

ROLE OF GENETIC SUSCEPTIBILITY IN NICOTINE ADDICTION AND CHRONIC OBSTRUCTIVE PULMONARY DISEASE

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ABSTRACT

Chronic obstructive pulmonary disease (COPD) is a leading cause of morbidity and mortality in developed countries. Although cigarette smoking is the major risk factor, only 10-20% of smokers develop COPD. The extent of cigarette smoking (pack-years and smoking duration) accounts for only 15% of the variation in lung function, indicating that differences in susceptibility to COPD must exist. We provide an overview of the complexity of nicotine addiction and COPD, with special attention to the involvement of genetic factors. The following aspects are discussed in the present article: (1) epidemiology in Mexico and (2) a review of the published literature on genetic association studies using the National Center for Biotechnology Information database of the United States as a search tool. COPD is unique among complex genetic diseases where an environmental risk factor is known and the level of exposure can be documented with some precision. The high morbidity and mortality associated with COPD and its chronic and progressive nature has prompted the use of molecular genetic studies to identify susceptibility factors for the disease. Biomedical research has a remarkable set of tools to aid in the discovery of genes and polymorphisms. We present a review of the most relevant genetic associations in nicotine addiction and COPD. (REV INVEST CLIN. 2019;71:36-54)

Key words: Chronic obstructive pulmonary disease. Exacerbations. Nicotine addiction. Genetic susceptibility. Single-nucleotide polymorphisms. HLA.

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INTRODUCTION

Smoking is the leading cause of preventable death worldwide and is considered a public health problem. The World Health Organization reports that smoking kills more than 5 million people each year. Moreover, 11% of deaths caused by ischemic cardiomyopathy and > 70% caused by lung cancer and chronic obstructive pulmonary disease (COPD) have smoking in common¹.

COPD is one of the ten main causes of morbidity and mortality in adult populations worldwide, with a prevalence of 9% in Europe, > 11% in Latin America, 8.6% in Japan, and 6% in the United States²⁻⁴. In Mexico, it is also one of the top ten causes of morbidity and mortality^{5,6}. A review study found that 54-77% of patients with COPD of moderate severity were smokers, while 38-51% of those with very severe stages of the disease also smoked⁷. Nicotine is among the substances found in cigarettes and produces addiction by first affecting the central nervous system. However, after chronic smoking, the smoker can present an inflammatory syndrome, which slowly progresses, causing multisystem damage⁸. It has been reported that there are different addiction levels. Some smokers smoke less throughout their lifetime and hardly show symptoms of addiction; they are known as chippers. On the other hand, there are heavy smokers who show high nicotine addiction. In general, the latter are less successful in quitting smoking⁹.

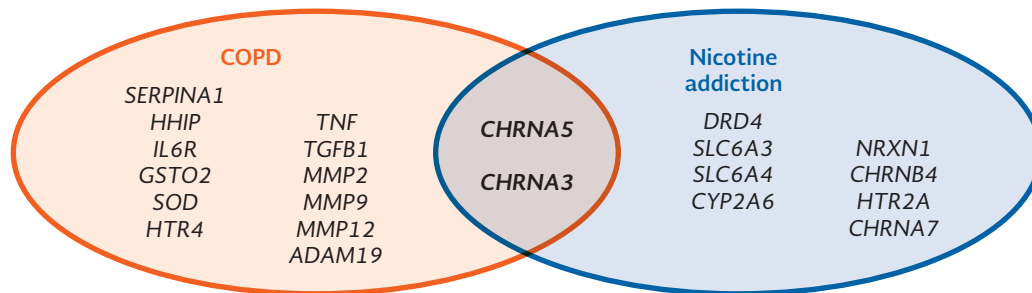
Globally, one billion people will die from tobacco-related illnesses this century if trends in tobacco use remain unchecked. In low–middle-income countries such as Mexico, the prevalence of tobacco smoking has not changed¹⁰. The classic model of addiction suggests that cigarette consumption increases to a level where regular nicotine administration helps smokers avoid withdrawal symptoms¹¹. Mexican smokers are more likely to be non-daily smokers and to consume a lower number of cigarettes per day (CPD) compared to smokers from the majority of ethnic groups in other countries. Little is known about their quit behaviors¹². Mexico was an early signer of the framework convention on tobacco control and is a world leader in public health approaches to tobacco control¹³. At the national level, the National Tobacco Control Agency (Oficina Nacional

