



Role of Endothelial Dysfunction in Cardiovascular Diseases: The Link Between Inflammation and Hydrogen Sulfide

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Specialty section:

This article was submitted to
Inflammation Pharmacology,
a section of the journal
Frontiers in Pharmacology

Received: 23 August 2019

Accepted: 03 December 2019

Published: 21 January 2020

Citation:

Sun H-J, Wu Z-Y, Nie X-W and
Bian J-S (2020) Role of Endothelial
Dysfunction in Cardiovascular
Diseases: The Link Between
Inflammation and Hydrogen Sulfide.
Front. Pharmacol. 10:1568.
doi: 10.3389/fphar.2019.01568

Endothelial cells are important constituents of blood vessels that play critical roles in cardiovascular homeostasis by regulating blood fluidity and fibrinolysis, vascular tone, angiogenesis, monocyte/leukocyte adhesion, and platelet aggregation. The normal vascular endothelium is taken as a gatekeeper of cardiovascular health, whereas abnormality of vascular endothelium is a major contributor to a plethora of cardiovascular ailments, such as atherosclerosis, aging, hypertension, obesity, and diabetes. Endothelial dysfunction is characterized by imbalanced vasodilation and vasoconstriction, elevated reactive oxygen species (ROS), and proinflammatory factors, as well as deficiency of nitric oxide (NO) bioavailability. The occurrence of endothelial dysfunction disrupts the endothelial barrier permeability that is a part of inflammatory response in the development of cardiovascular diseases. As such, abrogation of endothelial cell activation/inflammation is of clinical relevance. Recently, hydrogen sulfide (H₂S), an entry as a gasotransmitter, exerts diverse biological effects through acting on various targeted signaling pathways. Within the cardiovascular system, the formation of H₂S is detected in smooth muscle cells, vascular endothelial cells, and cardiomyocytes. Disrupted H₂S bioavailability is postulated to be a new indicator for endothelial cell inflammation and its associated endothelial dysfunction. In this review, we will summarize recent advances about the roles of H₂S in endothelial cell homeostasis, especially under pathological conditions, and discuss its putative therapeutic applications in endothelial inflammation-associated cardiovascular disorders.

Keywords: endothelial cell, gasotransmitters, hydrogen sulfide, inflammation, cardiovascular disease

INTRODUCTION

Currently, cardiovascular disease is identified to be a major cause of people death around the world, and this situation is estimated to remain for many years to come, thus bringing a considerable burden to the world's health resource (Mathers and Loncar, 2006). It is well known that poor diet, smoking, obesity, and physical inactivity are various modifiable risk factors for cardiovascular diseases, all of which lead to a proinflammatory state (Allende-Vigo, 2010). Actually, a wide range of

evidence supports a crucial role of inflammatory response in the pathogenesis of cardiovascular diseases through driving endothelial cell activation/dysfunction (Carter, 2012). Therefore, it is not unexpected that huge efforts have been made to identify therapeutically potential targets to halt endothelial cell inflammation.

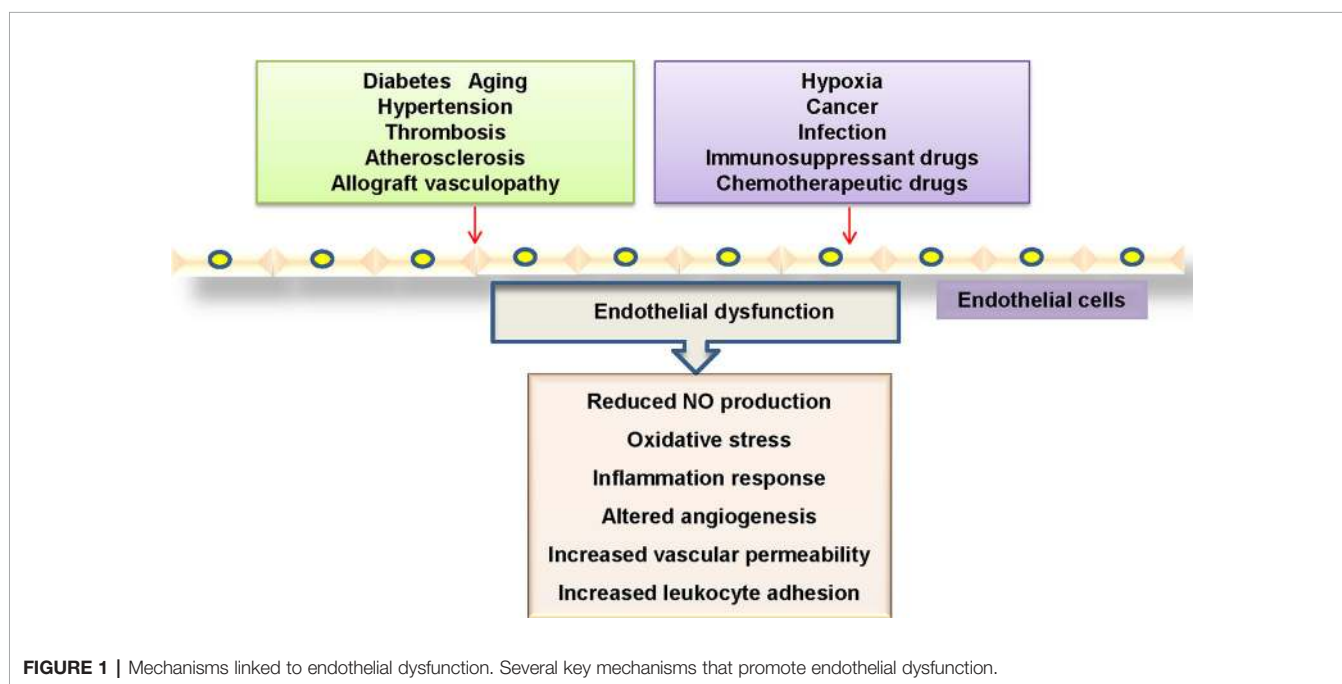
The blood vessels are composed of connective tissue, fibroblasts, endothelial cells, and vascular smooth muscle cells (VSMCs). On the innermost side of blood vessels, the normal endothelium is a semipermeable layer between blood stream and blood vessel wall. Due to its tight specialized cell-to-cell junctions, the endothelium forms a barrier that selectively limits the movement of macromolecules (Rahimi, 2017). The barrier is critically involved in vascular tone, fluid homeostasis, and host defense (Zhang et al., 2018b). Activated endothelial cells may release various cytokines, chemokines, and growth factors that promote the proliferation, migration, and permeability of endothelial cells (Park-Windhol and D'amore, 2016). The endothelial cells with inflammatory phenotype cause inflammation in the blood vessels, resulting in endothelial dysfunction and following progression of cardiovascular diseases (Sun et al., 2016). In accordance with this notion, endothelial cell inflammation is directly responsible for various cardiovascular diseases, such as hypertension, atherosclerosis, aging, stroke, heart disease, diabetes, obesity, venous thrombosis, and intimal hyperplasia (Sun et al., 2017; Castro-Ferreira et al., 2018; Haybar et al., 2019; Zhong et al., 2019).

In the endothelium, hydrogen sulfide (H_2S), the third endogenous gaseous molecule after nitric oxide (NO) and carbon monoxide (CO), is synthesized and observed (Pan et al., 2017). Over the last decade, the roles of H_2S in the pathogenesis of endothelial dysfunction have grown

exponentially. As a result, the current understanding of H_2S -mediated endothelial cell functions in both health and disease continues to deepen. However, the potential molecular mechanisms that underlie H_2S -mediated cardiovascular homeostasis, especially endothelial inflammation, are not comprehensively elucidated. The present review focuses on the current progress regarding the roles of H_2S in endothelial inflammation-related cardiovascular disorders including hypertension, atherosclerosis and diabetes. Furthermore, we will discuss the possible challenges for developing H_2S -derived therapeutics to treat endothelial dysfunction in cardiovascular disorders.

ENDOTHELIAL DYSFUNCTION AND INFLAMMATION

The dysfunction of endothelial cells in the vasculature is profoundly implicated in the pathogenesis of cardiovascular disorders (Boulanger, 2016). Mounting evidence has shown that endothelial cell dysfunction is characterized by disrupted vascular tone and redox balance, and increased inflammatory reactions within the blood vessel wall (Ooi et al., 2018) (Figure 1). Most commonly, the impaired endothelium-dependent vasodilatation is defined as a hallmark of endothelial dysfunction, which is critically responsible for several cardiovascular disorders including diabetes mellitus, hypertension, atherosclerosis, aging and heart failure (Leung and Vanhoutte, 2017; Suryavanshi and Kulkarni, 2017). More recently, endothelial activation is also a prominent alteration in endothelial dysfunction, which refers to the upregulations of chemokines and adhesion molecules and other proteins involved



in cell–cell interactions (Weber and Noels, 2011; Ng et al., 2018), thus leading to the prothrombotic and proinflammatory circumstance in the blood vessels.

In activated endothelial cells, the expressions of proinflammatory cytokines, chemokines, enzymes, and adhesion molecules are substantially upregulated (Baghai et al., 2018). It is highly possible that endothelial cell inflammation plays an important role in the pathogenesis of endothelial dysfunction in cardiovascular disorders. Therefore, identification of endothelial cell-derived inflammatory factors and its underlying mechanisms may be effective in preventing the progression of cardiovascular diseases.

REGULATION OF ENDOTHELIAL FUNCTION BY H₂S UNDER PHYSIOLOGICAL CONDITION

As the third endogenous gasotransmitter, H₂S is primarily synthesized in mammalian tissues through enzymatic or non-enzymatic pathways (Li et al., 2011; Liu et al., 2012). The majority of endogenous H₂S is produced by three enzymes including cystathionine γ -lyase (CSE), cystathionine β -synthase (CBS) and 3-mercaptopyruvate sulfurtransferase (3-MST) in mammalian tissues (Liu et al., 2011) (Figure 2). In the vascular endothelium, H₂S is synthesized *via* the enzymatic metabolism of

