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Ichiro Shiojima and Kenneth Walsh Circ. Res. 2002;90;1243-1250 DOI: 10.1161/01.RES.0000022200.71892.9F

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Review

Role of Akt Signaling in Vascular Homeostasis and Angiogenesis

Ichiro Shiojima, Kenneth Walsh

Abstract—Akt is a serine/threonine protein kinase that is activated by a number of growth factors and cytokines in a phosphatidylinositol-3 kinase—dependent manner. Although antiapoptotic activity of Akt is well known, it also regulates other aspects of cellular functions, including migration, glucose metabolism, and protein synthesis. In this review, Akt signaling in endothelial cells and its critical roles in the regulation of vascular homeostasis and angiogenesis will be discussed. (Circ Res. 2002;90:1243-1250.)

Key Words: Akt ■ endothelial cells ■ angiogenesis ■ statins ■ endothelial nitric oxide synthase

S ince the identification of several classes of receptor tyrosine kinases and their ligands as crucial mediators of vascular development, considerable progress has been made toward understanding the process of angiogenesis at sites of tissue growth and/or repair.^{1,2} A number of clinical trials are currently evaluating angiogenic ligands for their ability to induce neovascularization in ischemic tissues,^{3,4} and the intracellular signaling pathways that mediate the proangiogenic effects of these growth factors are being extensively investigated. This review specifically focuses on the role of phosphatidylinositol-3 kinase (PI3K)–Akt signaling axis in endothelial cells because it is activated by many angiogenic growth factors and it regulates downstream target molecules that are potentially involved in blood vessel growth and homeostasis.

PI3K-Akt Signaling Axis: Upstream Activators and Downstream Targets

Akt was originally identified as a cellular counterpart of the oncogene derived from murine AKT8 retrovirus.^{5,6} The same gene product was independently isolated as a protein kinase related to protein kinase A and C and was therefore named as protein kinase B (PKB) or RAC (related to protein kinase A and C).^{7,8} Mammalian genomes contain three Akt genes, Akt1/PKBα, Akt2/PKBβ, and Akt3/PKBγ, whereas Drosophila melanogaster and Caenorhabditis elegans contain one and two Akt genes, respectively.^{9,10} These genes encode proteins containing a pleckstrin homology (PH) domain in the amino terminus, a central kinase domain, and a carboxy terminal regulatory domain. All 3 mammalian Akt genes are widely expressed in various tissues but Akt1 is most abundant in brain, heart, and lung, whereas Akt2 is predominantly expressed in skeletal muscle and embryonic brown fat, and Akt3 is predominantly expressed in brain, kidney, and embryonic heart.7,11-13 In unstimulated cells, Akt protein exists in cytoplasm and the two regulatory phosphorylation sites at threonine at 308 and serine at 473 are in an unphosphorylated state. On growth factor stimulation, the PH domain binds to the lipid products of PI3K, and Akt is recruited to plasma membrane. Akt is then sequentially phosphorylated at T308 and S473 by upstream kinases referred to as 3-phosphoinositide-dependent protein kinase 1 (PDK1) and PDK2, respectively, to yield a fully activated kinase (Figure 1).14,15 PDK1 has been isolated and characterized,15 but the identity of PDK2 is still controversial. Several candidate molecules have been suggested to be a potential S473-kinase including integrin-linked kinase (ILK), MAP kinase-activated protein kinase 2 (MK2), PDK1 (conversion of substrate specificity in association with protein kinase C-related kinase-2 [PRK2]) and Akt itself (autophosphorylation). 16-19 Fully activated Akt becomes available to phosphorylate its downstream substrates and a portion of these molecules detach from the plasma membrane and translocate to various subcellular locations including nucleus.20 Akt is then dephosphorylated and inactivated by protein phosphatases such as protein phosphatase 2A (PP2A).21

Akt is a critical regulator of PI3K-mediated cell survival.22,23 A large number of studies have demonstrated in various cell types that constitutive activation of Akt signaling is sufficient to block cell death induced by a variety of apoptotic stimuli and that transduction of dominant-negative Akt inhibits growth factor-induced cell survival.²⁴⁻²⁶ The prosurvival function of Akt has also been demonstrated in the context of the intact organism. Mutation of Drosophila Akt leads to embryonic lethality due to massive apoptosis during embryogenesis,27 and Akt1 mutant mice exhibit increased spontaneous apoptosis in testis and thymus.²⁸ Several downstream targets of Akt are recognized to be apoptosisregulatory molecules including Bad, FKHR family of forkhead transcription factors, and IKK α , ^{29–35} and these findings are consistent with the notion that Akt functions as a survival kinase. However, other downstream effectors of Akt are

Original received March 27, 2002; revision received April 30, 2002; accepted May 6, 2002.

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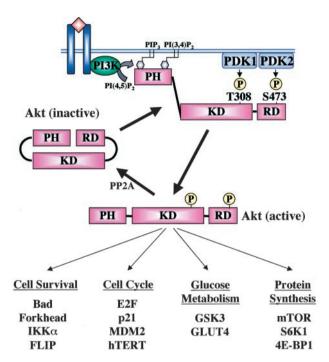


Figure 1. Mechanism of Akt activation and partial list of downstream molecules. Akt is activated by growth factors or cytokines in a PI3K-dependent manner, and phosphorylation of two residues by PDK1 (T308) and PDK2 (S473) is required for its full activation. Downstream target molecules are grouped according to their function. Note that these downstream molecules include both direct Akt substrates and indirect downstream effectors.

involved in different aspects of cellular regulation. For example, (1) Akt enhances glucose uptake by inducing membrane translocation of the glucose transporter GLUT4,36 (2) Akt promotes glycogen synthesis through the phosphorylation and inactivation of glycogen synthase kinase-3 (GSK-3),37 (3) Akt regulates cell cycle and cellular senescence, at least in part, through modulating the activities of E2F, p21, MDM2, and human telomerase reverse transcriptase subunit (hTERT),³⁸⁻⁴⁴ and (4) Akt enhances protein synthesis through increasing the phosphorylation of mammalian target of rapamycin (mTOR), eukaryotic initiation factor 4Ebinding protein 1 (4E-BP1), and 70-kDa S6 kinase (S6K1),^{15,45} although S6K1 may also be directly activated by PDK1 in a PI3K-dependent and Akt-independent fashion (Figure 1).46,47 Taken together, it is more appropriate to classify Akt as a multifunctional protein kinase rather than a simple regulator of cell survival.

Akt-Dependent Survival Signals in Endothelial Cells

Although originally identified as a factor that induces vascular permeability, vascular endothelial growth factor (VEGF) exhibits multiple biological activities in endothelial cells, including the enhancement of endothelial cell survival.⁴⁸ VEGF effects on cell survival have been shown to be mediated by Flk1/VEGFR2-PI3K-Akt pathway.^{49,50} Subsequently, it was also shown that several other endothelial cell stimuli including angiopoietin-1 (Ang-1),^{51,52} insulin,⁵³ insulin-like growth factor-I (IGF-I),⁵⁴ sphingosine-1-

phosphate (S1P),^{55,56} hepatocyte growth factor,⁵⁷ the small proteoglycan decorin,⁵⁸ fluid shear stress,⁵⁹ estrogen,^{59a} reactive oxygen species,^{59b} and corticosteroids^{59c} also activate PI3K-Akt signaling, illustrating the central role of this pathway in controlling endothelial cell viability.

