



Role of Cholesterol and Lipid Rafts in Cancer Signaling: A Promising Therapeutic Opportunity?

Rosa Vona, Elisabetta lessi and Paola Matarrese*

Center for Gender-Specific Medicine, Istituto Superiore di Sanità [Italian National Institute of Health], Rome, Italy

Cholesterol is a lipid molecule that plays an essential role in a number of biological processes, both physiological and pathological. It is an essential structural constituent of cell membranes, and it is fundamental for biosynthesis, integrity, and functions of biological membranes, including membrane trafficking and signaling. Moreover, cholesterol is the major lipid component of lipid rafts, a sort of lipid-based structures that regulate the assembly and functioning of numerous cell signaling pathways, including those related to cancer, such as tumor cell growth, adhesion, migration, invasion, and apoptosis. Considering the importance of cholesterol metabolism, its homeostasis is strictly regulated at every stage: import, synthesis, export, metabolism, and storage. The alterations of this homeostatic balance are known to be associated with cardiovascular diseases and atherosclerosis, but mounting evidence also connects these behaviors to increased cancer risks. Although there is conflicting evidence on the role of cholesterol in cancer development, most of the studies consistently suggest that a dysregulation of cholesterol homeostasis could lead to cancer development. This review aims to discuss the current understanding of cholesterol homeostasis in normal and cancerous cells, summarizing key findings from recent preclinical and clinical studies that have investigated the role of major players in cholesterol regulation and the organization of lipid rafts, which could represent promising therapeutic targets.

Keywords: cholesterol, cancer, lipid rafts, cell death, invasion, metastases, therapy

INTRODUCTION

Cholesterol is a primary lipid molecule that plays an essential role in a number of biological processes, both at physiological and pathological level (Maxfield and Tabas, 2005).

OPEN ACCESS

Edited by:

Giulio Preta, Vilnius University, Lithuania

Reviewed by:

Toshiyuki Murai, Osaka University, Japan Faustino Mollinedo, Consejo Superior de Investigaciones Científicas (CSIC), Spain

> *Correspondence: Paola Matarrese paola.matarrese@iss.it

Specialty section:

This article was submitted to Membrane Traffic, a section of the journal Frontiers in Cell and Developmental Biology

> Received: 29 October 2020 Accepted: 15 February 2021 Published: 19 March 2021

Citation:

Vona R, lessi E and Matarrese P (2021) Role of Cholesterol and Lipid Rafts in Cancer Signaling: A Promising Therapeutic Opportunity? Front. Cell Dev. Biol. 9:622908. doi: 10.3389/fcell.2021.622908

1

Abbreviations: ABCA1, ATP-binding cassette subfamily A member 1; ABCG1, ATP-binding cassette subfamily G member 1; ACAT1, acyl-CoA:cholesteryl acyltransferase 1; AMPK, 5'adenosine monophosphate-activated protein kinase; AMPK, AMP-activated protein kinase; ApoA-I, lipid-poor apolipoprotein A-I; CE, cholesteryl ester; ER, endoplasmic reticulum; FPP, farnesyl pyrophosphate; FPP, farnesyl pyrophosphate; FTase, farnesyl transferase; GGPP, geranylgeranyl pyrophosphate. GGTase, geranylgeranyl transferases; HDL, high-density lipoprotein; HMG-CoA, 3-hydroxy-3-methyl-glutaryl CoA; HMGR, HMG reductase; IGF, insulin-like growth factor; INSIGs, insulin-induced gene; LDL, low-density lipoprotein; LDLR, LDL receptor; LXR, liver X receptor; MβCD, methyl-β-cyclodextrin; miRNA, microRNA; mTOR, mammalian target of rapamycin complex 1; NPC1L1, Niemann-Pick type C1-like 1; PI3K, phosphatidylinositol 3-kinase; SCAP, SREBP cleavage activating protein; GTPase, small GTP-binding protein; SOAT, sterol-o-acyltransferase; SQLE, squalene epoxidase; SRE, sterol regulatory element; SREBP2, sterol regulatory element-binding protein 2; VLDL, very-low-density lipoprotein.

Cholesterol, in addition to being an important constituent of cell membranes, is fundamental for their biogenesis, and is indispensable for maintaining integrity and functions of biological membranes, including endocytosis, membrane trafficking, and signaling (Maxfield and Tabas, 2005; Yamauchi and Rogers, 2018). Inside the cell, cholesterol, heterogeneously distributed among the organelles, modulates the immune system, and represents a precursor of hormones such as sexual hormones and vitamin D (Mollinedo and Gajate, 2020; **Figure 1**).

Recently, cholesterol has played a key role in cancer research because of its potential therapeutic implications both in prevention and in treatment. However, the role of cholesterol in oncogenesis is still debated (DuBroff and de Lorgeril, 2015). Literature data reported a contradictory role of cholesterol depending on the type of tumor (Ding et al., 2019). Excess of cholesterol is related to breast, colon, rectal, prostatic, and testicular cancers (Llaverias et al., 2011; Pelton et al., 2012; Murai, 2015; Radisauskas et al., 2016), while some prospective cohort studies showed an inverse association in gastric and prostate cancers (Asano et al., 2008; Heir et al., 2016). This review aims to discuss the current knowledge of cholesterol homeostasis, critically analyzing the most recent preclinical and clinical studies investigating the role of the principal players of the cholesterol biosynthetic pathway, and of the cholesterol-based membrane structure's lipid rafts in the field of cancer.

CHOLESTEROL METABOLISM

Cholesterol is produced through a cascade of enzymatic reactions, namely, mevalonate pathway, which requires the participation of different enzymes localized on the membranes of the endoplasmic reticulum (ER). Briefly, the combination of three acetyl-CoA molecules leads to the formation of one 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) molecule. The latter, by the action of HMG-CoA reductase (HMGCR), is converted into mevalonate, in turn transformed into squalene (SQL), and ultimately into cholesterol by means of a number of reactions (**Figure 2**). Food can be a source of cholesterol too. In fact, the Niemann–Pick type C1-like 1 (NPC1L1) protein, present on the membrane of the intestinal enterocytes, is responsible for the absorption of cholesterol, which is released as chylomicrons,

Role of cholesterol

- ✓ Membranes: integrity, functions, biogenesis, trafficking and signaling
- ✓ Precursor for steroid hormones: estrogens and androgens
- Precursor for vitamin D
- ✓ Precursor for oxysterols
- Precursor for cholesteryl ester
- ✓ Precursor for bile acids

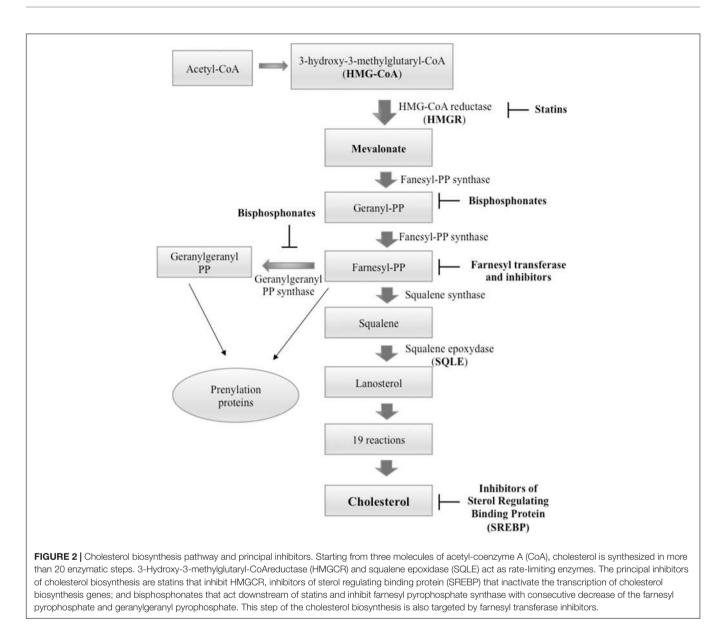
FIGURE 1 | Principal functions of cholesterol. Figure inspired by Mollinedo and Gajate (2020).

triglyceride-rich lipid particles, and is taken up by the liver (Altmann et al., 2004; Luo et al., 2020).

Cholesterol is mainly synthesized in the liver and free into the bloodstream as very-low-density lipoproteins (VLDLs). In the bloodstream, the VLDLs are transformed to produce lowdensity lipoproteins (LDLs), transported to peripheral cells by the bloodstream (Ikonen, 2008; Goldstein and Brown, 2009). LDLs enter cells via receptor (LDLR)-mediated endocytosis and are transported to lysosomes where they are hydrolyzed into free cholesterol molecules, which are transported to cellular membranes to carry out its multiple functions (Brown and Goldstein, 1986; Ikonen, 2008; Maxfield and van Meer, 2010; Kuzu et al., 2016).

Mevalonate pathway is tightly regulated by transcriptional and translational mechanisms capable of responding to physiological signals. Cholesterol biosynthesis is regulated by four principal players: (1) sterol regulatory element-binding protein 2 (SREBP2), which acts through a negative feedback mechanism (Sato, 2010), (2) liver X Receptors (LXRs), (3) HMGCR, and (4) squalene epoxidase (SQLE). HMGCR and SQLE are rate-limiting enzymes, which can regulate cholesterol biosynthesis, the reactions they catalyze being energetically expensive. When intracellular ATP levels are low, 5'adenosine monophosphate-activated protein kinase (AMPK) phosphorylates HMGCR inhibiting its function (Loh et al., 2019). Moreover, HMGCR is influenced by the presence of LDL in the medium; in fact, upon LDL starvation, HMGCR activity enhances, while it highly decreases when LDL is added back. On the other hand, once cholesterol has exhausted its function, its surplus is exported via ATP-binding cassette (ABC) subfamily A member 1 (ABCA1) or ABC subfamily G member 1 (ABCG1) to lipid-poor apolipoprotein A-I (ApoA-I), thus generating high-density lipoproteins (HDLs) (Gelissen et al., 2006; Lorenzi et al., 2008; Daniil et al., 2013; Phillips, 2014). The transcription of ABCA1 is upregulated by nuclear LXRs when the intracellular cholesterol level is high (Wang et al., 2008; Ouvrier et al., 2009; Kuzu et al., 2016). The CoA:cholesteryl acyltransferase 1 (ACAT1) transforms excess cholesterol into less toxic compounds, such as cholesteryl esters (CEs), which are stored as lipid droplets and used for the production of the main plasma lipoproteins (chylomicrons, VLDLs, LDLs, and HDLs). HDLs are then transported from peripheral tissues back to the liver and intestine, to recycle or eliminate cholesterol, and to steroidogenic organs, where cholesterol is used to generate steroid hormone (Chang et al., 2009; Luo et al., 2020).

Recent studies on microRNAs (miRNAs), a class of noncoding RNAs, highlighted their role in the cholesterol homeostasis by adjusting some important components of the system (Wagschal et al., 2015). For instance, under low sterol concentration conditions, higher transcription of miR-33a is required to control cholesterol export and HDL metabolism through ABCA1 inhibition (Wagschal et al., 2015). Conversely, miR-223 regulates cholesterol amount by inhibiting its production and ameliorating cholesterol efflux by enhancing ABCA1 levels (Vickers et al., 2014). miRNA-122, present mainly in hepatocytes, when inhibited significantly suppresses blood cholesterol level (Rotllan and Fernandez-Hernando, 2012).



miR-27a has been shown to control HMGCR level either by a posttranslational block or by mRNA degradation (Khan et al., 2020). Strikingly, other miRNAs have been discovered by meta-analyses to be associated with cholesterol–lipoprotein trafficking alterations, such as miR-128-1, miR-148a, miR-130b, and miR-301b. These miRNAs were able to increase circulating cholesterol by controlling the expression level of LDLR and ABCA1 (Wagschal et al., 2015).

These data suggest the participation of miRNAs in the control of cholesterol metabolism, underlining how they could contribute to the alteration of cholesterol levels when dysregulated.

Cholesterol Homeostasis in Normal and Cancer Cells

Considering the importance of cholesterol metabolism, its cellular homeostasis is strictly regulated at every stage: import,

synthesis, export, transport, and esterification (Ikonen, 2008). Sterol regulatory element-binding protein 2 (SREBF2) and LXRs act as key regulators of cholesterol homeostasis (Ikonen, 2008). In the ER, cholesterol itself regulates its homeostasis. Low cholesterol levels induce translocation of SREBP2 to the nucleus where it promotes activation of genes implicated in the biosynthesis (e.g., HMGCR) and uptake (e.g., LDLR) of cholesterol (Ikonen, 2008). High cholesterol levels inhibit the cholesterol synthesis and facilitate its export through the activation of LXRs by oxysterols, oxidized derivatives of cholesterol (Wang et al., 2008; Kuzu et al., 2016). Recent studies on LNCaP prostate cancer cells highlighted an important protective role of LXRs (Pommier et al., 2010; Fu et al., 2014). In fact, the activation of these transcription factors regulating cholesterol homeostasis was able to induce cell cycle arrest and to promote apoptosis (Pommier et al., 2010). The relationship among LXRs, cholesterol, and prostate cancer

highlights that LXRs have good chance to be targeted one day in this tumor.

During cholesterol biosynthesis, depending on the type of tissue, different intermediate sterols are formed, such as cholesteryl ester, oxysterols, bile acids, cholecalciferol/vitamin D, and various steroid hormones. All these sterols have important physiological functions in cells and in tissues (Simons and Ikonen, 2000). Some cholesterol metabolites may also contribute to the progression and metastatization of some types of cancer (Lin et al., 2013; McDonnell et al., 2014; Baek et al., 2017).

Under healthy conditions, the amount of cholesterol is the result of a balance between synthesis, uptake from extracellular milieu, removal of the cholesterol surplus from peripheral tissues, and metabolic conversion (Simons and Ikonen, 2000; **Figure 3**). The alterations of this balance, often caused by unbalanced diets and unhealthy lifestyles (Hu et al., 2012), are known to be associated with cardiovascular diseases and atherosclerosis, but mounting evidence also connects these behaviors to increased cancer risks (Luo et al., 2020).

Over the past few decades, literature data have demonstrated that metabolic alterations represent a hallmark of cancer.

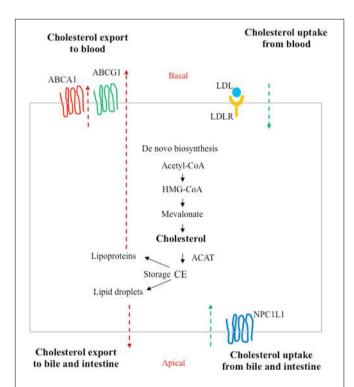
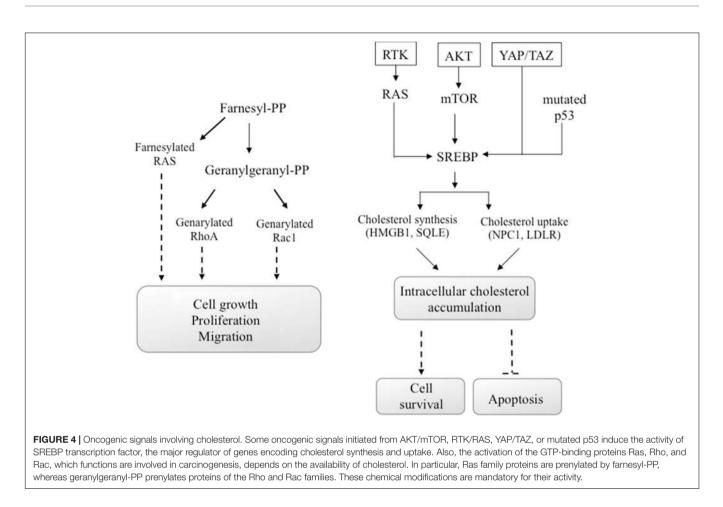


FIGURE 3 | Homeostasis of cholesterol in a polarized cell. In addition to *de novo* biosynthesis, cholesterol carried by low-density lipoprotein (LDLs) in the blood can be taken up by LDL receptors (LDLRs) at the basal surface of the polarized cells (such as enterocytes or hepatocytes). Niemann–Pick type C1-like 1 (NPC1L1) can absorb free cholesterol from dietary sources by enterocytes in the intestine and from bile in the biliary ducts by hepatocytes in the liver. Excess cholesterol is exported to the blood by ATP-binding cassette subfamily A member 1 (ABCA1) or the subfamily G member 1 (ABCG1). Cholesterol can also be converted to cholesteryl ester (CE) by acyl coenzyme A:cholesterol acyltransferase (ACAT) for storage in lipid droplets or for secretion as lipoproteins. Oncogenesis is an intricate process that involves, in addition to reprogramming energy metabolism, also reprogramming of genetic information, signaling mechanisms, and structural components, which are critical for the survival and growth of cancer cells (Hanahan and Weinberg, 2000, 2011). Reprogramming energy metabolism had already been described by Otto Warburg, which highlighted that cancer cells, even in the presence of abundant oxygen, preferably produce ATP (adenosine triphosphate) through the less efficient glycolytic pathway, rather than through oxidative phosphorylation (OXPHOS), thus producing more lactate than normal cells (Warburg, 1925). This metabolic switch of cancer cells, called Warburg effect, is also known as aerobic glycolysis. More recently, attention has been paid to metabolic heterogeneity within the tumor. Particular emphasis has been given to the cells present in the tumor stroma, which would also have the role of metabolically supporting the cancer cells. In this context fits the theory of the reverse Warburg effect according to which the aerobic glycolysis in the cells of the tumor stroma would have the role of providing, in addition to ATP, metabolites for the generation of further ATP through OXPHOS in cancer cells. Moreover, the metabolic coupling between stromal and tumor cell, with the exchange of useful metabolites, would also allow the tumor cell to increase its proliferation and reduce cell death (Xu et al., 2015; Wilde et al., 2017; Benny et al., 2020).

