Responses of human skin to intradermal injection of leukotrienes C_4 , D_4 and B_4

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- 1 The ability of intradermally injected leukotrienes C_4 (LTC₄), LTD₄ and LTB₄ to produce inflammatory changes in human skin alone and in combination with prostaglandin E_2 (PGE₂) has been investigated.
- 2 LTC₄ and D₄ (0.012-0.38 nmol) caused dose-related erythema and wealing. No evidence of synergism between PGE₂ and LTC₄ or LTD₄ was detected, although only single dose combinations were studied.
- 3 LTB₄ (0.15-1.5 nmol) caused areas of induration which persisted for more than 4 h and which showed perivascular neutrophil infiltrates on histological examination. Only slight synergism between PGE₂ and LTB₄ was found.
- 4 It was concluded that these pro-inflammatory properties of LTC₄, LTD₄ and LTB₄ are consistent with their proposed roles as mediators of inflammation in the skin and other tissues.

Introduction

Oxidative metabolism of arachidonic acid leads to the formation of a wide range of putative mediators of inflammation. Products of the cyclo-oxygenase pathway, including prostaglandin E₂ (PGE₂), PGD₂ and prostacyclin (PGI₂) may play a role as mediators of the vascular events of acute inflammation in human skin (see Camp, 1982, for review). Recently, other oxidative pathways of arachidonic acid metabolism have been described, which are catalysed by lipoxygenases. Of the several compounds formed by these pathways, the leukotrienes (Samuelsson & Hammarström, 1980) have generated interest and speculation as potential mediators of inflammation.

Leukotriene C₄ (LTC₄, 5S-hydroxy-6R-S-gluta-thionyl-7,9-trans-11,14-cis-eicosatetraenoic acid) and LTD₄ (5S-hydroxy-6R-S-cysteinylglycinyl-7,9-trans-11,14-cis-eicosatetraenoic acid) are components of the pharmacological substance, slow reacting substance of anaphylaxis (SRS-A) (see Sirois & Borgeat, 1980, for review) and both induce vasoconstriction in guinea-pig skin and cause increased vascular permeability in the presence of the vasodilator, PGE₂. LTC₄ was the more potent vasoconstrictor, and its permeability-increasing effect was partly masked by this property (Peck, Piper & Williams, 1981). LTC₄ and LTD₄ (1 nmol each) were injected intradermally in human skin (Soter, Lewis, Corey & Austen, 1983) and persistent weals with

central pallor and surrounding erythema were seen, but in this study little information about dose-related effects is given.

LTB₄ (5S, 12R-dihydroxy-6,14-cis-8, 10-transeicosatetraenoic acid) (Sirois, Roy, Borgeat, Picard & Corey, 1981) is one of the most potent known chemotactic agents for polymorphonuclear leucocytes (Ford-Hutchinson, Bray, Doig, Shipley & Smith, 1980; Palmer, Stepney, Higgs & Eakins, 1980; Nagy, Lee, Goetzl, Pickett & Kay, 1982). It has been shown to cause increased vascular permeability (Wedmore & Williams, 1981) and to induce leucocytic infiltration on intradermal injection in rabbits (Carr, Higgs, Salmon & Spayne, 1981). Soter et al. (1983) have also reported that intradermal injection of 1.6 nmol LTB4 in human skin caused an immediate weal and flare reaction followed by persistent induration in one subject, but no visible effect was seen in two other subjects. However, histological examination of biopsies of LTB₄ injection sites in each of the three subjects at 6 h showed perivenular neutrophil infiltrates. Both LTB4 and LTC4 were reported to enhance the expression of complement receptors on human neutrophils and eosinophils (Nagy et al., 1982). We have therefore investigated the pro-inflammatory activity of LTC4, LTD4 and LTB₄ in human skin, and we now describe the effects of intradermal injection of a range of doses of these

compounds. Some of this work was communicated to the British Pharmacological Society as a poster (Camp, Coutts, Greaves, Kay & Walport, 1982).

Methods

Subjects

The study was carried out on the skin of the backs of six healthy adult male subjects aged 33-47 years, all of whom were members of the medical or scientific staff of the Institute of Dermatology, and who were fully aware of the nature and objectives of the study. The protocol had previously been approved by the Institute of Dermatology Ethical Committee. None of the subjects had taken any systemic drugs within 24 h preceding the study, and none had any history of atopic disorders.