Growth factor activation of angiogenesis is dependent on proper endothelial cell-extracellular matrix attachment, 60 and in the absence of matrix attachment, cells undergo apoptotic cell death through a process termed anoikis (a Greek word for "homelessness").61 VEGF activation of Akt in endothelial cells is dependent on matrix attachment, and constitutively active Akt blocks cell detachment-induced apoptosis.50 These findings suggest that matrix attachment is required for growth factors to activate Akt and maintain endothelial cell viability. Cell attachment is mediated mainly through the engagement of extracellular matrix with integrin molecules. When integrins bind to extracellular matrix they become clustered and associate with the actin cytoskeleton through adaptor/signaling molecules, which further promotes integrin clustering and the assembly of actin filaments and leads to the formation of focal adhesion and activation of intracellular signaling.⁶² The α_v integrin combinations have been most extensively investigated in terms of their roles in angiogenesis.63 Endothelial cells stimulated with angiogenic growth factors or those in newly formed vessels express high levels of $\alpha_{\nu}\beta_{3}$ integrin, and antagonists against $\alpha_{\nu}\beta_{3}$ or $\alpha_{\nu}\beta_{5}$ integrin block the growth factor-induced angiogenesis. It has also been shown that $\alpha_v \beta_3$ integrin associates with VEGF and platelet-derived growth factor (PDGF) receptors and potentiates VEGF or PDGF signaling, respectively.63 Because several integrin signaling molecules including focal adhesion kinase (FAK), ILK, and Shc have been implicated in Akt activation,61 downregulation of Akt activity induced by cell detachment is likely due to the decrease in integrin-dependent Akt activation. Caspase-mediated cleavage of Akt is also implicated in the downregulation of Akt protein level during long-term suspension culture.⁶⁴ Collectively, these findings suggest that integrin signaling induced by cell attachment (outside-in signal) is an important regulator of growth factor dependent endothelial cell survival and angiogenesis through PI3K-Akt pathways. Furthermore, VEGF-induction of insideout signals has also been shown to activate integrins,65 suggesting that integrin and growth factor signaling are cooperative and synergistic with regard to activation of Akt signaling (Figure 2).

Currently, relatively little is known about the downstream mediators of Akt-dependent survival pathway in endothelial cells, although several candidate molecules has been identified including survivin,⁵¹ FLICE-inhibitory protein (FLIP),⁶⁶ and MEKK3.⁶⁷ Thus, possible combinations of these and other unidentified Akt target molecules may control endothelial cell survival depending on the context of pro- and antiapoptotic stimuli encountered in the cellular environment.

Regulation of Endothelial Nitric Oxide Synthase (eNOS) Activity by Akt

In addition to its antiapoptotic effects, VEGF induces hypotension in the intact organism, nitric oxide (NO)-dependent vasodilation in isolated coronary arteries, and NO release in

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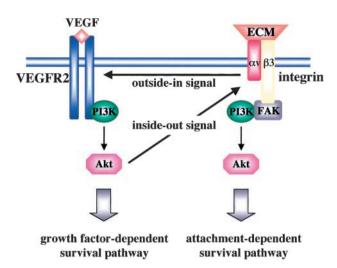


Figure 2. Growth factor- and cell attachment-dependent survival is mediated by PI3K/Akt signaling in endothelial cells. Integrin-dependent signals are prerequisite for growth factormediated activation of Akt. Akt-mediated crosstalk between these two signaling systems synergistically promotes endothelial cell survival.

isolated vessels and in cultured endothelial cells.68-70 Early studies demonstrated that VEGF-induced increase in NO release from endothelial cells is attenuated by PI3K inhibitors,71 and subsequently, it was demonstrated that VEGF stimulates Akt-mediated eNOS phosphorylation at Ser1177 (in human eNOS, equivalent to Ser1179 in bovine eNOS), leading to an increase in eNOS activity. 72,73 It is also reported that production of NO in response to fluid shear stress in cultured endothelial cells is controlled by Akt-dependent phosphorylation of eNOS,73 although another study has shown that shear stress induces eNOS phosphorylation predominantly through a protein kinase A-dependent, Aktindependent mechanism.74 Studies in intact animals have shown that overexpression of constitutively active Akt in the vascular endothelium increases resting diameter and blood flow, whereas transduction of dominant-negative Akt attenuates endothelium-dependent vasodilation induced by acetylcholine,75,75a demonstrating that Akt functions as a regulator of vasomotor tone in vivo. PI3K/Akt signaling has also been implicated in the control of endothelium-dependent vasorelaxation induced by adrenomedullin,76 and hyperglycemia has been shown to lead to the glycosylation of the Akt phosphorylation site in eNOS, resulting in an inhibition of eNOS activity.77

The activity of eNOS is also regulated by subcellular localization and/or protein-protein interactions. Of note, eNOS has been shown to be localized in a specific domain of plasma membrane called caveolae and to interact with caveolin-1 through caveolin-1 scaffolding domain, which inhibits eNOS activity.78-81 Although originally implicated in transmembrane trafficking of macromolecules, the finding that caveolae contain a variety of signaling molecules and caveolin-1 directly interacts with those caveolae-associated proteins have suggested that caveolae and caveolin-1 are involved in the compartmentalization and integration of signal transduction pathways at the cell membrane. Consis-

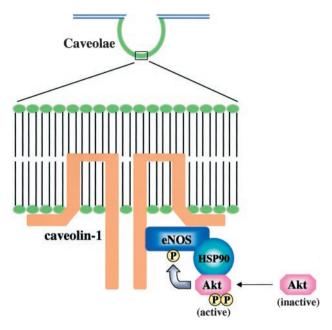


Figure 3. Schematic illustration of the Akt-eNOS interaction at caveolae. Caveolin-1 is localized to caveolae and associates with a number of regulatory molecules including eNOS. Association of eNOS with caveolin-1 negatively regulates eNOS activity, although targeting of eNOS to caveolae is required for proper eNOS function. Activated Akt and eNOS also associate with Hsp90. Hsp90 is believed to function as a scaffold protein for activation of eNOS by Akt-mediated phosphorylation.

tent with the inhibitory role of caveolin-1 on eNOS activity, administration of caveolin-1 scaffolding domain fused to cellular internalization sequences in vivo attenuates eNOS activity,82 and acetylcholine-induced vasorelaxation and NO production are enhanced in caveolin-1-deficient mice.83 The targeting of eNOS to caveolae, however, seems to be required for efficient and proper activation of eNOS on stimulation, because conditions that inhibit the localization of eNOS in caveolae also attenuate eNOS activity.84,85 It has also been shown that eNOS interacts with heat shock protein 90 (Hsp90) on stimulation with VEGF or shear stress, and this interaction enhances eNOS activity.86 Interestingly, Akt also interacts with Hsp90 on stimulation and this interaction enhances Akt enzymatic activity,87 suggesting that Hsp90 may serve as a scaffold protein for the efficient phosphorylation of eNOS by Akt at caveolae (Figure 3).88,89,89a

Regulation of Endothelial Cell Migration by Akt

The ability of endothelial cells to migrate and form capillarylike structures is essential for angiogenesis in vivo.1 VEGF enhances endothelial cell migration and capillary-like structure formation in vitro and these activities of VEGF are PI3K-Akt-dependent.90-92 S1P has also been shown to enhance endothelial cell migration and capillary formation in vitro through the activation of the endothelial differentiation gene (EDG) family of G protein-coupled receptors and PI3K-Akt-dependent pathways. 56,93,94 Conversely, oxidized LDL inhibits endothelial cell migration toward VEGF by promoting the dephosphorylation of Akt.95

Studies in other cell types have also implicated PI3K and Akt in the control of directional cell migration and the sensing of chemoattractant gradients by the cell. It has been shown that Akt transiently localizes to the leading edge membrane of migratory cells in a PI3K-dependent manner,96,97 and gene ablation studies in mice have demonstrated that PI3Ky is required for chemotaxis and chemoattractantdependent activation of Akt in macrophages and neutrophils. 98-100 Akt has been shown to be required for chemotaxis in Dictyostelium cells as well.96

