

## Rhizobia Catabolize *nod* Gene-Inducing Flavonoids via C-Ring Fission Mechanisms

JULURI RAGHAVENDRA RAO AND JAMES E. COOPER\*

Department of Applied Plant Science, The Queen's University of Belfast, Belfast BT9 5PX,  
Northern Ireland, United Kingdom

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Gas chromatographic and mass spectrometric analyses of derivatized culture medium extracts were used to identify the products of flavonoid metabolism by rhizobia. A number of *Rhizobium* species and biovars degraded their *nod* gene-inducing flavonoids by mechanisms which originated in a cleavage of the C-ring of the molecule and which yielded conserved A- and B-ring products among the metabolites. In contrast, *Pseudomonas putida* degraded quercetin via an initial fission in its A-ring, and *Agrobacterium tumefaciens* displayed a nonspecific mode of flavonoid degradation which yielded no conserved A- or B-ring products. When incubated with rhizobia, flavonoids with OH substitutions at the 5 and 7 positions yielded phloroglucinol as the conserved A-ring product, and those with a single OH substitution at the 7 position yielded resorcinol. A wider range of structures was found among the B-ring derivatives, including *p*-coumaric, *p*-hydroxybenzoic, protocatechuic, phenylacetic, and caffeic acids. The isoflavonoids genistein and daidzein were also degraded via C-ring fission by *Rhizobium fredii* and *Rhizobium* sp. strain NGR234, respectively. Partially characterized aromatic metabolites with potential *nod* gene-inducing activity were detected among the products of naringenin degradation by *Rhizobium leguminosarum* bv. *viciae*. The initial structural modification of *nod* gene-inducing flavonoids by rhizobia can generate chalcones, whose open C-ring system may have implications for the binding of inducers to the *nodD* gene product.

Flavonoids are polyphenolic secondary metabolites which are synthesized by plants via the expression of two multigene-encoded enzymes: phenylalanine ammonia lyase and chalcone synthase. Subgroups of compounds such as chalcones, flavanones, flavones, flavonols, and isoflavonoids occur in legume tissues, and they can be released (15) from roots into the rhizosphere, where some of them act as molecular signals to trigger the establishment of symbioses with bacteria in the family *Rhizobiaceae* (18, 19). Their principal function is to interact with the *nodD* gene products of rhizobia and the subsequent transcriptional activation of other *nod* genes (17). Other effects of flavonoids on rhizobia include promotion of chemotactic responses (1) and stimulation of growth rate by unspecified mechanisms (8).

Although rhizobia are known to utilize various aromatic compounds as carbon and/or energy sources by degrading them to catechol and protocatechuate and channelling these products after further enzymatic cleavage into the tricarboxylic acid cycle via the  $\beta$ -ketoacid pathway (3, 16), their capacity to degrade flavonoids has received little attention. Only two examples have been reported: the utilization of catechin by a *Rhizobium* sp. isolated from *Leucaena leucocephala*, with attendant formation of phloroglucinol carboxylic acid and protocatechuate (6), and a novel form of C-ring cleavage in a pentahydroxy flavone, quercetin, by *Rhizobium loti* (20). One report (4) has described an alteration in the types and amounts of formononetin derivatives in *Medicago sativa* root exudate during incubation with *Rhizobium meliloti*, but no biotransformation mechanism was proposed. Formononetin glycosides are *nod* gene inducers for *R. meliloti*.

The purpose of the present study was to investigate the

metabolic activity of rhizobia towards their *nod* gene-inducing flavonoids. We used gas chromatography and mass spectrometry (GC-MS) to facilitate the direct detection and identification of submilligram quantities of aromatic metabolites in supernatants of liquid media which had been supplemented with flavonoids and incubated with single-strain cultures of various *Rhizobium* species and biovars. We now report that rhizobia degrade *nod* gene-inducing flavonoids by mechanisms which originate in a cleavage of the C-ring of the molecule.

