

Regulation of InsP₃ receptor activity by neuronal Ca²⁺-binding proteins

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Inositol 1,4,5-trisphosphate receptors (InsP₃Rs) were recently demonstrated to be activated independently of InsP₃ by a family of calmodulin (CaM)-like neuronal Ca²⁺-binding proteins (CaBPs). We investigated the interaction of both naturally occurring long and short CaBP1 isoforms with InsP₃Rs, and their functional effects on InsP₃R-evoked Ca²⁺ signals. Using several experimental paradigms, including transient expression in COS cells, acute injection of recombinant protein into *Xenopus* oocytes and ⁴⁵Ca²⁺ flux from permeabilised COS cells, we demonstrated that CaBPs decrease the sensitivity of InsP₃-induced Ca²⁺ release (IICR). In addition, we found a Ca²⁺-independent interaction between CaBP1 and the NH₂-terminal 159 amino acids of the type 1 InsP₃R. This interaction resulted in decreased InsP₃ binding to the receptor reminiscent of that observed for CaM. Unlike CaM, however, CaBPs do not inhibit ryanodine receptors, have a higher affinity for InsP₃Rs and more potently inhibited IICR. We also show that phosphorylation of CaBP1 at a casein kinase 2 consensus site regulates its inhibition of IICR. Our data suggest that CaBPs are endogenous regulators of InsP₃Rs tuning the sensitivity of cells to InsP₃.

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Abbreviations: InsP₃, inositol 1,4,5-trisphosphate; InsP₃R, inositol 1,4,5-trisphosphate receptor; CaBP, Ca²⁺-binding protein; CaM, calmodulin; IICR, InsP₃-induced Ca²⁺ release; ER, endoplasmic reticulum; RyR, ryanodine receptor; IP, immunoprecipitation; co-IP, co-immunoprecipitation; aa, amino acids

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Introduction

Changes in intracellular Ca²⁺ control a diverse array of cellular processes, including fertilisation, memory formation and gene transcription (Berridge *et al*, 2003). Increases in cytosolic Ca²⁺ levels may arise as a result of either its release from the endoplasmic reticulum (ER) intracellular Ca²⁺ store or influx across the plasma membrane. The principal Ca²⁺ release channels on the ER belong to the ryanodine receptor (RyR) and inositol 1,4,5-trisphosphate receptor (InsP₃R) families (Ehrlich, 1995; Iino, 1999).

Both families of channels are biphasically regulated by Ca²⁺ (Bezprozvanny *et al*, 1991). In addition, inositol 1,4,5-trisphosphate (InsP₃) is a co-agonist for InsP₃Rs (Bezprozvanny *et al*, 1991). The mechanism by which Ca²⁺ exerts its effect has been the subject of intense investigation resulting in the identification of several Ca²⁺ binding sites in the InsP₃R (Sienaert *et al*, 1997). In addition to a direct action of Ca²⁺ on the receptor, Ca²⁺ may function through Ca²⁺-sensing proteins that either bind directly to or modify channel activity by the addition/removal of secondary modifications such as phosphate groups (Patel *et al*, 1999; Thrower *et al*, 2001; Nadif Kasri *et al*, 2002; Taylor and Laude, 2002; Roderick and Bootman, 2003). The tetra EF-hand-containing Ca²⁺ sensor protein calmodulin (CaM) has been shown to modulate the activity of InsP₃Rs in both a Ca²⁺-dependent and -independent manner (Reviewed in Nadif Kasri *et al*, 2002; Taylor and Laude, 2002). The Ca²⁺-independent CaM binding domain has been mapped to two noncontiguous sites in the NH₂-terminal 159 amino acids (aa) of the type 1 InsP₃R (InsP₃R1) (Sienaert *et al*, 2002). The interaction of CaM with these sites was shown to decrease the affinity of the receptor for InsP₃ (Sienaert *et al*, 2002). The inhibition of InsP₃-induced Ca²⁺ release (IICR) by CaM-like proteins may have great significance in neurons where it has been shown to underlie processes including axonal guidance, growth cone formation as well as excitability, which govern long-term potentiation and depression (LTD) associated with memory formation (Nakamura *et al*, 1999; Nagase *et al*, 2003). The propagation of Ca²⁺ release following repetitive stimulation of the parallel fibre/Purkinje cell synapses is generally restricted to the postsynaptic spines. This is surprising, since Purkinje cells abundantly express InsP₃Rs throughout the cell (Snyder and Supattapone, 1989; Volpe *et al*, 1991). Thus, it has been proposed that the high levels of CaM also present in these cells (~10 μM; Kakiuchi *et al*, 1982) decrease IICR and restrict excitability (Cardy and Taylor, 1998).

A family of neuronal Ca²⁺-binding proteins (CaBPs), sharing significant homology in sequence and structure with CaM, has recently been described in the retina and brain (Haeseleer *et al*, 2000). This family of proteins belongs to the superfamily of neuronal Ca²⁺ sensor proteins that also include NCS1 (or frequenin) and KChiPs, which have been shown to be involved in neuronal signalling (Burgoyne and

Weiss, 2001). Unlike CaM, one or two of the EF hand Ca^{2+} -binding domains in CaBPs are nonfunctional (Haeseleer *et al*, 2000). CaBPs have been shown to bind to the NH_2 -terminus of InsP_3 Rs in a Ca^{2+} -dependent manner. Furthermore, it was reported that CaBP1 activated Ca^{2+} release through InsP_3 Rs independent of InsP_3 binding (Yang *et al*, 2002). Thus, if CaBPs act as endogenous surrogates for InsP_3 , it is possible that InsP_3 Rs may be directly regulated by Ca^{2+} , allowing them to act as true Ca^{2+} -induced Ca^{2+} release channels in a manner similar to RyRs (Bootman *et al*, 2002).

In this study, we rigorously tested the functional relationship between CaBP- and InsP_3 -mediated Ca^{2+} release. We determined that CaBP1 interacted with InsP_3 Rs in a Ca^{2+} -independent manner. This interaction was localised to a peptide sequence in the NH_2 -terminal 159 aa of InsP_3 R1 to which CaM has also been shown to bind. CaBP1 binding resulted in a decrease in the sensitivity of IICR. This was demonstrated by measuring IICR in COS cells overexpressing CaBP1, in *Xenopus* oocytes microinjected with recombinant CaBP1 and permeabilised COS cells exposed to recombinant CaBP1. In addition, CaBP1 by itself could not activate Ca^{2+} release. We also show that CaBP1 is phosphorylated at a casein kinase 2 consensus sequence, which regulates its efficacy to inhibit IICR. The use of COS cells and *Xenopus* oocytes, which do not express endogenous CaBP1, provided an ideal null background to investigate the function of CaBP1. Our data suggest that CaBP1 can behave as an endogenous regulator of InsP_3 R activity, and may serve to tune the sensitivity of InsP_3 Rs to InsP_3 . Although structurally similar to CaM, CaBPs have distinct effects and provide an additional facet of InsP_3 R regulation.

