1 **Responses of grassland arthropods to an invasion by nonnative grasses**

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Abstract

In grassland ecosystems, invasions by nonnative grasses typically decrease floristic diversity and

structural heterogeneity in ways that alter the quantity and quality of habitat for animals. Grassland

arthropods that rely directly on herbaceous plants for food, shelter, or as substrates for reproduction

are especially vulnerable to these invasions because many have evolved specialized relationships with

host plants that might be displaced. We evaluated how invasions by nonnative grasses affected

abundance and richness of foliage-dwelling arthropods in semidesert grasslands of Arizona, USA. On 90,

3.1-ha plots established along a gradient of invasion where dominance of nonnative grasses ranged from

0 to nearly 100% of grass cover, we captured >90,000 arthropods from 11 orders during 270 surveys in

2014 and 2015. Although the invasion by nonnative grasses (primarily *Eragrostis lehmanniana* and

secondarily E. *curvula*) increased the amount of herbaceous foliage available to arthropods, richness of

arthropods decreased by an average of 2% and total abundance by an average of 7% for every 10%

increase in nonnative-grass dominance. Responses to the plant invasion, however, varied among taxa

and functional groups. As dominance of nonnative grasses increased, abundances of most predators and

specialist herbivores decreased, whereas abundances of most generalist herbivores were lowest at

intermediate points of the invasion gradient. The changes we observed in the arthropod community

have potential to alter broad-scale ecological processes, including energy flow and nutrient cycling, and

to reduce food resources for insectivores, which can have adverse, cascading effects on imperiled

grassland ecosystems.

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Introduction

Nonnative grasses have been introduced to almost every grassland or savanna ecosystem in the world, which has had significant impacts on native species (D'Antonio and Vitousek 1992; Steidl et al. 2013). Typically, as nonnative grasses invade grasslands and increase in dominance, floristic diversity and structural heterogeneity decrease (D'Antonio and Vitousek 1992; Vilà et al. 2011), and vegetation cover and biomass increase (Brooks et al. 2004; Geiger 2006). These changes in composition and structure of grassland plant communities alter ecosystem processes, including primary production, decomposition, nutrient and carbon cycling, and hydrologic and disturbance regimes (Vitousek et al. 1996; Levine et al. 2003; Bradley et al. 2006). Consequently, invasions by nonnative grasses pose a serious threat to remaining grasslands, which are among the most biologically diverse and most threatened ecosystems in the world (Wilson et al. 2012; Murphy et al. 2016).

Collectively, changes in ecosystem structure, composition, and processes associated with plant invasions can affect the quantity and quality of habitat for animals that inhabit grasslands (Steidl et al. 2013). Arthropods may be especially vulnerable to changes in the plant community that result from invasions because many have limited mobility or have evolved specialized relationships with host plants that provide food or substrates for reproduction (Bernays and Graham 1988). For example, approximately 90% of all phytophagous insects feed on plants in only one or a few plant lineages, and these host plants may be displaced or reduced in abundance when novel plants invade a community (Strong et al. 1984; Bernays and Graham 1988; Tallamy 2004). Additionally, survival, reproduction, or distributions of all arthropods, including those not associated closely with specific plants, can change in response to invasions by nonnative plants if vegetation structure is altered in ways that affect microclimatic conditions, including light intensity, temperature, and soil moisture (Wolkovich 2010; Schirmel and Buchholz 2013). These changes in abundance and community composition of arthropods can affect broad-scale ecological processes, including decomposition, nutrient cycling, and pollination

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67   (Hladik et al. 2015; Nitschke et al. 2015), and are likely to have disproportionately strong effects on food

68   webs because many taxa are key prey for insectivores. Grasshoppers (Orthoptera), for example, are the

69   primary prey of many breeding grassland birds (George and McEwen 1992), a group that has declined

70   more rapidly than any other group of birds in North America (Sauer and Link 2011).

71   Effects of plant invasions on arthropods may be strongest where invading plants have reduced

72   vegetation heterogeneity markedly (Hovick et al. 2015), such as in semidesert grasslands of North

73   America where two perennial C_4 grasses have come to dominate grasslands once characterized by a

74   diverse flora of native grasses (Litt and Steidl 2011). *Eragrostis lehmanniana* and *E. curvula* were

75   introduced to southeast Arizona from southern Africa in the 1930s and 1940s in an attempt to

76   revegetate degraded rangelands and mitigate soil erosion after prolonged drought (Anable et al. 1992).

77   These two species have spread rapidly and are predicted to expand to an area >71,000 km^2 in southern

78   Arizona and New Mexico alone (Anable et al. 1992; Schussman et al. 2006). Compared to most native

79   grasses, these nonnative species grow more rapidly, produce more seeds and up to four times more

80   aboveground biomass, and are less palatable to vertebrate herbivores (Cox et al. 1990; Geiger 2006).

81   These attributes can increase fire intensity and frequency, a primary ecological process that governs

82   spatial and temporal patterns of biodiversity in many grassland ecosystems (McPherson 1995; Brooks et

83   al. 2004; Steidl et al. 2013).

84   Although invasions by these nonnative grasses have been shown to alter richness and

85   abundance of many vertebrate taxa, effects on grassland arthropods are less well-understood (Steidl et

86   al. 2013; Litt et al. 2014). We know, however, that richness and abundance of epigeic (i.e., surface-

87   dwelling) insects decrease as dominance of *E. lehmanniana* increases, although responses vary among

88   taxa (Litt and Steidl 2010). Here, we sought to understand the effects of these invasions on arthropods

89   that use grasses or other herbaceous plants as their primary substrate (hereafter, foliage-dwelling

90   arthropods). This group may be especially vulnerable to changes in grassland structure or floristics that

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91   accompany invasions because of their direct reliance on herbaceous plants for food, shelter, or

92   reproduction (Gleditsch 2016). Specifically, we evaluated how abundance and richness of foliage-

93   dwelling arthropods changed along a gradient of dominance by nonnative grasses in semidesert

94   grasslands. This gradient represents a space-for-time substitution for the process by which nonnative-

95   grass invasions transform grassland plant communities once composed of a diverse assemblage of

96   native grass species into near-monocultures with few native species.

97

98   **Methods**

99   Study area

100   We surveyed arthropods and vegetation in southeastern Arizona, USA at three sites that span the range

101   of variation in vegetation structure and management of semidesert grasslands in North America:

102   Appleton-Whittell Research Ranch (31.6° N, 110.5° W), Fort Huachuca Military Reservation (31.6° N,

103   110.3° W), and Las Cienegas National Conservation Area (31.8° N, 110.6° W). Sites are located within a

104   40 x 55 km area where semidesert grasslands occur between 1350 and 1550 m elevation and annual

105   precipitation averages 350 to 450 mm, most of which falls during a monsoon season between July and

106   September that is preceded by two to three months that are hot and dry (McClaran 1995). Livestock

107   have been excluded from Fort Huachuca Military Reservation since 1950 and Appleton-Whittell

108   Research Ranch since 1968. Grasslands at Las Cienegas National Conservation Area are grazed by cattle,

109   therefore we excluded areas that were scheduled to be grazed during the study.

