



Responses of invasive silver and bighead carp to a carbon dioxide barrier in outdoor ponds

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Complete List of Authors:	Cupp, Aaron; USGS Upper Midwest Environmental Science Center Erickson, Richard A.; Upper Midwest Environmental Sciences Center Fredricks, Kim; USGS Upper Midwest Environmental Science Center Swyers, Nicholas; USGS Western Fisheries Research Center Hatton, Tyson; USGS Western Fisheries Research Center Amberg, Jon; USGS Upper Midwest Environmental Science Center
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3 **Author list:** Aaron R. Cupp^{1*}, Richard A. Erickson¹, Kim T. Fredricks¹, Nicholas M. Swyers², Tyson W.

4 Hatton², Jon J. Amberg¹

5 **Author email address:** acupp@usgs.gov, rerickson@usgs.gov, kfredricks@usgs.gov, nswyers@usgs.gov,

6 thatton@usgs.gov, jamberg@usgs.gov

7 **Author affiliation:**

8 ¹U.S. Geological Survey, Upper Midwest Environmental Sciences Center, 2630 Fanta Reed Rd., La Crosse,

9 WI 54603, USA

10 ²U.S. Geological Survey, Western Fisheries Research Center, 5501A Cook-Underwood Road, Cook,

11 Washington 98605, USA

12 ***Corresponding author:** Aaron R. Cupp, Telephone: (608)781-6266, Fax: (608)783-6066,

13 acupp@usgs.gov, 2630 Fanta Reed Rd., La Crosse, WI 54603, USA

14

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17 **Abstract**

18 Resource managers need for effective methods to prevent the movement of silver
19 (*Hypophthalmichthys molitrix*) and bighead carp (*H. nobilis*) from the Mississippi River basin into the
20 Laurentian Great Lakes. In this study, we evaluated dissolved carbon dioxide (CO₂) as a barrier and
21 deterrent to silver (278 ± 30.5 mm) and bighead (212 ± 7.7 mm) carp movement in continuous-flow
22 outdoor ponds. As a barrier, CO₂ significantly reduced upstream movement but was not 100% effective
23 at blocking fish passage. As a deterrent, we observed a significant shift away from areas of high CO₂
24 relative to normal movement before and after injection. Carbon dioxide concentrations varied across
25 the pond during injection and reached maximum concentrations of 74.5±1.9 mg/L CO₂; 29 532 – 41 393
26 µatm at the site of injection during three independent trials. We conclude that CO₂ altered silver and
27 bighead carp movement in outdoor ponds and recommend further research to determine barrier
28 effectiveness during field applications.

29

30 **Keywords**

31 Carbon dioxide; Asian carp; invasive species; barrier

32 Introduction

33 Range expansion of the invasive silver (*Hypophthalmichthys molitrix*) and bighead carp (*H.*
34 *nobilis*) throughout the Mississippi River basin has been documented by resource agencies since the
35 1970's (Kelly et al. 2011). These prolific fish cause harm to native aquatic ecosystems by competing with
36 native mussels, early life-stage fish, and other filter-feeding fish for food resources from a planktivorous
37 diet overlap (Irons et al. 2007; Sampson et al. 2009; Walleser et al. 2014). This increased competition
38 has been demonstrated through major shifts in zooplankton communities on the Illinois River where
39 silver and bighead carp are in high abundances (Sass et al. 2014). Recent detections using eDNA and
40 physical captures place the upper range of these invasive carp at tributaries adjacent to the southern
41 Great Lakes (Jerde et al. 2013; Parker et al. 2015). With direct hydrological connection of these
42 tributaries to the Great Lakes, research evaluating potential barriers to prevent further upstream
43 movement has become increasingly important.

44 Physical barriers are not an option on many navigational rivers due to economic losses from the
45 movement of goods and services. Closure of navigational locks for invasive species control on the
46 Chicago Area Waterway System (CAWS), the shipping channel connecting the Mississippi River basin to
47 the Great Lakes basin, was estimated to cost consumers and commercial traffic \$1.3 billion USD annually
48 (Schwieterman 2010). With obvious economic concerns, resource managers have started exploring non-
49 physical barriers that aim to prevent invasive species movement while concurrently allowing water-body
50 navigation (Noatch and Suski 2012).

51 Fisheries professionals have described a number of potential non-physical strategies for
52 preventing the spread of invasive carps in North America, but most agree that a general lack of data
53 addressing the effectiveness of these techniques will ultimately delay management actions (Wittman et
54 al. 2014). In response, recent studies have begun to address this gap in published literature. Experiments
55 using acoustic barriers, bubble barriers, hydro-guns, carbon dioxide (CO₂), and electricity have all shown

56 varying levels to efficacy to deter invasive carps (Taylor et al. 2005; Kates et al. 2012; Ruebush et al.
57 2012; Romine et al. 2015). Realizing the need for continued research, the Asian Carp Regional
58 Coordination Committee (ACRCC) prioritized promising technologies (ACRCC MRW 2014) and dissolved
59 CO₂ was identified as a strategy that requires further testing.

60 A review of non-physical barriers by Noatch and Suski (2012) highlights the potential for CO₂ to
61 work as a barrier for invasive fishes. Further, reviews by Ishimatsu et al. (2005) and Perry and Abdallah
62 (2012) describe a suite of negative physiological responses by fish when exposed to hypercapnia. Most
63 fish sense CO₂ through chemoreceptors located on the gills (Ishimatsu et al. 2005; Perry and Abdallah
64 2012) and the potential to exploit CO₂ sensitivity in fish suggests that this technique may also work to
65 deter invasive carp movement (Noatch and Suski 2012). In laboratory tests with silver carp, Kates et al
66 (2012) observed avoidance behavior and irregular movements at 70 – 90 mg/L CO₂. Further, recent tests
67 in a static pond confirmed that silver and bighead carp avoided approximately 60 mg/L CO₂ (Donaldson
68 et al. 2016). With evidence growing, the effectiveness of CO₂ as a barrier to invasive carp movement
69 needs to be tested.

70 The objective for our study was to evaluate silver and bighead carp movement in outdoor ponds
71 in response to a CO₂ barrier over three independent trials. We hypothesized that CO₂ would reduce
72 upstream passage and subsequently deter fish to seek refuge in areas of lower CO₂. Barrier effectiveness
73 was determined by quantifying upstream passage through the injection site. Deterrence effectiveness
74 was determined by quantifying shifts in fish locations relative to changes in CO₂. Using these behavioral
75 metrics, we compared fish movement collected before, during, and after CO₂ injection. Results are
76 focused on the applicability of CO₂ to deter silver and bighead carp passage with implications for
77 reducing the risk transfer between the Mississippi River and Great Lakes basins.