Results

CaBP1 inhibits agonist-induced Ca^{2+} signals

CaBP1 has previously been reported to increase the open probability of InsP_3 R independently of InsP_3 in nuclei isolated from *Xenopus* oocytes (Yang *et al*, 2002). To test whether this was also true in an intact cellular system, we first adopted a heterologous expression approach. Expression vectors for both the naturally occurring long and short CaBP1 isoforms were generated. In addition, to identify cells expressing these constructs, YFP was appended to the COOH-terminus of the CaBPs (Figure 1A). Figure 1B shows that when overexpressed, both S/CaBP1-YFP and L/CaBP1-YFP are targeted primarily to the plasma membrane, with some of the protein also residing on intracellular membranes, including the Golgi apparatus. A weaker, but significant, fluorescence was also observed in the cytosol. Transiently expressed S/CaBP1-YFP and L/CaBP1-YFP, detected using an anti-GFP antibody, migrated at the expected molecular mass (Figure 1C, top panel). Equivalent levels of expression of the endogenous type 3 InsP_3 R (InsP_3 R3) were detected (the major InsP_3 R isoform in COS-7 cells; Wojcikiewicz, 1995) in lysates from FACS-isolated CaBP1-expressing cells, indicating that CaBP1 expression did not impact on the levels of endogenous InsP_3 Rs (Figure 1C, bottom panel).

At 48 h post-transfection, ATP-evoked Ca^{2+} transients were monitored in Fura-2-loaded cells. Expression of either L/CaBP1 or S/CaBP1 resulted in a significant decrease in the number of responsive cells at both 0.5 and 1 μM ATP, and a decrease in the occurrence of agonist-evoked oscillations in

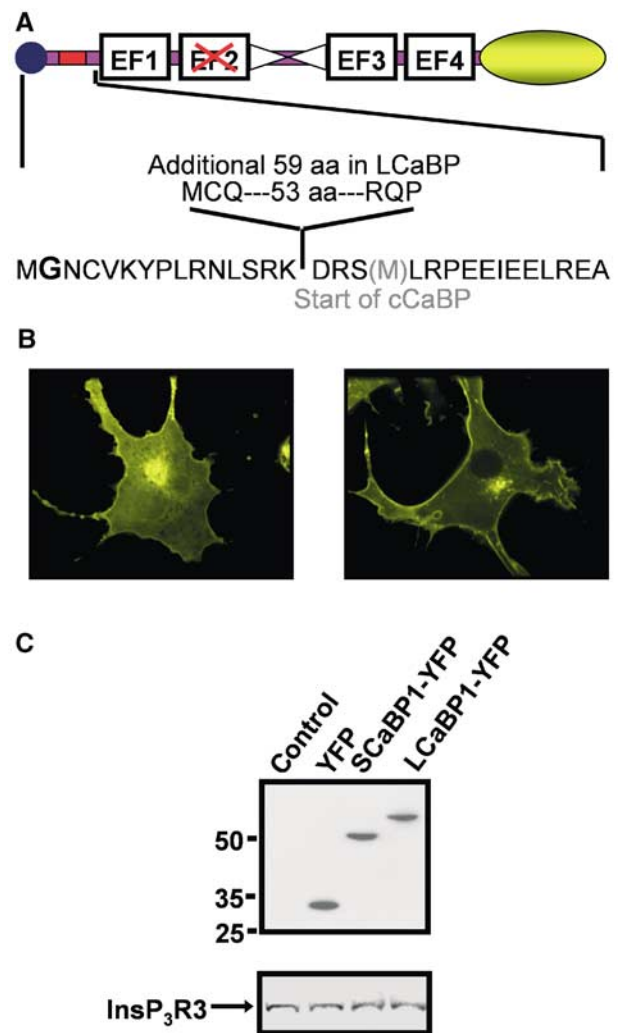


Figure 1 Design and expression of CaBP-YFP vectors. (A) Graphical representation of the L/CaBP1-YFP and S/CaBP1-YFP constructs used. The location of the additional aa in L/CaBP1 compared to S/CaBP1 is indicated by the red box. The central α -helix is indicated by the two triangles and the myristoylation sequence is represented by a circle. The four EF hand Ca^{2+} -binding domains are indicated and the nonfunctional EF hand is shown by the red cross. The yellow ellipse illustrates the COOH-terminal YFP. In the lower panel, aa boundaries for both S/LCaBP and truncated cCaBP (additional methionine at the start of cCaBP shown in grey) are shown. The myristoylated glycine is in bold. (B) COS-7 cells were transiently transfected with S/LCaBP-YFP constructs. At 48 h post-transfection, cells were imaged confocally through a $\times 100$, 1.4 n.a. oil immersion objective (Perkin-Elmer, Cambridge UK). The 488 nm line of the krypton/argon laser was used for excitation and emission collected > 505 nm. (C) (Upper panel) Mobility of YFP, S/CaBP1-YFP and L/CaBP1-YFP was determined in transiently transfected COS-7 cells by Western blot using anti-GFP antibody. (Lower panel) Abundance of InsP_3 R3 was determined in cells transfected with YFP, S/CaBP1, and L/CaBP1 isolated by FACS.

those cells that responded (Figure 2A and B). In addition, the integrated Ca^{2+} response at these agonist concentrations was reduced (Figure 2D). The integrated Ca^{2+} response to maximal agonist (100 μM ATP) was not significantly affected by CaBP1 expression, indicating that the Ca^{2+} content of the agonist-sensitive store was unchanged (Figure 2D). Although the amplitude and integrated Ca^{2+} response to maximal ATP were not altered by CaBP expression, the latency between agonist addition and the peak response was significantly

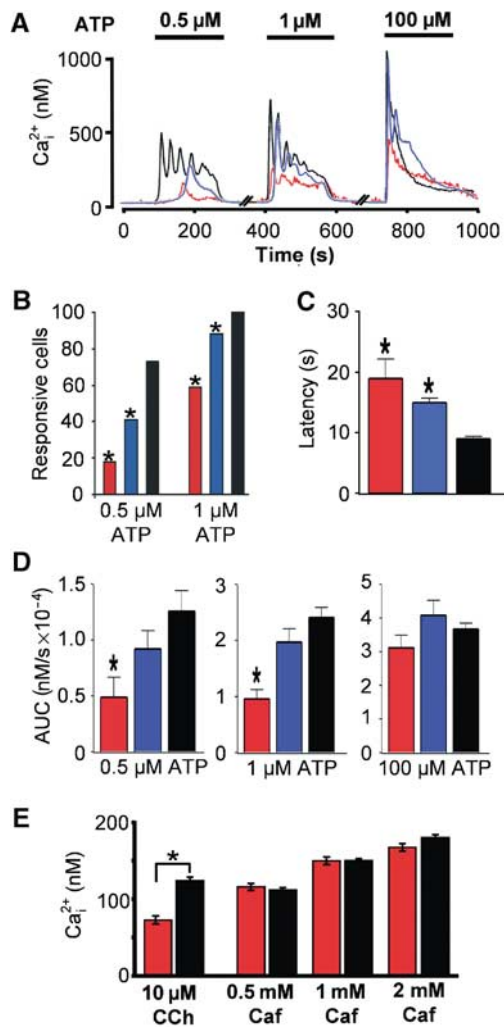


Figure 2 CaBP1 inhibits agonist-induced Ca^{2+} release. Responses to the InsP_3 -generating agonist ATP were examined in COS-7 cells transfected with either SCaBP1 (red trace and bars) or LCaBP1 (blue trace and bars), respectively. (A) Responses from transfected and nontransfected (control, black trace) cells were recorded from the same field of view. The concentrations of ATP used are shown by the solid bars above the traces. (B) Quantitative analysis of the experiments depicted in (A). (C) Latency between the application of agonist and the peak Ca^{2+} response in cells stimulated with 100 μM ATP. (D) Integrated Ca^{2+} signal observed with 0.5, 1 and 100 μM ATP. (E) Caffeine-induced Ca^{2+} release in HEK293 cells stably expressing RyR3 transiently transfected with SCaBP1-YFP. The inhibition of IICR in CaBP-expressing cells was confirmed by application of the InsP_3 -generating agonist carbachol (10 μM CCh, red bar). The peak amplitudes of the Ca^{2+} transients for control (black bars) and SCaBP1-YFP (red bars) induced by each caffeine concentration applied are indicated in the bar graph. Statistical significance is denoted by an asterisk.