Cellular movement requires the reorganization of actin cytoskeleton and distinct patterns of actin reorganization are required as cells establish leading edge and then generate contractile force to migrate forward. 101 Previous studies have implicated the Rho family of small G proteins as one of the major regulators of actin reorganization. Among Rho family members, Rho, Rac, and Cdc42 are most widely studied and each regulates specific aspects of cytoskeletal reorganization. Rho stimulates cytoplasmic stress fiber formation and actomyosin-based contractility, Rac induces membrane ruffling and extension of lamellipodia, and Cdc42 induces the extension of membrane protrusions (filopodia) and is also involved in chemoattractant gradient sensing. 102,103 In endothelial cells, it has been shown that VEGF-induced cell migration is dependent on Rho family GTPases. 104,105 However, the relationship between Akt and Rho family of G proteins is complicated and controversial. On one hand, Akt was shown to negatively regulate Rac1 activity by phosphorvlating Rac1 and inhibiting its GTP-binding activity. 106 In contrast, a recent study has demonstrated that Akt phosphorylates S1P receptor EDG-1 and induces Rac activation and cell migration in endothelial cells.¹⁰⁷ Other reports show that Rac and Cdc42 are situated upstream of Akt and that they promote Akt signaling. 108-110 Consistent with these findings. Akt has been shown to be required for cell motility induced by Rac or Cdc42 in fibroblasts.108

Another possible downstream effector of Akt that regulates cell motility is p21-activated protein kinase (PAK). PAK was originally identified as a Rac1-binding protein that specifically interacts with GTP-bound form of Rac.¹¹¹ Subsequently, it was shown that PAK is activated by Rac or Cdc42 and that it regulates polarized cytoskeletal reorganization.111 Recently it was shown in Dictyostelium cells that Akt regulates cell polarity and chemotaxis through the regulatory phosphorylation of PAK,112 suggesting a direct functional link between Akt and PAK in the regulation of cytoskeletal reorganization. In mammalian fibroblasts, it was also shown that Akt stimulates PAK1 activation and dominant-negative Akt inhibits Ras-induced activation of PAK1.113 However, the Akt phosphorylation site in Dictyostelium PAK is not conserved in mammalian PAK1, suggesting an indirect activation of mammalian PAK1 by Akt. Nonetheless, PAK family of protein kinases are attractive candidates for Akt effectors in the regulation of endothelial cell migration, and may be a convergence point of signals from Rac/Cdc42 and Akt.

Statins and Akt Signaling

The 3-hydroxyl-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors, or statins, are widely prescribed for the treatment of hypercholesterolemia, and several clinical trials have demonstrated that statins are effective for both primary and secondary prevention of coronary artery diseases.¹¹⁴ It has also been shown that statins rapidly improve vasomotor responses of atherosclerotic coronary arteries both in humans and in animal models,115-117 and studies in normocholesterolemic animals revealed that statins protect against stroke and myocardial ischemia/reperfusion injury possibly through NOdependent mechanisms. 118,119 These findings are consistent with the notion that cardioprotective effects of statins are partly independent of their serum lipid-lowering effects and may be due to the activation of eNOS in vascular endothelium.

Recent research has revealed a link between statins and Akt. Statins have been shown to rapidly promote the activation of Akt in endothelial cells leading to eNOS phosphorylation and increased NO production.91 Low statin concentrations have been shown to protect endothelial cells from serum deprivation-induced apoptosis and promote capillary-like structure formation on matrigel in an Akt-dependent manner, whereas higher concentrations are toxic.91 Consistent with their Akt-activating function, treatment with clinicallyrelevant doses of statins enhances angiogenesis in the ischemic hindlimbs of normocholesterolemic animals through an eNOS-dependent mechanism.91,120 It has also been shown that the activation of angiogenesis by statins is biphasic: low doses promote vessel formation, whereas high doses inhibit angiogenesis.¹²¹ More recently, it was shown that statins enhance the mobilization of endothelial progenitor cells (EPCs) from bone marrow to newly forming blood vessels in a PI3K-Akt-dependent manner, 122,123 suggesting another mechanism of Akt-dependent proangiogenic effects of statins. Moreover, it has been shown that statins promote EPC mobilization in patients with stable coronary heart diseases.124 Although there are numerous lines of evidence to suggest that statins promote endothelial cell function and angiogenesis, there is no evidence in clinical studies linking statin treatment to increase in cancer risk.125

Activation of Akt by statins is blocked by treatment with wortmannin or LY294002,91 suggesting that statin activation of Akt is mediated by PI3K. However, the mechanisms by which statins activate PI3K are unknown at present. In this regard, statins have been shown to decrease caveolin-1eNOS interaction and enhance the formation of eNOS-Hsp90-Akt complex in endothelial cells,89 although it is not clear whether these effects of statins are secondary to Akt activation or not. It should also be noted that endothelial cells are relatively unique in this response because activation of Akt by statins is not observed in cardiac or smooth muscle cells,91 suggesting an endothelial cell-specific pathway of PI3K-Akt activation. A recent report has shown that low, clinically relevant doses of statin activate endothelial Ras and promote Akt and eNOS phosphorylation.¹²⁶ It was also reported that higher statin doses are toxic to endothelial cells although they promote an increase in eNOS protein expression. Presumably, the toxicity results from an inhibition of protein prenylation,127 and this may explain the antiangiogenic effects observed in studies performed with higher statin concentrations. 128,129

In addition to their proangiogenic effects, statins have also been shown to exhibit antithrombotic actions in humans, which appears to be independent of their serum cholesterollowering effects. 130 Recent studies have shown that PI3K-Akt pathway inhibits the expression of tissue factor, 131,132 which is the primary cellular initiator of blood coagulation and whose expression is induced in endothelial cells and macrophages by a number of stimuli, including interleukin- 1β and tumor necrosis factor-α.¹³³ Although VEGF activates both tissue factor expression and PI3K-Akt signaling, administration of inhibitors of PI3K-Akt signaling further enhances VEGF-induced tissue factor expression. 131,132 Taken together, these data suggest that statins may inhibit blood coagulation, at least in part, through a selective activation of PI3K-Akt signaling in endothelial cells, leading to an inhibition of tissue factor expression.

Integrated Regulation of Growth and Angiogenesis by Akt

In addition to its role in angiogenesis, Akt has also been implicated as a general regulator of tissue and organ growth. Studies in *Drosophila* have demonstrated that components of the insulin/IGF signaling pathway including Akt are involved in the regulation of organ growth and body size in response to nutritional input.^{134,135} The role of this pathway in vertebrate growth control is indicated by the growth retardation observed after targeted disruption of various components of this pathway in mice.^{28,136,137} Importantly, it has recently been shown that the targeted disruption of Akt1 gene in mice results in general growth retardation.^{28,137}

In higher animals, organ growth is accompanied by the recruitment of new blood vessels. The dual role of Akt signaling in angiogenesis and tissue growth suggests that both processes can be coordinately regulated by this signaling step during organ enlargement. Consistent with this notion, it has been demonstrated that exercise training, a well-known stimulator of muscle hypertrophy, enhances VEGF expression and increases vessel density in skeletal muscles, and that exercise-induced increase in vessel density is blocked by a VEGF-neutralizing antibody.¹³⁸ Likewise, cardiac muscle cell-specific deletion of VEGF gene results in thin ventricular wall with fewer coronary vessels. 139 These results indicate that angiogenesis associated with physiological muscle tissue growth is dependent on paracrine VEGF secretion. Based on the notion that Akt positively regulates organ growth, we have examined the hypothesis that Akt may be involved in VEGF secretion associated with muscle hypertrophy. Indeed, overexpression of Akt in skeletal muscles in vivo induces skeletal muscle hypertrophy, local VEGF production, and angiogenesis.¹⁴⁰ Collectively, these findings suggest that Akt signaling in both muscle cells and endothelial cells coordinately regulate overall growth of muscle tissues in vertebrates. This concept may also be applicable to other organs as well.

