

# Regulation Mechanism of ERM (Ezrin/Radixin/Moesin) Protein/Plasma Membrane Association: Possible Involvement of Phosphatidylinositol Turnover and Rho-dependent Signaling Pathway

Motohiro Hirao,<sup>\*‡</sup> Naruki Sato,<sup>§</sup> Takahisa Kondo,<sup>\*||</sup> Shigenobu Yonemura,<sup>\*</sup> Morito Monden,<sup>‡</sup> Takuya Sasaki,<sup>¶</sup> Yoshimi Takai,<sup>¶</sup> Shoichiro Tsukita,<sup>\*</sup> and Sachiko Tsukita<sup>\*,\*\*</sup>

<sup>\*</sup>Department of Cell Biology, Faculty of Medicine, Kyoto University, Sakyo-ku, Kyoto 606, Japan; <sup>‡</sup>Department of Surgery II, Osaka University Medical School, Suita, Osaka 565, Japan; <sup>§</sup>Department of Biology, Faculty of Science, Chiba University, Chiba 260, Japan; <sup>||</sup>First Department of Internal Medicine, Nagoya University School of Medicine, Nagoya 466, Japan; <sup>¶</sup>Department of Molecular Biology and Biochemistry, Osaka University Medical School, Suita, Osaka 565, Japan; and <sup>\*\*</sup>College of Medical Technology, Kyoto University, Sakyo-ku, Kyoto 606, Japan

**Abstract.** The ERM proteins, ezrin, radixin, and moesin, are involved in the actin filament/plasma membrane interaction as cross-linkers. CD44 has been identified as one of the major membrane binding partners for ERM proteins. To examine the CD44/ERM protein interaction in vitro, we produced mouse ezrin, radixin, moesin, and the glutathione-S-transferase (GST)/CD44 cytoplasmic domain fusion protein (GST-CD44cyt) by means of recombinant baculovirus infection, and constructed an in vitro assay for the binding between ERM proteins and the cytoplasmic domain of CD44. In this system, ERM proteins bound to GST-CD44cyt with high affinity ( $K_d$  of moesin was  $9.3 \pm 1.6$  nM) at a low ionic strength, but with low affinity at a physiological ionic strength. However, in the presence of phosphoinositides (phosphatidylinositol [PI], phosphatidylinositol 4-monophosphate [4-PIP], and phosphatidylinositol 4,5-bisphosphate [4,5-PIP<sub>2</sub>]), ERM proteins bound with a relatively high affinity to GST-CD44cyt even at a physiological ionic strength: 4,5-PIP<sub>2</sub> showed a marked effect ( $K_d$  of moesin in the presence of 4,5-PIP<sub>2</sub> was  $9.3 \pm 4.8$  nM). Next, to examine the regulation mechanism of CD44/ERM interaction in vivo, we reexamined

the immunoprecipitated CD44/ERM complex from BHK cells and found that it contains Rho-GDP dissociation inhibitor (GDI), a regulator of Rho GTPase. We then evaluated the involvement of Rho in the regulation of the CD44/ERM complex formation. When recombinant ERM proteins were added and incubated with lysates of cultured BHK cells followed by centrifugation, a portion of the recombinant ERM proteins was recovered in the insoluble fraction. This binding was enhanced by GTP $\gamma$ S and markedly suppressed by C3 toxin, a specific inhibitor of Rho, indicating that the GTP form of Rho in the lysate is required for this binding. A mAb specific for the cytoplasmic domain of CD44 also markedly suppressed this binding, identifying most of the binding partners for exogenous ERM proteins in the insoluble fraction as CD44. Consistent with this binding analysis, in living BHK cells treated with C3 toxin, most insoluble ERM proteins moved to soluble compartments in the cytoplasm, leaving CD44 free from ERM. These findings indicate that Rho regulates the CD44/ERM complex formation in vivo and that the phosphatidylinositol turnover may be involved in this regulation mechanism.

ERM (Ezrin/Radixin/Moesin)<sup>1</sup> family proteins are thought to function as general cross-linkers between plasma membranes and actin filaments

Address all correspondence to Sachiko Tsukita, Ph.D., Department of Cell Biology, Faculty of Medicine, Kyoto University, Konoe-Yoshida, Sakyo-ku, Kyoto 606, Japan. Tel.: 81-75-753-4373. Fax: 81-75-753-4660. E-mail: atsukita@mfour.med.kyoto-u.ac.jp

1. *Abbreviations used in this paper:* C-moesin and N-moesin, carboxyl-terminal half and amino-terminal half of recombinant moesin; ERM, ezrin/

(Bretscher, 1983; Pakkanen et al., 1987; Lankes et al., 1988; Sa. Tsukita et al., 1989; Algrain et al., 1993; Arpin et al., 1994). The ERM family consists of three closely related proteins, ezrin, radixin, and moesin (Sato et al., 1992;

radixin/moesin; GDI, GDP dissociation inhibitor; GDS, GDP dissociation stimulator; GST, glutathione-S-transferase; PC, phosphatidylcholine; PI, phosphatidylinositol; 4-PIP, phosphatidylinositol 4-monophosphate; PIP<sub>2</sub>, phosphatidylinositol 4,5-bisphosphate.

Tsukita et al., 1992). In cultured epithelial and fibroblastic cells, they are expressed and localized at specialized regions where actin filaments are densely associated with plasma membranes, such as cleavage furrows, microvilli, ruffling membranes, and cell-cell/cell-matrix adhesion sites (especially at primordial forms of cadherin- and integrin-based cell adhesion sites), whereas among organs their expression varies, suggesting some dedifferentiation in terms of ERM expression in cultured cells (Bretscher, 1983; Pakkanen et al., 1987; Lankes et al., 1988; Sa. Tsukita et al., 1989; Sato et al., 1991, 1992; Sh. Tsukita et al., 1992; Berryman et al., 1993; Franck et al., 1993; Amieva et al., 1994; Takeuchi et al., 1994b; Henry et al., 1995). The suppression of ERM expression with antisense oligonucleotides destroys microvilli, cell-cell, and cell-matrix adhesion sites (Takeuchi et al., 1994b), and the introduction of a dominant-negative construct of ezrin and radixin impairs the cortical actin organization and cytokinesis, respectively (Henry et al., 1995; Martin et al., 1995).

Sequencing has revealed that the three ERM proteins are highly homologous; in the mouse, the identity is 75, 72, and 80% for ezrin/radixin, ezrin/moesin, and radixin/moesin, respectively (Gould et al., 1989; Turunen et al., 1989; Funayama et al., 1991; Lankes and Furthmayr, 1991; Sato et al., 1992). The sequences of their amino-terminal halves are highly conserved (~85% identity for any pair). A tumor suppressor molecule responsible for neurofibromatosis type 2, named merlin or schwannomin, was recently identified, which has significant sequence similarity to ERM proteins: ~49% identity overall and, especially, ~85% identity in their amino-terminal halves (Rouleau et al., 1993; Trofatter et al., 1993).

The highly conserved amino-terminal sequence in ERM proteins is also found in the amino-terminal end of some membrane-associated proteins such as band 4.1 protein, talin, PTPH1, and PTPMEG, indicating a band 4.1 superfamily (Conboy et al., 1986; Rees et al., 1990; Gu et al., 1991; Yang and Tonks, 1991; Arpin et al., 1994; Takeuchi et al., 1994a). Considering that this conserved domain in band 4.1 protein is reportedly responsible for its direct association with glycophorin C, an integral membrane glycoprotein of erythrocyte membranes (Bennett, 1989), the amino-terminal half domain of ERM proteins was thought to directly bind to some integral membrane proteins. Immunoprecipitation experiments using cultured cells subsequently revealed that CD44 is directly associated with ERM proteins (Tsukita et al., 1994). CD44 is a polymorphic cell-surface glycoprotein which is found on a wide variety of cells (Haynes et al., 1989, 1991; Lesley et al., 1993) that is also called Pgp-1 (Zhou et al., 1989), HCAM (Goldstein et al., 1989), Hermes antigen (Jalkanen et al., 1986), and ECMRIII (Wayner et al., 1988). In cultured fibroblasts, CD44 is precisely colocalized with ERM proteins, favoring the notion that CD44 is a membrane binding partner of ERM proteins (Tsukita et al., 1994). However, in the body, the expression of CD44 varies among tissues and its distribution is not necessarily identical to that of ERM proteins (Berryman et al., 1993; Nakamura et al., 1996). Furthermore, in some types of cells, integral membrane proteins such as CD43 and the H<sup>+</sup>/K<sup>+</sup> ATPase pump are precisely colocalized with ERM proteins (Hanzel et al., 1991; Yonemura et al., 1993). These findings

suggest that ERM proteins bind to a specific group of integral membrane proteins, including CD44. Considering that CD44 is rather highly and ubiquitously expressed, however, it could be a major binding partner of ERM proteins.

The carboxyl-terminal half of ERM proteins, especially the carboxy-terminal 34 amino acids, interacts with actin filaments (Turunen et al., 1994; Pestonjamas et al., 1995). The coexistence of plasma membrane- and actin filament-binding domains in individual molecules allows ERM proteins to function as a plasma membrane/actin filament cross-linker. Considering that the plasma membrane/actin filament association is dynamically regulated, the next question is how the membrane binding of ERM proteins is regulated *in vivo*.

In this study, we first analyzed the manner of binding between ERM proteins and the cytoplasmic domain of CD44 *in vitro* using recombinant ERM and a glutathione-S-transferase (GST)/CD44 cytoplasmic domain fusion protein produced by means of recombinant baculovirus infection. We found that the CD44 cytoplasmic domain showed very weak binding activity to any of the ERM proteins at physiological ionic strength, but that in the presence of phosphatidylinositol 4,5-bisphosphate (4,5-PIP<sub>2</sub>), it bound with high affinity to ERM proteins (the *K<sub>d</sub>* of moesin was 9.3 ± 4.8 nM). These findings are not only the first to show that ERM proteins can bind to the cytoplasmic domain of CD44 *in vitro*, but also suggests that the formation of CD44/ERM complex is dynamically regulated *in vivo*. Next, we reexamined the CD44/moesin immune complex from BHK cells and found that Rho-GDP dissociation inhibitor (GDI), a regulator of a small GTP-binding protein Rho, was tightly associated with the CD44/moesin complex.

Rho reportedly regulates the actin filament-plasma membrane association (Hall, 1990; Ridley and Hall, 1992; Ridley et al., 1992; Takai et al., 1995; Zigmond, 1995). The Rho family belongs to the small GTP-binding protein superfamily and consists of the Rho, Rac, and Cdc42 subfamilies. Interconversion between active and inactive forms (GTP-Rho and GDP-Rho, respectively) is regulated by guanine nucleotide exchange factors (GEFs) and GTPase-activating proteins (GAPs). Rho-GDI, Smg-GDP dissociation stimulator (GDS), Dbl, and Ost have been identified as GEFs. Therefore, the occurrence of Rho-GDI in the CD44/ERM complex suggests that this complex formation is regulated by Rho. We then established a cell-free system with which to measure the binding of the exogenous recombinant ERM proteins to the insoluble fraction of the lysate of BHK cells, and we found that the binding of ERM proteins to the cytoplasmic domain of CD44 was enhanced by GTPγS and remarkably suppressed by C3 toxin, a specific inhibitor of Rho GTPase (Aktories et al., 1988; Kikuchi et al., 1988; Narumiya et al., 1988; Braun et al., 1989). In good agreement with these binding results, when living BHK cells were incubated with C3 toxin, most insoluble ERM proteins moved to the soluble compartments in the cytoplasm, leaving free CD44. We thus concluded that Rho regulates the formation of CD44/ERM complex *in vivo* and that the elevation of 4,5-PIP<sub>2</sub> concentration in plasma membranes is possibly involved in this regulation pathway.

## Materials and Methods

### Cells and Antibodies

Sf9 and High Five cells were purchased from InVitrogen (San Diego, CA) and cultured in TC-100 medium (GIBCO BRL, Gaithersburg, MD) supplemented with tryptose phosphate broth (GIBCO BRL) and 10% FCS at 27°C. BHK cells were cultured in DME supplemented with 5% newborn calf serum and 10% tryptose phosphate broth. The mAbs, M11, R2-1, and M22, were raised in rats against recombinant mouse ezrin, radixin, and moesin, respectively (Takeuchi et al., 1994b). The pAbs, I1 and p-800, were raised in rabbits against purified rat radixin and recombinant mouse radixin, respectively (Sa. Tsukita et al., 1989; Funayama et al., 1991). The pAb I1 was used to detect all ERM proteins by immunoblotting (Sa. Tsukita et al., 1989; Sato et al., 1992). The mouse mAb CR-22 reacts specifically with moesin in immunoprecipitation (Sato et al., 1992; Tsukita et al., 1994). Anti-mouse rho-GDI mAb has been characterized (Shimizu et al., 1991). The rat mAb30189 is specific for hamster CD44 (Tsukita et al., 1994).

Monoclonal antibodies were produced in the rat using the GST fusion protein with the cytoplasmic domain of mouse CD44 (GST-CD44cyt) as described (Sh. Tsukita et al., 1989). The culture supernatant of each hybridoma was screened to select mAbs that recognize not only mouse CD44 (GST-CD44cyt and in mouse MTD-1A cells) but also hamster CD44 (in BHK cells) by immunoblotting.

### Production and Purification of Recombinant Ezrin, Radixin, and Moesin

Mouse ezrin, radixin, or moesin full-length cDNA was integrated into the baculovirus genome, and the recombinant virus containing ezrin, radixin, or moesin cDNA was isolated and condensed using a MAXBAC™ kit (InVitrogen). Insect Sf9 cells were infected with recombinant viruses, cultured in 200 ml spinner flasks at 27°C for 72 h, and then washed with PBS. After being suspended in PBS ( $3 \times 10^6$  cells/ml) containing 2 mM EGTA, 2 mM PMSF and 4 µg/ml of leupeptin, the cells were homogenized in a tight-fitting Dounce homogenizer and sonicated, followed by centrifugation at 100,000 g for 1 h at 4°C.

