1	Reproductive individuality of clonal fish raised in identical environments and its link
2	to early-life behavioural individuality
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#### 26 ABSTRACT

27 Genetic and environmental differences are by far the most studied drivers underlying 28 phenotypic variation. However, a growing number of studies finds among-individual 29 variation that is unexplained by genes or environment. Up to now, it remains an open 30 question whether such seemingly stochastic variation has fitness consequences. To address 31 this question, we performed a tightly controlled long-term life-history experiment with 32 naturally clonal fish (Poecilia formosa) separated directly after birth into identical 33 environments. Maintaining highly standardized conditions for 280 days, we first recorded 34 individuals for 10 hours per day over the first 28 days of their lives, characterizing entire 35 early-life behavioural profiles. We then measured the complete reproductive profiles of 36 these individuals over an average of 4.5 successive broods per individual, quantifying in 37 total 2522 offspring from 152 broods. We find that (i) individuals differ consistently in the 38 size of offspring and broods produced over consecutive broods – despite being genetically 39 identical and being raised in identical environments. (ii) These differences are observed 40 even when controlling for trade-offs between brood size, offspring size and reproductive 41 onset, indicating that individuals differ in life-history productivity. (iii) We also find strong 42 early-life behavioural individuality in both activity and feeding pattern, with among-43 individual differences in feeding being predictive of growth, and consequently reproduction: individuals that consistently spend more time feeding become larger and 44 45 larger individuals consistently produce larger offspring. Our findings provide experimental 46 evidence that processes unexplained by genetic and environmental differences translate 47 into predictable differences in life-history measures and ultimately reproductive fitness.

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### 49 SIGNIFICANCE STATEMENT

50 According to a long-standing paradigm, genetic and environmental differences are the key 51 drivers underlying variation. More recently, however, this paradigm has been 52 experimentally challenged, finding that substantial anatomical, behavioural and 53 neurobiological among-individual variation emerges in the absence of genetic and 54 environmental differences. But does such variation matter? Here, we show that genetically 55 identical individuals reared in identical environments develop substantial differences in 56 activity and feeding behavior, that individuality in feeding behavior then translates into 57 growth differences, and that ultimately these growth differences affect aspects of the phenotype that probably matter most for fitness: brood size, offspring size and life-history 58 59 productivity.

#### 60 INTRODUCTION

61 Phenotypic differences between individuals from the same species abound throughout the 62 animal kingdom (Bell et al., 2009; Laskowski et al., 2022; Sih et al., 2004), with substantial 63 consequences for fitness, ecology and evolution (Bolnick et al., 2011; Carere & Gherardi, 64 2013; Ingley & Johnson, 2014; Jolles et al., 2020; Wolf & Weissing, 2012). It is commonly 65 thought that such individuality (if not minor and inconsequential 'noise' or 'idiosyncrasies') is 66 primarily caused by genetic and/or experiential (i.e. environmental) differences. More 67 recently, however, this long-standing paradigm has been experimentally challenged, with 68 accumulating evidence that even genetically identical individuals raised individually and in 69 identical (i.e. highly standardized) environments develop substantial phenotypic differences 70 (Gärtner, 1990; Honegger & de Bivort, 2018; Werkhoven et al., 2021): isogenic fruit flies 71 reared under identical conditions, for example, differ in locomotor handedness and wing-72 folding, phototaxis, and object-fixated locomotion (Buchanan et al., 2015; Kain et al., 2012; 73 Linneweber et al., 2020); naturally clonal fish reared under identical conditions develop 74 repeatable differences in activity levels, that are already present from the first day of life 75 (Bierbach et al., 2017; Laskowski et al., 2022) (see Freund et al., 2013; Schuett et al., 2011, 76 for related fingings in genetically identical mice and clonal pea aphids).

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Such findings are important as they demonstrate that genetic and environmental differences are not the only potent source of variation between individuals, suggesting that factors like pre-birth processes (including epigenetics), development *per se* and/or minor experiential differences may represent other key drivers of phenotypic differences (Ehlman et al., 2022; Hiesinger & Hassan, 2018; Takagi & Benton, 2020; Vogt et al., 2008). Up to

now, however, all recent studies on the emergence of variation in the absence of genetic
and environmental differences have focused on characterizing emergent behavioural
differences (and their neurobiological underpinnings), but it remains an open question
whether or to what extent these differences really matter.

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88 One of the most direct ways to answer this question is to investigate whether phenotypic 89 differences that emerge under genetic and environmental standardization extend to those 90 aspects of the phenotype that directly affect fitness. The goal of the present study is to do 91 exactly this. We performed an experimental study with a live-bearing, naturally clonal 92 freshwater fish, the Amazon molly, Poecilia formosa. Directly after birth, we separated 93 genetically identical individuals (N = 34) into identical environments and reared them under 94 highly standardized conditions for 280 days (approx. 10 months). We utilized high-95 resolution automated long-term tracking of activity and feeding patterns to characterize 96 their early-life behavioural profiles over the first 28 days of their lives (daily recordings for 10 97 hours at 0.2 s resolution, amounting to a total of 9,520 recording hours and 171.4 million 98 data points) (Ehlman et al., 2023). We then characterized reproductive profiles; recording the onset of reproduction, the size of each brood produced (i.e. the number of offspring 99 100 per brood; in total, N = 152 broods), and the size of all offspring produced (N = 2522 total 101 offspring), thereby observing  $4.5 \pm 1.1$  broods (mean  $\pm$  SD) per female (gestation takes 102 approx. 30 days (Farr & Travis, 1986; Snelson et al., 1986). Individual body size was 103 measured weekly.