110   Vegetation composition at all sites was dominated by a variety of C_4 perennial grasses. Common

111   native grasses included *Aristida* spp., *Bothriochloa barbinodis*, *Bouteloua* spp., *Digitaria californica*, and

112   *Eragrostis intermedia*. At each site, composition of perennial grasses ranged from 0 to nearly 100%

113   nonnative species, with areas dominated by nonnative species occurring throughout each site. Of the

114   nonnative grasses, *E. lehmanniana* was the most widespread and often dominated large areas, although

E. curvula was abundant locally. Woody plants were composed almost entirely of native species (>99%

of woody cover), especially *Baccharis* spp., *Isocoma tenuisecta*, *Mimosa* spp., and *Prosopis velutina*,

which has encroached many grasslands in this region transforming them to shrub savannas (Archer et al.

2017).

Arthropod sampling

We established 90, 100-m-radius circular plots (3.1-ha) at random throughout upland areas of the three

sites. We excluded areas with obvious anthropogenic features (e.g., roads or buildings), those that were

scheduled to be grazed, and those that had been burned or otherwise manipulated to reduce woody

vegetation during the previous three years. We surveyed 50 plots in 2014 and 40 different plots in 2015;

12 plots were located at Appleton-Whittell, 29 at Fort Huachuca, and 49 at Las Cienegas.

We surveyed arthropods on each plot three times between mid-June and mid-August. This

period typically encompasses most monsoon precipitation, which coincides with the growing season for

grasses and peak abundance of many arthropod taxa in semidesert grasslands (Whitford et al. 1995). To

target foliage-dwelling arthropods, we used a 38-cm sweep net constructed of heavy sailcloth canvas to

capture arthropods along three 60-m transects radiating from plot center at 120° intervals. Sweep

sampling is well-suited for surveying these arthropods (Gardiner et al. 2005; Yi et al. 2012) and

characterizes relative abundance and community composition accurately (Larson et al. 1999; Kati et al.

2004). To minimize variation among surveyors and variation induced by heterogeneity in vegetation

density or height (Doxon et al. 2011), we standardized our sweeping procedure. Surveyors traversed

each transect at a constant pace and made 30 fast sweeps (one 2-m sweep per pace) as close to the

ground as permitted by vegetation (Neill et al. 2002). We emptied the contents of the net into a labeled

3.8 L freezer bag that we stored on ice soon after collection and transferred to a freezer within three

days.

In the lab, we counted and classified individuals to morphospecies based on recognizable visual characteristics, an efficient approach that produces estimates of richness similar to those obtained by specialists who identify specimens to species (Oliver and Beattie 1993, 1996). Additionally, we classified most (99.3%) individuals to order and individuals from the three most abundant orders to suborder (Hemiptera) or family (Coleoptera and Orthoptera). We classified taxa thought to be functionally homogeneous into functional groups for analysis (Table 1). Specifically, we classified taxa as generalist herbivores, specialist herbivores, and predators based on information provided in Triplehorn and Johnson (2005) and Litt et al. (2014).

Vegetation sampling

Although our primary objective was to evaluate responses of arthropods to variation in dominance of nonnative grasses, arthropod populations also reflect other vegetation features. Therefore, we characterized species composition and structure of the entire plant community on each plot using a design that complemented the approach we used to sample arthropods. We surveyed vegetation by establishing eight 10-m diameter subplots systematically on each plot, two along each cardinal axis with one 25 m and another 75 m from plot-center. On each subplot, we estimated canopy cover and composition (as percent of total cover) of grasses, canopy cover of woody plants and forbs (i.e., herbaceous dicots), and surface cover of ground litter and bare ground by combining visual estimates from four quadrants. Additionally, we measured grass height at eight points spaced systematically on the periphery of each subplot. We surveyed vegetation in September, when presence of grass inflorescences aided identification. Although we surveyed vegetation later in the season than arthropods, composition of C_4 perennial grasses remains stable during summer months (Geiger 2006), therefore dominance of nonnative grasses likely changed little between surveys. To reduce observer-induced variation, only two observers measured vegetation each year and each measured half of the

163   subplots on every plot. For analysis, we averaged estimates of vegetation cover, composition, and grass

164   height across the eight subplots on each plot.

165

166   Data analysis

167   For all analyses, we quantified dominance of nonnative grasses as the proportion of perennial

168   grass cover comprised by nonnative species. We used simple linear regression to evaluate how

169   vegetation features on plots varied with dominance of nonnative grasses; we used Shannon's Index to

170   characterize diversity of grasses. To reduce the number of non-focal vegetation variables in models, we

171   considered only cover of woody plants, forbs, and grasses as covariates because grass cover was

172   correlated with cover of bare ground (r = -0.90), litter (r = -0.53), and grass height (r = 0.61).

173   Our overarching approach to analysis was to evaluate the effect of dominance of nonnative

174   grasses on each arthropod response after accounting for other sources of variation among plots. As

175   responses, we considered richness of orders, richness of morphospecies for all taxa combined and the

176   three orders with highest abundances, and abundance of all orders combined (hereafter 'total

177   abundance'), each taxa individually, and each functional group. For each response, we began with a

178   generalized linear mixed model that included all vegetation covariates except for nonnative grasses, plus

179   site, plot, and three covariates to describe timing of surveys: year, Julian day, and time-of-day; we

180   considered both linear and quadratic effects for Julian day and time-of-day. We treated all terms as

181   fixed effects except for plot, which we treated as a random effect. We then used backwards elimination

182   to eliminate vegetation and temporal covariates with little explanatory power (P > 0.10 based on Wald

183   Z-tests). Lastly, we added terms for dominance of nonnative grasses (both linear and quadratic forms) to

184   a model that included all remaining covariates, which provided a direct test of the influence of

185   nonnative grasses on each response variable after accounting for other important features (Ramsey and

186   Schafer 2002). We standardized all covariates to Z-scores (mean = 0, standard deviation = 1) to improve

187   numerical convergence of models.

188   To model abundance, we specified a log link function and a negative binomial error distribution

189   because counts of arthropods were overdispersed. To model richness, we specified a log link function

190   and a Poisson error distribution. We fit models with the R package lme4 (Bates et al. 2015). To aid

191   interpretation, we report the percentage change in each response for every 10% increase in dominance

192   of nonnative grasses (Ramsey and Schafer 2002).

193

194   **Results**

195   Vegetation

196   Sites were dominated by perennial grasses and our plots spanned the gradient of invasion by nonnative

197   grasses on each site (Table 2, Fig. 1). Across all sites and plots, composition of perennial grasses

198   averaged 48% native species and 49% nonnative species (3% were unidentified), and ranged from 0 to

199   99% nonnative species (Table 2). Native grasses were represented by 27 species, none of which

200   comprised >10% of perennial grass cover, which illustrates the high degree of diversity in this category.