### MATERIALS AND METHODS

**Bacteria.** The bacterial strains used and their sources were as follows: *Rhizobium leguminosarum* bv. *trifolii* P3, this laboratory; *R. leguminosarum* bv. *viciae* RBL5601 (containing the wild-type plasmid pJB5JI), Department of Plant Molecular Biology, Leiden University, Leiden, The Netherlands; *R. leguminosarum* bv. *phaseoli* TAL182, *R. meliloti* RM41, *Rhizobium fredii* HH103, *Pseudomonas putida* DSM3226, and *Agrobacterium tumefaciens* C6-6, Botany Department, Marburg University, Marburg, Germany; and *Rhizobium* sp. strain NGR234, LBMPS, University of Geneva, Geneva, Switzerland.

Strains were grown individually in shake-flask cultures in flavonoid-free medium with mannitol replacing arabinose (20). Cultures were grown at 25°C for 72 h; this step was followed by centrifugation and washing (twice) and resuspension in sterile water.

**Test media and growth conditions.** For strain NGR234 the test medium was that of Broughton and Dilworth (2). For all other strains the test medium was as described by Rao et al. (20), except that mannitol was substituted for arabinose. High-pressure liquid chromatography (HPLC)-grade flavonoids were obtained from Apin Chemicals, Abingdon, United Kingdom, and rechecked for purity by GC. Flavonoids were added from stock solutions in methanol to give a final concentration of 10  $\mu$ M. Media were dispensed in 1-liter

\* Corresponding author. Mailing address: Department of Applied Plant Science, The Queen's University of Belfast, Newforge Ln., Belfast BT9 5PX, Northern Ireland, United Kingdom. Phone: 44 232 661166. Fax: 44 232 668375.

TABLE 1. Principal aromatic metabolites detected after incubation<sup>a</sup> of flavonoids with *Rhizobium*, *Pseudomonas*, and *Agrobacterium* species

Organism	Flavonoid substrate <sup>b</sup>	Ring cleavage mechanism	Principal degradation product(s) derived from:		Other partially characterized metabolites (MS data)	
			Conserved A-ring	Conserved B-ring	M <sup>+</sup>	Major ion peaks (m/z)
<i>R. leguminosarum</i> bv. <i>viciae</i>	Naringenin	C-ring	Phloroglucinol	<i>p</i> -Coumaric acid, <i>p</i> -hydroxybenzoic acid	300	65, 91, 134, 166, 193 (peak F [Fig. 1])
					300	69, 77, 121, 134, 166 (peak E [Fig. 1])
					314	55, 134, 180, 207, 286 (peak G [Fig. 1])
					314	69, 91, 134, 152, 180, 207, 271, 297 (peak H [Fig. 1])
<i>R. leguminosarum</i> bv. <i>phaseoli</i>	Quercetin	C-ring	Phloroglucinol	Protocatechuic acid		
<i>R. leguminosarum</i> bv. <i>trifolii</i>	7,4'-Dihydroxyflavone	C-ring	Resorcinol	<i>p</i> -Hydroxybenzoic acid		
<i>R. meliloti</i>	Luteolin	C-ring	Phloroglucinol, phloroglucinol carboxylic acid	Caffeic acid, protocatechuic acid, phenylacetic acid	298	69, 132, 135, 171, 269 (corresponds to apigenin)
<i>R. fredii</i>	Genistein	C-ring	Phloroglucinol carboxylic acid	<i>p</i> -Coumaric acid, <i>p</i> -hydroxybenzoic acid	174	51, 77, 87, 102, 115, 131, 159 (umbelliferoyl derivative)
<i>Rhizobium</i> sp. strain NGR234	Daidzein	C-ring	Resorcinol	<i>p</i> -Coumaric acid, <i>p</i> -hydroxybenzoic acid		
<i>Rhizobium</i> sp. strain NGR234	Apigenin	C-ring	Phloroglucinol	<i>p</i> -Hydroxybenzoic acid, phenylacetic acid		
<i>P. putida</i>	Quercetin	A-ring	Oxaloacetic acid (nonaromatic)	Protocatechuic acid		
<i>A. tumefaciens</i>	Naringenin	Nonspecific				

<sup>a</sup> The incubation period was 24 h for all metabolites depicted except the partially characterized products of naringenin degradation by *R. leguminosarum* bv. *viciae*, which were detected within 2 h.