Conclusions

PI3K-Akt signaling axis is activated by a variety of stimuli in endothelial cells and regulates multiple critical steps in angiogenesis, including endothelial cell survival, migration, and

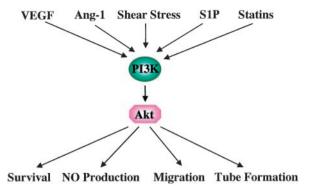


Figure 4. Activation of PI3K-Akt signaling axis in endothelial cells. Various growth factors, mechanical stimuli, and pharmacological interventions activate Akt signaling in endothelial cells, and Akt-dependent pathways control several events critical for cardiovascular homeostasis and angiogenesis.

capillary-like structure formation. Furthermore, this signaling pathway also regulates cardiovascular homeostasis and vessel integrity at least in part by controlling NO synthesis (Figure 4). Angiogenesis has been implicated in the pathophysiology of a number of diseases, and regulation of angiogenesis, both its increase and decrease, could be an important therapeutic strategy for those disease states. Thus, further dissection of the PI3K-Akt pathway and elucidation of the downstream effector molecules will lead to a better understanding of blood vessel growth and may provide avenues for the development of novel therapeutic interventions.

Acknowledgments

This work was supported in part by NIH grants AR-40197, AG-17241, HD-23681, AG-15052, and HL-50692. I.S. is supported by the Tanabe Medical Frontier Conference and American Heart Association New England Affiliate Fellowship Award.

References

- Carmeliet P. Mechanisms of angiogenesis and arteriogenesis. Nat Med. 2000:6:389–395.
- Yancopoulos GD, Davis S, Gale NW, Rudge JS, Wiegand SJ, Holash J. Vascular-specific growth factors and blood vessel formation. *Nature*. 2000;407:242–248.
- Isner JM, Asahara T. Angiogenesis and vasculogenesis as therapeutic strategies for postnatal neovascularization. *J Clin Invest*. 1999;103: 1231–1236.
- Freedman SB, Isner JM. Therapeutic angiogenesis for ischemic cardiovascular disease. J Mol Cell Cardiol. 2001;33:379–393.
- Staal SP. Molecular cloning of the Akt oncogene and its human homologues AKT1 and AKT2: amplification of AKT1 in a primary human gastric adenocarcinoma. *Proc Natl Acad Sci U S A*. 1987;84: 5034–5047.
- Bellacosa A, Testa JR, Staal SP, Tsichlis PN. A retroviral oncogene, Akt, encoding a serine-threonine kinase containing an SH2-like region. Science. 1991;254:274–277.
- Coffer PJ, Woodgett JR. Molecular cloning and characterisation of a novel putative protein-serine kinase related to the cAMP-dependent and protein kinase C families. Eur J Biochem. 1991;201:475–481.
- Jones PF, Jakubowicz T, Pitossi FJ, Maurer F, Hemmings BA. Molecular cloning and identification of a serine/threonine protein kinase of the second-messenger subfamily. *Proc Natl Acad Sci U S A*. 1991; 88:4171–4175.
- Datta SR, Brunet A, Greenberg ME. Cellular survival: a play in three Akts. Genes Dev. 1999;13:2905–2927.
- Scheid MP, Woodgett JR. Pkb/Akt. functional insights from genetic models. Nat Rev Mol Cell Biol. 2001;2:760–768.

- Altomare DA, Guo K, Cheng JQ, Sonoda G, Walsh K, Testa JR. Cloning, chromosomal localization and expression analysis of the mouse Akt2 oncogene. Oncogene. 1995;11:1055–1060.
- Altomare DA, Lyons GE, Mitsuuchi Y, Cheng JQ, Testa JR. Akt2 mRNA is highly expressed in embryonic brown fat and the AKT2 kinase is activated by insulin. *Oncogene*. 1998;16:2407–2411.
- Brodbeck D, Cron P, Hemmings BA. A human protein kinase Bγ with regulatory phosphorylation sites in the activation loop and in the C-terminal hydrophobic domain. J Biol Chem. 1999;274:9133–136.
- Hemmings BA. Akt signaling: linking membrane events to life and death decisions. Science. 1997;275:628–630.
- Downward J. Mechanisms and consequences of activation of protein kinase B/Akt. Curr Opin Cell Biol. 1998;10:262–267.
- Delcommenne M, Tan C, Gray V, Rue L, Woodgett J, Dedhar S. Phosphoinositide-3-OH kinase-dependent regulation of glycogen synthase kinase 3 and protein kinase B/AKT by the integrin-linked kinase. *Proc Natl Acad Sci U S A*. 1998;95:11211–1126.
- Rane MJ, Coxon PY, Powell DW, Webster R, Klein JB, Pierce W, Ping P, McLeish KR. p38 Kinase-dependent MAPKAPK-2 activation functions as 3-phosphoinositide-dependent kinase-2 for Akt in human neutrophils. *J Biol Chem.* 2001;276:3517–3523.
- Balendran A, Casamayor A, Deak M, Paterson A, Gaffney P, Currie R, Downes CP, Alessi DR. PDK1 acquires PDK2 activity in the presence of a synthetic peptide derived from the carboxyl terminus of PRK2. Curr Biol. 1999:9:393–404.
- Toker A, Newton AC. Akt/protein kinase B is regulated by autophosphorylation at the hypothetical PDK-2 site. J Biol Chem. 2000;275: 8271–8274.
- Camper-Kirby D, Welch S, Walker A, Shiraishi I, Setchell KD, Schaefer E, Kajstura J, Anversa P, Sussman MA. Myocardial Akt activation and gender: increased nuclear activity in females versus males. *Circ Res*. 2001:88:1020–1027.
- Andjelkovic M, Jakubowicz T, Cron P, Ming XF, Han JW, Hemmings BA. Activation and phosphorylation of a pleckstrin homology domain containing protein kinase (RAC-PK/PKB) promoted by serum and protein phosphatase inhibitors. *Proc Natl Acad Sci U S A*. 1996;93:5699–5704.
- Yao R, Cooper GM. Requirement for phosphatidylinositol-3 kinase in the prevention of apoptosis by nerve growth factor. Science. 1995;267:2003–2006.
- Franke TF, Yang SI, Chan TO, Datta K, Kazlauskas A, Morrison DK, Kaplan DR, Tsichlis PN. The protein kinase encoded by the Akt protooncogene is a target of the PDGF-activated phosphatidylinositol 3-kinase. Cell. 1995;81:727–736.
- Dudek H, Datta SR, Franke TF, Birnbaum MJ, Yao R, Cooper GM, Segal RA, Kaplan DR, Greenberg ME. Regulation of neuronal survival by the serine-threonine protein kinase Akt. Science. 1997;275:661–665.
- Fujio Y, Guo K, Mano T, Mitsuuchi Y, Testa JR, Walsh K. Cell cycle withdrawal promotes myogenic induction of Akt, a positive modulator of myocyte survival. *Mol Cell Biol*. 1999;19:5073–5082.
- Fujio Y, Nguyen T, Wencker D, Kitsis RN, Walsh K. Akt promotes survival of cardiomyocytes in vitro and protects against ischemiareperfusion injury in mouse heart. Circulation. 2000;101:660–667.
- Staveley BE, Ruel L, Jin J, Stambolic V, Mastronardi FG, Heitzler P, Woodgett JR, Manoukian AS. Genetic analysis of protein kinase B (AKT) in Drosophila. Curr Biol. 1998;8:599–602.
- Chen WS, Xu PZ, Gottlob K, Chen ML, Sokol K, Shiyanova T, Roninson I, Weng W, Suzuki R, Tobe K, Kadowaki T, Hay N. Growth retardation and increased apoptosis in mice with homozygous disruption of the Akt1 gene. *Genes Dev.* 2001;15:2203–2208.
- Datta SR, Dudek H, Tao X, Masters S, Fu H, Gotoh Y, Greenberg ME. Akt phosphorylation of BAD couples survival signals to the cell-intrinsic death machinery. Cell. 1997;91:231–241.
- del Peso L, Gonzalez-Garcia M, Page C, Herrera R, Nunez G. Interleukin-3-induced phosphorylation of BAD through the protein kinase Akt. Science. 1997;278:687–689.
- Brunet A, Bonni A, Zigmond MJ, Lin MZ, Juo P, Hu LS, Anderson MJ, Arden KC, Blenis J, Greenberg ME. Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. *Cell*. 1999;96:857–868.
- Kops GJ, de Ruiter ND, De Vries-Smits AM, Powell DR, Bos JL, Burgering BM. Direct control of the Forkhead transcription factor AFX by protein kinase B. *Nature*. 1999;398:630–634.
- Tang ED, Nunez G, Barr FG, Guan KL. Negative regulation of the forkhead transcription factor FKHR by Akt. J Biol Chem. 1999;274: 16741–16746.