78

79 **Materials and Methods**

80 *Study Animals* – We obtained silver carp ($n = 15$; total length: 278 ± 30.5 mm; wet weight: 229.2
81 ± 81.9 g; mean \pm standard deviation) and bighead carp ($n = 15$; total length: 212 ± 7.7 mm; wet weight:
82 101.4 ± 12.3 g; mean \pm standard deviation) from the Missouri River near Columbia, Missouri, USA and a
83 private aquaculture farm in Osage Beach, Missouri, USA . Fish were held at the U.S. Geological Survey
84 (USGS) Upper Midwest Environmental Sciences Center (UMESC) Invasive Species Complex (La Crosse, WI
85 USA) until the time of testing. During the week preceding each trial, five bighead and five silver carp
86 were removed from the 1500-L source tanks and placed in a single 130-L flow-through tank at $12 \pm 1^\circ\text{C}$
87 where diet was converted from dry feed to an algal feed mixture. Algal mix contained equal parts of 3.6
88 g/L Chlorella (Yaeyama Shokusan Co. Ltd., Shiraho, Ishigaki Island, Japan) and Spirulina (Stakich, Inc.,
89 Royal Oak, MI, USA) and was the same feed administered during the outdoor trials. Approval from
90 UMESC Animal Care and Use Committee (Protocol Number: AEH-12-PPTAC-01), using similar guidelines
91 as described in Guide for the Care and Use of Laboratory Animals (1996), was gained before study
92 initiation.

93
94 *Pond Setup* - Trials were conducted in outdoor concrete ponds at UMESC. Pond dimensions
95 were 10.0 m long x 4.9 m wide x 1.2 m deep with a volume of approximately 58 700 L. Influent water
96 was supplied directly to the pond from two on-site wells and effluent water was discharged from the
97 pond by spilling over at the top of the opposite wall. Setup was similar to a large choice chamber, with
98 the pond divided into two symmetrical halves by a partial concrete block wall and a 3.1 m opening at the
99 effluent end (Figure 1). Each half contained similar water volumes and a continuous supply of inflowing
100 water (300 L/min) down each side. Wood lattice was affixed to all pond walls, including the brick
101 partition, to reduce reflection of acoustic telemetry signals during fish tracking.

102 Three micro-bubble diffusers (Model: Point Four™ MBD 300, Pentair Aquatic Eco-Systems™,
103 Apopka, FL, USA) were placed at an equal distance across the opening to one half of the pond and were

104 used to inject CO₂ into the pond. To evaluate changes in fish locations, the pond was divided into two
105 areas (Figure 1). The side where CO₂ was injected was termed “High CO₂ Area” and the opposite side
106 “Low CO₂ Area”. Names for each area also describe relative CO₂ concentrations found during CO₂
107 injection.

108 For water chemistry sampling, plastic tubing (4.8 mm ID, Python Products Inc., Milwaukee, WI,
109 USA) was run from six locations near the pond bottom to a peristaltic pump (Model 7553-70, Cole-
110 Parmer®, Vernon Hills, IL, USA) outside of the pond area to prevent the potential influence of sample
111 collection on fish behavior (Figure 1). All tubing was placed before fish were stocked into the pond. In
112 the High CO₂ Area, shade was created near the inflowing water as refuge to entice carp to pass through
113 the area of elevated CO₂. Once-daily, 15 L of the algal feed mixture was injected using a small water
114 pump (Little Giant®, Oklahoma City, OK, USA) under the shaded area and served as an additional
115 attractant for fish to cross the High CO₂ area. Diffusers, shade and feeding were placed on the west half
116 of the pond during Trials 1 and 2. Trial 3 was run as a mirror-image of the first two trials; with diffuser,
117 shade, and feeding moved to the opposite (east) pond half. Study events were identical for all three
118 trials.

119
120 *Acoustic Telemetry* - Fish positions were continuously monitored and recorded using an acoustic
121 telemetry array. One Hydroacoustic Technology Inc. Model 290 acoustic tag receiver (HTI, Seattle, WA,
122 USA) was connected to 16 individual HTI Model 590-series hydrophones. Hydrophones were top-
123 mounted at the water’s surface by suspending each hydrophone from a grid of cable strung above the
124 pond supported with small foam floats. Locations for each hydrophone were manually surveyed from an
125 origin at the bottom of the southwest pond corner. All hydrophones were secured in the same fashion.

126 Ten modified HTI Model 795LD acoustic tags were used for each trial. To eliminate the need for
127 surgical implantation of acoustic tags, tags were modified to allow for external attachment using a T-bar

128 whisker (Floy Tag, Seattle, WA, USA) from methods described in Romine et al. (2015). Modifications
129 included a T-bar whisker, foam insulation (approximately 6 mm diameter) and an acoustic tag inserted
130 into heat shrink tubing (BuyHeatShrink.com, Deerfield, FL, USA) with the ceramic end of the acoustic tag
131 and T-bar whisker protruding from the tubing at the same end. Heat was carefully applied to the heat
132 shrink tubing until all components were securely held in place. Modified tags were placed in water and
133 the foam trimmed to achieve near neutral buoyancy. Individual tags were programmed using a HTI
134 Model 490-LP Acoustic Tag Programmer (ATP) with ping rates ranging from 2059 to 2675 ms. Once
135 activated, presence and persistence of pings were verified using the HTI Model 492-B Acoustic Tag
136 Detector before attachment to fish.

137 During post-processing of acoustic data, valid signal returns were filtered from ambient noise
138 and signal multipath using USGS-developed (public) software, FishCount. Position estimates were then
139 created using USGS-developed (public) software, GeneticFish, from the previous valid detections. Logic
140 filters (i.e., pond borders, distance between detections, velocity, vector angles, and absence of signal)
141 were used to identify and remove any unreasonable position estimates. We determined the accuracy of
142 our telemetry array to be < 0.2 m by comparing known tag locations to post-processed position
143 estimates.

144

145 *CO₂ Trials* - Feed was withheld 48 h before transfer and tagging of fish in the outdoor pond as
146 recommended when sedating fish. All fish were under light sedation using 50 – 100 mg/L AQUI-S®20E (5
147 – 10 mg/L active ingredient eugenol; AQUI-S New Zealand Ltd., Lower Hutt, New Zealand) during
148 transfer (Cupp et al. *in press*). Modified acoustic tags were disinfected for 15 min using a solution of 3%
149 (v/v) Nolvasan® (active ingredient: chlorohexidine diacetate [2%], Ford Dodge Animal Health, Ford
150 Dodge, IA, USA) and rinsed with deionized water. Five bighead and five silver carp were used during
151 each trial. Fish were removed from the sedative solution and the tag was inserted with a T-bar anchor

152 tagging gun at a 45° posterior angle into the dorsal musculature approximately 1 cm lateral and
153 posterior to the mid-line of the dorsal fin. This allowed streamline tag protrusion while swimming and
154 ensured that the T-end was securely implanted (Wydoski and Emergy 1983). All fish appeared to recover
155 in <1 min from the light sedation when placed in the pond. However, fish were given until the following
156 day to fully recover from sedation and handling. Water temperature during the stocking of fish was
157 typical of UMESC well water at $12 \pm 1^\circ\text{C}$. During experimentation, recorded pond temperature was $13.0 \pm$
158 3.0°C (Trial 1), $11.3 \pm 0.4^\circ\text{C}$ (Trial 2), and $8.2 \pm 2.5^\circ\text{C}$ (Trial 3) corresponding with air temperatures at the
159 time of testing. Pond water alkalinity measured throughout testing was $137 \pm 1 \text{ mg/L CaCO}_3$.