The supernatant was applied to a heparin column (5 ml in  $15 \times 25$  mm; Hi TrapHeparin; Pharmacia LKB Biotechnology AB, Uppsala, Sweden) equilibrated with DG buffer (10 mM Hepes, pH 7.5, 1 mM EGTA, 0.1 mM DTT). After thoroughly washing with DG containing 150 mM KCl, proteins were eluted with a linear concentration gradient of KCl (150–750 mM). Recombinant ERM proteins were eluted around 350 mM KCl. The eluate was dialyzed against DG followed by centrifugation at 100,000 g for 1 h. The supernatant was loaded onto a Q-Sepharose column (1 ml in  $8 \times 38$  mm; Econo-PacQ; Bio-Rad Laboratories, Hercules, CA) equilibrated with DG. The column was washed with DG and then proteins were eluted with a linear concentration gradient of KCl (0–200 mM). Purified ERM proteins were eluted around 50 mM KCl. The purity of each protein was determined by SDS-PAGE (see Fig. 1).

### Production and Purification of GST-Fusion Protein with the Cytoplasmic Domain of CD44

The cDNA encoding the full-length of the cytoplasmic domain of mouse CD44 (1180–1395 bp; CD44cyt) with BamHI and EcoRI sites at the ends was produced by PCR using mouse lung cDNA library as the template, and subcloned into pBluescript SK(–). The insert was then subcloned into the pGEX2T expression vector, which was designed to express a GST fusion protein in *Escherichia coli*. Using this GST-CD44cyt expression vector as a template, the GST-CD44cyt cDNA with NheI and BglII sites at the ends was produced by PCR and then subcloned into pBluescript SK(–). The insert was integrated into the baculovirus genome, and the recombinant virus containing GST-CD44cyt cDNA was isolated and condensed using a MAXBAC™ kit (InVitrogen). The correct sequence of the PCR products was confirmed by sequencing with a Taq terminator cycle sequencing kit (DyeDeoxy™; Applied Biosystems, Inc., Foster City, CA).

Insect Sf9 cells were infected, cultured, washed, homogenized, and centrifuged as described above. The supernatant was incubated with glutathione-Sepharose 4B beads (Pharmacia LKB Biotechnology AB), which had been washed with PBS for 30 min at room temperature. The glutathione-Sepharose beads bound to GST-CD44cyt were washed with W buffer (10 mM Hepes, pH 7.5, 2 mM PMSF, 4 µg/ml leupeptin) and used in the in vitro binding assay. In some experiments, GST-CD44cyt was purified by elution with H buffer containing 30 mM glutathione, dia-

lyzed against H buffer, and then used for the in vitro binding assay. The control experiments included GST expressed in *E. coli* using pGEX2T and purified using glutathione-Sepharose 4B beads.

### Production and Purification of the Amino- and Carboxyl-terminal Halves of Moesin

The cDNAs encoding the amino-terminal (1–930 bp) and carboxyl-terminal halves (931–1733 bp) of mouse moesin were produced by PCR using the moesin cDNA clone p22T6 (Sato et al., 1992) as the template and subcloned into pBluescript SK(–). These inserts were integrated into the baculovirus genome, and the recombinant virus containing the cDNA encoding the amino- or carboxyl-terminal half of moesin was isolated and condensed using a MAXBAC™ kit (InVitrogen). The sequences of the PCR products were confirmed using a Taq terminator cycle sequencing kit (DyeDeoxy™; Applied Biosystems Inc.).

To obtain a large amount of the amino-terminal half of recombinant moesin (N-moesin), we used insect High Five (HF) cells instead of Sf9 cells. HF cells were infected with recombinant virus carrying N-moesin cDNA, cultured on 19-cm dishes at 27°C for 48 h, and then washed with PBS. After being suspended in low salt buffer (2 mM Tris-HCl, pH 10, 1 mM EGTA, 0.5 mM PMSF), the cells were sonicated and centrifuged at 100,000 g for 10 min at 4°C. The pellet was resuspended and sonicated in TE (10 mM Tris-HCl, pH 8.0, 0.1 mM EDTA) containing 1% Triton X-100 followed by a 60 min incubation at 4°C. The sonicate was centrifuged at 100,000 g for 1 h, and the supernatant was dialyzed against DG. The dialysate was applied onto a heparin column (5 ml in  $15 \times 25$  mm; HiTrap Heparin) that had been equilibrated with DG. After thoroughly washing with DG, proteins were eluted with a linear concentration gradient of KCl (0–800 mM). Recombinant N-moesin was eluted at around 600 mM KCl.

To produce the carboxyl-terminal half of moesin (C-moesin), Sf9 cells were infected with recombinant viruses containing C-moesin cDNA. C-moesin was purified in the same manner as the full-length moesin.

### In Vitro Binding Assay between ERM Proteins and CD44 Using GST-CD44cyt Beads

After the glutathione-Sepharose beads containing GST-CD44cyt fusion protein or GST were pretreated with RIPA buffer (0.1% SDS, 0.5% deoxycholic acid, 1% Nonidet P-40, 50 mM Tris-HCl, pH 8.0, 150 mM NaCl) for 20 min at room temperature, they were washed with 20 vol of H buffer (10 mM Hepes, pH 7.5, 1 mM MgCl<sub>2</sub>). After a brief centrifugation, 30 µl of the glutathione-conjugated Sepharose bead slurry was suspended in 200 µl of H buffer containing 40 mM or 150 mM KCl. In some experiments, 150 mM KCl H buffer contained 50 µg/ml phosphatidylserine (PS), phosphatidylcholine (PC), phosphatidylinositol (PI), phosphatidylinositol 4-phosphate (4-PIP), or 4,5-PIP<sub>2</sub> (Sigma Chemical Co., St. Louis, MO). Each phospholipid was dissolved in distilled water to a final concentration of 1 mg/ml and sonicated three times each for 10 s.

Bead suspension (230 µl) was incubated with 4 µg of purified ezrin, radixin, moesin, N-moesin, or C-moesin in 10–20 µl of 40 or 150 mM KCl H buffer for 30 min at room temperature. The beads were washed three times with H buffer by a brief centrifugation, and bound proteins were eluted from the beads with H buffer containing 30 mM glutathione. The amount of GST-CD44cyt or GST in each eluate was determined by SDS-PAGE. An appropriate amount of each eluate was again applied to SDS-PAGE to contain the same amount of GST-CD44cyt or GST, and the amount of ezrin, radixin, moesin, N-moesin, or C-moesin in each eluate was evaluated by immunoblotting with specific mAbs.

### Estimation of the Dissociation Constant between Moesin and CD44cyt

To estimate the dissociation constant between CD44cyt and moesin, 30 µl of the glutathione-Sepharose bead slurry containing 40 µg of GST-CD44cyt was incubated in 240 µl of 40 or 150 mM KCl H buffer containing 0–100 pmol (0–13.3 µg) of purified moesin in the presence or absence of 4,5-PIP<sub>2</sub>. After the beads were washed three times with H buffer, bound moesin was eluted with H buffer containing 30 mM glutathione. The amount of moesin in each eluate was estimated by comparing the silver staining intensity of moesin band with that of various amounts of authentic purified moesin resolved and silver stained in the same gel. The staining intensity was measured by a densitometer using Adobe Photoshop TM 3.0J histogram and a Scatchard plot of the data was generated.

Experiments were repeated three times for each estimation of dissociation constant.

### ***In Vitro Binding between ERM Proteins and CD44 Using Ezrin/Radixin-coupled Beads***

Purified ezrin (10  $\mu$ g) was coupled to 30  $\mu$ l of Affigel-10 (Bio-Rad Laboratories). These ezrin-coupled beads were incubated with 1% BSA at 4°C for 2 h followed by 1 mg/ml GST in H buffer containing 150 mM KCl at 4°C overnight and then suspended in 200  $\mu$ l of H buffer containing 150 mM KCl in the presence or absence of 50  $\mu$ g/ml PS, PC, PI, 4-PIP<sub>2</sub> or 4,5-PIP<sub>2</sub>. They were then incubated with 5  $\mu$ g purified GST-CD44cyt in 40  $\mu$ l of 150 mM KCl H buffer for 30 min at room temperature. In the control experiment, the GST-fusion protein with the cytoplasmic domain of E-cadherin was used instead of the GST-CD44cyt. After washing three times with the incubation solution, bound proteins were eluted from beads with the sample buffer for SDS-PAGE. Each eluate was resolved by electrophoresis and stained with Coomassie brilliant blue.

In some experiments, 20  $\mu$ g of ezrin or radixin was coupled to 50  $\mu$ l of CNBr-activated Sepharose (Pharmacia LKB Biotechnology AB). These beads were blocked only by 1% BSA at 4°C for 2 h, and 50  $\mu$ l of beads were suspended in 200  $\mu$ l of 150 mM KCl H buffer in the presence or absence of 50  $\mu$ g/ml 4,5-PIP<sub>2</sub>. They were then incubated with GST-CD44cyt, and the amount of bound GST-CD44cyt was determined according to the same procedure as that used with ERM-coupled Affigel beads, except that purified GST was used as a control and that bound GST-CD44cyt was detected by immunoblotting with anti-GST pAb.

### ***Gel Filtration Chromatography***

Gel filtration was performed on Superose 12R 10/30 (24 ml; 1  $\times$  30 cm; equilibrated with DG) at 4°C. Each ERM protein or GST-CD44cyt (6  $\mu$ g) was mixed with no phospholipid, PS, PC, PI, 4-PIP, or 4,5-PIP<sub>2</sub> (20  $\mu$ g) in  $\sim$ 200  $\mu$ l of 10 mM Hepes, pH 7.5, containing 1 mM MgCl<sub>2</sub> and 150 mM KCl and then passed through the column. Eluted proteins were detected by ELISA using anti-ERM protein pAb, I1, or anti-GST pAb.

### ***Immunoprecipitation***

BHK cells were cultured as monolayers in 100-mm-diam plastic dishes to 80–90% confluence. After washing once with methionine-free medium supplemented with 2% fetal calf serum, the monolayers were incubated with 3 ml of the same medium containing 0.2 mCi [<sup>35</sup>S]methionine (Amersham International plc., Buckinghamshire, U.K.) for 3 h and washed three times with PBS. The labeled cells on one dish were lysed and incubated in 0.4–0.8 ml of RIPA buffer (0.1% SDS, 0.5% deoxycholate, 1% Nonidet P-40, 150 mM NaCl, 50 mM Tris, pH 8.0, 1 mM *p*-amidinoPMSF, and 10  $\mu$ g/ml leupeptin) for 5 min. The RIPA lysate was removed from the dish after fully dislodging any remaining cellular debris with a rubber policeman. After centrifugation at 12,000 *g* for 15 min, the RIPA-soluble supernatant was immunoprecipitated with 15  $\mu$ l of protein G-Sepharose 4B (Zymed Laboratories, Inc., South San Francisco, CA) conjugated with anti-moesin mAb CR-22 or control mouse IgG. In some experiments, to chase the labeled moesin, 100  $\mu$ l of 0.05 mg/ml purified nonlabeled recombinant moesin in 50 mM KCl/10 mM Hepes, pH 7.5, was added to 300  $\mu$ l of the RIPA lysate of [<sup>35</sup>S]methionine-labeled BHK cells. Sepharose 4B-bound immune complexes were washed five times with RIPA buffer. Immune complexes were then eluted by boiling in sample buffer that did not contain 2-mercaptoethanol and resolved by SDS-PAGE. The [<sup>35</sup>S]methionine signal was quantified by means of a Fujix Bioimage Analyzer Bas 2000 System (Fuji Film Co., Tokyo, Japan).

For reimunoprecipitation, the immune complexes were eluted from 15  $\mu$ l of Sepharose 4B into 1.5-ml test tubes with 50  $\mu$ l of a high salt solution consisting of 0.98 M KCl, 0.02 M NaCl, 1 mM MgCl<sub>2</sub>, 10 mM MOPS, pH 7.4. The eluates from five tubes were combined and diluted with 6 vol of distilled water and then reimunoprecipitated with 20  $\mu$ l of protein G-Sepharose 4B conjugated with mouse anti- $\rho$ -GDI mAb or control mouse IgG. After five washes with the high salt solution diluted with 6 vol of distilled water, immune complexes were eluted and analyzed as described above.

### ***In Vitro Translation of Mouse Rho-GDI***

Translation in vitro proceeded as reported (Pelham and Jackson, 1976). RNA was synthesized from the full-length cDNA of rho-GDI that was

subcloned into the KpnI-digested Bluescript SK(–) vector using an mCAP<sup>TM</sup> mRNA Capping Kit (Stratagene, La Jolla, CA). Capped RNA was translated in mRNA-dependent rabbit reticulocyte lysate in the presence of [<sup>35</sup>S]methionine using an *In Vitro* Express<sup>TM</sup> Translation kit (Stratagene). The [<sup>35</sup>S]methionine (1,300 Ci/mmol, Amersham International plc.) radiochemical concentration was 2.6 mCi/ml in a final reaction volume of 25  $\mu$ l. The reaction product in the in vitro translation reaction was then resolved by SDS-PAGE. The [<sup>35</sup>S]methionine signal was quantified by means of a Fujix Bioimage Analyzer Bas 2000 System (Fuji Film Co.).

### ***Binding of Purified Recombinant ERM Proteins to the Insoluble Fraction of BHK Cell Lysate***

BHK cell lysate was prepared from a monolayer in a 10-cm dish at 60–70% confluence at 4°C. Cells were washed three times with PBS, scraped off using a rubber policeman in lysis buffer (0.06% Nonidet P-40, 0.6% glycerol, 130 mM KCl, 20 mM NaCl, 1 mM sodium orthovanadate, 2 mM MgCl<sub>2</sub>, 0.1 mM EGTA, 1 mM EDTA, 1 mM DTT, 10 mM Hepes, pH 7.4, 1 mM PMSF, and 10  $\mu$ g/ml leupeptin), and then disrupted by three strokes in a loose-fitting teflon glass homogenizer. The protein concentration of each lysate was adjusted to 12 mg/ml using the lysis buffer. The protein concentration was estimated from the absorbance at 280 nm using BSA as the standard, after 5  $\mu$ l of the cell lysate was completely solubilized by 95  $\mu$ l of 2 N NaOH. The BHK cell lysate (5  $\mu$ l) was brought to 10  $\mu$ l with 1/15 PBS (1 mM sodium phosphate solution, pH 7.5, and 10 mM NaCl). To examine the effect of GTP $\gamma$ S, 2  $\mu$ l of 10 mM GTP $\gamma$ S, pH 7.5, and 3  $\mu$ l of 1/15 PBS were added to the BHK cell lysate (5  $\mu$ l), and to examine the effect of C3, 2  $\mu$ l of 10 mM GTP $\gamma$ S, pH 7.5, 2  $\mu$ l of 0.75 mg/ml of C3 toxin, and 1  $\mu$ l of 10 mM NAD<sup>+</sup> were added to the cell lysate. Thereafter, 5  $\mu$ l of purified ezrin, radixin, or moesin ( $\sim$ 0.16 mg/ml eluate from the monoQ column in  $\sim$ 50 mM KCl/10 mM Hepes, pH 7.5) was added and incubated for 8 h at 4°C, followed by centrifugation at 300,000 *g* for 10 min in a 7  $\times$  20 mm tube (Optima TL Ultracentrifuge; Beckman Instrs., Fullerton, CA). The insoluble fraction recovered as a pellet was washed with the lysis buffer without Nonidet P-40/glycerol and resolved by SDS-PAGE. The amount of ERM proteins in the insoluble fraction was estimated by comparing the staining intensity of each band in immunoblotting with that of the various amounts of authentic ERM proteins resolved by electrophoresis and immunoblotted on the same nitrocellulose sheet.