- Ozes ON, Mayo LD, Gustin JA, Pfeffer SR, Pfeffer LM, Donner DB. NF-κB activation by tumour necrosis factor requires the Akt serinethreonine kinase. *Nature*. 1999;401:82–85.
- Romashkova JA, Makarov SS. NF-κB is a target of AKT in anti-apoptotic PDGF signalling. Nature. 1999;401:86–90.
- Wang Q, Somwar R, Bilan PJ, Liu Z, Jin J, Woodgett JR, Klip A. Protein kinase B/Akt participates in GLUT4 translocation by insulin in L6 myoblasts. *Mol Cell Biol*. 1999;19:4008–4018.
- Cross DA, Alessi DR, Cohen P, Andjelkovich M, Hemmings BA. Inhibition of glycogen synthase kinase-3 by insulin mediated by protein kinase B. *Nature*. 1995;378:785–789.
- Brennan P, Babbage JW, Burgering BM, Groner B, Reif K, Cantrell DA. Phosphatidylinositol 3-kinase couples the interleukin-2 receptor to the cell cycle regulator E2F. *Immunity*. 1997;7:679–689.
- Kang SS, Kwon T, Kwon DY, Do SI. Akt protein kinase enhances human telomerase activity through phosphorylation of telomerase reverse transcriptase subunit. *J Biol Chem.* 1999;274:13085–13090.
- Zhou BP, Liao Y, Xia W, Spohn B, Lee MH, Hung MC. Cytoplasmic localization of p21Cip1/WAF1 by Akt-induced phosphorylation in HER-2/neu-overexpressing cells. *Nat Cell Biol*. 2001;3:245–252.
- Mayo LD, Donner DB. A phosphatidylinositol 3-kinase/Akt pathway promotes translocation of Mdm2 from the cytoplasm to the nucleus. Proc Natl Acad Sci U S A. 2001;98:11598–11603.
- Rossig L, Jadidi AS, Urbich C, Badorff C, Zeiher AM, Dimmeler S. Aktdependent phosphorylation of p21(Cip1) regulates PCNA binding and proliferation of endothelial cells. *Mol Cell Biol*. 2001;21:5644–5657.
- Breitschopf K, Zeiher AM, Dimmeler S. Pro-atherogenic factors induce telomerase inactivation in endothelial cells through an Akt-dependent mechanism. FEBS Lett. 2001;493:21–25.
- Rossig L, Badorff C, Holzmann Y, Zeiher AM, Dimmeler S. Glycogen synthase kinase-3 couples Akt-dependent signaling to the regulation of p21Cip1 degradation. *J Biol Chem*. 2002;277:9684–9689.
- Shah OJ, Anthony JC, Kimball SR, Jefferson LS. 4E-BP1 and S6K1: translational integration sites for nutritional and hormonal information in muscle. *Am J Physiol*. 2000;279:E715–E729.
- Alessi DR, Kozlowski MT, Weng QP, Morrice N, Avruch J. 3-Phosphoinositide-dependent protein kinase 1 (PDK1) phosphorylates and activates the p70 S6 kinase in vivo and in vitro. Curr Biol. 1998;8:69–81.
- Pullen N, Dennis PB, Andjelkovic M, Dufner A, Kozma SC, Hemmings BA, Thomas G. Phosphorylation and activation of p70s6k by PDK1. Science. 1998:279:707–710.
- Alon T, Hemo I, Itin A, Pe'er J, Stone J, Keshet E. Vascular endothelial growth factor acts as a survival factor for newly formed retinal vessels and has implications for retinopathy of prematurity. *Nat Med.* 1995;1:1024–1028.
- 49. Gerber HP, McMurtrey A, Kowalski J, Yan M, Keyt BA, Dixit V, Ferrara N. Vascular endothelial growth factor regulates endothelial cell survival through the phosphatidylinositol 3'-kinase/Akt signal transduction pathway: requirement for Flk-1/KDR activation. *J Biol Chem*. 1998;273:30336–30343.
- Fujio Y, Walsh K. Akt mediates cytoprotection of endothelial cells by vascular endothelial growth factor in an anchorage-dependent manner. *J Biol Chem.* 1999;274:16349–16354.
- Papapetropoulos A, Fulton D, Mahboubi K, Kalb RG, O'Connor DS, Li F, Altieri DC, Sessa WC. Angiopoietin-1 inhibits endothelial cell apoptosis via the Akt/survivin pathway. J Biol Chem. 2000;275:9102–105.
- Kim I, Kim HG, So JN, Kim JH, Kwak HJ, Koh GY. Angiopoietin-1 regulates endothelial cell survival through the phosphatidylinositol 3'-Kinase/Akt signal transduction pathway. Circ Res. 2000;86:24–29.
- Hermann C, Assmus B, Urbich C, Zeiher AM, Dimmeler S. Insulin-mediated stimulation of protein kinase Akt: a potent survival signaling cascade for endothelial cells. Arterioscler Thromb Vasc Biol. 2000;20:402–409.
- Michell BJ, Griffiths JE, Mitchelhill KI, Rodriguez-Crespo I, Tiganis T, Bozinovski S, de Montellano PR, Kemp BE, Pearson RB. The Akt kinase signals directly to endothelial nitric oxide synthase. *Curr Biol*. 1999:9:845–848.
- 55. Igarashi J, Bernier SG, Michel T. Sphingosine 1-phosphate and activation of endothelial nitric-oxide synthase. differential regulation of Akt and MAP kinase pathways by EDG and bradykinin receptors in vascular endothelial cells. *J Biol Chem.* 2001;276:12420–12426.
- Morales-Ruiz M, Lee MJ, Zollner S, Gratton JP, Scotland R, Shiojima I, Walsh K, Hla T, Sessa WC. Sphingosine 1-phosphate activates Akt, nitric oxide production, and chemotaxis through a Gi protein/phosphoinositide 3-kinase pathway in endothelial cells. *J Biol Chem.* 2001;276: 19672–19677.