160 After recovery, the first trial began and fish positions were continuously recorded for 72 h. All
161 trials were discretized into three sequential 24 h periods: (1) pre-; (2) during; and (3) post-CO₂ injection.
162 No human influence on fish behavior occurred at any time during these 72h trials. Fish positions
163 collected pre- and post-CO₂ were used to determine baseline occupancy and movement of fish
164 throughout the pond in the absence of CO₂. Study events across all three time periods were identical,
165 except for the date of CO₂ injection. No stimuli (e.g. injection of compressed gas) was added during the
166 pre- or post-injection.

167 During CO₂ injection, gas was supplied directly from the port on a single 180-L liquid CO₂ tank
168 (Airgas Inc., La Crosse, WI, USA) connected to a manifold of three flow meters (AngelAqua®, Busan,
169 Korea). Diffusers delivered CO₂ at a rate of 3 L/min. Two peristaltic pumps (Model 7553-70, Cole-
170 Parmer®, Vernon Hills, IL, USA) continuously extracted water through the plastic tubing from six
171 locations (Figure 1). Water samples were collected from pumps and analyzed immediately for carbon
172 dioxide by a modified HACH® Method 8205 digital titration method using sodium hydroxide. Briefly, the
173 100 mL water sample was poured into a 500-mL glass-beaker and placed on a stir plate with a magnetic
174 stir bar. Titrant ($0.3636 \pm 0.00200 \text{ N NaOH}$) was added to the sample using the HACH® Digital Titrator
175 until an endpoint of pH 8.3 (Beckman Coulter Model 410 pH meter, Beckman Coulter Inc., Chaska, MN,

176 USA) was reached. Partial pressures of CO₂ were calculated from pH, temperature, and alkalinity using
177 USGS CO2Calc (<http://pubs.usgs.gov/of/2010/1280/>). Alkalinity was measured using the pH 4.5
178 titrimetric method with 0.02 N H₂SO₄ at various times throughout all three trials from the water samples
179 (APHA 1995). Atmospheric carbon dioxide levels were intermittently measured during each trial using a
180 handheld pSense Model AZ-0001 meter (CO2 Meter Inc., Ormond Beach, FL, USA) to ensure human
181 safety.

182 At the conclusion of the trial, the pond was drained and fish were euthanized using an overdose
183 (200 mg/L) of MS-222. Trials were repeated using the same methods previously described, except for
184 the injection site, shade, and feed being placed on the opposite pond half during Trial 3. All trials began
185 at exactly 07:00 CDT and subsequent transitions between study phases occurred at exactly 24h
186 thereafter. Trials were conducted October 15, 2014 to November 1, 2014.

187
188 *Statistical Analysis* –Exploratory data analysis approaches was used to visualize data (Tukey
189 1977). Fish locations were visualized using density plots for pre-, during and post-CO₂ injection. Fish
190 location counts were also visualized for the same three time periods. Carbon dioxide and pH trends were
191 visualized through time. Fish crossings per hour were visualized using box-and-whisker plots before and
192 after CO₂ concentrations stabilized. All data visualization was done with the ggplot2 package (Wickham
193 2009) in R (R Core Team 2015). Confirmatory statistics were used for parameter estimation and to test
194 the research hypotheses. Generalized linear mixed models (GLMM) with Poisson error terms were used
195 for all statistical comparisons because the three exposure tanks created a nested experimental design
196 and individual fish are pseudo-replicates (Bolker 2008). Trial number and fish ID were included as a
197 random effect. Species and time period (pre, during, and post- injection) were included as fixed effects.
198 For the crossing per hour per fish GLMM, trial had an interaction with CO₂ concentration within the

199 model and was included as a fixed effect. The glmPQL package from the MASS (Venables and Ripley
200 2002) in R was used for all GLMM's. Statistical significance was declared at $\alpha < 0.05$.

201

202

203 **Results**

204 *Water Chemistry* – CO₂ was stable at ambient levels before and after injection (range: 3.8 - 7.6
205 mg/L CO₂; 1 776 – 4 688 μ atm). During injection, CO₂ increased and pH decreased (Figures 2-3). Highest
206 CO₂ concentrations (max = 74.5 \pm 1.9 mg/L; 29 532 – 41 393 μ atm) were found at the injection site
207 relative to the rest of the pond. Lowest CO₂ concentrations after 24 h of injection (max: 34.1 \pm 8.6 mg/L;
208 16 683 – 20 352 μ atm), were found farthest from injection and nearest to the inflowing fresh water. CO₂
209 stabilized at maximum levels approximately 4 – 6 h after injection began (Figure 2).

210

211 *Barrier and Deterrence Efficacy*– Signal transmission stopped from one tag in Trial 2 (n=9) and
212 two tags from Trial 3 (n=8) early in each trial and those tags were subsequently removed from the
213 dataset. No signals were lost during Trial 1 (n=10). Physical retention of tags by fish was 100% at the
214 completion of all three trials. After applying filters to the raw telemetry data, 679 248 (Trial 1), 777 807
215 (Trial 2), and 637 665 (Trial 3) valid fish positions were included in analyses. There were no mortalities
216 during any of the trials.

217 Silver and bighead carp were found significantly more times in the High CO₂ Area during pre- (t
218 =8.40, $p < 0.0001$, $df = 52$) and post-injection ($t = 7.20$, $p < 0.0001$, $df = 52$) compared to when CO₂ was
219 injected (Figure 4). Detections in the High CO₂ Area did not differ between species ($t = -1.329$, $p = 0.19$,
220 $df = 23$). Conversely, silver and bighead carp were detected significantly more times in the Low CO₂ Area
221 during CO₂ injection compared to pre- ($t = -20.63$, $p < 0.0001$, $df = 52$) and post-injection ($t = -20.80$, $p <$
222 0.0001 , $df = 52$; Figure 4). This did not differ by species ($t = -0.444$, $p = 0.66$, $df = 23$). Silver and bighead

223 carp crossed upstream through the injection site significantly fewer times during CO₂ injection
224 compared to pre- ($t = 14.46$, $p < 0.0001$, $df = 52$) and post-injection ($t = 6.76$, $p < 0.0001$, $df = 52$; Figure
225 4). Density plots visualized the change in fish locations for all three trials (Figures 5,6,7). Fish crossed the
226 injection site in both directions and there were no instances where a single fish remained upstream of
227 the CO₂ injection site. Silver carp crossed fewer times than bighead carp ($t = -2.10$, $p = 0.046$, $df = 23$).
228 Carbon dioxide levels began to stabilize at the injection site around 60 mg/L CO₂ (Figure 2). When CO₂
229 levels were > 60 mg/L (approximately $> 24\ 000\ \mu\text{atm}$), silver and bighead carp passed through the
230 injection site significantly fewer times per hour relative to concentrations ≤ 60 mg/L CO₂ ($t = -9.22$, $p <$
231 0.0001 , $df = 652$; Figure 8). Silver carp also passed through fewer times than bighead carp ($t = -2675$, $p =$
232 0.016 , $df = 16$). Trial 3 had the lowest number of crosses ($t = -6.48$, $p < 0.0001$, $df = 652$).

