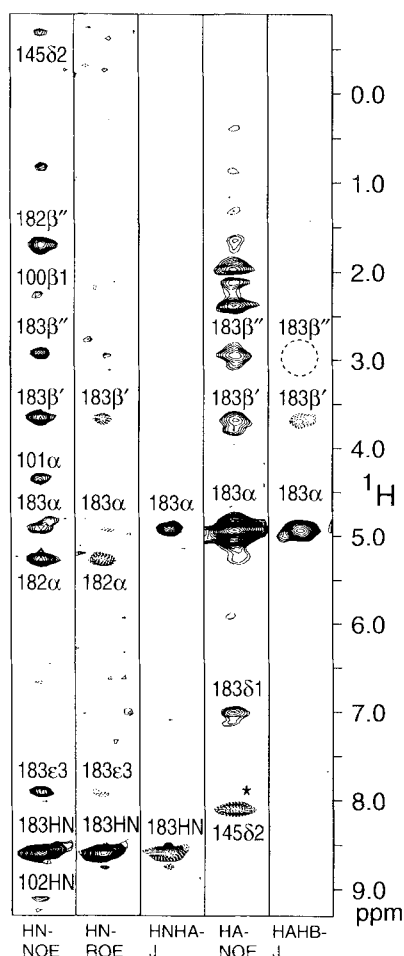


**Fig. 5.** Weighted average of the  $^1\text{H}^{\text{N}}$  and  $^{15}\text{N}$  chemical shift differences between NEF $^{\Delta 2-39}$  and NEF $^{\Delta 2-39, \Delta 159-173}$  as a function of residues number. Average chemical shift changes are calculated as  $[0.5 ((\Delta\delta_{\text{HN}})^2 + (\Delta\delta_{\text{N}}/5)^2)]^{1/2}$  where  $\Delta\delta_i$  is the chemical shift difference in ppm of nucleus  $i$  between NEF $^{\Delta 2-39}$  and NEF $^{\Delta 2-39, \Delta 159-173}$ .

#### Detailed description of Nef structure

Figure 7 summarizes the secondary structure information on NEF $^{\Delta 2-39, \Delta 159-173}$  contained in the  $^{15}\text{N}$ -edited NOESY, amide proton exchange rates,  $^3J_{\text{HNH}\alpha}$  values, and  $^{13}\text{C}^{\alpha}$  and  $^{13}\text{C}^{\beta}$  secondary shifts (Spera & Bax, 1991; Wishart et al., 1991). NOE contacts, which define the  $\beta$ -sheet topology, are shown in Figure 8. As a result of Nef's tendency to aggregate, the quality of the raw NMR data is relatively low, and the original set of Nef structures deposited in the Brookhaven Protein Data Bank (PDB, accession code 1NEF; Grzesiek et al., 1996a) was based on a rather limited set of structural constraints. This original structure has now been refined by including a substantial number of additional constraints, mainly related to side-chain orientations, but also for the backbone torsion angles. The majority of these constraints is derived from the additional analysis of  $^{15}\text{N}$ -edited ROESY data and quantitative three-





**Fig. 6.** Strip plots containing information relevant for the definition of side chain geometry and stereospecific assignments. Individual strips correspond to the  $^{15}\text{N}$ -edited NOESY (HN-NOE),  $^{15}\text{N}$ -edited ROESY (HN-ROE), the HNHA J-coupling experiment (HNHA-J), the  $^{13}\text{C}$ -edited NOESY (HA-NOE), and the HACAHB-COSY (HAHB-J). Strips are extracted from the three-dimensional data cubes of the individual experiments at the  $^1\text{H}$ - $^{15}\text{N}$  (HN-NOE, HN-ROE, HNHA-J) or  $^1\text{H}^\alpha$ - $^{13}\text{C}^\alpha$  (HA-NOE, HAHB-J) frequency pairs of W183 in NEF $^{\Delta 2-39, \Delta 159-173}$ . The resonance marked by an asterisk (HA-NOE) is aliased in the indirect  $^1\text{H}$  dimension.

bond J-coupling experiments, including HNCOCO ( $\phi_1$  angle), HACAHB-COSY ( $\chi_1$  angle), and HN(CO)(CG)/HN(CG) ( $\chi_1$  of aromatic residues) (Table 2). A refined set of 40 simulated annealing structures has been deposited in the PDB (accession code 2NEF).

A ribbon diagram of this refined structure is shown in Figure 9, and Table 3 provides a summary of the structural statistics. For the non-mobile residues defined in Table 3, the average RMSD from the mean for the non-hydrogen backbone atom positions improved from 0.98 Å (1NEF) to 0.81 Å (2NEF). For all non-hydrogen atoms, the RMSD decreased from 1.62 Å (1NEF) to 1.32 Å (2NEF). For the same residues, the standard deviation of the  $\chi_1$ -angle from ideal rotameric states improved from 21.7° to 17.5°. A 17.5° RMSD is evaluated to be good by PROCHECK-NMR quality criteria (Laskowski et al., 1996) and corresponds to a value typically found for a crystal structure solved at a resolution of 1.9 Å. Using the same program, 73% of the residues have  $\phi$  and  $\psi$  angles, which positions them in the most favored region of the Ramachandran map (64%

for 1NEF), indicating that the local quality of the backbone is lower and corresponds to what is typical for a 2.7 Å crystal structure.

