



Invited Review

Repurposing approved drugs for cancer therapy

Catherine H. Schein*

Department of Biochemistry and Molecular Biology Faculty, Institute for Human Infections and Immunity (IHII), University of Texas Medical Branch, Galveston 301 University Boulevard, Galveston, Texas 77555, USA

*Correspondence address. E-mail: chschein@utmb.edu

Received 26 August 2020; Revised 11 December 2020; Editorial Decision 17 December 2020; Accepted 17 December 2020

Abstract

Background: Many drugs approved for other indications can control the growth of tumor cells and limit adverse events (AE).

Data sources: Literature searches with keywords ‘repurposing and cancer’ books, websites: <https://clinicaltrials.gov/>, for drug structures: <https://pubchem.ncbi.nlm.nih.gov/>

Areas of agreement: Introducing approved drugs, such as those developed to treat diabetes (Metformin) or inflammation (Thalidomide), identified to have cytostatic activity, can enhance chemotherapy or even replace more cytotoxic drugs. Also, anti-inflammatory compounds, cytokines and inhibitors of proteolysis can be used to control the side effects of chemo- and immuno-therapies or as second-line treatments for tumors resistant to kinase inhibitors (KI). Drugs specifically developed for cancer therapy, such as interferons (IFN), the tyrosine KI abivertinib TKI (tyrosine kinase inhibitor) and interleukin-6 (IL-6) receptor inhibitors, may help control symptoms of Covid-19.

Areas of controversy: Better knowledge of mechanisms of drug activities is essential for repurposing. Chemotherapies induce ER stress and enhance mutation rates and chromosome alterations, leading to resistance that cannot always be related to mutations in the target gene. Metformin, thalidomide and cytokines (IFN, tumor necrosis factor (TNF), interleukin-2 (IL-2) and others) have pleiomorphic activities, some of which can enhance

tumorigenesis. The small and fragile patient pools available for clinical trials can cloud the data on the usefulness of cotreatments.

Growing points: Better understanding of drug metabolism and mechanisms should aid in repurposing drugs for primary, adjuvant and adjunct treatments.

Areas timely for developing research: Optimizing drug combinations, reducing cytotoxicity of chemotherapeutics and controlling associated inflammation.

Key words: thalidomide, metformin, cytokine storm, unfolded protein response (UPR), proteasome inhibitors, interferons (IFN), tumor necrosis factor (TNF), interleukin-2 (IL-2), Covid-19 adjuvant treatments

Introduction

Using compounds approved for one clinical use in another disease or syndrome is referred to as ‘repurposing’. Most of the drive for repurposing is the high cost of developing a drug, and the very long time it can take to determine the safety and specificity of a completely new drug. The timeline for any new cancer drug to go through enough clinical trials to obtain approval can be years, or even decades. Even for chronic myeloid leukemia (CML), which may affect about 0.2% of the population during their lifetime, over a decade was required for one of the first therapeutic KIs, Gleevec, to become standard of care. For smaller patient pools or drugs with less clear results in treatment, the delay can be even longer. Approval of omacetaxine, a plant alkaloid that inhibits protein translation, for treating TKI-resistant CML took more than 30 years.¹ A phase 1 trial (NCT02081378) for asciminib (an allosteric inhibitor of the ABL kinase) to treat TKI-resistant CML and Philadelphia chromosome-positive acute lymphoblastic leukemia (ALL), began in 2014, is ongoing in August of 2020 with an estimated completion date of 2024.

Repurposing, per se, is an old concept in oncology. Indeed, the first chemotherapy drugs might be regarded as repurposed chemical weapons, arising as they did from research on the ‘mustard gas’ (which has no relation to mustard and is liquid at room temperature) that caused so many deaths and

chronic illness in the wars of the 20th century. While treating survivors of these attacks, doctors realized that the toxins, in addition to their vesicant (i.e. blister-inducing) activity, might have antitumor potential. In the medical equivalent of beating guns into plowshares, in this case converting a toxic compound to a therapeutic one, chemists and doctors worked together in a decade-long search for compounds with lower toxicity and enhanced cytostatic activity, with the hope of finding treatments to prolong the lives of their cancer patients.² After many explorations of conjugates with different biological molecules, two alkylating agents, chlorambucil (Leukeran) and busulfan (Myleran) (Fig. 1), were developed to treat chronic lymphocytic and myeloid leukemias (CLL and CML). Despite the advent of many other chemotherapies, these simple drugs continue to be used to this day.

This review will discuss two basic meanings of repurposing in cancer therapy. The first is adapting drugs used in other areas, for example anti-infectives or treatments for chronic diseases, for their observed cytostatic activity.³ A second meaning is using drugs that were designed primarily to treat other illnesses to enhance the effects of chemotherapy or manage side effects. Examples of these two areas are shown below. Section ‘Repurposing cancer therapies as antivirals and specifically anti-Covid-19 treatments’ introduces the recent testing of cancer medications for treating Covid-19 infections and

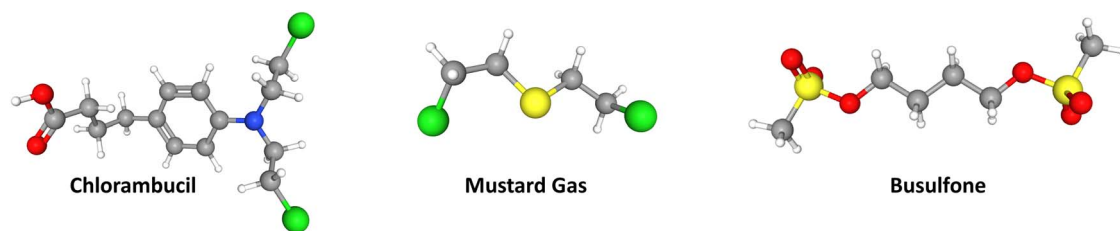


Fig. 1 Alkylating agents used for treating leukemias, chlorambucil (left) and busulfone (myleran, right) were patterned after toxic nitrogen mustard gas (center) during many design iterations. 3D chemical structures are from Pubchem; atom colors are: carbon, gray; hydrogens, small white; nitrogen, blue; sulfur, yellow; chloride, green; oxygen, red.

how this may have future benefit for repurposing in oncology.

Adapting common drugs with cytostatic potential for cancer therapy

The great dream of the many waves of drug design in cancer is to achieve a drug that will only kill cancer cells, leaving most normal cells untouched. No chemotherapy has ever achieved this lofty goal. Unlike the careful design of, for example, drugs targeting phosphorylation cascades^{4,5} and orally available TKIs,⁶ there is no clear cellular target for most early therapies. Chemotherapy infusions cannot be typically done at home, as there is risk of anaphylaxis and many are so toxic that accidental extravasation⁷ can lead to difficult to treat blisters. Directing treatments to specific organs often leads to the escape of a few wayward bandits, abnormal cells that will eventually find their way into another tissue and reinitiate tumorigenesis. Oncologists are thus always on the lookout for drugs with fewer side effects that can be used to treat cancer as a chronic disease or even prevent it.^{8,9} Drugs designed to treat many different indications have been introduced into cancer therapy¹⁰ and hundreds more have been reported to inhibit the growth of tumor cells in culture (see <https://depmap.org/repurposing> for the growth inhibitory activity of approved drugs against 578 human cancer cell lines.¹¹).

One area for repurposing is to replace current therapies with others that are cytostatic, rather than cytotoxic. Two repurposed drugs that have recently shown the most success are metformin, used since

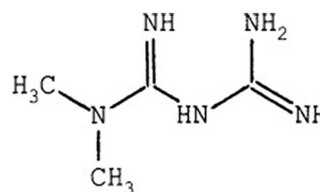


Fig. 2 Metformin (dimethylbiguanide) used for diabetes treatment.

1995 in the USA for diabetes, and thalidomide and its derivatives, which were developed to treat diseases such as psoriasis and inflammation related to infections.