233

234 Discussion

235 The potential for CO₂ to function as a non-physical deterrent has been recently described in the
236 literature (Noatch and Suski 2012; Kates et al. 2012), and our study was the first to quantify the
237 effectiveness of a CO₂ barrier to invasive carps movement. We found that that CO₂ injection was
238 effective to reduce the upstream movement of silver and bighead carp, but did not completely stop fish
239 passage. Similarly, we observed a significant shift in fish movement away from areas of high CO₂. Most
240 notably, results were consistent across all three trials, including when CO₂ was injected into the opposite
241 side of the pond during Trial 3, providing evidence that CO₂ modified silver and bighead carp behavior.
242 This study demonstrates the potential use of CO₂ to deter silver and bighead carp movement and results
243 contribute to a growing body of research evaluating the use of CO₂ as a deterrent to invasive fishes.

244 Many fish species have been shown to avoid CO₂ during laboratory experiments. For instance,
245 Clingerman et al. (2007) used 60 – 120 mg/L dissolved CO₂ for non-physical transfer of rainbow trout
246 (*Oncorhynchus mykiss*) between large aquaculture tanks with 100 percent effectiveness. A similar

247 approach was conducted by Kates et al. (2012), where avoidance responses were documented from
248 silver carp, bluegill (*Lepomis macrochirus*), and largemouth bass (*Micropterus salmoides*) at
249 concentrations ≥ 100 mg/L CO₂. Further, blacknose dace (*Rhinichthys atratulus*) and brook trout
250 (*Salvelinus fontinalis*) exhibited avoidance behavior when exposed to 120 mg/L CO₂ in small laboratory
251 tanks (Ross et al. 2001). When in our outdoor pond, we observed similar avoidance behavior of silver
252 and bighead carp at concentrations around 70 mg/L CO₂ and found that this technique could potentially
253 be used to deter upstream movement.

254 The sensitivity of fish to elevated CO₂ warrants continued investigation into the use of this non-
255 physical method. However, with any non-physical deterrence strategy, less than 100% deterrence must
256 be an acceptable outcome (Noatch and Suski 2012; Wittman et al. 2014). During periods of hypercapnia,
257 fish undergo physiological detriments such as ion imbalance (Claiborne et al. 2002), increased stress
258 response (Kates et al. 2012), acidosis (Ishimatsu et al. 2005), hyperventilation (Perry and Abdallah 2012),
259 hypoventilation (Kates et al. 2012), loss of sensory function (Nilsson et al. 2012), and changes in protein
260 composition (Dennis et al. 2015). In conjunction with elevated CO₂, a corresponding decrease in pH
261 occurs due to the hydration equilibrium of CO₂ to form bicarbonate (HCO₃⁻) and carbonic acid (H₂CO₃).
262 This decrease in pH may play a role in behavioral avoidance during hypercapnia due to the influence of
263 pH on biochemical processes, however, several studies have shown that fish specifically sense and avoid
264 CO₂ (Ishimatsu et al. 2005; Perry and Gilmour 2006; Perry and Abdallah 2012).

265 Consideration should be given to the methods used to quantify and units reported of dissolved
266 CO₂, for comparisons with previous literature. Our study quantified CO₂ using methods described in
267 Clingerman et al. (2007), Kates et al. (2012), Dennis et al. (2015), and Donaldson et al. (2016) to allow
268 direct comparisons of the results. Free CO₂ can be easily related with pH, which may also facilitate
269 transition to the field using common water chemistry data loggers (Kates et al. 2012). However, CO₂ gas
270 saturation is dependent on temperature, pressure, alkalinity, biological status and pH. Common units for

271 carbon dioxide are reported or derived in terms of pressure (e.g. PCO_2 , mmHg, μatm). While there is a
272 general lack of uniformity in units quantifying CO_2 across current research, recognizing units are
273 important for comparison of results.

274 Results from our study identify many of the consideration with CO_2 as a control tool in fisheries
275 management. For example, the U.S. Army Corps of Engineers operates and maintains an electrical
276 barrier in the CAWS near Romeoville, IL, USA to inhibit further upstream movement of silver and
277 bighead carp into the Great Lakes basin (Moy et al. 2011). While the electric barrier is currently the most
278 important management tool to deter upstream movement of carp in the Great Lakes, there is a need for
279 redundancy and multiple integrated deterrence mechanisms (Noatch and Suski 2012). Additionally,
280 Noatch and Suski (2012) expressed concern that electricity loses effectiveness on smaller fish due to a
281 reduction in total body voltage at smaller body sizes (Reynolds 1996; Dolan and Miranda 2003). The
282 potential for size-related effectiveness with the electric barrier further supports the need for alternative
283 and supplemental deterrence strategies. In our study, CO_2 deterred stock length silver carp and sub-
284 stock length bighead carp (Phelps and Willis 2013). With branchial CO_2 receptors present throughout the
285 life-history of most fish (Perry and Abdallah 2012), the absence of size-related effectiveness suggests the
286 CO_2 could be used to supplement other barriers to deter smaller fish passage.

287 Carbon dioxide may be useful to supplement other barriers (e.g. lock and dams or electrical
288 barriers). During scheduled usage or maintenance, CO_2 could be beneficial to deter silver and bighead
289 carp until primary barriers are restored. For instance, CO_2 could be applied to reduce fish from entering
290 or passing gates during planned vessel lockage. One important observation during our study was that
291 adequate time must be allowed for CO_2 to accumulate to target concentrations to be most effective at
292 deterring fish (e.g. Figures 5,6,7). We observed 4 -6 h for CO_2 levels to stabilize using a simple diffusion
293 injection system and recommend exploring improved designs to reduce build-up time. Reducing the
294 build-up time would expand the utility of CO_2 under different applications. Further, understanding

295 operational efficiency of individual CO₂ injection systems under different scenarios will be important
296 during rapid implementation (e.g. unexpected primary barrier failure) and for calculating operational
297 costs. For rapid implementation, other deterrent strategies (e.g. mobile electrical barriers, sound-
298 bubble-strobes, or acoustic noise) may be considered due to their on-and-off style of operation (Taylor
299 et al. 2005; Ruebush et al. 2012; Vetter et al. 2015).

300 Carbon dioxide was not an absolute barrier to silver and bighead carp in our outdoor ponds.
301 While the success of any barrier is binomial, it is possible that total crossings in our study may have been
302 inflated due to the relatively small spatial size of our test pond. With recorded burst speeds of 1.28 –
303 1.66 m/s (Hoover et al. 2012), silver and bighead carp in our 10 m pond could have moved upstream
304 through the injection site before sensing elevated CO₂. This hypothesis is further supported by the
305 reduced time that fish spent in the High CO₂ Area. In our study, fish consistently avoided areas of
306 elevated CO₂ when given an option of fresh-water. Further, when concentrations were greater than 60
307 mg/L CO₂ (~ 24 000 µatm) we also observed the most effective reduction in upstream movement
308 relative to lower concentrations, suggesting a potential dose-response effectiveness (Figure 8). Higher
309 concentrations may be considered for future research. Complementary to the statistical comparisons,
310 visualized distributions of fish in the pond shown in Figures 5,6,7 demonstrate the clear shift in fish
311 locations relative to CO₂ injection. Most notably, similar movement away from the injection site during
312 Trial 3 when CO₂ was moved to the opposite side of the pond confirmed that CO₂ deterred fish. Testing
313 on a larger spatial scale can likely address this shortcoming and application of CO₂ on a larger spatial
314 scale may increase its effectiveness as a deterrent, especially when access to fresh water is made readily
315 available. Locations such as the entrance to approach channels at lock and dams or tributaries on large
316 rivers where fish can choose to stay out in the main channel should be considered for further testing.