Alteration of energy metabolism is due to enhanced metabolic needs of cancer cells to support the proliferation, migration, and metastatic cancer activities (Warburg, 1956; Silvente-Poirot and Poirot, 2012). However, there are conflicting epidemiological evidence on the cholesterol function in cancer development, while important preclinical data suggested that a dysregulation of cholesterol homeostasis could lead to cancer progression. In particular, new data suggested that cholesterol amount in cells may have a greater role in cancer development than serum cholesterol (Kuzu et al., 2016). Some oncogenic pathways such as PI3K/AKT/mTOR, RTK/RAS, YAP/TAZ, and p53 are also able to regulate cholesterol production in cancer cells (**Figure 4**).

CHOLESTEROL AS COMPONENT OF LIPID RAFTS

Besides the classical role of cholesterol in determining the correct structure, fluidity, and functioning of eukaryotic cell membranes, cholesterol represents the prevalent lipid component of specific plasma membrane microdomains, known as lipid rafts, also contributing to their organization. Indeed, cholesterol accumulates in specialized region of the membrane and, by interacting with sphingolipids, forms lipid rafts (Brown and London, 2000; Pike, 2003, 2006). These membrane regions are heterogeneous and highly dynamic structures, ranging from 10 to 200 nm in size, that selectively recruit and concentrate different membrane proteins (i.e., receptors, adhesion molecules, etc.) and signaling molecules, forming a sort of platform for signal transduction (Simons and Toomre, 2000). Nitric oxide synthase, GPI-anchored proteins, Src-family tyrosine kinases, G-protein- α subunit, protein kinase C, and protein kinase A are typically



localized within lipid rafts (Song et al., 1997; Galbiati et al., 1999; Razani et al., 1999; Patel and Insel, 2009).

Membrane and signaling proteins reside transiently within lipid rafts. Thus, depending on whether the signal transduction needs to be enhanced or damped, they can be recruited or excluded from lipid rafts in a reversible manner. The mechanisms at the base of reversible raft-protein associations are not yet completely understood. Characteristic lipid posttranslational modifications have been described by researchers to be responsible for the recruitment of proteins within lipid rafts. One of the major raft-targeting signals is represented by S-palmitoylation of transmembrane proteins, consisting in palmitate moieties added to cysteine residues through thioester linkage (Resh, 1999, 2004; Petrova et al., 2013). For instance, death receptors such as Fas/CD95 (Chakrabandhu et al., 2007; Feig et al., 2007) and TRAIL receptor 1 (Rossin et al., 2009) are palmitoylated on specific cysteine residues, and this event is essential either for their recruitment within lipid rafts or for an efficient transmission of death signals. In human breast cancer cells, palmitoylation of the surface adhesion receptor CD44 relies on two cysteine residues. This event enhances CD44 affinity for cholesterol-rich lipid rafts, inducing CD44 recruitment into lipid microdomains (Babina et al., 2014), limiting breast cancer cell migration (Donatello et al., 2012). Moreover, proteins can be anchored to plasma membranes through a glycosylphosphatidylinositol (GPI) anchor or they can present a lysine-rich region within their amino acid sequence, which represents an additional signal beside palmitoylation targeting proteins within lipid rafts (Rossin et al., 2009).

Therefore, membrane lipid rafts represent a sort of lipidbased structures that regulate the assembly and functioning of numerous cell signaling, including those related to cancer (Staubach and Hanisch, 2011; Donatello et al., 2012), such as tumor cell growth, adhesion, migration, invasion, and apoptosis (Yang et al., 2014, 2018).

Lipid rafts were also found in mitochondria, ER, in the nuclear membrane, and phagosomes, respectively, where they form a sort of "raft-like microdomain" (Garofalo et al., 2005, 2016; Brunham et al., 2006; Cascianelli et al., 2008; Boslem et al., 2013; Matarrese et al., 2014). These regions, rich in gangliosides, present a content of cholesterol lower as compared with plasma membrane (Garofalo et al., 2015). They are involved in triggering specific events during apoptosis execution, i.e., mitochondria hyperpolarization and depolarization, with consequent release of apoptogenic factors (Garofalo et al., 2005; Scorrano, 2008; Sorice et al., 2009), and regulation of fusion–fission processes (Garofalo et al., 2007; Scorrano, 2008; Sorice et al., 2009; Ciarlo et al., 2010, 2018; Annunziata et al., 2018).

Taking into account all these considerations, the interest in targeting membrane raft cholesterol in cancer cells is increasing. Specific studies might be helpful to better understand the potential use of lipid rafts as innovative target in anticancer therapy.

Raft-Associated Cholesterol Involvement in Migration, Invasion, and Metastatic Processes of Cancer Cells

Mounting evidence of the literature supports a role for lipid rafts in several signaling transduction pathway related to malignancy. Therefore, the interest in studying lipid rafts and the modulation of lipid raft organization is growing. Approaches aimed at disorganizing membrane raft domains were reported by researchers to affect cancer cell proliferation, adhesion, migration, invasiveness, metastatic spread, and apoptosis (Simons and Toomre, 2000; Simons and Ehehalt, 2002; Zidovetzki and Levitan, 2007).

A critical feature of malignancy is represented by the acquisition of a more aggressive phenotype by cancer cells. Because of their malignancy, cancer cells became able to adhere to the extracellular matrix and therefore to migrate and to invade other tissues, leading to metastasis. The plasma membrane certainly represents the first structure of the tumor cell to be involved in the processes of invasion and metastasis. Invadopodia are highly specialized plasma membrane structures of tumor cells, corresponding to podosomes of untransformed cells, able to favor the adhesion and penetration of the tumor cell into the underlying extracellular matrix, stroma, and basement membrane. The formation of invadopodia, structures rich in cytoskeletal elements, adhesion molecules, and degradative enzymes, requires the presence of lipid rafts, which are decisive for the establishment of this invasive and dynamic structure (Nicolson, 2015). Yamaguchi et al. (2009) found that Cav-1, a typical component of caveolar membrane lipid raft structures, represents an essential regulator of invadopodia-mediated human breast cancer cell invasion (Yamaguchi et al., 2009). Disruption of lipid rafts by depletion of membrane cholesterol using methyl- β -cyclodextrin (M β CD) suppressed either invadopodia formation and tumor invasion, thus providing evidence that lipid raft formation was mandatory for invadopodia functioning, at least in human breast cancer (Yang et al., 2016). In bladder cancer cells, Cav-1 overexpression induced epithelial-to-mesenchymal transition that was mediated by PI3K/AKT activation (Liang et al., 2014).

Very interestingly, a reduced expression of Cav-1 was found frequently in human cancer-associated fibroblasts (Martinez-Outschoorn et al., 2014), where it was accompanied by a reduced mitochondrial function and an increased expression of glycolytic enzymes (Asterholm et al., 2012). Some studies have shown that tumor cells, inducing oxidative stress, were responsible for the autophagic degradation of Cav-1 in stromal fibroblast, thus favoring tumor–stroma co-evolution underlying the theory of the reverse Warburg effect (Martinez-Outschoorn et al., 2010).

The dysregulation of flotillin-2, a protein marker of noncaveolar plasma membrane microdomains, has been reported in a variety of tumors. In particular, a recent study related a high expression of flotillin-2 to the incidence of lymph node metastasis and TNM stage in intrahepatic cholangiocarcinoma cell lines (Xu et al., 2020), as also previously observed in a variety of human solid tumors (Liu et al., 2017). High expression of flotillins was also observed in drug-resistant strains of colorectal cancer cells (Ye et al., 2019). Data from Ye and coworkers clearly demonstrated that flotillin silencing using lentivirus-mediated RNAi approach, by inducing destruction of lipid rafts, inhibited the drug resistance of the colorectal cancer cell line HCT-15 (Ye et al., 2019).

Additional findings in prostate cancer cells suggested a potential positive role of membrane cholesterol in modulating the IL-6/STAT3 pathway and chemoattraction (Nakashima et al., 2000; Kim et al., 2004; Akashi et al., 2008; Chinni et al., 2008). The IL6-JAK-STAT3 pathway, known to be an important signaling pathway in prostate cancer, requires intact lipid rafts for sustaining pro-oncogenic signaling. Also in this case, disruption of lipid rafts by MBCD inhibited this pathway thus controlling prostate cancer growth (Dambal et al., 2020). In addition, it has also been reported that hypoxia, a prognostic predictor of poor clinical outcome (Vaupel et al., 2001), was able to increase cholesterol level and recruitment of Notch3 into lipid rafts. Notch3 activity sustained cell proliferation of different hormone-dependent and independent prostate cancer cell lines, and positively correlated with tumor aggressiveness (Danza et al., 2013). In prostate cancer cells, it has also been reported that the interaction of the tetraspanin CD82 with cholesterol in lipid rafts inhibited tumor cell movement and invasion by acting on plasma membrane/cytoskeleton linker ezrin, thus disengaging the plasma membrane-cytoskeleton connection and consequently repressing cell movement (Huang et al., 2020).

Several proteins responsible for development of a more aggressive and metastatic phenotype have been found associated with lipid rafts, including integrins and cell adhesion molecules, ion channels, potassium channels, and MUC1 (Staubach et al., 2009; Haddon and Hugh, 2015), strongly supporting a positive role for lipid rafts in tumor progression.

Several findings reported the localization within lipid rafts of the highly expressed cell surface adhesion receptor CD44, whose lipid raft localization was related to invasion and metastatic processes of cancer cells (Murai, 2015; Mollinedo and Gajate, 2020). In this context, it was reported that M β CD-mediated depletion of cholesterol induced CD44 shedding (Adler, 2011; Murai et al., 2011). Like M β CD, simvastatin, an agent that is able to target cholesterol with consequent disorganization of membrane rafts, also demonstrated to be effective in increasing shedding of CD44 and concomitantly in inhibiting tumor cell migration (Murai et al., 2004, 2006, 2009; Murai, 2015). In the same vein, it has also been shown that other statins induced disruption of lipid rafts, leading to impairment of tumor cell adhesion and migration (Murai et al., 2011; Murai, 2012).

Like other receptors, CD44 has been found into lipid rafts (Murai, 2015) or associated with the actin cytoskeleton through the ezrin protein in non-raft compartments (Martin et al., 2003; Yang et al., 2018). Once CD44 binds ezrin in non-raft compartments, it promotes cancer cell adhesion and migration in hepatocellular carcinoma cells (Yang et al., 2003). Similar results were also reported for breast cancer cells (Donatello et al., 2012).

Thus, CD44–ezrin interaction in non-raft compartments leads to cell migration (Martin et al., 2003; Mollinedo and Gajate, 2020). Taken together, these data point out the importance of lipid rafts in CD44-mediated migration and invasion processes, but the molecular mechanism involved remains to be fully elucidated. Taking into account all these evidences, lipid rafts might therefore represent an important modulator of the invasive and metastatic ability of cancer cells. Accordingly, it has demonstrated that suppressing lipid raft formation using the bioactive natural compound daphnane-type diterpenes from *Daphne genkwa* (GD) strongly impaired human hepatocellular carcinoma cell invasion and migration (Wu et al., 2020).

Raft-Associated Cholesterol in Cancer Cell Survival and Apoptosis

Many studies in the literature evidenced cholesterol-rich rafts as important players regarding cancer cell survival signaling and proliferative pathway. Indeed, it has been reported that survival signal pathways are compartmentalized into lipid rafts where membrane receptors can become in close contact with downstream signaling molecules, inducing the transmission of proliferative signals into the cells. This is the case of the activation of PI3K/AKT-mediated survival signals induced by insulinlike growth factor (Gao and Zhang, 2008, 2009; Gao et al., 2011; Reis-Sobreiro et al., 2013; Mollinedo and Gajate, 2020). In fact, it has been reported that activation of IGF receptors induced the recruitment of several signaling molecules involved in the PI3K/AKT pathway into lipid rafts, favoring activation by phosphorylation of AKT, with subsequent transmission of survival signals into the cells. Disruption of lipid rafts using MBCD or filipin III had demonstrated efficacy in inhibiting the activation of AKT and other survival signaling pathways in human lung adenocarcinoma and human T-cell leukemia, in favor of apoptosis activation (Motoyama et al., 2009).

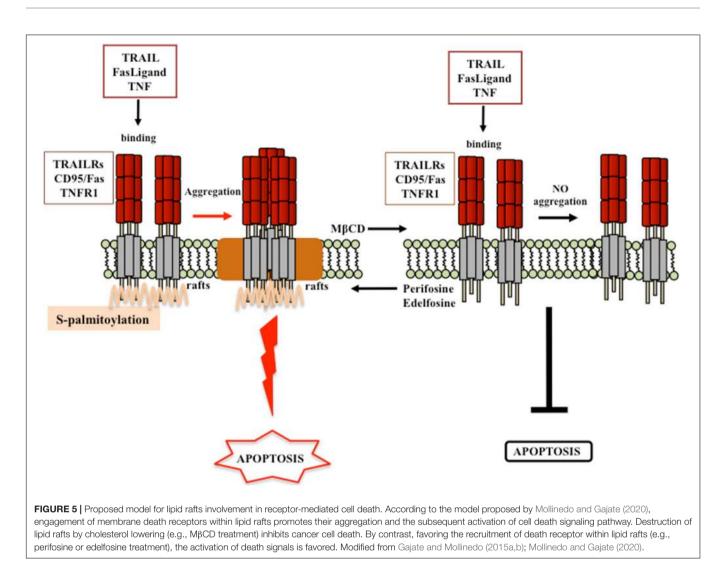
Data obtained in prostate cancer highlighted the importance of the epidermal growth factor (EGF) receptor recruitment into lipid rafts for an efficient transmission of survival signal within the cells (Zhuang et al., 2002; Zidovetzki and Levitan, 2007). Indeed, it has been reported that disruption of membrane rafts using statins, known cholesterol-lowering agents, impaired EGF receptor pathway, favoring apoptosis (Zhuang et al., 2005; Oh et al., 2007; Hryniewicz-Jankowska et al., 2019).

Particular attention has also been paid by researchers on the role of lipid rafts regarding the apoptotic signaling pathway, which could occur following one of the two classical pathways, the extrinsic membrane receptor-mediated or the intrinsic mitochondria-involving pathway. Interestingly, cholesterol has emerged as key modulator of both apoptotic pathways. Like for cancer survival signaling, lipid rafts have been described as scaffold plasma membrane regions, in which death receptors and downstream signaling molecules (FADD, pro-caspase-8/-10) aggregate and cluster in, favoring apoptosis induction and execution (Gajate and Mollinedo, 2001; Scheel-Toellner et al., 2002; Garofalo et al., 2003; Gajate et al., 2004; Malorni et al., 2007; Mollinedo and Gajate, 2010, 2020; Iessi et al., 2020). Findings from the last two decades evidenced that the localization

of death receptors (e.g., CD95/FAS and TRAIL) into these specialized regions is mandatory either for susceptibility to receptor-mediated death signal or for a correct and efficient initiation of the apoptotic signaling (Figure 5; Gajate and Mollinedo, 2001, 2007; Mollinedo et al., 2004; Mollinedo and Gajate, 2006, 2020; Marconi et al., 2013). Indeed, recruitment of the death receptor Fas/CD95 into lipid rafts has been described in several studies (Gajate and Mollinedo, 2001, 2015a; Gajate et al., 2004; Mollinedo and Gajate, 2020). Similarly, TRAIL death receptor 1, but not TRAIL death receptor 2, localized constitutively within the lipid rafts in human cancer B-cell lines, and this event was depicted as mandatory for an efficient death receptor-mediated signaling transduction (Marconi et al., 2013). Actin cytoskeleton plays a peculiar role in ensuring a correct lipid localization of membrane proteins and an efficient cell death signal transmission (Algeciras-Schimnich et al., 2002; Gajate and Mollinedo, 2005). The ERM protein ezrin, known linker between membrane proteins and the actin cytoskeleton, was described as a positive regulator of early steps of Fas receptor signaling in lymphoid T cells (Parlato et al., 2000; Lozupone et al., 2004). Moreover, aggregation of death receptors within lipid rafts, by promoting apoptosis induction, was favored by several agents, i.e., oxaliplatin (Xu et al., 2009), epirubicin (Xu et al., 2011), β-elemene (Xu et al., 2018), bufalin (Yan et al., 2014), resveratrol (Delmas et al., 2004; Reis-Sobreiro et al., 2009), ursodeoxycholic acid (Lim et al., 2011), edelfosine (Gajate and Mollinedo, 2001; Gajate et al., 2004; Lim et al., 2016; Mollinedo and Gajate, 2020), and perifosine (Gajate and Mollinedo, 2007; Mollinedo and Gajate, 2020). On the contrary, agents able to disaggregate lipid rafts through membrane cholesterol depletion induced resistance to apoptotic cell death (Delmas et al., 2004; Gajate and Mollinedo, 2007, 2014, 2015b; Gajate et al., 2009; Marconi et al., 2013). For instance, resistance to TRAIL may be promoted by MβCD (Delmas et al., 2004; Marconi et al., 2013), whereas enhancement of TRAIL-induced apoptosis could be observed following perifosine treatment, which favored the recruitment of DRs into lipid rafts (Gajate and Mollinedo, 2007; Marconi et al., 2013). Edelfosine-mediated upregulation of TRAIL-R2, and its subsequent increased recruitment within lipid rafts, turned resistant gastric cancer cells back to a TRAIL-sensitive phenotype (Lim et al., 2016). Enhanced Fas/CD95 recruitment and clustering within lipid rafts, and consequent increased Fas/CD95-mediated apoptosis, was observed after edelfosine treatment in several hematological malignancies (Gajate and Mollinedo, 2001, 2007; Gajate et al., 2004, 2009; Mollinedo and Gajate, 2006, 2015; Mollinedo et al., 2010a,b). As a consequence, depletion of membrane cholesterol by leading to disruption of lipid rafts strongly impaired Fas/CD95 death signaling (Gajate and Mollinedo, 2007, 2014, 2015b; Gajate et al., 2009).