### ***Exposing Living BHK Cells with C3 Toxin***

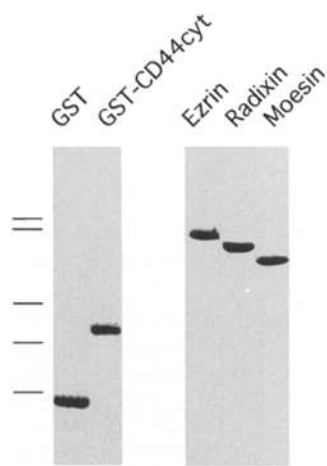
BHK cells cultured to 80–90% confluence were incubated with 100  $\mu$ g/ml C3 toxin in 1 ml of Opti-MEM medium (GIBCO BRL) in the presence of 10  $\mu$ l of lipofectamin (GIBCO BRL) for 4 h. Control cells were incubated with the same culture medium without C3 toxin for 6 h. To compare the amount of soluble and insoluble ERM proteins, cells exposed or not to C3 toxin were homogenized in a physiological solution consisting of 130 mM KCl, 20 mM NaCl, 1 mM MgCl<sub>2</sub>, 10 mM Hepes, pH 7.5, 1 mM *p*-amidinoPMSF, and 0.01 mg/ml of leupeptin. After centrifugation at 300,000 *g* for 20 min, the amount of ERM proteins in the pellet and supernatant was quantified by immunoblotting using anti-ERM pAb I1. The cells exposed or not to C3 toxin in 60-mm dishes were immunoprecipitated with anti-CD44 mAb (mAb30189) as described above, and the immunoprecipitates were separated by SDS-PAGE followed by immunoblotting with anti-ERM pAb I1.

### ***SDS-PAGE and Immunoblotting***

One-dimensional SDS-PAGE (7.5–18%) proceeded essentially according to Laemmli (1970), and the gels were stained with Coomassie brilliant blue R-250. For immunoblotting, proteins were electrophoretically transferred from gels to nitrocellulose membranes, which were incubated with the first antibody. Bound antibodies were visualized with alkaline phosphatase-conjugated goat anti-rabbit IgG and the appropriate substrates as described by the manufacturer (Amersham International).

### ***One-dimensional Peptide Mapping***

Protein bands were excised from the SDS-polyacrylamide gels, immersed in the sample buffer, and then subjected to limited proteolysis with *Staphylococcus aureus* V8 protease in 18% SDS-polyacrylamide gels as described by Cleveland et al. (1977). The [<sup>35</sup>S]methionine signal was quantified by means of a Fujix Bioimage Analyzer Bas 2000 System (Fuji Film Co.).



**Figure 1.** Recombinant proteins used in the *in vitro* binding assay. Fusion proteins between GST and the cytoplasmic domain of mouse CD44 (GST-CD44cyt), mouse ezrin, radixin, and moesin produced by means of recombinant baculovirus infection were purified to homogeneity. GST was produced in *E. coli*. The mobility of the molecular markers is shown on the left (110, 88, 48, 33, and 29-kD, from the top), and each band was stained with Coomassie brilliant blue.

## Results

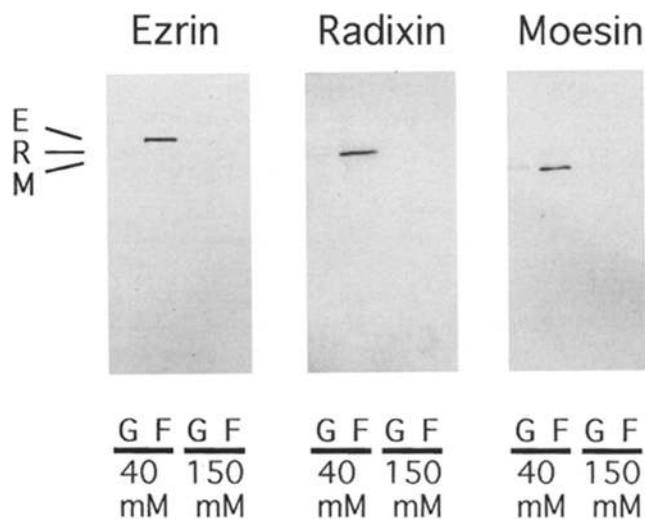
### *In Vitro* Binding Assay between GST-CD44cyt and ERM Proteins

To obtain a large amount of purified native ERM proteins for the *in vitro* binding assay, we overexpressed mouse ezrin, radixin, or moesin in insect Sf9 cells by means of recombinant baculovirus infection. The overexpressed ERM proteins were then purified to homogeneity through two chromatographic steps and used for the *in vitro* binding assay (Fig. 1). A fusion protein between GST and the cytoplasmic domain of mouse CD44 (GST-CD44cyt) was first expressed in *E. coli* and purified. However, this fusion protein was degraded into smaller fragments very rapidly in *E. coli*, making it very difficult to purify the full-length GST-CD44cyt required for the *in vitro* binding assay. We thus overexpressed GST-CD44cyt in Sf9 cells by recombinant baculovirus infection and found that it was stable in Sf9 cells. This product was used for the *in vitro* binding assay (Fig. 1).

GST-CD44cyt produced in the Sf9 system was incubated with glutathione-Sepharose beads. After washing, purified recombinant ezrin, radixin, or moesin was added to the beads. After incubation and washing, the proteins associated with GST-CD44cyt were eluted with a buffer containing glutathione, resolved by SDS-PAGE, and then immunoblotted with anti-ezrin, -radixin, or -moesin mAb. The control was GST produced in *E. coli* instead of GST-CD44cyt. All binding assays were performed at low and at physiological ionic strength (40 and 150 mM KCl, respectively). In some experiments, we used GST fusion proteins with the cytoplasmic domain of mouse E-cadherin as additional controls.

As shown in Fig. 2, at low ionic strength, ezrin, radixin, and moesin bound to the cytoplasmic domain of CD44. By contrast, at physiological ionic strength, they bound very weakly to GST-CD44cyt. Control experiments with GST as well as with GST/cadherin showed that the nonspecific background binding was negligible.

Using this assay, the interaction of moesin with CD44cyt was quantified by determining the relationship between the amount of "bound" moesin on beads and that of moesin in the incubation solution. The ratio of bound to total moesin over a range of 0–100 pmol was estimated by



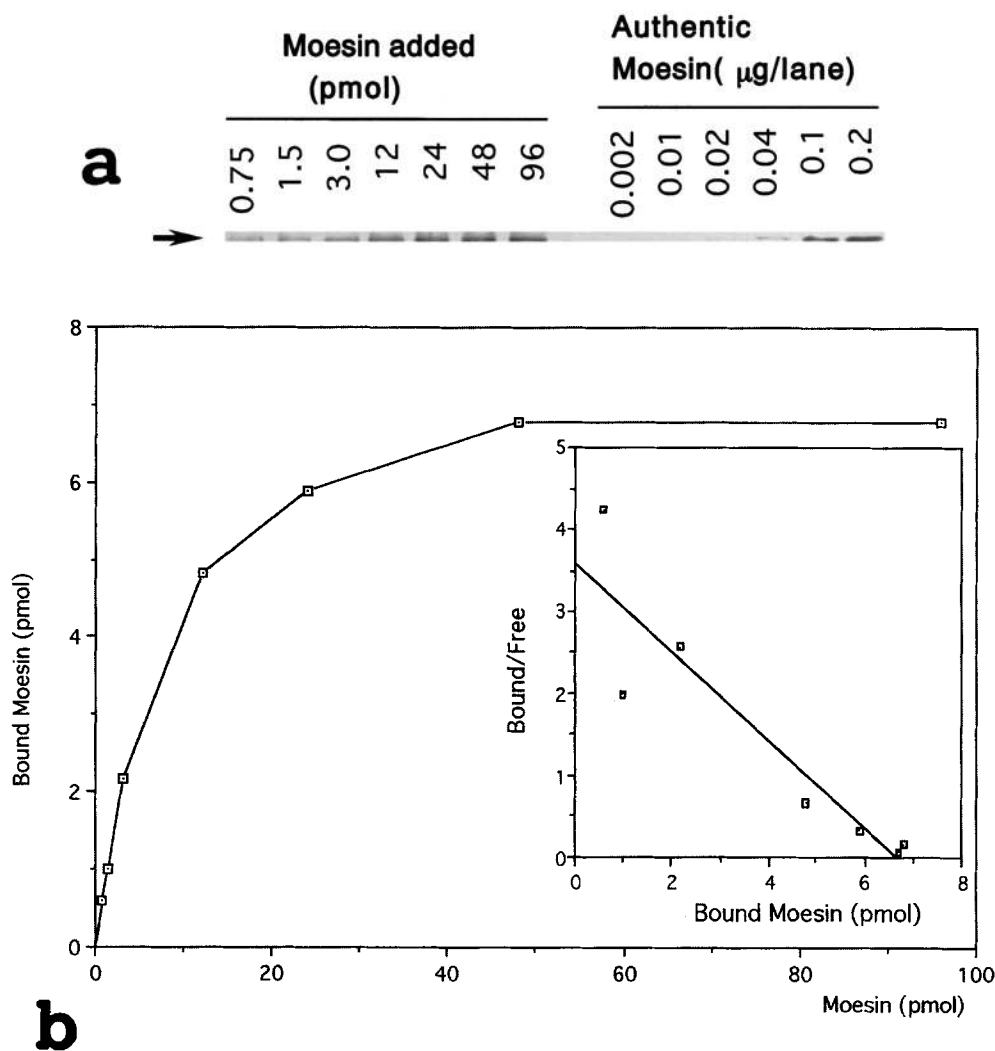
**Figure 2.** Association of ezrin, radixin, and moesin with GST-CD44cyt-bound beads. GST-CD44cyt or GST bound to glutathione-Sepharose beads was incubated with purified recombinant ezrin, radixin, or moesin at low (40 mM KCl) or physiological ionic strength (150 mM KCl). After washing, the protein associated with GST-CD44cyt (F) or GST (G) was eluted from the beads with a buffer containing glutathione. Proteins in the glutathione eluate were separated by SDS-PAGE followed by immunoblotting with anti-ezrin, anti-radixin, or anti-moesin mAbs. The bands of ezrin, radixin, and moesin are indicated by E, R, and M on the left.

subtracting the amount on control GST beads from that on GST-CD44cyt beads. As shown in Fig. 3, at low ionic strength, moesin binding to CD44cyt was saturable. Scatchard analysis determined the dissociation constant to be  $9.3 \pm 1.6$  nM (Fig. 3 *b*, *Inset*). The CD44cyt/moesin interaction at physiological ionic strength was too weak for quantitation.

### Effects of Phospholipids on the CD44cyt/ERM Binding

We investigated the regulation mechanism for the CD44/ERM interaction by means of this binding assay. Considering that many of the cellular events occurring just beneath the plasma membranes are regulated by phospholipids, we studied the effects of various phospholipids on the CD44/ERM interaction *in vitro*. As shown in Fig. 4, neither PS nor PC affected CD44/radixin or CD44/moesin interaction, whereas phosphoinositides significantly enhanced it at physiological ionic strength. PI, 4-PIP, and 4,5-PIP<sub>2</sub> apparently enhanced the binding of radixin or moesin to GST-CD44cyt, although there was a slight background binding to control GST. The effect of this enhancement of these three phosphoinositides was in the order of PI < 4-PIP < 4,5-PIP<sub>2</sub>, as shown in Fig. 4. The dissociation constant between CD44cyt and moesin at physiological ionic strength in the presence of 4,5-PIP<sub>2</sub> was determined to be  $9.3 \pm 4.8$  nM, according to the method described above.

The CD44/ezrin interaction also appeared to be enhanced by phosphoinositides. However, for unknown reasons, ezrin showed a relatively high affinity for Sepharose beads in the presence of phosphoinositides, making it difficult to conclusively evaluate the phosphoinositide depen-



**Figure 3.** Quantitative analysis of the binding between moesin and GST-CD44cyt at 40 mM KCl. (a) Estimation of the amount of “bound” moesin. Recombinant moesin (0.75–96 pmol) was added to the GST-CD44cyt or GST beads, and 15% of each glutathione eluate was applied to each lane of SDS-PAGE. The silver staining intensity of moesin band in glutathione eluates (*left*) was compared with that of the various amounts of authentic purified moesin resolved and silver stained in the same gel (*right*). (b) Binding mode of moesin to GST-CD44cyt beads. After subtraction of the GST-dependent background binding, the amount of bound versus added moesin (0–100 pmol) was plotted. Scatchard analysis (*Inset*) determined that the  $K_d$  was  $9.3 \pm 1.6$  nM.

dence of the CD44/ezrin interaction. To overcome this technical difficulty, we directly conjugated recombinant ezrin (and also radixin as a control) with CNBr-activated Sepharose beads and analyzed the interaction between those and soluble GST-CD44cyt in the absence or presence of 4,5-PIP<sub>2</sub> at physiological ionic strength. Fig. 5 *a* shows that in the absence of 4,5-PIP<sub>2</sub>, only a small amount of GST-CD44cyt was specifically bound to the ezrin beads, whereas in the presence of 4,5-PIP<sub>2</sub>, this binding was remarkably enhanced, an effect that was also observed between radixin beads and GST-CD44cyt. Using the ezrin-coupled Affigel beads, we further confirmed the similar phospholipid dependence of CD44/ezrin interaction to that of CD44/radixin and CD44/moesin, which was shown using GST-CD44cyt beads (Fig. 5 *b*). Taken all together, we conclude that the direct interaction of ezrin, radixin, and moesin with the cytoplasmic domain of CD44 is remarkably enhanced by phosphoinositides, especially 4,5-PIP<sub>2</sub>.