317 Carbon dioxide may provide a safer alternative than other non-physical methods, primarily the
318 electrical barrier in the CAWS. Measurements taken by the U. S. Coast Guard conclude that the

319 electrical barrier can result in a lethal shock to humans if they fall into the water near the barrier (ADRSC
320 2011). Carbon dioxide exists naturally in the atmosphere as a byproduct of aerobic respiration and
321 burning of fossil fuels at concentrations of 0.038 - 0.04 percent (NOAA 2015). Acceptable human
322 exposure limits of atmospheric CO₂ established by the Occupational Safety and Health Administration
323 (OSHA) were set at 5 000 ppm as a time weighted average (TWA) for an 8 h exposure and 30 000 ppm
324 for acute exposures (OSHA 29 CFR 1910.1000). Atmospheric CO₂ measured before and during our trials
325 were stable at a range of 433 – 498 ppm and did not increase during periods of CO₂ injection. While
326 these concentrations are well below levels generating safety concerns, adequate monitoring should be
327 incorporated when CO₂ is applied at a larger field-scale.

328

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447 **Figure Captions**

448 Figure 1: Outdoor concrete pond setup at the Upper Midwest Environmental Sciences Center in La
449 Crosse, WI during the evaluation of CO₂ as a barrier and deterrent to silver
450 (*Hypophthalmichthys molitrix*) and bighead carp (*H. nobilis*) conducted October 15, 2014 –
451 November 1, 2014. Graphical illustration shows setup during Trials 1 and 2. Trial 3 was a
452 reversed setup, with CO₂ injection moved to the opposite side of the center partition. Black
453 triangles show approximate pH and CO₂ spatial sampling locations within the pond: NW
454 (northwest), MW (middle-west), SW (southwest), NE (northeast), ME (middle-east), and SE
455 (southeast).

456

457 Figures 2,3: Carbon dioxide (CO₂) and pH levels in outdoor ponds during the evaluation of CO₂ as a
458 barrier and deterrent to silver (*Hypophthalmichthys molitrix*) and bighead carp (*H. nobilis*)
459 conducted October 15, 2014 – November 1, 2014 at the Upper Midwest Environmental
460 Sciences Center in La Crosse, WI. We observed increases in CO₂ levels (Figure 2) and decreases
461 in pH (Figure 3) throughout the entire pond during the entire 24h that CO₂ was injected into
462 the pond. Injection was located at MW during Trials 1 and 2. During Trial 3, CO₂ was injected
463 at ME. Locations where values were recorded in the pond are defined as: NW (northwest),
464 MW (middle-west), SW (southwest), NE (northeast), ME (middle-east), and SE (southeast).
465 Values describe a single measurement at each location and time.

466

467 Figures 4: Occupancy and behavior of silver (*Hypophthalmichthys molitrix*) and bighead carp (*H.*
468 *nobilis*) in outdoor ponds. Detections in the High CO₂ Area (top) decreased significantly while
469 CO₂ was injected into the pond, while detections the Low CO₂ Area (middle) increased

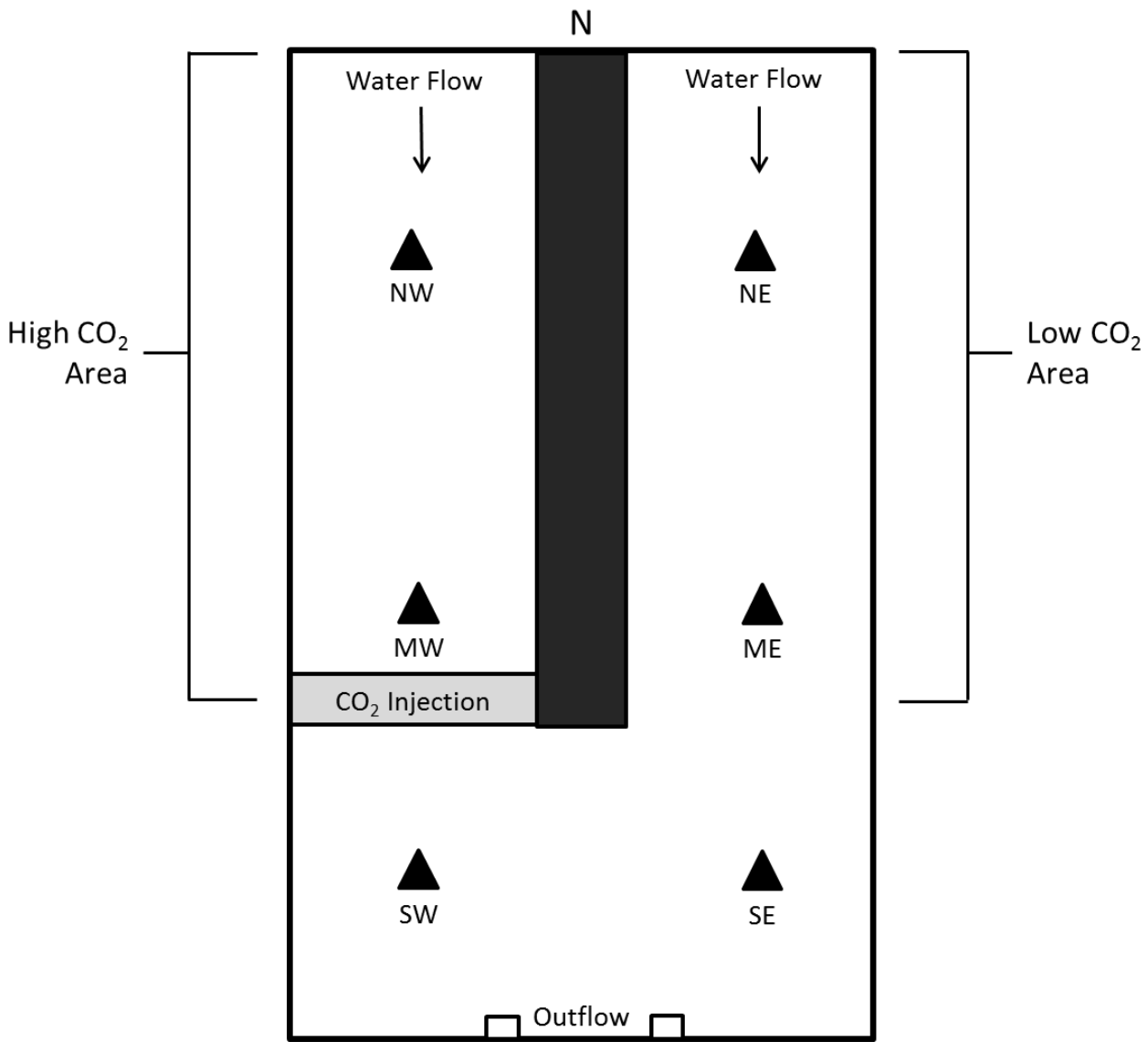
470 significantly. Crosses through the CO₂ injection site were significantly reduced during injection
471 (bottom), but CO₂ was not 100% effective to stop upstream fish passage.