prolonged. Therefore, even in the face of maximal InsP_3 increases, CaBP effected a delay in the development of the Ca^{2+} transient, consistent with a decrease in the sensitivity of InsP_3 Rs (Figure 2C). Similar results were obtained when SCaBP1 was expressed in HeLa, SH-SY5Y and HEK293 cells, indicating that CaBP1 could act on IICR from a variety of cell types expressing differing InsP_3 R isoform complements (data not shown). No effect on IICR was observed following expression of EYFP alone (data not shown). Since both

LCaBP1 and SCaBP1 were inhibitory to Ca^{2+} release, an NH_2 -terminally truncated cDNA construct spanning the conserved regions of these two isoforms (cCaBP) was expressed (Figure 1A). cCaBP resulted in a significant decrease in the percentage of responding cells at 0.5 μM ATP from $63.9 \pm 2.9\%$ in control cells to $26.4 \pm 3.7\%$ in cCaBP-expressing cells ($n = 11$, $P < 0.05$), which is comparable to that observed for long and short CaBP (Figure 2B). The specificity of the effect of CaBP1 upon IICR was further demonstrated by the absence of any inhibition of caffeine-induced Ca^{2+} release through RyRs in HEK293 cells in which they are stably expressed (Rossi *et al*, 2002) (Figure 2E). Carbachol-induced Ca^{2+} release measured in the same cells overexpressing CaBP1 was however inhibited, suggesting that InsP_3 Rs were specifically targeted.

To rule out that the effect of CaBP1 on IICR was mediated by Ca^{2+} buffering, the EF-hand-containing Ca^{2+} buffer protein calbindin was overexpressed and its effects on Ca^{2+} signals were examined. Unlike CaBP, calbindin overexpression did not significantly reduce the number of cells responding to ATP, even at the lowest agonist concentrations used (Supplementary Figure 1). Furthermore, overexpression of CaBP1 in which the three EF hands were disabled (CaBP1₁₃₄) (Yang *et al*, 2002) resulted in a similar level of inhibition of IICR as cells expressing wild-type CaBP1 ($13 \pm 6\%$ of cells expressing CaBP1₁₃₄ versus $17.6 \pm 4.5\%$ of cells for SCaBP1 responded to 0.5 μM ATP, $P > 0.05$). Thus, we conclude that the inhibition of IICR by CaBP1 is Ca^{2+} independent.

CaBP1 directly targets InsP_3 Rs to inhibit IICR

To test that the inhibition of IICR was due to a direct effect on InsP_3 Rs, we employed a number of strategies. Application of a cell-permeant InsP_3 ester (InsP_3BM) (Thomas *et al*, 2000) induced a robust Ca^{2+} transient preceded by Ca^{2+} oscillations in control COS-7 cells, which was not observed in CaBP1-overexpressing cells (Figure 3A). Confocal microscopy was also used to monitor the effect of CaBP on IICR in Fluo-4-loaded *Xenopus* oocytes. The oocytes were injected with either recombinant CaBP1 (8.5 μM final) or vehicle 30 min prior to imaging. In control oocytes, injection of 40 nM F- InsP_3 resulted in an accumulating increase in cytosolic Ca^{2+} levels punctuated by Ca^{2+} puffs (Figure 3B), which was not observed in the majority of oocytes injected with CaBP1 (Figure 3C). At 100 nM F- InsP_3 , however, Ca^{2+} release was observed in CaBP1-injected oocytes, although puffs prior to the Ca^{2+} tide were not apparent (data not shown). Furthermore, no calcium release was observed in oocytes imaged simultaneously with injection of recombinant CaBP1 alone (Supplementary Figure 2). These data indicated that CaBP1 did not induce Ca^{2+} release, did not irreversibly inhibit InsP_3 Rs, but significantly reduced the sensitivity of IICR.

In previous studies we have shown the affinity of the Ca^{2+} -independent CaM binding site on InsP_3 R1 to be 2 μM (Sienaert *et al*, 2002). In this study, based on the amount of excess peptide required to prevent migration of CaBP into gels (depicted in Figure 4C) compared to CaM under the same conditions, we conclude that CaBP has a three-fold greater affinity for the InsP_3 R than CaM. Thus, to investigate the effects of CaBP on Ca^{2+} release, we used concentrations of the protein up to one order of magnitude greater than its K_d . To quantitate specifically the unidirectional flux of Ca^{2+} , we

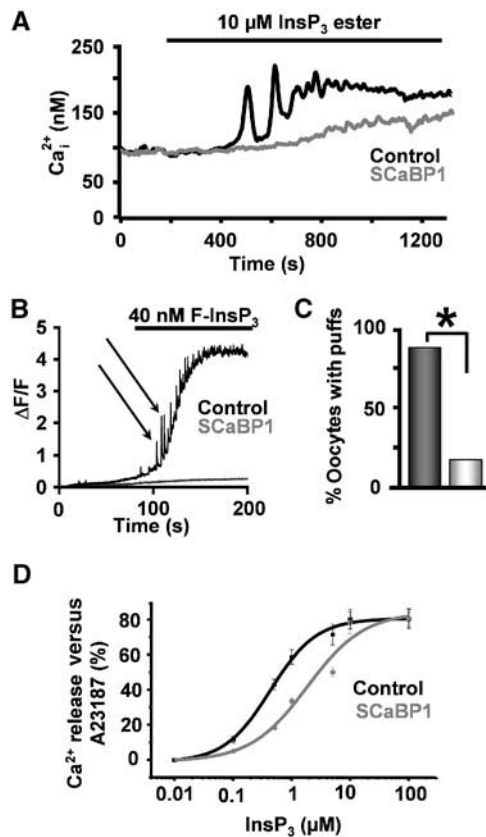


Figure 3 CaBP1 inhibits Ca^{2+} release induced by direct application of InsP_3 . (A) Ca^{2+} release following application of $10\ \mu\text{M}$ cell-permeant InsP_3 (InsP_3BM) observed in control (black trace) and CaBP1-transfected (grey trace) COS-7 cells. (B, C) F- InsP_3 -induced Ca^{2+} release in control oocytes (black trace) and oocytes preinjected with $8.5\ \mu\text{M}$ recombinant CaBP1 (grey trace). The black arrows in (B) indicate Ca^{2+} puffs. Oocytes were imaged as described in Materials and methods. (D) Effect of CaBP1 on InsP_3 -induced efflux of $^{45}\text{Ca}^{2+}$ from COS cells. After loading permeabilised COS-1 cells for 45 min in $150\ \text{nM}$ $^{45}\text{Ca}^{2+}$, efflux was started in a medium containing $1\ \mu\text{M}$ free Ca^{2+} , with (circles) or without (squares) $10\ \mu\text{M}$ SCaBP1. After 10 min of efflux, cells were challenged with InsP_3 for 2 min. The increase in fractional loss provoked by the InsP_3 challenge was plotted as a function of the applied InsP_3 concentration. The Ca^{2+} release was normalized to the total releasable fraction by $5\ \mu\text{M}$ A23187, which was taken as 100%. The results represent the mean \pm s.e.m. of three independent experiments, each performed in duplicate.