Materials

LTC₄ and LTD₄ were kindly supplied by Dr J. Rokach, Merck Frosst Canada Inc., Quebec, Canada. LTB₄ was biosynthesized as described by Camp, Woollard, Mallet, Fincham, Ford-Hutchinson & Bray (1982) using methods based on those of Borgeat & Samuelsson (1979). The LTB₄ fraction was finally purified by straight phase high performance liquid chromatography (h.p.l.c.) in order to separate the 5S,12S-trans-cis-trans isomer of LTB₄ (Borgeat, Picard, Vallerand and Sirois, 1981) which is a much less active stimulator of leucocyte motility (R.D.R. Camp and S.D. Brain, unpublished observations). The identity of LTB4 was confirmed by ultraviolet spectrophotometry and gas chromatography - mass spectrometry. PGE₂ (Prostin E₂) was obtained from Upjohn, Crawley. Histamine acid phosphate injection B.P. was obtained from McCarthy's, Romford. Phosphate buffered saline, pH 7.3, contained 0.01 M sodium phosphate and 0.154 M sodium chloride and was sterilized by Millipore filtration.

Dose-response curves

Skin was injected intradermally with 50 µl phosphate buffered saline alone or containing LTC₄, LTD₄, LTB₄, PGE₂, histamine or combinations of these agents. Injections were carried out between 11 h 00 min and 13 h 00 min in a temperature-controlled room (22-25°C). The area of erythema induced by each injection was delineated at 1 min and again at 5 min. These two areas were often quite different in outline and intensity. The outlines were traced on to paper and the areas of erythema measured planimetrically. Two perpendicular diameters of each weal, including the longest axis and ignoring small pseudopodia, were measured at intervals after injec-

tion. The size of the weal was given as the mean of the two diameters. Subjective sensations observed by the subjects in response to each injection were also recorded.

Statistics

The significance of differences between the mean area of erythema induced by injection of vehicle and that due to injection of each test substance was determined by the Mann-Whitney U-test. The mean maximum diameter of weals due to injection of vehicle was similarly compared with that due to test substances. Because sample numbers were low, tables of U for small numbers were consulted (Siegel, 1956).

Histological studies

Punch biopsies of injection sites (4 mm in diameter) were removed after intradermal infiltration of 1% lignocaine as a local anaesthetic. These were fixed in formalin, subjected to routine paraffin section and stained with haematoxylin and eosin before examination by light microscopy.

Results

Leukotrienes C4 and D4

Intradermal injections of solutions containing LTC₄ or LTD₄ caused similar responses (Figure 1 and 2). The first was an erythematous reaction in the surrounding skin. This appeared within 60 s of injection, reached a maximum between 1 to 3 min, was diminished by 5 min (as indicated in Figures 1 and 2) and faded completely over the following 60 min. As little as 0.012 nmol of each agent caused significant erythema. The extent of erythema was dose-related up to 0.19-0.38 nmol with the exception of a relative depression of the responses to 0.09 nmol LTD₄. In other experiments (data not shown) it was found that increasing the dose of LTC4 or LTD4 as high as 7.5 nmol in four subjects caused no further significant increase in erythema. Figures 1 and 2 also show the erythemal response to histamine for comparison. The maximum erythema response to either leukotriene was less than that due to 1.63 nmol histamine.

LTC₄ and LTD₄ also produced weal formation (Figures 1 and 2). The mean diameters of the weals were maximal at 15 min and were dose-related up to 0.19-0.38 nmol, with the exception of a relative depression of the response to 0.09 nmol LTD₄. The lowest dose of LTC₄ (0.012 nmol) and 0.047 nmol LTD₄ caused weals that were greater than the

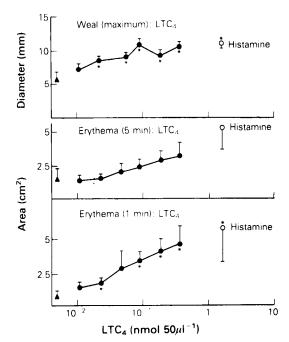


Figure 1 Dose-response curves (mean values with s.e. shown by vertical lines) for area of 1 min and 5 min erythema and for maximum diameter of weal evoked by intradermal injection of leukotriene C_4 (LTC₄), compared with responses to a single dose of histamine. (\triangle) = response to an equal volume of vehicle. Number of subjects = 4. *P < 0.05, on comparison with (\triangle) (Mann-Whitney U-test).

vehicle-induced weals. In other experiments (data not shown) further increase of the dosage of LTC₄ or LTD₄ up to 7.5 nmol caused no greater diameter of wealing than that due to 0.38 nmol of either agent. Maximum wealing due to these two leukotrienes was approximately equal in magnitude to that due to 1.63 nmol histamine. Wealing subsided 45–60 min after injection and no subsequent visible or palpable reactions developed at the injection sites. Central pallor was seen in the weals induced by LTC₄ and LTD₄, but this was no more prominent than that seen in the histamine-induced weals. The sensations caused by intradermal injection of LTC₄ and LTD₄ did not differ significantly in either quality or intensity from those due to injection of the vehicle.