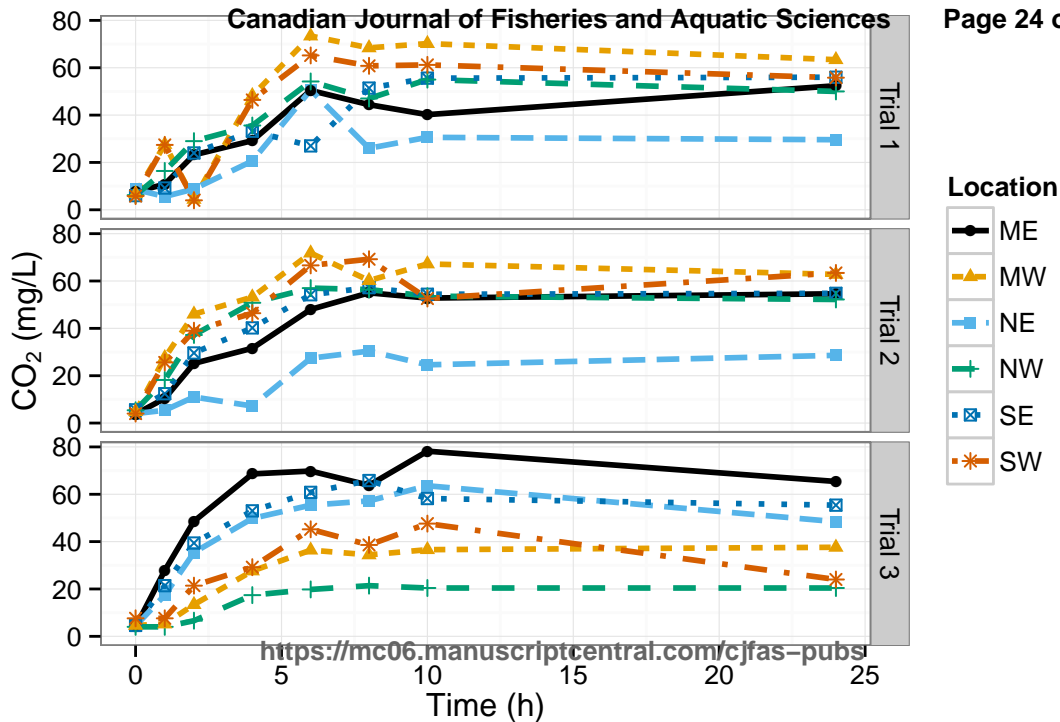
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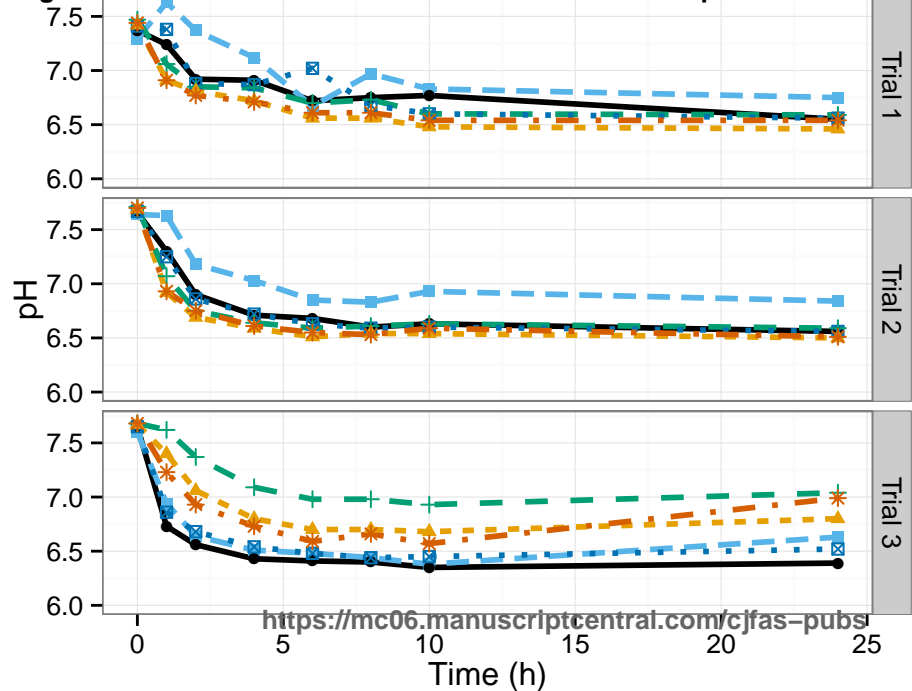
473 Figure 5,6,7: Density plots illustrating the spatial distribution of silver (*Hypophthalmichthys molitrix*)
474 and bighead carp (*H. nobilis*) during the evaluation of carbon dioxide (CO₂) as a barrier and
475 deterrent in outdoor ponds. Independent trials 1 (Figure 5), 2 (Figure 6), and 3 (Figure 7) were
476 discretized into three 24h periods: before (PreCO₂), during (DuringCO₂), and after (PostCO₂)
477 CO₂ injection. The red line represents the approximate location of the CO₂ barrier.

478

479 Figure 8: Box-and-whisker plots describing the movement of silver (*Hypophthalmichthys molitrix*) and
480 bighead carp (*H. nobilis*) upstream through a CO₂ injection site in outdoor concrete ponds. The
481 effectiveness of lower concentrations (≤ 60 mg/L CO₂) to impede upstream movement was
482 compared to higher concentrations (> 60 mg/L CO₂). Carp moved upstream significantly fewer
483 times when CO₂ was ≥ 60 mg/L CO₂ ($p < 0.01$) relative to lower concentrations. No differences
484 between species were observed ($p = 0.17$).

