# Risk-taking behavior in weight-compensating coho salmon, Oncorhynchus kisutch 

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#### Abstract

The effects of food restriction on predation risk-aking behavior were sudied in juvenile coho salmon, Oncorhynchus kisutch, during the period before seawater migration in the spring. A radiographic technique enabled the study of individual food intake in a safe and risky food patch before and after exposure to a piscivorous predator. The study revealed a significant increase in food intake and specific growth rate in the weeks following deprivation, resulting in compensatory growth and a recovery of the weight loss caused by dietary restriction. The increase in food intake resulted from a change in risk-aking behavior. Following a period of diet restriction, the fish habituated faster after predator exposure, and the proportion of fish in the risky patch was significantly higher than before deprivation. Deprived fish took greater shares of the group meal compared with control fish, and the pattern of the individual share of the group meal after food restriction indicated a change in the social hierarchy. This study indicates that risk-taking behavior is state dependent and changes temporarily to compensate for a period of food restriction. The results are discussed in terms of trade-offs between food and risk in a period leading up to an ontogenetic habitat shift. Key words: coho salmon, Oncorhynchus kisutch, predation, risk naking, Salmonidae, social hierarchy. [Behav Ecal 9:26-32 (1998)]


It has become increasingly clear that individuals are able to make behavioral trade-offs between food intake and risk of predation in a state-dependent fashion. Foraging at an increasing rate may be costly in terms of an increase in the risk of mortality due to predation (Godin and Smith, 1988). A hungry animal is more willing to take risks simply because an increase in the benefit of risk-taking behavior (food) results in an increase in the willingness to pay the potential cost (risk of injury or death). Such a compromise in a conflict situation has been demonstrated for several species, including mammals, birds, fishes, and invertebrates (reviewed by Lima and Dill, 1990). For example, prey attack distance declines with increasing satiation in coho salmon, Oncorhynchus kirutch (Dill and Fraser, 1984); parasite-infected three-spined sticklebacks, Gasterostrus aculeatus, are more willing to take risks compared with uninfected fish (Godin and Sproul, 1988); hunger level determines the time before Atlantic salmon, Salmo salar, resume feeding after exposure to a predator (Gotceitas and Godin, 1991); and hungry crucian carp (Carassius carassius) spend more time in an risky patch with food than in a safe patch without food (Pettersson and Brönmark, 1993).

Despite numerous descriptions of risk-taking behavior in the literature (Lima and Dill, 1990), the complex relationships between body size, food incake, and risk taking at an individual level are not completely understood. In nature, bigger prey fish are difficult for a piscivorous predator to catch and handle, confining the vulnerable prey to a size range dependent on the gape of the particular predators. Within this size range, relative vulnerability decreases as a function of prey body size, and beyond a certain size, depending on the size of the predators, the prey attain a preysize refuge (Damsgard, 1995; Hambright et al., 1991). Growing fish thus face a fun-

[^0]damental behavioral dilemma: a high growth rate will on one hand reduce the time a prey is vulnerable to a predator, but on the other hand a high food intake and growth rate can only be obtained through an increase in the risk of predation. The resulting size-dependent prey behavior can lead to complex patterns of habitat use and trophic interactions. For example, small, vulnerable bluegill sunfish, Lepowis macrochirus, shift their foraging behavior and reduce their use of open habitats in the presence of predators, while larger, nonvul nerable sunfish have a higher growth rate in the presence of predators than in their absence (Werner et al., 1983).

To examine the relationship between growth, food intake, and risk-raking behavior, we studied individually tagged coho salmon before their downstream spring migration to the sea Coho salmon in southern British Columbia (Canada) migrate to sea near the end of May (e.g., Fraser et al., 1983), normally after spending one winter in fresh water. However, some coho do not descend to the sea as yearlings, but spend 2,3, or even 4 years in fresh water before migration (Groot and Margolis, 1991). The frequency of older migrants increases in northern areas, but the size of coho smolts (i.e., seaward migrants) is fairly consistent throughout the species' geographic range (Groot and Margolis, 1991), indicating that age at seawater migration is related to growth in fresh water.

Smoltification involves a series of physiological, morphological, and behavioral changes that preadapts salmonids to a marine life (reviewed by Hoar, 1988). Many salmonid populations segregate into two subpopulations, displaying a bimodal growth pattern before smoltification (e.g., Thorpe, 1977). Individuals within Adlantic salmon populations that do not smolt enter a state of metabolic arrest and overwinter anorexia (sensu Mrosovsky and Sherry, 1980) and spend most of their time hiding in stony substrata (Metcalfe and Thorpe, 1992). A figh growth rate may thus be regarded as one result of the decision to become 2 smolt, and individual risk-taking behavior will depend on fish size, food availability, and the time remaining until migration. The high size-dependent mortality during seaward migration and the early seawater period in anadromous salmonids (Holtby et al., 1990) indicate
that body size at the time of migration, together with the occurrence of predators, will strongly affect fitness and thus select for animals reaching a minimum size before departure from the stream. In the present study, we tested the prediction that smoling coho salmon, deprived of food for a period of time, will be more willing to take risks in the presence of a predator, to compensate for the weight lost and increase their likelihood of reaching the threshold size by the time of migration.