Leukotriene B4

LTB₄ was injected at dosages of 0.15, 0.30 and 1.5 nmol in four subjects. In all subjects areas of induration developed at about 30 min after injection. The induration was ill-defined, erythematous and slightly raised, and achieved a maximum diameter of about

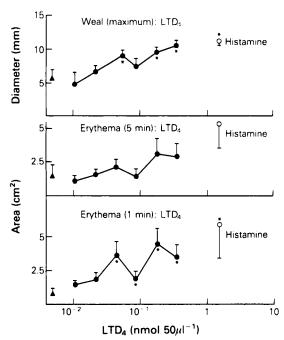


Figure 2 Dose-response curves (mean values with s.e. shown by vertical lines) for area of 1 min and 5 min erythema, and for maximum diameter of weal evoked by intradermal injection of leukotriene D_4 (LTD₄), compared with responses to a single dose of histamine. (\triangle) =response to an equal volume of vehicle. Number of subjects= 4. *P < 0.05, on comparison with (\triangle) (Mann-Whitney U-test).

8 mm at 60-120 min after injection. The diameters of these areas were not dose-related. These lesions persisted for at least 4 h, were tender and, in one subject, were associated with persistent itching. Neither the vehicle nor histamine (1.63 nmol) caused any delayed induration.

Prostaglandin E2

The effect of injection with PGE₂ alone and in combination with LTC₄, LTD₄ and LTB₄ was investigated in two subjects. PGE₂ alone (0.18 nmol) caused erythema and wealing which were approximately equal in extent to the maximal response to LTC₄ and LTD₄. A solution containing 0.18 nmol PGE₂ and 0.38 nmol LTC₄ or 0.75 nmol LTD₄ caused no greater erythema or wealing than would be expected from the sum of the responses individually. However, combined injections using lower doses of leukotrienes were not performed. Injection of a combination of LTB₄ (0.30 nmol) and PGE₂ (0.14 nmol) caused slight enhancement of the visible responses; at 4 h the diameters of the ill-defined swellings were

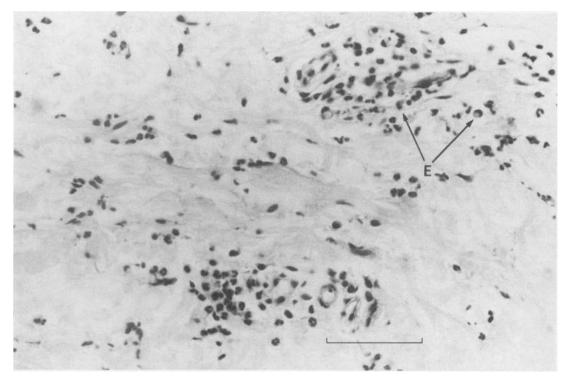


Figure 3 Histological appearance of human skin 4 h after intradermal injection of 0.3 nmol leukotriene B_4 (LTB₄), demonstrating a dermal perivascular neutrophil infiltrate with a few eosinophils. E = eosinophil. Bar = 100 μ m.

approximately 3 mm larger at the sites of the combined injections compared with the sites of injection of 0.30 nmol LTB₄ alone. PGE₂ (0.14 nmol) alone caused no detectable response at 4 h.

Histological appearances

In two subjects 4 mm punch biopsies were removed 4 h after injection of 0.30 nmol LTB₄, and were processed for light microscopy as described. Biopsies from both subjects showed a pronounced dermal neutrophil infiltrate (Figure 3), with perivascular accentuation, and containing a few eosinophils. In contrast, similar biopsies removed 2 h after injection of 7.5 nmol LTC₄ or LTD₄ in the same two subjects showed little or no evidence of polymorphonuclear infiltration.

Discussion

This study demonstrates that both LTC₄ and LTD₄ produce dose-related erythema and wealing in human skin at very low concentrations, although

dose-response curves are shallow and control values for weals and erythema are high. Although no attempt was made to make a full direct comparison of dose-response curves, consideration of our present and earlier results (Marks & Greaves, 1977) suggests that LTC₄ and LTD₄ are at least as potent as histamine in production of erythema and wealing at the lower dose range, especially in respect of weal production.