measured InsP_3 -induced $^{45}\text{Ca}^{2+}$ flux from permeabilised COS cells in the presence and absence of $10\ \mu\text{M}$ His-tagged CaBP1. We found that CaBP1 significantly increased the IC_{50} of IICR from 0.41 ± 0.13 to $1.83 \pm 0.21\ \mu\text{M}$ ($P < 0.005$) (Figure 3D).

Interaction between CaBP1 and the NH_2 -terminal region of $\text{InsP}_3\text{R1}$

To map the location of the CaBP-binding region within the NH_2 -terminus of $\text{InsP}_3\text{R1}$, pull-down experiments using GST fusions of different NH_2 -terminal domains of the $\text{InsP}_3\text{R1}$ were performed (Figure 4A). GST-(1–604) and GST-(1–225) but not GST-(226–604) or the GST control retained the purified SCaBP1. Similar results were obtained using the LCaBP-1 variant or proteins expressed without the HIS tag (data not shown). To identify the site(s) where CaBP1 interacts within the first 225 aa of the InsP_3R , we performed band-

shift experiments using a series of synthetic peptides spanning this region (Sienaert *et al*, 2002) (Figure 4Bi). A 15% nondenaturing gel illustrating the migration of SCaBP1 into the gel following incubation with a 10-fold molar excess of each of the five different peptides in either $200\ \mu\text{M}$ free Ca^{2+} or $1\ \text{mM}$ EGTA is shown (Figure 4Bii). The InsP_3R peptides alone did not enter the gel because they were positively charged. Figure 4Bii shows that CaBP1 binds to peptide B independently of Ca^{2+} , resulting in the formation of a complex that does not enter the gel. In a similar assay, CaM and apoCaM also interacted with peptide B (Sienaert *et al*, 2002), suggesting a common binding site for CaM and CaBP1. CaM and apoCaM, however, also interacted with peptide E (Sienaert *et al*, 2002). Since the peptide B/CaM interaction was Ca^{2+} independent (Sienaert *et al*, 2002), the Ca^{2+} dependence of the interaction of peptide B with SCaBP1 was investigated further. Band-shift analysis of this interaction in the presence of an increasing SCaBP1:peptide molar ratio revealed that the interaction was equally effective in the presence of $200\ \mu\text{M}$ Ca^{2+} or $1\ \text{mM}$ EGTA, and was thus Ca^{2+} independent (Figure 4C). Since our overexpression studies were carried out in COS-7 cells, we determined whether CaBP1 interacted with $\text{InsP}_3\text{R3}$. By co-immunoprecipitation (co-IP), we demonstrated that both CaBP1 isoforms interact with $\text{InsP}_3\text{R3}$ (Figure 4D). No co-IP of $\text{InsP}_3\text{R3}$ was observed from lysates of control cells or from cells expressing YFP alone. Furthermore, the interaction between CaBP1 and InsP_3Rs determined by co-IP was Ca^{2+} independent (Figure 4E). In parallel studies, co-IP of YFP-CaM with $\text{InsP}_3\text{R3}$ from transfected cells was more difficult to obtain (data not shown). The efficient co-IP of CaBP with $\text{InsP}_3\text{R1}$ from the brain demonstrates the physiological relevance of the CaBP- InsP_3R interaction (Figure 4F).

CaBP1 inhibits InsP_3 binding to InsP_3Rs

We next investigated the effects of CaBP1 on InsP_3 association with the ligand-binding domain of $\text{InsP}_3\text{R1}$ ($\text{InsP}_3\text{R1}$ aa 1–581, Lbs-1) (Sienaert *et al*, 2002). The presence of $5\ \mu\text{M}$ Ca^{2+} inhibited [^3H]- InsP_3 binding to Lbs-1 by $22 \pm 1.9\%$ as previously described ($n = 3$, $P < 0.05$) (Sienaert *et al*, 2002). SCaBP1 ($10\ \mu\text{M}$) inhibited InsP_3 binding to Lbs-1 by $36 \pm 5\%$ in the absence of Ca^{2+} . Inhibition by SCaBP1 was fully additive to inhibition by Ca^{2+} , and amounted to $58 \pm 4\%$ in the presence of Ca^{2+} ($n = 3$, $P < 0.05$). The inhibition of InsP_3 binding to Lbs-1 observed for SCaBP1 was comparable to that by CaM (K_i for CaBP $1\ \mu\text{M}$ compared with K_i of CaM $2\ \mu\text{M}$, data not shown). To exclude the possibility that the observed Ca^{2+} -independent SCaBP1 effect was due to a constitutively occupied Ca^{2+} -binding site with high affinity in the SCaBP1 protein, we used the EF hand mutant SCaBP1 $_{134}$. This non- Ca^{2+} -binding mutant ($10\ \mu\text{M}$) inhibited InsP_3 association with Lbs-1 by $33 \pm 4\%$ in the absence of Ca^{2+} and $49 \pm 5\%$ in the presence of Ca^{2+} . Similar results were obtained for full-length $\text{InsP}_3\text{R1}$ (data not shown). SCaBP1 had no effect on InsP_3 binding to Lbs-1 $\Delta 1$ –225 (data not shown). Taken together, these results confirm that SCaBP1, like CaM, inhibits InsP_3 binding to the InsP_3R in a Ca^{2+} -independent manner.