Metformin as a cytostatic agent

Metformin (Fig. 2) traces its roots to a plant extract whose primary ingredient was guanidine, used throughout the Middle ages to treat diabetes symptoms.¹² Metformin was first synthesized in 1922, but due to the advent of insulin, only advanced as ‘Glucophage’ 30 years later when it was approved in France. It has become the first-line treatment for type 2 diabetes. Pertinent to this review, metformin is playing an increasing role as a cytostatic cancer treatment, thanks to its low toxicity (its primary adverse event (AE) is the rarely occurring lactic acidosis.) The first cancer trials arose from reports that diabetics taking metformin daily had lower rates of breast¹³ and other cancers, augmented by studies showing cells from metformin-treated diabetics do not grow well in culture.¹⁴

The cytostatic effects reported led to introducing metformin as an adjunct therapy for different types

of cancers, some of which have reported remarkable success. In a recent phase II trial, 139 lung adenocarcinoma patients, whose tumors contained driver epidermal growth factor receptor (EGFR) mutations, were treated with TKIs (erlotinib hydrochloride, afatinib dimaleate or Gefitinib at standard dosage) plus or minus 500 mg/day of metformin (i.e. well within the normal dosage for treating diabetes). Adding metformin increased the progression-free survival (PFS) by about a third; it nearly doubled overall survival (OS).¹⁵

Metformin also improved PFS and OS in advanced, previously untreated non-small cell lung cancer (NSCLC) when used in combination with platinum-based chemotherapy with or without the anti-VEGF inhibiting antibody, bevacizumab (Avastin) in two phase II studies. These included 33 non-diabetic patients of whom 70% had some history of smoking, with *KRAS*, *EGFR* and *LKB1* mutation prevalence of 48, 26 and 8.3%, respectively. The PFS and OS for metformin-treated patients were especially improved in those with *KRAS* mutations. This suggests that determining molecular subgroups should be used to guide therapy in the future, especially in light of the paucity of direct *KRAS* inhibitors.¹⁶

As exciting as these results are, metformin does not always improve survival.¹⁷ The effects seen in the various clinical trials, many of which are ongoing, are patient specific. Several metabolic pathways altered by metformin treatment may account for the heterogeneity in response. It might be logical to assume that the overall effect of metformin, by lower circulating glucose concentration, would specifically starve tumor cells (which have an enhanced metabolic need for glucose), leading to decreased proliferation and metastasis. Various other explanations have been given for its observed effects in diabetes and in preventing cell growth,¹⁸ whereby the inability of metformin to enter many cells has not always been accounted for in studies of its effects on metabolism.¹⁹ One is that metformin can metabolically reprogram cancer cells by activating 5' AMP-activated kinase (AMPK), by increasing the ratio of AMP to ATP to some extent in cells (whereby

the ratio was much higher after Rosiglitazone treatment). Other studies attributed the lower rates of breast cancer to metformin's role in controlling fatty acid oxidation.²⁰

Identifying the patients most likely to be helped relies on accurately identifying the cells most affected by metformin and determining if those types predominate in the patient tumors. A study of normal murine mammary cells indicated that metformin had the highest effect on hormone receptor positive luminal cells, where it decreased the total cell number, progenitor capacity and DNA damage. The authors suggest that identifying this type of cells in humans would indicate those most likely to benefit from metformin treatment.²¹ To shed further light on this question, whole transcriptome RNA sequencing²² of 40 breast cancer patients before and after 13–21 days of dose escalating metformin (from 500 mg/day to 1500 by day 6, which is still within the dose range for diabetes treatment) revealed that patients' profiles correlated with an optimal antiproliferative response. One commonality was that metformin treatment increased glucose flux in the tumor (as measured by 18-fluorodeoxy glucose uptake via PET-CT) as well as in other tissues.

Thalidomide and derivatives in cancer therapy

Thalidomide derivatives have a variety of uses in modern cancer therapy. Indeed, someone who awoke suddenly from a 60-year sleep would be amazed that this notorious drug would be in such widespread use today. The clinical tragedy associated with its first introduction remains a cautionary tale for all involved in drug research.²³ Thalidomide was first developed to treat morning sickness and sold over the counter to pregnant women in Germany in the 1950s, with recommended doses in the range of aspirin treatments (300–500 mg). The drug's side effects, including peripheral neuropathy, stopped its approval in the USA. However, thalidomide was only withdrawn worldwide in 1962 after it was linked to severe birth defects. As discussed elsewhere,²³ even after this withdrawal, thalidomide remained in

clinical use for treating Hansen disease (leprosy). The major use of thalidomide and its derivatives for many years was immunomodulatory. There are now a variety of thalidomide-related compounds to choose from that have been designed to specifically control different pathways in immune cells. The relatively low cost of treatment for this family of drugs means it can play a role in many cancer therapies, both for its tumor growth inhibition and its anti-inflammatory activities.

The earliest introduction of thalidomide derivatives to cancer therapy was to control inflammation. Eventually, their ability to prevent the growth of certain cancer cells was recognized. Thalidomide and its more potent structural relatives, lenalidomide and pomalidomide,²³ are used to treat multiple myeloma,²⁴ mantle cell lymphoma, and myelodysplastic syndromes associated with the deletion 5q abnormality. On the other hand, apremilast (Otezla) was specifically designed to inhibit PDE4^{25,26} and is now used to control psoriasis, lupus erythematosus and rheumatoid arthritis.²⁷ PDE4 is a phosphatase that degrades cAMP, a small molecule that can modulate inflammatory responses. Targeted PDE4 inhibitors are in preclinical trials for cancer.^{28, 29}

At this point, the mechanistic basis for using the thalidomide drug family in cancer becomes confusing, as they have pleiomorphic effects. For example, their anti-inflammatory activity has been linked to their ability to inhibit secretion of tumor necrosis factor (TNF)- α and other cytokines.^{30,31} Anti-TNF antibodies such as Humira have revolutionized the treatment of psoriasis and rheumatoid arthritis; however, inhibiting TNF may enhance inflammatory central nervous system syndromes such as multiple sclerosis.³²

A recent discovery suggests another reason for thalidomide-related compounds' action in cancer cells: their ability to bind cereblon,³³ a protein involved in limb outgrowth. The devastating teratogenic effects of thalidomide when taken in early pregnancy have also been linked to this binding. Even more intriguing are current attempts to manipulate thalidomide's binding to cereblon to induce specific protein degradation in cancer cells.³⁴

Early results indicated that thalidomide induced specific degradation of repressors in T-cells that can lead to activation and increased IL-2 secretion, thus providing another way to stimulate the immune system to fight cancer cells.³⁵

Cytokine-based therapies

There are repeated trials of cytokines as co-therapies. The earliest of the cytokines to enter cancer trials were recombinant interferons (IFN), introduced in the 1980s.³⁶⁻³⁹ The IFNs were identified for their antiviral activity; their first use in cancer was for the control of leukemias. They have also been tested for a variety of blood and solid cancers. IFN- α was used for many years to treat hairy cell leukemia,⁴⁰ CML and myelofibrosis⁴¹; it may still be resorted to alone or as adjunct therapy for CML patients resistant to multiple TKIs.¹ A recent paper reported deep molecular remission in four of nine CML patients treated with Imatinib plus Ropeginterferon- α 2b, with few AE.⁴² IFNs have also been suggested to enhance treatment with temozolomide by inhibiting the MGMT repair enzyme.⁴³ However, chromosome instability in AML has been shown to directly upregulate IFN-stimulated genes,⁴⁴ suggesting that IFN itself will not be helpful in treatment. The cost, need for injection or infusion, and side effects of IFNs suggest they could be replaced with small molecule, intracellular inducers, such as STING activators,^{45,46} which may also require injection, or with compounds that may activate select steps in the IFN-induced pathways.

Other cytokines^{39,47} have been tried repeatedly as co-treatments. Two of these proteins, TNF⁴⁸ and IL-2, were highly anticipated as potential anticancer therapies. Early tests showed some successes, but their toxic effects thwarted widespread clinical use. As with IFNs, there are continuing attempts to repurpose IL-2 in cancer therapy,⁴⁹⁻⁵² as an additive to immunotherapy, or for treating patients who have failed to respond to immunotherapy.⁵³ More study on how to control the AEs in IL-2 treatments^{54,55} may lead to safer ways to use this molecule in cancer.

TNF is also problematic as a cancer treatment. Direct clinical use of TNF has been limited by side effects, such as cachexia and fever. TNF has been

implicated in the origin of many different types of tumors, possibly precluding its use as a treatment.⁵⁶ Furthermore, TNF levels in the body rise with age,^{57,58} along with increased incidence of cancer. TNF levels should certainly be checked in those who fail to respond to immune therapies.