Much existing information on the proinflammatory actions of LTC4 and LTD4 derives from studies on the microvasculature of guinea-pig skin (Peck et al., 1981). In this species, both LTC4 and LTD₄ cause vasoconstriction, the former being the more potent. LTD₄, but not LTC₄, caused local increase in vascular permeability which was greatly potentiated by addition of the vasodilator PGE₂. Evidence was obtained that LTC₄ had some permeability increasing activity, but this was masked by its potent vasoconstrictor properties. Soter et al. (1983) report the presence of pallor in the centre of weals due to intradermal injection of LTC4 and LTD4 (1 nmol each) in human subjects and interpret this as being due to intrinsic arteriolar constriction. While we have also observed pallor in the centre of weals

induced by LTC4 and LTD4, equivalent pallor was seen in histamine-induced weals. We believe that pallor in weals of human skin may simply be a function of the degree of dermal oedema, which, by extrinsic compression of the microvasculature, transiently interferes with dermal blood flow. We commonly observe pallor in wealing reactions induced experimentally and seen in clinical disease. Our interpretation is supported by the experiments of Bisgaard, Kristensen & Sondergaard (1982), in which the effects on cutaneous blood flow of intradermal injection of LTC₄ and LTD₄ (1 µg each) in humans were determined by a laser-Doppler technique. They found that wealing interfered with blood flow values, and therefore measured blood flow at a distance of 5 mm from injection sites. Increased blood flow was seen and there was no evidence of vasoconstriction.

In the study of Soter et al. (1983) erythema due to LTC₄ and LTD₄ was found to be maximal at 1 h and to persist for more than 4 h, whereas in the present study erythema was maximal between 1 and 3 min and had faded completely by 1 h. In addition, the study of Soter et al. indicated that wealing was maximal at 1 h and had dissipated by 4 h, whereas in the present study wealing was maximal at 15 min and had dissipated by 1 h. These differences may be explained by the higher doses of LTC₄ and LTD₄ used by Soter et al. (1983). It may also be relevant that their injections were carried out on the flexor aspect of the forearm, whereas ours were done on the back.

We were unable to demonstrate any synergism between PGE₂ and LTC₄ or LTD₄, but similar combination studies ought to be carried out using a wider leukotriene dose range and with other vasodilator mediators including PGI₂ and PGD₂. Direct evidence of increased generation of LTC₄ and LTD₄ in inflamed tissues has yet to be obtained. Nevertheless, our findings are consistent with the possibility that these compounds may behave as mediators of inflammation.

The results obtained with LTB4 show that sub-

nanomolar amounts of this agent are capable of inducing neutrophil infiltrates in human tissues in vivo. Although there is evidence that LTB₄ is chemokinetic (Bray, Ford-Hutchinson & Smith, 1981) and chemotactic (Nagy et al., 1982) towards eosinophils, only a few eosinophils were seen in the biopsies of LTB₄ injection sites in our experiments. The number of eosinophils infiltrating in response to LTB₄ injection may depend on the concentration of blood eosinophils, and may possibly be increased when the blood eosinophil count is high. Soter et al. (1983) have reported that PGD₂ enhanced the response to intradermal LTB₄. We have demonstrated only slight enhancement of the visible LTB₄ response in human skin by PGE₂.

There have been few conclusive reports of elevated LTB₄ levels in inflamed tissues. The report of Klickstein, Shapleigh & Goetzl (1980) that LTB₄ levels were raised in joint fluid of patients with inflammatory arthropathies appears to be based on analytical techniques of limited specificity, and is thus open to challenge. Their report has also been questioned by Davidson, Rae & Smith (1982), who analysed synovial fluid samples from patients with rheumatoid arthritis by h.p.l.c. and bioassay. They reported bioassayable concentrations of LTB4 of about 0.34 ng ml⁻¹ joint fluid, levels approximately 400 fold lower than those reported by Klickstein et al. (1980). Brain, Camp, Dowd, Kobza Black, Woollard, Mallet & Greaves (1982) have demonstrated the release of LTB₄ from the involved skin of patients with psoriasis, as determined by assay for chemokinetic activity in h.p.l.c. fractions of skin extracts.

The properties of the leukotrienes we have studied are entirely consistent with their proposed role as mediators of inflammation in skin and other tissues. Whilst specific integrated sequential measurements of the levels of these compounds in inflamed tissues need to be carried out, final establishment of their importance will have to await the development of specific antagonists or inhibitors.

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