CaBP1 is regulated by phosphorylation

Caldendrin, the NH_2 -terminal extended isoform of CaBP1, is phosphorylated, although the site of phosphorylation and its

function are not known (Seidenbecher *et al*, 1998). Using the PROSITE Database of protein families and domains (Sigrist *et al*, 2002), we identified a unique consensus site for phosphorylation by casein kinase 2 in SCaBP1 (S120), which is located at the COOH-terminus of its α -helical region. The functional consequence of phosphorylation of the structurally equivalent site in CaM (S101) is a decrease in its affinity for substrates (Quadroni *et al*, 1998). To test whether this was the case for CaBP1, the S120 residue was mutated to an alanine (S120A). By immunoprecipitation (IP) of SCaBP1-YFP from cells incubated with ^{32}P -orthophosphate, we found that wild-type SCaBP1-YFP was phosphorylated, whereas phosphorylation of SCaBP1-S120A-YFP was barely detectable (Figure 5Ai). Equivalent levels of SCaBP1-YFP and SCaBP1-S120A-YFP in the input lysate were determined by Western blotting (Figure 5Aii) and by FACS analysis of SCaBP1-YFP-expressing cells (data not shown). When stimulated with ATP, SCaBP1-S120A-expressing cells displayed a significantly lower percentage of responding cells, lesser integrated Ca^{2+} signals and lower peak Ca^{2+} transient amplitude in comparison to control cells (Figure 5B-E). Furthermore, the degree of inhibition was significantly greater than that observed for cells expressing wild-type SCaBP1-YFP ($P < 0.05$). No significant difference was observed in the amplitude of ionomycin-induced Ca^{2+} release between CaBP-S120A-expressing cells ($n = 14$) and controls ($n = 43$, $P > 0.5$), indicating that the ER calcium stores were intact (Figure 5F). As shown for the wild-type CaBP (Figure 3A), the SCaBP-S120A mutant also inhibi-

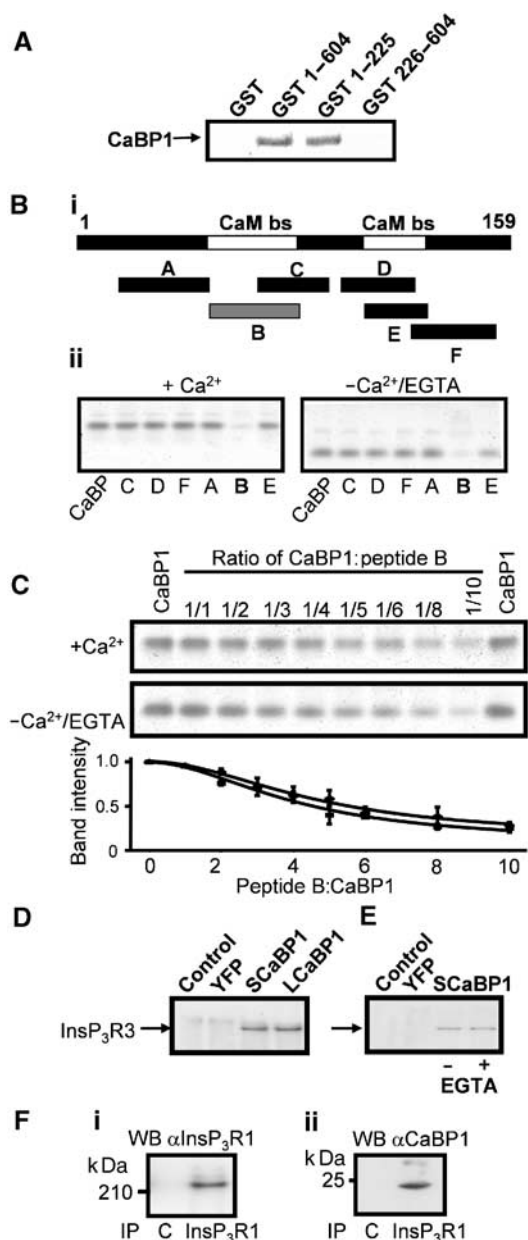
ted InsP_3 ester-induced Ca^{2+} release, indicating a direct effect on InsP_3Rs (Figure 5G and H).

Discussion

The neuronal CaBPs are a novel group of InsP_3R -regulating proteins. The results presented here demonstrate a clear functional interaction between CaBP1 and the InsP_3R Ca^{2+} release channel. Furthermore, we find that CaBP1 activity is regulated by phosphorylation. The consequences of InsP_3R inhibition by CaBP1 may have great significance for neuronal Ca^{2+} signalling, impacting on synaptic plasticity and neuronal growth.

In this study, we characterised a cellular function of CaBP1. Our findings contrast with a previous report using an *in vitro* assay, which suggested that CaBP1 stimulated Ca^{2+} release independent of InsP_3 (Yang *et al*, 2002). We found that both the long and short naturally occurring

Figure 4 CaBP1 binds directly to the InsP_3R . (A) SDS-PAGE gel, stained with SYPRO OrangeTM, showing the interaction between either 100 μg pGST, GST 1-604, GST 1-225, GST 226-604 or 100 μg SCaBP1 in a pull-down assay in the presence of 200 μM free Ca^{2+} . In lanes 2 and 3, SCaBP1 was retained by GST 1-604 and GST 1-225, respectively. (Bi) Map showing positions of synthetic peptides (A-F) used for binding experiments relative to the NH₂-terminal 159 aa region of the InsP_3R . Peptide A (S16-N48), peptide B (P49-N81), peptide C (Y66-K91), peptide D (D97-L123), peptide E (E106-S128) and peptide F (I121-L151). White bars indicating the Ca^{2+} -independent CaM binding sites on the InsP_3R 1 (Sienaert *et al*, 2002). Grey bar (B) showing a positive interaction of the peptide with SCaBP1. (Bii) Nondenaturing gel (15%) of 3.5 μM SCaBP1 in the presence of each of the InsP_3R 1 peptides (A-F) (35 μM) in 50 mM Tris buffer (pH 7.4) containing either 200 μM free Ca^{2+} or 1 mM EGTA stained with SYPRO OrangeTM. In lane 1, SCaBP1 alone was used as a control. SCaBP1 bound to the peptide diminishes the intensity of the SCaBP1 band. (C) SDS-PAGE gel showing the interaction of SCaBP1 with increasing amounts of peptide B (P49-N81) in the presence of 200 μM Ca^{2+} or 1 mM EGTA stained with SYPRO OrangeTM. (Lower panel) Densitometric analysis of the SCaBP1 bands by ImageQuant 5.2 of the interaction between SCaBP1 and peptide B (P49-N81) in the presence of 200 μM Ca^{2+} (triangles) or 1 mM EGTA (squares). The vertical axis denotes the intensity of the SCaBP1 band. (D) co-IP of InsP_3R 3 from COS-7 cells transfected with either SCaBP1 or LCaBP1-YFP. YFP-CaBP1 was immunoprecipitated with anti-GFP antibody. InsP_3R 3 immunoreactivity was detected using an anti- InsP_3R 3 monoclonal antibody and subsequent ECL. Transfected cDNA is indicated above the blot. (E) co-IP of InsP_3R 3 with SCaBP1-YFP was performed in the absence (lane 3) or presence of 1 mM EGTA (lane 4). Control IPs from YFP-transfected cells and nontransfected cells were also performed (lanes 1 and 2). (F) co-IP of CaBP with InsP_3R 1 from the brain. The antibody used for the IP is indicated below the blot and the antibody used for the Western blot (WB) is indicated above the blot. C indicates IP with a non-specific antibody. (Fi) IP of InsP_3R 1 subsequently detected by Western blot with anti- InsP_3R 1 antibody. (Fii) Parallel IP using an anti- InsP_3R 1 antibody and Western blot using an anti-CaBP antibody.