<sup>b</sup> Each flavonoid is an effective *nod* gene inducer for its corresponding *Rhizobium* sp.

amounts in 3-liter Erlenmeyer flasks and inoculated with single cultures of bacteria to give an initial concentration of  $1.5 \times 10^3$  cells per ml. Unless otherwise specified, incubation was at 25°C on a rotary shaker (200 rpm) for a maximum period of 24 h. Control treatments required for confirmation of bacterial flavonoid transformations were also included as follows: basal medium plus flavonoid with no inoculum and basal medium minus flavonoid with inoculum. Flasks were wrapped in aluminum foil to prevent photosensitive reactions from occurring.

**Analysis of culture extracts.** After incubation, 1-liter aliquots of media were centrifuged ( $8,000 \times g$  for 10 min) and the supernatants were extracted by hand (six times) with ethyl acetate after saturation with sodium chloride. This was followed by drying with  $\text{Na}_2\text{SO}_4$  and concentration on a rotary evaporator. Residues were taken up in 0.5 ml of methanol and treated with an excess of freshly prepared ethereal diazomethane using a Diazald kit (Aldrich Chemicals) at 0°C. After 12 h, the solvents, along with excess diazomethane, were evaporated, and the remaining residues were dissolved in 0.5 ml of diethyl ether prior to GC-MS analysis (20).

## RESULTS

**Principal degradation products.** The main aromatic products detected in the supernatants of flavonoid-supplemented media are presented in Table 1. Rhizobia consistently degraded flavonoids via C-ring cleavage, whereas *P. putida*, as expected, degraded quercetin by an initial fission in the A-ring of the molecule. *A. tumefaciens* displayed a nonspecific mode of degradation towards naringenin, which yielded no conserved A- or B-ring products. Analysis of mass spectral data from derivatized samples obtained from control flasks failed to detect any of the monocyclic, hydroxylated aromatics which

were found in the inoculated, flavonoid-supplemented treatments.

In the presence of *Rhizobium* spp., flavonoids with OH substitutions at the 5 and 7 positions yielded phloroglucinol as the conserved A-ring product. 7,4'-Dihydroxyflavone, which has a single OH substitution on its A-ring, yielded resorcinol as the conserved product from this part of the molecule. A wider range of metabolites was found among the B-ring derivatives. For example, *R. leguminosarum* bv. *viciae* yielded *p*-coumaric acid and its by-product, *p*-hydroxybenzoic acid. The same metabolites would be predicted from a C-ring fission of 7,4'-dihydroxyflavone by *R. leguminosarum* bv. *trifolii*, but only *p*-hydroxybenzoic acid was detected. *R. leguminosarum* bv. *phaseoli* generated protocatechuic acid, as expected from the OH substitution pattern of its *nod* gene inducer, quercetin. *R. meliloti* generated three compounds originating in the B-ring of luteolin: phenylacetic acid, caffeic acid, and protocatechuic acid. The isoflavonoids genistein and daidzein were metabolized by *R. fredii*, while the broad-host-range *Rhizobium* sp. strain NGR234 also degraded daidzein and apigenin.