TARGETING CHOLESTEROL PATHWAYS AS CANCER THERAPY

Cancer has often been coupled with modifications in lipid metabolism, in particular in cholesterol metabolism. Alterations in cholesterol homeostasis have been identified in a large number



of cancers. Both circulating and membrane associated cholesterol would appear to play a key role in the regulation of all cancer signaling pathways.

According with this, it has been suggested that several anticancer drugs may have anti-proliferative function, limiting the content or production of cholesterol. For instance, it has been shown that doxorubicin caused the death of cancer cells by promoting a reduction in HMGCR levels and causing a decrease in cholesterol content (Yun et al., 2019). Other data displayed that tamoxifen modulated cholesterol metabolism in breast cancer cells (Segala et al., 2013). Many natural compounds, including terpenoids, green tea, garlic extract, and curcumin, effective in cancer prevention and therapy, were identified to target cholesterol homeostasis in cancer cells (Mok and Lee, 2020). Thus, these natural compounds might be useful in regulating cholesterol homeostasis as adjuvants to complement current anticancer therapies.

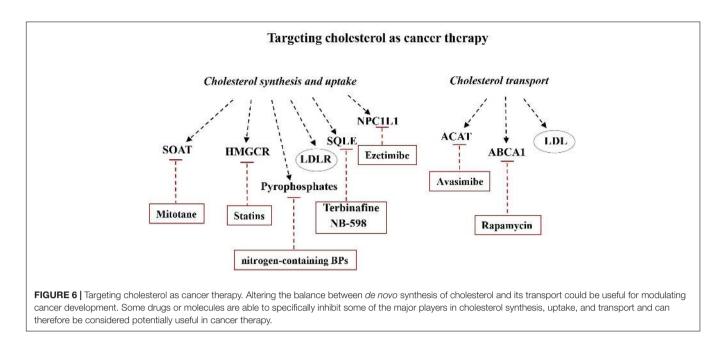
The cholesterol synthesis pathway consists of a number of enzymes, each of which can represent a possible target to disturb the mevalonate pathway in cancer cells. Many studies suggested that pharmacological modulation of intracellular cholesterol balance could be effective in controlling cancer progression (Figure 6).

Targeting Cholesterol Synthesis and Uptake

Cellular cholesterol levels are the result of a balance among the processes of production, transport, and storage. The biochemical events and the main players that in cells are underlying synthesis and trafficking cholesterol, and which, therefore, can represent potential therapeutic targets, are briefly mentioned in the following section.

HMGCR

The rate-limiting reduction of HMG-CoA to mevalonate is an important regulatory step in cholesterol synthesis. HMG-CoA is highly regulated at transcriptional, translational, and post-translational planes (Goldstein and Brown, 1990). The HMGCR is an ER-localized glycoprotein that converts HMG-CoA to mevalonate. HMGCR is genetically activated by nSREBP2 when sterol concentrations are low. Sterols, mainly oxysterols, and two members of the vitamin E family can induce its



degradation (Song et al., 2005; Chen et al., 2019). Moreover, ubiquitination and proteasome-mediated degradation are also known mechanisms regulating HMGCR. HMGCR changes from a phosphorylated to a dephosphorylated form. Phosphorylation inhibits HMGCR activity. AMP-activated protein kinase (AMPK) is the main actor for hepatic HMGCR phosphorylation, which in turn results in hampering of cholesterol production when low intracellular ATP levels are perceived. Conversely, HMGCRmediated cholesterol biosynthesis is triggered when AMPK suppression occurs (Zhang et al., 2015; Luo et al., 2017; Soto-Acosta et al., 2017). The interaction between HMGCR and the insulin-induced gene protein (INSIG-1 or -2) at ER membrane is pivotal to HMGCR ubiquitination and degradation (Gong et al., 2006). Overexpression of HMGCR has been well documented in gastric cancer cells, when it promoted growth and migration, while HMGCR knockdown inhibited growth, migration, and tumorigenesis (Chushi et al., 2016; Yang et al., 2020). An upregulation of HMGCR was also observed in glioblastoma and prostate cancer (Qiu et al., 2016; Longo et al., 2019).

Statins were first used in the treatment of dyslipidemic disorders as therapeutic target to limiting production of mevalonate. In fact, statins are competitive inhibitors of HMGCR, and act by inhibiting the mevalonate pathway and reducing the end-product amounts such as cholesterol, isoprenoids, ubiquinone, and isopentenyladenine (Clendening et al., 2010; Thurnher et al., 2012; Cruz et al., 2013). Recently, a role in anti-cancer treatments has been proposed for statins (Farwell et al., 2008). Literature data showed how the use of statins, alone or in combination with chemotherapeutic agents, could have anti-tumor effects and reduce the onset of multidrug resistance (Mollinedo et al., 2010a; Cruz et al., 2013; Warita et al., 2014; Kuzu et al., 2016). As they have been shown very effective against mesenchymal-like cancer cells, Warita et al. have hypothesized their possible use during the transition from epithelial to mesenchymal phenotype of

metastatic cells (Warita et al., 2014). It was reported that statin supply is effective in reducing the occurrence of a wide range of cancers (e.g., liver, gastric, colorectal, pancreatic, and prostate) and cancer-related mortality ratios (Gbelcova et al., 2008; Ginestier et al., 2012; Nielsen et al., 2013; Benakanakere et al., 2014; Mok and Lee, 2020).

SREBP

Lipid homeostasis is controlled by membrane-bound transcription factors. These include the family of sterol regulatory element-binding proteins (SREBPs), which consist of three isoforms (SREBP1a, SREBP1c, and SREBP2). Among them, SREBP2 modulates more than 30 genes specific to the synthesis and uptake of cholesterol, fatty acids, triglycerides, and phospholipids (Horton et al., 2002; Luo et al., 2020). SREBPs are inactive when high cholesterol amounts are sensed and bound to SREBP cleavage activating protein (SCAP) and insulin-induced genes (INSIGs). Under cholesterol depletion conditions, feedback responses determine the dissociation of INSIGs from the SREBP2/SCAP complex, which move from the ER toward the Golgi apparatus. Subsequently, SREBP2 is processed and addressed to the nucleus. Nuclear SREBP2 (nSREBP2) induces the transcription of target genes, such as HMGCR and SQLE, by the direct interaction with the sterol regulatory element (SRE) motifs in their promoters (Brown et al., 2018). The upregulation of SREBP-activated downstream genes has been observed in a plethora of cancers, including glioblastoma (Guo et al., 2009; Yamauchi and Rogers, 2018), prostate cancer (Ettinger et al., 2004), breast cancer (Pitroda et al., 2009), and melanoma (Yamauchi et al., 2011; Kuzu et al., 2016), and was often found associated with poor prognosis/survival. The increase of cholesterol synthesis is mediated by the activation of PI3K-AKT-mTORC1 signaling (Porstmann et al., 2008; Duvel et al., 2010). In fact, PI3K-AKT pathway induced expression of SREBP genes and increased the stability of nuclear SREBPs by preventing their proteasomal degradation. Similarly, the transcription of nSREBP2 can also be induced by the mammalian target of rapamycin complex 1 (mTORC1) via inhibition of lipin-1 (Peterson et al., 2011; Mok and Lee, 2020). Results from preclinical studies showed that the AKT/mTORC1/SREBP pathway takes part in cancer cell growth by inducing biosynthesis of cholesterol (Porstmann et al., 2008). Finally, direct inhibition of SREBP2 (e.g., using fatostatin), resulting in a decrease in cholesterol content, may represent an interesting target in the treatment of a wide range of cancers (Gholkar et al., 2016; Gao et al., 2018).

SQLE

Squalene epoxidase catalyzes the conversion of squalene to (S)-2,3-epoxysqualene (SQLE) and shares with HMGCR the low rates of catalysis in the cholesterol biosynthesis. Like HMGCR, SQLE abundance and activity are regulated at gene and protein levels. The *SQLE* gene responds to sterols via SREBP2 (Nagai et al., 2002; Howe et al., 2017; Luo et al., 2020) and the protein can be degraded in the presence of cholesterol by ubiquitination. HMGCR and SQLE are both transcriptionally controlled by nSREBP2, but they can lead to cholesterol biosynthesis independently of each other. In the presence of an excess of cholesterol, squalene epoxidase is degraded by the E3 ubiquitin ligase MARCH6 (Yoshioka et al., 2020).

Robust scientific evidence demonstrated that SQLE can be considered an important oncogene and, consequently, may represent a possible target in cancer therapy (Cirmena et al., 2018). In particular, in vitro and in vivo studies have shown that SQLE could promote tumor cell proliferation and migration, and that treatment with its inhibitors (i.e., terbinafine or NB-598) could induce cancer cell demise (Chua et al., 2018). Moreover, a further study on nasopharyngeal carcinoma (NPC) linked the oncogenic effect of SQLE to the cholesteryl ester accumulation, which promoted NPC cell proliferation by activating the PI3K/AKT pathway (Li et al., 2020). In the same vein, in hepatocellular carcinoma (HCC), the overexpression of SQLE was demonstrated to promote cell proliferation and migration by ERK signaling, while downregulation of SQLE was able to inhibit the tumorigenicity of HCC cells (Sui et al., 2015). In addition, SQLE overexpression seem to represent a negative prognostic factor both in breast and prostate cancers (Stopsack et al., 2016; Brown et al., 2019).

NPC1L1

Niemann–Pick type C1-like 1 is the principal responsible for cholesterol absorption from dietary sources. It is a membrane protein expressed on the apical surface of enterocytes and on the membrane of bile canaliculi of human hepatocytes (Altmann et al., 2004). Under normal growth conditions, NPC1L1 is present primarily in the endocytic recycling compartment and translocate rapidly to the plasma membrane upon cholesterol depletion (Yu et al., 2006; Ge et al., 2008). Supplying of cholesterol triggers the inward transport of NPC1L1 together with cholesterol from the plasma membrane to the endocytic recycling compartment (Ge et al., 2008). NPC1L1 can be sent back to the plasma membrane for recycling. This process

requires, in addition to other proteins, the small GTPase CDC42 and actin filaments (Zang et al., 2001; Xie et al., 2011; Luo et al., 2020). The human NPC1L1 gene is activated by SREBP2. Moreover, NPC1L1 protein amount can be modulated by degradation (Malhotra et al., 2019).

Recent literature data obtained both *in vivo* and *in vitro* indicated that NPC1L1 could represent a highly effective therapeutic target for treating pancreatic ductal adenocarcinoma (PDAC) because aberrant cholesterol uptake has been suggested to play a role in the proliferation and survival of pancreatic cancer cells (Guillaumond et al., 2015). Accordingly, the use of ezetimibe, an inhibitor of NPC1L1 clinically used for the hypercholesterolemia treatment, significantly reduced the survival capacity of PDAC cells (Nicolle et al., 2017) and tumorassociated blood vessel development (Solomon et al., 2009).

LDLR

Low-density lipoprotein receptor is a cell surface glycoprotein, which causes cellular absorption of cholesterol carried by the bloodstream. LDLR is the transcriptional target of SREBP2 and thyroid hormones (Lopez et al., 2007). LDLR binds circulating LDLs to clathrin-coated vesicles, which then accede the endocytic pathway (Goldstein and Brown, 2009). The endosome acidity induces conformational modifications that allow detaching LDLR from LDLs (Rudenko et al., 2002). LDLR is then redirected to the cell surface for further endocytic cycles (Bartuzi et al., 2016; Fedoseienko et al., 2018) or it is addressed to lysosomes for degradation. After endocytosis, lysosomal lipase hydrolyzes LDL-carried CE and produces cholesterol, which is finally exported by the coordinate actions of NPC2, NPC1, and LAMP2 (Kwon et al., 2009; Li and Pfeffer, 2016). Inhibition of the LDLR at any level (e.g., biosynthesis, membrane localization, internalization, recycling, and degradation) can affect its availability or functionality and, consequently, LDL removal. LXRs bind to promoter of a LDLR inhibitor (IDOL) and upregulate its expression (Zelcer et al., 2009). Accordingly, using synthetic LXR agonists decreased LDLR abundance and limited LDL uptake in cultured cells and livers of non-human primates (Zelcer et al., 2009; Luo et al., 2020). The increased PCSK9 binding with LDLR prevents LDLR recycling and promotes its degradation in lysosomes (Zhang et al., 2008; Chang et al., 2009; Luo et al., 2020). Several studies performed by using LXR agonists showed that cholesterol levels of glioblastoma cells decreased after stimulation of LXRs. The cholesterol decrement is due to enhanced cholesterol export and reduction of cholesterol import (Guo et al., 2011; Villa et al., 2016). LXR agonists also suppressed metastases of melanoma and prolonged animal survival in several in vivo models (Pencheva et al., 2014; Gabitova et al., 2015; Yamauchi and Rogers, 2018). In human cancers, the upregulation of PI3K-AKTmTORC1 signaling promoted cellular cholesterol accumulation trough stimulation of LDLR-mediated cholesterol import. In glioblastoma, the overexpression of LDLR was caused by AKT, and pharmacological inhibition of LDLR actually favored cancer cell death (Guo et al., 2011; Kuzu et al., 2016). As mentioned previously, the increase of oxysterol, a by-product of cholesterol biosynthesis, was able to activate LXRs, which have been proposed as the rapeutic targets in multiple cancer types (Lin et al., 2016; Komati et al., 2017).

SOAT

Sterol-o-acyltransferase (SOAT) is a protein of the rough surface ER responsible for cholesteryl ester (CE) production from cholesterol and long-chain fatty acids. SOAT plays an important function by regulating cellular cholesterol storage and free cholesterol content (Pramfalk et al., 2005). Because SOAT has a key role in regulating cellular cholesterol metabolism, it may represent a potential target for treatment of different sickness, including cancer (Rogers et al., 2015; Yang et al., 2020). For instance, it is well known that cholesterol esterification and lipid droplet formation are typical features of glioblastoma (GBM). In fact, accumulation of lipid droplets in the tumor in patients with glioma correlated with GBM progression and poor survival. Genetic inhibition of SOAT1 to block cholesterol esterification was observed to suppress GBM growth and prolong the survival in xenograft models (Geng et al., 2016). In addition, literature data also demonstrated that treating adrenocortical carcinoma (ACC) with mitotane reduced cancer cell survival by inhibiting SOAT1 activity (Sbiera et al., 2015). Very recently, Jiang and colleagues also highlighted that the high expression of SOAT1 was associated with a poor prognosis in hepatocellular carcinoma, whereas its inhibition suppressed cell proliferation and migration (Jiang et al., 2019).

Pyrophosphates

Different enzymes involved in the biosynthesis of cholesterol could be targeted by anticancer drugs. Farnesyl transferase (FTase) and geranylgeranyl transferases (GGTase) are enzymes that catalyzed the production of farnesyl pyrophosphate (FPP) and geranylgeranyl pyrophosphate (GGPP), both involved in cellular protein prenylation (Berndt et al., 2011; Sorrentino et al., 2014; Mok and Lee, 2020). The prenylation method consists in covalent bonds of hydrophobic molecules (both the C-15 isoprene farnesyl and the C-20 isoprene geranylgeranyl groups) to the C-terminal portion of different proteins, including the small GTP-binding proteins (sGTPases). sGTPases represent the greater and better described class of prenylated proteins. Each member of the Ras, Rho, and Rab GTPase families, many of which are involved in tumorigenesis, requires the structural alteration given by prenylation to bind to cell surface. Such posttranslational modifications and activation of GTP-binding proteins play a key function in most significant cellular processes. Inhibition of the mevalonate pathway reducing the isoprenylation of sGTPases could bring tumor cell death (Szajnman et al., 2005). Because the administration of GGPP and FPP prevented cancer cell death, it has been hypothesized that GGPP and FPP were necessary for the survival of cancer cells. It has been shown that bisphosphonates (BP) were able to block FPP synthase during the production of mevalonate, thus decreasing the formation of FPP and GGPP (Szajnman et al., 2005). Although BPs blocking osteoclast-mediated bone resorption are currently considered the most significant pharmaceutical treatment for metabolic bone disease, recently nitrogen-containing

BPs have aroused interest in their antitumor properties (Nooh et al., 2017).