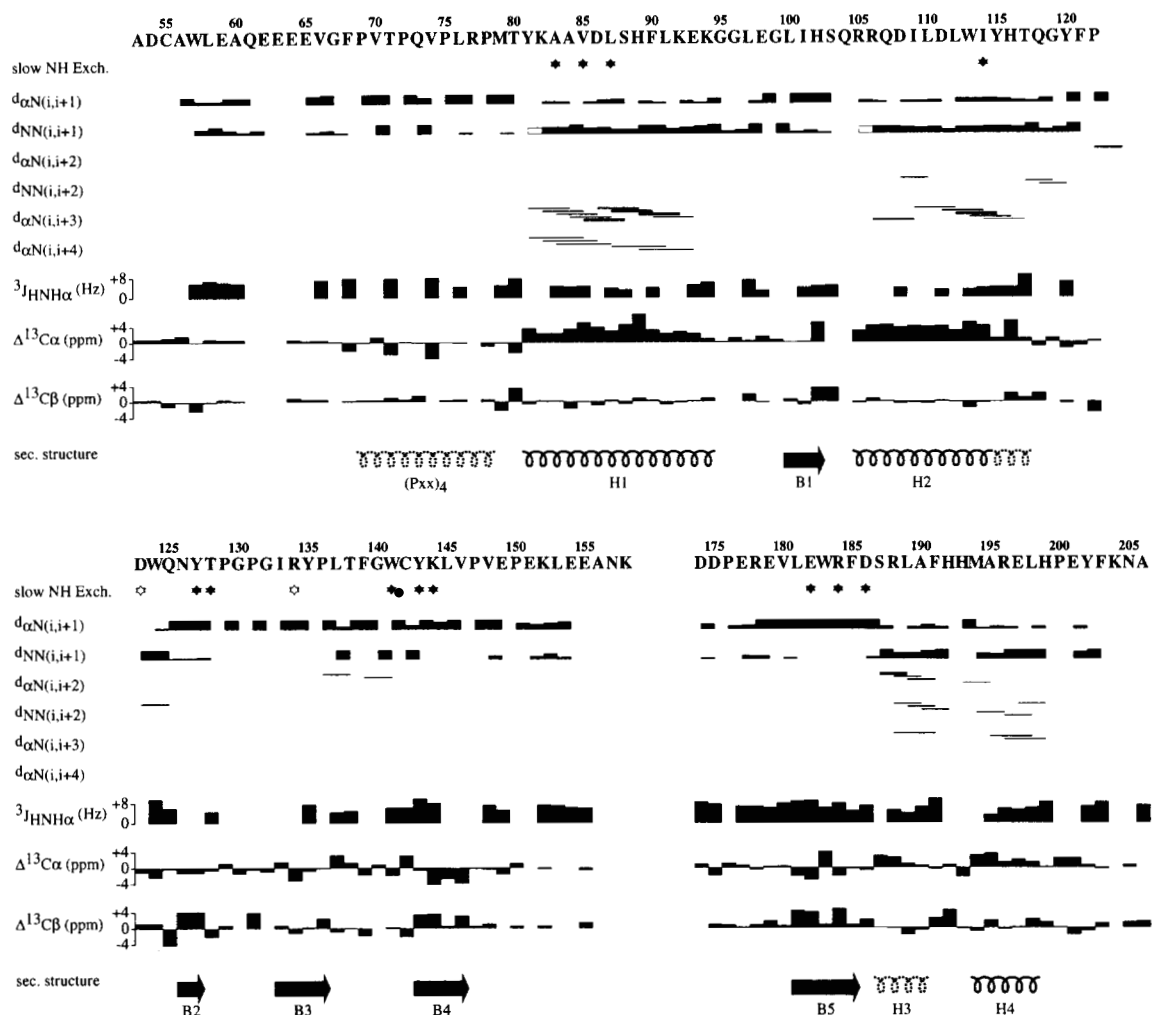
Figure 10 depicts the RMSD for the backbone and all non-hydrogen atoms as a function of residue number. The largest RMSD values are found for the loop regions between residues 59 and 72 as well as between 150 and 181, which exhibit flexibility on the ns time scale (see below). Two other regions, i.e., the C-terminal residues beyond P200 and the wide turn around residue P131, also have above average RMSD values, but these result from the limited number of experimental restraints and not from internal flexibility.

The first 14 residues of NEF $^{\Delta 2-39, \Delta 159-173}$  are disordered in solution as judged by their near-zero secondary  $^{13}\text{C}^\alpha$  and  $^{13}\text{C}^\beta$  shifts and fast backbone amide hydrogen exchange. Because of this rapid exchange, cross-peaks in the  $^{15}\text{N}$ -edited NOESY for these residues are weak and no interresidue contacts could be established. As expected for such a dynamically disordered region, aliphatic resonances for these residues are narrower than for the rest of the protein, indicative of fast internal motion (see also Fig. 11). The  $^{13}\text{C}$ -edited NOE spectra in this region did not reveal any contacts to the rest of the protein.