201   Nonnative grasses were represented by three species that were locally abundant (Table 2). *Eragrostis*

202   *lehmanniana* was by far the most common, comprising 95% of nonnative grass cover and 47% of

203   perennial grass cover across all plots (Table 2). As dominance of nonnative grasses increased, grass

204   diversity decreased markedly, simplifying the plant community (Fig. 1c). In addition to altering species

205   composition, nonnative grasses also altered vegetation structure and reduced vegetation heterogeneity.

206   For every 10% increase in dominance of nonnative grasses, mean grass height increased by 1.5 cm (SE =

207   0.34, t_{88} = 4.59, P < 0.0001) and total grass cover increased by 1.4% (SE = 0.58, t_{88} = 2.47, P = 0.015; Fig.

208   1a, b). Cover of bare ground averaged 18% (Table 2) and decreased by an average of 0.8% (SE = 0.40, t_{88}

209   = 1.98, P = 0.051) for every 10% increase in dominance of nonnative grasses. Cover of forbs averaged

210 10% and woody plants 8% across plots (Table 2), neither of which varied systematically with dominance

211 of nonnative grasses ($t_{88} < 0.95$, $P > 0.35$).

212

213 Arthropods

214 We captured 93,418 arthropods from 11 orders during 270 surveys on 90 plots (Table 1). Four orders

215 comprised 94% of individuals captured: 75% were Hemiptera (true bugs), 9% Orthoptera (grasshoppers,

216 crickets and katydids), 5% Coleoptera (beetles), and 5% Araneae (spiders).

217   Total abundance of arthropods decreased by an average of 7% for every 10% increase in

218 dominance of nonnative grasses (Table 1). Excluding the two most abundant Hemipteran

219 morphospecies, which represented 43% of all individuals captured, did not change the direction of this

220 effect but did reduce its magnitude to 5% (SE = 1.51, X^2 = 8.92, P = 0.003) for every 10% increase in

221 dominance of nonnative grasses.

222   Although total abundance of arthropods decreased as dominance of nonnative grasses

223 increased, effects varied widely among orders. Abundance of 3 of 11 (27%) orders decreased as

224 dominance of nonnative grasses increased (Table 1). Abundance decreased by an average of 9% in

225 Coleoptera, Hemiptera, and Diptera for every 10% increase in dominance of nonnative grasses (Table 1).

226 For Coleoptera, the decrease was driven by decreases in four of the five most common families (Table

227 1). Abundance decreased by an average of 16% in Melyridae (flower beetles), 11% in Cleridae

228 (checkered beetles), 8% in Curculionidae (weevils), and 6% in Chrysomelidae (leaf beetles) for every 10%

229 increase in dominance of nonnative grasses (Table 1, Fig. 2). Among suborders within Hemiptera,

230 abundance decreased by an average of 9% for Auchenorrhyncha (cicadas, hoppers, spittlebugs) and 8%

231 for Heteroptera (true bugs) for every 10% increase in dominance of nonnative grasses (Table 1, Fig. 2).

232   Abundance of 4 of 11 (36%) orders increased as dominance of nonnative grasses increased

233 (Table 1). Abundance increased by an average of 32% in Parasitiformes (ticks), 15% in Mantodea

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234   (mantises), 14% in Neuroptera (net-winged insects), and 5% in Hymenoptera (bees and ants) for every

235   10% increase in dominance of nonnative grasses (Table 1).

236   Abundances did not vary linearly with dominance of nonnative grasses for 3 of 11 (36%) orders:

237   Araneae, Phasmatodea (stick insects), Lepidoptera (butterflies and moths; 86% of which were larvae),

238   and Orthoptera (Table 1). Phasmatodea and Orthoptera, however, varied nonlinearly along the invasion

239   gradient, with abundances being similar at the gradient extremes and lowest at intermediate levels of

240   nonnative grass dominance (Fig. 2, Appendix 1 of Electronic Supplementary Material). For Orthoptera,

241   this pattern was driven by nonlinear responses in two of three families: Acrididae (grasshoppers) and

242   Tettigoniidae (bush crickets; Fig. 2, Appendix 1 of Electronic Supplementary Material).

243   Among functional groups, abundance decreased by an average of 9% for specialist herbivores

244   and 4% for predators for every 10% increase in dominance of nonnative grasses (Table 1, Fig. 3).

245   Abundance of generalist herbivores changed nonlinearly along the invasion gradient, with abundances

246   similar at the gradient extremes and lowest where composition of nonnative grasses was approximately

247   50% (Fig. 3, Appendix 1 of Electronic Supplementary Material).

248   Richness decreased as dominance of nonnative grasses increased for three of the five taxonomic

249   groups we considered (Table 3). Specifically, richness of morphospecies decreased by an average of 2%

250   for all taxa combined, 5% within Coleoptera, and 4% within Orthoptera for every 10% increase in

251   dominance of nonnative grasses (Table 3). Richness of orders and richness within Hemiptera did not

252   vary along the invasion gradient (Table 3).

253

254   **Discussion**

255   In the grasslands we studied, height and cover of grasses increased as nonnative grasses increased in

256   dominance, a pattern similar to that observed in many other invaded grasslands (Brooks et al. 2004).

257   Although these changes increased the quantity of substrate available to foliage-dwelling arthropods,

258   total abundance and richness decreased as dominance of nonnative grasses increased. This suggests

259   that any benefits from increased foliage to this community of arthropods were outweighed by costs

260   associated with changes in vegetation composition or structure, such as reduced structural

261   heterogeneity or grass diversity (Fig. 1).

262   Changes in abundance of foliage-dwelling arthropods along the invasion gradient varied

263   considerably among taxa, as some orders decreased in abundance as the invasion intensified

264   (Coleoptera, Diptera, Hemiptera), whereas others increased (Hymenoptera, Mantodea, Neuroptera,

265   Parasitiformes) or were unaffected (Araneae, Lepidoptera, Orthoptera, Phasmatodea). A spectrum of

266   responses has been reported for studies that considered the effects of plant invasions on multiple

267   arthropod taxa, which may be attributable to how different functional groups respond to changes in

268   plant composition and structure (Litt et al. 2014).

269   Relative to other groups, herbivorous insects are especially vulnerable to floristic changes from

270   plant invasions because 90% feed only on plants from one or a few lineages and may not use novel

271   plants as food, at least for species whose diets are known (Strong et al. 1984; Bernays and Graham 1988;

272   Tallamy 2004). In the grasslands we studied, as dominance of nonnative grasses increased, abundance

273   of specialist herbivores decreased more steeply than the other functional groups (Fig. 3). Among taxa in

274   this group, which are characterized largely by monophagous or oligophagous herbivores, we found that

275   abundance of Curculionidae and both suborders of Hemiptera decreased markedly as dominance of

276   nonnative grasses increased, a pattern consistent with other studies (reviewed in Litt et al. 2014).