#### **Binding Affinity of ERM Proteins for Phosphoinositides**

The enhancement of the CD44/ERM interaction by phosphoinositides suggested that either ERM proteins or the cytoplasmic domain of CD44, or both, have binding affin-

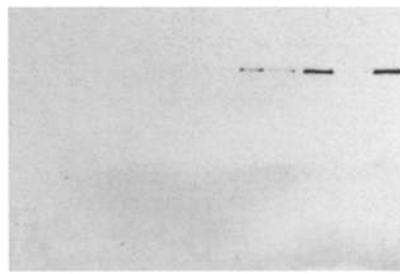
ity for phosphoinositides. We evaluated the binding affinity of ERM proteins and GST-CD44cyt for various phospholipids using gel filtration chromatography (Fig. 6).

Phospholipids were eluted in the void volume from a Superose 12R 10/30 column, due to micelle formation. ERM proteins were eluted at a position corresponding to an apparent molecular mass of ~60 kD. As shown in Fig. 6 *a*, purified recombinant moesin was incubated with various phospholipids and passed through the column. An incubation with PS, PC, or PI did not affect the behavior of moesin in the column. In sharp contrast, when moesin was incubated with 4-PIP or 4,5-PIP<sub>2</sub>, it coeluted with 4-PIP or 4,5-PIP<sub>2</sub> in the void volume. The same results were obtained using purified recombinant ezrin and radixin. A similar binding assay by means of gel filtration was performed between various lipids and GST-CD44cyt. None of PS, PC, PI, 4-PIP, or 4,5-PIP<sub>2</sub> affected the behavior of GST-CD44cyt in the column (Fig. 6 *b*).

#### **Phosphoinositide-independent Binding of the Amino-terminal Half of Moesin to the Cytoplasmic Domain of CD44**

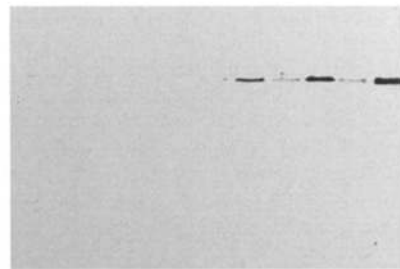
We produced amino- and carboxyl-terminal halves of

## Radixin



G F G F G F G F G F G F  
C PS PC PI PIP PIP<sub>2</sub>

## Moesin



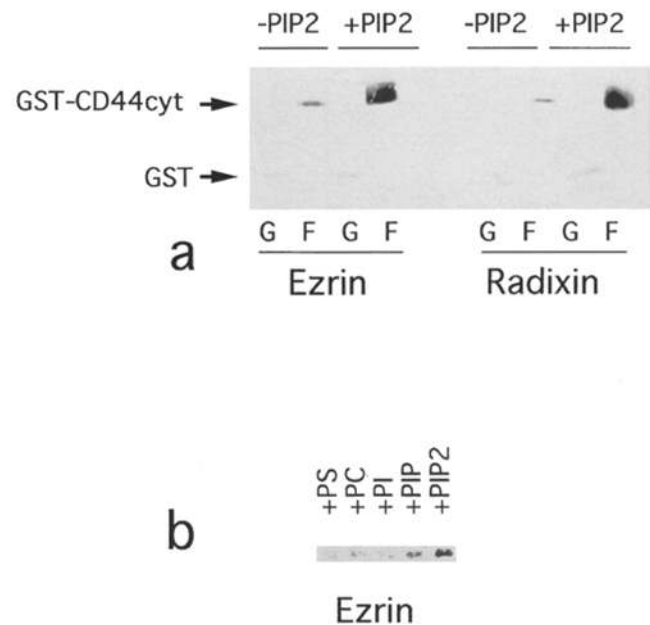
G F G F G F G F G F G F  
C PS PC PI PIP PIP<sub>2</sub>

**Figure 4.** Effects of phospholipids on the association of radixin and moesin with the cytoplasmic domain of CD44. As described in Fig. 2, radixin or moesin was incubated with GST-CD44cyt or GST bound to beads at physiological ionic strength in the absence (C) or presence of phosphatidylserine (PS), phosphatidylcholine (PC), phosphatidylinositol (PI), phosphatidylinositol 4-phosphate (PIP), or phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>). After washing, the protein associated with GST-CD44cyt (F) or GST (G) was eluted from the beads with a buffer containing glutathione. Proteins in the glutathione eluate were separated by SDS-PAGE and then immunoblotted with antiradixin or anti-moesin mAbs.

mouse moesin by means of recombinant baculovirus infection, purified them to homogeneity, then incubated them with the GST-CD44cyt beads at low or physiological ionic strength in the absence or presence of 4,5-PIP<sub>2</sub> (Fig. 7). In sharp contrast to full-length moesin, the amino-terminal half bound to GST-CD44cyt not only at low, but also at physiological ionic strength in the absence of 4,5-PIP<sub>2</sub>, indicating that the binding between GST-CD44cyt and the amino-terminal half of moesin at physiological ionic strength did not require 4,5-PIP<sub>2</sub>. The carboxyl-terminal half of moesin did not have any binding affinity for GST-CD44cyt under any of the conditions tested.

### *Rho-GDI Coimmunoprecipitated with the CD44/Moesin Complex*

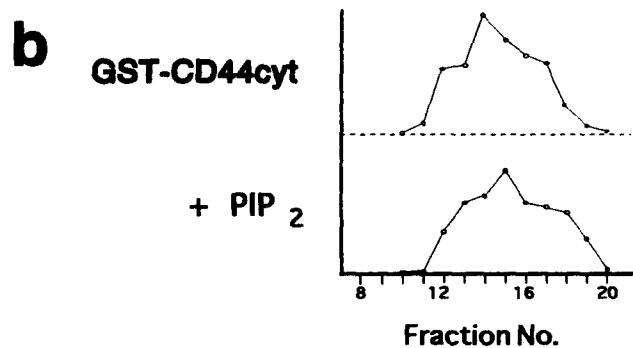
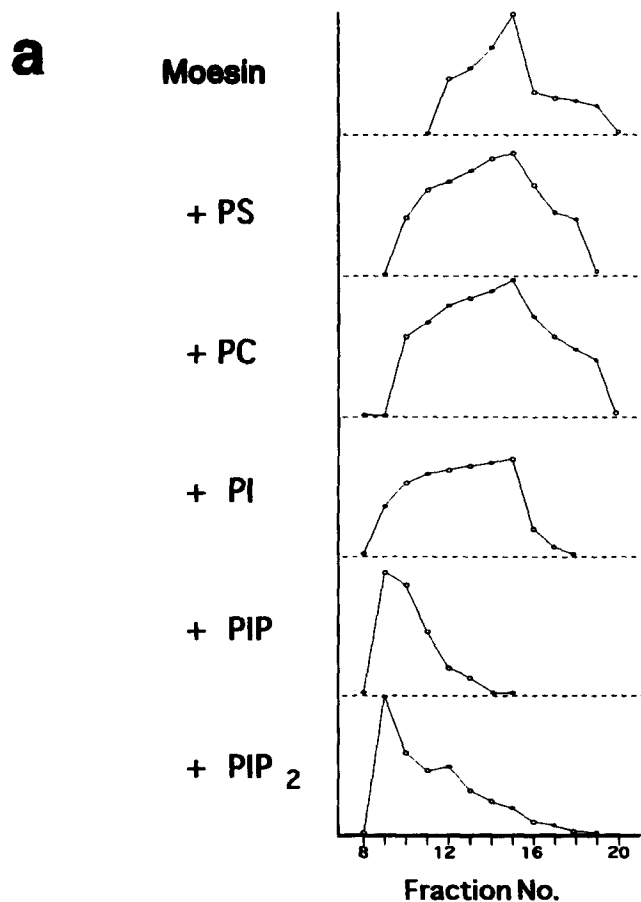
The above in vitro findings suggest that the formation of CD44/ERM complex is dynamically regulated in vivo. As a first step to examine the in vivo regulation mechanism of



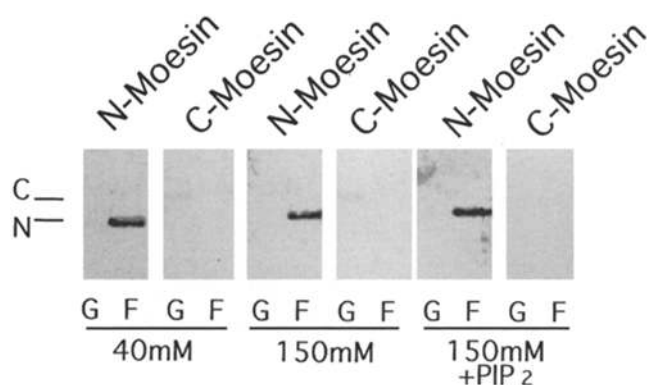
**Figure 5.** Association of GST-CD44cyt with ezrin- or radixin-coupled beads. (a) Ezrin- or radixin-coupled Sepharose beads (Ezrin or Radixin, respectively) were incubated with purified GST-CD44cyt (F) or GST (G) at physiological ionic strength (150 mM KCl) in the absence (-PIP<sub>2</sub>) or presence (+PIP<sub>2</sub>) of phosphatidylinositol 4,5-bisphosphate, and bound proteins were eluted with SDS-PAGE sample buffer. The eluates were separated by SDS-PAGE and then immunoblotted with anti-GST pAb. (b) Ezrin-coupled Affigel beads were incubated with purified GST-CD44cyt at physiological ionic strength (150 mM KCl) in the presence of phosphatidylserine (PS), phosphatidylcholine (PC), phosphatidylinositol (PI), phosphatidylinositol 4-phosphate (PIP), or phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>), and bound proteins were eluted with SDS-PAGE sample buffer. The eluates were resolved by electrophoresis and stained with Coomassie brilliant blue.

this complex formation, we immunoprecipitated the CD44/moesin complex from cultured BHK cells (Tsukita et al., 1994) and reexamined its constituents, focusing upon the small molecular mass range. Cultured BHK cells were metabolically labeled with [<sup>35</sup>S]methionine, solubilized with RIPA buffer, and then immunoprecipitated with anti-moesin mAb, CR-22. The immunoprecipitate was resolved in 12.5% SDS-polyacrylamide gels in the absence of mercaptoethanol to bring the IgG in the sample into the high molecular mass region. These conditions allowed small molecular mass constituents of the immunoprecipitates to be analyzed by autoradiography (Fig. 8 a).

As compared with the control immunoprecipitate, that of moesin reproducibly contained at least three low molecular mass polypeptides (55, 30, and 25 kD) in addition to moesin and CD44 (Tsukita et al., 1994). To exclude the possibility of nonspecific binding of these polypeptides to the immunoprecipitate, we added the excess amount of purified unlabeled recombinant moesin into the lysate from [<sup>35</sup>S]methionine-labeled BHK cells before immunoprecipitation (Fig. 8 b). As a result of competition, most [<sup>35</sup>S]methionine-labeled bands (three low molecular mass polypeptides [55, 30, and 25 kD], moesin, and CD44) were



**Figure 6.** Binding affinity of ERM proteins and the cytoplasmic domain of CD44 with phospholipids. (a) Interaction of moesin with phospholipids. Purified recombinant moesin was incubated without (*Moesin*) or with phosphatidylserine (*PS*), phosphatidylcholine (*PC*), phosphatidylinositol (*PI*), phosphatidylinositol 4-phosphate (*PIP*), or phosphatidylinositol 4,5-bisphosphate (*PIP<sub>2</sub>*) and passed through a column containing Superose 12R gel. Elution profiles of moesin were obtained by ELISA with anti-moesin mAb. An incubation of moesin with *PS*, *PC*, or *PI* did not affect its behavior in the column. By contrast, when moesin was incubated with 4-*PIP* or 4,5-*PIP<sub>2</sub>*, it coeluted with 4-*PIP* or 4,5-*PIP<sub>2</sub>* in the void volume. 4-*PIP* and 4,5-*PIP<sub>2</sub>* had the same effect upon ezrin and radixin (data not shown). (b) Binding affinity of GST-CD44cyt with 4,5-*PIP<sub>2</sub>*. Incubating GST-CD44cyt with 4,5-*PIP<sub>2</sub>* did not affect its binding behavior.



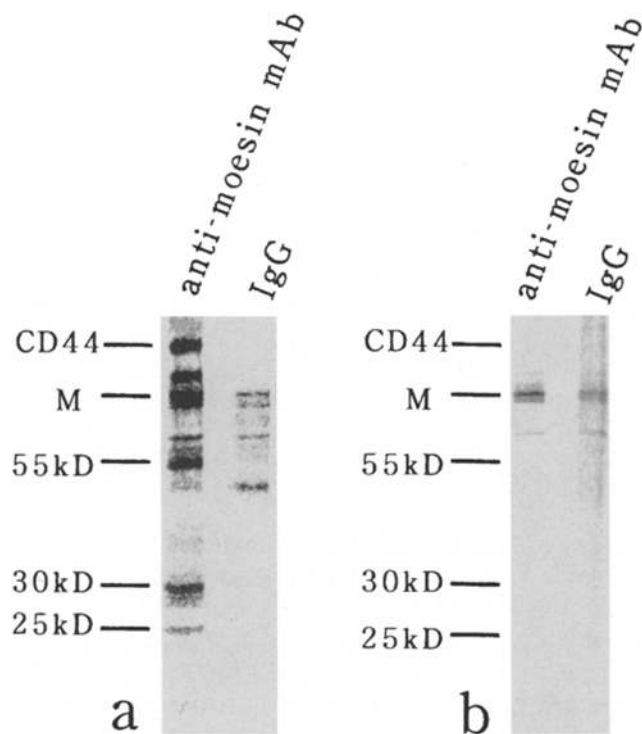
**Figure 7.** Interaction of amino- and carboxyl-terminal halves of moesin with GST-CD44cyt beads. As described in the legend to Fig. 2, the amino- or carboxyl-terminal half of moesin (*N-Moesin* and *C-Moesin*, respectively) was incubated with GST-CD44cyt- or GST-bound beads at low (40 mM) or at physiological ionic strength (150 mM) in the absence or presence (+*PIP<sub>2</sub>*) of 4,5-*PIP<sub>2</sub>*. After washing, the protein associated with GST-CD44cyt (*F*) or GST (*G*) was eluted from the beads with a buffer containing glutathione. Proteins in the glutathione eluate were separated by SDS-PAGE and then immunoblotted with anti-ERM pAbs (p-800 and I1) for the amino- and carboxyl-terminal halves, respectively. The bands of C- and N-moesin are indicated by *C* and *N* on the left.

removed from the immunoprecipitate, indicating that these low molecular mass polypeptides and CD44 were specifically precipitated with anti-moesin mAb through their association with moesin.