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105 We focus on three key research questions. First, do genetically identical individuals 106 separated at birth into identical environments develop significant between-individual 107 differences in reproductive traits, i.e. repeatable differences in offspring- or brood size? 108 Second, are between-individual differences in reproductive traits, if present, indicative of 109 differences in (i) life-history productivity, i.e. the ability to produce new biomass 110 (Adriaenssens & Johnsson, 2009; Biro & Stamps, 2008) and/or (ii) how individuals balance 111 the trade-off between brood size vs. offspring size (Charnov et al., 1995; Gomes-Jr & 112 Monteiro, 2007; Lim et al., 2014; Riesch et al., 2012)? Third, are reproductive differences, if 113 present, related to early-life behavioural differences? Such a link could be possible via 114 growth, where behaviour (in particular feeding behavior) could be linked to growth and 115 growth, in turn, to reproductive output (Furness et al., 2021; Gomes-Jr & Monteiro, 2007; 116 Reznick et al., 1996). As an underlying assumption, and as shown previously (Bierbach et al., 117 2017; Laskowski et al., 2022), we expected repeatable early-life behavioural variation. 118 Whenever appropriate, we focus on repeatability as a key parameter to quantify and test for 119 individuality (e.g. Bell et al., 2009; Biro & Stamps, 2015; Dingemanse & Dochtermann, 120 2013).

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### 122 RESULTS

### 123 Repeatable differences in early-life behaviour

We find that our genetically identical individuals separated into identical environments on the first day of life, exhibit strong behavioural individuality during the first four weeks of life, both in activity (R = 0.371, 95% CI = [0.329, 0.413]) and feeding behaviour (R = 0.183, 95% CI = [0.145, 0.224]). These repeatable differences even increase when controlling for within-

132	Tab. 1 in Suppl. 2).
131	negatively correlated (estimate = -9.134, CI [-10.550 – -7.719] p-value < 0.0001, $R^2$ = 0.164,
130	feeding = 0.238, 95% CI = [0.194, 0.285]). Daily activity and feeding behaviour are
129	observations (adjusted R for activity = $0.571$ , $95\%$ CI = $[0.532, 0.621]$ ; adjusted R for
128	individual variation caused by individuals growing and becoming older during our

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### 134 Repeatable differences in offspring size, brood size and life-history productivity

135 We find consistent among-individual differences in both the average size of offspring (Fig. 136 1a, R = 0.396, 95% CI [0.308, 0.484]) and the number of offspring produced (Fig. 1b, R = 137 0.177, 95% CI = [0.117, 0.238]) over consecutive broods. Interestingly, the trade-off 138 between brood size and offspring size explains only little of the variation observed (estimate 139 = -0.013, CI [-0.018, 0.007], p-value < 0.001, partial  $R^2$  = 0.115) (Fig. 1c, Tab. 2 in Suppl. 2). 140 And even when controlling for this trade-off, among-individual differences remain: given 141 the same brood size, onset of reproduction, and size at parturition, some individuals 142 consistently produce larger offspring than others (adjusted R for offspring size = 0.134, 95%143 CI = [0.085, 0.192]), and similarly, while controlling for onset of reproduction, size at 144 parturition, and offspring size, some individuals consistently produce larger broods than 145 others (adjusted R for brood size = 0.077, 95% CI = [0.050, 0.114]). These findings strongly 146 suggest that - next to developing repeatable among-individual differences in offspring size 147 and brood size - our genetically identical individuals, raised individually in highly 148 standardized environments, also differ in life-history productivity.

We stress that both offspring- and brood size are the most direct fitness components one can measure, and seemingly small - but repeatable - differences in these traits may have profound long-term consequences. This can be seen, for example, when considering the cumulative number of offspring produced, where even relatively minor individual differences in brood size, when expressed consistently, result in large among-individual differences in total reproductive output (**Fig. 1d**).

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### 158 Link between feeding behaviour and offspring size, mediated by growth

159 Despite no direct link between our two behavioural measures, activity and feeding, and our 160 three reproductive traits, i.e. offspring size (Fig. 2 a-b), brood size (Fig. 2 c-d), and onset of 161 reproduction (Fig. 2 e-f) (Tab. 3 in Suppl. 2), we find an indirect link between one of our 162 behavioural traits and reproduction: fish that spend more time feeding grow to a larger size 163 (estimate = 0.007, CI [0.003, 0.012], p-value = 0.002, partial R<sup>2</sup> = 0.266; Fig. 3b, Fig. 3d), and larger fish, in turn, produce larger offspring (Fig. 3c; estimate = 0.619, CI [0.299, 0.939], 164 165 p-value < 0.001, partial R<sup>2</sup> = 0.129). Larger fish also start reproducing later (estimate = 166 55.537, CI [25.189, 85.886], p-value = 0.001, partial  $R^2$  = 0.310; Fig. 3f), but there is no 167 effect of reproductive onset on brood size (Fig. 3e) (Tab. 5 in Suppl. 2).

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169 In contrast to feeding behaviour, we find no indirect link between activity and reproduction 170 (Fig. 3a, Tab. 3 and Tab. 5 in Suppl. 2), nor do we find an effect of our behavioural traits on 171 the growth rate (predicted from fit growth curves; see Methods) (Tab. 4 in Suppl. 2). We 172 note that growth rate and onset of reproduction are negatively correlated: fish that grow faster start reproducing later (estimate = 980.383, CI [235.137, 1725.629], p-value = 0.012, partial  $R^2 = 0.188$ ), but there is no effect of growth rate on brood- and offspring size (**Tab. 5** in **Suppl. 2**).

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#### 177 DISCUSSION

178 Employing an experimental 'twin study' approach, we find that genetically identical 179 individuals, raised separately and in highly standardized environments, develop repeatable 180 differences in key reproductive characteristics. In particular, when considering consecutive 181 broods, individuals differ consistently in how many offspring they produce and in how large 182 these offspring are. While we find evidence for a weak trade-off between offspring size and 183 number, repeatable among-individual differences are maintained even when controlling for 184 this tradeoff, as well as for body size and onset of reproduction, providing clear evidence 185 that individuals differ in life-history productivity. While previous studies have provided firm 186 evidence that substantial among-individual variation in anatomical-, behavioural- and 187 neurobiological traits can emerge even in the absence of genetic and environmental 188 differences (Bierbach et al., 2017; Buchanan et al., 2015; Honegger & de Bivort, 2018), the 189 current study builds on and substantially extends these studies by demonstrating that the 190 emerging variation extends to aspects of the phenotype that are directly associated with 191 fitness.