277   Similarly, many Lepidoptera require specific plants for feeding or reproduction, therefore their

278   abundances typically decrease when nonnative plants invade a community (Tallamy and Shropshire

279   2009; Burghardt et al. 2010). In Arizona grasslands, however, we and others observed no systematic

280   variation in abundance of lepidopterans with dominance of nonnative grasses (Litt and Steidl 2010). This

281   finding was not an artifact of a bias towards adults, which are vagile and more likely than larvae to be

captured on plants that they do not use for food or reproduction, because 86% of lepidopterans that we

captured were caterpillars. We acknowledge, however, that larvae of many species, including those that

bore into stems or feed within leaf sheaths, were likely underrepresented in our sample and might

respond to invasions differently than other groups. The lack of an effect for Lepidoptera may be

attributable to the way that grass invasions affect forbs, which are the focus of many lepidopteran-plant

specializations (Futuyma 1976). Typically, forbs decrease in abundance and diversity as dominance of

nonnative plants increase (Geiger 2006), but we found no systematic changes in forb cover along the

invasion gradient. The absence of a decrease in forbs along the invasion gradient might also explain why

abundance of Hymenoptera, an important pollinator taxa, did not decrease in our study. Like specialist

herbivores, pollinators typically are less common in invaded areas because the flowering plants they rely

on for food generally are less abundant (Montero-Castaño and Vilà 2012; Litt et al. 2014).

Generalist herbivores may be more likely to feed on novel plant species than forage-specialized

arthropods (Strong et al. 1984; Tallamy 2004). Consequently, taxa dominated by polyphagous species

might be less vulnerable to invasions that displace their native food sources, especially if nonnative

grasses lack effective defenses against evolutionarily novel herbivores (Parker et al. 2006; Avanesyan

and Culley 2015). They might even increase in abundance if nonnative plants produce more biomass

than native species. In our study, abundances of generalist herbivores were similar (Orthoptera,

Phasmatodea) or lower (Chrysomelidae) in areas dominated by nonnative grasses relative to areas

dominated by native grasses. Interestingly, though, we found that abundances of Phasmatodea and

Orthoptera (including two of three families, Acrididae and Tettigoniidae) were nonlinear, with lower

abundances at intermediate levels of nonnative dominance than at the gradient extremes (Fig. 2).

Nonlinear variation in abundance along the invasion gradient likely reflects species-level

variation in responses of generalist herbivores. The decrease in abundance associated with early stages

of invasion suggests that some species were unable to use nonnative grasses, which generally are

considered less palatable to herbivores than native species because of lower tissue nitrogen and higher

lignin content (Haddad et al. 2001; Geiger 2006). As nonnative grasses continued to increase in

dominance, however, abundance of generalist herbivores increased, suggesting that other species may

have been preadapted to use nonnative grasses, the most common of which are congeners of a locally

common native species (*Eragrostis intermedia*). These species may have benefitted from the increased

quantity of foliage in invaded areas or from structural changes that accompanied invasion by nonnative

grasses. For example, some orthopterans are associated closely with habitat features common to areas

dominated by nonnative grasses, such as increased litter depth or vegetation biomass, which increase

humidity and moderate temperature extremes (Samways 1990; Szinwelski et al. 2012). Conversely,

other species are associated with habitat features common to areas dominated by native grasses, such

as high structural heterogeneity, patches of bare ground, or areas open to direct sunlight (Whitford et

al. 1995; Chambers and Samways 1998). Collectively, these species-specific responses could explain the

nonlinear changes in abundance that we observed.

Because their diets are less specialized than herbivores, predaceous arthropods are thought to

respond more to changes in vegetation structure than to floristics (Pearson 2009; Litt et al. 2014).

Consequently, their abundance and richness often increase with height and complexity of vegetation

(Tscharntke 1995; Haddad et al. 2009). In our study, however, predator abundance decreased as

dominance of nonnative grasses increased, despite increases in grass height and density (Fig. 1a, b).

When taxa were considered individually, abundances of the four most common predator-dominated

taxa remained consistent (Araneae, Meloidae, which have predatory larvae) or decreased (Melyridae,

Cleridae) along the invasion gradient, likely reflecting decreased abundance of herbivore prey. This

suggests that decreases in abundance of herbivorous arthropods are unlikely the result of top-down

pressure from predators exploiting increased habitat along the invasion gradient.

329   Arthropods in semidesert grasslands of Arizona have been sampled with sweep nets (this study)

330   and pitfall traps (Litt and Steidl 2010), with each method chosen to target a subset of arthropods that

331   use different primary substrates. Taxonomic composition differed between these two subsets, but

332   among those taxa well represented in both studies, the direction of responses of foliage- and surface-

333   dwelling (i.e., epigeic) arthropods to invasions by nonnative grasses were generally similar (Litt and

334   Steidl 2010). For both groups abundances of Coleoptera and all taxa combined decreased similarly as

335   dominance of nonnative grasses increased, whereas abundances of foliage-dwelling Hemiptera and

336   Diptera decreased more sharply than their surface-dwelling counterparts. Responses to the plant

337   invasion differed directionally across studies for only two well-represented taxa. Abundance of surface-

338   dwelling Orthoptera decreased markedly as dominance of nonnative grasses increased (Litt and Steidl

339   2010) whereas abundance of foliage-dwelling Orthoptera was similar at the extremes of the invasion

340   gradient and lowest at intermediate levels of nonnative grass dominance. The decrease in abundance of

341   surface-dwelling Orthoptera may be attributable to the importance of open ground or microclimatic

342   conditions for many species, features that likely change when larger, denser-growing grasses invade

343   grasslands (Fig. 1a, b). Conversely, abundance of foliage-dwelling Hymenoptera increased as dominance

344   of nonnative grasses increased whereas abundance of ground-dwelling Hymenoptera did not change

345   (Litt and Steidl 2010). The response of ground-dwelling Hymenoptera was driven strongly by Formicidae

346   (ants), however, which comprised 63% of total individuals captured by pitfall trap (Litt and Steidl 2010)

347   but that we captured only rarely on foliage.

348   In summary, total abundance and richness of foliage-dwelling arthropods decreased as

349   dominance of nonnative grasses increased, although abundances of some groups increased or largely

350   were unaffected by invasions of nonnative grasses. As nonnative grasses continue to expand into

351   grassland and savanna systems as predicted (Schussman et al. 2006), the abundance and distribution of

352   many arthropods is likely to continue to change. Changes in arthropod communities in response to

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353 changes in the plant community could have far-reaching consequences for imperiled grassland

354 ecosystems and the native organisms they support. For example, changes in populations of pollinators

355 or herbivores could influence composition and structure of grassland plant communities (Bezemer et al.

356 2014). Additionally, decreases in abundance of foliage-dwelling arthropods have the potential to disrupt

357 food webs because these arthropods are the primary prey for many insectivorous species, including

358 groups of high-conservation concern, such as breeding grassland birds (George and McEwen 1992;

359 Rzanny and Voigt 2012). Further, given their key role in ecological processes, including energy flow and

360 nutrient cycling, changes to arthropod communities would be expected to alter how grassland

361 ecosystems function (Whiles and Charlton 2006). Consequently, understanding how arthropod

362 communities change in response to invasions by nonnative grasses will become increasingly important

363 for guiding conservation and restoration strategies for grasslands and the organisms and ecological

364 processes they support.