To identify these low molecular mass components, we analyzed the cross-reactivity of various antibodies specific to known cytoplasmic proteins. Considering that the Rho-signaling pathway plays a key role in the regulation of actin filament/plasma membrane association, we investigated whether or not there were Rho and/or Rho regulators in the above moesin immunoprecipitate. The molecular mass of Rho is around 21 kD, so we concluded that Rho itself was not included in the immunoprecipitate. Among Rho regulators, the molecular mass of Rho-GDI is reportedly ~30 kD (Fukumoto et al., 1990). Then we checked whether or not the 30-kD band in the moesin immunoprecipitate is identical to Rho-GDI.

The moesin immunoprecipitate was resolubilized and reimmunoprecipitated with anti-mouse Rho-GDI mAb. As shown in Fig. 9 a, most of the 30-kD band was specifically reimmunoprecipitated by this mAb, leaving the other bands in the supernatant. Since this mAb was raised against mouse Rho-GDI, we examined its specificity using hamster Rho-GDI. When BHK cells were immunoprecipitated with the anti-mouse Rho-GDI mAb, a 30-kD polypeptide was specifically recognized (Fig. 9 a). Furthermore, we compared the 30-kD band in the moesin immunoprecipitates with the *in vitro* translated mouse Rho-GDI by one-dimensional peptide mapping. As shown in Fig. 9 b, the peptide map of the 30-kD band was very similar to that of *in vitro* translated mouse Rho-GDI. We thus concluded that Rho-GDI is tightly associated with the CD44/moesin complex *in vivo*.



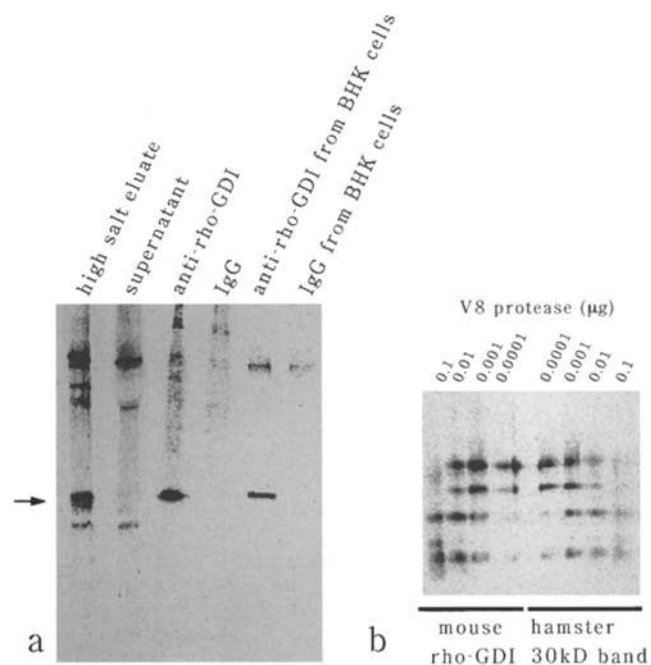


**Figure 8.** Coimmunoprecipitation of low molecular mass proteins with the CD44/ERM complex. (a) Cultured BHK cells were metabolically labeled with [<sup>35</sup>S]methionine, solubilized with RIPA buffer, and then immunoprecipitated with anti-moesin mAb CR-22 or nonimmune mouse IgG. The immunoprecipitate (*anti-moesin mAb* or *IgG*, respectively) was separated in a 15% SDS-polyacrylamide gel in the absence of mercaptoethanol to bring the IgG band to the high molecular mass regions and quantified by image analysis. In addition to CD44 (see Tsukita et al., 1994), three bands around 55, 30, and 25-kD coimmunoprecipitated with moesin (*M*). (b) Purified, unlabeled moesin was added to the RIPA lysate of [<sup>35</sup>S]methionine-labeled BHK cells and immunoprecipitated with anti-moesin mAb CR-22 or nonimmune mouse IgG (*anti-moesin mAb* or *IgG*, respectively). [<sup>35</sup>S]methionine-labeled bands of 55, 30, and 25-kD as well as CD44 were undetectable, although a weakly labeled moesin band was evident.

### Binding of Recombinant ERM Proteins to the Insoluble Fraction of Crude Cell Homogenate Is Suppressed by C3 Toxin

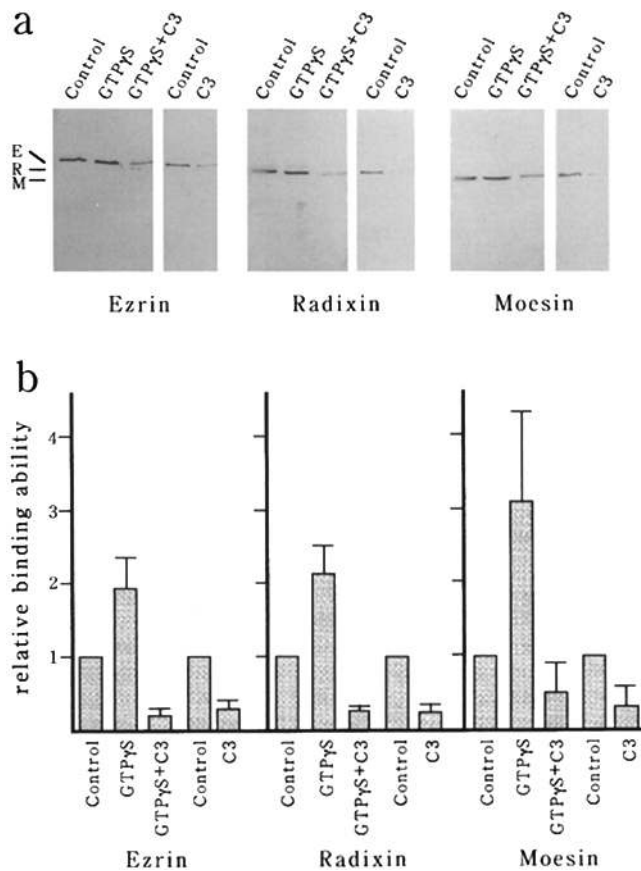
To initially evaluate the notion that the formation of CD44/ERM complex is regulated by the Rho-signaling pathway, we examined whether or not a GTP-binding protein is involved in regulating the binding of ERM proteins to the insoluble components of cells including plasma membranes. For this purpose, we established a cell-free system that allowed the estimation of ERM binding affinity to the insoluble components. Since we identified the CD44/ERM complex in BHK cells by immunoprecipitation, we used the same cells in this experiment.

BHK cells were lysed in lysis buffer at physiological ionic strength containing 0.06% Nonidet P-40, and the protein concentration in the lysate was adjusted to ~12 mg/ml. To estimate the binding affinity of ERM proteins to the insoluble fraction, recombinant ezrin, radixin, or moesin was added and incubated with the lysate. When



**Figure 9.** Identification of the 30-kD polypeptide in the moesin immunoprecipitate as a hamster Rho-GDI. (a) Specific recognition of the 30-kD peptide by anti-mouse Rho-GDI mAb. The moesin immunoprecipitate (see Fig. 8) was solubilized from the beads with high salt, and then the eluate (*high salt eluate*) was again immunoprecipitated with anti-mouse Rho-GDI mAb (*anti-rhoGDI*) or non-immune mouse IgG. The 30-kD band (*arrow*) was specifically immunoprecipitated, leaving the supernatant lacking the 30-kD band (*supernatant*). This anti-mouse Rho-GDI mAb specifically immunoprecipitated a polypeptide with a molecular mass of 30-kD from the BHK cells metabolically labeled with [<sup>35</sup>S]methionine (*anti-rhoGDI from BHK cells*). (b) One-dimensional peptide mapping. Mouse Rho-GDI, which was translated in a cell-free system then labeled with [<sup>35</sup>S]methionine using a rabbit reticulocyte lysate, and the 30-kD band in the moesin immunoprecipitate from BHK cells (see Fig. 8) were analyzed by one-dimensional peptide mapping using *Staphylococcus aureus* V8 protease. The map of mouse Rho-GDI was very similar to that of the hamster 30-kD band.

~0.8 μg of ezrin, radixin, or moesin was added to the lysate to a final volume of 15 μl, ~0.05 μg of each protein was recovered in the insoluble fraction after centrifugation (Fig. 10 a). The amount of endogenous ERM proteins recovered in the insoluble fraction was too small to be detected under our experimental conditions. The amount of recovered protein was estimated by comparing the staining intensity of each band in immunoblotting with that of the various amounts of authentic ERM proteins resolved by electrophoresis and immunoblotted on the same nitrocellulose sheet (Fig. 10 b). When the amount of ERM proteins added was increased, their binding to the insoluble fraction was saturable; at the level of ~0.8 μg added ERM proteins, binding was saturated sufficiently. Upon adding GTPγS to the incubation system, the amount of recovered ezrin, radixin, and moesin was increased 1.9 ± 0.5-fold, 2.1 ± 0.3-fold, and 3.1 ± 1.3-fold, respectively, averaged from four experiments. This suggested that the GTPγS ac-



**Figure 10.** Rho-dependent association of the recombinant ERM proteins with the insoluble fraction of BHK cell lysate. (a) Immunoblot analyses. After  $\sim 0.8$   $\mu$ g recombinant ezrin, radixin, or moesin was incubated with the BHK cell homogenate followed by centrifugation, the pellet was analyzed by immunoblotting with anti-ERM pAb I1. Under these conditions, the amount of endogenous ERM proteins was too small to be detected as compared to that of added ERM proteins, and  $\sim 0.05$   $\mu$ g of each ERM protein was recovered in the insoluble fraction (Control). When GTP $\gamma$ S was added to the incubation mixture, the amount of recovered ERM proteins in the insoluble fraction was significantly increased (GTP $\gamma$ S). C3 toxin markedly suppressed the binding of each ERM protein to the insoluble fraction in the presence (GTP $\gamma$ S+C3) as well as in the absence (C3) of GTP $\gamma$ S. The bands of ezrin, radixin, and moesin are indicated by E, R, and M, respectively, on the left. (b) Relative binding ability of ERM proteins to the insoluble fraction. The amount of recovered protein was estimated by comparing the staining intensity of each band in immunoblotting with that of the various amounts of authentic ERM proteins resolved by electrophoresis and immunoblotted on the same nitrocellulose sheet. A relative binding ability of 1 was equivalent to the amount of recovered ezrin, radixin, or moesin in the insoluble fraction for each experiment. Values represent relative binding abilities averaged from four experiments  $\pm$  SEMs.

tivation of some GTP-binding protein in cell lysates increases the ERM binding ability to the insoluble fraction.

C3 toxin, a specific Rho inactivator, was added to the incubation mixture with NAD<sup>+</sup>. As shown in Fig. 10 a, the binding ability of ERM proteins to the insoluble fraction of the lysate was remarkably suppressed by C3 toxin at a concentration of 0.1 mg/ml. In the presence of GTP $\gamma$ S, C3

toxin reduced the binding affinity of ezrin, radixin, and moesin to  $21 \pm 10\%$ ,  $29 \pm 5\%$ , and  $50 \pm 35\%$ , respectively, of the value in the absence of C3 toxin and GTP $\gamma$ S (control value). Also in the absence of GTP $\gamma$ S, the respective binding affinities were reduced by C3 toxin to  $23 \pm 11\%$ ,  $25 \pm 12\%$ , and  $32 \pm 30\%$  of the control value, suggesting that there was some stimulation by endogenous active Rho in the lysate. As a control, we added the same amount of BSA or IgG to the BHK cell lysate instead of ERM proteins, and no Rho-dependent binding of these proteins was identified. As another control, we added the recombinant ERM proteins to the same amount of buffer instead of the BHK cell lysate, and no GTP $\gamma$ S/C3 toxin-dependent precipitation of ERM proteins was observed. These results indicated that activated Rho is required for the interaction between the insoluble fraction and recombinant ERM proteins.