The total ion gas chromatogram of the extracted and derivatized supernatant from naringenin-supplemented medium after incubation for 2 h with *R. leguminosarum* bv. *viciae* is shown in Fig. 1. The retention times and mass spectra of peaks A, B, and C were identical to those of similarly derivatized authentic phloroglucinol, *p*-hydroxybenzoic acid, and *p*-coumaric acid, respectively. Peak D represents residual, unmetabolized naringenin, peaks E and F represent new flavanone ( $M^+$  300) structures, and peaks G and H represent new chalcone ( $M^+$  314) structures. The relative quantities of derivatives in the supernatant extracts may not truly reflect their concentrations in the culture medium. This is most likely

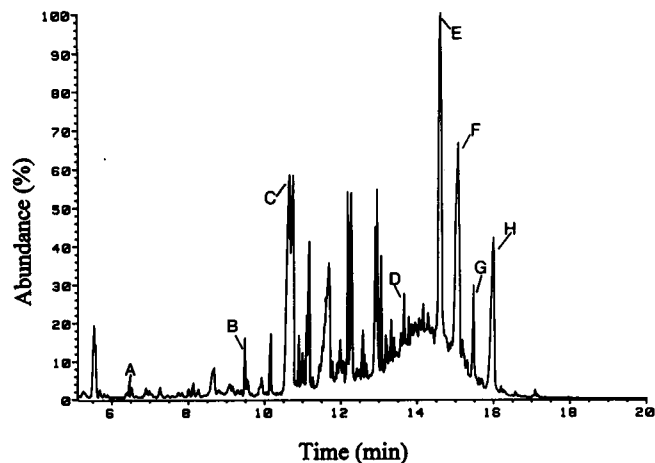


FIG. 1. Total ion gas chromatogram of methylated ethyl acetate extracts of naringenin-supplemented medium after incubation for 2 h with *R. leguminosarum* bv. *viciae*. Peaks: A, phloroglucinol; B, *p*-hydroxybenzoic acid; C, *p*-coumaric acid; D, naringenin; E and F, new flavanone structures; G and H, new chalcone structures. Compounds represented by peaks E, F, G, and H do not persist at detectable levels upon further incubation.

to be the case for phloroglucinol, which is highly water soluble and therefore not fully extractable with ethyl acetate.

**Partially characterized metabolites.** Other aromatic metabolites, some of which could be only partially characterized from the mass spectra of their methyl ether derivatives, were also present among the degradation products. In addition to the previously mentioned chalcone and flavanone structures generated from naringenin by *R. leguminosarum* bv. *viciae*, further examples are indicated in Table 1. A compound whose mass spectrum corresponded to that of a trimethyl ether derivative of apigenin was identified among the metabolites of luteolin after incubation with *R. meliloti*, and an umbelliferol derivative was detected among the products of genistein degradation by *R. fredii*.

**Degradation pathways.** Proposed degradation mechanisms for seven flavonoids are presented in Fig. 2. In each case the first product of C-ring fission is shown as an intermediate before further cleavage releases the conserved A- and B-ring metabolites. For example, the formation of phloroglucinol from ring A and of *p*-coumaric acid from ring B of naringenin indicates that the C-O bond in the C-ring undergoes an initial fission to generate a chalcone structure and that the C-2-C-3 bond is concomitantly reduced to a double bond. In the case of luteolin, the detection of apigenin and tetrahydroxy flavanone among the metabolites was indicative of C-ring closure and isomerization of a pentahydroxy chalcone structure in addition to the multiple C-ring fissions required to yield the array of conserved A- and B-ring products. The nature of the conserved B-ring products from genistein and daidzein metabolism suggests the operation of aryl shifts prior to C-ring fission in naringenin chalcone and trihydroxy chalcone. In all treatments, the low concentrations of flavonoid substrate, growth medium conditions, and rapid formation of end products limited our ability to detect the first, transient products of C-ring fission. Chalcone structures in particular would be expected to have a very weak persistency in such a physiologically active environment. HPLC analyses of large-scale extracts of inoculated media confirmed the presence of those products detected in the GC-MS analyses, but the additional

intermediate compounds predicted from the initial C-ring fissions were not recovered.