365

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Table 1 Number of individuals or taxa captured (No.), non-focal vegetation and temporal covariates included in final model, and linear effect of dominance of nonnative grasses on abundance and richness of arthropod taxa and functional groups (n = 90 plots). Estimates reported on the log scale and back transformed as the percentage change in abundance for every 10% increase in dominance of nonnative grasses (10% increase). Letters in parenthesis indicate functional groups (G = generalist herbivores, P = predators, S = specialist herbivores). Estimates for non-focal covariates provided in Appendix 1 of Electronic Supplementary Material

Response	Taxa or functional group	No.	Covariates	Estimate	SE	Z	P	10% increase
Abundance	Araneae (P)	4,690	year, day^2, time^2, forb	-0.03	0.05	-0.47	0.64	-0.8
	Coleoptera	4,765	year, day^2	-0.30	0.07	-4.47	<0.001	-9.4
	Chrysomelidae (G)	2,128	day^2, forb, grass	-0.20	0.09	-2.19	0.03	-6.2
	Cleridae (P)[a]	482	day, wood, grass	-0.35	0.13	-2.57	0.01	-10.7
	Curculionidae (S)	703	year, day^2, wood	-0.26	0.11	-2.48	0.01	-8.3
	Meloidae	116	day^2, wood	0.02	0.22	0.08	0.94	0.6
	Melyridae (P)	1,279	year, day	-0.53	0.18	-2.89	0.004	-16.1
	Diptera	861	year, day^2, time^2	-0.28	0.09	-3.05	0.002	-8.7
	Hemiptera (S)	69,698	year, forb, grass	-0.30	0.09	-3.16	0.002	-9.3
	Auchenorrhyncha (S)	56,945	year, day, time, forb, grass	-0.30	0.11	-2.70	0.007	-9.4
	Heteroptera (S)	12,753	year, day^2, grass	-0.24	0.09	-2.83	0.005	-7.7
	Hymenoptera	1,911	year, day^2, time^2, forb	0.15	0.09	1.69	0.09	5.0
	Lepidoptera (S)	1,532	year, day^2, grass	0.10	0.08	1.19	0.23	3.4
	Mantodea (P)	42	day, time, wood	0.43	0.21	1.99	0.05	15.1
	Neuroptera (P)	55	day, wood	0.39	0.18	-2.89	0.03	13.6
	Orthoptera (G)[a]	8,702	year, day, forb	-0.03	0.09	-0.35	0.73	-1.0
	Acrididae (G)[a]	5,869	year, day^2, time, wood, forb, grass	-0.05	0.09	-0.56	0.58	-1.7

	Taxa	N	Model terms					
	Gryllidae (G)	121	day^2, time, forb, grass	-0.04	0.22	-0.18	0.86	-1.3
	Tettigoniidae[a]	2,712	year, day^2, forb	0.04	0.15	-0.27	0.79	-1.3
	Parasitiformes	29	year, day^2, wood, grass	0.85	0.32	2.67	0.007	32.2
	Phasmatodea (G)[a]	512	year, time, wood, grass	-0.05	0.11	-0.50	0.62	-1.8
	All taxa combined	93,418	year, day, forb, grass	-0.21	0.07	-3.07	0.002	-6.8
	Generalist herbivores[a]	11,342	year, day, forb	-0.05	0.07	-0.64	0.52	-1.5
	Specialist herbivores	71,933	year, day^2, forb, grass	-0.27	0.09	-2.98	0.003	-8.5
	Predators	6,548	year, day^2, $time^2$	-0.13	0.06	-2.27	0.02	-4.2
Richness	Orders	11	year, day, time	0.01	0.03	0.54	0.59	0.5
	All taxa	131	year, day, $time^2$, forb, grass cover	-0.21	0.07	-3.07	0.002	-1.9
	Coleoptera	32	year, day^2, $time^2$, grass cover	-0.14	0.04	-4.08	<0.001	-4.6
	Hemiptera	31	year, day^2, forb, grass cover	-0.04	0.03	-1.36	0.18	-1.4
	Orthoptera	38	year, day, forb	-0.12	0.04	-2.69	0.007	-3.8

[a] Taxa for which there was evidence for a nonlinear response to nonnative dominance ($P < 0.05$); see Appendix 1 of Electronic Supplementary Material

Table 2 Mean, standard error, minimum and maximum cover of vegetation and composition of perennial grasses, including the three most common nonnative species across sites, plots, and years (n = 90 plots)

Measure	Category	Mean	SE	Min	Max
Cover (%)	Bare	18.4	1.2	2.5	51.7
	Forbs	9.8	1.0	0.3	52.5
	Litter	4.3	0.3	0.3	12.9
	Woody plants	7.8	0.8	0.0	33.5
	Annual grasses	6.6	1.1	0.0	44.2
	Perennial grasses	64.7	1.8	29.6	95.4
Composition (%)[a]	Native grasses	48.3	3.1	1.3	100.0
	Nonnative grasses	49.1	3.2	0.0	98.8
	Eragrostis lehmanniana	46.7	3.2	0.0	98.8
	Eragrostis curvula	1.9	0.9	0.0	66.3
	Bothriochloa ischaemum	0.5	0.2	0.0	14.3