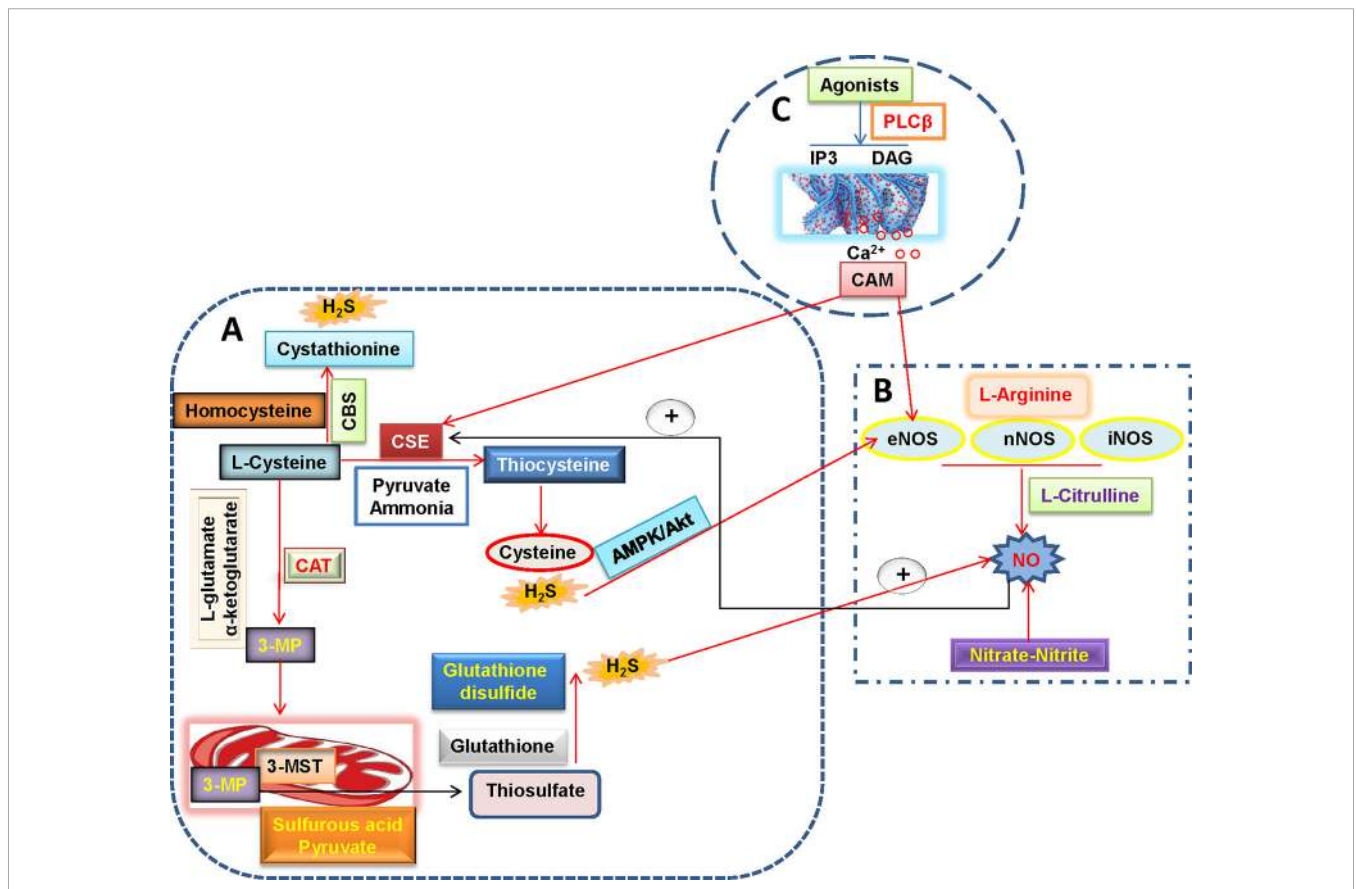


FIGURE 2 | H₂S and NO biosynthetic pathways in blood vessels. **(A)** L-cysteine is the substrate for the formation of H₂S through three H₂S-producing enzymes, L-cysteine is catalyzed by CSE to produce pyruvate, ammonia, and thiocysteine, the latter is then decomposed to cysteine and H₂S. The endogenous H₂S production by CBS is related with the condensation of homocysteine with L-cysteine, followed by the formation of cystathionine and H₂S. Direct reaction of L-cysteine and α -ketoglutarate by CAT yields the release of 3-MP and L-glutamate, 3-Mercaptopyruvate is transported into the mitochondria where it is catalyzed to sulfuric acid, pyruvate and thiosulfate by 3-MST. In the presence of reduced glutathione, the thiosulfate is reduced to glutathione disulfide and H₂S. It is well accepted that H₂S can increase eNOS activity and thereby subsequent NO production directly or through AMPK/Akt signaling pathway. **(B)** NO is produced in all tissues by NOS-dependent (L-arginine-NO pathway) and -independent (nitrate-nitrite-NO pathway) pathways. A recently discovered pathway for NO generation is the serial reduction of the inorganic anions nitrate and nitrite. With the assistance of three isoforms of NOS including nNOS, eNOS, and iNOS, L-arginine is oxidized into L-citrulline with NO. NO is found to increase CSE activity and expression and then stimulate H₂S production. **(C)** In endothelial cells, vasoconstrictor agonists stimulate the release of Ca²⁺ and cause formation of calcium-calmodulin (CaM) *via* the PLC β /IP₃/DAG pathway. Then, CaM can simultaneously activate eNOS and CSE that yield NO and H₂S, respectively. H₂S, hydrogen sulfide; NO, nitric oxide; 3-MP, 3-mercaptopyruvate; CAT, cysteine aminotransferase; CSE, cystathionine γ -lyase; CBS, cystathionine β -synthase; 3-MST, 3-mercaptopyruvate sulfurtransferase; CaM, calcium-calmodulin; PLC β , phospholipase C β ; IP₃, inositol-3-phosphate (IP₃); DAG, diacylglycerol (DAG); eNOS, endothelial NO synthase; iNOS, inducible NO synthase; nNOS, neuronal NO synthase.

CBS/CSE using cysteine as the substrates (Tao et al., 2017; Mitidieri and Gurgone, 2019). Likewise, the involvement of 3-MST and cysteine aminotransferase (CAT) in endothelial generation of H₂S has been demonstrated (Wang, 2012).

The regulation of vascular tone by H₂S may be dependent on endothelium-independent and -dependent manners (Wang et al., 2015b). In the vasculature, H₂S has been shown to induce vasodilation in aorta (Zhao et al., 2001), gastric artery (Kubo et al., 2007), mesenteric artery (Cheng et al., 2004), and internal mammary artery (Webb et al., 2008). The underlying mechanism by which H₂S relaxes blood vessels is related with activation of vascular smooth muscle ATP-sensitive K⁺ (KATP) channels (Zhao et al., 2001), independently of the endothelium. The involvement of KATP channels in H₂S-induced vasodilation is further confirmed by a finding that this relaxation is partially blocked by an inhibitor of KATP channels glibenclamide (Webb et al., 2008). Despite of these results, the exact mechanism of how KATP channels are directly activated by H₂S still remains unknown. It is also reported that 4-aminopyridine-sensitive K⁺ channels are involved in H₂S-induced relaxation in the rat coronary artery (Cheang et al., 2010). The H₂S donor sodium hydrosulfide (NaHS) induces concentration-dependent vasorelaxation in both mesenteric arteries and aortas, which is blocked by the KCNQ-type K_v channel inhibitor XE991, suggesting the involvement of KCNQ channels in H₂S-mediated peripheral artery relaxation (Schleifenbaum et al., 2010). Moreover, Ca²⁺ channels or sparks (Jackson-Weaver et al., 2015), Cl(-)/HCO(3)(-) channels (Kiss et al., 2008), the NO pathway (Ali et al., 2006), phospholipase A2 (D'emmanuele Di Villa Bianca et al., 2011), transient receptor potential (TRP) channels (White et al., 2013), and metabolic/mitochondrial effects (Kiss et al., 2008), are also suggested to be implicated in H₂S-induced vasorelaxation. H₂S appears to play an important role in vasorelaxation *via* multidimensional mechanisms. In the endothelium, recent studies have provided several lines of evidence to support that H₂S might function as an endothelium-derived relaxing factor (EDRF), which shares many common traits with other EDRFs (Wang, 2009). Interestingly, the vasorelaxation actions of H₂S are more remarkable in peripheral resistance arteries than in large-conduit arteries, the effects require the membrane hyperpolarization of both VSMCs and endothelial cells, as well as activation of endothelial intermediate conductance (IK(Ca)) and small conductance (SK(Ca)) potassium channels (Mustafa et al., 2011; Tang et al., 2013). The definition of H₂S as an endogenous EDHF might shed light on possible therapeutic effects of H₂S on pathological abnormalities in the vascular system. Still, more extensive and mechanistic studies are needed to determine whether H₂S is a new EDRF in the future.

The endothelial cells also orchestrate tube formation and angiogenesis (Watson et al., 2017). H₂S is reported to stimulate endothelial proliferation, migration, and angiogenesis (Wang et al., 2010b) (Figure 3). Furthermore, administration of H₂S promotes angiogenesis in the Matrigel plug assay (Cai et al., 2007). However, it should be pointed out that high dose of H₂S loses the ability to induce angiogenesis (Cai et al., 2007). In a rat

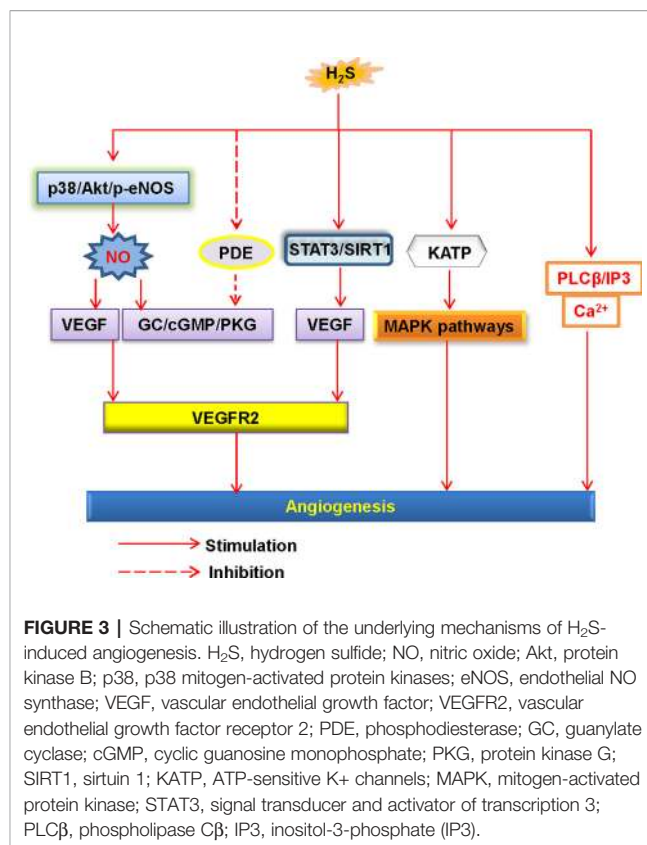


FIGURE 3 | Schematic illustration of the underlying mechanisms of H₂S-induced angiogenesis. H₂S, hydrogen sulfide; NO, nitric oxide; Akt, protein kinase B; p38, p38 mitogen-activated protein kinases; eNOS, endothelial NO synthase; VEGF, vascular endothelial growth factor; VEGFR2, vascular endothelial growth factor receptor 2; PDE, phosphodiesterase; GC, guanylate cyclase; cGMP, cyclic guanosine monophosphate; PKG, protein kinase G; SIRT1, sirtuin 1; KATP, ATP-sensitive K⁺ channels; MAPK, mitogen-activated protein kinase; STAT3, signal transducer and activator of transcription 3; PLCβ, phospholipase Cβ; IP3, inositol-3-phosphate (IP3).

model of chronic hindlimb ischaemia, intraperitoneal injection of the H₂S donor NaHS at the lower dose significantly improves capillary density, angiographic scores, thus improving hindlimb blood flow (Wang et al., 2010a). In line with the results discussed earlier, higher dose of the H₂S donor is found to be ineffective in this model (Wang et al., 2010a). On these grounds, we speculate that the effects of H₂S donors in angiogenesis may range from physiological, cytoprotective effects (low concentration) to cytotoxic effects (which are generally apparent at higher concentrations) (Szabo and Papapetropoulos, 2011). From a genetic perspective, mutant mice lacking CSE exhibit a variety of pathological features, including delayed wound healing secondary to inhibition of angiogenesis (Papapetropoulos et al., 2009). It has been reviewed that several cellular signaling pathways, such as the PI3K/Akt pathway, the mitogen activated protein kinase (MAPK) pathway, and ATP-sensitive potassium channels, are involved in H₂S-mediated angiogenic effects (Szabo and Papapetropoulos, 2011). In addition to this, further study has demonstrated that H₂S specifically disrupts cys1045-cys1024 disulfide bond in vascular endothelial growth factor receptor 2 (VEGFR2) and then stimulates its conformation for angiogenesis (Tao et al., 2013). As a molecular switch, H₂S is also reported to activate signal transducer and activator of transcription 3 (STAT3) (Kan et al., 2014), mammalian target of rapamycin (mTOR), and the VEGFR2 pathway (Zhou et al., 2016), then the endothelial cell proliferation and angiogenesis are observed. It is noteworthy that

due to its proangiogenic effects, H₂S might lead to pathological angiogenesis in atherosclerotic plaques, thus facilitating plaque vulnerability (Van Den Born et al., 2016). In spite of this, therapeutic angiogenesis is important for wound healing, organ ischaemia, or the reperfusion of previously ischaemic organs (Caporali and Emanuelli, 2011; Dulmovits and Herman, 2012; Ng et al., 2018). For this reason, the reparative angiogenesis by H₂S may provide novel therapeutic avenues for post-ischemic neovascularization. Due to the physiological importance of H₂S in the endothelium, further research is indispensable to examine the novel roles of endogenous H₂S in the regulation of cardiovascular functions.

ROLE OF H₂S IN ENDOTHELIAL INFLAMMATION

Using intravital microscopy, H₂S donors are found to attenuate the leukocyte adherence in rat mesenteric arteries induced by aspirin, this effect may be likely dependent on activation of KATP channels (Zanardo et al., 2006; Zuidema and Korthuis, 2015). In accordance with this, blockade of endogenous H₂S exacerbates leukocyte-mediated inflammation in the endothelium (Zanardo et al., 2006). By contrast, NaHS promotes leukocyte rolling and adherence in mesenteric venules of mice with cecal ligation and puncture (CLP)-induced sepsis (Zhang et al., 2007). These conflicting results imply that H₂S acts as a pivotal regulator of leukocyte activation under different inflammatory states. However, in recent years, more studies support that H₂S could inhibit the process of endothelial cell inflammation (Wen et al., 2018). For instance, specific endothelial deletion of CSE is associated with the development of endothelial inflammation and atherosclerosis, effects that are reversed on treatment with a polysulfide donor (Bibli et al., 2019). H₂S treatment reduces the increases in inflammatory mediators such as vascular cell adhesionmolecule-1 (VCAM-1), intercellular adhesionmolecule-1 (ICAM-1) and monocyte chemoattractant protein-1 (MCP-1) in endothelial cell induced by tumour necrosis factor- α (TNF- α), and the underlying mechanism of this protective effect is primarily mediated by inhibition of soluble TNF- α shedding and its relevant MCP-1 release (Perna et al., 2013). Similarly, exogenous H₂S attenuates Ang II-induced inflammation response *via* inhibition of the nuclear transcription factor- κ B (NF- κ B) signaling pathway in endothelial cells (Hu et al., 2016). Inhibition of the NF- κ B pathway is also required for H₂S to attenuate pulmonary endothelial cell inflammation and subsequent pulmonary hypertension (Feng et al., 2017b). Endogenous H₂S could directly induce sirtuin1 (SIRT1) sulfhydration and stability, thus reducing aortic inflammation and atherosclerotic plaque formation (Du et al., 2019). Deficiency of CSE increases endogenous sulfur dioxide (SO₂) level in endothelial cells, and blockade of endogenous SO₂ aggravates CSE knockdown-induced NF- κ B pathway and its downstream inflammatory factors release in endothelial cells (Zhang et al., 2018a), suggesting that the increased endogenous SO₂ generation might act as a compensatory mechanism for the downregulated CSE/H₂S

pathway in endothelial inflammatory response (Zhang et al., 2018a). It is concluded that the anti-inflammatory effects of H₂S donors show tremendous promise for the treatment of endothelial inflammation-related cardiovascular disorders.

In response to proinflammatory cytokines, the leukocyte or macrophages are activated and recruited to the endothelium, thus causing the development of endothelial dysfunction-related cardiovascular diseases (Fang et al., 2013). Fortunately, H₂S is found to alleviate vascular inflammation through various signaling pathways, including inhibition of NF- κ B and nucleotide-binding oligomerization domain, leucine rich repeat, and pyrin domain-containing protein 3 (NLRP3) inflammasome, activation of KATP channels and voltage- and calcium-gated potassium (BKCa) channels (Fiorucci et al., 2005; Zanardo et al., 2006; Zuidema et al., 2010; Altaany et al., 2014; Bourque et al., 2018; Li et al., 2019). These possible mechanisms of H₂S may explain that H₂S can diminish vascular inflammation and attenuate the vascular injury, suggesting that the anti-inflammatory effect of H₂S is a benefit for cardiovascular protection. Next, we will discuss the beneficial roles of H₂S-mediated suppression of endothelial dysfunction in cardiovascular disorders including atherosclerosis, diabetic cardiovascular complications and hypertension.