## DISCUSSION

Our results clearly show that rhizobia degrade a wide range of flavonoids by means of primary fissions located exclusively in the C-rings of the compounds. This type of flavonoid catabolism has previously been reported for members of two other bacterial genera: some intestinal *Clostridium* strains and an *Eubacterium* species (14, 25). While *A. tumefaciens* was able to catabolize naringenin, the fission mechanisms differed from those observed in *R. leguminosarum* bv. *viciae*, and no conserved A- or B-ring products were detected. The general catabolic mechanism displayed by rhizobia can be regarded as a reversal of plant flavonoid biosynthesis, which is characterized by the condensation of 3 U of malonyl coenzyme A (the phloroglucinol A-ring unit) and 1 U of *p*-coumaroyl coenzyme A (the B- and C-ring component) to yield the basic three-ring structure. More aromatic products were detected in the GC-MS analyses than are shown in Table 1 and Fig. 2. However, we have deliberately concentrated on those products associated directly with the ring cleavage mechanisms operating in these experiments and on other monocyclic aromatics which can be regarded as satellite molecules derived from the conserved A- or B-ring metabolites.

Two particularly interesting features of the rhizobial degradation pathways are the formation of chalcones as the first intermediates of C-ring fission in most cases and the appearance of new chalcone, flavanone, and flavone structures among the metabolites when naringenin and luteolin were incubated with *R. leguminosarum* bv. *viciae* and *R. meliloti*, respectively. Compounds with these structures are themselves potential *nod* gene inducers and could be involved in interactions with the *nodD* gene product. Chalcones especially have been reported to serve as more potent *nod* gene inducers than other flavonoid molecules (9), and this feature has largely been attributed to their open C-ring system (7), which offers increased spatial flexibility during interactions with NodD proteins (5). Chalcones emerging from an initial C-ring fission in the inducer flavonoid may provide additional conformational flexibility during binding to NodD proteins, thereby increasing transcriptional activation of other *nod* genes.

Our data suggest that flavonoid C-ring cleavage is conserved in all *Rhizobium* species tested and might be a form of natural catabolic response to the basic flavonoid ring system. This implies that such biotransformations could also be anticipated for other, non-gene-inducing flavonoids, which, because of their incompatible hydroxylation patterns or the steric arrangement of their ring systems, do not interact with NodD proteins (9). The preference of NodD proteins for certain flavonoids further suggests that the receptor protein is in direct contact with the inducer (7), perhaps in the cytoplasmic membrane wherein the NodD protein is located (24), and the inducer has also been shown to accumulate in *R. leguminosarum* bv. *viciae* (22). Further studies of inducer-NodD protein binding mechanisms would be required to ascertain the significance of flavonoid C-ring cleavage at the subcellular level during *nod* gene induction. Since all products were detected in culture supernatants, it is not possible to say whether they represent compounds that are released from cells into the medium or whether degradation is an extracellular process.

It is interesting to speculate on the significance of flavonoid degradation pathways for the general biology of rhizobia. As with other aromatic compounds (16), degradation could allow rhizobia to utilize flavonoids as carbon or energy sources.