## MATERIALS AND METHODS

Coho salmon (age 1+) from Inch Creek Hatchery, Fraser Vat ley, British Columbia, Canada, were acclimated to laboratory facilites at Simon Fraser University in December 1993. The fish were individually tagged (Fingerling Floy Tags), randomly divided into two groups of initially 100 fish each, and reared in $170-1$ holding taiks ( $10^{\circ} \mathrm{C}$ and a $12 \mathrm{~h}: 12 \mathrm{~h}$ lightdark regime). The fish were handfed dry pellets ( 2.4 mm ; MooreClark, Vancouver). Based on recommendations from MooreClark, the daily ration was calculated as 2.2 and $1.6 \%$ of total biomass, for the period when mean fish weight were smaller and larger than 12 g , respectively.

We conducted the study between week 5 (1 February) and week 16 (21 April) in 1994. During weeks 11-13 (11 March1 April), one of the groups was deprived of food by giving only $25 \%$ of the normal ration, while the other served as a control, receiving its full ration. Twice before dietary restriction (weeks 7 and 10) and twice after restriction (weeks 13 and 16), four replicate experiments were conducted in an observation tank. Each experiment included 22-25 fish from each group (depending upon the number of fish still alive), and thus totally approximately 50 fish. We randomly sampled the fish so each fish was tested once at every occasion, and groups were randomly reassembled for each testing period. In both weeks 7 and 10, one experiment was excluded due to aeration problems. The observation tank ( $270 \times 98 \mathrm{~cm}$ ) was divided with Plexiglas walls into three chambers. The central chamber ( $132 \times 98 \mathrm{~cm}$ ) served as the experimental arena for the prey fish. A mesh screen was located 5 cm above the bottom of the tank, and the water depth over the mesh was 25 cm, giving a total water volume of 320 L . One of the end chambers served as a predator comparment and had a clear Plexiglas window ( $30 \times 25 \mathrm{~cm}$ ) with an opaque sliding door behind it (operated with monofilament lines from outside the tank). The predator ( $235 \mathrm{~cm}, 0.5 \mathrm{~kg}$, rainbow trout, Oncorhynchus mykiss) was kept in an separate aquarium behind the door. To reduce the possibility that prey fish learned the position of the predator, the second end chamber was identical to the predator chamber.

We placed prey fish in the observation tank 1 day before the experiment and did not feed them; this ensured that all fish had equally empty stomachs at the time of testing. Throughout the experimental period, prey fish were fed with a belt feeder, providing the calculated amount of food at irregular intervals. The food was delivered at equal rates through Plexiglas tubes to two separate areas in the tank. The feeding stations were located $15-20 \mathrm{~cm}$ and $110-115 \mathrm{~cm}$ from the predator window, thus dividing the tank into 2 risky and a safe feeding patch of equal size. The bottom mesh prevented the fish from bottom feeding; thus, the fish had to stay at the feeding stations to be able to feed.
The experiment started at approximately 0900 h . We monitored the positions of the fish using a video camera mounted above the tank. After 1 h , the fish were visually exposed to the predator by removing the opaque partition between the chambers. The number of fish in the risky patch was recorded every 5 min from 45 min before to 90 min after exposure to
the predator (the predator remained visible throughout this $90-\mathrm{min}$ period). To control for the effects of partition movement, three identical experiments were conducted without the predator behind the Plexiglas window.

We examined food intake by individual coho using radiography (Jørgensen and Jobling, 1989; Talbot and Higgins, 1983). During the experiment, the normal food was replaced with pellets of identical composition but formulated to contain X-ray dense glass beads (Ballotini, Jencons Ltd., Leighton Buzzard, Bedfordshire, UK; size 7, 8.5, and 10). Three types of food with different sizes of glass beads were fed during the experiment; one before exposure and one in each patch after exposure to the predator.

After each experiment we anesthetized the coho using phenoxyethanol, weighed them to the nearest 0.1 g , and Xray photographed them (MicroVet MV 200 machine, 3.5 -s exposure, $63 \mathrm{kV}, 42 \mathrm{mAs}$, AGFA Structurix D7 film). Counts of the three different sizes of bead found in the stomachs enabled us to determine where, when, and how much each individual fish had eaten during the experiments. We calculated food intake of individual fish using the known relationship between numbers of glass beads and weight of food pellets. Weight-specific food intake was expressed as milligram dry weight of food per gram fresh weight of fish per hour (mg $d w \times \mathrm{gfw}^{-1} \times \mathrm{h}^{-1}$ ) after exposure to the predator. We classified individual fish as either nonfeeding fish, safe feeders (fish with more than $50 \%$ of their food from the safe patch), or risky feeders (fish with more than $50 \%$ from the risky patch) after the exposure to the predator. The individual share of a group meal (McCarthy et al., 1992) was calculated as food intake by single fish as a percentage of the total amount of food eaten in each experiment. The specific growth rates (SGR; \% body weight $\times$ day $^{-1}$ ) between two successive experiments were calculated as SGR $=100 \times(\ln$ W2 $-\ln$ W1) $/ t$, where W1 is the weight at time 1 , W2 is the weight at time 2, and $t$ is the time in days between the experiments (Jobling, 1994).