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193 Interestingly, we find no direct link between early-life behavioural differences and 194 differences in reproductive traits. It is conceivable, that a direct link between early-life 195 behavioural and later-in-life reproductive traits may only become apparent in non-benign

196 and/or more complex environments. To give a concrete example, in our experimental set-197 up, there are only minimal differential costs/benefits associated with behavioural 198 phenotypes: fish were presented with a stationary food resource, located at a standardized 199 position in tanks with no additional structures. There was no need to search for food and no 200 cost associated with exploiting a stationary food resource. In contrast, in a more naturalistic 201 context, activity might be linked to the ability to find food and the exploitation of a 202 stationary food resource can be risky (Brown, 1992; Verdolin, 2006). That said, it will be 203 interesting to see whether future studies, taking the above and other factors into account, 204 will be able to establish a link between early-life behavioural individuality and reproduction.

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206 While we do not find a direct link between reproduction and early-life behaviour, we find 207 the size of offspring produced to be linked to early-life feeding behavior: some individuals 208 feed consistently more than others, individuals that feed more grow to a larger predicted 209 final size, and individuals with a larger final size have larger offspring (but not larger broods). 210 The allocation of resources into increasing offspring size, rather than offspring number, may 211 represent an adaptation to environments in which larger offspring have higher fitness, e.g., 212 resource limitation, competition, or high juvenile mortality (cannibalism or size-dependent 213 predation) (reviewed in Riesch et al., 2012). All of the above factors may apply to our 214 experimental design: during the reproduction period, individuals were fed a standardized 215 amount of food that they shared with a P. mexicana male, which we kept in the female's 216 tank as a sperm donor, potentially causing both resource limitation and competition. 217 Furthermore, offspring were removed from the females' tanks directly after parturition, 218 which decreased the risk of cannibalism but may have caused perceived predation from the 219 female's perspective.

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221 It is intriguing to see that systematic (i.e. repeatable) among-individual differences in key 222 fitness components can emerge even in the absence of genetic and environmental 223 differences. Future work may compare the observed differences in repeatedly expressed 224 reproductive traits to other iteroparous species, taking genetic and/or environmental 225 variation into account, which will give us a better understanding of the magnitude of the 226 observed differences. Furthermore, we will need to evaluate both the causes and 227 consequences of the observed differences. First, all individuals in our study were exposed 228 to one (very specific) environment, and it will be important to investigate whether the 229 observed differences can also be detected in other environments. It will be particularly 230 informative to include predation risk, a key determinant of fitness and major factor shaping 231 life-history trade-offs (Lima & Dill, 1990; Lind & Cresswell, 2005; Reznick & Endler, 1982; 232 Reznick et al., 1996; Walsh & Reznick, 2009), and to investigate, for example, if high-233 productivity individuals are differently affected by predators than low-productivity 234 individuals. Second, in order to evaluate the consequences of the observed differences in 235 brood- and offspring size, it will be important to investigate whether and to what extent 236 these differences are heritable. Are offspring from mothers with large/small offspring (or 237 large/small brood size) more likely to produce large/small offspring (large/small brood size) 238 themselves? While all our individuals are genetically identical, such inheritance is still 239 conceivable, for example via epigenetic mechanisms (Bocock & Aagaard-Tillery, 2009; Klosin & Lehner, 2016; Kovalchuk, 2012). 240

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242 The study of between-individual variation is one of the central themes in ecology and 243 evolution. While such variation – if not minor and inconsequential "noise" – has traditionally 244 been thought to be caused by genetic and environmental differences, evidence is accumulating that even in the absence of such differences, substantial among-individual 245 246 variation can emerge. Here, we show that such emergent among-individual variation 247 extends to aspects of the phenotype that directly affect fitness. Put differently, we find that 248 the among-individual variation that arises under highly standardized conditions reflects 249 more than just 'idiosyncrasies' or 'noise' - it really matters.

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251 METHODS

### 252 Study species and holding conditions

253 Amazon mollies used here, were obtained from a stock kept at Humboldt-Universität zu 254 Berlin, (Berlin, Germany). Original stocks were housed in groups of approx. 20-50 fish under 255 standardized conditions: 50-100 liter tanks, 12:12h light:dark cycle, air temperature control 256 (approx.  $24\pm1^{\circ}$ C), weekly water changes. Twice a day, fish were fed with powder food (Sera 257 vipan baby). In the stock tanks, Amazon mollies were kept sympatrically with P. mexicana 258 males, allowing females to reproduce freely. Before the experiment, we separated potential 259 mothers (originating from the same ancestor) from the stock population in order for them to 260 give birth in individual tanks; this allowed us to track the mother ID (N = 3) of individuals 261 used in this experiment.

262

### 263 Early-life behaviour

264 Individuals were transferred to separate, identical observation tanks (Fig. 1 in Suppl. 1) on 265 the day they were born. Our behavioural observations started the next day, i.e. the first full 266 day of their life. We recorded individuals daily over the first 28 days of their life. Recordings were tracked using the software Biotracker (Mönck et al., 2018), and the movement data 267 268 obtained from Biotracker (csv-files with xy-coordinates over time) were processed 269 (visualization, calculation of metrics) with a costume repository we developed for this 270 purpose (Stärk et al., 2022) (see Suppl. 1 for further recording and tracking details). We 271 assessed individual activity as average daily swimming speed (cm/sec). Once a day, 272 individuals were fed with a stationary food resource, a 'food patch', that was positioned at a 273 standardized location in the tank for 2 hours. We assessed the time spent feeding as the 274 amount of time (min), individuals spent in immediate proximity to the food patch (Suppl. 1). 275 Food patches were prepared every 2 to 3 days using Sera vipan baby powder food and 276 agar (protocol is provided in Suppl. 1).