Steroid Hormones

Cholesterol is the precursor molecule of five different groups of steroid hormones synthesized by specific organs, the adrenal cortex and the gonads. Steroid hormones are widely known to control growth and differentiation. Moreover, steroid hormones are closely linked with the progression of ovarian, breast, and prostate cancers (Lukanova and Kaaks, 2005; Llaverias et al., 2011; Pelton et al., 2012; Murai, 2015; Radisauskas et al., 2016). According with this, inhibition of estrogen or androgen signaling represents the most used strategy in the treatment of these hormone-dependent tumors (Huang and Tindall, 2002; Lu-Yao et al., 2014; Fan et al., 2015; Watson et al., 2015; Simigdala et al., 2016; Shah et al., 2020). With regard to prostate cancer, although almost all patients with metastatic disease initially respond to androgen-ablation therapies, unfortunately the majority of patients progress to a castrate-resistant stage (Huang and Tindall, 2002; Lu-Yao et al., 2014; Watson et al., 2015; Shah et al., 2020). There is accumulating evidence suggesting that cholesterol, as a precursor of steroid hormones, effects on breast and prostate cancers. In this regard, it was observed that in vitro treatment of prostate and breast cancer cells with mevalonate was able to induce a high proliferation rate (Mokarram et al., 2017).

Targeting Cholesterol Transport

To keep the cellular cholesterol levels constant, it is necessary that the excess cholesterol is carried out of the cell, and this can take place through active export and passive diffusion. Active export requires energy and involves several ABC transporter proteins using ATP. Passive diffusion is caused by a difference in cholesterol content between the cell membrane and cholesterol acceptors such as HDL.

LDLs/HDLs

Cholesterol is a highly insoluble molecule that is transported in the bloodstream via lipoproteins LDLs and HDLs. Cells can acquire exogenous cholesterol from LDLs that carry abundant cholesterol esters. LDL initially binds to the LDLR receptor on the plasma membrane and is subsequently internalized by clathrin vesicles (Brown and Goldstein, 1986). In LDL, the lipase activity degrades esterified cholesterol producing free cholesterol, which is internalized by the plasma membrane and reaches the endosome/lysosome (Sugii et al., 2003; Yamauchi et al., 2007; Yamauchi and Rogers, 2018). ApoA-I is one of the best known HDL apolipoproteins, capable of binding cholesterol and phospholipids. HDLs bind and carry excess cholesterol from peripheral tissues to the liver where cholesterol is transformed into bile acids for excretion (reverse cholesterol transport) (Yamauchi and Rogers, 2018). Lipoproteins are involved in the elaboration and transport of dietary cholesterol to tissues and aid in keeping a constant level of cholesterol by removing its excess to carry it to the liver. Therefore, lipoproteins could be involved in tumor progression by promoting the availability of cholesterol to tumor cells (Silvente-Poirot and Poirot, 2012; Cruz et al., 2013).

ABCA1

ATP-binding cassette transporters are drug efflux pumps that can cause multidrug resistance and tumor treatment failure (Deeley et al., 2006; Borel et al., 2012). Regarding ABCA1, it is responsible for the bidirectional transport of cholesterol across the cell surface, regulating its availability within the cell. In addition, it mediates the export of cholesterol and phospholipids in ApoA-I for HDL assembly (Oram and Heinecke, 2005). Some research works with Abca1 knockout mice have shown that both hepatic and intestinal ABCA1 concur to the production of total plasma HDL, although to varying degrees (Timmins et al., 2005; Brunham et al., 2006). ABCA1 localizes both at the plasma membrane, where it mediates the assembly of nascent HDLs (Denis et al., 2008), and at the endocytic compartments, where it is degraded or recycled from the plasmatic membrane (Lu et al., 2008). An excess of ABCA1 generates deformation sites in the plasma membrane, where apoA-I binds (Vedhachalam et al., 2007). This could be a result of the disturbing action of ABCA1 on lipid raft organization (Landry et al., 2006). In contrast, ABCA1 deficiency has been found to increase lipid rafts and cholesterol contents in the plasma membrane (Koseki et al., 2007; Zhu et al., 2008; Yamauchi et al., 2015). ABCA1 is closely regulated both at gene and protein levels. LXRs are mainly responsible for activating the ABCA1 gene (Costet et al., 2000; Venkateswaran et al., 2000). Accordingly, data obtained in vitro by using LXR agonists revealed that the activation of LXR reduced intracellular cholesterol amount by increasing its exportation and decreasing its absorption therefore determining a growth inhibition of glioblastoma cells (Guo et al., 2011; Villa et al., 2016). Moreover, LXR agonists suppressed melanoma metastases and prolonged animal survival in several in vivo models (Pencheva et al., 2014; Gabitova et al., 2015; Yamauchi and Rogers, 2018).

On the other hand, it was also reported the PI3K/AKT signaling, through the inhibition of ABCA1 mTORC1-dependent pathway, resulted in an increase of intracellular level of cholesterol (Porstmann et al., 2008; Dong et al., 2014). Rapamycin, the mTORC1 inhibitor, repressed the cholesterol export due to the action of ABCA1 (Dong et al., 2014).

Finally, the regulation of *ABCA1* gene was also modulated by some miRNAs, including miR-33a/b and miR-148 (Najafi-Shoushtari et al., 2010; Rayner et al., 2011; Feinberg and Moore, 2016). These miRNAs suppressed *ABCA1* gene expression and reduced plasma HDL levels in mice and non-human primates (Najafi-Shoushtari et al., 2010; Yamauchi and Rogers, 2018).

ACAT

Acetyl-CoA acetyltransferase 1 and 2 (ACAT1 and ACAT2, respectively) play an important role in lipid metabolism. After esterification into CEs by ACAT1, surplus cholesterol can be stored as lipid droplets (Chang et al., 2009). ACAT1 overexpression and CE accumulation were found to play pro-tumor functions. In fact, it was observed that ACAT1 induced an excessive storage of cholesterol in lipid droplets in prostate (Yue et al., 2014), pancreatic cancers (Li et al., 2016), and glioblastoma (Geng et al., 2016). According with this, ACAT1 inhibition was found to reduce migration of breast cancer cells (Antalis et al., 2010) and decrease prostate cancer

progression (Saraon et al., 2014). Moreover, ACAT1 inactivation determined abundant ER cholesterol content, which is acting on SREBP-1 reduced cholesterol synthesis and uptake (Yue et al., 2014). Recently, pharmacological or genetic ACAT1 inhibition has been associated with attenuated cancer cell proliferation (Yang et al., 2020).

In light of all this, ACAT1 can be considered a possible therapeutic target for the treatment of some neoplasms linked to the abundant presence of cholesterol esters. The use of avasimibe, an effective inhibitor of ACAT1, remarkably decreased cholesteryl esters stocked in lipid droplets and enhanced free cholesterol amount into the cell in human prostate, pancreatic, lung, glioblastoma, and colon cancer cells leading to inhibition of proliferation, cell cycle arrest, and apoptosis (Yang et al., 2020).

CONCLUSION

As the metabolic needs of cancer cells appear significantly different from non-cancer cells, the metabolic pathways could represent potential innovative targets for a more selective anticancer therapy.

The analysis of literature data confirmed a fundamental role of cholesterol and its metabolism in many and different human pathologies, including atherosclerosis, Alzheimer's disease, and other neurodegenerative disorders, and mounting evidence connected cholesterol homeostasis disorder and cancer. Moreover, dysregulation of the mevalonate pathway has been found to promote cell transformation, thus suggesting the role of oncogene for *HMG-CoA reductase* (Clendening et al., 2010). All of this makes cholesterol a potential attractive pharmacological target also in oncology context. Accordingly, although quite controversial, preclinical and epidemiological data seem to show an antitumor activity of statins, at least against some types of cancer (Solomon and Freeman, 2008).

At present, several drugs targeting cholesterol metabolism besides statins are clinically used to inhibit cholesterol production and increase LDL absorption, to hinder NPC1L1 absorption of dietary cholesterol and LDLR degradation, and to reduce plasma LDLs. For instance, the cholesterol uptake-blocking agent ezetimibe, by decreasing circulating cholesterol levels, has been found to reduce prostate tumor growth (Solomon et al., 2009).

Moreover, an increasing number of preclinical and clinical studies are underway to define as new potential therapeutic target for cancer therapy SREBP and its regulators LXRs, the transporters ABCs, and ACATs.

Increasing evidence showed that also cholesterol-rich raft membrane domains could represent a promising target in cancer therapy (Mollinedo and Gajate, 2006, 2020; Mollinedo et al., 2010a). Indeed, acting as scaffolds for signaling pathways in the cell membrane, lipid rafts promote and facilitate the triggering of both survival and apoptosis by recruiting signaling molecules and death receptors. In this regard, understanding how the different rafts that promote cell survival or death are temporarily formed and disrupted may be an important goal to understand if and how lipid rafts could represent a specific target in cancer therapy.

Finally, it is important to underline the key role played by the intestinal microbiota in the reduction and elimination of cholesterol, thus contributing significantly to its homeostasis (Zimber et al., 2000; Antharam et al., 2016). An alteration of the bacterial flora could constitute a further risk factor, especially for those tumors particularly dependent on cholesterol homeostasis. Thus, acting on microbiota might also represent a further possibility of therapeutic intervention in the treatment and/or prevention of certain types of cancer.

AUTHOR CONTRIBUTIONS

RV contributed to conceive the idea, drafted the article, and drew figures relative to cholesterol metabolism. EI

REFERENCES

- Adler, E.M. (2011). Lost without a raft. *Sci. Sign.* 4:c24. doi: 10.1126/scisignal. 4157ec24 s
- Akashi, T., Koizumi, K., Tsuneyama, K., Saiki, I., Takano, Y., and Fuse, H. (2008). Chemokine receptor CXCR4 expression and prognosis in patients with metastatic prostate cancer. *Cancer Sci.* 99, 539–542. doi: 10.1111/j.1349-7006. 2007.00712.x
- Algeciras-Schimnich, A., Shen, L., Barnhart, B.C., Murmann, A.E., Burkhardt, J.K., and Peter, M.E. (2002). Molecular ordering of the initial signaling events of CD95. *Mol. Cell. Biol.* 22, 207–220. doi: 10.1128/mcb.22.1.207-220.2002
- Altmann, S.W., Davis, H.R., Jr., Zhu, L.J., Yao, X., Hoos, L.M., Tetzloff, G., et al. (2004). Niemann-pick C1 like 1 protein is critical for intestinal cholesterol absorption. *Science* 303, 1201–1204. doi: 10.1126/science.1093131
- Annunziata, I., Sano, R., and d'Azzo, A. (2018). Mitochondria-associated ER membranes (MAMs) and lysosomal storage diseases. *Cell. Death Dis.* 9:328. doi: 10.1038/s41419-017-0025-4
- Antalis, C.J., Arnold, T., Rasool, T., Lee, B., Buhman, K.K., and Siddiqui, R.A. (2010). High ACAT1 expression in estrogen receptor negative basal-like breast cancer cells is associated with LDL-induced proliferation. *Breast Cancer Res. Treat* 122, 661–670. doi: 10.1007/s10549-009-0594-8
- Antharam, V.C., McEwen, D.C., Garrett, T.J., Dossey, A.T., Li, E.C., Kozlov, A.N., et al. (2016). An integrated metabolomic and microbiome analysis identified specific gut microbiota associated with fecal cholesterol and coprostanol in clostridium difficile infection. *PLoS One* 11:e0148824. doi: 10.1371/journal. pone.0148824
- Asano, K., Kubo, M., Yonemoto, K., Doi, Y., Ninomiya, T., Tanizaki, Y., et al. (2008). Impact of serum total cholesterol on the incidence of gastric cancer in a population-based prospective study: the hisayama study. *Int. J. Cancer* 122, 909–914. doi: 10.1002/ijc.23191
- Asterholm, I.W., Mundy, D.I., Weng, J., Anderson, R.G., and Scherer, P.E. (2012). Altered mitochondrial function and metabolic inflexibility associated with loss of caveolin-1. *Cell. Metab.* 15, 171–185. doi: 10.1016/j.cmet.2012. 01.004
- Babina, I.S., McSherry, E.A., Donatello, S., Hill, A.D., and Hopkins, A.M. (2014). A novel mechanism of regulating breast cancer cell migration via palmitoylationdependent alterations in the lipid raft affiliation of CD44. *Breast Cancer Res.* 16:R19. doi: 10.1186/bcr3614
- Baek, A.E., Yu, Y.A., He, S., Wardell, S.E., Chang, C.Y., Kwon, S., et al. (2017). The cholesterol metabolite 27 hydroxycholesterol facilitates breast cancer metastasis through its actions on immune cells. *Nat. Commun.* 8:864. doi: 10.1038/s41467-017-00910-z
- Bartuzi, P., Billadeau, D.D., Favier, R., Rong, S., Dekker, D., Fedoseienko, A., et al. (2016). CCC- and WASH-mediated endosomal sorting of LDLR is required for normal clearance of circulating LDL. *Nat. Commun.* 7:10961. doi: 10.1038/ ncomms10961
- Benakanakere, I., Johnson, T., Sleightholm, R., Villeda, V., Arya, M., Bobba, R., et al. (2014). Targeting cholesterol synthesis increases chemoimmunosensitivity in chronic lymphocytic leukemia cells. *Exp. Hematol. Oncol.* 3:24. doi: 10.1186/2162-3619-3-24

contributed to design and to draft the article and drew figures relative to lipid rafts. PM conceived the idea, draft and revised the article, coordinated the activity, and concurred to the final version of the review. All authors contributed to the article and approved the submitted version.

FUNDING

The authors gratefully acknowledge the financial support obtained from the Italian Association for Cancer Research (18526 to PM) and Peretti Foundation (Grant number NaEPF 2019-042).

- Benny, S., Mishra, R., Manojkumar, M.K., and Aneesh, T.P. (2020). From warburg effect to reverse warburg effect; the new horizons of anti-cancer therapy. *Med. Hypotheses* 144:110216. doi: 10.1016/j.mehy.2020.110216
- Berndt, N., Hamilton, A.D., and Sebti, S.M. (2011). Targeting protein prenylation for cancer therapy. *Nat. Rev. Cancer* 11, 775–791. doi: 10.1038/nrc3151
- Borel, F., Han, R., Visser, A., Petry, H., van Deventer, S.J., Jansen, P.L., et al. (2012). Adenosine triphosphate-binding cassette transporter genes up-regulation in untreated hepatocellular carcinoma is mediated by cellular microRNAs. *Hepatology* 55, 821–832. doi: 10.1002/hep.24682
- Boslem, E., Weir, J.M., MacIntosh, G., Sue, N., Cantley, J., Meikle, P.J., et al. (2013). Alteration of endoplasmic reticulum lipid rafts contributes to lipotoxicity in pancreatic beta-cells. J. Biol. Chem. 288, 26569–26582. doi: 10.1074/jbc.M113. 489310
- Brown, A.J., Chua, N.K., and Yan, N. (2019). The shape of human squalene epoxidase expands the arsenal against cancer. *Nat. Commun.* 10:888. doi: 10. 1038/s41467-019-08866-y
- Brown, D.A., and London, E. (2000). Structure and function of sphingolipid- and cholesterol-rich membrane rafts. J. Biol. Chem. 275, 17221–17224. doi: 10.1074/ jbc.R000005200
- Brown, M.S., and Goldstein, J.L. (1986). A receptor-mediated pathway for cholesterol homeostasis. *Science* 232(4746), 34–47. doi: 10.1126/science.351 3311
- Brown, M.S., Radhakrishnan, A., and Goldstein, J.L. (2018). Retrospective on cholesterol homeostasis: the central role of scap. Annu. Rev. Biochem. 87, 783–807. doi: 10.1146/annurev-biochem-062917-011852
- Brunham, L.R., Kruit, J.K., Iqbal, J., Fievet, C., Timmins, J.M., Pape, T.D., et al. (2006). Intestinal ABCA1 directly contributes to HDL biogenesis in vivo. J. Clin. Invest. 116, 1052–1062. doi: 10.1172/JCI27352
- Cascianelli, G., Villani, M., Tosti, M., Marini, F., Bartoccini, E., Magni, M.V., et al. (2008). Lipid microdomains in cell nucleus. *Mol. Biol. Cell.* 19, 5289–5295. doi: 10.1091/mbc.E08-05-0517
- Chakrabandhu, K., Herincs, Z., Huault, S., Dost, B., Peng, L., Conchonaud, F., et al. (2007). Palmitoylation is required for efficient Fas cell death signaling. *EMBO J.* 26, 209–220. doi: 10.1038/sj.emboj.7601456
- Chang, T.Y., Li, B.L., Chang, C.C., and Urano, Y. (2009). Acyl-coenzyme a:cholesterol acyltransferases. *Am. J. Physiol. Endocrinol. Metab.* 297, E1–E9. doi: 10.1152/ajpendo.90926.2008
- Chen, L., Ma, M.Y., Sun, M., Jiang, L.Y., Zhao, X.T., Fang, X.X., et al. (2019). Endogenous sterol intermediates of the mevalonate pathway regulate HMGCR degradation and SREBP-2 processing. *J. Lipid Res.* 60, 1765–1775. doi: 10.1194/ jlr.RA119000201
- Chinni, S.R., Yamamoto, H., Dong, Z., Sabbota, A., Bonfil, R.D., and Cher, M.L. (2008). CXCL12/CXCR4 transactivates HER2 in lipid rafts of prostate cancer cells and promotes growth of metastatic deposits in bone. *Mol. Cancer Res.* 6, 446–457. doi: 10.1158/1541-7786.MCR-07-0117
- Chua, N.K., Coates, H.W., and Brown, A.J. (2018). Cholesterol, cancer, and rebooting a treatment for athlete's foot. *Sci. Transl. Med.* 10:eaat3741. doi: 10.1126/scitranslmed.aat3741
- Chushi, L., Wei, W., Kangkang, X., Yongzeng, F., Ning, X., and Xiaolei, C. (2016). HMGCR is up-regulated in gastric cancer and promotes the growth

and migration of the cancer cells. *Gene* 587, 42–47. doi: 10.1016/j.gene.2016. 04.029