[a] 2.6% of perennial grasses could not be classified as native or nonnative
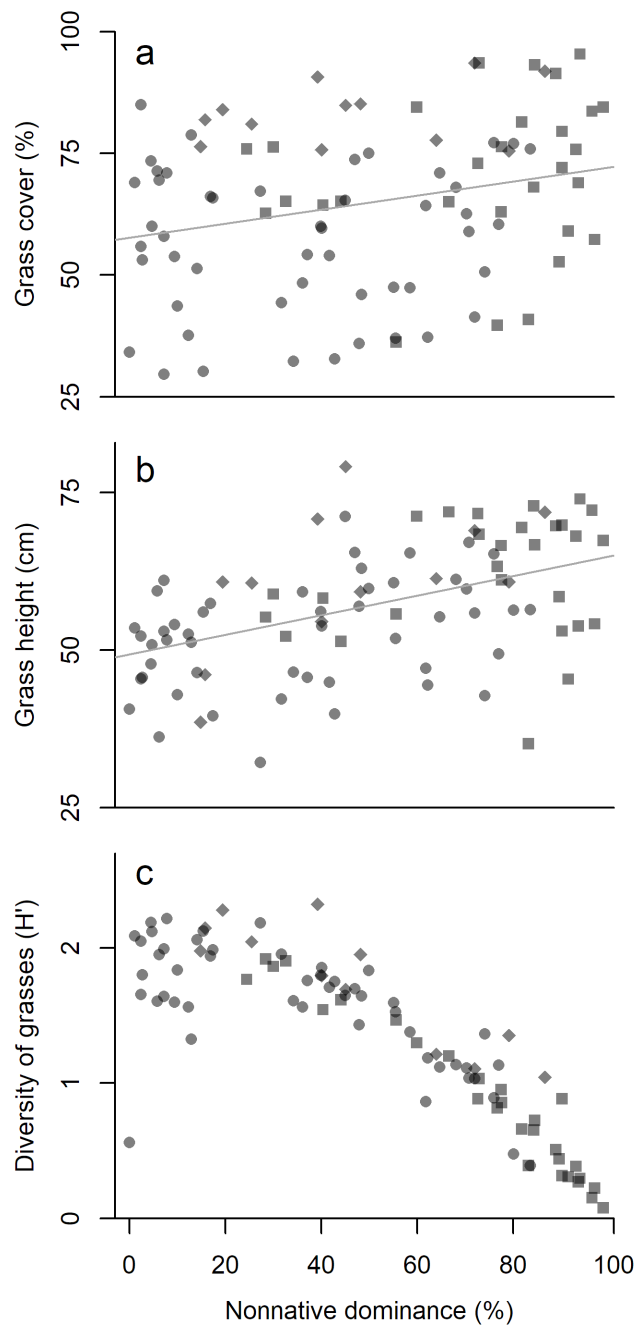
Fig. 1 Grass cover (a), grass height (b), and diversity (Shannon's Index) of perennial grasses (c) versus dominance of nonnative grasses. Point shapes represent the three different sites we surveyed and demonstrate the range of nonnative grass dominance at each site
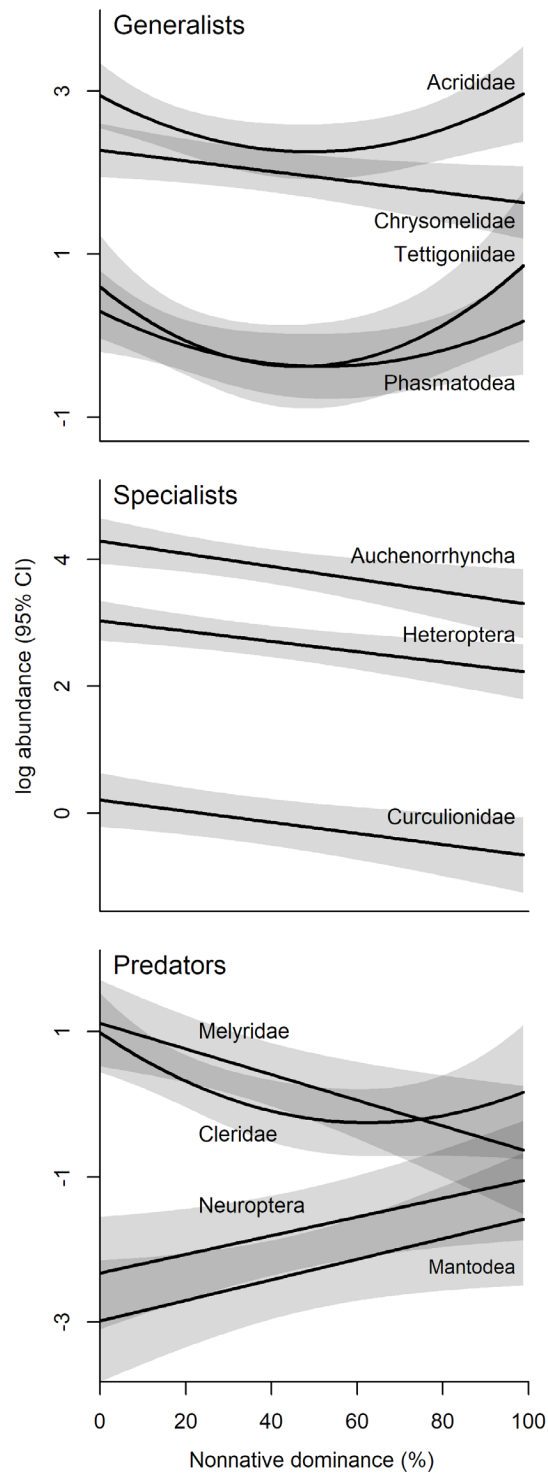
Fig. 2 Log abundance (95% CI) of arthropod taxa versus dominance of nonnative grasses for generalist herbivores (top panel), specialist herbivores (middle panel) and predators (bottom panel). Only those taxa where abundance varied appreciably are shown (Table 1)
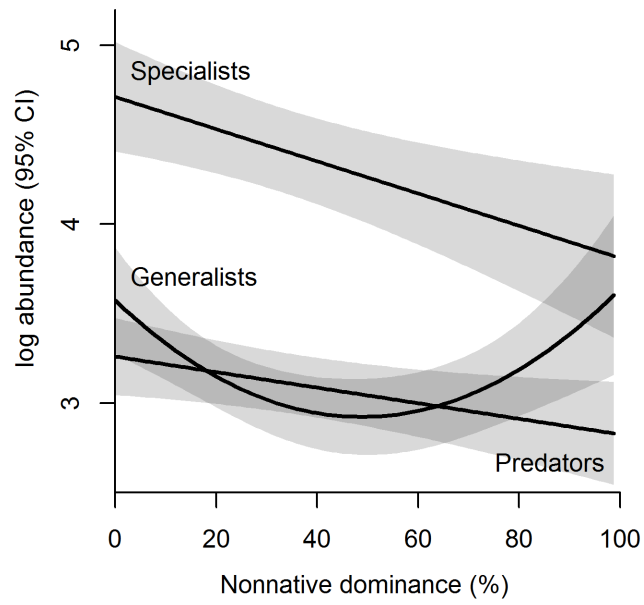
Fig. 3 Log abundance (95% CI) of arthropod functional groups versus dominance of nonnative grasses.

Taxa comprising each group are shown in Table 1

Appendix 1 Estimates reported on the log scale with standard errors (SE), test statistics (*Z*), and *P*-values for fixed effects from final models of arthropod abundance and richness by functional group and taxa. All models included plot as a random effect. For nonnative-dominance effect, quadratic term included when supported (*P* < 0.05). For site effect, FH = Fort Huachuca Military Reservation, LC = Las Cienegas National Conservation Area, and the reference level is Appleton-Whittell Research Ranch