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#### 278 Reproductive profiles

As the Amazon molly is a gynogenetic species, i.e. sperm from one of the parental species (*P. mexicana*, *P. latipinna*) is needed to trigger embryogenesis (but the male's DNA is not incorporated into the offspring's genome (Lamatsch et al., 2005; Schlupp, 2005)), we ensured that females had access to various *P. mexicana* males throughout the reproductive phase.

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285 We initiated the reproduction phase of the experiment directly after our early-life 286 behavioural observations. We transferred test individuals to individual breeding tanks (11 287 liter, flow-through water system, visual separation between individual tanks) where they 288 remained until the end of the experiment (total observation time per individual is 280 days, 289 approx. 10 months). Breeding tanks were equipped with one P. mexicana male as sperm 290 donor, a small plastic pipe (length = 4 cm, diameter = 2 cm), and 'sera biofibres' (a loose 291 bundle of green plastic fibers, structurally comparable to thread algae) as a refuge. Females 292 were swapped between breeding tanks once a week (i.e. males remained in the tank) in a 293 randomized manner to control for a potential tank or male effect. Over the course of the 294 experiment, females had access to approx. 20 different males. Feeding was standardized: 295 twice a day for 5 days a week, fish received 1/64 tsp (up to the age of 70 days) or 1/32 tsp 296 (from the age of 70 to 280 days) of powder food.

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Breeding tanks were checked for offspring daily. Once a female gave birth to a brood, all offspring were photographed and counted. Offspring standard length (i.e. the length from the tip of the snout to the end of the caudal peduncle) was later measured from the photos using ImageJ (Schneider et al., 2012). In total, we recorded 152 broods and measured the size of 2522 offspring from 144 broods (no measurements for 8 broods). Individuals produced on average 4.5  $\pm$  1.1 broods (mean  $\pm$  SD). We excluded all individuals from our analyses with no or partial reproductive data (*N* = 11 females).

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### 306 Long-term growth

307 Standard lengths of focal individuals were measured from photos once a week, using 308 ImageJ (Schneider et al., 2012) (34 ± 1 measurements per individual, mean ± SD). We fit 309 individual growth curves using the von Bertalanffy growth model (von Bertalanffy, 1938), a 310 logistic function commonly used to model fish growth. Estimated parameters in this 311 function are the theoretical age when size is zero (t<sub>0</sub>), the growth coefficient (K), and the 312 maximum predicted (i.e. asymptotic) size (L<sub>inf</sub>). For all analyses, we used predicted sizes 313 estimated using individual growth curves rather than raw measurements and characterized 314 individual growth via K and Linf obtained from those individual growth curves.

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#### 316 Statistical analysis

317 General details

318 Data were analyzed in R version 4.2.1 (R Core Team, 2022). Most parsimonious LMs (linear 319 models) and LMMs (Linear mixed-effect models; models built using the lme4-package; 320 Bates et al., 2015) were fit via stepwise-backward removal of non-significant predictors. 321 Model assumptions were visually assured using residual- and q-q plots. In the main text, we 322 report estimates and p-values for significant predictors only. The effects of covariates are 323 presented in Suppl. 2. Most importantly, individual size at parturition is related to both the 324 number and size of offspring produced (model summaries in Tab. 3 in Supp. 2, figure in 325 Suppl. 3); and we found mother ID (N = 3) to be related to individual growth and reproductive output (Suppl. 2). We therefore included mother ID as a covariate in all 326 327 models (LMs and LMMs), and individual size at parturition wherever appropriate. In all 328 LMMs, individual ID was included as a random term. Individual size on the first day of life 329 did not affect early-life behavior (Tab. 1 in Suppl. 4), growth (Tab. 21 in Suppl. 4), or 330 reproductive output (Tab. 31 in Suppl. 4); and was therefore not considered during 331 analyses. For significant predictors, we calculated partial R<sup>2</sup> using the sensemakr- and 332 r2qlmm-package (Cinelli et al., 2021; Jaeger, 2017) (for LMs and LMMs, respectively). Complete model summaries of all full (containing all predictors) and final models 333 334 (containing significant predictors only) are provided in Suppl. 2. Model summary tables 335 (including marginal and conditional R<sup>2</sup> following Nakagawa et al., 2017) were built using the 336 package siPlot (Lüdecke, 2022).

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338 Repeatabilities

339 We estimated repeatabilities with 95% CIs (confidence intervals) in two ways: first by 340 building LMMs with only the target variable as response and female ID as random term, but 341 no predictors (i.e. normal repeatability), and then by adding fixed effects to the model, 342 allowing us to estimate the amount of variation caused by consistent between-individual differences while controlling for variation explained by other factors (i.e. adjusted 343 344 repeatability). The significance of consistent among-individual differences was derived from 345 the 95% CI being distinctly different to 0 (95% CI based on 1000 model simulations) (Hertel et al., 2020). 346

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To test for 'reproductive individuality', we calculated normal repeatabilities in the size of broods (N = 152 broods) and the size of offspring (N = 144 broods, average offspring size per brood) over all broods produced. To test for between-individual differences in productivity, we adjusted repeatabilities of both brood- and offspring size for onset, female

size at parturition, and mother ID. For the model on brood size, we additionally included offspring size as predictor and vice versa (i.e. we accounted for brood size, offspring size, and onset trade-offs).