H₂S-RELATED ENDOTHELIAL DYSFUNCTION IN ATHEROSCLEROSIS

Atherosclerosis, a chronic vascular disease of large and medium arteries, involves various risk factors including lipid deposition, hypertension, inflammatory factors, and hyperhomocysteinemia, which synergistically elicit endothelial dysfunction (Baszczuk et al., 2014). Biochemical effects of these factors on the endothelium could lead to endothelial cell damage and vascular remodeling (Baszczuk et al., 2014). This important event induces endothelial inflammation, macrophage differentiation, foam cell formation, platelet deposition, and thrombus formation (Chistiakov et al., 2017; Rahman and Woollard, 2017). As such, correction of endothelial dysfunction could be a therapeutic strategy for management of atherosclerosis.

In recent years, considerable evidence indicates that the downregulated CSE/H₂S pathway plays a pathophysiologic role in the development of atherosclerosis (Kanagy et al., 2017; Fan et al., 2019). CSE-knockout mice fed with atherogenic diet exhibit more severe atherosclerosis, suggesting that the disturbed CSE/H₂S pathway predisposes the animals to the development of atherosclerosis (Mani et al., 2013). Macrophage inflammation directly contributes to necrotic core formation and plaque instability in atherosclerosis (Kavurma et al., 2017). In oxidized low density lipoprotein (ox-LDL)-treated macrophage, the levels of CSE mRNA and protein expression, as well as H₂S production are remarkably decreased, thus, this finding indicates that alterations of the CSE/H₂S pathway plays an important role in ox-LDL-stimulated macrophage inflammation and atherosclerosis (Wang et al., 2013b). It is worth noting that

CBS deficiency may cause hyperhomocysteinemia, which is an independent risk factor for the development of atherosclerosis (Zhang et al., 2012a; Yuan et al., 2017). In transgenic CBS-deficient mice, the emergence of hypercholesterolemia accelerates atherosclerotic lesions via oxidative stress and inflammatory monocyte generation (Zhang et al., 2009). On the other hand, vascular calcification and neointimal hyperplasia are also involved in the progression of atherosclerosis (Durham et al., 2018; Yu et al., 2018). Not surprisingly, the production of H₂S and CSE protein expression are obviously decreased in rats with vascular calcification (Wu et al., 2006). The CSE expression and H₂S production are impaired during the development of balloon injury-induced neointimal hyperplasia in rats, and this effect is obviously reversed by H₂S treatment (Meng et al., 2007). In mice with high fat diet for 16 weeks, it is found that CSE protein level is downregulated in the liver, the lung, and the aortic endothelium, 3-MST was also reduced in the liver (Peh et al., 2014). By contrast, CBS expression was higher in the liver and the kidney (Peh et al., 2014). These results suggest that an abnormal H₂S pathway may be an important factor for the pathophysiology of metabolic disorders and atherosclerosis.

The anti-atherosclerotic mechanisms of H₂S have been gradually described, including anti-inflammatory response, anti-oxidative action, endothelial function preservation, inhibition of foam cell formation and regulation of ion channels (Altaany et al., 2014; Mani et al., 2014; Xu et al., 2014; Wang et al., 2017; Barton and Meyer, 2019). Reduced CSE expressions at both mRNA and protein levels are detected in ox-LDL-treated endothelial cells and in aortas from apolipoprotein E knockout (ApoE^{-/-}) mice (Leucker et al., 2017). In this study, the authors demonstrated that increased histone deacetylase 6 (HDAC6) downregulated CSE and H₂S production *via* posttranslational modifications, thus leading to endothelial cell dysfunction and the development of atherosclerosis (Leucker et al., 2017). In cultured vascular endothelial cells, the expressions of miR-455-3p, endothelial nitric oxide synthase (eNOS) protein and NO production are augmented by H₂S. Besides, H₂S levels and miR-455-3p expressions are also increased in human atherosclerosis plaque, suggesting that the miR-455-3p/eNOS/NO axis is required to H₂S to circumvent the development of atherosclerosis (Li et al., 2017). Genetic deletion of CSE exaggerates atherosclerosis in ApoE^{-/-} mice, and treatment of CSE-knockout mice with H₂S inhibits the development of atherosclerosis (Mani et al., 2013), pinpointing that endogenous H₂S may be of benefit in the treatment of atherosclerosis. In addition, the augmented expressions of selectins (P-selectin and E-selectin) and cell adhesion molecules (ICAM-1 and VCAM-1) are observed in vascular endothelial cells from CSE knockout mice (Mani et al., 2013). GYY4137, a novel slow-releasing H₂S compound, retards atherosclerotic plaque formation and partially restores endothelium-dependent relaxation in ApoE^{-/-} mice through decreasing vascular inflammation and oxidative stress (Liu et al., 2013). Preconditioning with NaHS also grants a protection in atherosclerosis, as manifested by decreased atherosclerotic plaque size and aortic ICAM-1 levels (Wang

et al., 2009). Supplementation with H₂S ameliorates, while inhibition of H₂S formation intensified aortic CX3CR1 and CX3CL1 expressions and the formation of atherosclerosis (Zhang et al., 2012b). Recently, H₂S induces S-sulfhydration of kelch-like ECH-associated protein 1 (Keap1) and nuclear factor erythroid 2-related factor 2 (Nrf2) dissociation from Keap1, followed by Nrf2 nuclear translocation and anti-oxidize effects in endothelial cells, contributing to the ameliorating effect of H₂S on atherosclerosis in the context of diabetes (Xie et al., 2016). Furthermore, in a mouse model of disturbed flow-induced atherosclerosis, application of H₂S donor NaHS considerably attenuates the severity of atherosclerosis via upregulating expressions of angiotensin converting enzyme 2 (ACE2), thus converting pro-atherosclerotic Ang II to anti-atherosclerotic angiotensin 1-7 (Ang-(1-7)) (Lin et al., 2017). At the cellular level, NaHS promotes the expression of ACE2 to exert anti-inflammatory properties in lipopolysaccharide (LPS)-stimulated endothelial cells, as pretreatment with a selective ACE2 inhibitor DX600 abrogates the anti-inflammatory effect of NaHS (Lin et al., 2017). The results showed that endogenous H₂S system was involved in the development of atherosclerosis. Exogenous H₂S could confer beneficial effects on the pathogenesis of atherosclerosis.

Also, H₂S is involved in shear stress and blood viscosity. The occurrence of atherosclerosis may be initiated due to changed patterns of blood flow and ensuing shear stress (Dunn et al., 2015). It is well established that atherosclerotic plaque formation in the endothelium is site specific, and disturbed blood flow formed at the lesser curvature of the aortic arch and branch points promotes plaque formation, whereas steady laminar flow at the greater curvature is indicated to be atheroprotective (Heo et al., 2016). The branches and curvatures of the blood vessels are predisposed to endothelial dysfunction and atherosclerosis progression (Zhou et al., 2014). Under oscillatory shear stress, H₂S treatment inhibits monocyte adhesion to endothelial cells *via* activating the NO-producing Akt/eNOS signaling pathway (Go et al., 2012). Conversely, H₂S impairs shear stress-induced dilation of isolated mouse coronary arteries by inhibition of NO generation (Chai et al., 2015). It is likely that both H₂S and NO are implicated in the shear stress-induced atherosclerosis. However, further investigation is required to help us obtain more novel insights into the underlying mechanisms.

H₂S-RELATED ENDOTHELIAL DYSFUNCTION IN DIABETIC VASCULAR COMPLICATIONS

Circulating levels of H₂S are markedly reduced in diabetic animal models, such as diabetic rats (Jain et al., 2010; Suzuki et al., 2011), diabetic mice (Brancaleone et al., 2008), and also in diabetic patients (Jain et al., 2013; Suzuki et al., 2017). However, the mRNA level of CSE in the aortas of diabetic rats is not altered (Denizalti et al., 2011). Likewise, the expressions of CSE, CBS, and 3-MST are unaltered in either high glucose-treated endothelial cells or in the aortas of diabetic rats (Jain et al.,

2010; Coletta et al., 2015). On the contrary, it has been demonstrated that both high glucose and palmitate inhibit CSE expression and H₂S production in rat aortic endothelial cells, while exogenous H₂S could protect endothelial cells against apoptosis under high glucose and palmitate stimulation *via* suppressing oxidative stress, decreasing mitochondrial fragments and promoting mitophagy (Liu et al., 2017). The CSE expression and H₂S content are significantly reduced in granulation tissues of wounds in obese diabetic mice when compared with control mice (Zhao et al., 2017). The expression of CSE and H₂S level are reduced after renal ischemia/reperfusion injury in diabetes mellitus (Chen et al., 2018). In comparison with control mice, the H₂S content and CSE expression in heart tissues of diabetic rats are also markedly lower (Guo et al., 2017). In progressive diabetic nephropathy, CSE expression is markedly reduced, whereas CBS expression is unaffected (Yamamoto et al., 2013). By contrast, the protein and mRNA expression of CBS are specifically decreased in the kidney, while CSE expression remains unchanged in obese diabetic mice (Liu et al., 2018). Interestingly, CSE expression is upregulated in cerebral microvessels of type I diabetic rats (Streeter et al., 2013). Although the data are conflicting, they raise the possibility that H₂S may be a double-edged sword under diabetic pathophysiology. Certainly, more research is needed to determine the molecular mechanisms underlying the changed or unchanged expressions of H₂S-generating enzymes/H₂S under diabetic conditions.

Despite of the aforementioned results, recent study has demonstrated that 3-MST activity is inhibited in endothelial cells during hyperglycemia, leading to reduced H₂S level, impaired angiogenesis, and suppressed mitochondrial function (Coletta et al., 2015). It is highly probable that inactivation of 3-MST and elevated H₂S depletion are putative mechanisms for the decreased circulating H₂S levels in hyperglycemic endothelial cells. The high glucose-incubated vascular rings exhibit impaired endothelium-dependent relaxation, and this effect is rescued by CSE overexpression or H₂S supplementation (Suzuki et al., 2011). In the same study, they have also shown that the vascular rings from mice with gene knockout of CSE display an aggravated impairment of endothelium-dependent relaxation in response to hyperglycemia (Suzuki et al., 2011). It is anticipated that genetic modulations of CSE, CBS or 3-MST levels are effective approaches to experimentally investigate the roles of H₂S in diabetic vascular complications.

Exposure to high glucose results in elevated ROS production and apoptosis, as well as decreased superoxide dismutase activity in endothelial cells, and all the above responses could be eliminated by pretreatment with H₂S (Guan et al., 2012). Exogenous H₂S alleviated the ROS overproduction and apoptosis in hyperglycemic endothelial cells through inhibiting necroptosis (Lin et al., 2018). In the aortas of diabetic rats, the connexin (Cx) 43 and 40 expressions are downregulated, while protein kinase C (PKC) and nicotinamide adenine dinucleotide phosphate-oxidase (NADPH) oxidase subunits are upregulated, H₂S appears to be effective in attenuating these abnormalities (Zheng et al., 2010). The novel mitochondria-targeted H₂S

donors AP123 and AP39 are proven to prevent hyperglycemia-triggered oxidative stress and metabolic abnormalities in microvascular endothelial cells, suggesting that these compounds could be useful for the treatment of diabetic vascular complications (Gero et al., 2016). Induction of H₂S by Ginkgolide B alleviates endothelial dysfunction *via* inhibiting oxidative stress and increasing NO bioavailability in diabetic rats (Wang et al., 2015a). It is likely that the cardiovascular protective effects of H₂S in diabetes may be mediated by inhibition of oxidative stress.

Inhibition of the leptin/leptin receptor signal pathway contributes to the protective effects of H₂S on high-glucose-induced injuries in endothelial cells (Wu et al., 2016). Pretreatment with H₂S prevents high glucose-induced ICAM-1 levels as well as NF- κ B activation in endothelial cells (Guan et al., 2013). Besides, stimulation of endothelial cells with high glucose significantly promotes the secretion of endothelin-1 with the concomitant suppression of H₂S production, and administration of H₂S attenuates the release of endothelin-1 induced by high glucose (Guan et al., 2015). The increasing recognitions of protective effects of H₂S in high glucose-induced endothelial inflammation provide a new avenue of antagonism towards diabetic vascular complications.

In addition, high glucose/palmitate-induced excessive autophagy in endothelial cells is rectified by H₂S, this may be mediated by the Nrf2-ROS-adenosine 5'-monophosphate (AMP)-activated protein kinase (AMPK) signaling pathway (Liu et al., 2016). However, another group demonstrates that exogenous H₂S inhibits mitochondrial apoptosis and promotes mitochondrial autophagy, thus protecting endothelial cells against apoptosis induced by high glucose and palmitate (Liu et al., 2017). These contradictory results suggest that additional research is necessary to ascertain the role of autophagy in H₂S-mediated protective actions on diabetic endothelial dysfunction.