Residues A53 to A60 show more pronounced deviations from the random coil values for their  $^{13}\text{C}^\alpha$  and  $^{13}\text{C}^\beta$  chemical shifts (Fig. 7). However, the amide protons in this region exchange relatively rapidly with the solvent and are difficult to observe.  $^{15}\text{N}$  relaxation measurements (Fig. 11) indicate that this region is relatively rigid on the nanosecond time scale. Three-bond  $J_{\text{HNH}\alpha}$  couplings, which could only be measured for residues 57 to 60, range from 6 to 7 Hz and restrain the  $\phi$  angle for these residues to values of about 75°. A set of unambiguous NOE contacts between residues W57/L58 and G95, L97, L100, I109, and L110 establishes a connection between this region and a concave hydrophobic pocket, formed by the C-terminal end of helix H1, the N-terminal end of helix H2, and the region connecting these two helices. The aromatic side chain of W57 inserts itself into this pocket with hydrophobic contacts to the leucine and isoleucine side chains. We have shown previously that W57/L58 and neighboring residues form a binding site for a peptide derived from the C-terminal tail of CD4 (Grzesiek et al., 1996b). A di-leucine motif within this peptide is critical both for the peptide binding to Nef as well as for the down-regulation of CD4 by Nef (Aiken et al., 1994; Anderson et al., 1994; Salghetti et al., 1995). In the final set of calculated Nef structures, the aromatic side chain of W57 does not completely fill the hydrophobic pocket between the C- and N-termini of helices H1 and H2. Presumably, the hydrophobic side chains of the two critical leucines of CD4 can insert themselves into the remaining space. The existence of this hydrophobic crevice was also noted in the crystal structure of the Nef/Fyn complex (Lee et al., 1996). However, the contact of this crevice to W57/L58 was not reported, and the region of residues 54 to 69 was described as disordered. This is probably caused by the flexibility of the region immediately following W57 and L58.

Residues 60 to 68 are quite flexible on the nanosecond time scale (vide infra), have nearly random coil shifts for  $^{13}\text{C}^\alpha$ ,  $^{13}\text{C}^\beta$ ,  $^1\text{H}^\text{N}$ , and  $^{15}\text{N}$ , and also show rapid amide hydrogen exchange. This region, therefore, does not adopt an ordered conformation in solution and forms a solvent-exposed loop. As a result of fast hydrogen exchange, the amide signals of residues 62 to 64 could not be observed.

Interestingly, the HIV-1 protease cleaves Nef between residues W57 and L58 (Freund et al., 1994b). The HIV-1 protease is thus able to remove Nef's N-terminal membrane anchor, and HIV-1



**Fig. 7.** Summary of the sequential and medium-range NOEs involving  $H^N$  and  $H^\alpha$  protons, amide exchange data,  $^3J_{\text{HNH}\alpha}$  coupling constants, and  $^{13}\text{C}^\alpha$  and  $^{13}\text{C}^\beta$  secondary shifts observed for NEF $^{\Delta 2-39, \Delta 159-173}$ . Amide protons which are still present in the 2D  $^1\text{H}$ - $^{15}\text{N}$  spectrum after exchange to  $\text{D}_2\text{O}$  (see text) are marked by stars. The thickness of the lines for the NOE connectivities corresponds to the intensity of the crosspeaks. Open symbols depict tentative assignments due to overlap in the 2D  $^1\text{H}$ - $^{15}\text{N}$  spectrum. Helices drawn as solid lines are  $\alpha$ -helical. The dashed helix marked as (Pxx) $_4$  corresponds to a type II polyproline helix, whereas the dashed helix H3 adopts a  $^3_10$  conformation.

viral particles have been shown to contain the resulting 18 kDa Nef core protein (Welker et al., 1996). This virion-associated Nef core protein without the membrane anchor might, therefore, play a role in the early stages of infection prior to viral gene expression (Welker et al., 1996). Removal of W57 and its preceding residues by the protease also removes the anchor by which the C-terminal end of the flexible region between residues 60 to 68 is attached to the rest of the protein at the hydrophobic pocket between helices H1 and H2. Experimentally, we observed that a NEF $^{\Delta 2-59}$  deletion mutant has amide proton  $T_2$  values of  $\sim 9$  ms, i.e., considerably shorter than the  $\sim 13$  ms observed both for NEF $^{\Delta 2-39}$  and an additionally investigated NEF $^{\Delta 2-52}$  deletion mutant. However, the NEF $^{\Delta 2-59}$  deletion mutant retains the same structure as NEF $^{\Delta 2-39}$ , as judged by the nearly identical chemical shifts for the amide  $^1\text{H}^N$  and  $^{15}\text{N}$  nuclei of amino acids common to both mutants. Presumably, in the absence of W57 the very hydrophilic region between residues 60 and 68 protrudes freely into the surrounding water, thereby exposing the underlying hydrophobic region between the C- and N-termini

of helices H1 and H2. This, in turn, leads to additional aggregation and shortening of  $T_2$ s.

The residues of the polyproline region starting at P69 and ending at P78 are all characterized by positive  $\psi$  angles. Evidence for this comes from strong  $d_{\alpha\text{N}}(i-1, i)$  ROEs in cases where  $i$  is not a Pro residue, and from strong  $d_{\alpha\delta}(i-1, i)$  NOEs for  $i = \text{Pro}$ . These strong  $d_{\alpha\delta}(i-1, i)$  NOEs and the absence of  $d_{\alpha\alpha}(i-1, i)$  NOEs also restrict the  $i-1$  to  $i$  prolyl peptide bond to a *trans* conformation. Three-bond  $J_{\text{HNH}\alpha}$  couplings of 4 to 8 Hz restrict the  $\phi$  angles for all the non-Pro residues between P69 and P78 to values in the range of  $-90^\circ$  to  $-60^\circ$ . However, as will be discussed later, significant internal motions on the nanosecond time scale for the  $\text{H}^N$ -N bond vectors of residues 68, 71, and 74 indicate that the first half of the polyproline stretch is somewhat less rigid than the core of the protein. The restraints for both  $\phi$  and  $\psi$  for residues P69 to P78 agree with a left-handed poly-L-proline II helix conformation (Adzubei & Sternberg, 1993). Additional NOEs observed from residues 72-77 to residues 116-120 confirm this classification.