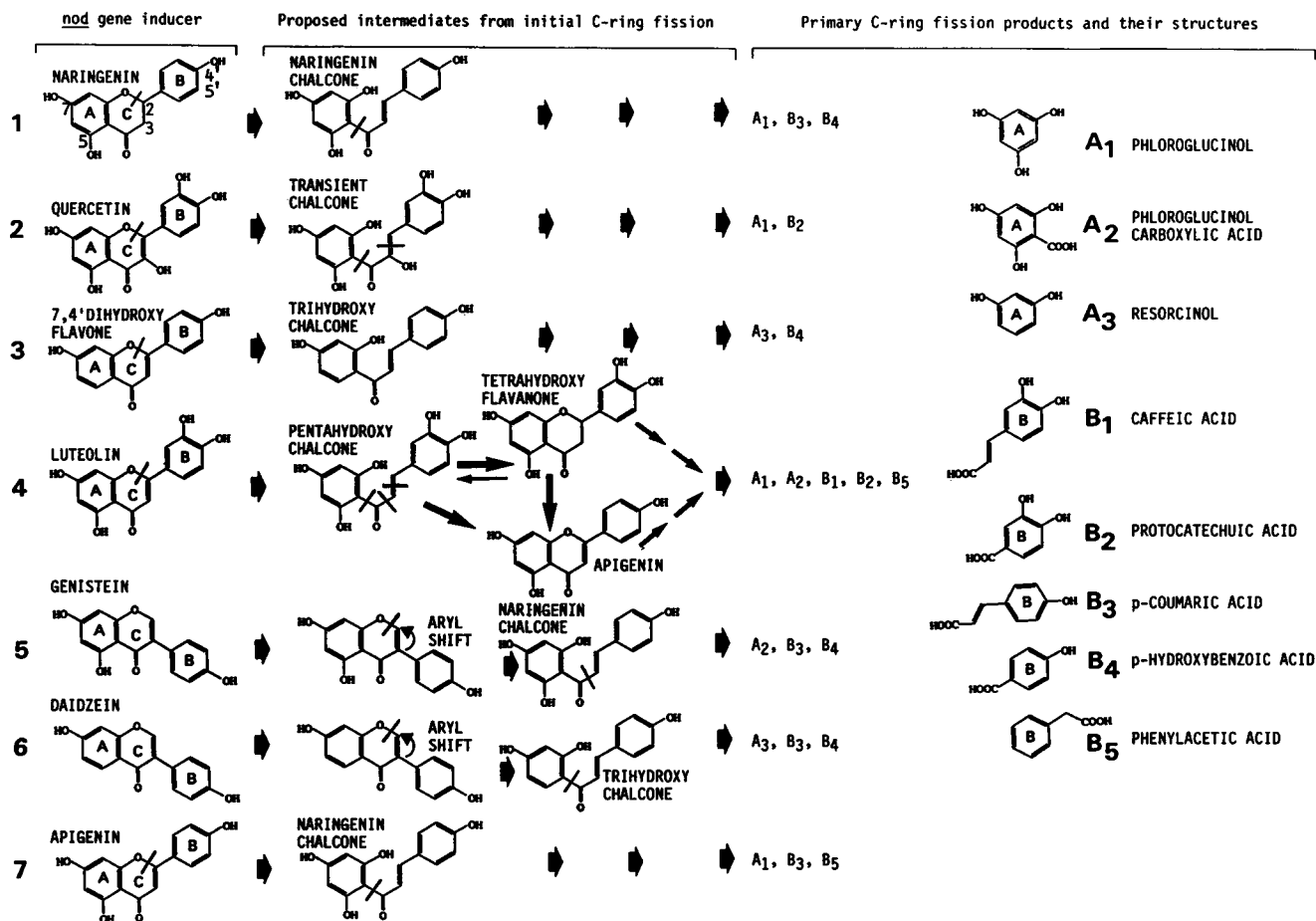


FIG. 2. Proposed degradation pathways for seven flavonoids incubated with rhizobia. Compounds: 1, *R. leguminosarum* bv. *viciae*; 2, *R. leguminosarum* bv. *phaseoli*; 3, *R. leguminosarum* bv. *trifolii*; 4, *R. meliloti*; 5, *R. fredii*; 6 and 7, *Rhizobium* sp. strain NGR234.

Degradation of this type could also provide an effective means of detoxifying certain phytoalexin-like molecules.