The state of the smoltification process during the experimental period was measured as the frequency of fish showing externally visible smolt characters, according to Gorbman et al. (1982). Before the experiment, $78.3 \%$ of the fish were classified as parr (with parr marks and without silvering), 21.7\% as silvery parr (with partial parr marks and silvering), and none of the fish as smolts (without parr marks and with full sitvering). During the course of the experiment, the frequency of parr decreased, and the frequency of smolts increased, and by week 16 only $1.1 \%$ of the fish were classified as parr, $65.1 \%$ were silvery parr, and $33.7 \%$ were smolts.

## RESULTS

There was no significant difference in weight, food intake or specific growth rates between the deprived group and the control group (Figure 1 ; week 10, $t$ tests, $p>.05$ ) before the food restriction that followed the experiment in week 10 . During the deprivation period between weeks 10 and 13, the mean weight of fish in the deprived group leveled off (Figure 1a). The specific growth rate was significantly lower ( $t$ test, $p<$ .001) in the deprived group ( mean $\pm$ SE, $0.33 \pm 0.04 \% \times$ $d^{-1} y^{-1}, n=89$ ) than in the control group ( $1.15 \pm 0.05 \% \times$ day $^{-1}, n=88$; Figure 1c). After the restriction period ended, food intake was significantly greater ( $t$ test, $p<.001$ ) in the deprived group ( $3.2 \pm 0.2 \mathrm{mg} \mathrm{dw} \times \mathrm{g} \mathrm{fw}^{-1} \times \mathrm{h}^{-1}, \boldsymbol{n}=90$ ) than in the control group ( $0.6 \pm 0.1 \mathrm{mg} \mathrm{dw} \times \mathrm{g} \mathrm{fw}^{-1} \times \mathrm{h}^{-1}$, $n=88$; Figure 1b). Accordingly, between weeks 13 and 16 the deprived group had a significantly higher ( $t$ test, $p<.001$ ) specific growth rate ( $1.64 \pm 0.04 \% \times$ day $^{-1}, n=88$ ) than the control group ( $1.25 \pm 0.03 \% \times$ day $^{-1}, n=86$, Figure 1c). As


Figure 1
Differences between experimentally food-deprived coho salmon and the control group without deprivation: (a) weight (g, $n=85-100$ ), (b) food intake (mg dw $\times \mathrm{g} \mathrm{fw}^{-1} \times \mathrm{h}^{-1} ; n=64-90$, and (c) specific growth rate ( $\% \times \mathrm{day}^{-1} ; n=84-91$ ). Weights and food intakes represent means and SE at the sampling day; growth rates represent means and SE for the period prior to the ampling dayThe deprivation period is indicated with a horizoncal bar in panel 2. ${ }^{*} p<.05,{ }^{* * *} p<.001$.
a result of this compensatory growth, the deprived group caught up with the control group, and there was no difference in mean fish weight by the end of the experiment ( $15.3 \pm 0.4$ $\mathrm{g}, n=88$ and $15.4 \pm 0.4 \mathrm{~g}, n=87$ in the deprived and control groups, respectively, Fugure 12).

Before predator exposure, the mean proportion of fish in the risky patch ranged from 48.4 to $64.8 \%$ (Table 1). In all experiments, the number of fish in the risky patch sharply decreased after the predator became visible and thereafter increased slowty. During this time, both solitary and group predator inspection was frequently observed. A typical inspection involved successive movements toward the predator, often ending with lateral displays in front of the Plexiglas window, followed by rapid movement away from the predator. Before food restriction (week 10 ), the mean proportion of fish in the risky patch decreased from 57.1 to $9.2 \%$ after predator exposure and increased to $37.7 \%$ 65-90 min after exposure to the predator (Table 1, Figure 2a). After the period of food restriction (week 13), the proportion of fish in the risky patch did not decrease to zero as it had in week 10 and increased to $56.5 \% 35-60 \mathrm{~min}$ after predator exposure (Table 1, Figure 2a). Four weeks before restriction (week 7), and 3 weeks after the restriction (week 16), the numbers of fish in the risky patch were similar to the results in week 10 (Table 1). There was no significant difference between the numbers of fish in the risky patch before predator exposure in weeks 10 and 13, whereas the differences $0-30 \mathrm{~min}, 35-60 \mathrm{~min}$, and $65-90 \mathrm{~min}$ after exposure were highly significant (Mann-Whitney, $p<$ $.001, p<.001$, and $p<.01$, respectively).