However, things may turn around for this protein; a recent paper using mouse models suggests that the TLR-5 antagonist entolimod may control the toxic effects of TNF without affecting its antitumor activity.⁵⁹ If this proves true in human trials, this may unveil a new future use of TNF in cancer therapy.

Other drugs in testing

Other agents designed to treat a variety of different diseases are in testing against cancers. Statins have been tested as anticancer agents, as they can inhibit the activity of many GTPase oncogenes. While statins have not performed well as cancer drugs,⁶⁰ disulfiram (Antabuse), developed to treat alcoholism, is in testing with copper to treat metastatic breast cancer (NCT03323346). Nelfinavir, an AKT inhibitor developed to treat HIV, is in phase I trials for treating solid tumors,⁶¹ (NCT01445106).

Gamma-secretase inhibitors, developed to treat Alzheimer disease (AD), are in multiple tests as anti-cancer drugs (alone [NCT01981551, NCT03785964, NCT03691207] or in combination with Car-T therapy [NCT 03502577]) as they also inhibit Notch 1 and signal peptidases.⁶² And in turn, the cancer drug saracatinib (AZD-0530), designed to inhibit the SRC and BCR-ABL kinases, is now being tested for its effects on AD,⁶³ based on its inhibition of the Fyn Kinase, which may contribute to synaptotoxicity.

Repurposing drugs to control the effects of or enhance chemotherapy

Oncologists are also combining chemotherapeutics with compounds to control their effects on normal cells or improve their overall activity. Many FDA-approved chemotherapy drugs have severe side effects, ranging from blistering at the site of infusion to hair and teeth loss. While cooling the scalp or

chewing ice during the infusion can partially control these side effects,⁶⁴ additional anti-inflammatory compounds are being sought. The orally available inhibitors of poly (ADP-ribose) polymerase (PARP) and kinases are easy to administer, but they may induce side effects such as nausea,^{65,66} which may require co-treatment with antiemetics. However, use of proton pump inhibitors, used to control GERD symptoms in as many as 30% of cancer patients, has been shown to limit the effect of chemotherapy and worsen OS.⁶⁷

Especially patients with cardiac problems may benefit from co-treatment during chemotherapy with drugs such as β -blockers and aspirin.^{68,69} Low-dose aspirin has long been recommended both to control inflammation and inhibit coagulation and was previously recommended to control stroke and heart attack incidence. However, ASPREE, a 4.7-year placebo controlled trial of >19 000 individuals older than 65–70 determined there was no advantage of taking aspirin. The risk of being diagnosed with stage 3 or 4 cancers and increased mortality was higher in the aspirin-treated patients than in the placebo group.⁷⁰

The usefulness of β -blockers to control tumor growth especially has been explored as propranolol decreased proliferation, migration and invasion of triple negative breast cancer cells *in vitro*.⁷¹ Topical treatment with the β -blockers propranolol and timolol is a validated treatment for complicated infantile hemangiomas.^{72,73} There are also many studies indicating that β -blockers can control the growth of vascular sarcomas and other endothelial cell tumors.⁷⁴ Their use in other cancers has shown less benefit. Multiple retrospective studies of patients treated with combination therapies including β -blockers have shown little indication of overall efficacy in ovarian cancer,⁷⁵ lung cancer⁷⁶ or in preventing cancer recurrence.⁷⁷

However, β -blocker co-treatment with anthracyclines can significantly reduce chemotherapy cardiotoxicity and preserve left ventricle function.^{78,79} Furthermore, topical treatments with propranolol and timolol, such as those developed for infantile hemangiomas, can also shorten the recovery time

when applied to painful swelling around the nails that can develop after treatment with EGFR inhibitors.⁸⁰

Another repurposed drug, dexrazoxane, may be superior for this use. Originally developed as an antimetabolic, dexrazoxane, the (+)-enantiomer of razoxane, has now been approved by the FDA for repurposing as a cotreatment to prevent anthracycline-induced extravasation injuries⁸¹ and cardiomyopathy. Dexrazoxane's effect may be due to its ability to inhibit the formation of a toxic iron-anthracycline complex.⁸² A recent multicenter study⁸³ of over 1000 AML patients treated with daunorubicin or mitoxantrone showed that cotreatment with dexrazoxane significantly lowered cardiac problems and also reduced treatment-related mortality. Dexrazoxane cotreatment is also used for immunosuppressive purposes.

The blood pressure medication, Mibefradil, which slows the excretion of many common drugs, can be used short term to enhance the activity of several different cancer drugs.⁸⁴ However, this may be counterproductive as increases in a drug's plasma concentration can induce cytokine-release syndrome (also called 'cytokine storm'). Drugs that can prevent the release of several inflammatory cytokines are hence useful. Abivertinib⁸⁵ is a novel small molecule TKI targeting mutant forms of both EGFR and Bruton's tyrosine kinase (BTK). In addition to slowing cell growth, abivertinib binds irreversibly and prevents phosphorylation of the BTK receptor, thus inhibiting the release of pro-inflammatory cytokines, including IL-1 β , IL-6 and TNF- α .

Leucovorin (folinic acid), developed to treat pernicious and megaloblastic anemias,⁸⁶ can control the side effects of methotrexate and chemotherapy drugs. Combinations of leucovorin with 5-fluorouracil and either oxaliplatin or irinotecan (the FOLFOX or FOLFIRI regimens) are standard treatments for colorectal cancers. However, alternatives to oxaliplatin should be sought, as there are severe and long-term neurotoxic AEs associated with this combination.⁸⁷ Artemisinin (malaria treatment) derivatives may be an alternative to oxaliplatin, as they are cytotoxic against colon cancer cells lines at low concentrations when used

in combination with leucovorin and 5-fluorouracil (FOLNSC combination).⁸⁸

Another problem arising in cancer is enhanced coagulation, leading to stroke and heart attacks, due to chemotherapy or disease progression. While aspirin can reduce coagulation, other compounds specifically designed to control clotting, such as apixaban (Eliquis), a factor Xa inhibitor developed to treat atrial fibrillation,⁸⁹ may be preferable as cancer progresses. Apixaban does not have the same effect on platelet interaction as warfarin and heparin and may thus be a safer alternative.

One problem with combining inhibitors is that treatment with a wide variety of cytotoxic agents enhances mutations and treatment resistance by inducing ER stress and the unfolded protein response (UPR). UPR-induced autophagy supports tumorigenesis and the development of resistance to treatment.^{90,91} One way to handle the unfolded protein response to chemotherapy in general is to control the proteasome, which regulates protein expression by removing ubiquitylated proteins. Proteasome inhibitors such as ixazomib (Ninlaro[®]) can be combined with lenalidomide and dexamethasone for the treatment of patients with multiple myeloma who have received at least one prior therapy. Bortezomib (Velcade) is used in multiple myeloma and mantle cell lymphoma. Bortezomib caused a rapid and dramatic change in the levels of intracellular peptides that are produced by the proteasome,⁹² due to the inhibitor's direct interaction with subunits of the proteasome. Bortezomib has been suggested as an alternative to vincristine (and to treat neuropathy associated with vincristine treatment) for pediatric ALL.⁹³

Of course, no drug is without AEs. A recent report⁹⁴ suggests that administration of the antihistamine, ketotifen, can control the ocular effects of bortezomib.

Repurposing cancer therapies as antivirals and specifically anti-Covid-19 treatments

There is a clear overlap between the needs of cancer and severe Covid-19 patients for drugs to control inflammation and coagulation. Thus, there has been

a recent spate of papers on repurposing drugs, including chemotherapy agents, to treat Covid-19 (and before the current pandemic, Ebola virus). The reader will not be surprised that there are trials planned of metformin and thalidomide as possible adjuvant treatments (e.g. NCT04510194, NCT04273529). A recent review summarizes the plethora of ongoing clinical trials of anti-cancer drugs being tested.⁹⁵ Among these are many phase 1–3 trials of IFNs and Janus associated kinase (JAK) inhibitors, alone or in combination with antiviral drugs. The JAK inhibitor, baricitinib, has also been reported to be efficacious in treating Covid-19.⁹⁶ There are also intriguing reports that those with mutations in IFN-related genes⁹⁷ or auto-antibodies against IFNs have a more severe disease course.⁹⁸ Covid-19, along with other β -coronaviruses, is known to interfere with the early immune response that is based on IFN and the genes it stimulates.