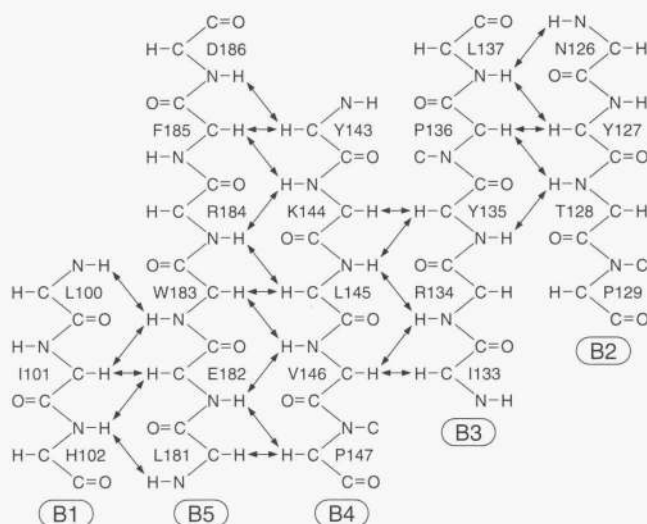


Fig. 8. NOE contacts defining the  $\beta$ -sheet topology for NEF $^{\Delta 2-39, \Delta 159-173}$ . Observed NOEs are drawn as arrows.

The first helix, H1, starts at Y81 and ends at K94, judged by the presence of consecutive  $d_{NN}(i, i + 1)$  and  $d_{\alpha N}(i, i + 3)$  NOEs,  $^3J_{\text{HNH}\alpha}$  values of about 4 Hz, and a consecutive set of large positive  $^{13}\text{C}^\alpha$  secondary shifts. The secondary shifts of T80 (negative  $\Delta\delta^{13}\text{C}^\alpha$  and positive  $\Delta\delta^{13}\text{C}^\beta$ ) are indicative of an N-cap residue (Gronenborn & Clore, 1994) and the amide proton of A83 is within hydrogen bonding distance to the side chain oxygen of T80 in the final set of calculated structures without using explicit hydrogen bond distance restraints. However, unlike the classical N-cap configuration (Harper & Rose, 1993), the side chain of A83 cannot function as an acceptor for a second hydrogen bond from the amide proton of T80. Helix H1 is  $\alpha$ -helical, as indicated by the presence of a large number of  $d_{\alpha N}(i, i + 3)$  and  $d_{\alpha N}(i, i + 4)$  NOEs.

H1 is followed by an irregular stretch of residues that precedes the first  $\beta$ -strand, B1, consisting of L100, I101, and H102. B1 forms one edge of an extended, five-stranded anti-parallel  $\beta$ -sheet, consisting consecutively of strands B1, B5, B4, B3, and B2 (Fig. 8). The  $\beta$ -strand backbone conformation of residues 100–102 is defined by a set of unambiguous NOEs from the backbone protons of L100 to H102 to the backbone protons of L181 to W183 in strand B5 (Fig. 8).

Due to fast hydrogen exchange it was not possible to assign the amide protons of residues Q104 and R105, which follow strand B1. Helix H2 starts at R105 and continues as a regular  $\alpha$ -helix to I114, as judged from a contiguous set of large positive  $^{13}\text{C}^\alpha$  secondary shifts,  $d_{NN}(i, i + 1)$  and  $d_{\alpha N}(i, i + 3)$  NOEs, and  $^3J_{\text{HNH}\alpha}$  values of about 4 Hz. Residues Y115 to T117 are located at the C-terminal end of H2 and do not adopt a regular  $\alpha$ -helical geometry. Although the  $d_{\alpha N}(i, i + 3)$  NOEs continue to T117, the large and positive  $^{13}\text{C}^\alpha$  secondary shifts already stop at I114 and the

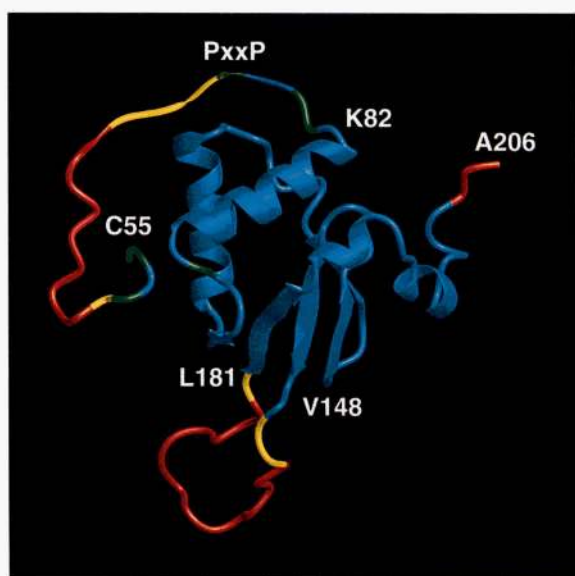


Fig. 9. Backbone representation of the refined structure of NEF $^{\Delta 2-39, \Delta 159-173}$ . Color coding depicts heteronuclear  $\{^1\text{H}\}-^{15}\text{N}$  NOE values:  $<0.43$  (red),  $<0.54$  (yellow),  $<0.65$  (green),  $\geq 0.65$  (blue). This graphic was generated with the program MOLMOL (Koradi et al., 1996).