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To test for early-life behavioural individuality, we calculated normal repeatabilites for activity 356 357 (N = 941 observations of 34 individuals) and the time spent feeding (LMM with N = 932358 observations of 34 individuals), observed daily over the first 28 days of life. We also 359 adjusted repeatabilities for size and age (week 1-4, categorical variable), i.e. we accounted 360 for variation that was caused by individuals growing and aging over the observation period. 361 We further included a size-age interaction term as predictor (activity was differently affected by size, depending on age). For repeatability calculations, activity was log-transformed for 362 363 normality. We tested if early-life activity (response) and feeding (predictor) are correlated (LMM N = 931 observations of 34 individuals). 364

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366 Link between early-life behaviour and reproduction

To test for a behaviour-reproduction link, we built three models with either brood size (LMM with N = 152 broods), offspring size (LMM with N = 144 broods), or reproductive onset (LM with N = 34 individuals) as response. In all models, activity and feeding (averaged over 28 days) were modeled as predictors.

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To test for the link between behaviour and growth, we fit one model on each  $L_{inf}$  (LM with N373 = 34 individuals) and K (LM with N = 34 individuals), with activity and feeding as predictors 374 (average behaviour over 28 days). When having K as the response, we additionally included bioRxiv preprint doi: https://doi.org/10.1101/2023.04.05.535730; this version posted April 6, 2023. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

- 375 L<sub>inf</sub> as a covariate to control for the effect L<sub>inf</sub> has on K; i.e. bigger fish grow slower to their
- 376 final size L<sub>inf</sub>, (LM with L<sub>inf</sub> as response and K as well as mother ID as predictors: intercept [CI]

377 = 6.09 [5.8, 6.4]; estimate of K [CI] = -16.9 [-20.0, -13.8], R<sup>2</sup> = 0.89).

- 378
- 379 To test for the link between growth and reproduction, we fit a model on each brood size
- 380 (LMM with N = 152 broods), offspring size (LMM with N = 144 broods), and onset (LM with
- N = 34 individuals); including K and L<sub>inf</sub> as predictors and female age at parturition (not in
- 382 the onset-model) as a covariate.
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### 540 FIGURES



Fig. 1 (a-b) Individuals differ consistently in the size- and number of offspring produced over successive broods; boxplots sorted by median offspring size. (c) The brood size vs. offspring size trade-off explains only very little of the variation; shown are individual averages in brood/offspring size ± standard deviation. (d) Differences in reproductive productivity have profound long-term consequences because reproductive output accumulates over time; shown is the cumulative number of offspring produced by each individual over the first 280 days of life.



549

550 **Fig. 2** We find no effect of early-life behaviour (activity and feeding) on offspring size (a-b),

551 brood size (c-d), or onset of reproduction. (e-f) Shown are individual averages (data points)

552 ± standard deviation (error bars); individuals are coloured differently.



Fig. 3 (a) There is no effect of early-life activity on the final predicted size, L<sub>inf</sub> [cm] feeding behaviour and reproductive output are indirectly connected via growth: (b, d) individuals that feed more grow to a larger size, and (c) larger fish produce larger offspring (f). Larger fish also start reproducing later but (e) final size and brood size are not linked. Shown are (b, d) regression lines (black) with 95% Cls, (d), estimates for individual von Bertalanffy growth curves (lines) with raw data (points). (a-c, e) individual averages (points) with standard deviation (error bars).

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562 SUPPLEMENT 1

563

564 Behavioural observations

565

For our early-life behavioural observations, individual observation tanks (Fig. 1) were filmed from above with a Basler acA5472 camera (5 frames per second). Activity was recorded over the first 8 hours of each day, followed by a 2-hour feeding. In total, we collected 952 recordings of daily activity and feeding behaviour (34 test fish recorded over 28 days). Recordings were tracked using the software Biotracker (Mönck et al., 2018). Quality of tracks was assured via plotting each individual's movement data (i.e., xy-coordinates over time) in 30 min chunks and visually inspecting the trajectory plots for errors.

573

574 To assess individual, daily activity, we used 941 recording days (mean ± SD recording 575 length: 471.7 ± 29.9 min; 11 days were removed from the data due to technical issues). 576 Individual activity was calculated from the xy-coordinates over time in steps of 0.2 sec as the average distance moved (cm) in one second. To calculate how much time individuals 577 578 spent feeding, we processed feeding recordings of 932 days (mean ± SD recording length 579 = 120.2 ± 11.2 min; 20 recordings were removed due to technical issues). Individual time 580 spent feeding was calculated as the amount of time an individual spent in the 'feeding 581 zone', a 5 x 13 cm large zone surrounding the food patch. For the visualization of 582 movement data and the calculation of metrics (activity, time spent feeding) we a costume 583 repository we developed for this purpose (Stärk et al., 2022).

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584



585

Fig. 1 One experimental unit with 2 observation tanks. Water level in the tanks: 7 cm. Tanks
were externally filtered. Food patches were present during the feeding time slots only.

588 There was no visual contact between observation tanks, but tanks were connected via a

589 flow-through water system (24 observation tanks split into 4 flow-through systems).

- 590
- 591

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593

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# 602 Food patch preparation

Ingred	lients
-	0.5g Agar (Bio Agar-Agar, Ruf)
-	100 ml tap water
-	14 g powder food (Sera vipan baby)
<u>Materi</u>	als
-	One fine scale
-	One 150 ml beaker
-	One small pot
-	One stove plate
-	One pipette
-	Food patch cups (max. volume = 2ml)
Instruc	ctions
1.	Prepare clean and dry food patch forms.
2.	Weigh the amount of agar needed in a 150 ml beaker.
3.	Add 100 ml of water.
4.	Fill the water-agar mixture into a small pot and cook for 2 min at medium heat.
5.	Remove pot from stove and add powder food, mix thoroughly.
6.	Let cool down for approx. 5 min at room temperature until agar starts binding.
7.	Quickly fill the agar-food mixture into the cups using the pipette.
8.	Let the food patches harden in the fridge for approx. 10 min without a lid.
9.	Store food patches in a container with a lid (e.g., petri dishes) in the fridge until
	further proceeding (can be stored for at least 4 days when stored at 4°C).
<u>Notes</u>	
The p	rotocol can be modified regarding the amount of food or agar added. Adding more
	Ingred - - - - - - - - - - - - - - - - - - -

631 agar will give the mixture a firmer texture. Using less agar is not advised. The amount of

- 632 agar used here is the least amount possible (resulting in a very soft texture) in order to allow
- 633 juvenile fish to feed on the food patches. The amount of food can be both decreased and
- 634 increased allowing for variations in food quality; food variations may require adjustments in
- 635 the amount of agar used.