New flavanones and chalcones have also been observed in the root exudates of *Vicia sativa* subsp. *nigra* after inoculation with *R. leguminosarum* bv. *viciae* (21), and their origin was attributed to de novo biosynthesis in root tissue followed by rapid release into the rhizosphere (23). Our results suggest that the new compounds could arise from the degradation of naringenin in root exudate by free-living rhizobia. Although the new chalcones and flavanones formed by *R. leguminosarum* bv. *viciae* from naringenin in the present study could not be fully characterized, it was established that their GC retention times and the mass spectra of their derivatives were different from those of authentic naringenin and naringenin chalcone. In inoculated, naringenin-supplemented medium, a mixture of naringenin and naringenin chalcone was detected within 1 h of incubation. The addition of naringenin chalcone alone to a sterile basal medium resulted in cyclization of the C-ring and complete isomerization to naringenin after 2 h, thus confirming that no spontaneous isomerization in the reverse direction occurred under the test conditions and that the initial C-ring cleavage in naringenin was bacterially mediated.

When incubated with *R. meliloti*, luteolin yielded several other closed C-ring metabolites (e.g., a tetrahydroxy flavanone and apigenin), despite the concurrent existence of C-ring fission (Fig. 2). Speculation on additional roles for phenolic

compounds as signal molecules in legume-*Rhizobium* interactions has increased since the discovery that some flavonoids function as natural auxin transport regulators in plants by competing with naphthylphthalamic acid for binding to the naphthylphthalamic acid receptor in plant cells (13) and acting as auxin transport inhibitors. An extra dimension has been added to this concept following reports that auxin transport inhibitors induce nodule-like structures on legume roots (10, 12) and that *R. meliloti*, in both wild-type and pSym-cured forms, produces a luteolin-induced metabolite which competes with naphthylphthalamic acid for its binding site on plant membranes (11). The fact that this compound is produced by rhizobia which are devoid of *nod* genes suggests that it is unlikely to possess a structure of the type found in Nod factors and that it could be either a novel microbial metabolite or a product derived from luteolin degradation.