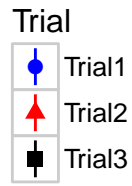
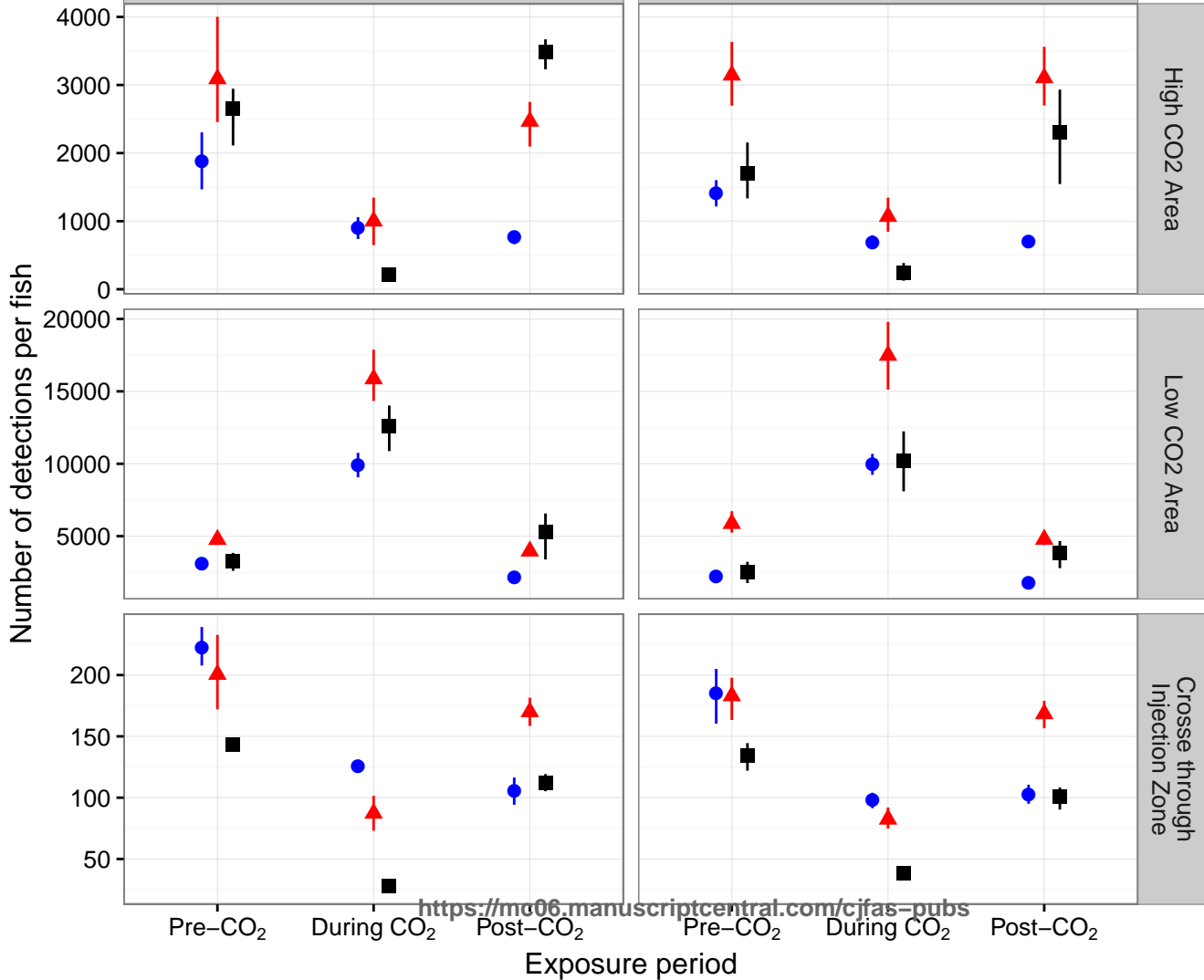