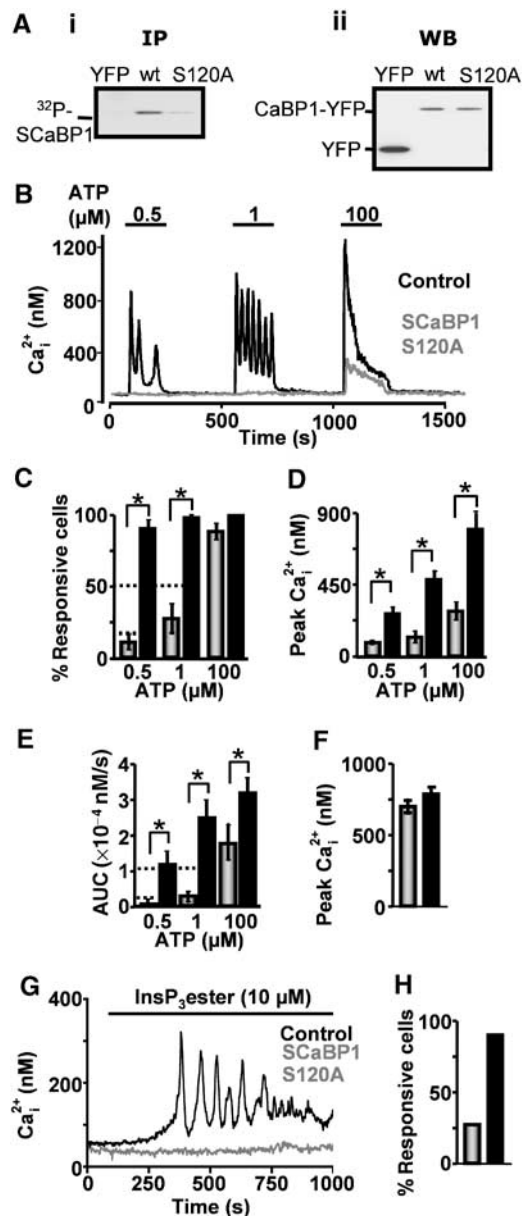


Figure 5 Phosphorylation of S120 regulates CaBP1 activity. **(Ai)** IP using an anti-GFP antibody of SCaBP1-YFP and SCaBP1-S120A-YFP from ^{32}P -labelled cells. Control IPs of YFP-transfected cells are also shown. **(Aii)** Equivalent levels of protein in the input lysate are shown in the Western blot using an anti-GFP antibody. The content of each lane is indicated in the figure. **(B)** Ca^{2+} imaging of control cells (black trace) and cells expressing SCaBP1-S120A-YFP (grey trace). **(C)** Number of responsive cells. **(D)** Peak amplitude of responses. **(E)** Integrated Ca^{2+} response (AUC). Control and CaBP-transfected cells are represented by black and grey bars, respectively. **(F)** Peak amplitude of Ca^{2+} release following ionomycin treatment ($10\mu\text{M}$) for control cells (black bars) and SCaBP-S120A-YFP-transfected cells (grey bars). **(G)** Ca^{2+} release following stimulation with $10\mu\text{M}$ InsP_3 ester for control (black trace) and SCaBP-S120A-YFP-transfected cells (grey trace). **(H)** Percentage of control (black bars) and SCaBP-S120A-YFP-transfected cells (grey bars) responding to InsP_3 ester stimulation is shown in the bar graph. Statistical significance is indicated by an asterisk.

isoforms of CaBP1 as well as the NH_2 -terminally truncated cCaBP inhibited Ca^{2+} release induced by application of an InsP_3 -generating agonist when expressed in COS-7 cells. Since CaBP1 also inhibited Ca^{2+} release induced by applica-

tion of cell-permeant InsP_3 in transfected COS-7 cells, IICR from permeabilised COS cells and from intact *Xenopus* oocytes, we concluded that CaBP1 was directly targeting InsP_3 Rs. The effects of CaBP1 were not due to Ca^{2+} buffering, since CaBP1₁₃₄, in which the three functional EF hands had been disabled, had a similar effect as the wild-type protein. Furthermore, the effects of CaBP1 on Ca^{2+} signalling were unlike those observed for calbindin, another EF-hand-containing protein that functions solely as a Ca^{2+} buffer (John *et al*, 2001). In addition, CaBP1 expression did not affect caffeine-induced Ca^{2+} release through RyRs. As well as showing specificity for IICR, these data support our conclusions that CaBP1 is not causing an artefactual adaptive response in overexpressing cells. The lack of an effect on RyRs is not surprising, since the affinity of the interaction between CaBP1 and RyRs, unlike that between CaM and RyRs, is very low (S Hamilton, personal communication). Our band-shift and co-IP data, together with the data of Yang *et al* (2002), which demonstrated that CaM was unable to displace CaBP1 from InsP_3 Rs, suggest that the affinity of the CaBP1- InsP_3 R interaction is greater than that between CaM and InsP_3 Rs. We also found that when overexpressed in COS-7 cells, CaM did not inhibit IICR to the same degree as CaBP1 (MD Bootman and HL Roderick, unpublished observations). Thus, in neurons that express InsP_3 Rs and RyRs, CaBP1 may serve to inhibit IICR specifically whereas CaM may target RyRs. Indeed, by co-IP from brain tissue we readily observe an interaction between CaBP and InsP_3 Rs, whereas an interaction between InsP_3 Rs and CaM is more difficult to detect (Figure 4F; K Rietdorf, MD Bootman and HL Roderick, unpublished observations).

CaM has a dual role in regulating IICR. It binds in a Ca^{2+} -dependent manner to the regulatory domain of InsP_3 Rs, where it has been suggested to inactivate the receptor following Ca^{2+} release (Michikawa *et al*, 1999; Adkins *et al*, 2000). It also binds independently of Ca^{2+} to the NH_2 -terminus of InsP_3 Rs, inhibiting InsP_3 binding and resultant Ca^{2+} release (Patel *et al*, 1997; Cardy and Taylor, 1998; Sipma *et al*, 1999; Adkins *et al*, 2000; Sienaert *et al*, 2002). Interestingly, in this study we observe that CaBP1 also binds to the NH_2 -terminus of the InsP_3 R resulting in inhibition of InsP_3 binding. Unlike CaM however, which binds to two noncontiguous sequences in the first 159 aa of InsP_3 R1 (Sienaert *et al*, 2002), CaBP1 bound to the first of these sequences alone.

Like other members of the neuronal Ca^{2+} -binding protein family of proteins, CaBP1 is myristoylated at its NH_2 -terminus (Haeseleer *et al*, 2000; Burgoyne and Weiss, 2001). In the case of hippocalcin and recoverin, an increase in cytosolic Ca^{2+} results in exposure of their myristoyl moieties, which results in their translocation to intracellular membranes (Burgoyne and Weiss, 2001). We find that CaBP1 is primarily localised to the plasma membrane and Golgi, but is also present in the cytosol (Figure 1B). This distribution did not change following an increase in cytosolic Ca^{2+} (data not shown). Furthermore, myristoylation is not required for CaBP1 activity since IICR is inhibited by recombinant CaBP and the NH_2 -terminally truncated CaBP1 (cCaBP). The similar levels of inhibition of IICR observed in cells expressing a myristoylation mutant (SCaBP-G2A-YFP) (Supplementary Figure 3), despite expression levels approximately two-fold higher than the wild-type protein (determined by FACS), suggest that myristoylation and thus membrane localisation

increase the potency of CaBP in inhibiting IICR. Although concentrated at the plasma membrane and Golgi apparatus, CaBP clearly affects InsP_3 Rs. Since the Golgi apparatus is a functional InsP_3 -sensitive Ca^{2+} store (Pinton *et al*, 1998), CaBP is strategically located to inhibit release from this organelle. In addition, it is possible that in the dendrites where CaBP1 is endogenously expressed, myristoylation serves to target CaBP1 to the cellular membranes where its function in regulating IICR is performed. In neurons, regions of the ER lie in close proximity to the plasma membrane, which may thus facilitate an interaction between InsP_3 Rs and CaBP (Blaustein and Golovina, 2001). Furthermore, CaBPs may modulate other processes at the plasma membrane, for example Ca^{2+} influx into neurons through $\text{Ca}_v2.1$ channels (Lee *et al*, 2002).