H₂S-RELATED ENDOTHELIAL DYSFUNCTION IN HYPERTENSION

The abnormal levels of H₂S have been found to be correlated with hypertension (Szabo, 2007; Whiteman and Winyard, 2011). Specifically, in a clinical study, patients with severe hypertension exhibited lower plasma H₂S level (Meng et al., 2015). In subjects with pulmonary hypertension, both CSE expression and H₂S level are significantly lower than those in healthy population (Sun et al., 2014). *In situ* hybridization analysis has shown that the expression of CSE mRNA is downregulated in the pulmonary arteries of rats with pulmonary hypertension (Xiaohui et al., 2005). Likewise, the reduced protein contents of CSE and CBS are detected in pulmonary artery endothelial cells from tobacco smoke-induced emphysema and pulmonary hypertension (Han et al., 2011). It has been revealed a marked reduction in CBS and CSE expression as well as H₂S production in mesenteric artery and carotid artery from dexamethasone-induced hypertensive rats (D'Emmanuele Di Villa Bianca et al., 2015). By contrast, the suppressed CBS expression and reduced H₂S concentration in the kidney are

observed in high salt-induced hypertension in Dahl rats (Huang et al., 2015). A decreased CSE/H₂S activity is a potential contributor to the pathogenesis of maternal hypertension in preeclampsia (Wang et al., 2013a). Also, the plasma H₂S level and CSE protein expression in thoracic aorta are all suppressed in spontaneously hypertensive rats (SHR) in comparison with normotensive rats (Yan et al., 2004; Ahmad et al., 2014). An intriguing study has illustrated that the blood pressure is enhanced by treatment with the combination of CSE inhibitor DL-propargylglycine (PAG) or the CBS inhibitor aminooxyacetic acid (AOA) in rats, while either compound alone has no any effect on the arterial pressure, suggesting that H₂S plays a critical role in regulating blood pressure (Roy et al., 2012). Despite that the expression of 3-MST is still uncertain under hypensive condition, 3-MST gene therapy improves renovascular dysfunction in response to hyperhomocysteinemia (Sen et al., 2012). Thus, a better understanding of the biochemical functions of the H₂S-producing enzyme 3-MST as well as its roles in hypertension may lead to new therapeutic targets based on modulation of H₂S production. Overall, these studies suggest that endogenous H₂S dysregulation plays an important role in regulating hypertension-associated pathological processes.

As mentioned above, a close relationship between H₂S-related endothelial dysfunction and hypertension is confirmed by an observation that genetic deletion of CSE causes the development of hypertension in mice (Yang et al., 2008). In these CSE knockout mice, the endothelium-dependent relaxation of resistance mesenteric arteries is particularly impaired (Yang et al., 2008). In a mouse model of Ang II-induced hypertension, both aortic endothelial function and NO bioavailability are significantly attenuated, and these are reversed by treatment with H₂S (Al-Magableh et al., 2015). Conversely, blockade of endogenous H₂S exacerbates these abnormalities (Al-Magableh et al., 2015). In other studies, application of H₂S donors decrease blood pressure, reverse vascular remodeling *via* suppressing VSMC proliferation, and collagen deposition in the blood vessels (Zhao et al., 2001; Li et al., 2008; Wang, 2012; Meng et al., 2015; Tomasova et al., 2015). H₂S therapy markedly restores eNOS function and NO bioavailability in N^ω-nitro-L-arginine methyl ester (L-NAME)-induced hypertensive rats (Ji et al., 2014). In agreement with this, the improvement of endothelial function by H₂S is largely attributed to inhibition of oxidative stress, suppression of renin angiotensin system (RAS), downregulation of BMP4/COX-2 pathway, or activation of the PPAR δ /PI3K/Akt/AMPK/eNOS pathway, thus contributing to the antihypertensive mechanism of H₂S in renovascular hypertensive rats (Xue et al., 2015; Xiao and Dong, 2016; Xiao et al., 2018). In SHR, exogenous H₂S administration significantly reduces blood pressure and abrogated damaged endothelial dysfunction *via* inactivation of NLRP3 inflammasome and oxidative stress (Li et al., 2019). H₂S treatment blunts increases in systolic blood pressure and ameliorates endothelial dysfunction by inhibiting oxidative stress in lead-induced hypertensive rats (Possomato-Vieira and Goncalves-Rizzi, 2018). These results demonstrate that the H₂S

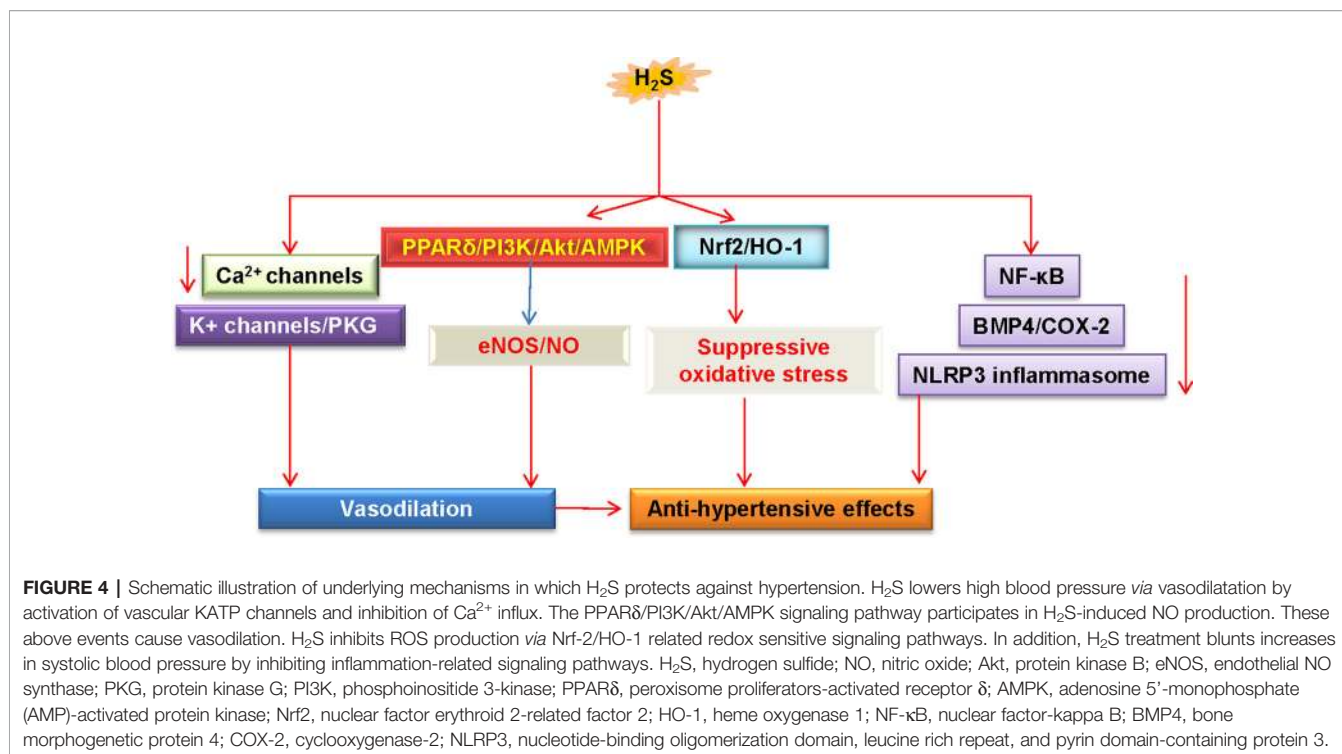
pathway may provide potential therapeutic target for treating different hypertension models.

Importantly, supplementation with S-zofenopril ameliorates vascular endothelial dysfunction by potentiating the H₂S pathway in spontaneously hypertensive models (Bucci et al., 2014). Moreover, exercise training counteracts hypertension, ameliorates vascular remodeling, and endothelial dysfunction *via* restoring bioavailability of H₂S and NO in hypertensive rats (Gu et al., 2013). HDAC6 inhibitor tubastatin A alleviates Ang II-induced high blood pressure and vasoconstriction by preventing the protein degradation of CSE (Chi et al., 2019). Overall, these studies suggest that upregulation of H₂S may be considered as a promising strategy for preventing the progression of hypertension and its associated endothelial dysfunction (Figure 4). However, further in-depth research is still required to understand the precise underlying mechanisms, and this will be helpful to develop better therapeutic employment of H₂S in the treatment of hypertension.

EVIDENCE FOR H₂S/NO CROSSTALK IN ENDOTHELIAL INFLAMMATION

Due to the importance of H₂S and NO in cardiovascular disease, the interactive regulatory functions of H₂S and NO in endothelial dysfunction-associated cardiovascular disease may be a very attractive subject. In other words, the biological interactions of H₂S with NO could influence each other's fate in the endothelium as described previously (Wang et al., 2015b; Nagpure and Bian, 2016; Wu and Hu, 2018) (Figure 2).

Studies on H₂S/NO interaction in inflammation response, especially in endothelial cell inflammation, have been less extensive. Administration of LPS increases H₂S synthesis, upregulates CSE and iNOS expressions, and promotes myeloperoxidase activity in the liver, whereas the effects are inhibited by the NO donor, nitroflurbiprofen (Anuar et al., 2006). These results suggest that downregulation of H₂S biosynthesis is responsible for the augmented anti-inflammatory activity of nitroflurbiprofen in the liver (Anuar et al., 2006). In turn, pretreatment with H₂S is able to inhibit LPS-induced iNOS expression and NO production *via* heme oxygenase 1 (HO-1) expression in macrophages (Oh et al., 2006). In accordance with this finding, H₂S donor inhibits the release of the pro-inflammatory mediators and NO production, potentially *via* NF- κ B inactivation in LPS-treated macrophages (Whiteman et al., 2010). The pulmonary CSE expression and H₂S levels are downregulated in a model of inflammatory lung disease (Chen et al., 2009; Whiteman and Winyard, 2011). NaHS, a donor for H₂S, significantly attenuates pulmonary iNOS activation in ovalbumin-treated rats (Chen et al., 2009). Moreover, H₂S is found to act as an anti-inflammatory agent contributing to gastrointestinal mucosal defense through NO-dependent pathway (Jensen et al., 2017). In cardiovascular system, vasodilation is impaired and endothelial H₂S content is decreased in vessels from obese mice, this may be attributed to



the increased iNOS activity in proinflammatory macrophages (Candela et al., 2017). This finding suggests that macrophage-derived iNOS promotes microvascular endothelial dysfunction through reducing the bioavailability of H₂S in the blood vessel (Candela et al., 2017). GYY4137, a novel slow-releasing H₂S compound, is reported to attenuate vascular inflammation and improve endothelial function *via* activating aortic eNOS phosphorylation in ApoE^{-/-} mice (Liu et al., 2013). A number of studies have showed that H₂S strengthens endothelial NO production *via* activating eNOS phosphorylation (Predmore et al., 2011; Xiao et al., 2018), which results in ameliorating the development of hypertension (Xiao et al., 2018). Taken together, the above studies imply that a complex interaction between H₂S and NO might serve as an important regulator for endothelial inflammation and associated endothelial dysfunction. However, the potential mechanisms of the interactions between H₂S and NO in endothelial inflammation remain unclear. As research in this area progresses and more data are available, it will help us to better understand the underlying mechanisms.

ROLE OF H₂S IN INTESTINAL MICROBIOTA AND CIRCADIAN RHYTHMS

It should be emphasized that intestinal microbiota is also an emerging factor for human health and disease, including cardiovascular diseases (Tang et al., 2017). In recent years, both human and animal experiments have established that alterations in the composition, function, and metabolites of intestinal flora might induce gut microflora dysbiosis, contributing to the pathogenesis

of cardiovascular disorders (Tang et al., 2019). Circadian rhythmicity is a characteristic of mammalian metabolism that orchestrates metabolic processes in living organisms based on day/night light cycles (Liu and Chang, 2017). Disturbance of circadian rhythmicity is associated with increased risk for metabolic obesity, diabetes, and cardiovascular dysfunction (Crnko et al., 2018). Similarly, the intestinal microbiota exhibits their own circadian rhythmicity in terms of composition and functions (Tahara et al., 2017). Circadian disorganization may affect the intestinal microbiota which may result in metabolic syndrome and cardiovascular diseases (Voigt et al., 2016). Accumulating evidence has indicated that circadian rhythm disruption in intestinal microbiota is involved in various human diseases, including cardiovascular diseases (Jin et al., 2019). Thereafter, interfering with the composition, function, and metabolites of the intestinal flora or recovery of the normal circadian rhythm in the intestinal flora may provide valuable insights into potential therapeutic strategies for cardiovascular diseases.

Notably, the cysteine degradation by the microbiota is taken as a dominant pathway for H₂S generation (Basic et al., 2017). Intestinal microbiota is a potential target of H₂S, and H₂S acts on gastrointestinal epithelium to modify the gut microbiota (Wallace et al., 2018). It has been reviewed that H₂S is a double-edge sword for the intestinal epithelium with beneficial effect at low concentration (nanomolar to low micromolar), but deleterious effects at higher concentrations (high micromolar to millimolar) (Blachier et al., 2019). Considering the critical importance of intestinal microbiota and H₂S in maintaining cardiovascular homeostasis, it is believed that intestinal microbiota-derived H₂S integrates microbial and circadian cues

for regulation of diurnal metabolic rhythms, thereby influencing the endothelial dysfunction in cardiovascular system. However, it is still largely unknown with respect to the roles of H₂S in intestinal microbiota-mediated endothelial dysfunction. The relationship between H₂S and intestinal microbiota in cardiovascular regulation may be a very interesting topic. As the gut microbiota leads to much more H₂S production from cysteine than endogenous metabolism, it is likely that H₂S from the bacterial or intestinal epithelium may be a critical determinant for cardiovascular health or disease. However, additional investigation is warranted to identify the exact roles of H₂S in intestinal microbiota and circadian rhythms. Our current understanding of the relationship between H₂S and intestinal microbiota in endothelial inflammation-related cardiovascular disorders is expanding continuously. The interaction between their signaling pathways is increasingly recognized as the future direction for the research in the gasotransmitters field.

CONCLUDING REMARKS AND FUTURE PERSPECTIVES

In addition to NO deficiency and intestinal flora, other factors including oxidative stress (Nakahira et al., 2011), endoplasmic reticulum stress (Battson et al., 2017; Luchetti et al., 2017), mitochondrial dysfunction (Gao et al., 2018), hypoxia (Feng et al., 2017a), homocysteine (Esse and Barroso, 2019), and immune activation (Pan et al., 2017) are also closely related with endothelial inflammation and dysfunction in cardiovascular diseases. With in-depth research, our knowledge on the underlying mechanisms of H₂S-mediated suppression of endothelial cell inflammation is expanding and it is now apparent that interactions between H₂S and endothelial inflammation-regulated pathways may be proposed as a promising approach for cardiovascular disease therapy. A better understanding of such interactions will be favorable to develop novel therapeutic strategies for endothelial dysfunction-related cardiovascular diseases.