### 636 SUPPLEMENT 2

- 637
- 638 **Tab.1** Full (left) and final (right) models testing if early-life feeding behaviour and activity are

### 639 correlated.

Response			Full model		Final model					
	Predictors	Estimate	CI	P-value	Estimate	CI	P-value			
	Intercept	55.554	48.433 - 62.675	-	60.734	55.675 – 65.792	-			
	Activity [cm/sec]	-9.222	-10.644 – -7.800	<0.001	-9.134	-10.550 – -7.719	<0.001			
	Mother ID [m2]	3.103	-11.814 – 18.020	0.088	-	-	-			
	Mother ID [m3]	10.495	0.907 – 20.084	0.000	-	-	-			
	Random Effects									
Feeding [min]	σ²		721.68		721.69					
[]	<b>T</b> 00		150.82 Test fish ID		165.52 Test fish ID					
	ICC		0.17		0.19					
	Ν		34 Test fish ID		34 Test fish ID					
	Observations		931		931					
	Marginal R <sup>2</sup> / Conditional R <sup>2</sup>		0.174 / 0.317		0.164 / 0.320					

			Full model         Final mode           stimate         Cl         P-value         Estimate         Cl         P-value $5.083$ $4.238 - 5.927$ - $4.466$ $3.827 - 5.104$ - $0.011$ $-0.0180.005$ $<0.001$ $-0.013$ $-0.0180.007$ $<0.001$ $0.002$ $-0.006 - 0.002$ $0.335$ -         -         - $0.002$ $-0.006 - 0.002$ $0.335$ -         -         - $0.919$ $0.730 - 1.108$ $<0.001$ $0.988$ $0.830 - 1.146$ $<0.001$ $0.038$ $-0.273 - 0.196$ $0.061$ -         -         - $0.197$ $-0.3590.035$ $0.061$ -         -         - $0.197$ $-0.3590.035$ $0.061$ -         -         - $0.10$ $0.10$ $0.10$ $0.10$ -         - $0.10$ $0.13$ $34$ Test fish ID $34$ Test fish ID $34$ Test fish ID						
Response	Predictors	Estimate	CI	P-value	Estimate	CI	P-value		
	Intercept	5.083	4.238 – 5.927	-	4.466	3.827 – 5.104	-		
	Brood size	-0.011	-0.018 – -0.005	<0.001	-0.013	-0.018 – -0.007	<0.001		
	Onset [days]	-0.002	-0.006 – 0.002	0.335	-	-	-		
	Size at parturition [cm]	0.919	0.730 – 1.108	<0.001	0.988	0.830 – 1.146	<0.001		
	Mother ID [m2]	-0.038	-0.273 – 0.196	0.061	-	-	-		
Offspring	Mother ID [m3]	-0.197	-0.359 – -0.035		-	-	-		
(brood	Random Effects								
average, mm)	σ²		0.10		0.10				
	<b>T</b> 00		0.01 Test fish ID		0.01 Test fish ID				
	ICC		0.10		0.13				
	Ν		34 Test fish ID		34 Test fish ID				
	Observations		144		144				
	Marginal R <sup>2</sup> / Conditional R <sup>2</sup>		0.580 / 0.623			0.545 / 0.602			

# **Tab. 2** Full (left) and final (right) models testing for an offspring size vs. brood size trade-off.

# 643 Tab. 3 Full (left) and final (right) models testing if early-life behaviour (activity, feeding)

# 644 predicts reproduction (brood size, offspring size, onset).

			Full model			Final model		
Response	Predictors	Estimate	CI	P-value	Estimate	CI	P-value	
	Intercept	-15.029	-35.583 – 5.524	-	-19898	-39.823 – 0.027	-	
	Activity [cm/sec]	-2.317	-4.881 – 0.248	0.082	-	-	-	
	Feeding [min]	-0.071	-0.207 – 0.066	0.312	-	-	-	
	Size at parturition [cm]	8.002	3.307 – 12.697	0.001	7820	3.157 – 12.484	0.001	
	Mother ID [m2]	6.108	0.048 – 12.169	<0.001	4324	-1.608 – 10.256	<0.001	
	Mother ID [m3]	10.208	5.706 - 14.710	<0.001	8444	4.353 – 12.535	<0.001	
Brood size			Random	Effects				
	σ²		70.72			70.53		
	<b>T</b> 00		7.54 Test fish ID			10.07 Test fish ID		
	ICC		0.10			0.12		
	Ν		34 Test fish ID			34 Test fish ID		
	Observations		152		152			
	Marginal R <sup>2</sup> / Conditional R <sup>2</sup>		0.160 / 0.241		0.138 / 0.246			
	Intercept	5.168	4.359 – 5.978	-	5.147	4.366 – 5.928	-	
	Activity [cm/sec]	-0.047	-0.149 – 0.056	0.369	-	_	-	
	Feeding [min]	0.002	-0.004 - 0.007	0.513	-	_	-	
	Size at parturition [cm]	0.791	0.607 – 0.975	<0.001	0.804	0.622 – 0.987	<0.001	
	Mother ID [m2]	-0.058	-0.305 – 0.190	0.018	-0.119	-0.356 – 0.118	<0.001	
Offspring	Mother ID [m3]	-0.265	-0.4460.084	0.010	-0.270	-0.434 – -0.107	0.001	
size (brood average,			Random	Effects				
mm)	σ²		0.10		0.10			
	<b>T</b> 00		0.01 Test fish ID		0.02 Test fish ID			
	ICC		0.12			0.14		
	Ν		34 Test fish ID		34 Test fish ID			
	Observations		144			144		
	Marginal R <sup>2</sup> / Conditional R <sup>2</sup>		0.552 / 0.607			0.539 / 0.605		
	Intercept	126.473	94.420 – 158.526	-	133.786	123.545 – 144.027	-	
	Activity [cm/sec]	4.000	-6.139 – 14.138	0.426	-	-	-	
Onset of	Feeding [min]	0.072	-0.458 - 0.602	0.784	-	-	-	
reproduction	Mother ID [m2]	15.777	-9.037 – 40.590	0.004	19.964	-1.760 – 41.689	0.002	
լագչօյ	Mother ID [m3]	-19.908	-35.944 – -3.872	0.004	-17.411	-31.434 – -3.387	0.002	
Brood size Offspring size (brood average, mm) Onset of reproduction [days]	Observations		34			34		
	R <sup>2</sup> / R <sup>2</sup> adjusted		0.341 / 0.250		0.326 / 0.282			