The existence of a constitutive inhibitor of InsP_3 Rs in neurons suggests that IICR is strictly governed. As suggested for CaM, this effect may contribute to the low sensitivity of InsP_3 Rs reported in Purkinje neurons, which require InsP_3 concentrations several orders of magnitude greater than that needed for channel opening in peripheral tissues (Khodakhah and Ogden, 1993; Cardy and Taylor, 1998; Fujiwara *et al*, 2001). A mechanism to allow Ca^{2+} release under conditions of repetitive stimulation such as during memory formation would, however, be advantageous to the cell. We identified a casein kinase 2 consensus site in SCaBP1 (S120) that is structurally conserved with a previously identified site in CaM (S101). Phosphorylation of this serine in CaM has been shown to decrease its affinity for substrates (Quadroni *et al*, 1998). In this study, we find that S120 is the predominant site for phosphorylation in CaBP1. Furthermore, the potency of CaBP1 in inhibiting IICR was increased by mutation of S120 to alanine. Thus, the phosphorylation status of CaBP1 may contribute to the dynamic regulation of InsP_3 R sensitivity in cells where it is expressed.

The data presented here contrast with that previously published (Yang *et al*, 2002). In that study, a truncated form of CaBP1 (cCaBP) spanning the conserved regions of long and short CaBP1 and caldendrin was used (Figure 1A). Since the NH_2 -terminus of other neuronal Ca^{2+} -binding proteins is known to be important in their function, we speculated that this might also be the case for CaBP1. cCaBP, however, had a similar effect on Ca^{2+} signalling as the full-length wild-type protein. We also considered that a possible drawback of the heterologous expression approach used in this study is that the cell may adapt to CaBP1 overexpression, possibly by decreasing the abundance of InsP_3 Rs. This appeared not to be the case since IICR in *Xenopus* oocytes and COS cell microsomes was inhibited by recombinant CaBP. Furthermore, when IICR was investigated at earlier time points following transfection, similar, yet less dramatic, effects on Ca^{2+} release were observed (data not shown). In addition, as shown in Figure 1, we were also unable to detect any change in InsP_3 R levels.

In summary, in this study we have characterised a novel mechanism of action of a newly described InsP_3 R-associated neuronal Ca^{2+} -binding protein. CaBP1 forms part of a wide array of proteins associated with InsP_3 Rs, which enable them to integrate multiple signalling inputs modulating Ca^{2+} release activity and subsequent intracellular Ca^{2+} homeostasis (Roderick and Bootman, 2003). In addition, CaBP1 may prove to be an important determinant of neuronal function

regulating the activity of multiple partners yet to be identified. Future studies involving gene knockout and transgenic approaches will be required to resolve these issues.

Materials and methods

Generation of YFP expression vectors

cDNAs for both long and short CaBP1 were amplified by PCR from pSh-CaBP1-GFP and pLh-CaBP1-GFP, respectively (Haeseleer *et al*, 2000), using primers containing an *Nhe*I site at the 5' end and a *Hind*III site at the 3' end. The resultant PCR products were digested with *Nhe*I and *Hind*III, gel purified and ligated into similarly digested pEYFP-N1 (Clontech). CaBP1₁₃₄ (EF hand mutant) was amplified by PCR using pGEX-CaBP1-c1m3EFH as template and primers to introduce *Eco*R1 restriction sites at both the 5' and 3' ends of the sequence. The PCR product was digested with *Eco*R1, gel purified and ligated into similarly digested pEYFP-N1-SCaBP1. The G2A (myristoylation site) mutant and the casein kinase 2 mutant CaBP1-S120A were generated using the Quick Change mutagenesis protocol (Stratagene). The presence of both mutations was confirmed by sequencing. To generate YFP-CaM and YFP-CaM₁₂₃₄, cDNAs were amplified by PCR using pAED4-hCaM and pET21B-CaM₁₂₃₄, respectively, as templates and primers to introduce *Bam*HI sites at either end of the sequence. The PCR products were digested with *Bam*HI and ligated into similarly digested pEYFP-C1 vector (Clontech). The calbindin cDNA was generously provided by Dr B Schwaller (University of Freiburg).

Expression of CaBP-YFP fusion proteins

COS-7 cells were plated on 22 mm coverslips in 35 mm dishes at 50–60% confluency 24 h prior to transfection. Cells were transfected using the GeneJuice™ transfection reagent (Novagen) using 1 µg of DNA per well. Experiments were performed at 48 h post-transfection unless otherwise stated.

Fura-2 imaging of transfected cells

Videoimaging of Fura-2-loaded cells was performed as previously described using a Sutter (Lambda Technologies, Brattleboro) filter wheel-based imaging system (Peppiatt *et al*, 2003), except that the cells were loaded with 1 µM Fura-2 AM (Molecular Probes). In addition, images of YFP-positive and -negative cells were captured using excitation at 488 nm and emission >520 nm and saved as reference.

Imaging of *Xenopus* oocytes

Ovaries were extracted from albino *Xenopus laevis* following euthanasia with 0.4% tricaine methane sulphonate (MS222). Oocytes were isolated and maintained as previously described (Camacho and Lechleiter, 2000). Oocytes were injected with Fluo-4 (Molecular Probes) to a final concentration of 40 µM 30 min prior to imaging. Imaging was performed using a Noran Oz confocal attached to a Nikon TE200 microscope equipped with a 40 × 1.4 n.a. oil immersion S-Fluor objective. The Ca^{2+} dye was excited by laser illumination at 488 nm. Images of 512 × 512 pixels at a 0.8 zoom were collected at 15 frames per second with a jump average of 2 and a slit width of 25 µm. Ca^{2+} release was initiated by injection with the nonmetabolisable InsP_3 analogue F- InsP_3 (Calbiochem) at final concentrations of 40 or 100 nM. Recombinant CaBP1 (described below) was either coinjected with the indicator dye 30 min prior to imaging or during image acquisition to a final concentration of 8.5 µM. This protocol has previously been successfully used to inject other Ca^{2+} -binding proteins into oocytes (John *et al*, 2001). Image analysis was performed using the public domain software ImageJ (NIH, <http://rsb.info.nih.gov/ij/>).