The result whs not due to disturbance caused by opening the partition between the prey and the predator. The fish did move away from the risky patch after the removal of the Plexigias window in the experiments without a predator, and the proportion of fish there decreased from $63.4 \%$ before to $18.8 \%$ after. However, the fish habituated sooner than in the predator experiments, and $35-60 \mathrm{~min}$ after removal of the partition $58.0 \%$ of fish were in the risky patch (Table 1, Figure 2b).

Most of the feeding fish in both the deprived and the control groups in all experiments ate in only one of the patches $(72.3 \pm 3.0 \%, n=8)$; the rest of the fish had food from both patches in their stomachs. The distribution of fish among the three feeding types did not differ between the deprived group and the control group before restriction (week 10; Figure 3a). More than $50 \%$ of the fish did not feed, while $15-25 \%$ fed in each of the safe and risky patches. From week 10 to week 13, the frequency of nonfeeding fish in the deprived group decreased significantly (Mann-Whitney, $p<.05$ ) from 54.3\% ( $n$ - 3) to $19.3 \%(n=4)$, whereas significantly more fish (MannWhitney, $p<.05$ ) fed in the risky patch after deprivation ( $49.1 \%, n=4$, versus $21.7 \%, n=3$ ). After the period of food

Table 1
Percentage of fish in riaky patch before (weeks 7 and 10) and after (weeks 13 and 16) a period of food deprivation, compared with an experiment without a predator

Mean $\pm$ SE percentage of fish in risky patch

|  | Before predator exposure | After predator exposure (min) |  |  |
| :---: | :---: | :---: | :---: | :---: |
|  |  | 0-30 | 35-60 | 65-90 |
| Week 7 | $64.8 \pm 3.3$ | $11.3 \pm 3.1$ | $54.2 \pm 5.2$ | $29.9 \pm 3.5$ |
| Week 10 | $57.1 \pm 3.5$ | $9.2 \pm 2.1$ | $18.4 \pm 4.8$ | $37.7 \pm 7.0$ |
| Week 13 | $62.1 \pm 2.3$ | $28.7 \pm 4.6$ | $56.5 \pm 3.3$ | $64.1 \pm 2.7$ |
| Week 16 | $48.4 \pm 4.0$ | $15.7 \pm 4.3$ | $21.5 \pm 4.6$ | $31.6 \pm 4.9$ |
| Without predator | $63.4 \pm 2.7$ | $18.8 \pm 4.4$ | $58.0 \pm 4.6$ | $53.1 \pm 4.6$ |

[^1]

Figure 2
Mean percentage of coho salmon in the risky patch (a) before (week 10, $n=3$ ) and after (week 13, $n=4$ ) a period of food deprivation. (b) Experiments without a predator present ( $n=3$ ).
restriction (week 13), the distribution of fish among patches changed dramatically in the deprived group, whereas only small changes occurred in the control group, and the frequency of nonfeeding fish, fish in the safe patch, the deprived group and the control group (Table 2, Figure 3b). Four weeks before restriction (week 7) and 3 weeks after restriction (week 16), the frequency of nonfeeding fish and the proportions of fish feeding in the safe and risky patches in both the deprived and the control groups were similar to the results in week 10 (Table 2).
Food restriction changed the social hierarchy, and a relatively greater number of deprived fish fed after the period of food restriction. Before restriction (week 10 ), a small number of fish monopolized the feeding patches and took a large share of the meals (Figure 4a). There was no difference between the number of fish that ate more than the mean share in the deprived group and the control group [ 6 (24\%) and 7 ( $28 \%$ ) of 25 fish, respectively]. After the food restriction


Figure 3
Feeding in food-deprived coho salmon and in a control group. Frequency of nonfeeding fish and fish feeding in safe and risky patches (a) Before deprivation (week $10, n=3$ ) and (b) after deprivation (week 13, $n=4$ ). The data represent means and SE * $p$ $<.05$
(week 13), the deprived group increased, while the control group decreased, their share of the food eaten (Figure 4b). In the deprived group, 15 of 25 fish ( $60 \%$ ) ate more than the mean share, compared with only 2 fish (8\%) in the control group.