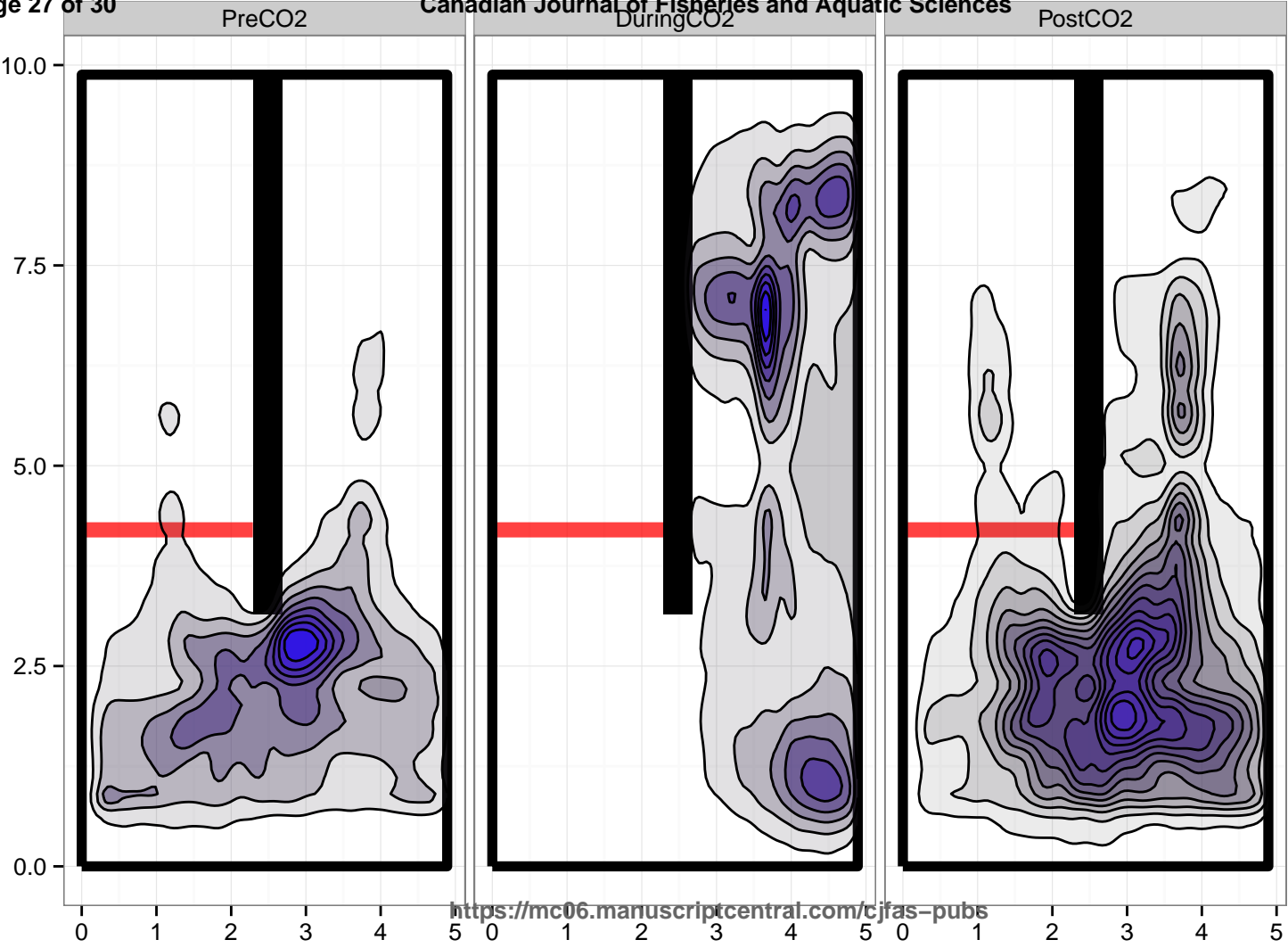


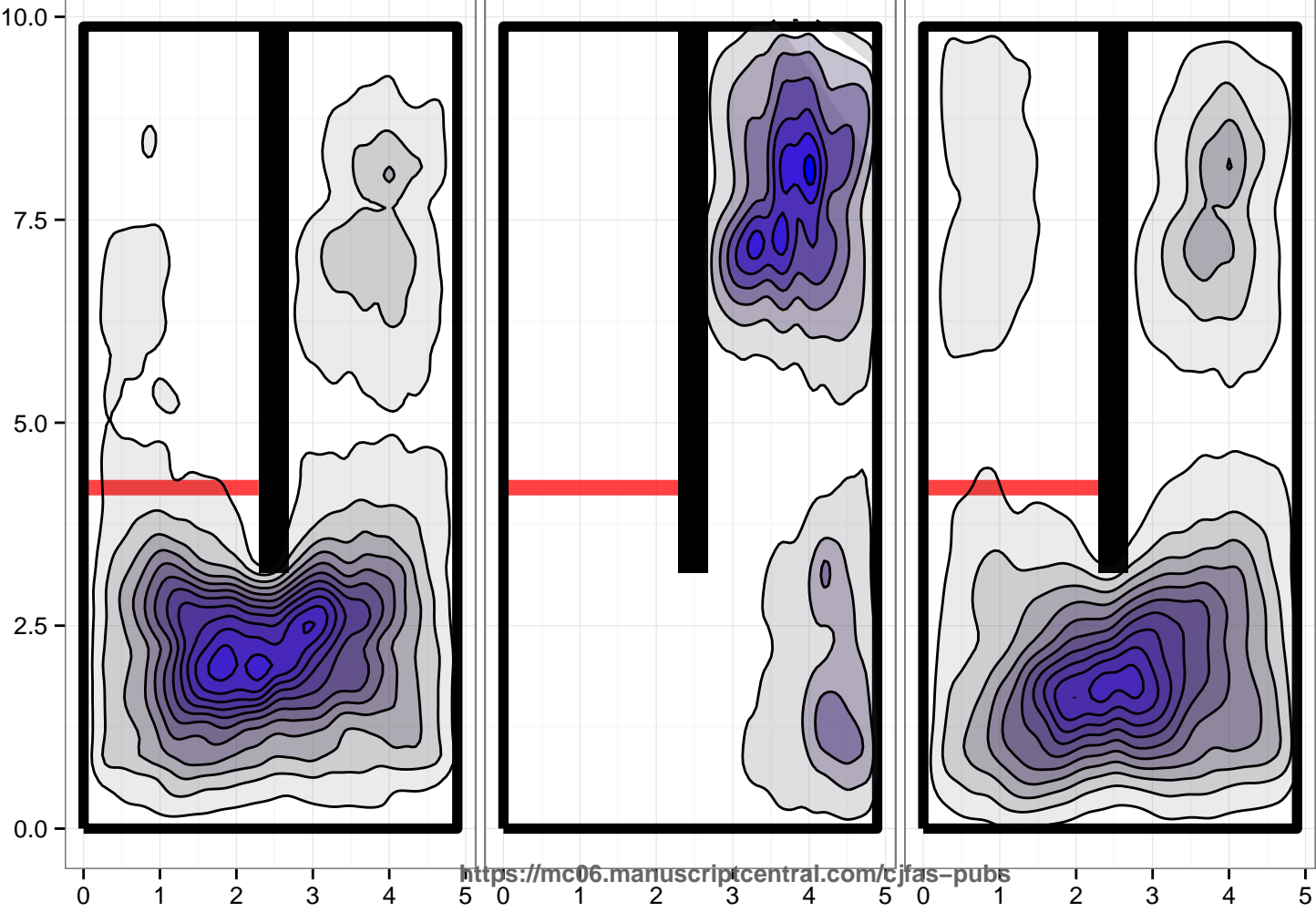


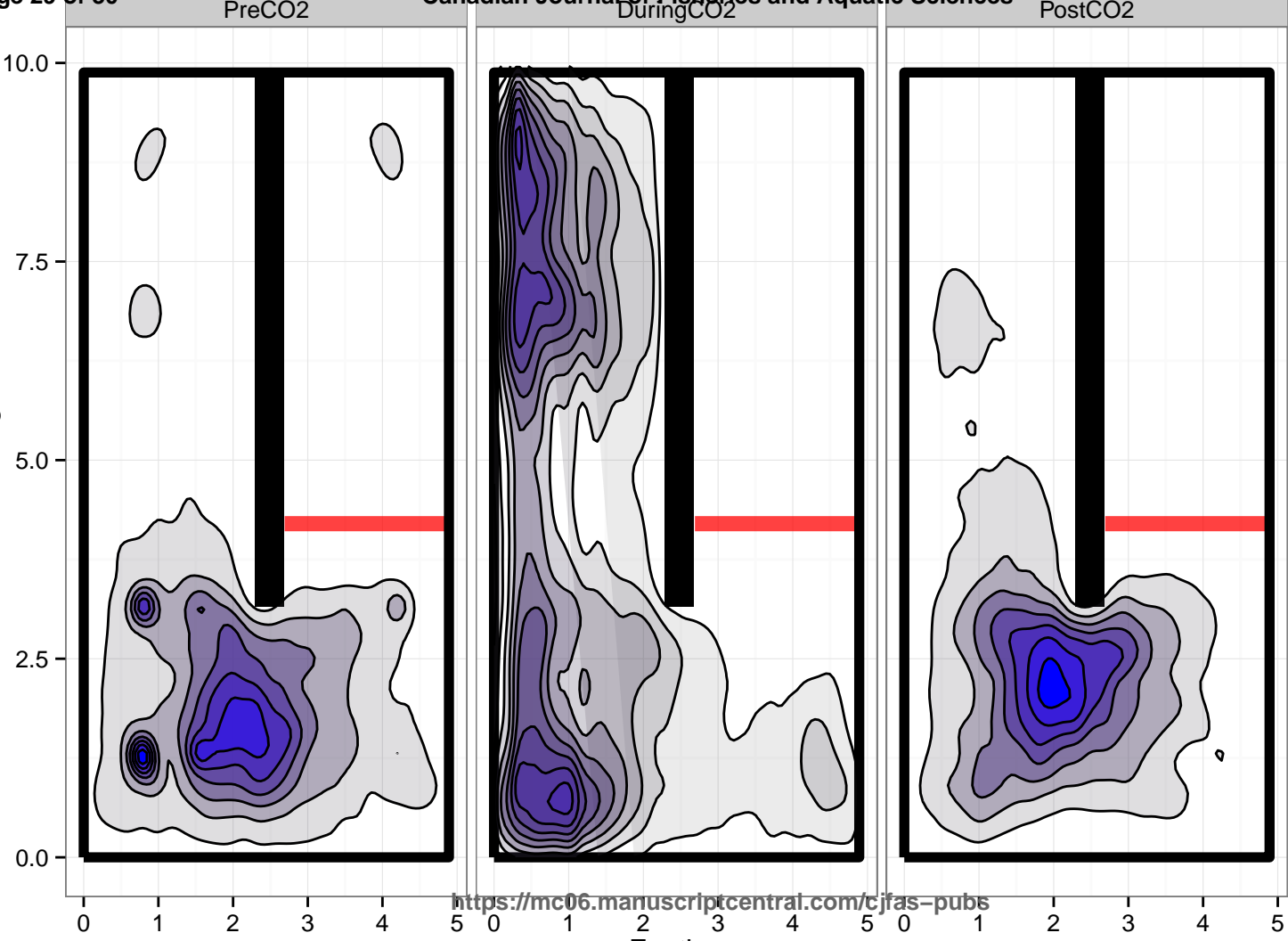
Location

- ME
- MW
- NE
- NW
- SE
- SW





PreCO₂DuringCO₂PostCO₂



Crossing per hour per fish

25
20
15
10
5
0 ≤ 60 (mg/L) > 60 (mg/L) ≤ 60 (mg/L) > 60 (mg/L) ≤ 60 (mg/L) > 60 (mg/L)CO₂ concentration<https://mc.manuscriptcentral.com/cjfas-pubs>