However, while early treatment with IFN (types I and III) may have benefit, later in the disease course, it can cause damage to the lung epithelia that can lead to superinfections.⁹⁹ This is because IFNs can also play a role in cytokine release syndrome or ‘cytokine storm’, which may be responsible for mortality associated with Covid-19. Other tests are ongoing to determine whether inhibition of specific inflammatory cytokines is beneficial. A recent report found high levels of TNF in T-cells from Covid-19 patients with a fatal outcome and suggested the cytokine may have inhibited the immune response to the virus.¹⁰⁰ As this increase may inhibit normal humoral responses,¹⁰⁰ it is possible that TNF inhibitors, such as those developed for psoriasis, might be beneficial in treatment.

There are also many ongoing clinical trials for tocilizumab and other inhibitors of interleukin-6, a cytokine associated with inflammation. These IL-6 inhibitors were previously approved to treat multiple myeloma, lymphoproliferative disorders and Castleman’s syndrome. Abivertinib, a cancer therapy TKI, is in phase 2 clinical trials for repurposing to prevent cytokine storm in Covid-19 patients (NCT04440007).

While these trials are dedicated to finding better treatments for Covid-19 in this hour of emergency, it is clear that the results can have impact on the future course of cancer therapy. There are particular ramifications for how to control the inflammatory AEs of immune therapy, which often limit its usefulness in treating fragile patients who have endured many types of chemotherapy. Companies should be conscious of the advantages of combining repurposed compounds with novel therapies, during their clinical trials. Any additional costs will be more than paid for if the therapy is then more acceptable to patients.

A few words in parting

While the examples here show the positive side of repurposing, it should be noted that not all chemotherapy agents can be re-used for every cancer. Combining therapies does not always bring better results. For example, recent results of a breast cancer trial¹⁰¹ indicate that adding the anthracycline Epirubicin (+5-fluorouracil and cyclophosphamide) to neoadjuvant chemotherapy (including trastuzumab and pertuzumab) had little effect on event-free survival but increased AE, including two women who developed acute leukemia. As chemotherapy may primarily act in breast cancer by inducing terminal menopause, adding it to treatments designed to reduce estrogen levels may be superfluous. A 1999 study of metastatic (stage IV) melanoma showed there was no survival advantage to combining dacarbazine with cisplatin, carmustine and tamoxifen compared to high-dose dacarbazine alone. Only 25% of the patients on either regimen survived more than a year.¹⁰² Fortunately, new immunotherapy drugs, such as a combination of CTLA-4 and PD-1 inhibitors (ipilimumab and nivolumab), have revolutionized the outlook for patients with metastatic melanoma. A recent report showed 1-year survival exceeded 80%, with 4-year rates >50%.¹⁰³

A second caution is that while many FDA-approved compounds have been found to have antitumor cell activity *in vitro* in ‘high-throughput’

screening, moving them into human treatments is difficult. Not every new combination of drugs can be tested in a random control trial; the costs, not to mention the lack of suitable patients, would be prohibitive. Improving selection from 'shot in the dark' assays of whole cells requires better understanding of why a given drug is cytostatic, and skepticism is called for when effects in culture require unrealistically high concentrations. For example, emetine, better known as an active ingredient of Ipecac syrup used to induce vomiting, binds ribosomes¹⁰⁴ and has been reported to selectively kill AML cells.¹⁰⁵ Whether its side effects can be overcome sufficiently to justify testing as a cancer therapeutic is not currently clear.

Investigating the metabolic basis for a 'hit' can be time consuming and costly. Funding is often lacking for such trials,¹⁰⁶ especially for off-patent drugs. In this respect, there should be more public and pharmaceutical coalition funding available for off-label testing. Another major difficulty in repurposing compounds is the need to assemble a proper patient pool for a blinded control study. One must take into account that cancer patients, especially those who are older, have additional disease, or have survived many different treatments and accompanying AE, are fragile. For example, a paper reporting a complete response to Ipilumab therapy following treatment with BRAF/MEK inhibitors ended by reporting the patient's death from side effects of the therapy.¹⁰⁷ Many trials, even of new drugs, are abandoned because they do not meet their target patient pool within a reasonable time frame, or the company funding it changes direction. Dosages that may be well tolerated in trials conducted using healthy subjects or when used for treating the diseases, the drug was originally intended for may not be achievable in cancer patients. Combination therapies only complicate these problems. However, individual treatments with older drugs can lead to remarkable cures.¹⁰⁸ Still, the best approach for a patient who has no clear alternative therapy may be to recommend an ongoing trial. Another alternative, for patients with genetic markers that correlate with the anticipated activity of a test intervention, is the N-of-1 trial.¹⁰⁹

Here, the patient serves as his/her own control. The success of the therapy, or basis for continuation, can be based for example on blood levels of selected metabolites and proteins that should lead to reduced disease or tumor growth.

Conclusions

The examples included in this review and related references show that the pantheon of approved drugs is a rich source of solutions for many problems in oncology. Replacing cytotoxic with cytostatic drugs targeting specific cellular pathways promises to further enhance treatment while limiting AEs. Combinations with repurposed inhibitors designed to control inflammation can control the toxicity of chemotherapeutics or cytokines to normal cells and provide new treatments for resistant tumors.

The pace of research on repurposing compounds from all clinical areas is breathtaking and has contributed to increasing survival and easing the effects of chemotherapy on cancer patients. In the proper setting, established drugs can be smart therapies that can replace untargeted toxins relied upon in the past.

Data Availability Statement

No new data were generated or analyzed in support of this research.

Funding

This work was supported in part by grants from the National Institute of Allergy and Infectious Diseases, USA, R21 AI105985-01 (to CHS) and R01 AI137332-01.

Conflict of interest statement

The authors have no potential conflicts of interest.

References

1. Winer ES, DeAngelo DJ. A review of omacetaxine: a chronic myeloid leukemia treatment resurrected. *Oncol Ther* 2018;6:9–20. doi: [10.1007/s40487-018-0058-6](https://doi.org/10.1007/s40487-018-0058-6) [published Online First: Epub Date].