## DISCUSSION

We have demonstrated experimentally that coho salmon are able to compensate for a period of low food availability with an increase in food intake after the deprivation period. In nature, variation in food availability causes many organisms to experience periods of low growth. As an adaptation to this variation, many species grow faster than normal after a period of undernutrition or famine. Such "compensatory growth" or "recovery growth" has been observed in invertebrates (Bradley et al., 1991), several species of mammals and birds (Ashworth, 1986; Thornton et al., 1979; Wilson and Osbourne,

Table 2
Mean $\pm$ SE frequency of nonfeeding fish and fish feeding in safe and risky patches after exposure to a predator before (weeks 7 and 10 , a : 3) and after (weeks 13 and 16, $n=4$ ) a period of food deprivation, compared with fish from the control group without depilvathon

|  | Deprived fish |  |  | Control fish |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Nonfeeding | Safe patch | Risky patch | Nonfeeding | Safe patch | Risky patch |
| Week 7 | $77.4 \pm 7.1$ | $14.4 \pm 3.6$ | $8.2 \pm 4.2$ | $75.8 \pm 5.2$ | $12.5 \pm 4.0$ | $11.7 \pm 2.4$ |
| Week 10 | $54.3 \pm 16.4$ | $24.0 \pm 12.2$ | $21.7 \pm 5.5$ | $61.2 \pm 10.7$ | $15.2 \pm 5.9$ | $23.6 \pm 5.0$ |
| Week 19 | $19.3 \pm 5.8$ | $31.7 \pm 4.2$ | $49.1 \pm 7.8$ | $64.5 \pm 9.2$ | $11.7 \pm 5.0$ | $23.8 \pm 5.5$ |
| Week 16 | $44.9 \pm 8.4$ | $33.4 \pm 10.4$ | $21.7 \pm 4.1$ | $79.5 \pm 4.3$ | $13.6 \pm 4.6$ | $6.9 \pm 3.1$ |



Figure 4
Individual share of meal as percentage of total food eaten by fooddeprived coho salmon and by individuals in the control group: (a) Before deprivation (week $10, n=3$ ), and (b) after deprivation (week 13, $n=4$ ). The dotted lines represent the mean share of the meal.
1960) and fish species, such as rainbow trout (Dobson and Holmes, 1984; Quinton and Blake, 1990 Weatherley and Gill, 1981), sockeye salmon, Oncorhynchus nerka (Bilton and Robins, 1973), Adantic salmon (Mortensen and Damsgård, 1993), and Arctic charr, Satvelinus alpinus (Miglavs and Jobling, 1989, Mortensen and Damsgard, 1993). According to these studies, the control of food intake and growth can be explained in terms of an individual "set-point" weight (Le Magnen, 1985; Mrosovsky and Sherry, 1980). Animals "remember" their nutritional history and meet all deviation from a predetermined growth trajectory by compensating as soon as food is avaitable.
Our study demonstrates that compensatory growth involves a change in the trade-off between food intake and risk of mortality due to predation, explaining an individual's increase in food intake in terms of an increase in risk taking. The willingness to risk exposure to an predator varies between species of fish (Abrahams and Healey, 1993; Magnhagen, 1988) and with fish size (Johnsson, 1993), life history, and sex (Abrahams and Dill, 1989; Gilliam and Fraser, 1987; Huntingford et al., 1988). The behavioral flexibility we have demonstrated indicates that individual fish may temporarily alter their behavior to become risk prone. All fish in the current study had the same gastric emptiness at the time of the experiments, and the regulatory mechanism must therefore represent a metabolic hunger rathor than a gastric one. These regulatory mechanisms allow fish to optimize their behavior during longterm shifts in food availability. In nature, such flexibility is selectively favored as compared with having a constant riskprone or risk-averse behavior (Lima and Dill, 1990).
The existence of a "shy-bold" continuum as a fundamental behavioral axis has been demonstrated in several species in-
cluding humans, fish, and invertebrates, having been measured, for example, as the response to novel objects (reviewed by Wilson et al., 1994). The evolutionary implications of boldness are still poorly understood, and the existence of "high risk-high gain" phenotypes probably involves complex physiological and hormonal regulation mechanisms in addition to a genetic component. Our findings suggest that an individual's position along the shy-bold continuum is state dependent. A norm of reaction to a predator must be related to environmental conditions and factors such as hunger, sex, maturation state, and other life-history characters.

The theory of ideal free distribution (IFD) describes the spatial distribution of competitors between patches with vary ing suitability (Abrahams, 1989; Fretwell, 1972; Fretwell and Lucas, 1970; Parker and Sutherland, 1986). Despite the fact that assumptions in IFD models may be violated, the theory has been useful in understanding the mechanisms undertying patch choice. In the current study, we provided an identical amount of food in each patch and, accordingly, the fish distributed approximately evenly in the two patches. We observed that most of the feeding fish only fed in one of the two patches, possibly due to travel costs between patches (Godin and Reenleyside, 1984), perceptual constraints (Abrahams, 1986), or the establishment of a feeding hierarchy in each patch. In many of our experiments, however, more fish preferred the risky patch before the exposure to the predator, indicating a possible side bias.