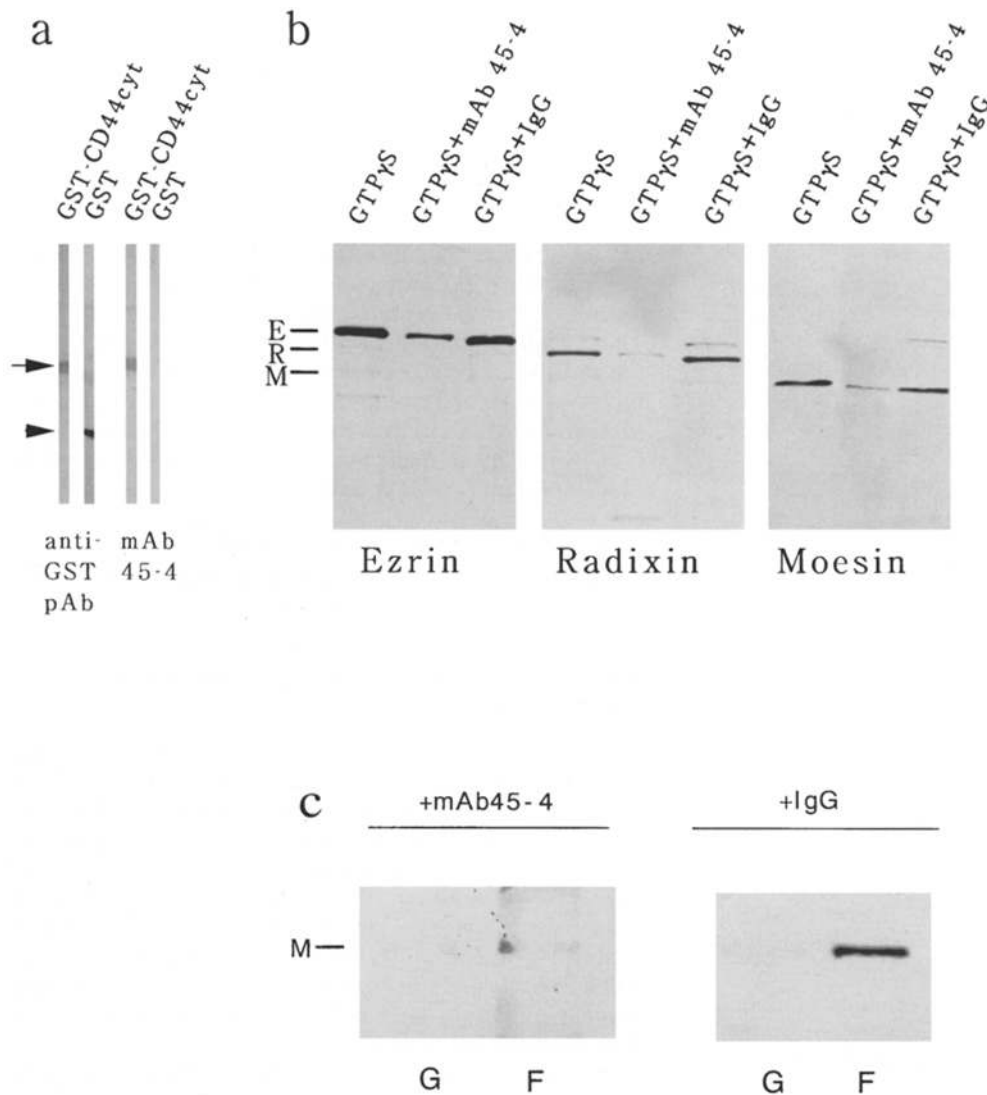
#### **Binding of Recombinant ERM Proteins to the Insoluble Fraction of BHK Cell Lysate Is Suppressed by an Anti-CD44 Cytoplasmic Domain mAb**

We studied whether the Rho-dependent association of exogenous ERM proteins with the insoluble fraction can be attributed to the CD44/ERM or other unknown interactions. We raised mAbs using the GST fusion protein with the cytoplasmic domain of mouse CD44 as the antigen in rats and then studied whether or not they interfere with the Rho-dependent association of ERM proteins in the presence of GTP $\gamma$ S. We obtained one mAb (45-4), that recognized the cytoplasmic domain of hamster CD44 as well as mouse CD44 on immunoblots (Fig. 11 a). When 0.5  $\mu$ g of purified mAb 45-4 was added to 15  $\mu$ l of the incubation solution, the binding affinity of ERM proteins to the insoluble fraction was suppressed (Fig. 11 b) to a level that was almost equivalent to that of C3 toxin. In contrast, nonimmune rat IgG did not affect the association. Furthermore, this mAb effectively interfered with the *in vitro* binding of recombinant moesin with GST-CD44cyt at physiological ionic strength in the presence of 4,5-PIP<sub>2</sub>, while control nonimmune IgG did not affect this interaction (Fig. 11 c).

#### **C3 Toxin Interferes with ERM-Plasma Membrane Interaction in Living Cells**

We examined the effect of C3 toxin on the ERM-plasma membrane interaction in living BHK cells. This strategy is however, technically difficult. C3 toxin-induced morphological changes (rounding up of cells) are undetectable for 12 h in living BHK cells, and the effect is not particularly synchronized among treated cells. To circumvent this difficulty, we introduced C3 toxin into cells using the lipofectamin, which rather clarified its effect.

After a 6-h incubation with the lipofectamin-conjugated C3 toxin, BHK cells were homogenized in PBS and then divided into soluble and insoluble fractions by centrifugation. From the untreated BHK cells,  $\sim 30\%$  of the endogenous ERM proteins were recovered in the insoluble fraction, whereas only a trace amount was recovered in the insoluble fraction obtained from C3 toxin-treated cells (Fig. 12 a). The untreated and C3 toxin-treated BHK cells were lysed with RIPA buffer and immunoprecipitated



**Figure 11.** The cytoplasmic domain of CD44 as a binding partner in the insoluble fraction of BHK cell lysate for recombinant ERM proteins. (a) Specificity of mAb 45-4 for the cytoplasmic domain of mouse CD44. Immunoblotting of the lysate of Sf9 cells expressing the GST fusion protein with the cytoplasmic domain of mouse CD44 (*GST-CD44cyt*) or the lysate of *E. coli* expressing the GST protein with anti-GST pAb or mAb 45-4. Anti-GST pAb recognized both *GST-CD44cyt* (arrow) and GST (arrowhead), whereas mAb 45-4 recognized only *GST-CD44cyt*. (b) The same binding experiments were performed as those shown in Fig. 10, except that C3 toxin was replaced with the purified mAb 45-4. As controls, non-immune rat IgG was added instead of mAbs (IgG). The mAb 45-4 markedly suppressed the binding of recombinant ERM proteins. The bands of ezrin, radixin, and moesin are indicated by E, R, and M respectively, on the left. (c) Effects of the mAb 45-4 on the association of recombinant moesin with the *GST-CD44cyt* beads at physiological ionic strength in the presence of 4, 5-PIP<sub>2</sub>. Purified mAb 45-4 (+mAb45-4)

or control nonimmune IgG (+IgG) was added to the incubation mixture at a final concentration of 30  $\mu\text{g/ml}$ . Moesin bound to *GST-CD44cyt* (F) or GST (G) was eluted and detected by immunoblotting with anti-moesin mAb.

with anti-CD44 antibody (Fig. 12 b). From untreated BHK cells, some ERM proteins were recovered and detected by immunoblotting in the CD44 immunoprecipitate, whereas ERM proteins were virtually undetectable in the CD44 immunoprecipitate from the C3 toxin-treated cells.

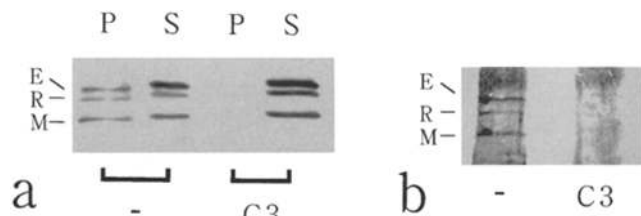
## Discussion

### *ERM Proteins Directly Bind to the Cytoplasmic Domain of CD44 as a 4,5-PIP<sub>2</sub>-dependent Manner In Vitro*

We found that ERM proteins coimmunoprecipitate with CD44 (Tsukita et al., 1994). However, so far in vitro evidence for a direct association between the cytoplasmic domain of CD44 and ERM proteins has not been obtained. Here, using an in vitro binding assay, we revealed that at low ionic strength (40 mM KCl), ERM proteins directly bind to the cytoplasmic domain of CD44 at relatively high

affinity ( $K_d$  of moesin is  $9.3 \pm 1.6$  nM). Although this direct binding became undetectable at physiological ionic strength (150 mM KCl), 4,5-PIP<sub>2</sub> facilitated the CD44 binding of ERM proteins ( $K_d$  of moesin is  $9.3 \pm 4.8$  nM). Thus, we concluded that ERM proteins directly bind to the cytoplasmic domain of CD44 in a regulated manner and that ezrin, radixin, and moesin are essentially indistinguishable with respect to their binding capacity with the cytoplasmic domain of CD44.

Many actin-modulating proteins, such as gelsolin (Janmey and Stossel, 1987), gCap39 (Yu et al., 1990), profilin (Lassing and Lindberg, 1985), cofilin (Yonezawa et al., 1991), and  $\alpha$ -actinin (Fukami et al., 1992) reportedly bind to 4,5-PIP<sub>2</sub>. 4,5-PIP<sub>2</sub> induces the dissociation of profilin-actin and gelsolin-actin complexes, and the interaction of cofilin with G- and F-actin is also inhibited by PI, 4-PIP, and 4,5-PIP<sub>2</sub>. Through these mechanisms, the structural integrity of actin-based cytoskeletons is thought to be regulated depending on the level of phosphoinositides in vivo.



**Figure 12.** Behavior of ERM proteins in the C3 toxin-treated living BHK cells. (a) BHK cells incubated in the presence (C3) or absence (-) of C3 toxin were homogenized in physiological solution (see Materials and Methods) and then divided into soluble supernatant (S) and insoluble pellet (P) fractions by centrifugation. Each fraction was resolved by SDS-PAGE, and ERM proteins were detected by immunoblotting with anti-ERM pAb I1. In the insoluble fraction of untreated BHK cells (-), ~30% of the total ERM proteins was recovered, whereas only a trace amount was recovered in the insoluble fraction of C3 toxin-treated cells (C3). (b) The RIPA lysates of C3 toxin-treated (C3) or untreated (-) BHK cells were immunoprecipitated by mAb 30189, which is specific for the extracellular domain of hamster CD44, and each immunoprecipitate was immunoblotted with anti-ERM pAb I1. ERM proteins were reproducibly detected in the CD44 immunoprecipitate in the absence of C3 toxin as described (Tsukita et al., 1994), whereas they were hardly detectable in the immunoprecipitate in the presence of C3 toxin.

The interaction of ERM proteins with the cytoplasmic domain of CD44 is enhanced by 4,5-PIP<sub>2</sub>, increasing the number of actin filament-association sites on the cytoplasmic surface of plasma membranes. This may also markedly affect the structural integrity of actin-based cytoskeletons. ERM proteins localize and function just beneath the plasma membrane, so compared with other actin-modulating proteins, they have many topological opportunities to interact with phosphoinositides, which are components of plasma membranes.

#### **The Mechanism by Which 4,5-PIP<sub>2</sub> Enhances the Association between the Cytoplasmic Domain of CD44 and ERM Proteins**

This study revealed that ERM proteins bind to 4,5-PIP<sub>2</sub> with high affinity. Considering that the cytoplasmic domain of CD44 does not have any affinity for 4,5-PIP<sub>2</sub>, the facilitation of the CD44/ERM interaction by 4,5-PIP<sub>2</sub> is derived not from artifactual cross-linking of CD44 and ERM by 4,5-PIP<sub>2</sub> micelles, but from the 4,5-PIP<sub>2</sub>-induced conformational change of ERM proteins. During the course of this study, Niggli et al. (1995) independently reported, using different techniques, that 4,5-PIP<sub>2</sub> binds ezrin with high affinity and that its binding domain of ezrin has been narrowed down to the amino-terminal half.

Several studies have indicated that the carboxyl- and amino-terminal halves of ERM proteins have distinct functions. The respective halves bind to actin-based cytoskeletons and plasma membranes (Algrain et al., 1993). Some *in vitro* and *in vivo* data suggested that the amino-terminal half of native ERM proteins interferes with the interaction of their carboxyl-terminal half with actin filaments (Turunen et al., 1994; Martin et al., 1995). Actually,

the amino-terminal half of ezrin binds to its carboxyl-terminal half directly (Gary and Bretscher, 1995).

In this study, at physiological ionic strength in the absence of phosphoinositides, full-length moesin very weakly bound the cytoplasmic domain of CD44, whereas the amino-terminal half of moesin bound with high affinity. This indicated not only that the amino-terminal half of moesin is responsible for its association with plasma membranes (CD44) but also that in native moesin, the carboxyl-terminal half suppresses the direct interaction of the amino-terminal half with the cytoplasmic domain of CD44. These findings indicate that the amino- and carboxyl-terminal halves of ERM proteins suppress the functions of their carboxyl- and amino-terminal halves; that is, binding to actin-based cytoskeletons and plasma membranes, respectively. This mutual suppression mechanism would be very important for ERM proteins to function as molecular cross-linkers between actin filaments and plasma membranes. This study showed that 4,5-PIP<sub>2</sub> can function as a key modulator to release this mutual suppression immediately below the plasma membrane.

#### **Rho-GDI Is Associated with the CD44/Moesin Complex *In Vivo***

To examine the *in vivo* regulation mechanism of the CD44/ERM complex formation, we reexamined the immunoprecipitated CD44/moesin complex from BHK cells. Immunoprecipitation and peptide mapping revealed that this immunoprecipitate contained not only moesin and CD44 but also three smaller molecular mass proteins around 55, 30, and 25 kD, and that the 30-kD protein is identical to hamster Rho-GDI. So far, the physiological function of Rho-GDI is not clear. Rho-GDI was originally thought to form a tight complex with the GDP-form of Rho to form a cytoplasmic pool (Araki et al., 1990). However, it may recruit cytoplasmic Rho to the plasma membrane to be activated there as it is in Rab-GDI (Novick and Garret, 1994; Horiuchi et al., 1995; Takai et al., 1995). The Rab molecule undergoes a cycle of localization coupled with that of nucleotide exchange and hydrolysis (Novick and Garrett, 1994); Rab remains soluble until the Rab/Rab-GDI complex interacts with the appropriate donor membrane, promoting dissociation from Rab-GDI, membrane attachment and the exchange of GDP for GTP (Soldati et al., 1994; Ullrich et al., 1994; Horiuchi et al., 1995). If this recycling pathway of Rab is also true of Rho, the occurrence of Rho-GDI in the CD44/moesin complex would indicate that this complex is a site where active Rho regulates the plasma membrane/actin filament interaction. The immunoprecipitation results agree with the findings that in MDCK cells expressing myc-tagged RhoA, activated Rho is reportedly translocated from the cytoplasm to ERM-concentrated regions of plasma membranes (Takaishi et al., 1995).

In our immunoprecipitates, probably because of the severe solubilizing conditions, Rho itself was not detected, leaving Rho-GDI bound to the complex. As far as we examined, none of the available antibodies specific for Rho-related proteins such as Rho-GDS and Rho-GAP recognized the other 55- and 25-kD bands in the immunoprecipitate. The identification of these proteins will lead to a better

understanding of the physiological functions of CD44/ERM complex. However, the small amount of these proteins recovered as immunoprecipitates makes this identification technically challenging.

### ***The CD44/ERM Complex Formation Is Regulated by Rho In Vivo***

To evaluate the involvement of Rho in the CD44/ERM complex formation, we developed a cell-free system with which to examine the binding affinity of exogenous ERM proteins to the insoluble fraction of the BHK cell lysate. Using this system, we found that GTP $\gamma$ S facilitated the binding and that C3 toxin remarkably suppressed it in the presence as well as in the absence of GTP $\gamma$ S. This indicated that the GTP form of Rho included in the cell lysate is required for this binding. Furthermore, we found that one of the anti-CD44 cytoplasmic domain mAbs also effectively suppressed the ERM binding to the insoluble fraction and that this mAb interfered with the association between ERM proteins and CD44 in vitro. We then concluded that most of the membrane binding partners in this BHK cell-free system are CD44 molecules and that the association between ERM proteins and the cytoplasmic domain of CD44 is regulated by Rho. As described in the Introduction, there are other potential ERM membrane binding partners. It remains elusive whether or not the binding of these possible partners to ERM proteins are also Rho dependent.