## 646 **Tab. 4** Full (left) and final (right) models testing for an effect of early-life behaviour on the

			Full model			Final model		
Response	Predictors	Estimates	CI	P-value	Estimates	CI	P-value	
	Intercept	4.465	4.159 – 4.771	-	4.375	4.141 – 4.608	-	
	Activity [cm/sec]	-0.045	-0.141 – 0.052	0.354	-	-	-	
	Feeding [min]	0.006	0.001 – 0.011	0.015	0.007	0.003 - 0.012	0.002	
L <sub>inf</sub> [cm]	Mother ID [m2]	0.144	-0.093 – 0.380	<0.001	0.098	-0.116 – 0.313	<0.001	
	Mother ID [m3]	-0.349	-0.502 – -0.196	<0.001	-0.378	-0.517 – -0.239	<0.001	
	Observations		34		34			
	R <sup>2</sup> / R <sup>2</sup> adjusted	0.592 / 0.535			0.579 / 0.537			
	Intercept	0.296	0.247 – 0.344	-	0.305	0.263 – 0.346	-	
	Activity [cm/sec]	0.001	-0.001 - 0.004	0.326	-	-	-	
	Feeding [min]	0.000	-0.000 - 0.000	0.963	-	-	-	
K	L <sub>inf</sub> [cm]	-0.046	-0.0560.035	<0.001	-0.047	-0.0560.039	<0.001	
ĸ	Mother ID [m2]	0.004	-0.003 – 0.010	<0.001	0.005	-0.001 – 0.011	<0.001	
	Mother ID [m3]	-0.019	-0.0240.013	<0.001	-0.018	-0.0230.014	_ \0.001	
	Observations		34		34			
	R <sup>2</sup> / R <sup>2</sup> adjusted		0.814 / 0.781			0.807 / 0.788		

## 647 maximum predicted size (L<sub>inf</sub>) and growth rate (K) of individuals.

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- 649 Tab 5 Full (left) and final (right) models testing if von Bertalanffy growth parameters (the
- 650 growth rate K and maximum predicted size L<sub>inf</sub>) predict reproduction (brood size, offspring
- 651 size, onset of reproduction).

			Full model								
Response	Predictors	Estimates	CI	P-value	Estimates	CI	P-value				
	Intercept	43.171	-71.768 – 158.110	-	7.187	1.217 – 13.157	-				
	L <sub>inf</sub> [cm]	-4.576	-23.302 – 14.151	0.630	-	-	-				
	К	-173.702	-530.501 – 183.096	0.338	-	-	-				
	Age at parturition [weeks]	0.205	0.015 – 0.394	0.035	0.215	0.026 – 0.404	0.026				
	Mother ID [m2]	5.630	-0.429 – 11.690	0 111	5.085	-0.828 – 10.999	0.011				
	Mother ID [m3]	3.808	-4.004 – 11.620	0.111	5.875	2.167 – 9.583					
Brood size			Rando	m Effects		L					
	σ²		73.73			73.90					
	<b>T</b> 00		8.36 Test fish ID			9.32 Test fish ID					
	ICC		0.10			0.11					
	N		34 Test fish ID			34 Test fish ID					
	Observations		152			152					
	Marginal R <sup>2</sup> / Conditional R <sup>2</sup>		0.120 / 0.210		0.105 / 0.206						
	Intercept	5.739	1.240 – 10.239	-	4.808	3.310 – 6.307	-				
	L <sub>inf</sub> [cm]	0.475	-0.257 – 1.206	0.170	0.619	0.299 – 0.939	<0.001				
	К	-3.076	-17.062 – 10.909	0.638	-	-	-				
	Age at parturition [weeks]	0.029	0.022 – 0.037	<0.001	0.030	0.022 – 0.037	<0.001				
Offspring	Mother ID [m2]	-0.081	-0.318 – 0.155	0.026	-0.096	-0.320 – 0.128	<0.001				
size	Mother ID [m3]	-0.385	-0.689 – -0.080	0.020	-0.329	-0.497 – -0.162	<0.00T				
(brood	Random Effects										
average,	σ²		0.10		0.10						
mm)	<b>T</b> 00		0.01 Test fish ID		0.01 Test fish ID						
	ICC		0.12		0.11						
	N		34 Test fish ID		34 Test fish ID						
	Observations		144		144						
	Marginal R <sup>2</sup> / Conditional R <sup>2</sup>		0.568 / 0.618			0.570 / 0.616					
	Intercept	-141.259	-595.812 – 313.294	-	-204.285	-383.856 – -24.714	-				
	L <sub>inf</sub> [cm]	45.090	-29.090 – 119.270	0.224	55.537	25.189 – 85.886	0.001				
Onset of	К	768.659	-631.930 – 2169.249	0.271	980.383	235.137 – 1725.629	0.012				
repro- duction	Mother ID [m2]	15.631	-7.523 – 38.785	0 202	-	-	-				
[days]	Mother ID [m3]	-0.368	-31.794 – 31.058	0.373	-	-	-				
	Observations		34	-		34					
	R <sup>2</sup> / R <sup>2</sup> adjusted		0.360 / 0.271			0.317 / 0.273					

## 653 SUPPLEMENT 3



655 **Suppl. 3** Female size is positively correlated with (a) offspring size (shown is average 656 offspring size per brood  $\pm$  SD) and (b) brood size. Data points are coloured by individuals.