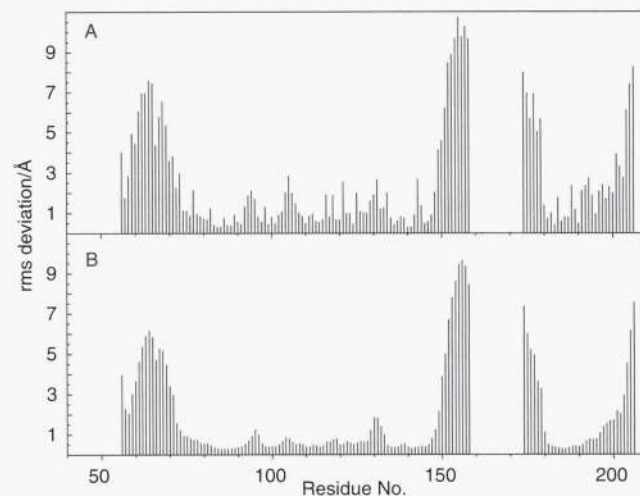


Fig. 10. RMSDs from the mean position for the final set of 40 structures as a function of residue number. (A) All non-hydrogen atoms; (B) backbone non-hydrogen atoms.

**Table 3.** Structural statistics<sup>a</sup>

RMSDs from experimental distance restraints (Å)	
All (818)	0.064 ± 0.003
Intraresidue (70)	0.042 ± 0.006
Interresidue sequential ( $ i - j  = 1$ ) (338)	0.069 ± 0.007
Interresidue short range ( $1 <  i - j  \leq 5$ ) (101)	0.088 ± 0.006
Interresidue long range ( $ i - j  > 5$ ) (245)	0.053 ± 0.006
H-bonds (64) <sup>b</sup>	0.043 ± 0.010
RMSDs from $^3J_{\text{HNH}\alpha}$ coupling constants (Hz) (91)	
RMSDs from experimental dihedral restraints (°) (157) <sup>c</sup>	0.40 ± 0.15
RMSDs from experimental secondary shifts (ppm)	
$^{13}\text{C}^\alpha$ (93)	1.31 ± 0.05
$^{13}\text{C}^\beta$ (91)	1.29 ± 0.07
Deviations from idealized covalent geometry <sup>d</sup>	
Bonds (Å) (2270)	0.004 ± 0.0001
Angles (°) (4101)	0.47 ± 0.01
Improvers (°) (1260)	0.39 ± 0.02
$E_{\text{LJ}}$ (kcal/mol) <sup>e</sup>	-558 ± 16
Coordinate precision (Å) <sup>f</sup>	
Backbone non-hydrogen atoms	0.81 ± 0.19
All non-hydrogen atoms	1.32 ± 0.18
PROCHECK quality indicators <sup>g</sup>	
Percentage of non-gly, non-pro residues in most favored region of Ramachandran plane	
$\chi_1$ pooled standard deviation (°)	72.5
$\chi_2$ pooled standard deviation (°)	17.5
	15.6

<sup>a</sup>These statistics comprise the ensemble of the final 40 simulated annealing structures. Individual simulated annealing structures are fitted to each other using residues 76 to 94, 97 to 102, 106 to 147, 181 to 191, and 194 to 199. The number of the various constraints is given in parentheses.

<sup>b</sup>For each backbone hydrogen bond constraint, there are two distance restraints:  $r_{\text{NH}\cdots\text{O}}$ , 1.7–2.5 Å,  $r_{\text{N}\cdots\text{O}}$ , 2.3–3.5 Å.

<sup>c</sup>These comprise 78  $\phi$  restraints from  $^{13}\text{C}'\text{-}^{13}\text{C}'$  J couplings (which partially duplicate the  $^3J_{\text{HNH}\alpha}$ -derived restraints), 10  $\psi$ , 55  $\chi_1$ , and 14 aromatic  $\chi_2$  angles.

<sup>d</sup>The improper torsion restraints serve to maintain planarity and chirality.

<sup>e</sup> $E_{\text{LJ}}$  is the Lennard-Jones van der Waals energy calculated with the CHARMM empirical energy function (Brooks et al., 1983) and is not included in the target function for simulated annealing and restrained minimization.

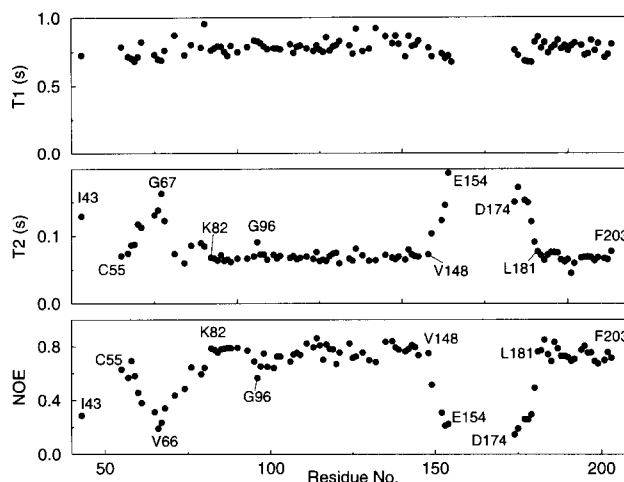
<sup>f</sup>The coordinate precision is defined as the average rms difference between the individual simulated annealing structures and the mean coordinates. Values are reported for residues 58, 72–95, 97–102, 106–148, 181–191, 194–202, i.e., for residues that do not exhibit large amplitude internal motions on the nanosecond time scale (Fig. 9)

<sup>g</sup>These values are calculated with the program PROCHECK-NMR (Laskowski et al., 1996). Values are reported for the non-mobile residues.<sup>f</sup>

$^3J_{\text{HNH}\alpha}$  value of 9.2 Hz for T117 constrains the  $\phi$ -angle for this residue to  $-120 \pm 20^\circ$ .