Due to a myriad of biological functions of H₂S, there has been a growing interest regarding the enormously therapeutic potential of H₂S in various diseases including cardiovascular diseases. However, our current knowledge on cardiovascular protective effects of H₂S is mainly from animal or cell experiments using H₂S donors or inhibitors of H₂S-producing enzymes. Whether the promising effects of these chemicals in animal studies can be

transferable to clinical studies warrants further studies. As such, it should be mentioned that clinical trial results will also pave the way to a better understanding of the effectiveness of H₂S in human diseases. In one completed clinical trial in healthy volunteers and subjects with impaired renal function received known concentrations of sodium sulfide (clinicaltrials.gov, NCT00879645). Despite that only some results have been announced so far, but the treatment could be considered safe because no serious adverse effects are occurred in the involved patients. However, some caution can be warranted as another clinical trial regarding the potential of H₂S in coronary artery bypass graft patients was terminated without undisclosed reasons (NCT00858936) and one had been withdrawn before enrollment (NCT01007461). Some other completed trials are completed to test the role of H₂S in inflammatory diseases such as ulcerative colitis (NCT01282905) or septic shock and stroke (NCT01088490). However, to date, no results are posted due to unknown reasons. Therefore, further results and information from those ongoing and future trials will help to elucidate the physiological and pathophysiological importance of H₂S in various diseases.

Until now, endothelial inflammation and dysfunction remain mortal factors for cardiovascular diseases. It is anticipated that a full understanding of the modulatory mechanisms of the link between endothelial inflammation and destructive H₂S bioavailability might promote the translation of H₂S biology to clinical management of endothelial dysfunction-related cardiovascular diseases. To achieve this, more original work remains to be experimentally evaluated in the future.

AUTHOR CONTRIBUTIONS

H-JS and J-SB designed the contents of this review article. H-JS, Z-YW, and X-WN conducted initial search of literature and prepared the figures. H-JS and J-SB drafted the manuscript. J-SB critically helped to revise the manuscript. All authors have read and approved the final manuscript.

FUNDING

This work was supported by the Ministry of Education of Singapore Tier 2 Research grant (MOE2017-T2-2-029), and the China Jiangsu Nature Science Foundation (BK20181185).

REFERENCES

- Ahmad, F. U., Sattar, M. A., Rathore, H. A., Tan, Y. C., Akhtar, S., Jin, O. H., et al. (2014). Hydrogen sulphide and tempol treatments improve the blood pressure and renal excretory responses in spontaneously hypertensive rats. *Ren. Fail* 36, 598–605. doi: 10.3109/0886022X.2014.882218
- Ali, M. Y., Ping, C. Y., Mok, Y. Y., Ling, L., Whiteman, M., Bhatia, M., et al. (2006). Regulation of vascular nitric oxide *in vitro* and *in vivo*; a new role for endogenous hydrogen sulphide? *Br. J. Pharmacol.* 149, 625–634. doi: 10.1038/sj.bjp.0706906
- Allende-Vigo, M. Z. (2010). Pathophysiologic mechanisms linking adipose tissue and cardiometabolic risk. *Endocr. Pract.* 16, 692–698. doi: 10.4158/EP09340.RA
- Al-Magableh, M. R., Kemp-Harper, B. K., and Hart, J. L. (2015). Hydrogen sulfide treatment reduces blood pressure and oxidative stress in angiotensin II-induced hypertensive mice. *Hypertens. Res.* 38, 13–20. doi: 10.1038/hr.2014.125

- Altaany, Z., Moccia, F., Munaron, L., Mancardi, D., and Wang, R. (2014). Hydrogen sulfide and endothelial dysfunction: relationship with nitric oxide. *Curr. Med. Chem.* 21, 3646–3661. doi: 10.1097/SHK.0000000000000599
- Anuar, F., Whiteman, M., Siau, J. L., Kwong, S. E., Bhatia, M., and Moore, P. K. (2006). Nitric oxide-releasing flurbiprofen reduces formation of proinflammatory hydrogen sulfide in lipopolysaccharide-treated rat. *Br. J. Pharmacol.* 147, 966–974. doi: 10.1038/sj.bjp.0706696
- Baghai, T. C., Varallo-Bedarida, G., Born, C., Hafner, S., Schule, C., Eser, D., et al. (2018). Classical risk factors and inflammatory biomarkers: one of the missing biological links between cardiovascular disease and major depressive disorder. *Int. J. Mol. Sci.* 19, E1740. doi: 10.3390/ijms19061740
- Barton, M., and Meyer, M. R. (2019). HuR-ry Up: how hydrogen sulfide protects against atherosclerosis. *Circulation* 139, 115–118. doi: 10.1161/CIRCULATIONAHA.118.036854
- Basic, A., Blomqvist, M., Dahlen, G., and Svensater, G. (2017). The proteins of *Fusobacterium* spp. involved in hydrogen sulfide production from L-cysteine. *BMC Microbiol.* 17, 61. doi: 10.1186/s12866-017-0967-9
- Baszczuk, A., Kopczyński, Z., and Thielemann, A. (2014). [Endothelial dysfunction in patients with primary hypertension and hyperhomocysteinemia]. *Postepy Hig. Med. Dosw (Online)* 68, 91–100. doi: 10.5604/17322693.1087521
- Battson, M. L., Lee, D. M., and Gentile, C. L. (2017). Endoplasmic reticulum stress and the development of endothelial dysfunction. *Am. J. Physiol. Heart Circ. Physiol.* 312, H355–H367. doi: 10.1152/ajpheart.00437.2016
- Bibli, S. I., Hu, J., Sigala, F., Wittig, I., Heidler, J., Zukunft, S., et al. (2019). Cystathionine gamma lyase sulfhydrates the RNA binding protein human antigen r to preserve endothelial cell function and delay atherogenesis. *Circulation* 139, 101–114. doi: 10.1161/CIRCULATIONAHA.118.034757
- Blachier, F., Beaumont, M., and Kim, E. (2019). Cysteine-derived hydrogen sulfide and gut health: a matter of endogenous or bacterial origin. *Curr. Opin. Clin. Nutr. Metab. Care* 22, 68–75. doi: 10.1097/MCO.0000000000000526
- Boulanger, C. M. (2016). Endothelium. *Arterioscler. Thromb. Vasc. Biol.* 36, e26–e31. doi: 10.1161/ATVBAHA.116.306940
- Bourque, C., Zhang, Y., Fu, M., Racine, M., Greasley, A., Pei, Y., et al. (2018). H2S protects lipopolysaccharide-induced inflammation by blocking NFkappaB transactivation in endothelial cells. *Toxicol. Appl. Pharmacol.* 338, 20–29. doi: 10.1016/j.taap.2017.11.004
- Brancaleone, V., Rovietto, F., Vellecco, V., De Gruttola, L., Bucci, M., and Cirino, G. (2008). Biosynthesis of H2S is impaired in non-obese diabetic (NOD) mice. *Br. J. Pharmacol.* 155, 673–680. doi: 10.1038/bjp.2008.296
- Bucci, M., Vellecco, V., Cantalupo, A., Brancaleone, V., Zhou, Z., Evangelista, S., et al. (2014). Hydrogen sulfide accounts for the peripheral vascular effects of zofenopril independently of ACE inhibition. *Cardiovasc. Res.* 102, 138–147. doi: 10.1093/cvr/cvu026
- Cai, W. J., Wang, M. J., Moore, P. K., Jin, H. M., Yao, T., and Zhu, Y. C. (2007). The novel proangiogenic effect of hydrogen sulfide is dependent on Akt phosphorylation. *Cardiovasc. Res.* 76, 29–40. doi: 10.1016/j.cardiores.2007.05.026
- Candela, J., Wang, R., and White, C. (2017). Microvascular endothelial dysfunction in obesity is driven by macrophage-dependent hydrogen sulfide depletion. *Arterioscler. Thromb. Vasc. Biol.* 37, 889–899. doi: 10.1161/ATVBAHA.117.309138
- Caporali, A., and Emanueli, C. (2011). MicroRNA regulation in angiogenesis. *Vascul. Pharmacol.* 55, 79–86. doi: 10.1016/j.vph.2011.06.006
- Carter, A. M. (2012). Complement activation: an emerging player in the pathogenesis of cardiovascular disease. *Scientifica (Cairo)* 2012, 402783. doi: 10.6064/2012/402783
- Castro-Ferreira, R., Cardoso, R., Leite-Moreira, A., and Mansilha, A. (2018). The role of endothelial dysfunction and inflammation in chronic venous disease. *Ann. Vasc. Surg.* 46, 380–393. doi: 10.1016/j.avsg.2017.06.131
- Chai, Q., Lu, T., Wang, X. L., and Lee, H. C. (2015). Hydrogen sulfide impairs shear stress-induced vasodilation in mouse coronary arteries. *Pflugers Arch.* 467, 329–340. doi: 10.1007/s00424-014-1526-y
- Cheang, W. S., Wong, W. T., Shen, B., Lau, C. W., Tian, X. Y., Tsang, S. Y., et al. (2010). 4-aminopyridine-sensitive K⁺ channels contributes to NaHS-induced membrane hyperpolarization and relaxation in the rat coronary artery. *Vascul. Pharmacol.* 53, 94–98. doi: 10.1016/j.vph.2010.04.004
- Chen, Y. H., Wu, R., Geng, B., Qi, Y. F., Wang, P. P., Yao, W. Z., et al. (2009). Endogenous hydrogen sulfide reduces airway inflammation and remodeling in a rat model of asthma. *Cytokine* 45, 117–123. doi: 10.1016/j.cyto.2008.11.009
- Chen, Y., Zhao, L., Jiang, S., Hu, Z., Hu, B., Tong, F., et al. (2018). Cystathionine gamma-lyase is involved in the renoprotective effect of brief and repeated ischemic postconditioning after renal ischemia/reperfusion injury in Diabetes Mellitus. *Transplant. Proc.* 50, 1549–1557. doi: 10.1016/j.transproceed.2018.03.011
- Cheng, Y., Ndisang, J. F., Tang, G., Cao, K., and Wang, R. (2004). Hydrogen sulfide-induced relaxation of resistance mesenteric artery beds of rats. *Am. J. Physiol. Heart Circ. Physiol.* 287, H2316–H2323. doi: 10.1152/ajpheart.00331.2004
- Chi, Z., Byeon, H. E., Seo, E., Nguyen, Q. T., Lee, W., Jeong, Y., et al. (2019). Histone deacetylase 6 inhibitor tubastatin A attenuates angiotensin II-induced hypertension by preventing cystathionine gamma-lyase protein degradation. *Pharmacol. Res.* 146, 104281. doi: 10.1016/j.phrs.2019.104281
- Chistiakov, D. A., Melnichenko, A. A., Myasoedova, V. A., Grechko, A. V., and Orekhov, A. N. (2017). Mechanisms of foam cell formation in atherosclerosis. *J. Mol. Med. (Berl)* 95, 1153–1165. doi: 10.1007/s00109-017-1575-8
- Coletta, C., Modis, K., Szczesny, B., Brunyanski, A., Olah, G., Rios, E. C., et al. (2015). Regulation of vascular tone, angiogenesis and cellular bioenergetics by the 3-mercaptopyruvate sulfurtransferase/H2S Pathway: functional impairment by hyperglycemia and restoration by DL-alpha-lipoic acid. *Mol. Med.* 21, 1–14. doi: 10.2119/molmed.2015.00035
- Crnko, S., Cour, M., Van Laake, L. W., and Lecour, S. (2018). Vasculature on the clock: circadian rhythm and vascular dysfunction. *Vascul. Pharmacol.* 108, 1–7. doi: 10.1016/j.vph.2018.05.003
- D'emmanuele Di Villa Bianca, R., Sorrentino, R., Coletta, C., Mitidieri, E., Rossi, A., Vellecco, V., et al. (2011). Hydrogen sulfide-induced dual vascular effect involves arachidonic acid cascade in rat mesenteric arterial bed. *J. Pharmacol. Exp. Ther.* 337, 59–64. doi: 10.1124/jpet.110.176016
- D'emmanuele Di Villa Bianca, R., Mitidieri, E., Donnarumma, E., Tramontano, T., Brancaleone, V., Cirino, G., et al. (2015). Hydrogen sulfide is involved in dexamethasone-induced hypertension in rat. *Nitric. Oxide* 46, 80–86. doi: 10.1016/j.niox.2014.11.013
- Denizalti, M., Bozkurt, T. E., Akpulat, U., Sahin-Erdemli, I., and Abacioglu, N. (2011). The vasorelaxant effect of hydrogen sulfide is enhanced in streptozotocin-induced diabetic rats. *Naunyn Schmiedeberg's Arch. Pharmacol.* 383, 509–517. doi: 10.1007/s00210-011-0601-6
- Du, C., Lin, X., Xu, W., Zheng, F., Cai, J., Yang, J., et al. (2019). Sulfhydrated Sirtuin-1 increasing its deacetylation activity is an essential epigenetics mechanism of anti-atherogenesis by hydrogen sulfide. *Antioxid. Redox Signal.* 30, 184–197. doi: 10.1089/ars.2017.7195
- Dulmovits, B. M., and Herman, I. M. (2012). Microvascular remodeling and wound healing: a role for pericytes. *Int. J. Biochem. Cell Biol.* 44, 1800–1812. doi: 10.1016/j.biocel.2012.06.031
- Dunn, J., Simmons, R., Thabet, S., and Jo, H. (2015). The role of epigenetics in the endothelial cell shear stress response and atherosclerosis. *Int. J. Biochem. Cell Biol.* 67, 167–176. doi: 10.1016/j.biocel.2015.05.001
- Durham, A. L., Speer, M. Y., Scatena, M., Giachelli, C. M., and Shanahan, C. M. (2018). Role of smooth muscle cells in vascular calcification: implications in atherosclerosis and arterial stiffness. *Cardiovasc. Res.* 114, 590–600. doi: 10.1093/cvr/cvy010
- Esse, R., and Barroso, M. (2019). The contribution of homocysteine metabolism disruption to endothelial dysfunction: state-of-the-Art. *Int. J. Mol. Sci.* 20, E867. doi: 10.3390/ijms20040867
- Fan, J., Zheng, F., Li, S., Cui, C., Jiang, S., Zhang, J., et al. (2019). Hydrogen sulfide lowers hyperhomocysteinemia dependent on cystathionine gamma lyase S-sulfhydration in ApoE-knockout atherosclerotic mice. *Br. J. Pharmacol.* 176, 3180–3192. doi: 10.1111/bph.14719
- Fang, F., Chen, D., Yu, L., Dai, X., Yang, Y., Tian, W., et al. (2013). Proinflammatory stimuli engage Brahma related gene 1 and Brahma in endothelial injury. *Circ. Res.* 113, 986–996. doi: 10.1161/CIRCRESAHA.113.301296
- Feng, S., Bowden, N., Fragiadaki, M., Souilhol, C., Hsiao, S., Mahmoud, M., et al. (2017a). Mechanical activation of hypoxia-inducible factor 1alpha drives