- Ciarlo, L., Manganelli, V., Garofalo, T., Matarrese, P., Tinari, A., Misasi, R., et al. (2010). Association of fission proteins with mitochondrial raft-like domains. *Cell. Death Differ*. 17, 1047–1058. doi: 10.1038/cdd.2009.208
- Ciarlo, L., Vona, R., Manganelli, V., Gambardella, L., Raggi, C., Marconi, M., et al. (2018). Recruitment of mitofusin 2 into "lipid rafts" drives mitochondria fusion induced by Mdivi-1. *Oncotarget* 9, 18869–18884. doi: 10.18632/oncotarget. 24792
- Cirmena, G., Franceschelli, P., Isnaldi, E., Ferrando, L., De Mariano, M., Ballestrero, A., et al. (2018). Squalene epoxidase as a promising metabolic target in cancer treatment. *Cancer Lett.* 425, 13–20. doi: 10.1016/j.canlet.2018.03.034
- Clendening, J.W., Pandyra, A., Boutros, P.C., El Ghamrasni, S., Khosravi, F., Trentin, G.A., et al. (2010). Dysregulation of the mevalonate pathway promotes transformation. *Proc. Natl. Acad. Sci. U S A.* 107, 15051–15056. doi: 10.1073/ pnas.0910258107
- Costet, P., Luo, Y., Wang, N., and Tall, A.R. (2000). Sterol-dependent transactivation of the ABC1 promoter by the liver X receptor/retinoid X receptor. J. Biol. Chem. 275, 28240–28245. doi: 10.1074/jbc.M003337200
- Cruz, P.M., Mo, H., McConathy, W.J., Sabnis, N., and Lacko, A.G. (2013). The role of cholesterol metabolism and cholesterol transport in carcinogenesis: a review of scientific findings, relevant to future cancer therapeutics. *Front. Pharmacol.* 4:119. doi: 10.3389/fphar.2013.00119
- Dambal, S., Alfaqih, M., Sanders, S., Maravilla, E., Ramirez-Torres, A., Galvan, G.C., et al. (2020). 27-Hydroxycholesterol impairs plasma membrane lipid raft signaling as evidenced by inhibition of IL6-JAK-STAT3 signaling in prostate cancer cells. *Mol. Cancer Res.* 18, 671–684. doi: 10.1158/1541-7786.MCR-19-0974
- Daniil, G., Zannis, V.I., and Chroni, A. (2013). Effect of apoA-I mutations in the capacity of reconstituted HDL to promote ABCG1-mediated cholesterol efflux. *PLoS One* 8:e67993. doi: 10.1371/journal.pone.0067993
- Danza, G., Di Serio, C., Ambrosio, M.R., Sturli, N., Lonetto, G., Rosati, F., et al. (2013). Notch3 is activated by chronic hypoxia and contributes to the progression of human prostate cancer. *Int. J. Cancer* 133, 2577–2586. doi: 10.1002/ijc.28293
- Deeley, R.G., Westlake, C., and Cole, S.P. (2006). Transmembrane transport of endo- and xenobiotics by mammalian ATP-binding cassette multidrug resistance proteins. *Physiol Rev.* 86, 849–899. doi: 10.1152/physrev.00035.2005
- Delmas, D., Rebe, C., Micheau, O., Athias, A., Gambert, P., Grazide, S., et al. (2004). Redistribution of CD95, DR4 and DR5 in rafts accounts for the synergistic toxicity of resveratrol and death receptor ligands in colon carcinoma cells. Oncogene 23, 8979–8986. doi: 10.1038/sj.onc.1208086
- Denis, M., Landry, Y.D., and Zha, X. (2008). ATP-binding cassette A1-mediated lipidation of apolipoprotein A-I occurs at the plasma membrane and not in the endocytic compartments. *J. Biol. Chem.* 283, 16178–16186. doi: 10.1074/jbc. M709597200
- Ding, X., Zhang, W., Li, S., and Yang, H. (2019). The role of cholesterol metabolism in cancer. Am. J. Cancer Res. 9, 219–227.
- Donatello, S., Babina, I.S., Hazelwood, L.D., Hill, A.D., Nabi, I.R., and Hopkins, A.M. (2012). Lipid raft association restricts CD44-ezrin interaction and promotion of breast cancer cell migration. *Am. J. Pathol.* 181, 2172–2187. doi: 10.1016/j.ajpath.2012.08.025
- Dong, F., Mo, Z., Eid, W., Courtney, K.C., and Zha, X. (2014). Akt inhibition promotes ABCA1-mediated cholesterol efflux to ApoA-I through suppressing mTORC1. *PLoS One* 9:e113789. doi: 10.1371/journal.pone.0113789
- DuBroff, R., and de Lorgeril, M. (2015). Cholesterol confusion and statin controversy. World J. Cardiol. 7, 404–409. doi: 10.4330/wjc.v7.i7.404
- Duvel, K., Yecies, J.L., Menon, S., Raman, P., Lipovsky, A.I., Souza, A.L., et al. (2010). Activation of a metabolic gene regulatory network downstream of mTOR complex 1. *Mol. Cell.* 39, 171–183. doi: 10.1016/j.molcel.2010.06.022
- Ettinger, S.L., Sobel, R., Whitmore, T.G., Akbari, M., Bradley, D.R., Gleave, M.E., et al. (2004). Dysregulation of sterol response element-binding proteins and downstream effectors in prostate cancer during progression to androgen independence. *Cancer Res* 64(6), 2212–2221. doi: 10.1158/0008-5472.can-2148-2
- Fan, W., Chang, J., and Fu, P. (2015). Endocrine therapy resistance in breast cancer: current status, possible mechanisms and overcoming strategies. *Future Med. Chem.* 7, 1511–1519. doi: 10.4155/fmc.15.93

- Farwell, W.R., Scranton, R.E., Lawler, E.V., Lew, R.A., Brophy, M.T., Fiore, L.D., et al. (2008). The association between statins and cancer incidence in a veterans population. *J. Natl. Cancer Inst.* 100, 134–139. doi: 10.1093/jnci/djm286
- Fedoseienko, A., Wijers, M., Wolters, J.C., Dekker, D., Smit, M., Huijkman, N., et al. (2018). The COMMD family regulates plasma LDL levels and attenuates atherosclerosis through stabilizing the CCC complex in endosomal LDLR trafficking. *Circ. Res.* 122, 1648–1660. doi: 10.1161/CIRCRESAHA.117.312004
- Feig, C., Tchikov, V., Schutze, S., and Peter, M.E. (2007). Palmitoylation of CD95 facilitates formation of SDS-stable receptor aggregates that initiate apoptosis signaling. *EMBO J.* 26, 221–231. doi: 10.1038/sj.emboj.7601460
- Feinberg, M.W., and Moore, K.J. (2016). MicroRNA regulation of atherosclerosis. *Circ. Res.* 118, 703–720. doi: 10.1161/CIRCRESAHA.115.306300
- Fu, W., Yao, J., Huang, Y., Li, Q., Li, W., Chen, Z., et al. (2014). LXR agonist regulates the carcinogenesis of PCa via the SOCS3 pathway. *Cell. Physiol. Biochem.* 33, 195–204. doi: 10.1159/000356662
- Gabitova, L., Restifo, D., Gorin, A., Manocha, K., Handorf, E., Yang, D.H., et al. (2015). Endogenous sterol metabolites regulate growth of EGFR/KRASdependent tumors via LXR. *Cell. Rep.* 12, 1927–1938. doi: 10.1016/j.celrep.2015. 08.023
- Gajate, C., Del Canto-Janez, E., Acuna, A.U., Amat-Guerri, F., Geijo, E., Santos-Beneit, A.M., et al. (2004). Intracellular triggering of Fas aggregation and recruitment of apoptotic molecules into Fas-enriched rafts in selective tumor cell apoptosis. J. Exp. Med. 200, 353–365. doi: 10.1084/jem.20040213
- Gajate, C., Gonzalez-Camacho, F., and Mollinedo, F. (2009). Involvement of raft aggregates enriched in Fas/CD95 death-inducing signaling complex in the antileukemic action of edelfosine in jurkat cells. *PLoS One* 4:e5044. doi: 10.1371/ journal.pone.0005044
- Gajate, C., and Mollinedo, F. (2001). The antitumor ether lipid ET-18-OCH(3) induces apoptosis through translocation and capping of Fas/CD95 into membrane rafts in human leukemic cells. *Blood* 98, 3860–3863. doi: 10.1182/ blood.v98.13.3860
- Gajate, C., and Mollinedo, F. (2005). Cytoskeleton-mediated death receptor and ligand concentration in lipid rafts forms apoptosis-promoting clusters in cancer chemotherapy. J. Biol. Chem. 280, 11641–11647. doi: 10.1074/jbc.M411781200
- Gajate, C., and Mollinedo, F. (2007). Edelfosine and perifosine induce selective apoptosis in multiple myeloma by recruitment of death receptors and downstream signaling molecules into lipid rafts. *Blood* 109, 711–719. doi: 10. 1182/blood-2006-04-016824
- Gajate, C., and Mollinedo, F. (2014). Lipid rafts, endoplasmic reticulum and mitochondria in the antitumor action of the alkylphospholipid analog edelfosine. *Anticancer Agents Med. Chem.* 14, 509–527. doi: 10.2174/187152 0614666140309222259
- Gajate, C., and Mollinedo, F. (2015a). Lipid raft-mediated Fas/CD95 apoptotic signaling in leukemic cells and normal leukocytes and therapeutic implications. *J. Leukoc Biol.* 98, 739–759. doi: 10.1189/jlb.2MR0215-055R
- Gajate, C., and Mollinedo, F. (2015b). Lipid rafts and raft-mediated supramolecular entities in the regulation of CD95 death receptor apoptotic signaling. *Apoptosis* 20, 584–606. doi: 10.1007/s10495-015-1104-6
- Galbiati, F., Volonte, D., Meani, D., Milligan, G., Lublin, D.M., Lisanti, M.P., et al. (1999). The dually acylated NH2-terminal domain of gi1alpha is sufficient to target a green fluorescent protein reporter to caveolin-enriched plasma membrane domains. Palmitoylation of caveolin-1 is required for the recognition of dually acylated g-protein alpha subunits in vivo. *J. Biol. Chem.* 274(9), 5843–5850. doi: 10.1074/jbc.274.9.5843
- Gao, S., Shi, Z., Li, X., Li, W., Wang, Y., Liu, Z., et al. (2018). Fatostatin suppresses growth and enhances apoptosis by blocking SREBP-regulated metabolic pathways in endometrial carcinoma. *Oncol. Rep.* 39, 1919–1929. doi: 10.3892/or.2018.6265
- Gao, X., Lowry, P.R., Zhou, X., Depry, C., Wei, Z., Wong, G.W., et al. (2011). PI3K/Akt signaling requires spatial compartmentalization in plasma membrane microdomains. *Proc. Natl. Acad. Sci. U S A.* 108, 14509–14514. doi: 10.1073/ pnas.1019386108
- Gao, X., and Zhang, J. (2008). Spatiotemporal analysis of differential Akt regulation in plasma membrane microdomains. *Mol. Biol. Cell.* 19, 4366–4373. doi: 10. 1091/mbc.E08-05-0449
- Gao, X., and Zhang, J. (2009). Akt signaling dynamics in plasma membrane microdomains visualized by FRET-based reporters. *Commun. Integr. Biol.* 2, 32–34. doi: 10.4161/cib.2.1.7420

- Garofalo, T., Giammarioli, A.M., Misasi, R., Tinari, A., Manganelli, V., Gambardella, L., et al. (2005). Lipid microdomains contribute to apoptosisassociated modifications of mitochondria in T cells. *Cell. Death Differ.* 12, 1378–1389. doi: 10.1038/sj.cdd.4401672
- Garofalo, T., Manganelli, V., Grasso, M., Mattei, V., Ferri, A., Misasi, R., et al. (2015). Role of mitochondrial raft-like microdomains in the regulation of cell apoptosis. *Apoptosis* 20, 621–634. doi: 10.1007/s10495-015-1100-x
- Garofalo, T., Matarrese, P., Manganelli, V., Marconi, M., Tinari, A., Gambardella, L., et al. (2016). Evidence for the involvement of lipid rafts localized at the ERmitochondria associated membranes in autophagosome formation. *Autophagy* 12, 917–935. doi: 10.1080/15548627.2016.1160971
- Garofalo, T., Misasi, R., Mattei, V., Giammarioli, A.M., Malorni, W., Pontieri, G.M., et al. (2003). Association of the death-inducing signaling complex with microdomains after triggering through CD95/Fas. Evidence for caspase-8ganglioside interaction in T cells. J. Biol. Chem. 278, 8309–8315. doi: 10.1074/ jbc.M207618200
- Garofalo, T., Tinari, A., Matarrese, P., Giammarioli, A.M., Manganelli, V., Ciarlo, L., et al. (2007). Do mitochondria act as "cargo boats" in the journey of GD3 to the nucleus during apoptosis? *FEBS Lett.* 581, 3899–3903. doi: 10.1016/j.febslet. 2007.07.020
- Gbelcova, H., Lenicek, M., Zelenka, J., Knejzlik, Z., Dvorakova, G., Zadinova, M., et al. (2008). Differences in antitumor effects of various statins on human pancreatic cancer. *Int. J. Cancer* 122, 1214–1221. doi: 10.1002/ijc.23242
- Ge, L., Wang, J., Qi, W., Miao, H.H., Cao, J., Qu, Y.X., et al. (2008). The cholesterol absorption inhibitor ezetimibe acts by blocking the sterol-induced internalization of NPC1L1. *Cell. Metab.* 7, 508–519. doi: 10.1016/j.cmet.2008. 04.001
- Gelissen, I.C., Harris, M., Rye, K.A., Quinn, C., Brown, A.J., Kockx, M., et al. (2006). ABCA1 and ABCG1 synergize to mediate cholesterol export to apoA-I. *Arterioscler Thromb Vasc. Biol.* 26, 534–540. doi: 10.1161/01.ATV.0000200082. 58536.e1
- Geng, F., Cheng, X., Wu, X., Yoo, J.Y., Cheng, C., Guo, J.Y., et al. (2016). Inhibition of SOAT1 suppresses glioblastoma growth via blocking SREBP-1-mediated lipogenesis. *Clin. Cancer Res.* 22, 5337–5348. doi: 10.1158/1078-0432.CCR-15-2973
- Gholkar, A.A., Cheung, K., Williams, K.J., Lo, Y.C., Hamideh, S.A., Nnebe, C., et al. (2016). Fatostatin inhibits cancer cell proliferation by affecting mitotic microtubule spindle assembly and cell division. J. Biol. Chem. 291, 17001– 17008. doi: 10.1074/jbc.C116.737346
- Ginestier, C., Monville, F., Wicinski, J., Cabaud, O., Cervera, N., Josselin, E., et al. (2012). Mevalonate metabolism regulates Basal breast cancer stem cells and is a potential therapeutic target. *Stem Cells* 30, 1327–1337. doi: 10.1002/stem.1122
- Goldstein, J.L., and Brown, M.S. (1990). Regulation of the mevalonate pathway. *Nature* 343, 425–430. doi: 10.1038/343425a0
- Goldstein, J.L., and Brown, M.S. (2009). The LDL receptor. Arterioscler Thromb Vasc. Biol. 29, 431-438. doi: 10.1161/ATVBAHA.108.179564
- Gong, Y., Lee, J.N., Lee, P.C., Goldstein, J.L., Brown, M.S., and Ye, J. (2006). Sterol-regulated ubiquitination and degradation of Insig-1 creates a convergent mechanism for feedback control of cholesterol synthesis and uptake. *Cell. Metab.* 3, 15–24. doi: 10.1016/j.cmet.2005.11.014
- Guillaumond, F., Bidaut, G., Ouaissi, M., Servais, S., Gouirand, V., Olivares, O., et al. (2015). Cholesterol uptake disruption, in association with chemotherapy, is a promising combined metabolic therapy for pancreatic adenocarcinoma. *Proc. Natl. Acad. Sci. U S A.* 112, 2473–2478. doi: 10.1073/pnas.1421601112
- Guo, D., Prins, R.M., Dang, J., Kuga, D., Iwanami, A., Soto, H., et al. (2009). EGFR signaling through an Akt-SREBP-1-dependent, rapamycin-resistant pathway sensitizes glioblastomas to antilipogenic therapy. *Sci. Sign.* 2:ra82. doi: 10.1126/ scisignal.2000446
- Guo, D., Reinitz, F., Youssef, M., Hong, C., Nathanson, D., Akhavan, D., et al. (2011). An LXR agonist promotes glioblastoma cell death through inhibition of an EGFR/AKT/SREBP-1/LDLR-dependent pathway. *Cancer Discov.* 1, 442– 456. doi: 10.1158/2159-8290.CD-11-0102
- Haddon, L., and Hugh, J. (2015). MUC1-mediated motility in breast cancer: a review highlighting the role of the MUC1/ICAM-1/Src signaling triad. *Clin. Exp. Metastasis* 32, 393–403. doi: 10.1007/s10585-015-9711-8
- Hanahan, D., and Weinberg, R.A. (2000). The hallmarks of cancer. *Cell* 100, 57–70. doi: 10.1016/s0092-8674(00)81683-9