Helix H2 is followed by a stretch of irregular structure containing a helical turn at D123, which is immediately followed by the second  $\beta$ -strand, B2. It consists of residues N126, Y127, and T128, and forms the other edge of the antiparallel  $\beta$ -sheet (Fig. 8). Its  $\beta$ -strand secondary structure is derived from a set of NOEs from its backbone protons to the backbone protons of B3 (Fig. 8), as well as from the strongly downfield-shifted  $^{13}\text{C}^\beta$  resonances of N126 and Y127 (Fig. 7). B2 is followed by an irregular, wide turn consisting of residues P129, G130, P131, and G132.

The third anti-parallel  $\beta$ -strand, B3, consists of residues I133 to L137, which are hydrogen bonded to both B2 and B4, as evidenced by a series of NOE connectivities between backbone protons in all three strands (Fig. 8). Only two to three residues per strand par-



**Fig. 11.**  $^{15}\text{N}$   $T_1$ ,  $T_2$  and heteronuclear  $\{^1\text{H}\}\text{-}^{15}\text{N}$  NOE relaxation data for NEF $^{\Delta 2-39, \Delta 159-173}$  as a function of residue number.

ticipate in the hydrogen bond network between  $\beta$ -strands B2, B3, and B4. This was also shown in the crystal structure of the Nef-Fyn complex (Lee et al., 1996). The presence of two highly conserved Pro residues (Lee et al., 1996) at position 136 (in B3) and 147 (in B4) and of a Pro at position 129 (in B2) seems to prevent the formation of a more extended hydrogen bond network along the strands.

Strands B3 and B4 are connected by a type II  $\beta$ -turn, consisting of residues T138, F139, G140, and W141. Glycine 140 is found in the preferred ( $i + 2$ ) position of the turn (Wilmot & Thornton, 1988). Experimental evidence for this type of turn comes from a characteristic set of NOEs [strong  $d_{\text{NN}}(140,141)$ , strong  $d_{\alpha\text{N}}(139,140)$ , moderate  $d_{\alpha\text{N}}(139,141)$ , and absence of  $d_{\text{NN}}(139,140)$ ], which indicates type II over type I (Wüthrich, 1986). Furthermore, the slow hydrogen exchange of W141- $\text{H}^{\text{N}}$  is indicative of a hydrogen bond, presumably to the carbonyl oxygen of F139.

The fourth  $\beta$ -strand, B4, consists of residues Y143 to P147, and is marked by a contiguous set of large  $^3J_{\text{HNH}\alpha}$  values as well as negative  $^{13}\text{C}^\alpha$  and positive  $^{13}\text{C}^\beta$  secondary shifts. Numerous inter-strand backbone NOEs connect B4 to B5, whereas the region of contact to strand B3 is more limited (Fig. 8). P147 terminates B4, and  $^{15}\text{N}$  relaxation data (Fig. 11) indicate that after this residue the protein backbone becomes increasingly flexible and forms a long, solvent-exposed loop, connecting strands B4 and B5.

B5 consists of residues L181 to D186 and is identified by large  $^3J_{\text{HNH}\alpha}$  values, mostly negative  $^{13}\text{C}^\alpha$  and positive  $^{13}\text{C}^\beta$  secondary shifts, and numerous inter-strand backbone NOEs to both B1 and B4. Apparently, hydrogen bonding of the amide protons in B5 to the carbonyl oxygens in strand B4 is stronger than to strand B1, because the amide protons of W183 and F185 exchange considerably faster with the solvent than the amides of E182, R184, and D186. The connection of strands B5 and B1 (Fig. 8) exerts a strong constraint on the topology of Nef and on the position of helix H2, which connects strand B1 to B2.

B5 is followed by a third short helix, H3, consisting of residues S187 to A190. H3 adopts a  $3_{10}$  conformation, as judged from a consecutive set of  $d_{\alpha\text{N}}(i, i + 2)$  NOEs,  $^3J_{\text{HNH}\alpha}$  values of 4–6 Hz, and the absence of a contiguous set of strong positive  $^{13}\text{C}^\alpha$  secondary shifts. The helical structure ends at F191, which has a

$^3J_{\text{HNH}\alpha}$  value of 9.9 Hz. This constrains the F191  $\phi$ -angle to  $\sim -120^\circ$ . Residues F191 to H193 form a kink connecting helix H3 to H4. The amide proton of H193 in this kink is readily accessible to water and could not be observed because of fast hydrogen exchange.

Helix H4 (M194 to L198) immediately follows this kink. The absence of  $d_{\alpha\text{N}}(i, i+2)$  NOEs, two  $d_{\alpha\text{N}}(i, i+3)$  NOEs, and  $^3J_{\text{HNH}\alpha}$  values of 4–6 Hz characterize it as  $\alpha$ -helical.  $^{15}\text{N}$  relaxation data indicate that the backbone of the protein remains relatively rigid through residue 203 (Fig. 11).

#### Backbone dynamics

Figure 11 shows the results of  $^{15}\text{N}$   $T_1$ ,  $T_2$ , and heteronuclear  $\{^1\text{H}\}$ - $^{15}\text{N}$  NOE relaxation measurements carried out on NEF $\Delta 2-39, \Delta 159-173$ . The flexibility on the pico- to nanosecond time scale revealed by the heteronuclear  $\{^1\text{H}\}$ - $^{15}\text{N}$  NOE values has been color coded onto a backbone ribbon diagram of Nef in Figure 9. Quantitative analysis of the relaxation data according to the Lipari and Szabo model-free approach (Lipari & Szabo, 1982a, 1982b) is complicated by the effect of Nef's aggregation on the relaxation rates. Hence, we only present a qualitative discussion of Nef's internal mobility. However, a brief quantitative description of the effect of aggregation on  $T_1$  and  $T_2$  will also be presented.