- endothelial dysfunction at atheroprone sites. *Arterioscler. Thromb. Vasc. Biol.* 37, 2087–2101. doi: 10.1161/ATVBAHA.117.309249
- Feng, S., Chen, S., Yu, W., Zhang, D., Zhang, C., Tang, C., et al. (2017b). H2S inhibits pulmonary arterial endothelial cell inflammation in rats with monocrotaline-induced pulmonary hypertension. *Lab. Invest.* 97, 268–278. doi: 10.1038/labinvest.2016.129
- Fiorucci, S., Antonelli, E., Distrutti, E., Rizzo, G., Mencarelli, A., Orlandi, S., et al. (2005). Inhibition of hydrogen sulfide generation contributes to gastric injury caused by anti-inflammatory nonsteroidal drugs. *Gastroenterology* 129, 1210–1224. doi: 10.1053/j.gastro.2005.07.060
- Gao, F., Chen, J., and Zhu, H. (2018). A potential strategy for treating atherosclerosis: improving endothelial function via AMP-activated protein kinase. *Sci. China Life Sci.* 61, 1024–1029. doi: 10.1007/s11427-017-9285-1
- Gero, D., Torregrossa, R., Perry, A., Waters, A., Le-Trionnaire, S., Whatmore, J. L., et al. (2016). The novel mitochondria-targeted hydrogen sulfide (H2S) donors AP123 and AP39 protect against hyperglycemic injury in microvascular endothelial cells *in vitro*. *Pharmacol. Res.* 113, 186–198. doi: 10.1016/j.phrs.2016.08.019
- Go, Y. M., Lee, H. R., and Park, H. (2012). H(2)S inhibits oscillatory shear stress-induced monocyte binding to endothelial cells via nitric oxide production. *Mol. Cells* 34, 449–455. doi: 10.1007/s10059-012-0200-5
- Gu, Q., Wang, B., Zhang, X. F., Ma, Y. P., Liu, J. D., and Wang, X. Z. (2013). Contribution of hydrogen sulfide and nitric oxide to exercise-induced attenuation of aortic remodeling and improvement of endothelial function in spontaneously hypertensive rats. *Mol. Cell Biochem.* 375, 199–206. doi: 10.1007/s11010-012-1542-1
- Guan, Q., Zhang, Y., Yu, C., Liu, Y., Gao, L., and Zhao, J. (2012). Hydrogen sulfide protects against high-glucose-induced apoptosis in endothelial cells. *J. Cardiovasc. Pharmacol.* 59, 188–193. doi: 10.1097/FJC.0b013e31823b4915
- Guan, Q., Wang, X., Gao, L., Chen, J., Liu, Y., Yu, C., et al. (2013). Hydrogen sulfide suppresses high glucose-induced expression of intercellular adhesion molecule-1 in endothelial cells. *J. Cardiovasc. Pharmacol.* 62, 278–284. doi: 10.1097/FJC.0b013e31829875ef
- Guan, Q., Liu, W., Liu, Y., Fan, Y., Wang, X., Yu, C., et al. (2015). High glucose induces the release of endothelin-1 through the inhibition of hydrogen sulfide production in HUVECs. *Int. J. Mol. Med.* 35, 810–814. doi: 10.3892/ijmm.2014.2059
- Guo, R., Wu, Z., Jiang, J., Liu, C., Wu, B., Li, X., et al. (2017). New mechanism of lipotoxicity in diabetic cardiomyopathy: Deficiency of Endogenous H2S Production and ER stress. *Mech. Ageing Dev.* 162, 46–52. doi: 10.1016/j.mad.2016.11.005
- Han, W., Dong, Z., Dimitropoulou, C., and Su, Y. (2011). Hydrogen sulfide ameliorates tobacco smoke-induced oxidative stress and emphysema in mice. *Antioxid. Redox Signal.* 15, 2121–2134. doi: 10.1089/ars.2010.3821
- Haybar, H., Shokuhian, M., Bagheri, M., Davari, N., and Saki, N. (2019). Involvement of circulating inflammatory factors in prognosis and risk of cardiovascular disease. *J. Mol. Cell Cardiol.* 132, 110–119. doi: 10.1016/j.yjmcc.2019.05.010
- Heo, K. S., Berk, B. C., and Abe, J. (2016). Disturbed flow-induced endothelial proatherogenic signaling via regulating post-translational modifications and epigenetic events. *Antioxid. Redox Signal.* 25, 435–450. doi: 10.1089/ars.2015.6556
- Hu, H. J., Jiang, Z. S., Zhou, S. H., and Liu, Q. M. (2016). Hydrogen sulfide suppresses angiotensin II-stimulated endothelin-1 generation and subsequent cytotoxicity-induced endoplasmic reticulum stress in endothelial cells via NF-kappaB. *Mol. Med. Rep.* 14, 4729–4740. doi: 10.3892/mmr.2016.5827
- Huang, P., Chen, S., Wang, Y., Liu, J., Yao, Q., Huang, Y., et al. (2015). Down-regulated CBS/H2S pathway is involved in high-salt-induced hypertension in Dahl rats. *Nitric. Oxide* 46, 192–203. doi: 10.1016/j.niox.2015.01.004
- Jackson-Weaver, O., Osmond, J. M., Naik, J. S., Gonzalez Bosc, L. V., Walker, B. R., and Kanagy, N. L. (2015). Intermittent hypoxia in rats reduces activation of Ca2+ sparks in mesenteric arteries. *Am. J. Physiol. Heart Circ. Physiol.* 309, H1915–H1922. doi: 10.1152/ajpheart.00179.2015
- Jain, S. K., Bull, R., Rains, J. L., Bass, P. F., Levine, S. N., Reddy, S., et al. (2010). Low levels of hydrogen sulfide in the blood of diabetes patients and streptozotocin-treated rats causes vascular inflammation? *Antioxid. Redox Signal.* 12, 1333–1337. doi: 10.1089/ars.2009.2956
- Jain, S. K., Manna, P., Micinski, D., Lieblong, B. J., Kahlon, G., Morehead, L., et al. (2013). In African American type 2 diabetic patients, is vitamin D deficiency associated with lower blood levels of hydrogen sulfide and cyclic adenosine monophosphate, and elevated oxidative stress? *Antioxid. Redox Signal.* 18, 1154–1158. doi: 10.1089/ars.2012.4843
- Jensen, A. R., Drucker, N. A., Khaneki, S., Ferkowicz, M. J., and Markel, T. A. (2017). Hydrogen sulfide improves intestinal recovery following ischemia by endothelial nitric oxide-dependent mechanisms. *Am. J. Physiol. Gastrointest. Liver Physiol.* 312, G450–g456. doi: 10.1152/ajpgi.00444.2016
- Ji, W., Liu, S., Dai, J., Yang, T., Jiang, X., Duan, X., et al. (2014). Hydrogen sulfide defends against the cardiovascular risk of Nw-nitro-L-argininemethyl ester-induced hypertension in rats via the nitric oxide/endothelial nitric oxide synthase pathway. *Chin. Med. J. (Engl)* 127, 3751–3757. doi: 10.3760/cma.j.issn.0366-6999.20141573
- Jin, M., Qian, Z., Yin, J., Xu, W., and Zhou, X. (2019). The role of intestinal microbiota in cardiovascular disease. *J. Cell Mol. Med.* 23, 2343–2350. doi: 10.1111/jcmm.14195
- Kan, J., Guo, W., Huang, C., Bao, G., Zhu, Y., and Zhu, Y. Z. (2014). S-propargyl-cysteine, a novel water-soluble modulator of endogenous hydrogen sulfide, promotes angiogenesis through activation of signal transducer and activator of transcription 3. *Antioxid. Redox Signal.* 20, 2303–2316. doi: 10.1089/ars.2013.5449
- Kanagy, N. L., Szabo, C., and Papapetropoulos, A. (2017). Vascular biology of hydrogen sulfide. *Am. J. Physiol. Cell Physiol.* 312, C537–C549. doi: 10.1152/ajpcell.00329.2016
- Kavurma, M. M., Rayner, K. J., and Karunakaran, D. (2017). The walking dead: macrophage inflammation and death in atherosclerosis. *Curr. Opin. Lipidol.* 28, 91–98. doi: 10.1097/MOL.0000000000000394
- Kiss, L., Deitch, E. A., and Szabo, C. (2008). Hydrogen sulfide decreases adenosine triphosphate levels in aortic rings and leads to vasorelaxation via metabolic inhibition. *Life Sci.* 83, 589–594. doi: 10.1016/j.lfs.2008.08.006
- Kubo, S., Kajiwara, M., and Kawabata, A. (2007). Dual modulation of the tension of isolated gastric artery and gastric mucosal circulation by hydrogen sulfide in rats. *Inflammopharmacology* 15, 288–292. doi: 10.1007/s10787-007-1590-4
- Leucker, T. M., Nomura, Y., Kim, J. H., Bhatta, A., Wang, V., Wecker, A., et al. (2017). Cystathionine gamma-lyase protects vascular endothelium: a role for inhibition of histone deacetylase 6. *Am. J. Physiol. Heart Circ. Physiol.* 312, H711–H720. doi: 10.1152/ajpheart.00724.2016
- Leung, S. W., and Vanhoutte, P. M. (2017). Endothelium-dependent hyperpolarization: age, gender and blood pressure, do they matter? *Acta Physiol. (Oxf)* 219, 108–123. doi: 10.1111/apha.12628
- Li, L., Whiteman, M., Guan, Y. Y., Neo, K. L., Cheng, Y., Lee, S. W., et al. (2008). Characterization of a novel, water-soluble hydrogen sulfide-releasing molecule (GYY4137): new insights into the biology of hydrogen sulfide. *Circulation* 117, 2351–2360. doi: 10.1161/CIRCULATIONAHA.107.753467
- Li, L., Rose, P., and Moore, P. K. (2011). Hydrogen sulfide and cell signaling. *Annu. Rev. Pharmacol. Toxicol.* 51, 169–187. doi: 10.1146/annurev-pharmtox-010510-100505
- Li, X. H., Xue, W. L., Wang, M. J., Zhou, Y., Zhang, C. C., Sun, C., et al. (2017). H2S regulates endothelial nitric oxide synthase protein stability by promoting microRNA-455-3p expression. *Sci. Rep.* 7, 44807. doi: 10.1038/srep44807
- Li, J., Teng, X., Jin, S., Dong, J., Guo, Q., Tian, D., et al. (2019). Hydrogen sulfide improves endothelial dysfunction by inhibiting the vicious cycle of NLRP3 inflammasome and oxidative stress in spontaneously hypertensive rats. *J. Hypertens.* 37, 1633–1643. doi: 10.1097/HJH.0000000000002101
- Lin, Y., Zeng, H., Gao, L., Gu, T., Wang, C., and Zhang, H. (2017). Hydrogen sulfide attenuates atherosclerosis in a partially ligated carotid artery mouse model via regulating angiotensin converting enzyme 2 expression. *Front. Physiol.* 8, 782. doi: 10.3389/fphys.2017.00782
- Lin, J., Chen, M., Liu, D., Guo, R., Lin, K., Deng, H., et al. (2018). Exogenous hydrogen sulfide protects human umbilical vein endothelial cells against high glucose-induced injury by inhibiting the necroptosis pathway. *Int. J. Mol. Med.* 41, 1477–1486. doi: 10.3892/ijmm.2017.3330
- Liu, F., and Chang, H. C. (2017). Physiological links of circadian clock and biological clock of aging. *Protein Cell* 8, 477–488. doi: 10.1007/s13238-016-0366-2
- Liu, Y. H., Yan, C. D., and Bian, J. S. (2011). Hydrogen sulfide: a novel signaling molecule in the vascular system. *J. Cardiovasc. Pharmacol.* 58, 560–569. doi: 10.1097/FJC.0b013e31820eb7a1
- Liu, Y. H., Lu, M., Hu, L. F., Wong, P. T., Webb, G. D., and Bian, J. S. (2012). Hydrogen sulfide in the mammalian cardiovascular system. *Antioxid. Redox Signal.* 17, 141–185. doi: 10.1089/ars.2011.4005