2. Haddow A. On the biological alkylating agents. *Perspect Biol Med* 1973;16:503–24. doi: [10.1353/pbm.1973.0029](https://doi.org/10.1353/pbm.1973.0029) [published Online First: Epub Date].
3. Berns A, Ringborg U, Celis JE, et al. Towards a cancer mission in horizon Europe: recommendations. *Mol Oncol* 2020;14:1589–615. doi: [10.1002/1878-0261.12763](https://doi.org/10.1002/1878-0261.12763) [published Online First: Epub Date].
4. Sugiyama Y, Kameshita I. Multi-PK antibodies: powerful analytical tools to explore the protein kinase world. *Biochem Biophys Res Commun* 2017;11:40–5. doi: [10.1016/j.bbrep.2017.06.005](https://doi.org/10.1016/j.bbrep.2017.06.005) [published Online First: Epub Date].
5. Braun W, Schein CH. Membrane interaction and functional plasticity of inositol polyphosphate 5-phosphatases. *Structure* 2014;22:664–6. doi: [10.1016/j.str.2014.04.008](https://doi.org/10.1016/j.str.2014.04.008) [published Online First: Epub Date].
6. Duenas-Gonzalez A, Garcia-Lopez P, Herrera LA, et al. The prince and the pauper. A tale of anticancer targeted agents. *Mol Cancer* 2008;7:82. doi: [10.1186/1476-4598-7-82](https://doi.org/10.1186/1476-4598-7-82) [published Online First: Epub Date].
7. Chang R, Murray N. Management of anthracycline extravasation into the pleural space. *Oxf Med Case Rep* 2016;2016:omw079. doi: [10.1093/omcr/omw079](https://doi.org/10.1093/omcr/omw079) [published Online First: Epub Date].
8. Turanli B, Grotli M, Boren J, et al. Drug repositioning for effective prostate cancer treatment. *Front Physiol* 2018;9:500. doi: [10.3389/fphys.2018.00500](https://doi.org/10.3389/fphys.2018.00500) [published Online First: Epub Date].
9. Jeter JM, Bowles TL, Curiel-Lewandrowski C, et al. Chemoprevention agents for melanoma: a path forward into phase 3 clinical trials. *Cancer* 2019;125:18–44. doi: [10.1002/cncr.31719](https://doi.org/10.1002/cncr.31719) [published Online First: Epub Date].
10. Gupta SC, Sung B, Prasad S, et al. Cancer drug discovery by repurposing: teaching new tricks to old dogs. *Trends Pharmacol Sci* 2013;34:508–17. doi: [10.1016/j.tips.2013.06.005](https://doi.org/10.1016/j.tips.2013.06.005) [published Online First: Epub Date].
11. Corsello SM, Nagari RT, Spangler RD, et al. Discovering the anti-cancer potential of non-oncology drugs by systematic viability profiling. *Nature Cancer* 2020;1:235–48. doi: [10.1038/s43018-019-0018-6](https://doi.org/10.1038/s43018-019-0018-6) [published Online First: Epub Date].
12. Thomas I, Gregg B. Metformin; a review of its history and future: from lilac to longevity. *Pediatr Diabetes* 2017;18:10–6. doi: [10.1111/pedi.12473](https://doi.org/10.1111/pedi.12473) [published Online First: Epub Date].
13. Decensi A, Puntoni M, Goodwin P, et al. Metformin and cancer risk in diabetic patients: a systematic review and meta-analysis. *Cancer Prev Res* 2010;3:1451–61. doi: [10.1158/1940-6207.CAPR-10-0157](https://doi.org/10.1158/1940-6207.CAPR-10-0157) [published Online First: Epub Date].
14. Palazzolo G, Mollica H, Lusi V, et al. Modulating the distant spreading of patient-derived colorectal cancer cells via aspirin and metformin. *Transl Oncol* 2020;13:100760. doi: [10.1016/j.tranon.2020.100760](https://doi.org/10.1016/j.tranon.2020.100760) [published Online First: Epub Date].
15. Arrieta O, Barron F, Padilla MS, et al. Effect of metformin plus tyrosine kinase inhibitors compared with tyrosine kinase inhibitors alone in patients with epidermal growth factor receptor-mutated lung adenocarcinoma: a phase 2 randomized clinical trial. *JAMA Oncol* 2019;e192553. doi: [10.1001/jamaoncol.2019.2553](https://doi.org/10.1001/jamaoncol.2019.2553) [published Online First: Epub Date].
16. Parikh AB, Marrone KA, Becker DJ, et al. A pooled analysis of two phase II trials evaluating metformin plus platinum-based chemotherapy in advanced non-small cell lung cancer. *Cancer Treat Res Commun* 2019;20:100150. doi: [10.1016/j.ctarc.2019.100150](https://doi.org/10.1016/j.ctarc.2019.100150) [published Online First: Epub Date].
17. De A, Kuppusamy G. Metformin in breast cancer: preclinical and clinical evidence. *Curr Probl Cancer* 2020;44:100488. doi: [10.1016/j.currprobcancer.2019.06.003](https://doi.org/10.1016/j.currprobcancer.2019.06.003) [published Online First: Epub Date].
18. Pacal L, Kankova K. Metformin in oncology—how far is its repurposing as an anticancer drug? *Klin Onkol* 2020;33:107–13. doi: [10.14735/amko2020107](https://doi.org/10.14735/amko2020107) [published Online First: Epub Date].
19. Spiering MJ. The mystery of metformin. *J Biol Chem* 2019;294:6689–91. doi: [10.1074/jbc.CL119.008628](https://doi.org/10.1074/jbc.CL119.008628) [published Online First: Epub Date].
20. Lord SR, Collins JM, Cheng WC, et al. Transcriptomic analysis of human primary breast cancer identifies fatty acid oxidation as a target for metformin. *Br J Cancer* 2020;122:258–65. doi: [10.1038/s41416-019-0665-5](https://doi.org/10.1038/s41416-019-0665-5) [published Online First: Epub Date].
21. Shehata M, Kim H, Vellanki R, et al. Identifying the murine mammary cell target of metformin exposure. *Commun Biol* 2019;2:192. doi: [10.1038/s42003-019-0439-x](https://doi.org/10.1038/s42003-019-0439-x) [published Online First: Epub Date].
22. Lord SR, Cheng WC, Liu D, et al. Integrated pharmacodynamic analysis identifies two metabolic adaptation pathways to metformin in breast cancer. *Cell Metab* 2018;28:679, e4–88. doi: [10.1016/j.cmet.2018.08.021](https://doi.org/10.1016/j.cmet.2018.08.021) [published Online First: Epub Date].
23. Schein CH. Repurposing approved drugs on the pathway to novel therapies. *Med Res Rev* 2020;40:586–605. doi: [10.1002/med.21627](https://doi.org/10.1002/med.21627) [published Online First: Epub Date].

24. Zhu YX, Kortuem KM, Stewart AK. Molecular mechanism of action of immune-modulatory drugs thalidomide, lenalidomide and pomalidomide in multiple myeloma. *Leuk Lymphoma* 2013;54:683–7. doi: [10.3109/10428194.2012.728597](https://doi.org/10.3109/10428194.2012.728597) [published Online First: Epub Date].
25. Corral LG, Kaplan G. Immunomodulation by thalidomide and thalidomide analogues. *Ann Rheum Dis* 1999;58:1107–13.
26. Corral LG, Haslett PA, Muller GW, et al. Differential cytokine modulation and T cell activation by two distinct classes of thalidomide analogues that are potent inhibitors of TNF-alpha. *J Immunol* 1999;163:380–6.
27. De Souza A, Strober BE, Merola JF, et al. Apremilast for discoid lupus erythematosus: results of a phase 2, open-label, single-arm, pilot study. *J Drugs Dermatol* 2012;11:1224–6.
28. Massimi M, Ragusa F, Cardarelli S, et al. Targeting cyclic AMP signalling in hepatocellular carcinoma. *Cell* 2019;8. doi: [10.3390/cells8121511](https://doi.org/10.3390/cells8121511) [published Online First: Epub Date].
29. Youghare I, Belemnaba L, Morin C, et al. NCS 613, a potent PDE4 inhibitor, displays anti-inflammatory and anti-proliferative properties on A549 lung epithelial cells and human lung adenocarcinoma explants. *Front Pharmacol* 2020;11:1266. doi: [10.3389/fphar.2020.01266](https://doi.org/10.3389/fphar.2020.01266) [published Online First: Epub Date].
30. Martiniuk F, Giovanazzo J, Tan AU, et al. Lessons of leprosy: the emergence of TH17 cytokines during type II reactions (ENL) is teaching us about T-cell plasticity. *Journal of drugs in dermatology: JDD* 2012;11:626–30.
31. Petzold G, Fischer ES, Thoma NH. Structural basis of lenalidomide-induced CK1alpha degradation by the CRL4(CRBN) ubiquitin ligase. *Nature* 2016;532:127–30. doi: [10.1038/nature16979](https://doi.org/10.1038/nature16979) [published Online First: Epub Date].
32. Kunchok A, Aksamit AJ Jr, Davis JM 3rd, et al. Association between tumor necrosis factor inhibitor exposure and inflammatory central nervous system events. *JAMA Neurol* 2020;77:937–46. doi: [10.1001/jamaneurol.2020.1162](https://doi.org/10.1001/jamaneurol.2020.1162) [published Online First: Epub Date].
33. Mori T, Ito T, Liu S, et al. Structural basis of thalidomide enantiomer binding to cereblon. *Sci Rep* 2018;8:1294. doi: [10.1038/s41598-018-19202-7](https://doi.org/10.1038/s41598-018-19202-7) [published Online First: Epub Date].
34. Chamberlain PP, Cathers BE. Cereblon modulators: low molecular weight inducers of protein degradation. *Drug discovery today. Dent Tech* 2019;31:29–34. doi: [10.1016/j.ddtec.2019.02.004](https://doi.org/10.1016/j.ddtec.2019.02.004) [published Online First: Epub Date].
35. Gandhi AK, Kang J, Havens CG, et al. Immunomodulatory agents lenalidomide and pomalidomide co-stimulate T cells by inducing degradation of T cell repressors Ikaros and Aiolos via modulation of the E3 ubiquitin ligase complex CRL4(CRBN). *Br J Haematol* 2014;164:811–21. doi: [10.1111/bjh.12708](https://doi.org/10.1111/bjh.12708) [published Online First: Epub Date].
36. Schellekens H, de Reus A, Bolhuis R, et al. Comparative antiviral efficiency of leukocyte and bacterially produced human alpha-interferon in rhesus monkeys. *Nature* 1981;292:775–6.
37. Weissmann C, Nagata S, Boll W, et al. Structure and expression of human alpha-interferon genes. In: Merrigan TC (ed.). *Interferons*. Academic Press, 1982;295–396
38. Palva I, Lehtovaara P, Kaariainen L, et al. Secretion of interferon by *Bacillus subtilis*. *Gene* 1983;22:229–35.
39. Schein CH. The shape of the messenger: using protein structure information to design novel cytokine-based therapeutics. *Curr Pharm Des* 2002;8:2113–29.
40. Silva WFDJ, Teixeira LLC, Rocha V, et al. Current role of interferon in hairy cell leukemia therapy: a timely decision. *Hematol Transfus Cell Ther* 2019;41:88–90. doi: [10.1016/j.htct.2018.04.004](https://doi.org/10.1016/j.htct.2018.04.004) [published Online First: Epub Date].
41. Bewersdorf JP, Giri S, Wang R, et al. Interferon therapy in myelofibrosis: systematic review and meta-analysis. *Clin Lymphoma Myeloma Leuk* 2020;20:e712–23. doi: [10.1016/j.clml.2020.05.018](https://doi.org/10.1016/j.clml.2020.05.018) [published Online First: Epub Date].
42. Heibl S, Buxhofer-Ausch V, Schmidt S, et al. A phase 1 study to evaluate the feasibility and efficacy of the addition of ropeginterferon alpha 2b to imatinib treatment in patients with chronic phase chronic myeloid leukemia not achieving a deep molecular response (MR 4.5)—AGMT_CML 1. *Hematol Oncol* 2020. doi: [10.1002/hon.2786](https://doi.org/10.1002/hon.2786) [published Online First: Epub Date].
43. Vazquez-Blomquist D, Leenstra S, van der Kaaij M, et al. A co-formulation of interferons type I and II enhances temozolomide response in glioblastoma with unmethylated MGMT promoter status. *Mol Biol Rep* 2020;47:5263–71. doi: [10.1007/s11033-020-05604-2](https://doi.org/10.1007/s11033-020-05604-2) [published Online First: Epub Date].
44. Jin N, Lera RF, Yan RE, et al. Chromosomal instability upregulates interferon in acute myeloid leukemia. *Genes Chromosomes Cancer* 2020;59:627–38. doi:

- 10.1002/gcc.22880 [published Online First: Epub Date].
45. Harabuchi S, Kosaka A, Yajima Y, et al. Intratumoral STING activations overcome negative impact of cisplatin on antitumor immunity by inflaming tumor microenvironment in squamous cell carcinoma. *Biochem Biophys Res Commun* 2020;522:408–14. doi: [10.1016/j.bbrc.2019.11.107](https://doi.org/10.1016/j.bbrc.2019.11.107) [published Online First: Epub Date].
 46. Ritter JL, Zhu Z, Thai TC, et al. Phosphorylation of RAB7 by TBK1/IKKepsilon regulates innate immune Signaling in triple-negative breast cancer. *Cancer Res* 2020;80:44–56. doi: [10.1158/0008-5472.CAN-19-1310](https://doi.org/10.1158/0008-5472.CAN-19-1310) [published Online First: Epub Date].
 47. Schein CH. From interleukin families to glycans: relating cytokine structure to function. *Curr Pharm Des* 2004;10:3853–5. doi: [10.2174/1381612043382512](https://doi.org/10.2174/1381612043382512) [published Online First: Epub Date].
 48. Aggarwal BB, Gupta SC, Kim JH. Historical perspectives on tumor necrosis factor and its superfamily: 25 years later, a golden journey. *Blood* 2012;119:651–65. doi: [10.1182/blood-2011-04-325225](https://doi.org/10.1182/blood-2011-04-325225) [published Online First: Epub Date].
 49. Li G, Sachdev U, Peters K, et al. The VE-PTP inhibitor AKB-9778 improves antitumor activity and diminishes the toxicity of interleukin 2 (IL-2) administration. *J Immunother* 2019;42:237–43. doi: [10.1097/CJI.0000000000000290](https://doi.org/10.1097/CJI.0000000000000290) [published Online First: Epub Date].
 50. Pachella LA, Madsen LT, Dains JE. The toxicity and benefit of various dosing strategies for interleukin-2 in metastatic melanoma and renal cell carcinoma. *J Adv Pract Oncol* 2015;6:212–21.
 51. Payne R, Glenn L, Hoen H, et al. Durable responses and reversible toxicity of high-dose interleukin-2 treatment of melanoma and renal cancer in a community hospital biotherapy program. *J Immunother Cancer* 2014;2. doi: [10.1186/2051-1426-2-13](https://doi.org/10.1186/2051-1426-2-13) [published Online First: Epub Date].
 52. Alwan LM, Grossmann K, Sageser D, et al. Comparison of acute toxicity and mortality after two different dosing regimens of high-dose interleukin-2 for patients with metastatic melanoma. *Target Oncol* 2014;9:63–71. doi: [10.1007/s11523-013-0276-7](https://doi.org/10.1007/s11523-013-0276-7) [published Online First: Epub Date].
 53. Buchbinder EI, Dutcher JP, Daniels GA, et al. Therapy with high-dose interleukin-2 (HD IL-2) in metastatic melanoma and renal cell carcinoma following PD1 or PDL1 inhibition. *J Immunother Cancer* 2019;7:49. doi: [10.1186/s40425-019-0522-3](https://doi.org/10.1186/s40425-019-0522-3) [published Online First: Epub Date].
 54. Carey PD, Wakefield CH, Guillou PJ. Neutrophil activation, vascular leak toxicity, and cytotoxicity during interleukin-2 infusion in human cancer. *Surgery* 1997;122:918–26. doi: [10.1016/s0039-6060\(97\)90333-0](https://doi.org/10.1016/s0039-6060(97)90333-0) [published Online First: Epub Date].
 55. Maybauer DM, Maybauer MO, Szabo C, et al. Lung-protective effects of the metalloporphyrinic peroxy-nitrite decomposition catalyst WW-85 in interleukin-2 induced toxicity. *Biochem Biophys Res Commun* 2008;377:786–91. doi: [10.1016/j.bbrc.2008.10.066](https://doi.org/10.1016/j.bbrc.2008.10.066) [published Online First: Epub Date].
 56. Donia M, Kjeldsen JW, Svane IM. The controversial role of TNF in melanoma. *Onco Targets Ther* 2016; 5:e1107699. doi: [10.1080/2162402X.2015.1107699](https://doi.org/10.1080/2162402X.2015.1107699) [published Online First: Epub Date].
 57. Puchta A, Naidoo A, Verschoor CP, et al. TNF drives monocyte dysfunction with age and results in impaired anti-pneumococcal immunity. *PLoS Pathog* 2016;12:e1005368. doi: [10.1371/journal.ppat.1005368](https://doi.org/10.1371/journal.ppat.1005368) [published Online First: Epub Date].
 58. Verschoor CP, Loukov D, Naidoo A, et al. Circulating TNF and mitochondrial DNA are major determinants of neutrophil phenotype in the advanced-age, frail elderly. *Mol Immunol* 2015;65:148–56. doi: [10.1016/j.molimm.2015.01.015](https://doi.org/10.1016/j.molimm.2015.01.015) [published Online First: Epub Date].
 59. Haderski GJ, Kandar BM, Brackett CM, et al. TLR5 agonist entolimod reduces the adverse toxicity of TNF while preserving its antitumor effects. *PLoS One* 2020;15:e0227940. doi: [10.1371/journal.pone.0227940](https://doi.org/10.1371/journal.pone.0227940) [published Online First: Epub Date].
 60. Abdullah MI, de Wolf E, Jawad MJ, et al. The poor design of clinical trials of statins in oncology may explain their failure—lessons for drug repurposing. *Cancer Treat Rev* 2018;69:84–9. doi: [10.1016/j.ctrv.2018.06.010](https://doi.org/10.1016/j.ctrv.2018.06.010) [published Online First: Epub Date].
 61. Blumenthal GM, Gills JJ, Ballas MS, et al. A phase I trial of the HIV protease inhibitor nelfinavir in adults with solid tumors. *Oncotarget* 2014;5:8161–72. doi: [10.18632/oncotarget.2415](https://doi.org/10.18632/oncotarget.2415) [published Online First: Epub Date].
 62. Ran Y, Hossain F, Pannuti A, et al. Gamma-secretase inhibitors in cancer clinical trials are pharmacologically and functionally distinct. *EMBO Mol Med* 2017;9:950–66. doi: [10.15252/emmm.201607265](https://doi.org/10.15252/emmm.201607265) [published Online First: Epub Date].