Metric	Functional group or taxa	Covariates	Estimate	SE	*Z*	*P*
Abundance	Generalist herbivores	intercept	3.17	0.17	18.94	<0.001
		site FH	-0.08	0.21	-0.37	0.714
		site LC	-0.24	0.18	-1.35	0.177
		year	0.27	0.06	4.45	8.410
		Julian day	0.86	0.05	17.59	<0.001
		forb cover	0.21	0.06	3.57	<0.001
		nonnative dominance	0.00	0.07	0.07	0.946
		nonnative dominance^2	0.25	0.07	3.43	<0.001
	Specialist herbivores	intercept	5.61	0.22	25.56	<0.001
		site FH	-0.15	0.26	-0.58	0.560
		site LC	-1.34	0.26	-5.24	<0.001
		year	0.73	0.09	7.82	<0.001
		Julian day	0.09	0.04	2.28	0.023
		Julian day^2	0.11	0.05	2.33	0.020
		forb cover	0.13	0.09	1.42	0.155
		grass cover	0.17	0.11	1.54	0.124
		nonnative dominance	-0.27	0.09	-2.98	0.003
	Predators	intercept	3.65	0.14	25.44	<0.001
		site FH	-0.30	0.16	-1.85	0.065
		site LC	-0.60	0.15	-4.08	<0.001
		year	0.43	0.05	8.77	<0.001
		Julian day	0.19	0.04	5.41	<0.001
		Julian day^2	-0.17	0.04	-4.11	<0.001
		time-of-day	-0.02	0.04	-0.42	0.676
		time-of-day^2	-0.07	0.04	-1.92	0.055
		nonnative dominance	-0.13	0.06	-2.27	0.023
	Araneae	intercept	3.45	0.13	25.91	<0.001

	site FH	-0.33	0.16	-2.12	0.034
	site LC	-0.54	0.14	-3.97	<0.001
	year	0.49	0.05	10.67	<0.001
	Julian day	0.01	0.03	0.27	0.791
	Julian day^2	-0.29	0.04	-7.39	<0.001
	time-of-day	0.03	0.04	0.69	0.494
	time-of-day^2	-0.09	0.04	-2.59	0.010
	forb cover	0.08	0.05	1.80	0.072
	nonnative dominance	-0.03	0.05	-0.47	0.642
Coleoptera	intercept	3.00	0.17	17.79	<0.001
	site FH	-0.22	0.19	-1.16	0.246
	site LC	-0.60	0.17	-3.48	<0.001
	year	0.10	0.06	1.70	0.090
	Julian day	0.77	0.05	14.73	<0.001
	Julian day^2	-0.12	0.06	-1.84	0.066
	nonnative dominance	-0.30	0.07	-4.47	<0.001
Chrysomelidae	intercept	2.09	0.23	9.25	<0.001
	site FH	-0.20	0.25	-0.80	0.421
	site LC	-0.13	0.25	-0.54	0.588
	Julian day	0.73	0.07	10.11	<0.001
	Julian day^2	-0.30	0.08	-3.69	<0.001
	forb cover	0.19	0.09	2.13	0.034
	grass cover	0.18	0.09	2.00	0.046
	nonnative dominance	-0.20	0.09	-2.19	0.029
Cleridae	intercept	1.01	0.29	3.44	<0.001
	site FH	-1.36	0.38	-3.57	<0.001
	site LC	-1.21	0.34	-3.53	<0.001
	year	0.22	0.13	1.68	0.092
	Julian day	0.53	0.11	4.77	<0.001
	woody plant cover	-0.19	0.13	-1.48	0.138
	grass cover	0.24	0.14	1.66	0.097
	nonnative dominance	-0.26	0.14	-1.86	0.063
	nonnative dominance^2	0.29	0.14	2.09	0.036
Curculionidae	intercept	0.89	0.27	3.35	<0.001
	site FH	0.00	0.30	0.01	0.992
	site LC	-1.12	0.26	-4.36	<0.001

	year	0.34	0.09	3.72	<0.001
	Julian day	0.70	0.09	7.87	<0.001
	Julian day^2	0.23	0.11	2.15	0.032
	woody plant cover	0.43	0.09	4.83	<0.001
	nonnative dominance	-0.26	0.11	-2.48	0.013
Meloidae	intercept	-3.85	0.76	-5.06	<0.001
	site FH	2.59	0.74	3.51	<0.001
	site LC	0.58	0.72	0.80	0.423
	Julian day	5.92	1.40	4.23	<0.001
	Julian day^2	-3.62	1.03	-3.51	<0.001
	woody plant cover	-0.60	0.21	-2.79	0.005
	nonnative dominance	0.02	0.22	0.08	0.937
Melyridae	intercept	0.32	0.44	0.72	0.470
	site FH	-0.18	0.52	-0.34	0.734
	site LC	-0.07	0.48	-0.14	0.886
	year	0.25	0.16	1.57	0.116
	time-of-day	0.07	0.13	0.53	0.597
	Julian day	1.04	0.14	7.58	<0.001
	nonnative dominance	-0.53	0.18	-2.89	0.004
Diptera	intercept	0.46	0.26	1.77	0.077
	site FH	0.37	0.26	1.41	0.159
	site LC	0.16	0.24	0.64	0.521
	year	0.34	0.08	4.46	<0.001
	Julian day	0.94	0.09	10.36	<0.001
	Julian day^2	-0.29	0.10	-2.82	0.005
	time-of-day	-0.19	0.08	-2.42	0.016
	time-of-day^2	0.25	0.07	3.63	<0.001
	nonnative dominance	-0.28	0.09	-3.05	0.002
Hemiptera	intercept	5.70	0.22	25.85	<0.001
	site FH	-0.19	0.27	-0.72	0.472
	site LC	-1.42	0.26	-5.38	<0.001
	year	0.79	0.10	8.12	<0.001
	forb cover	0.14	0.09	1.46	0.145
	grass cover	0.17	0.12	1.41	0.158
	nonnative dominance	-0.30	0.09	-3.16	0.002
Auchenorrhyncha	intercept	5.48	0.26	21.03	<0.001

	site FH	-0.36	0.32	-1.12	0.261
	site LC	-1.68	0.31	-5.38	<0.001
	year	0.79	0.12	6.82	<0.001
	Julian day	-0.22	0.04	-6.31	<0.001
	time-of-day	-0.07	0.04	-1.75	0.080
	forb cover	0.19	0.11	1.71	0.088
	grass cover	0.26	0.14	1.90	0.057
	nonnative dominance	-0.30	0.11	-2.70	0.007
Heteroptera	intercept	2.75	0.22	12.26	<0.001
	site FH	0.63	0.25	2.54	0.011
	site LC	-0.12	0.25	-0.46	0.644
	year	1.02	0.08	12.11	<0.001
	Julian day	0.75	0.06	12.85	<0.001
	Julian day^2	0.22	0.07	3.34	<0.001
	grass cover	0.24	0.09	2.62	0.009
	nonnative dominance	-0.24	0.09	-2.83	0.005
Hymenoptera	intercept	2.16	0.22	9.65	<0.001
	site FH	-0.36	0.25	-1.43	0.153
	site LC	-0.11	0.22	-0.50	0.618
	year	-0.27	0.08	-3.52	<0.001
	Julian day	0.12	0.07	1.69	0.091
	Julian day^2	-0.45	0.08	-5.29	<0.001
	time-of-day	-0.11	0.08	-1.40	0.160
	time-of-day^2	0.18	0.07	2.38	0.017
	nonnative dominance	0.15	0.09	1.69	0.090
Lepidoptera	intercept	0.16	0.23	0.68	0.494
	site FH	0.68	0.25	2.71	0.007
	site LC	0.35	0.25	1.41	0.160
	year	0.18	0.08	2.14	0.032
	Julian day	0.97	0.08	12.73	<0.001
	Julian day^2	0.50	0.10	5.22	<0.001
	grass cover	0.37	0.10	3.84	<0.001
	nonnative dominance	0.10	0.08	1.19	0.233
Mantodea	intercept	-2.06	0.51	-4.03	<0.001
	site FH	0.07	0.60	0.11	0.914
	site LC	-0.23	0.58	-0.40	0.691