- Liu, Z., Han, Y., Li, L., Lu, H., Meng, G., Li, X., et al. (2013). The hydrogen sulfide donor, GYY4137, exhibits anti-atherosclerotic activity in high fat fed apolipoprotein E(-/-) mice. *Br. J. Pharmacol.* 169, 1795–1809. doi: 10.1111/bph.12246
- Liu, J., Wu, J., Sun, A., Sun, Y., Yu, X., Liu, N., et al. (2016). Hydrogen sulfide decreases high glucose/palmitate-induced autophagy in endothelial cells by the Nrf2-ROS-AMPK signaling pathway. *Cell Biosci.* 6, 33. doi: 10.1186/s13578-016-0099-1
- Liu, N., Wu, J., Zhang, L., Gao, Z., Sun, Y., Yu, M., et al. (2017). Hydrogen Sulphide modulating mitochondrial morphology to promote mitophagy in endothelial cells under high-glucose and high-palmitate. *J. Cell Mol. Med.* 21, 3190–3203. doi: 10.1111/jcmm.13223
- Liu, M., Deng, M., Su, J., Lin, Y., Jia, Z., Peng, K., et al. (2018). Specific downregulation of cystathionine beta-synthase expression in the kidney during obesity. *Physiol. Rep.* 6, e13630. doi: 10.14814/phy2.13630
- Luchetti, F., Crinelli, R., Cesarini, E., Canonico, B., Guidi, L., Zerbinati, C., et al. (2017). Endothelial cells, endoplasmic reticulum stress and oxysterols. *Redox Biol.* 13, 581–587. doi: 10.1016/j.redox.2017.07.014
- Mani, S., Li, H., Untereiner, A., Wu, L., Yang, G., Austin, R. C., et al. (2013). Decreased endogenous production of hydrogen sulfide accelerates atherosclerosis. *Circulation* 127, 2523–2534. doi: 10.1161/CIRCULATIONAHA.113.002208
- Mani, S., Untereiner, A., Wu, L., and Wang, R. (2014). Hydrogen sulfide and the pathogenesis of atherosclerosis. *Antioxid. Redox Signal.* 20, 805–817. doi: 10.1089/ars.2013.5324
- Mathers, C. D., and Loncar, D. (2006). Projections of global mortality and burden of disease from 2002 to 2030. *PloS Med.* 3, e442. doi: 10.1371/journal.pmed.0030442
- Meng, Q. H., Yang, G., Yang, W., Jiang, B., Wu, L., and Wang, R. (2007). Protective effect of hydrogen sulfide on balloon injury-induced neointima hyperplasia in rat carotid arteries. *Am. J. Pathol.* 170, 1406–1414. doi: 10.2353/ajpath.2007.060939
- Meng, G., Ma, Y., Xie, L., Ferro, A., and Ji, Y. (2015). Emerging role of hydrogen sulfide in hypertension and related cardiovascular diseases. *Br. J. Pharmacol.* 172, 5501–5511. doi: 10.1111/bph.12900
- Mitidieri, E., and Gurgone, D. (2019). L-cysteine/cystathionine-beta-synthase-induced relaxation in mouse aorta involves a L-serine/sphingosine-1-phosphate/NO pathway. *Br. J. Pharmacol.* doi: 10.1111/bph.14654
- Mustafa, A. K., Sikka, G., Gazi, S. K., Steppan, J., Jung, S. M., Bhunia, A. K., et al. (2011). Hydrogen sulfide as endothelium-derived hyperpolarizing factor sulphydrates potassium channels. *Circ. Res.* 109, 1259–1268. doi: 10.1161/CIRCRESAHA.111.240242
- Nagpure, B. V., and Bian, J. S. (2016). Interaction of hydrogen sulfide with nitric oxide in the cardiovascular system. *Oxid. Med. Cell. Longev.* 2016, 6904327. doi: 10.1155/2016/6904327
- Nakahira, K., Haspel, J. A., Rathinam, V. A., Lee, S. J., Dolinay, T., Lam, H. C., et al. (2011). Autophagy proteins regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by the NALP3 inflammasome. *Nat. Immunol.* 12, 222–230. doi: 10.1038/ni.1980
- Ng, H. H., Leo, C. H., Parry, L. J., and Ritchie, R. H. (2018). Relaxin as a therapeutic target for the cardiovascular complications of diabetes. *Front. Pharmacol.* 9, 501. doi: 10.3389/fphar.2018.00501
- Oh, G. S., Pae, H. O., Lee, B. S., Kim, B. N., Kim, J. M., Kim, H. R., et al. (2006). Hydrogen sulfide inhibits nitric oxide production and nuclear factor-kappaB via heme oxygenase-1 expression in RAW264.7 macrophages stimulated with lipopolysaccharide. *Free Radic. Biol. Med.* 41, 106–119. doi: 10.1016/j.freeradbiomed.2006.03.021
- Ooi, B. K., Chan, K. G., Goh, B. H., and Yap, W. H. (2018). The role of natural products in targeting cardiovascular diseases via Nrf2 pathway: novel molecular mechanisms and therapeutic approaches. *Front. Pharmacol.* 9, 1308. doi: 10.3389/fphar.2018.01308
- Pan, L. L., Qin, M., Liu, X. H., and Zhu, Y. Z. (2017). The role of hydrogen sulfide on cardiovascular homeostasis: an overview with update on immunomodulation. *Front. Pharmacol.* 8, 686. doi: 10.3389/fphar.2017.00686
- Papapetropoulos, A., Pyriochou, A., Altaany, Z., Yang, G., Marazioti, A., Zhou, Z., et al. (2009). Hydrogen sulfide is an endogenous stimulator of angiogenesis. *Proc. Natl. Acad. Sci. U. S. A.* 106, 21972–21977. doi: 10.1073/pnas.0908047106
- Park-Windhol, C., and D'Amore, P. A. (2016). Disorders of Vascular Permeability. *Annu. Rev. Pathol.* 11, 251–281. doi: 10.1146/annurev-pathol-012615-044506
- Peh, M. T., Anwar, A. B., Ng, D. S., Atan, M. S., Kumar, S. D., and Moore, P. K. (2014). Effect of feeding a high fat diet on hydrogen sulfide (H2S) metabolism in the mouse. *Nitric. Oxide* 41, 138–145. doi: 10.1016/j.niox.2014.03.002
- Perna, A. F., Sepe, I., Lanza, D., Capasso, R., Zappavigna, S., Capasso, G., et al. (2013). Hydrogen sulfide reduces cell adhesion and relevant inflammatory triggering by preventing ADAM17-dependent TNF-alpha activation. *J. Cell Biochem.* 114, 1536–1548. doi: 10.1002/jcb.24495
- Possomato-Vieira, J. S., and Goncalves-Rizzi, V. H. (2018). Clinical and experimental evidences of hydrogen sulfide involvement in lead-induced hypertension. *BioMed. Res. Int.* 2018, 4627391. doi: 10.1155/2018/4627391
- Predmore, B. L., Julian, D., and Cardounel, A. J. (2011). Hydrogen sulfide increases nitric oxide production from endothelial cells by an akt-dependent mechanism. *Front. Physiol.* 2, 104. doi: 10.3389/fphys.2011.00104
- Rahimi, N. (2017). Defenders and challengers of endothelial barrier function. *Front. Immunol.* 8, 1847. doi: 10.3389/fimmu.2017.01847
- Rahman, M. S., and Woollard, K. (2017). Atherosclerosis. *Adv. Exp. Med. Biol.* 1003, 121–144. doi: 10.1007/978-3-319-57613-8_7
- Roy, A., Khan, A. H., Islam, M. T., Prieto, M. C., and Majid, D. S. (2012). Interdependency of cystathione gamma-lyase and cystathione beta-synthase in hydrogen sulfide-induced blood pressure regulation in rats. *Am. J. Hypertens.* 25, 74–81. doi: 10.1038/ajh.2011.149
- Schleifenbaum, J., Kohn, C., Voblova, N., Dubrovskaya, G., Zavarirskaya, O., Gloe, T., et al. (2010). Systemic peripheral artery relaxation by KCNQ channel openers and hydrogen sulfide. *J. Hypertens.* 28, 1875–1882. doi: 10.1097/HJH.0b013e32833c20d5
- Sen, U., Sathnur, P. B., Kundu, S., Givvimani, S., Coley, D. M., Mishra, P. K., et al. (2012). Increased endogenous H2S generation by CBS, CSE, and 3MST gene therapy improves ex vivo renovascular relaxation in hyperhomocysteinemia. *Am. J. Physiol. Cell Physiol.* 303, C41–C51. doi: 10.1152/ajpcell.00398.2011
- Streeter, E. Y., Badoer, E., Woodman, O. L., and Hart, J. L. (2013). Effect of type 1 diabetes on the production and vasoactivity of hydrogen sulfide in rat middle cerebral arteries. *Physiol. Rep.* 1, e00111. doi: 10.1002/phy2.111
- Sun, L., Sun, S., Li, Y., Pan, W., Xie, Y., Wang, S., et al. (2014). Potential biomarkers predicting risk of pulmonary hypertension in congenital heart disease: the role of homocysteine and hydrogen sulfide. *Chin. Med. J. (Engl)* 127, 893–899. doi: 10.3760/cma.j.issn.0366-6999.20132694
- Sun, H. J., Hou, B., Wang, X., Zhu, X. X., Li, K. X., and Qiu, L. Y. (2016). Endothelial dysfunction and cardiometabolic diseases: Role of long non-coding RNAs. *Life Sci.* 167, 6–11. doi: 10.1016/j.lfs.2016.11.005
- Sun, H. J., Zhu, X. X., Cai, W. W., and Qiu, L. Y. (2017). Functional roles of exosomes in cardiovascular disorders: a systematic review. *Eur. Rev. Med. Pharmacol. Sci.* 21, 5197–5206. doi: 10.26355/eurrev_201711_13840
- Suryavanshi, S. V., and Kulkarni, Y. A. (2017). NF-kappabeta: a potential target in the management of vascular complications of diabetes. *Front. Pharmacol.* 8, 798. doi: 10.3389/fphar.2017.00798
- Suzuki, K., Olah, G., Modis, K., Coletta, C., Kulp, G., Gero, D., et al. (2011). Hydrogen sulfide replacement therapy protects the vascular endothelium in hyperglycemia by preserving mitochondrial function. *Proc. Natl. Acad. Sci. U. S. A.* 108, 13829–13834. doi: 10.1073/pnas.1105121108
- Suzuki, K., Sagara, M., Aoki, C., Tanaka, S., and Aso, Y. (2017). Clinical implication of plasma hydrogen sulfide levels in Japanese patients with type 2 diabetes. *Intern. Med.* 56, 17–21. doi: 10.2169/internalmedicine.56.7403
- Szabo, C., and Papapetropoulos, A. (2011). Hydrogen sulphide and angiogenesis: mechanisms and applications. *Br. J. Pharmacol.* 164, 853–865. doi: 10.1111/j.1476-5381.2010.01191.x
- Szabo, C. (2007). Hydrogen sulphide and its therapeutic potential. *Nat. Rev. Drug Discovery* 6, 917–935. doi: 10.1038/nrd2425
- Tahara, Y., Aoyama, S., and Shibata, S. (2017). The mammalian circadian clock and its entrainment by stress and exercise. *J. Physiol. Sci.* 67, 1–10. doi: 10.1007/s12576-016-0450-7
- Tang, G., Yang, G., Jiang, B., Ju, Y., Wu, L., and Wang, R. (2013). H(2)S is an endothelium-derived hyperpolarizing factor. *Antioxid. Redox Signal.* 19, 1634–1646. doi: 10.1089/ars.2008.2117
- Tang, W. H., Kitai, T., and Hazen, S. L. (2017). Gut microbiota in cardiovascular health and disease. *Circ. Res.* 120, 1183–1196. doi: 10.1161/CIRCRESAHA.117.309715