The effects of predation risk were measured as deviations from the fish distribution before predator introduction. These deviations resulted both from an initial decrease in numbers of fish in the risky patch and a time lag before the fish habituated to the predator and began using the patch again. Before food deprivation, the recovery time in our study was approximately $60-90 \mathrm{~min}$. In comparison, juvenile Atlantic salmon recovered during $\mathbf{2 h}$ after a brief exposure of a predatory fish (Metcalfe et al., 1987). The increasing numbers of fish in the risky patch after the period of food restriction may be explained in terms of a decrease in the time lag caused by predator inspections. The observed predator inspections were similar to those reported in other studies (e.g., Godin and Crossmann, 1994; Pitcher et al., 1986), but we were not able to evaluate whether inspecting coho had a higher food intake or spent more time in the risky patch.

Coho clearly are unequal competitors, and a small number of fish monopolized each feeding patch. Feeding hierarchies have been demonstrated in a wide range of fish species (Damsgard et al., 1997; Li and Brocksen, 1977; McCarthy et al., 1992; Metcalfe, 1986). Social hierarchies are size dependent (e.g. McCarthy et al., 1992) and dominant fish grow faster than subordinates (Abbott and Dill, 1989), but food acquisition will also be strongly affected by factors such as hunger and, according to our study, long-term energetic need. After food restriction, food-deprived fish took a greater share of meal, whereas nondeprived fish fed little or nothing. Both food-deprived and non-deprived fish were tested simultaneousty. Food intake in the two groups was thus not independent of each other, and the decrease in the nondeprived group may be explained as a change in the social interactions. The aggressive behavior required to assure success in intra specific competition may increase risk of predation (Martel and Dill, 1993); so the interaction between deprivation level, predation risk and food share is likely to be complex.

The externally visible smott characters of the fish ehangod during the course of the study, suggesting that they were likely to smoluify and migrate seaward approximately at the end of the study. Hypothesized risk-taking behavior in a period leading up to an ontogenetic habitat shift, such as migration from rivers to the sea, is illustrated in Figure 5. An individual animal


Figure 5
Graphical model of riak-aking behavior before an ontogenetic habitat shift ( $T_{0}$ ). The solid line represente a predetermined growth trajectory. A deviation from this trajectory will cause the animal to adopt a low-risk strategy or a high-risk strategy (shaded area), depending on the weight-deviation and the time unil $T_{0}$ The dotued line represents the limit below which the animal is no longer able to reach the minimum weight by $T_{0}$ and will adopt a low-risk strategy and exhibit temporary anorexia.
has a predetermined growth trajectory in order to reach 2 minimum size at the time of the habitat shift ( $T_{0}$ ). If its weight is above this trajectory, the individual adopts in a low-risk strategy, aking few risks in the presence of a predator. On the other hand, if the individual's weight is below the set-point weight for that point in time, the animal increases its food intake by adopting a high-risk behavioral strategy until the setpoint weight is reached. However, if the weight is forced well below the trajectory, the animal is no longer capable of reaching the minimum weight before $T_{0}$, and will adopt a low-risk strategy and wait for the next opportunity to migrate. This change in the preparation for seaward migration corresponds to the E2 developmental switch proposed by Mangel (1994), and the occurrence of a window of opportunity for this decision (approximately 1 March ), indicating the existence of " 2 point of no return." The change in risk-taking strategy in our study is in agreement with a model of seasonal foraging behavior, developed to study long-term energy requirements and fat storage in anorexic salmon (Bull et al., 1996): According to their model, fish in good condition will forage little or not at all in early winter, whereas fish in poor condition will maintain a high appetite, and the fish will respond differently to food deprivation at different times of the season, anticipating future energy requirements.
It is important to stress that compensatory trade-offs between food and risk may not be a general mechanism in fish. As pointed out by Pettersson and Brönmark (1993), many ectothermic animals are normally not in danger of starvation in a period of low food availability. A low growth rate does thus not necessarily mean that risk-caking must be increased, particularly if conditions are expected to improve in the future. Accordingly, both Allantic salmon that would become smolts the following spring and fish that would require another year to reach the smolt stage adopted a low-risk strategy in December (Huntingford et al., 1988). Our study concerns how coho respond in the spring before an ontogenetic habitat shift, and we expect that such behavioral decisions may change during the life span of a fish. Although ectothermic animals may be risk prone during the warm growing season and risk averse the rest of the year, it is likely that endotherms will behave in an opposite fashion, taking the greatest risks during the cold season, simply because they, in contrast to fish, face a greater chance of famine and death at that time. However, a study of age-0 largernouth bass, Micropterus salmoides, indicated that a
failure to reach a minimum level of energy reserves before winter may be fatal to the fish (Miranda and Hubbard, 1994), suggesting that the annual changes in risk-taking behavior may be complex.

In summary, our study indicates that compensatory growth in coho salmon involves a temporary state-dependent change in the nature of the trade-off between foraging and predation risk. The fish compensated for a period of low food availability with a period of high risk taking, enabling them to return to a predetermined growth trajectory.