- Hanahan, D., and Weinberg, R.A. (2011). Hallmarks of cancer: the next generation. *Cell* 144, 646–674. doi: 10.1016/j.cell.2011.02.013
- Heir, T., Falk, R.S., Robsahm, T.E., Sandvik, L., Erikssen, J., and Tretli, S. (2016). Cholesterol and prostate cancer risk: a long-term prospective cohort study. *BMC Cancer* 16:643. doi: 10.1186/s12885-016-2691-5
- Horton, J.D., Goldstein, J.L., and Brown, M.S. (2002). SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver. J Clin Invest 109(9), 1125-1131. doi: 10.1172/JCI15593
- Howe, V., Sharpe, L.J., Prabhu, A.V., and Brown, A.J. (2017). New insights into cellular cholesterol acquisition: promoter analysis of human HMGCR and SQLE, two key control enzymes in cholesterol synthesis. *Biochim. Biophys. Acta. Mol. Cell. Biol. Lipids* 1862, 647–657. doi: 10.1016/j.bbalip.2017.03.009
- Hryniewicz-Jankowska, A., Augoff, K., and Sikorski, A.F. (2019). The role of cholesterol and cholesterol-driven membrane raft domains in prostate cancer. *Exp. Biol. Med. (Maywood)* 244, 1053–1061. doi: 10.1177/1535370219870771
- Hu, J., La Vecchia, C., de Groh, M., Negri, E., Morrison, H., Mery, L., et al. (2012). Dietary cholesterol intake and cancer. Ann. Oncol. 23, 491–500. doi: 10.1093/annonc/mdr155
- Huang, C., Hays, F.A., Tomasek, J.J., Benyajati, S., and Zhang, X.A. (2020). Tetraspanin CD82 interaction with cholesterol promotes extracellular vesiclemediated release of ezrin to inhibit tumour cell movement. *J. Extracell Vesicles* 9:1692417. doi: 10.1080/20013078.2019.1692417
- Huang, H., and Tindall, D.J. (2002). The role of the androgen receptor in prostate cancer. *Crit. Rev. Eukaryot Gene. Expr.* 12, 193–207. doi: 10.1615/ critreveukaryotgeneexpr.v12.i3.30
- Iessi, E., Marconi, M., Manganelli, V., Sorice, M., Malorni, W., Garofalo, T., et al. (2020). On the role of sphingolipids in cell survival and death. *Int. Rev. Cell. Mol. Biol.* 351, 149–195. doi: 10.1016/bs.ircmb.2020.02.004
- Ikonen, E. (2008). Cellular cholesterol trafficking and compartmentalization. Nat. Rev. Mol. Cell. Biol. 9, 125–138. doi: 10.1038/nrm2336
- Jiang, Y., Sun, A., Zhao, Y., Ying, W., Sun, H., Yang, X., et al. (2019). Proteomics identifies new therapeutic targets of early-stage hepatocellular carcinoma. *Nature* 567, 257–261. doi: 10.1038/s41586-019-0987-8
- Khan, A.A., Agarwal, H., Reddy, S.S., Arige, V., Natarajan, B., Gupta, V., et al. (2020). MicroRNA 27a is a key modulator of cholesterol biosynthesis. *Mol. Cell. Biol.* 40:e470-19. doi: 10.1128/MCB.00470-19
- Kim, J., Adam, R.M., Solomon, K.R., and Freeman, M.R. (2004). Involvement of cholesterol-rich lipid rafts in interleukin-6-induced neuroendocrine differentiation of LNCaP prostate cancer cells. *Endocrinology* 145, 613–619. doi: 10.1210/en.2003-0772
- Komati, R., Spadoni, D., Zheng, S., Sridhar, J., Riley, K.E., and Wang, G. (2017). Ligands of therapeutic utility for the liver X receptors. *Molecules* 22:88. doi: 10.3390/molecules22010088
- Koseki, M., Hirano, K., Masuda, D., Ikegami, C., Tanaka, M., Ota, A., et al. (2007). Increased lipid rafts and accelerated lipopolysaccharide-induced tumor necrosis factor-alpha secretion in Abca1-deficient macrophages. J. Lipid Res. 48, 299–306. doi: 10.1194/jlr.M600428-JLR200
- Kuzu, O.F., Noory, M.A., and Robertson, G.P. (2016). The role of cholesterol in cancer. *Cancer Res.* 76, 2063–2070. doi: 10.1158/0008-5472.CAN-15-2613
- Kwon, H.J., Abi-Mosleh, L., Wang, M.L., Deisenhofer, J., Goldstein, J.L., Brown, M.S., et al. (2009). Structure of N-terminal domain of NPC1 reveals distinct subdomains for binding and transfer of cholesterol. *Cell* 137, 1213–1224. doi: 10.1016/j.cell.2009.03.049
- Landry, Y.D., Denis, M., Nandi, S., Bell, S., Vaughan, A.M., and Zha, X. (2006). ATP-binding cassette transporter A1 expression disrupts raft membrane microdomains through its ATPase-related functions. J. Biol. Chem. 281, 36091– 36101. doi: 10.1074/jbc.M602247200
- Li, J., Gu, D., Lee, S.S., Song, B., Bandyopadhyay, S., Chen, S., et al. (2016). Abrogating cholesterol esterification suppresses growth and metastasis of pancreatic cancer. Oncogene 35, 6378–6388. doi: 10.1038/onc.2016.168
- Li, J., and Pfeffer, S.R. (2016). Lysosomal membrane glycoproteins bind cholesterol and contribute to lysosomal cholesterol export. *Elife* 5:e21635. doi: 10.7554/ eLife.21635
- Li, L., Zhang, Q., Wang, X., Li, Y., Xie, H., and Chen, X. (2020). Squalene epoxidaseinduced cholesteryl ester accumulation promotes nasopharyngeal carcinoma development by activating PI3K/AKT signaling. *Cancer Sci.* 111, 2275–2283. doi: 10.1111/cas.14426

- Liang, W., Hao, Z., Han, J.L., Zhu, D.J., Jin, Z.F., and Xie, W.L. (2014). CAV-1 contributes to bladder cancer progression by inducing epithelial-tomesenchymal transition. Urol. Oncol. 32, 855–863. doi: 10.1016/j.urolonc.2014. 01.005
- Lim, S.C., Duong, H.Q., Choi, J.E., Lee, T.B., Kang, J.H., Oh, S.H., et al. (2011). Lipid raft-dependent death receptor 5 (DR5) expression and activation are critical for ursodeoxycholic acid-induced apoptosis in gastric cancer cells. *Carcinogenesis* 32, 723–731. doi: 10.1093/carcin/bgr038
- Lim, S.C., Parajuli, K.R., and Han, S.I. (2016). The alkyllysophospholipid edelfosine enhances TRAIL-mediated apoptosis in gastric cancer cells through death receptor 5 and the mitochondrial pathway. *Tumour Biol.* 37, 6205–6216. doi: 10.1007/s13277-015-4485-9
- Lin, C.Y., Huo, C., Kuo, L.K., Hiipakka, R.A., Jones, R.B., Lin, H.P., et al. (2013). Cholestane-3beta, 5alpha, 6beta-triol suppresses proliferation, migration, and invasion of human prostate cancer cells. *PLoS One* 8:e65734. doi: 10.1371/ journal.pone.0065734
- Lin, C.Y., Vedin, L.L., and Steffensen, K.R. (2016). The emerging roles of liver X receptors and their ligands in cancer. *Exp. Opin. Ther. Targets* 20, 61–71. doi: 10.1517/14728222.2015.1081169
- Liu, F.T., Qu, Q.G., and Zhu, Z.M. (2017). Up-regulation of Flot-2 protein is related to lymph node metastasis and poor prognosis in human solid tumors. *Minerva Chir.* 72, 146–156. doi: 10.23736/S0026-4733.16.07261-8
- Llaverias, G., Danilo, C., Mercier, I., Daumer, K., Capozza, F., Williams, T.M., et al. (2011). Role of cholesterol in the development and progression of breast cancer. *Am. J. Pathol.* 178, 402–412. doi: 10.1016/j.ajpath.2010.11.005
- Loh, K., Tam, S., Murray-Segal, L., Huynh, K., Meikle, P.J., Scott, J.W., et al. (2019). Inhibition of adenosine monophosphate-activated protein kinase-3-hydroxy-3methylglutaryl coenzyme a reductase signaling leads to hypercholesterolemia and promotes hepatic steatosis and insulin resistance. *Hepatol. Commun.* 3, 84–98. doi: 10.1002/hep4.1279
- Longo, J., Mullen, P.J., Yu, R., van Leeuwen, J.E., Masoomian, M., Woon, D.T.S., et al. (2019). An actionable sterol-regulated feedback loop modulates statin sensitivity in prostate cancer. *Mol. Metab.* 25, 119–130. doi: 10.1016/j.molmet. 2019.04.003
- Lopez, D., Abisambra Socarras, J.F., Bedi, M., and Ness, G.C. (2007). Activation of the hepatic LDL receptor promoter by thyroid hormone. *Biochim. Biophys. Acta* 1771, 1216–1225. doi: 10.1016/j.bbalip.2007.05.001
- Lorenzi, I., von Eckardstein, A., Radosavljevic, S., and Rohrer, L. (2008). Lipidation of apolipoprotein A-I by ATP-binding cassette transporter (ABC) A1 generates an interaction partner for ABCG1 but not for scavenger receptor BI. *Biochim. Biophys. Acta* 1781, 306–313. doi: 10.1016/j.bbalip.2008.04.006
- Lozupone, F., Lugini, L., Matarrese, P., Luciani, F., Federici, C., Iessi, E., et al. (2004). Identification and relevance of the CD95-binding domain in the N-terminal region of ezrin. J. Biol. Chem. 279, 9199–9207. doi: 10.1074/jbc. M305561200
- Lu-Yao, G.L., Albertsen, P.C., Moore, D.F., Shih, W., Lin, Y., DiPaola, R.S., et al. (2014). Fifteen-year survival outcomes following primary androgendeprivation therapy for localized prostate cancer. *JAMA Intern. Med.* 174, 1460–1467. doi: 10.1001/jamainternmed.2014.3028
- Lu, R., Arakawa, R., Ito-Osumi, C., Iwamoto, N., and Yokoyama, S. (2008). ApoA-I facilitates ABCA1 recycle/accumulation to cell surface by inhibiting its intracellular degradation and increases HDL generation. *Arterioscler Thromb Vasc. Biol.* 28, 1820–1824. doi: 10.1161/ATVBAHA.108.16 9482
- Lukanova, A., and Kaaks, R. (2005). Endogenous hormones and ovarian cancer: epidemiology and current hypotheses. *Cancer Epidemiol. Biomark. Prev.* 14, 98–107.
- Luo, J., Yang, H., and Song, B.L. (2020). Mechanisms and regulation of cholesterol homeostasis. Nat. Rev. Mol. Cell. Biol. 21, 225–245. doi: 10.1038/s41580-019-0190-7
- Luo, X., Cheng, C., Tan, Z., Li, N., Tang, M., Yang, L., et al. (2017). Emerging roles of lipid metabolism in cancer metastasis. *Mol. Cancer* 16:76. doi: 10.1186/s12943-017-0646-3
- Malhotra, P., Soni, V., Yamanashi, Y., Takada, T., Suzuki, H., Gill, R.K., et al. (2019). Mechanisms of niemann-pick type C1 Like 1 protein degradation in intestinal epithelial cells. Am. J. Physiol. Cell. Physiol 316, C559–C566. doi: 10.1152/ajpcell.00465.2018

- Malorni, W., Giammarioli, A.M., Garofalo, T., and Sorice, M. (2007). Dynamics of lipid raft components during lymphocyte apoptosis: the paradigmatic role of GD3. *Apoptosis* 12(5), 941–949. doi: 10.1007/s10495-007-0757-1
- Marconi, M., Ascione, B., Ciarlo, L., Vona, R., Garofalo, T., Sorice, M., et al. (2013). Constitutive localization of DR4 in lipid rafts is mandatory for TRAILinduced apoptosis in B-cell hematologic malignancies. *Cell. Death Dis.* 4:e863. doi: 10.1038/cddis.2013.389
- Martin, T.A., Harrison, G., Mansel, R.E., and Jiang, W.G. (2003). The role of the CD44/ezrin complex in cancer metastasis. *Crit. Rev. Oncol. Hematol.* 46, 165–186. doi: 10.1016/s1040-8428(02)00172-5
- Martinez-Outschoorn, U.E., Lisanti, M.P., and Sotgia, F. (2014). Catabolic cancerassociated fibroblasts transfer energy and biomass to anabolic cancer cells, fueling tumor growth. *Semin. Cancer Biol.* 25, 47–60. doi: 10.1016/j.semcancer. 2014.01.005
- Martinez-Outschoorn, U.E., Trimmer, C., Lin, Z., Whitaker-Menezes, D., Chiavarina, B., Zhou, J., et al. (2010). Autophagy in cancer associated fibroblasts promotes tumor cell survival: role of hypoxia, HIF1 induction and NFkappaB activation in the tumor stromal microenvironment. *Cell. Cycle* 9, 3515–3533. doi: 10.4161/cc.9.17.12928
- Matarrese, P., Garofalo, T., Manganelli, V., Gambardella, L., Marconi, M., Grasso, M., et al. (2014). Evidence for the involvement of GD3 ganglioside in autophagosome formation and maturation. *Autophagy* 10, 750–765. doi: 10. 4161/auto.27959
- Maxfield, F.R., and Tabas, I. (2005). Role of cholesterol and lipid organization in disease. *Nature* 438, 612–621. doi: 10.1038/nature04399
- Maxfield, F.R., and van Meer, G. (2010). Cholesterol, the central lipid of mammalian cells. *Curr. Opin. Cell. Biol.* 22, 422–429. doi: 10.1016/j.ceb.2010. 05.004
- McDonnell, D.P., Park, S., Goulet, M.T., Jasper, J., Wardell, S.E., Chang, C.Y., et al. (2014). Obesity, cholesterol metabolism, and breast cancer pathogenesis. *Cancer Res.* 74, 4976–4982. doi: 10.1158/0008-5472.CAN-14-1756
- Mok, E.H.K., and Lee, T.K.W. (2020). The pivotal role of the dysregulation of cholesterol homeostasis in cancer: implications for therapeutic targets. *Cancers* (*Basel*) 12:1410. doi: 10.3390/cancers12061410
- Mokarram, P., Alizadeh, J., Razban, V., Barazeh, M., Solomon, C., and Kavousipour, S. (2017). Interconnection of estrogen/testosterone metabolism and mevalonate pathway in breast and prostate cancers. *Curr. Mol. Pharmacol.* 10, 86–114. doi: 10.2174/1874467209666160112125631
- Mollinedo, F., de la Iglesia-Vicente, J., Gajate, C., Estella-Hermoso de Mendoza, A., Villa-Pulgarin, J.A., Campanero, M.A., et al. (2010a). Lipid raft-targeted therapy in multiple myeloma. Oncogene 29, 3748–3757. doi: 10.1038/onc.2010.131
- Mollinedo, F., de la Iglesia-Vicente, J., Gajate, C., Estella-Hermoso de Mendoza, A., Villa-Pulgarin, J.A., de Frias, M., et al. (2010b). In vitro and In vivo selective antitumor activity of Edelfosine against mantle cell lymphoma and chronic lymphocytic leukemia involving lipid rafts. *Clin. Cancer Res.* 16, 2046–2054. doi: 10.1158/1078-0432.CCR-09-2456
- Mollinedo, F., and Gajate, C. (2006). Fas/CD95 death receptor and lipid rafts: new targets for apoptosis-directed cancer therapy. *Drug Resist. Updat.* 9(1-2), 51–73. doi: 10.1016/j.drup.2006.04.002
- Mollinedo, F., and Gajate, C. (2010). Lipid rafts and clusters of apoptotic signaling molecule-enriched rafts in cancer therapy. *Future Oncol.* 6, 811–821. doi: 10. 2217/fon.10.34
- Mollinedo, F., and Gajate, C. (2015). Lipid rafts as major platforms for signaling regulation in cancer. Adv. Biol. Regul. 57, 130–146. doi: 10.1016/j.jbior.2014. 10.003
- Mollinedo, F., and Gajate, C. (2020). Lipid rafts as signaling hubs in cancer cell survival/death and invasion: implications in tumor progression and therapy. *J. Lipid Res.* 61, 611–635. doi: 10.1194/jlr.TR119000439
- Mollinedo, F., Gajate, C., Martin-Santamaria, S., and Gago, F. (2004). ET-18-OCH3 (edelfosine): a selective antitumour lipid targeting apoptosis through intracellular activation of Fas/CD95 death receptor. *Curr. Med. Chem.* 11, 3163–3184. doi: 10.2174/0929867043363703
- Motoyama, K., Kameyama, K., Onodera, R., Araki, N., Hirayama, F., Uekama, K., et al. (2009). Involvement of PI3K-Akt-Bad pathway in apoptosis induced by 2,6-di-O-methyl-beta-cyclodextrin, not 2,6-di-O-methyl-alpha-cyclodextrin, through cholesterol depletion from lipid rafts on plasma membranes in cells. *Eur. J. Pharm. Sci.* 38(3), 249–261. doi: 10.1016/j.ejps.2009.07.010