- Tang, W. H. W., Backhed, F., Landmesser, U., and Hazen, S. L. (2019). Intestinal microbiota in cardiovascular health and disease: JACC state-of-the-art review. *J. Am. Coll. Cardiol.* 73, 2089–2105. doi: 10.1016/j.jacc.2019.03.024
- Tao, B. B., Liu, S. Y., Zhang, C. C., Fu, W., Cai, W. J., Wang, Y., et al. (2013). VEGFR2 functions as an H₂S-targeting receptor protein kinase with its novel Cys1045-Cys1024 disulfide bond serving as a specific molecular switch for hydrogen sulfide actions in vascular endothelial cells. *Antioxid. Redox Signal.* 19, 448–464. doi: 10.1089/ars.2012.4565
- Tao, B., Wang, R., Sun, C., and Zhu, Y. (2017). 3-Mercaptopyruvate sulfurtransferase, not cystathionine beta-synthase nor cystathionine gamma-lyase, mediates hypoxia-induced migration of vascular endothelial cells. *Front. Pharmacol.* 8, 657. doi: 10.3389/fphar.2017.00657
- Tomasova, L., Pavlovicova, M., Malekova, L., Misak, A., Kristek, F., Grman, M., et al. (2015). Effects of AP39, a novel triphenylphosphonium derivatised anethole dithiolethione hydrogen sulfide donor, on rat haemodynamic parameters and chloride and calcium Cav3 and RyR2 channels. *Nitric. Oxide* 46, 131–144. doi: 10.1016/j.niox.2014.12.012
- Van Den Born, J. C., Mencke, R., Conroy, S., Zeebregts, C. J., Van Goor, H., and Hillebrands, J. L. (2016). Cystathionine gamma-lyase is expressed in human atherosclerotic plaque microvessels and is involved in micro-angiogenesis. *Sci. Rep.* 6, 34608. doi: 10.1038/srep34608
- Voigt, R. M., Forsyth, C. B., Green, S. J., Engen, P. A., and Keshavarzian, A. (2016). Circadian rhythm and the gut microbiome. *Int. Rev. Neurobiol.* 131, 193–205. doi: 10.1016/bs.irn.2016.07.002
- Wallace, J. L., Motta, J. P., and Buret, A. G. (2018). Hydrogen sulfide: an agent of stability at the microbiome-mucosa interface. *Am. J. Physiol. Gastrointest. Liver Physiol.* 314, G143–G149. doi: 10.1152/ajpgi.00249.2017
- Wang, Y., Zhao, X., Jin, H., Wei, H., Li, W., Bu, D., et al. (2009). Role of hydrogen sulfide in the development of atherosclerotic lesions in apolipoprotein E knockout mice. *Arterioscler. Thromb. Vasc. Biol.* 29, 173–179. doi: 10.1161/ATVBAHA.108.179333
- Wang, M. J., Cai, W. J., Li, N., Ding, Y. J., Chen, Y., and Zhu, Y. C. (2010a). The hydrogen sulfide donor NaHS promotes angiogenesis in a rat model of hind limb ischemia. *Antioxid. Redox Signal.* 12, 1065–1077. doi: 10.1089/ars.2009.2945
- Wang, M. J., Cai, W. J., and Zhu, Y. C. (2010b). Mechanisms of angiogenesis: role of hydrogen sulphide. *Clin. Exp. Pharmacol. Physiol.* 37, 764–771. doi: 10.1111/j.1440-1681.2010.05371.x
- Wang, K., Ahmad, S., Cai, M., Rennie, J., Fujisawa, T., Crispi, F., et al. (2013a). Dysregulation of hydrogen sulfide producing enzyme cystathionine gamma-lyase contributes to maternal hypertension and placental abnormalities in preeclampsia. *Circulation* 127, 2514–2522. doi: 10.1161/CIRCULATIONAHA.113.001631
- Wang, X. H., Wang, F., You, S. J., Cao, Y. J., Cao, L. D., Han, Q., et al. (2013b). Dysregulation of cystathionine gamma-lyase (CSE)/hydrogen sulfide pathway contributes to ox-LDL-induced inflammation in macrophage. *Cell Signal* 25, 2255–2262. doi: 10.1016/j.cellsig.2013.07.010
- Wang, G. G., Chen, Q. Y., Li, W., Lu, X. H., and Zhao, X. (2015a). Ginkgolide B increases hydrogen sulfide and protects against endothelial dysfunction in diabetic rats. *Croat. Med. J.* 56, 4–13. doi: 10.3325/cmj.2015.56.4
- Wang, R., Szabo, C., Ichinose, F., Ahmed, A., Whiteman, M., and Papapetropoulos, A. (2015b). The role of H₂S bioavailability in endothelial dysfunction. *Trends Pharmacol. Sci.* 36, 568–578. doi: 10.1016/j.tips.2015.05.007
- Wang, Z. J., Wu, J., Guo, W., and Zhu, Y. Z. (2017). Atherosclerosis and the hydrogen sulfide signaling pathway - therapeutic approaches to disease prevention. *Cell Physiol. Biochem.* 42, 859–875. doi: 10.1159/000478628
- Wang, R. (2009). Hydrogen sulfide: a new EDRF. *Kidney Int.* 76, 700–704. doi: 10.1038/ki.2009.221
- Wang, R. (2012). Physiological implications of hydrogen sulfide: a whiff exploration that blossomed. *Physiol. Rev.* 92, 791–896. doi: 10.1152/physrev.00017.2011
- Watson, E. C., Grant, Z. L., and Coultas, L. (2017). Endothelial cell apoptosis in angiogenesis and vessel regression. *Cell Mol. Life Sci.* 74, 4387–4403. doi: 10.1007/s00018-017-2577-y
- Webb, G. D., Lim, L. H., Oh, V. M., Yeo, S. B., Cheong, Y. P., Ali, M. Y., et al. (2008). Contractile and vasorelaxant effects of hydrogen sulfide and its biosynthesis in the human internal mammary artery. *J. Pharmacol. Exp. Ther.* 324, 876–882. doi: 10.1124/jpet.107.133538
- Weber, C., and Noels, H. (2011). Atherosclerosis: current pathogenesis and therapeutic options. *Nat. Med.* 17, 1410–1422. doi: 10.1038/nm.2538
- Wen, Y. D., Wang, H., and Zhu, Y. Z. (2018). The drug developments of hydrogen sulfide on cardiovascular disease. *Oxid. Med. Cell. Longev.* 2018, 4010395. doi: 10.1155/2018/4010395
- White, B. J., Smith, P. A., and Dunn, W. R. (2013). Hydrogen sulphide-mediated vasodilatation involves the release of neurotransmitters from sensory nerves in pressurized mesenteric small arteries isolated from rats. *Br. J. Pharmacol.* 168, 785–793. doi: 10.1111/j.1476-5381.2012.02187.x
- Whiteman, M., and Winyard, P. G. (2011). Hydrogen sulfide and inflammation: the good, the bad, the ugly and the promising. *Expert Rev. Clin. Pharmacol.* 4, 13–32. doi: 10.1586/ecp.10.134
- Whiteman, M., Li, L., Rose, P., Tan, C. H., Parkinson, D. B., and Moore, P. K. (2010). The effect of hydrogen sulfide donors on lipopolysaccharide-induced formation of inflammatory mediators in macrophages. *Antioxid. Redox Signal.* 12, 1147–1154. doi: 10.1089/ars.2009.2899
- Wu, D., and Hu, Q. (2018). An update on hydrogen sulfide and nitric oxide interactions in the cardiovascular system. *Oxid. Med. Cell. Longev.* 2018, 4579140. doi: 10.1155/2018/4579140
- Wu, S. Y., Pan, C. S., Geng, B., Zhao, J., Yu, F., Pang, Y. Z., et al. (2006). Hydrogen sulfide ameliorates vascular calcification induced by vitamin D3 plus nicotine in rats. *Acta Pharmacol. Sin.* 27, 299–306. doi: 10.1111/j.1745-7254.2006.00283.x
- Wu, D. B., Chen, J. F., Xu, Q., Lin, J. Q., Liao, J. Q., and Wu, W. (2016). [Exogenous hydrogen sulfide inhibits high-glucose-induced injuries via regulating leptin/leptin receptor signaling pathway in human umbilical vein endothelial cells]. *Nan Fang Yi Ke Da Xue Xue Bao* 36, 1055–1061.
- Xiao, L., and Dong, J. H. (2016). Hydrogen sulfide improves endothelial dysfunction via downregulating BMP4/COX-2 pathway in rats with hypertension. *Oxid. Med. Cell. Longev.* 2016, 8128957. doi: 10.1155/2016/8128957
- Xiao, L., Dong, J. H., Teng, X., Jin, S., Xue, H. M., Liu, S. Y., et al. (2018). Hydrogen sulfide improves endothelial dysfunction in hypertension by activating peroxisome proliferator-activated receptor delta/endothelial nitric oxide synthase signaling. *J. Hypertens.* 36, 651–665. doi: 10.1097/HJH.0000000000001605
- Xiaohui, L., Junbao, D., Lin, S., Jian, L., Xiuying, T., Jianguang, Q., et al. (2005). Down-regulation of endogenous hydrogen sulfide pathway in pulmonary hypertension and pulmonary vascular structural remodeling induced by high pulmonary blood flow in rats. *Circ. J.* 69, 1418–1424. doi: 10.1253/circj.69.1418
- Xie, L., Gu, Y., Wen, M., Zhao, S., Wang, W., Ma, Y., et al. (2016). Hydrogen sulfide induces Keap1-S-sulphydration and suppresses diabetes-accelerated atherosclerosis via Nrf2 activation. *Diabetes* 65, 3171–3184. doi: 10.2337/db16-0020
- Xu, S., Liu, Z., and Liu, P. (2014). Targeting hydrogen sulfide as a promising therapeutic strategy for atherosclerosis. *Int. J. Cardiol.* 172, 313–317. doi: 10.1016/j.ijcard.2014.01.068
- Xue, H., Zhou, S., Xiao, L., Guo, Q., Liu, S., and Wu, Y. (2015). Hydrogen sulfide improves the endothelial dysfunction in renovascular hypertensive rats. *Physiol. Res.* 64, 663–672.
- Yamamoto, J., Sato, W., Kosugi, T., Yamamoto, T., Kimura, T., Taniguchi, S., et al. (2013). Distribution of hydrogen sulfide (H₂S)-producing enzymes and the roles of the H₂S donor sodium hydrosulfide in diabetic nephropathy. *Clin. Exp. Nephrol.* 17, 32–40. doi: 10.1007/s10157-012-0670-y
- Yan, H., Du, J., and Tang, C. (2004). The possible role of hydrogen sulfide on the pathogenesis of spontaneous hypertension in rats. *Biochem. Biophys. Res. Commun.* 313, 22–27. doi: 10.1016/j.bbrc.2003.11.081
- Yang, G., Wu, L., Jiang, B., Yang, W., Qi, J., Cao, K., et al. (2008). H₂S as a physiologic vasorelaxant: hypertension in mice with deletion of cystathionine gamma-lyase. *Science* 322, 587–590. doi: 10.1126/science.1162667
- Yu, B., Chen, Q., Le Bras, A., Zhang, L., and Xu, Q. (2018). Vascular Stem/Progenitor Cell Migration and Differentiation in Atherosclerosis. *Antioxid. Redox Signal.* 29, 219–235. doi: 10.1089/ars.2017.7171
- Yuan, S., Shen, X., and Kevil, C. G. (2017). Beyond a gasotransmitter: hydrogen sulfide and polysulfide in cardiovascular health and immune response. *Antioxid. Redox Signal.* 27, 634–653. doi: 10.1089/ars.2017.7096
- Zanardo, R. C., Brancaleone, V., Distrutti, E., Fiorucci, S., Cirino, G., and Wallace, J. L. (2006). Hydrogen sulfide is an endogenous modulator of

- leukocyte-mediated inflammation. *FASEB J.* 20, 2118–2120. doi: 10.1096/fj.06-6270fje
- Zhang, H., Zhi, L., Mochhala, S. M., Moore, P. K., and Bhatia, M. (2007). Endogenous hydrogen sulfide regulates leukocyte trafficking in cecal ligation and puncture-induced sepsis. *J. Leukoc. Biol.* 82, 894–905. doi: 10.1161/CIRCULATIONAHA.109.866889
- Zhang, D., Jiang, X., Fang, P., Yan, Y., Song, J., Gupta, S., et al. (2009). Hyperhomocysteinemia promotes inflammatory monocyte generation and accelerates atherosclerosis in transgenic cystathionine beta-synthase-deficient mice. *Circulation* 120, 1893–1902. doi: 10.1161/CIRCULATIONAHA.109.866889
- Zhang, D., Fang, P., Jiang, X., Nelson, J., Moore, J. K., Kruger, W. D., et al. (2012a). Severe hyperhomocysteinemia promotes bone marrow-derived and resident inflammatory monocyte differentiation and atherosclerosis in LDLr/CBS-deficient mice. *Circ. Res.* 111, 37–49. doi: 10.1161/CIRCRESAHA.112.26947
- Zhang, H., Guo, C., Wu, D., Zhang, A., Gu, T., Wang, L., et al. (2012b). Hydrogen sulfide inhibits the development of atherosclerosis with suppressing CX3CR1 and CX3CL1 expression. *PLoS One* 7, e41147. doi: 10.1371/journal.pone.0041147
- Zhang, D., Wang, X., Tian, X., Zhang, L., Yang, G., Tao, Y., et al. (2018a). The increased endogenous sulfur dioxide acts as a compensatory mechanism for the downregulated endogenous hydrogen sulfide pathway in the endothelial cell inflammation. *Front. Immunol.* 9, 882. doi: 10.3389/fimmu.2018.00882
- Zhang, X., Sessa, W. C., and Fernandez-Hernando, C. (2018b). Endothelial transcytosis of lipoproteins in atherosclerosis. *Front. Cardiovasc. Med.* 5, 130. doi: 10.3389/fcvm.2018.00130
- Zhao, W., Zhang, J., Lu, Y., and Wang, R. (2001). The vasorelaxant effect of H₂S as a novel endogenous gaseous K(ATP) channel opener. *EMBO J.* 20, 6008–6016. doi: 10.1093/emboj/20.21.6008
- Zhao, H., Lu, S., Chai, J., Zhang, Y., Ma, X., Chen, J., et al. (2017). Hydrogen sulfide improves diabetic wound healing in ob/ob mice via attenuating inflammation. *J. Diabetes Complications* 31, 1363–1369. doi: 10.1016/j.jdiacomp.2017.06.011
- Zheng, Y. F., Dai, D. Z., and Dai, Y. (2010). NaHS ameliorates diabetic vascular injury by correcting depressed connexin 43 and 40 in the vasculature in streptozotocin-injected rats. *J. Pharm. Pharmacol.* 62, 615–621. doi: 10.1211/jpp/62.05.0009
- Zhong, S., Li, L., Shen, X., Li, Q., Xu, W., Wang, X., et al. (2019). An update on lipid oxidation and inflammation in cardiovascular diseases. *Free Radic. Biol. Med.* 144, 266–278. doi: 10.1016/j.freeradbiomed
- Zhou, J., Li, Y. S., and Chien, S. (2014). Shear stress-initiated signaling and its regulation of endothelial function. *Arterioscler. Thromb. Vasc. Biol.* 34, 2191–2198. doi: 10.1161/ATVBAHA.114.303422
- Zhou, Y., Li, X. H., Zhang, C. C., Wang, M. J., Xue, W. L., Wu, D. D., et al. (2016). Hydrogen sulfide promotes angiogenesis by downregulating miR-640 via the VEGFR2/mTOR pathway. *Am. J. Physiol. Cell Physiol.* 310, C305–C317. doi: 10.1152/ajpcell.00230.2015
- Zuidema, M. Y., and Korthuis, R. J. (2015). Intravital microscopic methods to evaluate anti-inflammatory effects and signaling mechanisms evoked by hydrogen sulfide. *Methods Enzymol.* 555, 93–125. doi: 10.1016/bs.mie.2014.11.022
- Zuidema, M. Y., Yang, Y., Wang, M., Kalogeris, T., Liu, Y., Meininger, C. J., et al. (2010). Antecedent hydrogen sulfide elicits an anti-inflammatory phenotype in postischemic murine small intestine: role of BK channels. *Am. J. Physiol. Heart Circ. Physiol.* 299, H1554–H1567. doi: 10.1152/ajpheart.01229.2009

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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