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## REFERENCES

Abbot JC, Dill LM, 1989. The relative growth of dominant and subordinate juvenile steelhead trout (Salmo gairdnoni) fed equal re tions. Behaviour 108:104-113.
Abrahams MV, 1986. Patch choice under percepoul constrainte: a cause for departures from an ideal free distribution. Behav Ecol Sociobiol 19:409-415.
Abrahams MV, 1989. Foraging guppies and the ideal free distribution: the influence of information on patch choice. Ethology 82:116126.

Abrahams MV. Dill LM, 1989. A determination of the energetic equivalence of the risk of predation. Ecology 70:999-1007.
Abrahams MV, Healey MC, 1993. A comparison of the willingness of four species of Pacific salmon to risk exposure to a predator. Oikos 66:439-446.
Ashworth A, 1986. Catch-up growth in children. Nutr Rev 44:157-163.
Bilton HT, Robins CL, 1973. The effects of starvation and subsequent feeding on survival and growth of Fulton channel sockeye salmon fry (Oncorhynchus nerka). J Fish Res Board Can 30.1-5.
Bradley MC, Perrin N, Calow P, 1991. Energy allocation in the cladoceran Daphnia magna Suraus, under starvation and refeeding. Oecologia 86:414-418.
Bull CD, Metcalfe NB, Mangel M, 1996. Seasonal matching of foraging to anticipated energy requirement in anorexic juvenile salmon. Proc R Soc Lond B 263:13-18.
Damsgărd B, 1995. Arctic charr, Satuelinus alpinus (L), as prey for piscivorous fish - 2 model to predict prey vulnerabilities and prey size refuges. Nordic J Freahw Res 71:190-196.
Damsgärd B, Arnesen AM, Baardvik BM, Jobling M, 1997. State-dependent feed acquisition among two strains of hatchery-reared Aretic charr. J Fish Biol 50:859-869.
Dill LM, Fraser AHG, 1984. Risk of predation and the feeding behar ior of juvenile coho salmon (Oncorhynchus kisulch). Behav Ecol Sociobiol 16:65-71.
Dobson SH, Holmes RM, 1984. Compensatory growth in rainbow trout, Salmo grairdneri Richardson. J Fish Biol 25:649-656.
Fraser FJ, Perry EA, Lightly DT, 1983. Big Qualicum River salmon development project Can Tech Rep Fish Aquat Sci 1189:1-198.
Fretwell SD, 1972. Populations in a seasonal environment. Princeton, New Jersey. Princeton University Press.
Fretwell SD, Lucas HL, 1970. On territorial behaviour and other factors influencing habitat distribution in birds. Acta Biotheor 19:1636.

Gilliam JF, Fraser DF, 1987. Habitat selection under predation hazard: test of a model with foraging minnows. Ecology 68:1856-1862.
Godin J-GJ, Crussman SL, 1994. Hunger-dependent predator inspection and foraging behaviours in the threespine stickleback (Caster-
astrus aculeatus) under predation risk Behav Ecol Sociobiol 34: 359-366.
Codin J-GJ, Keenleyside MHA, 1984. Foraging on patchily distributed prey by a cichlid fish (Teleostei, Cichlidac): a test of the ideal free distribution theory. Anim Behav 32:120-131.
Godin J-GJ, Smith SA, 1988. A fimess cost of foraging in the guppy. Nature 333:69-71.
Godin J-GJ, Sproul CD, 1988. Risk taking in parasitized stictlebacks under threat of predation: effects of energetic need and food avail ability. Can J Zool 66:2360-2367.
Corbman A, Dickhoff WW, Mighell JL, Prentice EF, Waknita FW, 1982. Morphological indices of developmental progress in the parremolt coho almon, Oncorhynchus kisutch. Aquaculure 28:1-19.
Gotceitas V, Codin J-GJ, 1991. Foraging under the riak of predation in Juvenile Aulantic salmon (Sabmo salar L.): effects of socinl status and hunger. Behav Ecol Sociobiod 29:255-261.
Groot C, Margolis L, 1991. Pacific salmon life histories. Vencouver: University of British Columbia Press.
Hambright KD, Drenner RW, McComas SR, Hairston NG, 1991. Capelimited piscivores, planktivore size refuges, and the trophic cascade hypothesis. Arch Hydrobiol 121:389-404.
Hoar WS, 1988. The phyiology of smolting-salmonide In: Fish phyt iology, vol. 11B (Hoar WS, Randall DJ, eds). New York: Academic Press; 275-343.
Holtby LB, Andersen BC, Kadowald RK, 1990. Importance of amolt size and earty ocean growth to interannual variability in marine survival of coho silmon (Oncortynchus gorbuscha). Can J Fish Aquat Sci 47:2181-2194.
Huntingford FA, Metcalfe NB, Thorpe JE, 1988. Choice of feeding station in Allantic salmon, Satmo salar, parr. effects of predation risk, season and life history strategy. J Fish Biol 33:917-924.
Jobling M, 1994. Fish bioenergetica. London: Chapman and Hall.
Johnsson JI, 1993. Big and brave: size selection affects foraging under risk of predation in juvenile rainbow trout, Oncontynches mykiss. Anim Behav 45:1219-1225.
Jergensen E, Jobling M. 1989. Patterns of food intake in Arctic charr, Satudinus alpinus, monitored by radiography. Aquaculure 81:155160.