- Murai, T. (2012). The role of lipid rafts in cancer cell adhesion and migration. *Int. J. Cell. Biol.* 2012;763283. doi: 10.1155/2012/763283
- Murai, T. (2015). Cholesterol lowering: role in cancer prevention and treatment. Biol. Chem. 396, 1–11. doi: 10.1515/hsz-2014-0194
- Murai, T., Hokonohara, H., Takagi, A., and Kawai, T. (2009). Atomic force microscopy imaging of supramolecular organization of hyaluronan and its receptor CD44. *IEEE Trans. Nanobiosci.* 8, 294–299. doi: 10.1109/TNB.2009. 2035251
- Murai, T., Maruyama, Y., Mio, K., Nishiyama, H., Suga, M., and Sato, C. (2011). Low cholesterol triggers membrane microdomain-dependent CD44 shedding and suppresses tumor cell migration. *J. Biol. Chem.* 286, 1999–2007. doi: 10. 1074/jbc.M110.184010
- Murai, T., Miyauchi, T., Yanagida, T., and Sako, Y. (2006). Epidermal growth factor-regulated activation of Rac GTPase enhances CD44 cleavage by metalloproteinase disintegrin ADAM10. *Biochem. J.* 395, 65–71. doi: 10.1042/ BJ20050582
- Murai, T., Miyazaki, Y., Nishinakamura, H., Sugahara, K.N., Miyauchi, T., Sako, Y., et al. (2004). Engagement of CD44 promotes Rac activation and CD44 cleavage during tumor cell migration. *J. Biol. Chem.* 279, 4541–4550. doi: 10.1074/jbc. M307356200
- Nagai, M., Sakakibara, J., Nakamura, Y., Gejyo, F., and Ono, T. (2002). SREBP-2 and NF-Y are involved in the transcriptional regulation of squalene epoxidase. *Biochem. Biophys. Res. Commun.* 295, 74–80. doi: 10.1016/s0006-291x(02) 00623-x
- Najafi-Shoushtari, S.H., Kristo, F., Li, Y., Shioda, T., Cohen, D.E., Gerszten, R.E., et al. (2010). MicroRNA-33 and the SREBP host genes cooperate to control cholesterol homeostasis. *Science* 328, 1566–1569. doi: 10.1126/science.1189123
- Nakashima, J., Tachibana, M., Horiguchi, Y., Oya, M., Ohigashi, T., Asakura, H., et al. (2000). Serum interleukin 6 as a prognostic factor in patients with prostate cancer. *Clin. Cancer Res.* 6, 2702–2706.
- Nicolle, R., Blum, Y., Marisa, L., Loncle, C., Gayet, O., Moutardier, V., et al. (2017). Pancreatic adenocarcinoma therapeutic targets revealed by tumorstroma cross-talk analyses in patient-derived xenografts. *Cell. Rep.* 21, 2458– 2470. doi: 10.1016/j.celrep.2017.11.003
- Nicolson, G.L. (2015). Cell membrane fluid-mosaic structure and cancer metastasis. *Cancer Res.* 75, 1169–1176. doi: 10.1158/0008-5472.CAN-14-3216
- Nielsen, S.F., Nordestgaard, B.G., and Bojesen, S.E. (2013). Statin use and reduced cancer-related mortality. N. Engl. J. Med. 368, 576–577. doi: 10.1056/ NEJMc1214827
- Nooh, A., Zhang, Y.L., Sato, D., Rosenzweig, D.H., Tabaries, S., Siegel, P., et al. (2017). Intra-tumor delivery of zoledronate mitigates metastasis-induced osteolysis superior to systemic administration. J. Bone Oncol 6, 8–15. doi: 10. 1016/j.jbo.2017.01.001
- Oh, H.Y., Lee, E.J., Yoon, S., Chung, B.H., Cho, K.S., and Hong, S.J. (2007). Cholesterol level of lipid raft microdomains regulates apoptotic cell death in prostate cancer cells through EGFR-mediated Akt and ERK signal transduction. *Prostate* 67, 1061–1069. doi: 10.1002/pros.20593
- Oram, J.F., and Heinecke, J.W. (2005). ATP-binding cassette transporter A1: a cell cholesterol exporter that protects against cardiovascular disease. *Physiol. Rev.* 85, 1343–1372. doi: 10.1152/physrev.00005.2005
- Ouvrier, A., Cadet, R., Vernet, P., Laillet, B., Chardigny, J.M., Lobaccaro, J.M., et al. (2009). LXR and ABCA1 control cholesterol homeostasis in the proximal mouse epididymis in a cell-specific manner. *J. Lipid Res.* 50, 1766–1775. doi: 10.1194/jlr.M800657-JLR200
- Parlato, S., Giammarioli, A.M., Logozzi, M., Lozupone, F., Matarrese, P., Luciani, F., et al. (2000). CD95 (APO-1/Fas) linkage to the actin cytoskeleton through ezrin in human T lymphocytes: a novel regulatory mechanism of the CD95 apoptotic pathway. *EMBO J.* 19, 5123–5134. doi: 10.1093/emboj/19.19. 5123
- Patel, H.H., and Insel, P.A. (2009). Lipid rafts and caveolae and their role in compartmentation of redox signaling. *Antioxid Redox Signal* 11, 1357–1372. doi: 10.1089/ARS.2008.2365
- Pelton, K., Freeman, M.R., and Solomon, K.R. (2012). Cholesterol and prostate cancer. Curr. Opin. Pharmacol. 12, 751–759. doi: 10.1016/j.coph.2012.07.006
- Pencheva, N., Buss, C.G., Posada, J., Merghoub, T., and Tavazoie, S.F. (2014). Broad-spectrum therapeutic suppression of metastatic melanoma through nuclear hormone receptor activation. *Cell* 156, 986–1001. doi: 10.1016/j.cell. 2014.01.038

- Peterson, T.R., Sengupta, S.S., Harris, T.E., Carmack, A.E., Kang, S.A., Balderas, E., et al. (2011). mTOR complex 1 regulates lipin 1 localization to control the SREBP pathway. *Cell* 146, 408–420. doi: 10.1016/j.cell.2011.06.034
- Petrova, E., Rios-Esteves, J., Ouerfelli, O., Glickman, J.F., and Resh, M.D. (2013). Inhibitors of hedgehog acyltransferase block Sonic hedgehog signaling. *Nat. Chem. Biol.* 9, 247–249. doi: 10.1038/nchembio.1184
- Phillips, M.C. (2014). Molecular mechanisms of cellular cholesterol efflux. J. Biol. Chem. 289, 24020–24029. doi: 10.1074/jbc.R114.583658
- Pike, L.J. (2003). Lipid rafts: bringing order to chaos. J. Lipid Res. 44, 655–667. doi: 10.1194/jlr.R200021-JLR200
- Pike, L.J. (2006). Rafts defined: a report on the keystone symposium on lipid rafts and cell function. J. Lipid Res. 47, 1597–1598. doi: 10.1194/jlr.E600002-JLR200
- Pitroda, S.P., Khodarev, N.N., Beckett, M.A., Kufe, D.W., and Weichselbaum, R.R. (2009). MUC1-induced alterations in a lipid metabolic gene network predict response of human breast cancers to tamoxifen treatment. *Proc. Natl. Acad Sci.* U S A. 106, 5837–5841. doi: 10.1073/pnas.0812029106
- Pommier, A.J., Alves, G., Viennois, E., Bernard, S., Communal, Y., Sion, B., et al. (2010). Liver X Receptor activation downregulates AKT survival signaling in lipid rafts and induces apoptosis of prostate cancer cells. *Oncogene* 29, 2712–2723. doi: 10.1038/onc.2010.30
- Porstmann, T., Santos, C.R., Griffiths, B., Cully, M., Wu, M., Leevers, S., et al. (2008). SREBP activity is regulated by mTORC1 and contributes to Aktdependent cell growth. *Cell. Metab.* 8, 224–236. doi: 10.1016/j.cmet.2008. 07.007
- Pramfalk, C., Davis, M.A., Eriksson, M., Rudel, L.L., and Parini, P. (2005). Control of ACAT2 liver expression by HNF1. J. Lipid Res. 46, 1868–1876. doi: 10.1194/ jlr.M400450-JLR200
- Qiu, Z., Yuan, W., Chen, T., Zhou, C., Liu, C., Huang, Y., et al. (2016). HMGCR positively regulated the growth and migration of glioblastoma cells. *Gene* 576, 22–27. doi: 10.1016/j.gene.2015.09.067
- Radisauskas, R., Kuzmickiene, I., Milinaviciene, E., and Everatt, R. (2016). Hypertension, serum lipids and cancer risk: a review of epidemiological evidence. *Medicina (Kaunas)* 52, 89–98. doi: 10.1016/j.medici.2016.03.002
- Rayner, K.J., Esau, C.C., Hussain, F.N., McDaniel, A.L., Marshall, S.M., van Gils, J.M., et al. (2011). Inhibition of miR-33a/b in non-human primates raises plasma HDL and lowers VLDL triglycerides. *Nature* 478, 404–407. doi: 10.1038/ nature10486
- Razani, B., Rubin, C.S., and Lisanti, M.P. (1999). Regulation of cAMP-mediated signal transduction via interaction of caveolins with the catalytic subunit of protein kinase A. J. Biol. Chem. 274, 26353–26360. doi: 10.1074/jbc.274.37. 26353
- Reis-Sobreiro, M., Gajate, C., and Mollinedo, F. (2009). Involvement of mitochondria and recruitment of Fas/CD95 signaling in lipid rafts in resveratrol-mediated antimyeloma and antileukemia actions. Oncogene 28, 3221–3234. doi: 10.1038/onc.2009.183
- Reis-Sobreiro, M., Roue, G., Moros, A., Gajate, C., de la Iglesia-Vicente, J., Colomer, D., et al. (2013). Lipid raft-mediated Akt signaling as a therapeutic target in mantle cell lymphoma. *Blood Cancer J.* 3:e118. doi: 10.1038/bcj.2013.15
- Resh, M.D. (1999). Fatty acylation of proteins: new insights into membrane targeting of myristoylated and palmitoylated proteins. *Biochim. Biophys. Acta* 1451, 1–16. doi: 10.1016/s0167-4889(99)00075-0
- Resh, M.D. (2004). Membrane targeting of lipid modified signal transduction proteins. Subcell. Biochem. 37, 217–232. doi: 10.1007/978-1-4757-5806-1_6
- Rogers, M.A., Liu, J., Song, B.L., Li, B.L., Chang, C.C., and Chang, T.Y. (2015). Acyl-CoA:cholesterol acyltransferases (ACATs/SOATs): enzymes with multiple sterols as substrates and as activators. *J. Steroid Biochem. Mol. Biol.* 151, 102–107. doi: 10.1016/j.jsbmb.2014.09.008
- Rossin, A., Derouet, M., Abdel-Sater, F., and Hueber, A.O. (2009). Palmitoylation of the TRAIL receptor DR4 confers an efficient TRAIL-induced cell death signalling. *Biochem J*. 419, 185–192. doi: 10.1042/BJ20081212
- Rotllan, N., and Fernandez-Hernando, C. (2012). MicroRNA regulation of cholesterol metabolism. *Cholesterol* 2012:847849. doi: 10.1155/2012/847849
- Rudenko, G., Henry, L., Henderson, K., Ichtchenko, K., Brown, M.S., Goldstein, J.L., et al. (2002). Structure of the LDL receptor extracellular domain at endosomal pH. *Science* 298, 2353–2358. doi: 10.1126/science.1078124
- Saraon, P., Trudel, D., Kron, K., Dmitromanolakis, A., Trachtenberg, J., Bapat, B., et al. (2014). Evaluation and prognostic significance of ACAT1 as a marker of prostate cancer progression. *Prostate* 74, 372–380. doi: 10.1002/pros.22758

- Sato, R. (2010). Sterol metabolism and SREBP activation. Arch. Biochem. Biophys. 501, 177–181. doi: 10.1016/j.abb.2010.06.004
- Sbiera, S., Leich, E., Liebisch, G., Sbiera, I., Schirbel, A., Wiemer, L., et al. (2015). Mitotane inhibits sterol-O-acyl transferase 1 triggering lipid-mediated endoplasmic reticulum stress and apoptosis in adrenocortical carcinoma cells. *Endocrinology* 156, 3895–3908. doi: 10.1210/en.2015-1367
- Scheel-Toellner, D., Wang, K., Singh, R., Majeed, S., Raza, K., Curnow, S.J., et al. (2002). The death-inducing signalling complex is recruited to lipid rafts in Fas-induced apoptosis. *Biochem. Biophys. Res. Commun.* 297, 876–879. doi: 10.1016/s0006-291x(02)02311-2
- Scorrano, L. (2008). Caspase-8 goes cardiolipin: a new platform to provide mitochondria with microdomains of apoptotic signals? J. Cell. Biol. 183, 579– 581. doi: 10.1083/jcb.200810125
- Segala, G., de Medina, P., Iuliano, L., Zerbinati, C., Paillasse, M.R., Noguer, E., et al. (2013). 5,6-Epoxy-cholesterols contribute to the anticancer pharmacology of tamoxifen in breast cancer cells. *Biochem. Pharmacol.* 86, 175–189. doi: 10.1016/j.bcp.2013.02.031
- Shah, K., Gagliano, T., Garland, L., O'Hanlon, T., Bortolotti, D., Gentili, V., et al. (2020). Androgen receptor signaling regulates the transcriptome of prostate cancer cells by modulating global alternative splicing. *Oncogene* 39, 6172–6189. doi: 10.1038/s41388-020-01429-2
- Silvente-Poirot, S., and Poirot, M. (2012). Cholesterol metabolism and cancer: the good, the bad and the ugly. *Curr. Opin. Pharmacol.* 12, 673–676. doi: 10.1016/j. coph.2012.10.004
- Simigdala, N., Gao, Q., Pancholi, S., Roberg-Larsen, H., Zvelebil, M., Ribas, R., et al. (2016). Cholesterol biosynthesis pathway as a novel mechanism of resistance to estrogen deprivation in estrogen receptor-positive breast cancer. *Breast Cancer Res.* 18:58. doi: 10.1186/s13058-016-0713-5
- Simons, K., and Ehehalt, R. (2002). Cholesterol, lipid rafts, and disease. J. Clin. Invest. 110, 597–603. doi: 10.1172/JCI16390
- Simons, K., and Ikonen, E. (2000). How cells handle cholesterol. *Science* 290, 1721–1726. doi: 10.1126/science.290.5497.1721
- Simons, K., and Toomre, D. (2000). Lipid rafts and signal transduction. Nat. Rev. Mol. Cell. Biol. 1, 31–39. doi: 10.1038/35036052
- Solomon, K.R., and Freeman, M.R. (2008). Do the cholesterol-lowering properties of statins affect cancer risk? *Trends Endocrinol. Metab.* 19, 113–121. doi: 10. 1016/j.tem.2007.12.004
- Solomon, K.R., Pelton, K., Boucher, K., Joo, J., Tully, C., Zurakowski, D., et al. (2009). Ezetimibe is an inhibitor of tumor angiogenesis. *Am. J. Pathol.* 174, 1017–1026. doi: 10.2353/ajpath.2009.080551
- Song, B.L., Javitt, N.B., and DeBose-Boyd, R.A. (2005). Insig-mediated degradation of HMG CoA reductase stimulated by lanosterol, an intermediate in the synthesis of cholesterol. *Cell. Metab.* 1, 179–189. doi: 10.1016/j.cmet.2005. 01.001
- Song, K.S., Sargiacomo, M., Galbiati, F., Parenti, M., and Lisanti, M.P. (1997). Targeting of a G alpha subunit (Gi1 alpha) and c-Src tyrosine kinase to caveolae membranes: clarifying the role of N-myristoylation. *Cell. Mol. Biol.* 43, 293–303.
- Sorice, M., Manganelli, V., Matarrese, P., Tinari, A., Misasi, R., Malorni, W., et al. (2009). Cardiolipin-enriched raft-like microdomains are essential activating platforms for apoptotic signals on mitochondria. *FEBS Lett.* 583, 2447–2450. doi: 10.1016/j.febslet.2009.07.018
- Sorrentino, G., Ruggeri, N., Specchia, V., Cordenonsi, M., Mano, M., Dupont, S., et al. (2014). Metabolic control of YAP and TAZ by the mevalonate pathway. *Nat. Cell. Biol.* 16, 357–366. doi: 10.1038/ncb2936
- Soto-Acosta, R., Bautista-Carbajal, P., Cervantes-Salazar, M., Angel-Ambrocio, A.H., and Del Angel, R.M. (2017). DENV up-regulates the HMG-CoA reductase activity through the impairment of AMPK phosphorylation: a potential antiviral target. *PLoS Pathog.* 13:e1006257. doi: 10.1371/journal.ppat. 1006257
- Staubach, S., and Hanisch, F.G. (2011). Lipid rafts: signaling and sorting platforms of cells and their roles in cancer. *Exp. Rev. Proteomics* 8, 263–277. doi: 10.1586/ epr.11.2
- Staubach, S., Razawi, H., and Hanisch, F.G. (2009). Proteomics of MUC1containing lipid rafts from plasma membranes and exosomes of human breast carcinoma cells MCF-7. *Proteomics* 9, 2820–2835. doi: 10.1002/pmic. 200800793