63. Nygaard HB, Wagner AF, Bowen GS, et al. A phase Ib multiple ascending dose study of the safety, tolerability, and central nervous system availability of AZD0530 (saracatinib) in Alzheimer's disease. *Alzheimers Res Ther* 2015;7:35. doi: [10.1186/s13195-015-0119-0](https://doi.org/10.1186/s13195-015-0119-0) [published Online First: Epub Date].
64. Vasconcelos I, Wiesske A, Schoenegg W. Scalp cooling successfully prevents alopecia in breast cancer patients undergoing anthracycline/taxane-based chemotherapy. *Breast* 2018;40:1–3. doi: [10.1016/j.breast.2018.04.012](https://doi.org/10.1016/j.breast.2018.04.012) [published Online First: Epub Date].
65. Veneris JT, Matulonis UA, Liu JF, et al. Choosing wisely: selecting PARP inhibitor combinations to promote anti-tumor immune responses beyond BRCA mutations. *Gynecol Oncol* 2020;156:488–97. doi: [10.1016/j.ygyno.2019.09.021](https://doi.org/10.1016/j.ygyno.2019.09.021) [published Online First: Epub Date].
66. Deangelo DJ. Managing chronic myeloid leukemia patients intolerant to tyrosine kinase inhibitor therapy. *Blood Cancer J* 2012;2:e95. doi: [10.1038/bcj.2012.30](https://doi.org/10.1038/bcj.2012.30) [published Online First: Epub Date].
67. Chalabi M, Cardona A, Nagarkar DR, et al. Efficacy of chemotherapy and atezolizumab in patients with non-small-cell lung cancer receiving antibiotics and proton pump inhibitors: pooled post hoc analyses of the OAK and POPLAR trials. *Ann Oncol* 2020;31:525–31. doi: [10.1016/j.annonc.2020.01.006](https://doi.org/10.1016/j.annonc.2020.01.006) [published Online First: Epub Date].
68. Moran TB, Plana JC. Management of Patients with acute coronary syndrome and cancer. *Curr Cardiol Rep* 2020;22:159. doi: [10.1007/s11886-020-01409-8](https://doi.org/10.1007/s11886-020-01409-8) [published Online First: Epub Date].
69. Regulska K, Regulski M, Karolak B, et al. Beyond the boundaries of cardiology: still untapped anticancer properties of the cardiovascular system-related drugs. *Pharmacol Res* 2019;147:104326. doi: [10.1016/j.phrs.2019.104326](https://doi.org/10.1016/j.phrs.2019.104326) [published Online First: Epub Date].
70. McNeil JJ, Gibbs P, Orchard SG, et al. Effect of aspirin on cancer incidence and mortality in older adults. *J Natl Cancer Inst* 2020. doi: [10.1093/jnci/djaa114](https://doi.org/10.1093/jnci/djaa114) [published Online First: Epub Date].
71. Spini A, Roberto G, Gini R, et al. Evidence of beta-blockers drug repurposing for the treatment of triple negative breast cancer: a systematic review. *Neoplasma* 2019;66:963–70. doi: [10.4149/neo_2019_190110N34](https://doi.org/10.4149/neo_2019_190110N34) [published Online First: Epub Date].
72. Hagen R, Ghareeb E, Jalali O, et al. Infantile hemangiomas: what have we learned from propranolol? *Curr Opin Pediatr* 2018;30:499–504. doi: [10.1097/MOP.0000000000000650](https://doi.org/10.1097/MOP.0000000000000650) [published Online First: Epub Date].
73. Price A, Rai S, McLeod RWJ, et al. Topical propranolol for infantile haemangiomas: a systematic review. *J Eur Acad Dermatol Venereol* 2018;32:2083–9. doi: [10.1111/jdv.14963](https://doi.org/10.1111/jdv.14963) [published Online First: Epub Date].
74. Wagner MJ, Cranmer LD, Loggers ET, et al. Propranolol for the treatment of vascular sarcomas. *J Exp Pharmacol* 2018;10:51–8. doi: [10.2147/JEP.S146211](https://doi.org/10.2147/JEP.S146211) [published Online First: Epub Date].
75. Cho MA, Jeong SY, Sohn I, et al. Impact of angiotensin receptor blockers, beta blockers, calcium channel blockers and thiazide diuretics on survival of ovarian cancer patients. *Cancer Res Treat* 2020;52:645–54. doi: [10.4143/crt.2019.509](https://doi.org/10.4143/crt.2019.509) [published Online First: Epub Date].
76. Coelho M, Squizzato A, Cassina N, et al. Effect of beta-blockers on survival of lung cancer patients: a systematic review and meta-analysis. *Eur J Cancer Prev* 2020;29:306–14. doi: [10.1097/CEJ.0000000000000544](https://doi.org/10.1097/CEJ.0000000000000544) [published Online First: Epub Date].
77. Yap A, Lopez-Olivo MA, Dubowitz J, et al. Effect of beta-blockers on cancer recurrence and survival: a meta-analysis of epidemiological and perioperative studies. *Br J Anaesth* 2018;121:45–57. doi: [10.1016/j.bja.2018.03.024](https://doi.org/10.1016/j.bja.2018.03.024) [published Online First: Epub Date].
78. Kheiri B, Abdalla A, Osman M, et al. Meta-analysis of carvedilol for the prevention of anthracycline-induced cardiotoxicity. *Am J Cardiol* 2018;122:1959–64. doi: [10.1016/j.amjcard.2018.08.039](https://doi.org/10.1016/j.amjcard.2018.08.039) [published Online First: Epub Date].
79. Shah P, Garris R, Abboud R, et al. Meta-analysis comparing usefulness of beta blockers to preserve left ventricular function during anthracycline therapy. *Am J Cardiol* 2019;124:789–94. doi: [10.1016/j.amjcard.2019.05.046](https://doi.org/10.1016/j.amjcard.2019.05.046) [published Online First: Epub Date].
80. Sibaud V, Casassa E, D'Andrea M. Are topical beta-blockers really effective "in real life" for targeted therapy-induced paronychia. *Supportive Care Cancer* 2019;27:2341–3. doi: [10.1007/s00520-019-04690-8](https://doi.org/10.1007/s00520-019-04690-8) [published Online First: Epub Date].
81. Jordan K, Behlendorf T, Mueller F, et al. Anthracycline extravasation injuries: management with dexrazoxane. *Ther Clin Risk Manag* 2009;5:361–6. doi: [10.2147/tcrm.s3694](https://doi.org/10.2147/tcrm.s3694) [published Online First: Epub Date].