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## REFERENCES

1. Aguilar, J. M. M., A. M. Ashby, A. J. H. Richards, G. J. Loake, M. D. Watson, and C. H. Shaw. 1988. Chemotaxis of *Rhizobium leguminosarum* biovar *phaseoli* towards flavonoid inducers of the symbiotic nodulation genes. *J. Gen. Microbiol.* **134**:2741–2746.
2. Broughton, W. J., and M. J. Dilworth. 1971. Control of leghaemoglobin synthesis in snake beans. *Biochem. J.* **125**:1075–1080.
3. Chen, Y. P., A. R. Glenn, and M. J. Dilworth. 1984. Uptake and oxidation of aromatic substrates by *Rhizobium leguminosarum* MNF3841 and *Rhizobium trifolii* TA1. *FEMS Microbiol. Lett.* **21**:201–205.
4. Dakora, F. D., C. M. Joseph, and D. A. Phillips. 1993. Alfalfa root exudates contain isoflavonoids in the presence of *Rhizobium meliloti*. *Plant Physiol.* **101**:819–824.
5. Djordjevic, M. A., and J. J. Weinman. 1991. Factors determining host recognition in the clover-*Rhizobium* symbiosis. *Aust. J. Plant Physiol.* **18**:543–557.
6. Gajendiran, N., and A. Mahadevan. 1988. Utilization of catechin by *Rhizobium* sp. *Plant Soil* **108**:263–266.
7. Gottfert, M. 1993. Regulation and functions of rhizobial nodulation genes. *FEMS Microbiol. Rev.* **104**:39–64.
8. Hartwig, U. A., C. M. Joseph, and D. A. Phillips. 1991. Flavonoids released naturally from alfalfa seeds enhance growth rate of *Rhizobium meliloti*. *Plant Physiol.* **95**:797–803.
9. Hartwig, U. A., C. A. Maxwell, C. M. Joseph, and D. A. Phillips. 1990. Effects of alfalfa *nod* gene-inducing flavonoids on *nodABC* transcription in *Rhizobium meliloti* strains containing different *nodD* genes. *J. Bacteriol.* **172**:2769–2773.
10. Hirsch, A. M., T. V. Bhuvanawari, J. G. Torrey, and T. Bisseling. 1989. Early nodulin genes are induced in alfalfa root outgrowths elicited by auxin transport inhibitors. *Proc. Natl. Acad. Sci. USA* **86**:1244–1248.
11. Hirsch, A. M., A. S. Fang, H. I. McKhann, B. Niner, K. Wycoff, and M. Jacobs. 1992. Molecular interactions during nodule development, abstr. 21. 9th Int. Congr. Nitrogen Fixation 1992.
12. Hirsch, A. M., H. I. McKhann, and M. Löbner. 1992. Bacterial-induced changes in plant form and function. *Int. J. Plant Sci.* **153**:171–181.
13. Jacobs, M., and P. H. Rubery. 1988. Naturally occurring auxin transport regulators. *Science* **241**:346–349.
14. Krumholz, L. R., and M. P. Bryant. 1986. *Eubacterium oxidoreducens* sp. nov. requiring H<sub>2</sub> or formate to degrade gallate, pyrogallol, phloroglucinol and quercetin. *Arch. Microbiol.* **144**:8–14.
15. Maxwell, C. A., and D. A. Phillips. 1990. Concurrent synthesis and release of *nod* gene-inducing flavonoids from alfalfa roots. *Plant Physiol.* **93**:1552–1558.
16. Parke, D., and L. N. Ornston. 1984. Nutritional diversity of *Rhizobiaceae* revealed by auxanography. *J. Gen. Microbiol.* **130**:1743–1750.
17. Peters, N. K., J. N. Frost, and S. R. Long. 1986. A plant flavone, luteolin, induces expression of *Rhizobium meliloti* nodulation genes. *Science* **233**:977–980.
18. Peters, N. K., and D. P. S. Verma. 1990. Phenolic compounds as regulators of gene expression in plant-microbe interactions. *Mol. Plant-Microbe Interact.* **3**:4–8.
19. Phillips, D. A. 1992. Flavonoids: plant signals to soil microbes. *Annu. Rev. Phytochem.* **26**:201–231.
20. Rao, J. R., N. D. Sharma, J. T. G. Hamilton, D. R. Boyd, and J. E. Cooper. 1991. Biotransformation of the pentahydroxy flavone quercetin by *Rhizobium loti* and *Bradyrhizobium* strains (*Lotus*). *Appl. Environ. Microbiol.* **57**:1563–1565.
21. Recourt, K., J. Schripsema, J. W. Kijne, A. A. N. Van Brussel, and B. J. J. Lugtenberg. 1991. Inoculation of *Vicia sativa* subsp. *nigra* with *Rhizobium leguminosarum* biovar *viciae* results in release of *nod* gene activating flavanones and chalcones. *Plant Mol. Biol.* **16**:841–852.
22. Recourt, K., A. A. N. van Brussel, A. J. M. Driessen, and B. J. J. Lugtenberg. 1989. Accumulation of a *nod* gene inducer, the flavonoid naringenin, in the cytoplasmic membrane of *Rhizobium leguminosarum* biovar *viciae* is caused by the pH-dependent hydrophobicity of naringenin. *J. Bacteriol.* **171**:4370–4377.
23. Recourt, K., M. Verkerke, J. Schripsema, A. A. N. Van Brussel, B. J. J. Lugtenberg, and J. W. Kijne. 1992. Major flavonoids in uninoculated and inoculated roots of *Vicia sativa* subsp. *nigra* are four conjugates of the nodulation gene inhibitor kaempferol. *Plant Mol. Biol.* **18**:505–513.
24. Schlaman, H. R. M., H. P. Spaink, R. J. H. Okker, and B. J. J. Lugtenberg. 1989. Subcellular localization of the *nodD* gene product in *Rhizobium leguminosarum*. *J. Bacteriol.* **171**:4686–4693.
25. Winter, J., L. H. Moore, V. R. Dowell, Jr., and V. D. Bokkenheuser. 1989. C-ring cleavage of flavonoids by human intestinal bacteria. *Appl. Environ. Microbiol.* **55**:1203–1208.