- Stopsack, K.H., Gerke, T.A., Sinnott, J.A., Penney, K.L., Tyekucheva, S., Sesso, H.D., et al. (2016). Cholesterol metabolism and prostate cancer lethality. *Cancer Res.* 76, 4785–4790. doi: 10.1158/0008-5472.CAN-16-0903
- Sugii, S., Reid, P.C., Ohgami, N., Du, H., and Chang, T.Y. (2003). Distinct endosomal compartments in early trafficking of low density lipoproteinderived cholesterol. *J. Biol. Chem.* 278, 27180–27189. doi: 10.1074/jbc.M30054 2200
- Sui, Z., Zhou, J., Cheng, Z., and Lu, P. (2015). Squalene epoxidase (SQLE) promotes the growth and migration of the hepatocellular carcinoma cells. *Tumour Biol.* 36, 6173–6179. doi: 10.1007/s13277-015-3301-x
- Szajnman, S.H., Ravaschino, E.L., Docampo, R., and Rodriguez, J.B. (2005). Synthesis and biological evaluation of 1-amino-1,1-bisphosphonates derived from fatty acids against *Trypanosoma* cruzi targeting farnesyl pyrophosphate synthase. *Bioorg. Med. Chem. Lett.* 15, 4685–4690. doi: 10.1016/j.bmcl.2005. 07.060
- Thurnher, M., Nussbaumer, O., and Gruenbacher, G. (2012). Novel aspects of mevalonate pathway inhibitors as antitumor agents. *Clin. Cancer Res.* 18, 3524–3531. doi: 10.1158/1078-0432.CCR-12-0489
- Timmins, J.M., Lee, J.Y., Boudyguina, E., Kluckman, K.D., Brunham, L.R., Mulya, A., et al. (2005). Targeted inactivation of hepatic Abcal causes profound hypoalphalipoproteinemia and kidney hypercatabolism of apoA-I. J. Clin. Invest. 115, 1333–1342. doi: 10.1172/JCI23915
- Vaupel, P., Kelleher, D.K., and Hockel, M. (2001). Oxygen status of malignant tumors: pathogenesis of hypoxia and significance for tumor therapy. *Semin. Oncol.* 28, 29–35. doi: 10.1016/s0093-7754(01)90210-6
- Vedhachalam, C., Duong, P.T., Nickel, M., Nguyen, D., Dhanasekaran, P., Saito, H., et al. (2007). Mechanism of ATP-binding cassette transporter A1mediated cellular lipid efflux to apolipoprotein A-I and formation of high density lipoprotein particles. J. Biol. Chem. 282, 25123–25130. doi: 10.1074/jbc. M704590200
- Venkateswaran, A., Laffitte, B.A., Joseph, S.B., Mak, P.A., Wilpitz, D.C., Edwards, P.A., et al. (2000). Control of cellular cholesterol efflux by the nuclear oxysterol receptor LXR alpha. *Proc. Natl. Acad. Sci. U S A.* 97, 12097–12102. doi: 10.1073/ pnas.200367697
- Vickers, K.C., Landstreet, S.R., Levin, M.G., Shoucri, B.M., Toth, C.L., Taylor, R.C., et al. (2014). MicroRNA-223 coordinates cholesterol homeostasis. *Proc. Natl. Acad. Sci. U S A.* 111, 14518–14523. doi: 10.1073/pnas.1215767111
- Villa, G.R., Hulce, J.J., Zanca, C., Bi, J., Ikegami, S., Cahill, G.L., et al. (2016). An LXR-cholesterol axis creates a metabolic co-dependency for brain cancers. *Cancer Cell.* 30, 683–693. doi: 10.1016/j.ccell.2016.09.008
- Wagschal, A., Najafi-Shoushtari, S.H., Wang, L., Goedeke, L., Sinha, S., deLemos, A.S., et al. (2015). Genome-wide identification of microRNAs regulating cholesterol and triglyceride homeostasis. *Nat. Med.* 21, 1290–1297. doi: 10.1038/ nm.3980
- Wang, Y., Rogers, P.M., Su, C., Varga, G., Stayrook, K.R., and Burris, T.P. (2008). Regulation of cholesterologenesis by the oxysterol receptor, LXRalpha. J. Biol. Chem. 283, 26332–26339. doi: 10.1074/jbc.M804808200
- Warburg, O. (1925). The metabolism of carcinoma cells. J. Cancer Res. 9, 148–163. Warburg, O. (1956). On the origin of cancer cells. Science 123, 309–314. doi:
- 10.1126/science.123.3191.309 Warita, K., Warita, T., Beckwitt, C.H., Schurdak, M.E., Vazquez, A., Wells, A., et al. (2014). Statin-induced mevalonate pathway inhibition attenuates the growth of mesenchymal-like cancer cells that lack functional E-cadherin mediated cell cohesion. *Sci. Rep.* 4:7593. doi: 10.1038/srep07593
- Watson, P.A., Arora, V.K., and Sawyers, C.L. (2015). Emerging mechanisms of resistance to androgen receptor inhibitors in prostate cancer. *Nat. Rev. Cancer* 15, 701–711. doi: 10.1038/nrc4016
- Wilde, L., Roche, M., Domingo-Vidal, M., Tanson, K., Philp, N., Curry, J., et al. (2017). Metabolic coupling and the reverse warburg effect in cancer: implications for novel biomarker and anticancer agent development. *Semin. Oncol.* 44, 198–203. doi: 10.1053/j.seminoncol.2017.10.004
- Wu, J., Guo, L., Qiu, X., Ren, Y., Li, F., Cui, W., et al. (2020). Genkwadaphnin inhibits growth and invasion in hepatocellular carcinoma by blocking DHCR24-mediated cholesterol biosynthesis and lipid rafts formation. *Br. J. Cancer* 123, 1673–1685. doi: 10.1038/s41416-020-01085-z
- Xie, C., Li, N., Chen, Z.J., Li, B.L., and Song, B.L. (2011). The small GTPase Cdc42 interacts with niemann-pick C1-like 1 (NPC1L1) and controls its movement

from endocytic recycling compartment to plasma membrane in a cholesteroldependent manner. *J. Biol. Chem.* 286, 35933–35942. doi: 10.1074/jbc.M111. 270199

- Xu, L., Guo, T., Qu, X., Hu, X., Zhang, Y., Che, X., et al. (2018). beta-elemene increases the sensitivity of gastric cancer cells to TRAIL by promoting the formation of DISC in lipid rafts. *Cell. Biol. Int.* 42, 1377–1385. doi: 10.1002/ cbin.11023
- Xu, L., Qu, X., Luo, Y., Zhang, Y., Liu, J., Qu, J., et al. (2011). Epirubicin enhances TRAIL-induced apoptosis in gastric cancer cells by promoting death receptor clustering in lipid rafts. *Mol. Med. Rep.* 4, 407–411. doi: 10.3892/mmr.2011.439
- Xu, L., Qu, X., Zhang, Y., Hu, X., Yang, X., Hou, K., et al. (2009). Oxaliplatin enhances TRAIL-induced apoptosis in gastric cancer cells by CBL-regulated death receptor redistribution in lipid rafts. *FEBS Lett.* 583, 943–948. doi: 10. 1016/j.febslet.2009.02.014
- Xu, X.D., Shao, S.X., Jiang, H.P., Cao, Y.W., Wang, Y.H., Yang, X.C., et al. (2015). Warburg effect or reverse warburg effect? A review of cancer metabolism. Oncol. Res. Treat 38, 117–122. doi: 10.1159/000375435
- Xu, Z., Wang, T., Song, H., and Jiang, X. (2020). Flotillin-2 predicts poor prognosis and promotes tumor invasion in intrahepatic cholangiocarcinoma. *Oncol. Lett.* 19, 2243–2250. doi: 10.3892/ol.2020.11349
- Yamaguchi, H., Takeo, Y., Yoshida, S., Kouchi, Z., Nakamura, Y., and Fukami, K. (2009). Lipid rafts and caveolin-1 are required for invadopodia formation and extracellular matrix degradation by human breast cancer cells. *Cancer Res.* 69, 8594–8602. doi: 10.1158/0008-5472.CAN-09-2305
- Yamauchi, Y., Furukawa, K., Hamamura, K., and Furukawa, K. (2011). Positive feedback loop between PI3K-Akt-mTORC1 signaling and the lipogenic pathway boosts Akt signaling: induction of the lipogenic pathway by a melanoma antigen. *Cancer Res.* 71, 4989–4997. doi: 10.1158/0008-5472.CAN-10-4108
- Yamauchi, Y., Iwamoto, N., Rogers, M.A., Abe-Dohmae, S., Fujimoto, T., Chang, C.C., et al. (2015). Deficiency in the lipid exporter ABCA1 impairs retrograde sterol movement and disrupts sterol sensing at the endoplasmic reticulum. *J. Biol. Chem.* 290, 23464–23477. doi: 10.1074/jbc.M115.662668
- Yamauchi, Y., Reid, P.C., Sperry, J.B., Furukawa, K., Takeya, M., Chang, C.C., et al. (2007). Plasma membrane rafts complete cholesterol synthesis by participating in retrograde movement of precursor sterols. *J. Biol. Chem.* 282, 34994–35004. doi: 10.1074/jbc.M703653200
- Yamauchi, Y., and Rogers, M.A. (2018). Sterol metabolism and transport in atherosclerosis and cancer. *Front. Endocrinol. (Lausanne)* 9:509. doi: 10.3389/ fendo.2018.00509
- Yan, S., Qu, X., Xu, L., Che, X., Ma, Y., Zhang, L., et al. (2014). Bufalin enhances TRAIL-induced apoptosis by redistributing death receptors in lipid rafts in breast cancer cells. *Anticancer Drugs* 25 683–689. doi: 10.1097/CAD. 000000000000095
- Yang, B., Guo, M., Herman, J.G., and Clark, D.P. (2003). Aberrant promoter methylation profiles of tumor suppressor genes in hepatocellular carcinoma. *Am. J. Pathol.* 16, 1101–1107. doi: 10.1016/S0002-9440(10)63469-4
- Yang, H., Guan, L., Li, S., Jiang, Y., Xiong, N., Li, L., et al. (2016). Mechanosensitive caveolin-1 activation-induced PI3K/Akt/mTOR signaling pathway promotes breast cancer motility, invadopodia formation and metastasis in vivo. *Oncotarget* 7, 16227–16247. doi: 10.18632/oncotarget.7583
- Yang, J., Wang, L., and Jia, R. (2020). Role of de novo cholesterol synthesis enzymes in cancer. J. Cancer 11, 1761–1767. doi: 10.7150/jca.38598
- Yang, Y.F., Jan, Y.H., Liu, Y.P., Yang, C.J., Su, C.Y., Chang, Y.C., et al. (2014). Squalene synthase induces tumor necrosis factor receptor 1 enrichment in lipid rafts to promote lung cancer metastasis. *Am. J. Respir. Crit. Care Med.* 190, 675–687. doi: 10.1164/rccm.201404-0714OC
- Yang, Z., Qin, W., Chen, Y., Yuan, B., Song, X., Wang, B., et al. (2018). Cholesterol inhibits hepatocellular carcinoma invasion and metastasis by promoting CD44 localization in lipid rafts. *Cancer Lett.* 429, 66–77. doi: 10.1016/j.canlet.2018. 04.038
- Ye, D.M., Ye, S.C., Yu, S.Q., Shu, F.F., Xu, S.S., Chen, Q.Q., et al. (2019). Drug-resistance reversal in colorectal cancer cells by destruction of flotillins,

the key lipid rafts proteins. *Neoplasma* 66, 576–583. doi: 10.4149/neo_2018_180820N633

- Yoshioka, H., Coates, H.W., Chua, N.K., Hashimoto, Y., Brown, A.J., and Ohgane, K. (2020). A key mammalian cholesterol synthesis enzyme, squalene monooxygenase, is allosterically stabilized by its substrate. *Proc. Natl. Acad. Sci.* USA. 117, 7150–7158. doi: 10.1073/pnas.1915923117
- Yu, L., Bharadwaj, S., Brown, J.M., Ma, Y., Du, W., Davis, M.A., et al. (2006). Cholesterol-regulated translocation of NPC1L1 to the cell surface facilitates free cholesterol uptake. *J. Biol. Chem.* 281, 6616–6624. doi: 10.1074/jbc. M511123200
- Yue, S., Li, J., Lee, S.Y., Lee, H.J., Shao, T., Song, B., et al. (2014). Cholesteryl ester accumulation induced by PTEN loss and PI3K/AKT activation underlies human prostate cancer aggressiveness. *Cell. Metab.* 19, 393–406. doi: 10.1016/j. cmet.2014.01.019
- Yun, U.J., Lee, J.H., Shim, J., Yoon, K., Goh, S.H., Yi, E.H., et al. (2019). Anticancer effect of doxorubicin is mediated by downregulation of HMG-Co A reductase via inhibition of EGFR/Src pathway. *Lab. Invest.* 99, 1157–1172. doi: 10.1038/s41374-019-0193-1
- Zang, M., Waelde, C.A., Xiang, X., Rana, A., Wen, R., and Luo, Z. (2001). Microtubule integrity regulates Pak leading to Ras-independent activation of Raf-1. insights into mechanisms of Raf-1 activation. J. Biol. Chem. 276, 25157–25165. doi: 10.1074/jbc.M100152200
- Zelcer, N., Hong, C., Boyadjian, R., and Tontonoz, P. (2009). LXR regulates cholesterol uptake through Idol-dependent ubiquitination of the LDL receptor. *Science* 325, 100–104. doi: 10.1126/science.1168974
- Zhang, D.W., Garuti, R., Tang, W.J., Cohen, J.C., and Hobbs, H.H. (2008). Structural requirements for PCSK9-mediated degradation of the low-density lipoprotein receptor. *Proc. Natl. Acad. Sci. U S A.* 105, 13045–13050. doi: 10. 1073/pnas.0806312105
- Zhang, X., Song, Y., Feng, M., Zhou, X., Lu, Y., Gao, L., et al. (2015). Thyroidstimulating hormone decreases HMG-CoA reductase phosphorylation via AMP-activated protein kinase in the liver. J. Lipid Res. 56, 963–971. doi: 10. 1194/jlr.M047654
- Zhu, X., Lee, J.Y., Timmins, J.M., Brown, J.M., Boudyguina, E., Mulya, A., et al. (2008). Increased cellular free cholesterol in macrophage-specific Abca1 knockout mice enhances pro-inflammatory response of macrophages. J. Biol. Chem. 283, 22930–22941. doi: 10.1074/jbc.M801408200
- Zhuang, L., Kim, J., Adam, R.M., Solomon, K.R., and Freeman, M.R. (2005). Cholesterol targeting alters lipid raft composition and cell survival in prostate cancer cells and xenografts. J. Clin. Invest. 115, 959–968. doi: 10.1172/JCI19935
- Zhuang, L., Lin, J., Lu, M.L., Solomon, K.R., and Freeman, M.R. (2002). Cholesterol-rich lipid rafts mediate akt-regulated survival in prostate cancer cells. *Cancer Res.* 62, 2227–2231.
- Zidovetzki, R., and Levitan, I. (2007). Use of cyclodextrins to manipulate plasma membrane cholesterol content: evidence, misconceptions and control strategies. *Biochim. Biophy.s Acta* 1768, 1311–1324. doi: 10.1016/j.bbamem. 2007.03.026
- Zimber, A., Chedeville, A., Abita, J.P., Barbu, V., and Gespach, C. (2000). Functional interactions between bile acids, all-trans retinoic acid, and 1,25dihydroxy-vitamin D3 on monocytic differentiation and myeloblastin gene down-regulation in HL60 and THP-1 human leukemia cells. *Cancer Res.* 60, 672–678.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Vona, Iessi and Matarrese. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.