Residues K82 through V148 and L181 through F203 show remarkably little variation in their  $T_2$  and NOE values (Fig. 11). These two regions comprise all the  $\alpha$ -helical and  $\beta$ -strand elements of Nef. The average  $T_2$  and NOE values for residues in this region are  $69 \pm 6$  ms and  $0.74 \pm 0.07$ , respectively. With the exception of G96, all heteronuclear NOE values in this region are higher than 0.6, indicating a rather low degree of internal mobility on the sub-nanosecond time scale. Residue G96 is affected to some extent by internal motions, as the low NOE value of 0.57 is accompanied by an increase in  $T_2$  to 91 ms. Interestingly, G96 is part of the irregular "hinge" region that connects helix H1 to strand B1.

For residue I43 in the N-terminal tail, hydrogen exchange is sufficiently slow to permit reliable measurement of relaxation data. Its NOE value of 0.29 and  $T_2$  of 129 ms clearly indicate a high degree of mobility. This also manifests itself in very narrow resonances for the aliphatic side chains of the residues in this N-terminal region. In contrast, residues in the vicinity of the HIV-1 protease cleavage site, i.e., C55, W57, L58, and E59, are restricted in their motion. This is reflected in NOE values of  $\sim 0.6$  and  $^{15}\text{N}$   $T_2$  values in the 70–88 ms range. These residues are followed by an increasingly mobile region, as evidenced by a monotonous decrease in NOE values and an increase in  $T_2$ . The internal mobility of this region reaches a maximum at residues V66 and G67 with NOEs of  $\sim 0.2$  and  $T_2$  values  $\geq 140$  ms. This mobile region, which also is characterized by fast amide proton exchange, corresponds to a highly conserved acidic region in the Nef primary sequence (Shugars et al., 1993).

From V66 on through the poly-Pro region (P69 to P78) until the beginning of helix H1, heteronuclear NOE values increase almost linearly to values of  $\sim 0.8$ . The polyproline stretch exhibits, therefore, some internal motion on the sub-nanosecond time scale, but this motion becomes increasingly restricted toward its C-terminal end, where most of the residues have NOE values larger than 0.6. Thus, although the backbone of this stretch is more flexible than the core of the protein, it is by no means "unstructured." Numerous NOEs from V74 and L76 to backbone and side-chain protons of residues H116–Y120 confirm that the poly-Pro region is anchored

to the core of the protein. Nef is the first protein for which the structure of an SH3 target site poly-Pro helix has been determined. As expected, in the complex with Fyn-SH3 it adopts the same conformation as is observed for poly-Pro model peptides, complexed to SH3 domains. Our results indicate that this poly-Pro region adopts a similar, albeit more flexible, conformation in the absence of the SH3 domain.

The solvent exposed loop between strands B4 and B5 is the third highly flexible region. Although this loop is considerably shortened in NEF $\Delta 2-39, \Delta 159-173$ , the flexibility in this region markedly increases from the last well-structured residue before the loop, V148, to its center at residues E154 to D174, with a minimum NOE of 0.15 and a maximum  $T_2$  of 194 ms. The first well-ordered residue following this loop is L181, at the beginning of strand B5.

#### Stability of the Nef core

As noted by Lee et al. (1996), the arrangement of strands and helices in Nef appears rather irregular. In particular, the large separation between helices H1 and H2 forms a hydrophobic solvent-exposed crevice. When complexed to the SH3 domain of either Hck (Grzesiek et al., 1996a) or Fyn (mutant R96I; Lee et al., 1996), the part of this crevice that is close to the N-terminal end of H1 can accommodate a hydrophobic isoleucine side chain from the SH3 RT-loop (the RT-loop connects strands 1 and 2 of the SH3 domains). This SH3 Ile residue is essential for the strong SH3-Nef binding affinity (Lee et al., 1995; Saksela et al., 1995). The part of the crevice near the C-terminal end of H1 accommodates the side chains of W57 and L58 and also serves as a binding pocket for the leucine side chains of the CD4 C-terminal peptide (Grzesiek et al., 1996b). Because of the "open" appearance of Nef, it had been suggested that its core might not be particularly stable (Lee et al., 1996). However, the present  $^{15}\text{N}$  relaxation data do not support this suggestion. As mentioned above, all residues that are part of Nef's helices or strands have heteronuclear NOEs of  $\sim 0.74$ , which is close to the values observed for the well-ordered regions in "stable" proteins such as staphylococcal nuclease (Kay et al., 1989) or HIV-1 protease/inhibitor complexes (Nicholson et al., 1995). Such high heteronuclear NOE values are typically found for residues with  $^1\text{H}$ - $^{15}\text{N}$  bond vector order parameters,  $S^2$ , of  $\geq 0.8$  and internal correlation times for the  $^1\text{H}$ - $^{15}\text{N}$  motion  $\leq 100$  picoseconds. Besides the absence of large amplitude motions on the sub-nanosecond time scale, very uniform  $T_2$  values indicate that there are no significant conformational exchange contributions. Thus, the Nef core is also rather stable in the millisecond regime. Evidence for the stability of Nef on the time scale of hours is obtained from backbone amide hydrogen exchange data. Indeed, a substantial number of amide protons is observed even several hours after the bulk water of the Nef solution has been exchanged to  $\text{D}_2\text{O}$  (Fig. 7). At  $35^\circ\text{C}$  and a pH value of 8.0, used during these experiments, unprotected amide protons exchange with the bulk water at a rate of about  $100 \text{ s}^{-1}$  (Bai et al., 1993).