	Julian day	-0.34	0.16	-2.18	0.029
	time-of-day	-0.34	0.19	-1.83	0.067
	forb cover	0.18	0.13	1.43	0.152
	woody plant cover	0.28	0.15	1.87	0.061
	nonnative dominance	0.43	0.21	1.99	0.047
Neuroptera	intercept	-2.21	0.50	-4.43	<0.001
	site FH	-0.18	0.56	-0.32	0.748
	site LC	0.52	0.51	1.01	0.311
	Julian day	0.73	0.17	4.27	<0.001
	woody plant cover	0.30	0.13	2.34	0.019
	nonnative dominance	0.39	0.18	2.14	0.033
Orthoptera	intercept	2.57	0.21	12.37	<0.001
	site FH	0.08	0.26	0.29	0.774
	site LC	-0.08	0.23	-0.36	0.721
	year	0.39	0.08	5.11	<0.001
	Julian day	1.01	0.06	17.24	<0.001
	forb cover	0.23	0.07	3.12	0.002
	nonnative dominance	0.02	0.09	0.26	0.797
	nonnative dominance^2	0.25	0.09	2.82	0.005
Acrididae	intercept	2.08	0.23	9.12	<0.001
	site FH	0.00	0.27	-0.01	0.989
	site LC	0.17	0.26	0.66	0.508
	year	0.27	0.10	2.66	0.008
	Julian day	0.86	0.06	13.68	<0.001
	Julian day^2	0.12	0.07	1.69	0.092
	time-of-day	0.14	0.06	2.33	0.020
	forb cover	0.29	0.09	3.21	0.001
	woody plant cover	0.16	0.08	2.08	0.038
	grass cover	0.14	0.12	1.20	0.230
	nonnative dominance	0.00	0.09	0.02	0.980
	nonnative dominance^2	0.26	0.09	2.76	0.006
Gryllidae	intercept	-2.35	0.58	-4.04	<0.001
	site FH	0.42	0.57	0.74	0.459
	site LC	-2.33	0.66	-3.55	<0.001
	Julian day	1.17	0.13	8.98	<0.001
	Julian day^2	0.80	0.20	4.06	<0.001

	time-of-day	0.23	0.12	1.89	0.058
	forb cover	-0.44	0.20	-2.19	0.028
	grass cover	-1.21	0.25	-4.91	<0.001
	nonnative dominance	-0.04	0.22	-0.18	0.861
Tettigoniidae	intercept	0.71	0.37	1.91	0.056
	site FH	-0.07	0.44	-0.17	0.864
	site LC	-1.09	0.39	-2.81	0.005
	year	0.85	0.13	6.49	<0.001
	Julian day	1.87	0.15	12.48	<0.001
	forb cover	0.22	0.12	1.77	0.076
	nonnative dominance	0.07	0.14	0.51	0.612
	nonnative dominance^2	0.41	0.15	2.75	0.006
Parasitiformes	intercept	-1.36	0.55	-2.48	0.013
	site FH	-1.75	0.71	-2.46	0.014
	site LC	-1.36	0.77	-1.76	0.079
	year	0.89	0.30	2.98	0.003
	Julian day	-0.19	0.26	-0.76	0.450
	Julian day^2	-0.80	0.25	-3.20	0.001
	woody plant cover	0.58	0.29	2.00	0.046
	grass cover	0.85	0.43	1.99	0.047
	nonnative dominance	0.85	0.32	2.67	0.007
Phasmatodea	intercept	0.32	0.25	1.28	0.202
	site FH	0.12	0.30	0.40	0.689
	site LC	-0.70	0.30	-2.31	0.021
	year	0.32	0.11	2.85	0.004
	time-of-day	-0.33	0.08	-4.28	<0.001
	woody plant cover	-0.21	0.11	-2.00	0.046
	grass cover	0.21	0.13	1.63	0.104
	nonnative dominance	-0.04	0.11	-0.39	0.700
	nonnative dominance^2	0.23	0.12	1.98	0.047
All taxa	intercept	6.00	0.16	36.60	<0.001
	site FH	-0.10	0.20	-0.52	0.601
	site LC	-0.97	0.20	-4.92	<0.001
	year	0.59	0.07	8.20	<0.001
	Julian day	0.18	0.03	5.42	<0.001
	forb cover	0.18	0.07	2.54	0.011

		grass cover	0.18	0.09	2.02	0.044
		nonnative dominance	-0.21	0.07	-3.07	0.002
Richness	Orders	intercept	2.03	0.06	33.43	<0.001
		site FH	-0.05	0.08	-0.68	0.496
		site LC	-0.10	0.07	-1.39	0.163
		year	0.05	0.02	2.04	0.041
		Julian day	0.07	0.02	3.17	0.002
		time-of-day	-0.05	0.02	-2.13	0.034
		nonnative dominance	0.01	0.03	0.54	0.590
	Morphospecies	intercept	6.00	0.16	36.60	<0.001
		site FH	-0.10	0.20	-0.52	0.601
		site LC	-0.97	0.20	-4.92	<0.001
		year	0.59	0.07	8.20	<0.001
		Julian day	0.18	0.03	5.42	<0.001
		forb cover	0.18	0.07	2.54	0.011
		grass cover	0.18	0.09	2.02	0.044
		nonnative dominance	-0.21	0.07	-3.07	0.002
	Coleoptera	intercept	1.56	0.09	16.85	<0.001
		site FH	0.04	0.10	0.39	0.697
		site LC	-0.16	0.10	-1.71	0.087
		year	0.09	0.03	2.60	0.009
		Julian day	0.40	0.03	11.91	<0.001
		Julian day^2	-0.15	0.04	-3.77	<0.001
		time-of-day	-0.01	0.03	-0.25	0.805
		time-of-day^2	0.04	0.03	1.55	0.121
		grass cover	0.07	0.04	1.91	0.056
		nonnative dominance	-0.14	0.04	-4.08	<0.001
	Hemiptera	intercept	1.69	0.08	20.92	<0.001
		site FH	0.07	0.09	0.81	0.418
		site LC	-0.20	0.09	-2.19	0.029
		year	0.14	0.03	4.23	<0.001
		Julian day	0.20	0.03	7.64	<0.001
		Julian day^2	0.08	0.03	2.43	0.015
		forb cover	0.08	0.03	2.60	0.009
		grass cover	0.07	0.04	1.54	0.123
		nonnative dominance	-0.04	0.03	-1.36	0.175

Orthoptera	intercept	1.34	0.10	12.97	<0.001
	site FH	0.32	0.13	2.50	0.012
	site LC	0.06	0.11	0.51	0.609
	year	0.07	0.04	2.00	0.046
	Julian day	0.45	0.03	15.05	<0.001
	forb cover	0.13	0.03	3.83	<0.001
	nonnative dominance	-0.12	0.04	-2.69	0.007