$^{45}\text{Ca}^{2+}$ fluxes

$^{45}\text{Ca}^{2+}$ fluxes were performed on saponin-permeabilised COS-1 cells as previously described (Missiaen *et al*, 2001). Here, 2 µM thapsigargin was added during the efflux to block the ER Ca^{2+} pumps. The efflux medium was replaced every 2 min during 18 min, and the efflux was performed at 25°C. The additions of $^{45}\text{Ca}^{2+}$ and InsP_3 are indicated in the legends to the figures. At the end of the experiment, the $^{45}\text{Ca}^{2+}$ remaining in the stores was released by incubation with 1 ml of a 2% sodium dodecyl sulphate solution for 30 min. Ca^{2+} release is plotted as the fractional loss, that is, the

amount of Ca²⁺ released in 2 min divided by the total store Ca²⁺ content at that time. The latter value was calculated by summing in retrograde order the amount of tracer remaining in the cells at the end of the efflux and the amount of tracer collected during the successive time intervals.

Preparation of His-tagged CaBP1

Recombinant His-tagged long and short CaBP1 isoforms were prepared as described previously (Nadif Kasri *et al*, 2003).

Expression of GST fusion proteins encoding the NH₂-terminal part of InsP₃R1

cDNA fragments encoding different regions of the NH₂-terminal part of InsP₃R1 (aa 1–159, 1–225, 1–604 and 224–604) were subcloned by PCR amplification using the mouse InsP₃R1 (p400C1 plasmid kindly provided by Dr K Mikoshiba, University of Tokyo) as a template into the pGEX6p2 vector. The PCR products were digested with *Bam*HI and *Eco*RI, cloned in the pGEX6p vector and expressed in BL21(DE3) host cells. Expression and purification of GST fusion proteins was carried out as described previously (Sienaert *et al*, 1997).

Interaction between SCaBP1 and the NH₂-terminal part of InsP₃R1

For the pull-down assay, 100 µg of purified and dialysed GST fusion protein was rebound to glutathione-sepharose 4B beads for 2 h at 4°C. After washing (TBS/lysis buffer, Pierce, Belgium), 100 µg His-tagged SCaBP1 was added to the immobilised GST fusion proteins in the presence of 200 µM free Ca²⁺. Following an incubation of 2 h at 4°C, the protein complexes were washed extensively with binding buffer containing 200 µM of free Ca²⁺, and the retained protein was eluted by 100 mM glutathione in TBS/lysis buffer. Analysis of the eluted proteins was performed on NuPAGE[®] gels, 4–12% Bis-Tris in MES-SDS buffer (Invitrogen Life Technologies) and detected by Sypro[™] Orange staining (Amersham Pharmacia Biotech).

Band-shift assays by nondenaturing gel electrophoresis

The band-shift assays were performed as described previously (Sienaert *et al*, 2002).

InsP₃ binding assay

³H-InsP₃ binding to the NH₂-terminal (aa 1–581) part of InsP₃R1 (Lbs-1) was performed as described previously (Sipma *et al*, 1999). In all, 4 µg of purified Lbs-1 protein was used.

Western blotting

For CaBP expression analysis, cell lysates were prepared as described previously (Roderick *et al*, 2000). Extracts were prepared from either subconfluent cells isolated by trypsinisation or from YFP-positive cells isolated by FACS. A measure of 10–20 µg of the cell extract was analysed using the NuPAGE[®] gel system on 4–12% linear gradient gels, transferred to nitrocellulose and probed for CaBP-YFP using either polyclonal anti-GFP antibody (dilution 1:5000; Roderick *et al*, 1998) or a polyclonal anti-CaBP1 antibody (dilution 1:5000, UW72; Haeseleer *et al*, 2000). Enhanced chemiluminescence (ECL, Pierce) was used to detect immunoreactive bands after incubation of secondary antibodies conjugated to horseradish peroxidase (HRP) (Jackson ImmunoResearch, dilution 1:10 000). For InsP₃R analysis, COS-7 cell lysates were boiled for 2 min in Laemmli sample buffer, analysed on NuPAGE[®] Tris-acetate 3–8% gradient gels, transferred to Immobilon-P and probed with an isoform-specific antibody against InsP₃R3 (I31220, Transduction Laboratories, dilution 1:5000). Quantification of the immunoreactive bands was performed after incubation with secondary antibodies coupled to alkaline phosphatase, detection using Vistra[™] ECF (Amersham Pharmacia Biotech) (Vanlingen *et al*, 1997).

³²P labelling of CaBP-YFP

At 24 h postseeding in 60 mm Petri dishes, COS-7 cells were transfected with the appropriate CaBP1-YFP vectors. At 40 h post-

transfection, cells were washed × 1 in phosphate-free MEM (ICN, cat. #1642349) containing 10% dialysed calf serum, 2 mM L-glutamine, 100 U/ml penicillin and 100 µg/ml streptomycin (labelling media). A measure of 1.5 ml of labelling media containing 0.375 µCi/ml ³²P-phosphorous (orthophosphate in acid-free aqueous solution, Amersham, cat. #PBS13) was added to the cells and incubated for 6 h at 37°C. Following washing in ice-cold TBS, cells were scraped into 0.6 ml of RIPA buffer and processed for IP with 5 µl of a polyclonal anti-GFP antibody used per tube. Protein A/G sepharose (Santa Cruz) was used to capture immune complexes. Isolated proteins were incubated at 95°C for 5 min in 1 × Laemmli buffer containing β-mercaptoethanol prior to analysis through Bis-Tris NuPAGE[®] 4–12% linear gradient gels. Gels were fixed, dried and exposed to the film for 24 h.

Co-IP of InsP₃Rs and CaBP

Lysates were prepared as described for Western blotting from 10 cm dishes of COS-7 cells expressing appropriate CaBP1-YFP constructs. Following clarification, 30 µl of each lysate was retained for Western blot analysis and the remainder diluted four-fold in lysis buffer without detergent. IP using 5 µl of anti-GFP antibody was performed and immune complexes were captured by incubation with 30 µl protein A/G sepharose. Immunoprecipitates were boiled in 30 µl sample buffer and analysed by SDS-PAGE and Western blot using a monoclonal antibody directed against the InsP₃R3. Immunoreactive bands were visualised by ECL following incubation with an HRP-conjugated secondary antibody. For IP from rat brain, whole rat brain was disrupted with a Dounce homogeniser in cell lysis buffer described above. Following 1 h incubation on ice, the insoluble matter was isolated by centrifugation at 5000g for 10 min. A measure of 2 mg of cell lysate was used for each IP. A polyclonal antibody against InsP₃R1 was used to immunoprecipitate InsP₃Rs (Parys *et al*, 1995).

Data analysis

For the percentage of responding cells, an average of the responding cells per coverslip was taken. A response was characterised as a deflection greater than 25 nM from baseline. The integrated Ca²⁺ response is the area under the Ca²⁺ transient minus baseline. Statistical analysis was by Student's *t*-test or χ^2 test. Data are presented as mean ± s.e.m. Significance was accepted at *P* < 0.05 and is indicated by an asterisk.

Supplementary data

Supplementary data are available at *The EMBO Journal* Online.

Note added in proof

During review of this manuscript, a paper appeared in the press that described an inhibitory effect of CaBP on IICR in PC12 and HeLa cells (Haynes *et al*, 2003). Their findings complement our data showing an inhibition of Ca²⁺ release using direct stimulation of InsP₃Rs.

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