82. Langer SW. Dexrazoxane for the treatment of chemotherapy-related side effects. *Cancer Manag Res* 2014;6:357–63. doi: [10.2147/CMAR.S47238](https://doi.org/10.2147/CMAR.S47238) [published Online First: Epub Date].
83. Getz KD, Sung L, Alonzo TA, et al. Effect of dexrazoxane on left ventricular systolic function and treatment outcomes in patients with acute myeloid leukemia: a report from the Children's Oncology Group. *J Clin Oncol* 2020;38:2398–406. doi: [10.1200/JCO.19.02856](https://doi.org/10.1200/JCO.19.02856) [published Online First: Epub Date].
84. Krouse AJ, Gray L, Macdonald T, et al. Repurposing and rescuing of Mibefradil, an antihypertensive, for cancer: a case study. *Assay Drug Dev Technol* 2015;13:650–3. doi: [10.1089/adt.2015.29014.ajkdr](https://doi.org/10.1089/adt.2015.29014.ajkdr) [published Online First: Epub Date].
85. Wang H, Pan R, Zhang X, et al. Abivertinib in patients with T790M-positive advanced NSCLC and its subsequent treatment with osimertinib. *Thoracic cancer* 2020;11:594–602. doi: [10.1111/1759-7714.13302](https://doi.org/10.1111/1759-7714.13302) [published Online First: Epub Date].
86. Girdwood RH. The relationships between vitamin B12, folic acid and folinic acid. *Br J Nutr* 1952;6:315–24. doi: [10.1079/bjn19520033](https://doi.org/10.1079/bjn19520033) [published Online First: Epub Date].
87. Selvy M, Pereira B, Kerckhove N, et al. Long-term prevalence of sensory chemotherapy-induced peripheral neuropathy for 5 years after adjuvant FOLFOX chemotherapy to treat colorectal cancer: a Multicenter cross-sectional study. *J Clin Med* 2020;9. doi: [10.3390/jcm9082400](https://doi.org/10.3390/jcm9082400) [published Online First: Epub Date].
88. Elhassanny AEM, Soliman E, Marie M, et al. Heme-dependent ER stress apoptosis: a mechanism for the selective toxicity of the dihydroartemisinin, NSC735847, in colorectal cancer cells. *Front Oncol* 2020;10:965. doi: [10.3389/fonc.2020.00965](https://doi.org/10.3389/fonc.2020.00965) [published Online First: Epub Date].
89. Fazeel ZA. Apixaban: an oral anticoagulant having unique mechanism of action with better safety and efficacy profile. *MAMC J Med Sci* 2016;2:63–8. doi: [10.4103/2394-7438.182723](https://doi.org/10.4103/2394-7438.182723) [published Online First: Epub Date].
90. Bhardwaj M, Leli NM, Koumenis C, et al. Regulation of autophagy by canonical and non-canonical ER stress responses. *Semin Cancer Biol* 2019. doi: [10.1016/j.semcancer.2019.11.007](https://doi.org/10.1016/j.semcancer.2019.11.007) [published Online First: Epub Date].
91. Martelli AM, Paganelli F, Chiarini F, et al. The unfolded protein response: a novel therapeutic target in acute leukemias. *Cancer* 2020;12. doi: [10.3390/cancers12020333](https://doi.org/10.3390/cancers12020333) [published Online First: Epub Date].
92. Gelman JS, Sironi J, Berezniuk I, et al. Alterations of the intracellular peptidome in response to the proteasome inhibitor bortezomib. *PLoS One* 2013;8:e53263. doi: [10.1371/journal.pone.0053263](https://doi.org/10.1371/journal.pone.0053263) [published Online First: Epub Date].
93. Joshi J, Tanner L, Gilchrist L, et al. Switching to bortezomib may improve recovery from severe vincristine neuropathy in pediatric acute lymphoblastic Leukemia. *J Pediatr Hematol Oncol* 2019;41:457–62. doi: [10.1097/MPH.0000000000001529](https://doi.org/10.1097/MPH.0000000000001529) [published Online First: Epub Date].
94. Dennis M, Maoz A, Hughes D, et al. Bortezomib ocular toxicities: outcomes with ketotifen. *Am J Hematol* 2019;94:E80–2. doi: [10.1002/ajh.25382](https://doi.org/10.1002/ajh.25382) [published Online First: Epub Date].
95. Borcherding N, Jethava Y, Vikas P. Repurposing anti-cancer drugs for COVID-19 treatment. *Drug Des Devel Ther* 2020;14:5045–58. doi: [10.2147/DDDT.S282252](https://doi.org/10.2147/DDDT.S282252) [published Online First: Epub Date].
96. Stebbing J, Sanchez Nieves G, Falcone M, et al. JAK inhibition reduces SARS-CoV-2 liver infectivity and modulates inflammatory responses to reduce morbidity and mortality. *Sci Adv* 2020. doi: [10.1126/sciadv.abe4724](https://doi.org/10.1126/sciadv.abe4724) [published Online First: Epub Date].
97. Zhang Q, Bastard P, Liu Z, et al. Inborn errors of type I IFN immunity in patients with life-threatening COVID-19. *Science* 2020;370. doi: [10.1126/science.abd4570](https://doi.org/10.1126/science.abd4570) [published Online First: Epub Date].
98. Bastard P, Rosen LB, Zhang Q, et al. Autoantibodies against type I IFNs in patients with life-threatening COVID-19. *Science* 2020;370. doi: [10.1126/science.abd4585](https://doi.org/10.1126/science.abd4585) [published Online First: Epub Date].
99. Major J, Crotta S, Llorian M, et al. Type I and III interferons disrupt lung epithelial repair during recovery from viral infection. *Science* 2020;369:712–7. doi: [10.1126/science.abc2061](https://doi.org/10.1126/science.abc2061) [published Online First: Epub Date].
100. Kaneko N, Kuo H-H, Boucau J, et al. Loss of Bcl-6-expressing T follicular helper cells and germinal centers in COVID-19. *Cell* 2020;183:143–57 e13. doi: [10.1016/j.cell.2020.08.025](https://doi.org/10.1016/j.cell.2020.08.025) [published Online First: Epub Date].
101. Van der Voort A, Van Ramshorst MS, Van Werkhoven ED, et al. Three-year follow-up of neoadjuvant chemotherapy with or without anthracyclines in the

- presence of dual HER2-blockade for HER2-positive breast cancer (TRAIN-2): a randomized phase III trial. *J Clin Oncol* 2020;38:501–1. doi: [10.1200/JCO.2020.38.15_suppl.501](https://doi.org/10.1200/JCO.2020.38.15_suppl.501) [published Online First: Epub Date].
102. Chapman PB, Einhorn LH, Meyers ML, et al. Phase III multicenter randomized trial of the Dartmouth regimen versus dacarbazine in patients with metastatic melanoma. *J Clin Oncol* 1999;17:2745–51. doi: [10.1200/JCO.1999.17.9.2745](https://doi.org/10.1200/JCO.1999.17.9.2745) [published Online First: Epub Date].
103. Smylie MG. Use of immuno-oncology in melanoma. *Curr Oncol* 2020;27:S51–8. doi: [10.3747/co.27.5135](https://doi.org/10.3747/co.27.5135) [published Online First: Epub Date].
104. Wong W, Bai XC, Brown A, et al. Cryo-EM structure of the plasmodium falciparum 80S ribosome bound to the anti-protozoan drug emetine. *Elife* 2014;3. doi: [10.7554/eLife.03080](https://doi.org/10.7554/eLife.03080) [published Online First: Epub Date].
105. Cornet-Masana JM, Moreno-Martinez D, Lara-Castillo MC, et al. Emetine induces chemosensitivity and reduces clonogenicity of acute myeloid leukemia cells. *Oncotarget* 2016;7:23239–50. doi: [10.18632/oncotarget.8096](https://doi.org/10.18632/oncotarget.8096) [published Online First: Epub Date].
106. Pantziarka P, Verbaanderd C, Huys I, et al. Repurposing drugs in oncology: from candidate selection to clinical adoption. *Semin Cancer Biol* 2020. doi: [10.1016/j.semcancer.2020.01.008](https://doi.org/10.1016/j.semcancer.2020.01.008) [published Online First: Epub Date].
107. Gonzalez-Cao M, Boada A, Teixido C, et al. Fatal gastrointestinal toxicity with ipilimumab after BRAF/MEK inhibitor combination in a melanoma patient achieving pathological complete response. *Oncotarget* 2016;7:56619–27. doi: [10.18632/oncotarget.10651](https://doi.org/10.18632/oncotarget.10651) [published Online First: Epub Date].
108. Magge T, Shaikh H, Chaudhary R. Complete response to temozolomide in metastatic melanoma after failure of 5 lines of treatment. *Am J Ther* 2020. doi: [10.1097/MJT.0000000000001186](https://doi.org/10.1097/MJT.0000000000001186) [published Online First: Epub Date].
109. Kronish IM, Cheung YK, Shimbo D, et al. Increasing the precision of hypertension treatment through personalized trials: a pilot study. *J Gen Intern Med* 2019;34:839–45. doi: [10.1007/s11606-019-04831-z](https://doi.org/10.1007/s11606-019-04831-z) [published Online First: Epub Date].