Recently, ezrin was reported to be oligomerized possibly through phosphorylation (Berryman et al., 1995; Gary and Bretscher, 1995). This nature of ezrin is probably shared by radixin and moesin (Andreoli et al., 1994). The effective suppression of ERM binding to the insoluble fraction by an anti-CD44 cytoplasmic domain mAb, however, indicates that our cell-free system detected CD44/ERM interaction rather than ERM oligomerization.

The results obtained from the incubating living BHK cells with C3 toxin also favored the notion that the formation of CD44/ERM complex is dependent on active Rho. C3 toxin in living cells recruited most insoluble ERM proteins to the soluble pool, leaving CD44 free from ERM proteins. Considering that the effect of C3 toxin in living cells is not sharp in terms of its time course and synchronization, the effect observed in living cells is highly compatible to that observed in our cell-free system.

Ridley and Hall (1992) and Ridley et al. (1992) have found using serum-starved Swiss 3T3 cells that Rho plays a central role in the coordinated assembly of focal adhesions and stress fibers induced by growth factors, and that Rac stimulates actin filament accumulation at the plasma membrane, forming membrane ruffles. Furthermore, Cdc42 controls the formation of filopodia (Kozma et al., 1995; Nobes and Hall, 1995). However, the relationship between these three small GTP-binding proteins and actin-based cellular events does not appear simple. For example, in KB and MDCK cells, membrane ruffle formation is reportedly regulated by Rho (Takaishi et al., 1993; Nishiyama et al., 1994), and the cleavage furrow formation is also Rho dependent in *Xenopus* and sea urchin eggs (Kishi et al., 1993; Mabuchi et al., 1993). The Rho-dependent enhancement of the CD44/ERM complex formation would provide an

increasing number of actin filament attachment sites on the cytoplasmic surface of plasma membranes at microvilli, cleavage furrows, ruffling membranes, and cell-cell/cell-matrix adhesion sites. This speculation could explain the diversified functions of Rho in the regulation mechanism of actin filament/plasma membrane interaction in general.

### ***The Elevation of 4,5-PIP<sub>2</sub> Level in Plasma Membranes Is Possibly Involved in the Rho-dependent Regulation of the CD44/ERM Complex Formation***

The question then arises as to how Rho regulates the formation of the CD44/ERM complex in vivo. One explanation is that Rho directly regulates CD44/ERM association. This possibility was not considered likely. An alternative is that through one or several Rho downstream components, this complex formation is Rho dependent. Several intensive studies have been performed to identify the direct target of Rho. However, the list of activities stimulated by Rho as well as Rac and Cdc42 is long, and includes cascades of kinases that regulate gene transcription and cell growth, none of which have been linked to the actin filament/plasma membrane interactions (Zigmond, 1995; Watanabe et al., 1996; Amano et al., 1996). Rho reportedly regulates PI turnover (Chong et al., 1994; Divecha and Irvine, 1995; Zhang et al., 1995). Furthermore, considering that Rac also rearranges the actin filaments also through 4,5-PIP<sub>2</sub> synthesis (Hartwig et al., 1995), Rho might control actin-based cellular events by similar mechanisms. The enzymatic activities of phosphatidylinositol 3-kinase (Zhang et al., 1993), phosphatidylinositol 4-phosphate 5-kinase (Chong et al., 1994), and phospholipase D (Bowman et al., 1993; Malcolm et al., 1994) are reportedly regulated by Rho. Among them, the activation of phosphatidylinositol 4-phosphate 5-kinase elevates the 4,5-PIP<sub>2</sub> concentration in membranes.

At least as one of the signaling pathways that links Rho to the actin-based cytoskeleton, we speculate the following. (a) Activation of Rho directly or indirectly upregulates PIP5-kinase. (b) Activated PIP 5-kinase elevates the 4,5-PIP<sub>2</sub> level in plasma membranes. (c) The CD44/ERM complex formation is induced by 4,5-PIP<sub>2</sub>. The elevated 4,5-PIP<sub>2</sub> level in the plasma membranes could be derived from mechanisms other than PIP 5-kinase activation. Regardless, considering that lovastatin, a general inhibitor of small GTP-binding proteins, suppresses the elevation of 4,5-PIP<sub>2</sub> in the plasma membrane in vivo (Chong et al., 1994), Rho is a likely upstream regulator of this elevation.

Rho also reportedly regulates the activity of some serine/threonine kinases (Zigmond, 1995; Amano et al., 1996; Watanabe et al., 1996). ERM proteins are highly serine/threonine phosphorylated (Gould et al., 1986; Urushidani et al., 1989; Nakamura et al., 1995), and their phosphorylation is suggested to be involved in the regulation of the ERM/membrane association (Chen et al., 1995). We should examine the possibility that Rho regulates the ERM/membrane association through the serine/threonine phosphorylation of ERM proteins. The in vitro binding system developed in this study would be useful for this purpose. Further molecular studies on how active Rho facilitates the formation of CD44/ERM complex in vivo and how actin filament association with plasma membranes is

facilitated through the formation of this complex are now underway in our laboratory.

We thank all the members of our laboratory (Department of Cell Biology, Faculty of Medicine, Kyoto University, Kyoto, Japan) for their helpful discussions. Our thanks are also due to Miss Naomi Sekiguchi for technical assistance. C3 toxin was kindly supplied from Drs. S. Narumiya and Y. Saito (Kyoto University). M. Hirao thanks Dr. H. Shiozaki (Osaka University) for providing him with the opportunity to work in the Department of Cell Biology, Faculty of Medicine, Kyoto University.

This work was supported in part by a Grant-in-Aid for Scientific Research (B) (to Sa. Tsukita), a Grant-in-Aid for Cancer Research, and a Grant-in-Aid for Scientific Research (A) from the Ministry of Education, Science and Culture of Japan, and by research grants from the Yamada Science Foundation, the Mitsubishi Foundation, and the Toray Science Foundation (to Sh. Tsukita).

Received for publication 16 August 1995 and in revised form 19 June 1996.

## References

Aktorjes, K., S. Rosener, U. Blaschke, and G. S. Chhatwal. 1988. Botulinum ADP-ribosyltransferase C3: purification of the enzyme and characterization of the ADP-ribosylation reaction in platelet membranes. *Eur. J. Biochem.* 172:445–450.

Algrain, M., O. Turunen, A. Vaheri, D. Louvard, and M. Arpin. 1993. Ezrin contains cytoskeleton and membrane binding domains accounting for its proposed role as a membrane-cytoskeletal linker. *J. Cell Biol.* 120:129–140.

Amano, M., H. Mukai, Y. Ono, K. Chihara, T. Matsui, Y. Hamajima, K. Okawa, A. Iwamatsu, and K. Kaibuchi. 1996. Identification of a putative target for rho as the serine-threonine kinase protein kinase N. *Science (Wash. DC)*. 271:648–650.

Amieva, M.R., K.K. Wilgenbus, and H. Furthmayr. 1994. Radixin is a component of hepatocyte microvilli in situ. *Exp. Cell Res.* 210:140–144.

Andreoli, C., M. Martin, R. Le-Borgne, H. Reggio, and P. Mangeat. 1994. Ezrin has properties to self-associate at the plasma membrane. *J. Cell Sci.* 107: 2509–2521.

Araki, S., A. Kikuchi, Y. Hata, M. Isomura, and Y. Takai. 1990. Regulation of reversible binding of smg p25A, a ras p21-like GTP-binding protein, to synaptic plasma membranes and vesicles by its specific regulatory protein, GDP dissociation inhibitor. *J. Biol. Chem.* 265:13007–13015.

Arpin, M., M. Algrain, M., and D. Louvard. 1994. Membrane-actin microfilament connections: an increasing diversity of players related to band 4.1. *Curr. Opin. Cell Biol.* 6:136–141.

Bennett, V. 1989. The spectrin-actin junction of erythrocyte membrane skeletons. *Biochem. Biophys. Acta.* 988:107–121.

Berryman, M., Z. Franck, and A. Bretscher. 1993. Ezrin is concentrated in the apical microvilli of a wide variety of epithelial cells, whereas moesin is found primarily in endothelial cells. *J. Cell Sci.* 105:1025–1043.

Berryman, M., R. Gary, and A. Bretscher. 1995. Ezrin oligomers are major cytoskeletal components of placental microvilli: a proposal for their involvement in cortical morphogenesis. *J. Cell Biol.* 131:1231–1242.

Bowman, E.P., D.J. Uhlinger, and J.D. Lambeth. 1993. Neutrophil phospholipase D is activated by a membrane-associated Rho family small molecular weight GTP-binding protein. *J. Biol. Chem.* 268:21509–21512.

Braun, U., B. Habermann, I. Just, K. Aktories, and J. Vandekerckhove. 1989. Purification of the 22kDa protein substrate of botulinum ADP-ribosyltransferase C3 from porcine brain cytosol and its characterization as a GTP-binding protein highly homologous to the Rho gene product. *FEBS Lett.* 243:70–76.

Bretscher, A. 1983. Purification of an 80,000-dalton protein that is a component of isolated microvillus cytoskeleton, and its localization in nonmuscle cells. *J. Cell Biol.* 97:425–432.

Chen, J., J.A. Cohn, and L.J. Mandel. 1995. Dephosphorylation of ezrin as an early event in renal microvillar breakdown and anoxic injury. *Proc. Natl. Acad. Sci. USA.* 92:7495–7499.

Chong, L.D., A. Traynor-Kaplan, G.M. Bokoch, and M.A. Schwartz. 1994. The small GTP-binding protein Rho regulates a phosphatidylinositol 4-phosphate 5-kinase in mammalian cells. *Cell.* 79:507–513.

Cleveland, D.W., S.G. Fischer, M.W. Kirschner, and U.K. Laemmli. 1977. Peptide mapping by limited proteolysis in sodium dodecyl sulfate and analysis by gel electrophoresis. *J. Biol. Chem.* 252:1102–1106.

Conboy, J., Y.W. Kan, S.B. Shohet, and N. Mohandas. 1986. Molecular cloning of protein 4.1, a major structural element of the human erythrocyte membrane skeleton. *Proc. Natl. Acad. Sci. USA.* 83:9512–9516.

Divecha, N., and R.F. Irvine. 1995. Phospholipid signaling. *Cell.* 80:269–278.

Franck, Z., R. Gary, and A. Bretscher. 1993. Moesin, like ezrin, colocalizes with actin in the cortical cytoskeleton in cultured cells, but its expression is more variable. *J. Cell Sci.* 105:219–231.

Fukami, K., K. Furuhashi, M. Inagaki, T. Endo, S. Hatano, and T. Takenawa.

1992. Requirement of phosphatidylinositol 4, 5-bisphosphate for  $\alpha$ -actinin function. *Nature (Lond.)*. 359:150–152.

Fukumoto, Y., K. Kaibuchi, Y. Hori, H. Fujioaka, S. Araki, T. Ueda, A. Kikuchi, and Y. Takai. 1990. Molecular cloning and characterization of a novel type of regulatory protein (GDI) for the Rho proteins, ras p21-like small GTP-binding proteins. *Oncogene.* 5:1321–1328.

Funayama, N., A. Nagafuchi, N. Sato, Sa. Tsukita, and Sh. Tsukita. 1991. Radixin is a novel member of the band 4.1 family. *J. Cell Biol.* 115:1039–1048.

Gary, R., and A. Bretscher. 1995. Ezrin self-association involves binding of an N-terminal domain to a normally masked C-terminal domain that includes the F-actin binding site. *Mol. Biol. Cell.* 6:1061–1075.

Goldstein, L.A., D.F.H. Zhou, L.J. Picker, C.N. Minty, R.B. Bargatze, J.F. Ding, and E.C. Butcher. 1989. A human lymphocyte homing receptor, the Hermes antigen, is related to cartilage proteoglycan core and link proteins. *Cell.* 56:1063–1072.

Gould, K.L., J.A. Cooper, A. Bretscher, and T. Hunter. 1986. The protein-tyrosine kinase substrate, p81, is homologous to a chicken microvillar core protein. *J. Cell Biol.* 102:660–669.

Gould, K.L., A. Bretcher, F.S. Esch, and T. Hunter. 1989. cDNA cloning and sequencing of the protein-tyrosine kinase substrate, ezrin, reveals homology to band4.1. *EMBO (Eur. Mol. Biol. Organ.) J.* 8:4133–4142.

Gu, M., J.D. Ydrk, I. Warshawsky, and P.W. Majerus. 1991. Identification, cloning, and expression of a cytosolic megakaryocyte protein-tyrosine-phosphatase with sequence homology to cytoskeletal protein 4.1. *Proc. Natl. Acad. Sci. USA.* 88:5867–5871.

Hall, A. 1990. The cellular functions of small GTP-binding proteins. *Science (Wash. DC)*. 249:635–640.

Hanzel, D., H. Reggio, A. Bretcher, J.G. Forte, and P. Mangeat. 1991. The secretion-stimulated 80K phosphoprotein of a parietal cells is ezrin, and has properties of a membrane cytoskeletal linker in the induced apical microvilli. *EMBO (Eur. Mol. Biol. Organ.) J.* 10:2363–2373.

Hartwig, J.H., G.M. Bokoch, C.L. Carpenter, P.A. Janmey, L.A. Taylor, A. Toker, and T.P. Stossel. 1995. Thrombin receptor ligation and activated Rac uncouple actin filament barbed ends through phosphoinositide synthesis in permeabilized human platelets. *Cell.* 82:643–653.

Haynes, B.F., M.J. Telen, L.P. Hale, and S.M. Denning. 1989. CD44—a molecule involved in leukocyte adherence and T-cell activation. *Immunol. Today.* 10:423–428.

Haynes, B.F., H.-X. Liao, and K.L. Patton. 1991. The transmembrane hyaluronate receptor (CD44): multiple functions, multiple forms. *Cancer Cells (Cold Spring Harbor)*. 3:347–350.