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### 657 SUPPLEMENT 4

658

659 Tab. 1 Full (left) and final (right) models testing if size on the first day of life predicts early-

# 660 life behavior.

			Full model			Final model		
Response	Predictors	Estimates	CI	P-value	Estimates	CI	P-value	
	Intercept	1.546	-1.112 – 4.204	-	0.996	0.577 – 1.416	-	
	Size at birth [cm]	-0.656	-3.786 – 2.475	0.672	-	-	-	
Activity	Mother ID [m2]	1.210	0.301 – 2.118	0.025	1.188	0.298 – 2.077	0.011	
[cm/sec]	Mother ID [m3]	0.595	-0.079 – 1.269	0.025	0.524	-0.050 - 1.098	0.072	
	Observations		34		34			
	R <sup>2</sup> / R <sup>2</sup> adjusted		0.218 / 0.139			0.213 / 0.162		
	(Intercept)	10.888	-38.316 – 60.093	-	48.058	42.847 – 53.268	-	
	Size at birth [cm]	42.305	-15.642 – 100.252	0.146	-	-	-	
Feeding	Mother ID [m2]	-9.241	-26.064 – 7.581	0.454	-	-	-	
[min]	Mother ID [m3]	1.021	-11.450 – 13.493	0.454	-	-	-	
	Observations		34		34			
	R <sup>2</sup> / R <sup>2</sup> adjusted		0.151 / 0.066			0.000 / 0.000		

## 662 Tab. 2 Full (left) and final (right) models testing if size on the first day of life predicts

## 663 reproductive output.

			Full model			Final model		
Response	Predictors	Estimates	CI	P-value	Estimates	CI	P-value	
	(Intercept)	-1.708	-18.716 – 15.300	-	7.187	1.217 – 13.157	-	
	Size at birth [cm]	10.904	-8.548 – 30.356	0.270	-	-	-	
	Age at parturition [weeks]	0.208	0.019 – 0.398	0.031	0.215	0.026 - 0.404	0.027	
	Mother ID [m2]	4.675	-1.168 – 10.518	0.00(	5.085	-0.828 – 10.999	0.011	
	Mother ID [m3]	4.561	0.254 – 8.868	0.066	5.875	2.167 – 9.583	0.011	
Prood size		Random Effects						
BIOOD SIZE	$\sigma^2$		74.06			73.90		
	<b>T</b> 00		8.19 Test fish ID			9.32 Test fish ID		
	ICC		0.10		0.11			
	Ν		34 Test fish ID		34 Test fish ID			
	Observations		152			152		
	Marginal R <sup>2</sup> / Conditional R <sup>2</sup>		0.116 / 0.204		0.105 / 0.206			
	Intercept	8.036	7.306 – 8.766	-	7.685	7.448 – 7.921	-	
	Size at birth [cm]	-0.431	-1.276 – 0.414	0.315	-	-	-	
	Age at parturition [weeks]	0.031	0.024 – 0.038	<0.001	0.031	0.024 – 0.038	<0.001	
	Mother ID [m2]	-0.035	-0.288 – 0.218	<0.001	-0.053	-0.307 – 0.201	<0.001	
Offspring	Mother ID [m3]	-0.462	-0.649 – -0.276	<0.001	-0.514	-0.674 – -0.353	<0.001	
size (brood			Random	Effects				
average,	σ²		0.10			0.10		
mm)	<b>T</b> 00		0.02 Test fish ID			0.02 Test fish ID		
	ICC		0.18			0.19		
	Ν		34 Test fish ID			34 Test fish ID		
	Observations		144			144		
	Marginal R <sup>2</sup> / Conditional R <sup>2</sup>		0.525 / 0.612		0.519 / 0.612			

665 Tab. 3 Full (left) and final (right) models testing if size on the first day of life predicts the

666 growth rate K and predicted final size L<sub>inf</sub>.

			Full model			Final model	
Response	Predictors	Estimates	CI	P-value	Estimates	CI	P-value
	Intercept	4.171	3.473 – 4.870	-	4.716	4.602 - 4.831	-
	Size at birth [cm]	0.650	-0.172 – 1.473	0.117	-	-	-
L. A	Mother ID [m2]	0.019	-0.220 – 0.258	<0.001	0.041	-0.202 – 0.284	<0.001
Lint	Mother ID [m3]	-0.407	-0.584 – -0.230	<0.001	-0.337	-0.493 – -0.180	<0.001
	Observations		34		34		
	R2 / R <sup>2</sup> adjusted		0.472 / 0.419		0.426 / 0.389		
	Intercept	0.309	0.268 – 0.350	-	0.305	0.263 – 0.346	-
	Linf	-0.045	-0.054 – -0.036	<0.001	-0.047	-0.0560.039	<0.001
	Size at birth [cm]	-0.017	-0.037 – 0.004	0.107	-	-	-
К	Mother ID [m2]	0.006	-0.000 - 0.011	<0.001	0.005	-0.001 – 0.011	<0.001
	Mother ID [m3]	-0.016	-0.0210.010	<0.001	-0.018	-0.0230.014	<0.001
	Observations		34		34		
	R2 / R <sup>2</sup> adjusted		0.824 / 0.799			0.807 / 0.788	