- Nakagami H, Morishita R, Yamamoto K, Taniyama Y, Aoki M, Matsumoto K, Nakamura T, Kaneda Y, Horiuchi M, Ogihara T. Mitogenic and antiapoptotic actions of hepatocyte growth factor through ERK, Stat3, and Akt in endothelial cells. *Hypertension*. 2001;37:581–586.
- Schonherr E, Levkau B, Schaefer L, Kresse H, Walsh K. Decorinmediated signal transduction in endothelial cells. Involvement of Akt/ protein kinase B in up-regulation of p21(WAF1/CIP1) but not p27(KIP1). *J Biol Chem.* 2001;276:40687–40692.
- Dimmeler S, Assmus B, Hermann C, Haendeler J, Zeiher AM. Fluid shear stress stimulates phosphorylation of Akt in human endothelial cells: involvement in suppression of apoptosis. Circ Res. 1998;83:334–341.
- 59a. Simoncini T, Hafezi-Moghadam A, Brazil DP, Ley K, Chin WW, Liao JK. Interaction of oestrogen receptor with the regulatory subunit of phosphatidylinositol-3-OH kinase. *Nature*. 2000;407:538–341.
- 59b. Thomas SR, Chen K, Keaney JFJ. Hydrogen peroxide activates endothelial nitric-oxide synthase through coordinated phoshorylation and dephosphorylation via a phosphoinositide 3-kinase-dependent signaling pathway. J Biol Chem. 2002;277:6017–6024.
- 59c. Hafezi-Moghadam A, Simoncini T, Yang E, Limbourg FP, Plumier JC, Rebsamen MC, Hsieh CM, Chui DS, Thomas KL, Prorock AJ, Laubach VE, Moskowitz MA, French BA, Ley K, Liao JK. Acute cardiovascular protective effects of corticosteroids are mediated by non-transcriptional activation of endothelial nitric oxide synthase. *Nat Med.* 2002;8:473–479.
- Brooks PC, Clark RA, Cheresh DA. Requirement of vascular integrin α_vβ₃ for angiogenesis. *Science*. 1994;264:569–571.
- Frisch SM, Screaton RA. Anoikis mechanisms. Curr Opin Cell Biol. 2001:13:555–562.
- Giancotti FG, Ruoslahti E. Integrin signaling. Science. 1999;285: 1028–1032.
- Eliceiri BP. Integrin and growth factor receptor crosstalk. Circ Res. 2001;89:1104–1110.
- 64. Bachelder RE, Wendt MA, Fujita N, Tsuruo T, Mercurio AM. The cleavage of Akt/protein kinase B by death receptor signaling is an important event in detachment-induced apoptosis. *J Biol Chem.* 2001; 276:34702–34707.
- Byzova TV, Goldman CK, Pampori N, Thomas KA, Bett A, Shattil SJ, Plow EF. A mechanism for modulation of cellular responses to VEGF: activation of the integrins. *Mol Cell*. 2000;6:851–860.
- Suhara T, Mano T, Oliveira BE, Walsh K. Phosphatidylinositol 3-kinase/Akt signaling controls endothelial cell sensitivity to Fas-mediated apoptosis via regulation of FLICE-inhibitory protein (FLIP). Circ Res. 2001;89:13–19.
- 67. Gratton JP, Morales-Ruiz M, Kureishi Y, Fulton D, Walsh K, Sessa WC. Akt down-regulation of p38 signaling provides a novel mechanism of vascular endothelial growth factor-mediated cytoprotection in endothelial cells. *J Biol Chem.* 2001;276:30359–30365.
- Ku DD, Zaleski JK, Liu S, Brock TA. Vascular endothelial growth factor induces EDRF-dependent relaxation in coronary arteries. Am J Physiol. 1993;265:H586–H592.
- Yang R, Thomas GR, Bunting S, Ko A, Ferrara N, Keyt B, Ross J, Jin H. Effects of vascular endothelial growth factor on hemodynamics and cardiac performance. *J Cardiovasc Pharmacol*. 1996;27:838-844.
- van der Zee R, Murohara T, Luo Z, Zollmann F, Passeri J, Lekutat C, Isner JM. Vascular endothelial growth factor/vascular permeability factor augments nitric oxide release from quiescent rabbit and human vascular endothelium. *Circulation*. 1997;95:1030–1037.
- Papapetropoulos A, Garcia-Cardena G, Madri JA, Sessa WC. Nitric oxide production contributes to the angiogenic properties of vascular endothelial growth factor in human endothelial cells. *J Clin Invest*. 1997;100:3131–3139.
- Fulton D, Gratton JP, McCabe TJ, Fontana J, Fujio Y, Walsh K, Franke TF, Papapetropoulos A, Sessa WC. Regulation of endothelium-derived nitric oxide production by the protein kinase Akt. *Nature*. 1999;399:597–601.
- Dimmeler S, Fleming I, Fisslthaler B, Hermann C, Busse R, Zeiher AM. Activation of nitric oxide synthase in endothelial cells by Akt-dependent phosphorylation. *Nature*. 1999;399:601–605.
- Boo YC, Sorescu G, Boyd N, Shiojima I, Walsh K, Du J, Jo H. Shear stress stimulates phosphorylation of endothelial nitric-oxide synthase at Ser1179 by Akt-independent mechanisms: role of protein kinase A. *J Biol Chem.* 2002;277:3388–3396.
- Luo Z, Fujio Y, Kureishi Y, Rudic RD, Daumerie G, Fulton D, Sessa WC, Walsh K. Acute modulation of endothelial Akt/PKB activity alters nitric oxidedependent vasomotor activity in vivo. J Clin Invest. 2000;106:493–499.
- 75a. Scotland RS, Morales-Ruiz M, Chen Y, Yu J, Rudic RD, Fulton D, Gratton JP, Sessa WC. Functional reconstitution of endothelial nitric