Henry, M.D., C.G. Agosti, and F. Solomon. 1995. Molecular dissection of radixin: Distinct and interdependent functions of the amino- and carboxyl-terminal domains. *J. Cell Biol.* 129:1007–1022.

Horiuchi, H., A. Giner, B. Hoflack, and M. Zerial. 1995. A GDP/GTP exchange-stimulatory activity for the Rab5-RabGDI complex on clathrin-coated vesicles from bovine brain. *J. Biol. Chem.* 270:11257–11262.

Jalkanen, S.T., R.F. Bargatze, L.R. Herron, and E.C. Butcher. 1986. A lymphoid cell surface glycoprotein involved in endothelium recognition and lymphocyte homing in man. *Eur. J. Immunol.* 16:1195–1202.

Janmey, P.A., and T.P. Stossel. 1987. Modulation of gelsolin function by phosphatidylinositol 4, 5-bisphosphate. *Nature (Lond.)*. 325:362–364.

Kikuchi, A., K. Yamamoto, T. Fujita, and Y. Takai. 1988. ADP-ribosylation of the bovine brain Rho protein by botulinum toxin type C1. *J. Biol. Chem.* 263:16303–16308.

Kishi, K., T. Sasaki, S. Kuroda, T. Itoh, and Y. Takai. 1993. Regulation of cytoplasmic division of *Xenopus* embryo by Rhop21 and its inhibitory GDP/GTP exchange protein (RhoGDI). *J. Cell Biol.* 120:1187–1195.

Kozma, R., S. Ahmed, A. Best, and L. Lim. 1995. The Ras-related protein Cdc42Hs and bradykinin promote formation of peripheral actin microspikes and filopodia in Swiss 3T3 fibroblasts. *Mol. Cell Biol.* 15:1942–1952.

Laemmli, U. K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature (Lond.)*. 227:680–685.

Lankes, W., and H. Furthmayr. 1991. Moesin: a member of the protein 4.1-talin-ezrin family of proteins. *Proc. Natl. Acad. Sci. USA.* 88:8297–8301.

Lankes, W., A. Griesmacher, J. Grunwald, R. Schwarts-Albiez, and R. Keller. 1988. A heparin-binding protein involved in inhibition of smooth-muscle cell proliferation. *Biochem. J.* 251:831–842.

Lassing, I., and U. Lindberg. 1985. Specific interaction between phosphatidylinositol 4, 5-bisphosphate and profilin. *Nature (Lond.)*. 314:472–474.

Lesley, J., R. Hyman, and P.W. Kincade. 1993. CD44 and its interaction with extracellular matrix. *Adv. Immunol.* 54:271–335.

Mabuchi, I., Y. Hamaguchi, H. Fujimoto, N. Morii, M. Mishima, and S. Narumiya. 1993. A rho-like protein is involved in the organization of the contractile ring in dividing sand dollar eggs. *Zyote.* 1:325–331.

Malcolm, K.C., A.H. Ross, R.G. Qiu, M. Symons, and J.H. Exton. 1994. Activation of rat liver phospholipase D by the small GTP-binding protein Rho A. *J. Biol. Chem.* 269:25951–25954.

Martin, M., C. Andreoli, A. Sahuquet, P. Montcourrier, M. Algrain, and P. Mangeat. 1995. Ezrin NH<sub>2</sub>-terminal domain inhibits the cell extension activity of the COOH-terminal domain. *J. Cell Biol.* 28:1081–1093.

Nakamura, F., M.R. Amieva, and H. Furthmayr. 1995. Phosphorylation of threonine 558 in the carboxyl-terminal actin-binding domain of moesin by thrombin activation of human platelets. *J. Biol. Chem.* 270:31377–31385.

Nakamura, H., and H. Ozawa. 1996. Immunolocalization of CD44 and ERM

- family in osteoblasts and osteoclasts in mouse tibiae. *J. Bone Miner. Res.* In press.
- Narumiya, S., A. Sekine, and M. Fujiwara. 1988. Substrate for botulinum ADP-ribosyltransferase, Gb, has an amino acid sequence homologous to a putative Rho gene product. *J. Biol. Chem.* 263:17255-17257.
- Niggli, V., C. Andreoli, C. Roy, and P. Mangeat. 1995. Identification of a phosphatidylinositol-4,5-bisphosphate-binding domain in the N-terminal region of ezrin. *FEBS Lett.* 376:172-176.
- Nishiyama, T., T. Sasaki, K. Takaishi, M. Kato, H. Yaku, K. Araki, Y. Matsuura, and Y. Takai. 1994. Rac2p1 is involved in insulin-induced membrane ruffling and Rhop21 is involved in hepatocyte growth factor- and 12-O-tetradecanoyl-phorbol-13-acetate (TPA)-induced membrane ruffling in KB cells. *Mol. Cell. Biol.* 14:2447-2456.
- Nobes, C.D., and A. Hall. 1995. Rho, rac, and cdc42 GTPases regulate the assembly of multimolecular focal complexes associated with actin stress fibers, lamellipodia, and filopodia. *Cell.* 81:53-62.
- Novick, P., and M. D. Garret. 1994. No exchange without receipt. *Nature (Lond.)*. 369:18-19.
- Pakkanen, R., K. Hedman, O. Turunen, T. Wahlstrom, and A. Vaheri. 1987. Microvillus-specific Mr 75,000 plasma membrane protein of human choriocarcinoma cells. *J. Histochem. Cytochem.* 135:809-816.
- Pelham, H.R. and R.J. Jackson. 1976. An efficient mRNA-dependent translation system from reticulocyte lysates. *Eur. J. Biochem.* 67:247-256.
- Pestonjampas, K., M.R. Amieva, C.P. Strassel, W.M. Nauseef, H. Furthmayr, and E.J. Luna. 1995. Moesin, ezrin, and p205 are actin-binding proteins associated with neutrophil plasma membranes. *Mol. Biol. Cell.* 6:247-259.
- Rees, D.J.G., S.E. Ades, S.J. Singer, and R.O. Hynes. 1990. Sequence and domain structure of talin. *Nature (Lond.)*. 347:685-689.
- Ridley, A.J., and A. Hall. 1992. The small GTP-binding protein Rho regulates the assembly of focal adhesions and actin stress fibers in response to growth factors. *Cell.* 70:389-399.
- Ridley, A.J., H.F. Paterson, C.L. Johnston, D. Diekmann, and A. Hall. 1992. The small GTP-binding protein rac regulates growth factor-induced membrane ruffling. *Cell.* 70:401-410.
- Rouleau, G.A., P. Merel, M. Lutchman, M. Sanson, J. Zucman, C. Marineau, K. Hoang-Xuan, S. Demczuk, C. Desmaze, B. Plougastel et al. 1993. Alteration in a new gene encoding a putative membrane-organizing protein causes neuro-fibromatosis type 2. *Nature (Lond.)*. 363:515-521.
- Sato, N., S. Yonemura, T. Obinata, Sa. Tsukita, and Sh. Tsukita. 1991. Radixin, a barbed end-capping actin-modulating protein, is concentrated at the cleavage furrow during cytokinesis. *J. Cell Biol.* 113:321-330.
- Sato, N., N. Funayama, A. Nagafuchi, S. Yonemura, Sa. Tsukita, and Sh. Tsukita. 1992. A gene family consisting of ezrin, radixin, and moesin. Its specific localization at actin filament/plasma membrane association sites. *J. Cell Sci.* 103:131-143.
- Shimizu, K., K. Kaibuchi, H. Nonaka, J. Yamamoto, and Y. Takai. 1991. Tissue and subcellular distribution of an inhibitory GDP/GTP exchange protein (GDI) for the Rho proteins by use of its specific antibody. *Biochem. Biophys. Res. Commun.* 175:199-205.
- Soldati, T., A.D. Shapiro, A.B. Svejstrup, and S.R. Pfeffer. 1994. Membrane targeting of the small GTPase Rab9 is accompanied by nucleotide exchange. *Nature (Lond.)*. 369:18-19.
- Takai, Y., T. Sasaki, K. Tanaka, and H. Nakanishi. 1995. Rho as a regulator of the cytoskeleton. *Trends Biochem. Sci.* 20:227-231.
- Takaishi, K., A. Kikuchi, S. Kuroda, K. Kotani, T. Sasaki, and Y. Takai. 1993. Involvement of Rhop21 and its inhibitory GDP/GTP exchange protein (RhoGDI) in cell motility. *Mol. Cell Biol.* 13:72-79.
- Takaishi, K., T. Sasaki, T. Kameyama, Sa. Tsukita, Sh. Tsukita, and Y. Takai. 1995. Translocation of activated Rho from the cytoplasm to membrane ruffling area, cell-cell adhesion sites and cleavage furrows. *Oncogene.* 11:39-48.
- Takeuchi, K., A. Kawashima, A. Nagafuchi, and Sh. Tsukita. 1994a. Structural diversity of band 4.1 superfamily members. *J. Cell Sci.* 107:1921-1928.
- Takeuchi, K., N. Sato, H. Kasahara, N. Funayama, A. Nagafuchi, S. Yonemura, Sa. Tsukita, and Sh. Tsukita. 1994b. Perturbation of cell adhesion and microvilli formation by antisense oligonucleotides to ERM family members. *J. Cell Biol.* 125:1371-1384.
- Trofatter, J.A., M.M. MacCollin, J.L. Rutter, J.R. Murrell, M.P. Duyao, D.M. Parry, R. Eldridge, N. Kley, A.G. Menon, K. Pulaski et al. 1993. A novel moesin-, ezrin-, radixin-like gene is a candidate for the neurofibromatosis 2 tumor suppressor. *Cell.* 72:791-800.
- Tsukita, Sa., Y. Hieda, and Sh. Tsukita. 1989. A new 82kD-barbed end capping protein localized in the cell-to-cell adherens junction: purification and characterization. *J. Cell Biol.* 108:2369-2382.
- Tsukita, Sa., K. Oishi, N. Sato, J. Sagara, A. Kawai, and Sh. Tsukita. 1994. ERM family members as molecular linkers between the cell surface glycoprotein CD44 and actin-based cytoskeletons. *J. Cell Biol.* 126:391-401.
- Tsukita, Sh. and Sa. Tsukita. 1989. Isolation of cell-to-cell adherens junction from rat liver. *J. Cell Biol.* 108:31-41.
- Tsukita, Sh., M. Itoh, and Sa. Tsukita. 1989. A new 400-kD protein from isolated adherens junctions: its localization at the undercoat of adherens junctions and at microfilament bundles such as stress fibers and circumferential bundles. *J. Cell Biol.* 109:2905-2915.
- Tsukita, Sh., Sa. Tsukita, A. Nagafuchi, and S. Yonemura. 1992. Molecular linkage between cadherins and actin filaments in cell-to-cell adherens junctions. *Curr. Opin. Cell Biol.* 4:834-839.
- Turunen, O., R. Winqvist, R. Pakkanen, K. H. Grzeschik, T. Wahlstrom, and A. Vaheri. 1989. Cytovillin, a microvillar Mr 75,000 protein. cDNA sequence, pro-karyotic expression, and chromosomal localization. *J. Biol. Chem.* 264:16727-16732.
- Turunen, O., T. Wahlstrom, and A. Vaheri. 1994. Ezrin has a COOH-terminal actin-binding site that is conserved in the ezrin protein family. *J. Cell Biol.* 126:1445-1453.
- Ullrich, O., H. Horiuchi, C. Bucci, and M. Zerial. 1994. Membrane association of Rab5 mediated by GDP-dissociation inhibitor and accompanied by GDP/GTP exchange. *Nature (Lond.)*. 368:157-160.
- Urushidani, T., D.K. Hanzel, and J.G. Forte. 1989. *Am. J. Physiol.* 256:G1070-G1081.
- Vandekerckhove, J. 1990. Actin-binding proteins. *Curr. Opin. Cell Biol.* 2:41-51.
- Watanabe, G., Y. Saito, P. Madaule, T. Ishizaki, K. Fujisawa, N. Morii, H. Mukai, Y. Ono, A. Kakizuka, and S. Narumiya. 1996. Protein kinase N (PKN) and PKN-related protein rhophilin as targets of small GTPase Rho. *Science (Wash. DC)*. 271:645-648.
- Wayner, E.A., W.G. Carter, R.S. Piotrowicz, and T.J. Kunicki. 1988. The function of multiple extracellular matrix receptors in mediating cell adhesion to extracellular matrix: Preparation of monoclonal antibodies to the fibronectin receptor that specifically inhibit cell adhesion to fibronectin and react with platelet glycoproteins Ic-IIa. *J. Cell Biol.* 107:1881-1891.
- Yang, Q., and N.K. Tonks. 1991. Isolation of a cDNA clone encoding a human protein-tyrosine phosphatase with homology to the cytoskeletal-associated proteins band 4.1, ezrin, and talin. *Proc. Natl. Acad. Sci. USA.* 88:5949-5953.
- Yonemura, S., A. Nagafuchi, N. Sato, and Sh. Tsukita. 1993. Concentration of an integral membrane protein, CD43 (leukosialin, sialophorin), in the cleavage furrow through the interaction of its cytoplasmic domain with actin-based cytoskeletons. *J. Cell Biol.* 120:437-449.
- Yonezawa, N., Y. Homma, I. Yahara, H. Sakai, and E. Nishida. 1991. A short sequence responsible for both phosphoinositide binding and actin binding activities of cofilin. *J. Biol. Chem.* 266:17218-17221.
- Yu, F.X., P.A. Johnston, T.C. Sudhof, and H.L. Yin. 1990. gCap39, a calcium ion- and polyphosphoinositide-regulated actin capping protein. *Science (Wash. DC)*. 250:1413-1415.
- Zhang, J., W.G. King, S. Dillon, A. Hall, L. Feig, and S.E. Rittenhouse. 1995. Activation of platelet phosphatidylinositol 3-kinase requires the small GTP-binding protein Rho. *J. Biol. Chem.* 268:22251-22254.
- Zhou, D.F., J.F. Ding, L.J. Picker, R.F. Bargatzke, E.C. Butcher, and D.V. Goeddel. 1989. Molecular cloning and expression of Pgp-1. The mouse homolog of the human H-CAM (hermes) lymphocyte homing receptor. *J. Immunol.* 143:3390-3395.
- Zigmond, S.H. 1995. Signal transduction and actin filament organization. *Curr. Opin. Cell Biol.* 8:66-73.