#### Effect of aggregation on $^{15}\text{N}$ $T_1$ and $T_2$

In an equilibrium situation, where exchange between monomeric and multimeric states of Nef is fast compared to the relaxation rates but slow compared to the rotational correlation time, the observed relaxation rates correspond to the weighted average of relaxation rates for the different Nef species in solution. As the rotational diffusion rate of monomeric Nef is within the slow-tumbling limit, aggregation increases  $T_1$  and decreases  $T_2$ . For the

sake of simplicity, we assume isotropic tumbling with a rotational correlation time  $\tau_c$  proportional to the molecular weight and a simple monomer-dimer equilibrium. Then the relaxation rates can be expressed as:

$$R_{obs} = xR(\tau_c) + (1 - x)R(2\tau_c) \quad (1)$$

where  $R$  corresponds to a relaxation rate, e.g.,  $1/T_1$  or  $1/T_2$ ,  $R_{obs}$  is the observed rate,  $x$  the fraction of molecules in the monomeric state,  $R(\tau_c)$  the monomer relaxation rate, and  $R(2\tau_c)$  the dimer relaxation rate. An estimate of  $x$  and  $\tau_c$  can then be obtained by solving Equation 1 simultaneously for observed values of  $T_1$  and  $T_2$ . The average of all measured  $T_1$  and  $T_2$  values for residues in Nef with a heteronuclear  $^{15}\text{N}$ - $\{^1\text{H}\}$  NOE of  $\geq 0.7$  are 790 and 69 ms, respectively. For such residues of the Nef core, the predominant effect of internal motions on the relaxation rates  $1/T_1$  and  $1/T_2$  is a scaling by the factor  $S^2$  (Kay et al., 1989). Using standard equations for  $T_1$  and  $T_2$  (Kay et al., 1989), scaling by  $S^2 = 0.85$  (which is typical for well-ordered residues in globular proteins), a value of  $-160$  ppm for the  $^{15}\text{N}$  chemical shielding anisotropy, and a  $^1\text{H}$ - $^{15}\text{N}$  distance of  $1.02 \text{ \AA}$ , the simultaneous solution of Equation 1 for  $T_1$  and  $T_2$  yields  $\tau_c = 7.5$  ns and  $x = 0.54$ . This  $\tau_c$  value is consistent with the rotational correlation time expected for the 17.8 kDa monomeric NEF $^{\Delta 2-39, \Delta 159-173}$  mutant at  $35^\circ\text{C}$ . For a Nef concentration of 0.6 mM, the value of  $x$  corresponds to a dissociation constant for the Nef dimer of 0.7 mM. This agrees with the observation that the amide proton  $T_2$  values of NEF $^{\Delta 2-39, \Delta 159-173}$  at 0.1 mM are in the range expected for a 18 kDa monomer, whereas values at a 1 mM concentration are approximately twofold smaller. The value of the dimerization constant is close to the 0.6 mM concentration of Nef, which was used throughout the experiments. This means that  $\sim 50\%$  of the molecules are in the monomeric form, and 50% are dimeric. The apparent molecular weight of Nef under those conditions is, therefore,  $\sim 27$  kDa. At this stage, it remains unclear whether Nef exerts its biological function(s) as a monomer, or whether the tendency to form dimers or higher order aggregates is functionally significant.

## Materials and methods

### Sample preparation

Expression, labeling, and purification of the various Nef constructs, i.e., full-length Nef, deletion of residues 2–39 (NEF $^{\Delta 2-39}$ ), 2–52 (NEF $^{\Delta 2-52}$ ), 2–59 (NEF $^{\Delta 2-59}$ ), and 2–39 + 159–173 (NEF $^{\Delta 2-39, 159-173}$ ) was carried out as described previously (Grzesiek et al., 1996a). A summary of the sample preparations is provided in Table 1.

In order to identify slowly exchanging amide protons, a sample of  $^{15}\text{N}$ -labeled NEF $^{\Delta 2-39}$  was lyophilized at pH 8.0 and redissolved in  $\text{D}_2\text{O}$ . After a period of 19 h at  $6^\circ\text{C}$  and 1 h at  $35^\circ\text{C}$ , a 2D  $^1\text{H}$ - $^{15}\text{N}$  spectrum with water flip-back (Grzesiek & Bax, 1993b) was recorded. Amide resonances still present in this spectrum are marked by stars in Figure 7.

### NMR spectroscopy

All NMR spectra were recorded at  $35^\circ\text{C}$  on a Bruker AMX600 or DMX600 spectrometer, equipped with an 8 mm self-shielded  $z$ -gradient probe (Nalorac). Relevant parameters and literature references for the various experiments carried out on Nef are collected in Table 2.

All NMR data were processed with the software package NMRPipe (Delaglio et al., 1995). Manual peak picking of the processed spectra was achieved with the program PIPP (Garrett et al., 1991). Combination of the frequency information and analysis of the intensity information contained in the cross-peaks of the various spectra was performed using in-house routines.

### Structure calculation

Structures were calculated using a modified version (Garrett et al., 1994; Kuszewski et al., 1995) of the program X-PLOR (Brünger, 1992) and hybrid distance geometry/simulated annealing protocols (Nilges et al., 1988) as described previously (Grzesiek et al., 1996a).

### Supplementary material in Electronic Appendix

The electronic appendix contains three tables of chemical shift assignments for backbone and side chain  $^1\text{H}$ ,  $^{15}\text{N}$ , and  $^{13}\text{C}$  resonances for the constructs HIV-1 NEF $^{\Delta 2-39}$ , NEF $^{\Delta 2-39, \Delta 159-173}$ , and NEF $^{\Delta 2-39, \Delta 159-173}$  in complex with Hck SH3.

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