- oxide synthase reveals the importance of serine 1179 in endothelium-dependent vasomotion. *Circ Res.* 2002;90:904–910.
- Nishimatsu H, Suzuki E, Nagata D, Moriyama N, Satonaka H, Walsh K, Sata M, Kangawa K, Matsuo H, Goto A, Kitamura T, Hirata Y. Adrenomedullin induces endothelium-dependent vasorelaxation via the Phosphatidylinositol 3-Kinase/Akt-dependent pathway in rat aorta. Circ Res. 2001;89:63–70.
- Du XL, Edelstein D, Dimmeler S, Ju Q, Sui C, Brownlee M. Hyperglycemia inhibits endothelial nitric oxide synthase activity by posttranslational modification at the Akt site. *J Clin Invest*. 2001;108:1341–1348.
- Garcia-Cardena G, Oh P, Liu J, Schnitzer JE, Sessa WC. Targeting of nitric oxide synthase to endothelial cell caveolae via palmitoylation: implications for nitric oxide signaling. *Proc Natl Acad Sci U S A*. 1996; 93:6448–6453.
- Shaul PW, Smart EJ, Robinson LJ, German Z, Yuhanna IS, Ying Y, Anderson RG, Michel T. Acylation targets endothelial nitric-oxide synthase to plasmalemmal caveolae. *J Biol Chem.* 1996;271:6518–6522.
- Feron O, Belhassen L, Kobzik L, Smith TW, Kelly RA, Michel T. Endothelial nitric oxide synthase targeting to caveolae: specific interactions with caveolin isoforms in cardiac myocytes and endothelial cells. *J Biol Chem.* 1996;271:22810–22814.
- Ju H, Zou R, Venema VJ, Venema RC. Direct interaction of endothelial nitric-oxide synthase and caveolin-1 inhibits synthase activity. *J Biol Chem.* 1997;272:18522–18525.
- Bucci M, Gratton JP, Rudic RD, Acevedo L, Roviezzo F, Cirino G, Sessa WC.
 In vivo delivery of the caveolin-1 scaffolding domain inhibits nitric oxide synthesis and reduces inflammation. *Nat Med.* 2000:6:1362–1367.
- Drab M, Verkade P, Elger M, Kasper M, Lohn M, Lauterbach B, Menne J, Lindschau C, Mende F, Luft FC, Schedl A, Haller H, Kurzchalia TV. Loss of caveolae, vascular dysfunction, and pulmonary defects in caveolin-1 gene-disrupted mice. *Science*. 2001;293:2449–2452.
- 84. Feron O, Dessy C, Opel DJ, Arstall MA, Kelly RA, Michel T. Modulation of the endothelial nitric-oxide synthase-caveolin interaction in cardiac myocytes: implications for the autonomic regulation of heart rate. *J Biol Chem.* 1998;273:30249–30254.
- Govers R, Rabelink TJ. Cellular regulation of endothelial nitric oxide synthase. Am J Physiol Renal Physiol. 2001;280:F193–F206.
- Garcia-Cardena G, Fan R, Shah V, Sorrentino R, Cirino G, Papapetropoulos A, Sessa WC. Dynamic activation of endothelial nitric oxide synthase by Hsp90. *Nature*. 1998;392:821–824.
- Sato S, Fujita N, Tsuruo T. Modulation of Akt kinase activity by binding to Hsp90. Proc Natl Acad Sci U S A. 2000;97:10832–10837.
- 88. Brouet A, Sonveaux P, Dessy C, Balligand JL, Feron O. Hsp90 ensures the transition from the early Ca²⁺-dependent to the late phosphorylation-dependent activation of the endothelial nitric-oxide synthase in vascular endothelial growth factor-exposed endothelial cells. *J Biol Chem.* 2001;276:32663–32669.
- Brouet A, Sonveaux P, Dessy C, Moniotte S, Balligand JL, Feron O. Hsp90 and caveolin are key targets for the proangiogenic nitric oxidemediated effects of statins. Circ Res. 2001;89:866–873.
- 89a. Fontana J, Fulton D, Chen Y, Fairchild TA, McCabe TJ, Fujita N, Tsuruo T, Sessa WC. Domain mapping studies reveal that the M domain of hsp90 serves as a molecular scaffold to regulate Akt-dependent phosphorylation of endothelial nitric oxide synthase and NO release. Circ Res. 2002;90:866–873.
- Morales-Ruiz M, Fulton D, Sowa G, Languino LR, Fujio Y, Walsh K, Sessa WC. Vascular endothelial growth factor-stimulated actin reorganization and migration of endothelial cells is regulated via the serine/ threonine kinase Akt. Circ Res. 2000;86:892–896.
- Kureishi Y, Luo Z, Shiojima I, Bialik A, Fulton D, Lefer DJ, Sessa WC, Walsh K, The HMG-CoA reductase inhibitor simvastatin activates the protein kinase Akt and promotes angiogenesis in normocholesterolemic animals. *Nat Med.* 2000;6:1004–1010.
- Dimmeler S, Dernbach E, Zeiher AM. Phosphorylation of the endothelial nitric oxide synthase at ser-1177 is required for VEGF-induced endothelial cell migration. FEBS Lett. 2000;477:258–262.
- Lee MJ, Thangada S, Claffey KP, Ancellin N, Liu CH, Kluk M, Volpi M, Sha'afi RI, Hla T. Vascular endothelial cell adherens junction assembly and morphogenesis induced by sphingosine-1-phosphate. *Cell*. 1999;99:301–312.
- Panetti TS, Nowlen J, Mosher DF. Sphingosine-1-phosphate and lysophosphatidic acid stimulate endothelial cell migration. Arterioscler Thromb Vasc Biol. 2000;20:1013–1019.
- Chavakis E, Dernbach E, Hermann C, Mondorf UF, Zeiher AM, Dimmeler S.
 Oxidized LDL inhibits vascular endothelial growth factor-induced endothelial

- cell migration by an inhibitory effect on the Akt/endothelial nitric oxide synthase pathway. *Circulation*. 2001;103:2102–2107.
- Meili R, Ellsworth C, Lee S, Reddy TB, Ma H, Firtel RA. Chemoattractant-mediated transient activation and membrane localization of Akt/PKB is required for efficient chemotaxis to cAMP in Dictyostelium. EMBO J. 1999;18:2092–2105.
- Servant G, Weiner OD, Herzmark P, Balla T, Sedat JW, Bourne HR. Polarization of chemoattractant receptor signaling during neutrophil chemotaxis. Science. 2000;287:1037–1040.
- Hirsch E, Katanaev VL, Garlanda C, Azzolino O, Pirola L, Silengo L, Sozzani S, Mantovani A, Altruda F, Wymann MP. Central role for G protein–coupled phosphoinositide 3-kinase γ in inflammation. *Science*. 2000;287:1049–1053.
- 99. Li Z, Jiang H, Xie W, Zhang Z, Smrcka AV, Wu D. Roles of PLC- β 2 and - β 3 and PI3K γ in chemoattractant-mediated signal transduction. *Science*. 2000;287:1046–1049.
- 100. Sasaki T, Irie-Sasaki J, Jones RG, Oliveira-dos-Santos AJ, Stanford WL, Bolon B, Wakeham A, Itie A, Bouchard D, Kozieradzki I, Joza N, Mak TW, Ohashi PS, Suzuki A, Penninger JM. Function of PI3Kγ in thymocyte development, T cell activation, and neutrophil migration. *Science*. 2000;287:1040–1046.
- Lauffenburger DA, Horwitz AF. Cell migration: a physically integrated molecular process. Cell. 1996;84:359–369.
- Ridley AJ. Rho proteins, PI 3-kinases, and monocyte/macrophage motility. FEBS Lett. 2001;498:168–171.
- 103. van Nieuw Amerongen GP, van Hinsbergh VW. Cytoskeletal effects of rho-like small guanine nucleotide-binding proteins in the vascular system. Arterioscler Thromb Vasc Biol. 2001;21:300–311.
- 104. Garcia JG, Liu F, Verin AD, Birukova A, Dechert MA, Gerthoffer WT, Bamberg JR, English D. Sphingosine 1-phosphate promotes endothelial cell barrier integrity by Edg-dependent cytoskeletal rearrangement. *J Clin Invest.* 2001;108:689–701.
- 105. Soga N, Namba N, McAllister S, Cornelius L, Teitelbaum SL, Dowdy SF, Kawamura J, Hruska KA. Rho family GTPases regulate VEGF-stimulated endothelial cell motility. Exp Cell Res. 2001;269:73–87.
- 106. Kwon T, Kwon DY, Chun J, Kim JH, Kang SS. Akt protein kinase inhibits Rac1-GTP binding through phosphorylation at serine 71 of Rac1. J Biol Chem. 2000;275:423–428.
- 107. Lee MJ, Thangada S, Paik JH, Sapkota GP, Ancellin N, Chae SS, Wu M, Morales-Ruiz M, Sessa WC, Alessi DR, Hla T. Akt-mediated phosphorylation of the G protein–coupled receptor EDG-1 is required for endothelial cell chemotaxis. *Mol Cell*. 2001;8:693–704.
- Higuchi M, Masuyama N, Fukui Y, Suzuki A, Gotoh Y. Akt mediates Rac/Cdc42-regulated cell motility in growth factor-stimulated cells and in invasive PTEN knockout cells. Curr Biol. 2001;11:1958–1962.
- Djuder N, Schmidt G, Frings M, Cavalie A, Thelen M, Aktories K. Rac and phosphatidylinositol 3-kinase regulate the protein kinase B in Fc epsilon RI signaling in RBL 2H3 mast cells. *J Immunol*. 2001;166:1627–1634.
- 110. Genot EM, Arrieumerlou C, Ku G, Burgering BM, Weiss A, Kramer IM. The T-cell receptor regulates Akt (protein kinase B) via a pathway involving Rac1 and phosphatidylinositide 3-kinase. Mol Cell Biol. 2000;20:5469–5478.
- Daniels RH, Bokoch GM. p21-activated protein kinase: a crucial component of morphological signaling? Trends Biochem Sci. 1999;24:350–355.
- Chung CY, Potikyan G, Firtel RA. Control of cell polarity and chemotaxis by Akt/PKB and PI3 kinase through the regulation of PAKa. Mol Cell. 2001;7:937–947.
- Tang Y, Zhou H, Chen A, Pittman RN, Field J, The Akt proto-oncogene links Ras to Pak and cell survival signals. *J Biol Chem.*;2000;275:9106–9109.
- Maron DJ, Fazio S, Linton MF. Current perspectives on statins. Circulation. 2000:101:207–213.
- 115. Anderson TJ, Meredith IT, Yeung AC, Frei B, Selwyn AP, Ganz P. The effect of cholesterol-lowering and antioxidant therapy on endotheliumdependent coronary vasomotion. N Engl J Med. 1995;332:488–493.
- 116. Treasure CB, Klein JL, Weintraub WS, Talley JD, Stillabower ME, Kosinski AS, Zhang J, Boccuzzi SJ, Cedarholm JC, Alexander RW. Beneficial effects of cholesterol-lowering therapy on the coronary endothelium in patients with coronary artery disease. N Engl J Med. 1995;332:481–487
- Williams JK, Sukhova GK, Herrington DM, Libby P. Pravastatin has cholesterol-lowering independent effects on the artery wall of atherosclerotic monkeys. J Am Coll Cardiol. 1998;31:684