Le Magnen J, 1985. Hunger. Cambridge: Cambridge University Presa.
Li HW, Brocksen RW, 1977. Approaches to the analyxis of energetic costs of intraspecific comperition for space by rainbow trout (Satmo gairdren). J Fish Biol 11:329-941.
Lima SL, Dill LM, 1990. Behavioural decisions made under the risk of predation. Can J Zood 68:619-640.
Magnhagen C, 1988. Predation risk and foraging in juvenile pink (Oncorhynchus gorbuscha) and chum salmon (O. heta). Can J Fish Aquat Sci 45:592-596.
Mangel M, 1994. Climate change and life history variation. Deep-rea Res II 41:75-106.
Martel G, Dill LM, 1993. Feeding and aggressive behaviours in juvenile coho salmon (Oncorkynchus kisutch) under chernically-mediated risk of predation. Behav Ecol Sociobiol 32:365-370.

McCarthy ID, Carter CG, Houlihan DF, 1992. The effect of feeding hierarchy on individual variability in daily feeding of rainbow trout, Oncorkynchus mektiss (Walbaum). J Fish Biol 41:257-263.
Metcalfe NB, 1986. Intraspecific variation in comperivive ability and food intake in salmonids: consequences for energy budgets and growth rates. J Fish Biol 28:525-531.
Metcalfe NB, Huntingford FA, Thorpe JE, 1987. The influence of predation risk on the feeding motivation and foraging strategy of juvenile Atantic salmon. Anim Behav 35:901-911.
Metcalfe NB, Thorpe JE, 1992 . Anorexia and defended energy in overwintering juvenile salmon. J Anim Ecol 61:175-181.
Miglave I, Jobling M, 1989. Effects of feeding regime on food consumption, growth rates and tissue nucleic acids in juvenile Arctic charr, Sahatimus alpinus, with particular respect to compensatory growth. J Fish Biol 34:947-957.
Miranda LF, Hubbard WD, 1994. Length-dependent winter survival and lipid composition of age-0 largemouth bass in Bay Springa Res ervoir, Missisalppi. Trans Am Fish Soc 123:80-87.
Mortensen A, Damsgird B, 1993. Compensatory growth and weight segregation following light and temperature maniputaion of juvenile Adantic salmon (Satmo salar L) and Arcic charr (Sabuetinus atpinus L). Aquaculture 114:261-272.
Mrosoviky N, Sherry DF, 1980. Animal anorexias. Science 207:837842.

Parker GA. Sutherkand WJ, 1986. Ideal free distributions when individuals differ in competidive ability: phenotype-limited ideal free models. Anim Behav 94:1222-1242.
Pettersoon LB, Bronmart C, 1993. Trading off safety against food: state dependent habitat choice and foraging in crucian carp. Oecologia 95:353-357.
Pitcher TJ, Green DA. Magurran AE, 1986. Dicing with death: predator inspection behaviour in minnow ahoals. J Fish Biol 28:439-448.
Quinton JC, Blake RW, 1990. The effect of feed cyeling and ration level on the compensatory growth response in rainbow trout, Orcorkynchus mokiss J Fish Biol 37:33-41.
Talbot C. Higgins PJ, 1983. A radiographic method for feeding studies on fish using metallic iron powder as a marker. J Fish Biol 23:211220.

Thornton RF, Hood RL, Jones PN, Re VM, 1979. Compensatory growth in sheep. Aus J Agric Res 30:135-151.
Thorpe JE, 1977. Bimodal distribution of length of juvenile Adantic salmon (Salmo salar L_). J Fish Biol 11:175-184.
Weatherley AH, Gill HS, 1981. Recovery growth following periods of restricted rations and starvation in rainbow trout Salmo gairdmeri Richardson. J Fish Biol 18:195-208.
Werner EE, Gilliam JF, Hall DJ, Mittelbach GG, 1983. An experimencal test of the effects of predation risk on habitat use in fish. Ecology 64:1540-1548.
Wilson DS, Clark AB, Coleman K, Dearstyne T, 1994. Shyness and boldness in humans and other animals. Trend Ecol Evol 11:442446.

Wilson PN, Osbourne DF, 1960. Compensatory growth after undernutrition in mammals and birds. Biol Rev 37:324-969.


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[^1]:    Data represent means $\pm$ SE every 5 min from 45 min before predator exposure and during three $\mathbf{3 0}$-min intervals after predator exposure.