 –691.
- Endres M, Laufs U, Huang Z, Nakamura T, Huang P, Moskowitz MA, Liao JK. Stroke protection by 3-hydroxy-3-methylglutaryl (HMG)-CoA

- reductase inhibitors mediated by endothelial nitric oxide synthase. *Proc Natl Acad Sci U S A*. 1998;95:8880–8885.
- Lefer AM, Campbell B, Shin YK, Scalia R, Hayward R, Lefer DJ. Simvastatin preserves the ischemic-reperfused myocardium in normocholesterolemic rat hearts. Circulation. 1999:100:178–184.
- 120. Sata M, Nishimatsu H, Suzuki E, Sugiura S, Yoshizumi M, Ouchi Y, Hirata Y, Nagai R. Endothelial nitric oxide synthase is essential for the HMG-CoA reductase inhibitor cerivastatin to promote collateral growth in response to ischemia. FASEB J. 2001;15:2530–2532.
- 121. Weis M, Heeschen C, Glassford AJ, Cooke JP. Statins have biphasic effects on angiogenesis. *Circulation*. 2002;105:739–745.
- 122. Dimmeler S, Aicher A, Vasa M, Mildner-Rihm C, Adler K, Tiemann M, Rutten H, Fichtlscherer S, Martin H, Zeiher AM. HMG-CoA reductase inhibitors (statins) increase endothelial progenitor cells via the PI 3-kinase/Akt pathway. *J Clin Invest*. 2001;108:391–397.
- Llevadot J, Murasawa S, Kureishi Y, Uchida S, Masuda H, Kawamoto A, Walsh K, Isner JM, Asahara T. HMG-CoA reductase inhibitor mobilizes bone marrow-derived endothelial progenitor cells. *J Clin Invest*. 2001;108:399–405.
- 124. Vasa M, Fichtlscherer S, Adler K, Aicher A, Martin H, Zeiher AM, Dimmeler S. Increase in circulating endothelial progenitor cells by statin therapy in patients with stable coronary artery disease. *Circulation*. 2001;103:2885–2890.
- 125. Davidson MH. Safety profiles for the HMG-CoA reductase inhibitors: treatment and trust. *Drugs*. 2001;61:197–206.
- Urbich C, Dernbach E, Zeiher AM, Dimmeler S. Double-edged role of statins in angiogenesis signaling. Circ Res. 2002;90:737–744.
- Laufs U, La Fata V, Plutzky J, Liao JK. Upregulation of endothelial nitric oxide synthase by HMG CoA reductase inhibitors. *Circulation*. 1998;97:1129–1135.
- 128. Vincent L, Chen W, Hong L, Mirshahi F, Mishal Z, Mirshahi-Khorassani T, Vannier JP, Soria J, Soria C. Inhibition of endothelial cell migration by cerivastatin, an HMG-CoA reductase inhibitor: contribution to its anti-angiogenic effect. FEBS Lett. 2001;495:159–166.
- 129. Vincent L, Soria C, Mirshahi F, Opolon P, Mishal Z, Vannier JP, Soria J, Hong L. Cerivastatin, an inhibitor of 3-hydroxy-3-methylglutaryl coenzyme a reductase, inhibits endothelial cell proliferation induced by angiogenic factors in vitro and angiogenesis in in vivo models. Arterioscler Thromb Vasc Biol. 2002;22:623–629.
- Dangas G, Smith DA, Unger AH, Shao JH, Meraj P, Fier C, Cohen AM, Fallon JT, Badimon JJ, Ambrose JA. Pravastatin: an antithrombotic effect independent of the cholesterol-lowering effect. *Thromb Haemost*. 2000;83:688–692.
- 131. Blum S, Issbruker K, Willuweit A, Hehlgans S, Lucerna M, Mechtcheriakova D, Walsh K, von der Ahe D, Hofer E, Clauss M. An inhibitory role of the phosphatidylinositol 3-kinase-signaling pathway in vascular endothelial growth factor-induced tissue factor expression. J Biol Chem. 2001;276:33428–33434.
- 132. Kim I, Oh JL, Ryu YS, So JN, Sessa WC, Walsh K, Koh GY. Angiopoietin-1 negatively regulates expression and activity of tissue factor in endothelial cells. FASEB J. 2002;16:126–128.
- Moons AH, Levi M, Peters RJ. Tissue factor and coronary artery disease. Cardiovasc Res. 2002;53:313–325.
- Verdu J, Buratovich MA, Wilder EL, Birnbaum MJ. Cell-autonomous regulation of cell and organ growth in *Drosophila* by Akt/PKB. *Nat Cell Biol.* 1999;1:500–506.
- Weinkove D, Leevers SJ. The genetic control of organ growth: insights from *Drosophila*. Curr Opin Genet Dev. 2000;10:75–80.
- Rother KI, Accili D. Role of insulin receptors and IGF receptors in growth and development. *Pediatr Nephrol*. 2000;14:558–561.
- Cho H, Thorvaldsen JL, Chu Q, Feng F, Birnbaum MJ. Akt1/pkbα is required for normal growth but dispensable for maintenance of glucose homeostasis in mice. *J Biol Chem.* 2001;276:38349–38352.
- Amaral SL, Papanek PE, Greene AS. Angiotensin II and VEGF are involved in angiogenesis induced by short-term exercise training. Am J Physiol Heart Circ Physiol. 2001;281:H1163–H1169.
- 139. Giordano FJ, Gerber HP, Williams SP, VanBruggen N, Bunting S, Ruiz-Lozano P, Gu Y, Nath AK, Huang Y, Hickey R, Dalton N, Peterson KL, Ross J Jr, Chien KR, Ferrara N. A cardiac myocyte vascular endothelial growth factor paracrine pathway is required to maintain cardiac function. *Proc Natl Acad Sci U S A*. 2001;98:5780–5785.
- Takahashi A, Kureishi Y, Yang J, Luo Z, Guo K, Mukhopadhyay D, Ivashchenko Y, Branellec D, Walsh K. Myogenic Akt signaling regulates blood vessel recruitment during myofiber growth. *Mol Cell Biol*. 2002;22:4803–4814.