para el Control del Tabaco) has instituted health warnings on cigarette packages, pursued a vigorous prevention and cessation media campaign, and produced detailed economic evaluations of the costs of the tobacco epidemic¹⁴. However, despite all these efforts, the smoking prevalence in Mexico has remained unchanged¹⁵. According to the Global Burden of Disease 2015, while the smoking prevalence declined between 1990 and 2005, these gains were not maintained. In 2010, we began to see an increase in the prevalence of daily smokers, particularly among those between 25 and 34 years of age and who are female¹³. Consequently, over 14.3 million Mexican adults (16.4%) currently smoke, and it is expected that 4 million will die from tobacco-related diseases in the next decade if the smoking prevalence remains stable. As described in the most-recent Global Adult Tobacco Survey in Mexico, the static smoking prevalence is in part attributable to the fact that fewer than 10% of Mexican smokers take advantage of evidence-based approaches to smoking cessation¹⁵. The potential for reducing the projected morbidity and mortality associated with smoking depends greatly on reaching and treating current smokers.

The combination of counseling and pharmacotherapy has been shown to be one of the most effective and feasible interventions for smoking cessation¹⁶. Primary care systems in Mexico follow established guidelines for the identification and treatment of smokers. Although health-care providers are usually encouraged to address smoking with their patients at every visit, most providers fail to initiate cessation treatment, even when cessation resources are available, as they are in Mexico. The lack of time during routine patient care, inadequate training of personnel, and competing patient demands are other factors related to the limited implementation of these cessation guidelines¹⁷.

In Mexico, tobacco use is causally related to up to 5% of the annual reported mortality¹⁸. For the Mexican Social Security Institute (Instituto Mexicano del Seguro Social), smoking generates an annual cost of up to 7082 million pesos (376 million US dollars) simply to address three of the related diseases (lung cancer, chronic obstructive lung disease, and brain-cardiovascular diseases), which represents 4.3% of the total health-care cost of this institution⁵. To continue in

Figure 1. Main genes associated with chronic obstructive pulmonary disease and nicotine addiction, as well as those that have been associated in both pathologies.



the path to decrease the smoking prevalence in Mexico, it is essential to expand scientific efforts to understand nicotine dependence beyond what we currently know.

NICOTINE ADDICTION AND COPD ARE COMPLEX AND MULTIFACTORIAL DISEASES

The reinforcement effect of nicotine is the main reason for its consumption. However, between 10% and 20% of smokers never show addiction. In addition to the environmental and sociocultural context, genetic variants associated with a higher risk of cigarette smoking have been described mainly in Caucasian and African American populations. Studies in twins have demonstrated that the genetic component for cigarette smoking accounts for up to 35%¹⁹.

In some cases, nicotine addiction causes an increase in cigarette smoking. Damage to lung function occurs in approximately 40% of smokers, which could result in the development and progression of COPD²⁰. Studies in twins report that 60% of the individual susceptibility to develop COPD depends on genetic factors²¹. On the other hand, 18.6% of COPD patients have at least one family member with the same pathology and have a higher number of exacerbations and a poorer quality of life²².

Nicotine addiction and COPD are complex and multifactorial diseases, where the genetic component contributes to their development (Fig. 1). To identify genetic variants associated with these types of diseases, there are different genetic strategies, including

cohort analysis, twin or family studies, and case/control group analysis.

Candidate gene studies are one of the older strategies used in genetic association studies. This method evaluates genes that encode proteins involved in biological pathways known in a particular illness. In the case of nicotine addiction, genes encoding for brain neurotransmitter receptors and involved in nicotine metabolism have been evaluated.

With new technological advances in genomics, other methods have been developed to accelerate the identification of genes associated with complex diseases, such as genome-wide association studies (GWASs). GWAS analyze hundreds of thousands to millions of single-nucleotide polymorphisms (SNPs), a strategy that provides a unique opportunity for exploring the genome without any hypothesis²³.

POLYMORPHISMS ASSOCIATED WITH NICOTINE ADDICTION IDENTIFIED BY THE CANDIDATE FUNCTIONAL GENE STRATEGY

In the studies of candidate genes, polymorphisms associated with nicotine addiction have been identified in gene encoding proteins involved in the reward and pleasure mechanisms in the central nervous system. The dopaminergic pathway was the first one found among the pathways explored.

The dopamine D4 receptor, specifically a variable number tandem repeats (VNTR) polymorphism, is found among the genes with higher evidence of

association with the nicotine addiction process. The biological importance translates to alterations in the length of the receptor. The 48-base pair (bp) VNTR in exon 3 encodes for the third intracellular domain of the protein. Short alleles, known as S (4 repeats), have less signaling efficiency compared to long alleles, known as L (7 repeats). A study of African-American smokers and non-smokers found a significant association between elevated risk of smoking and those individuals encoding the long allele (L). However, the same study analyzed a Caucasian population and did not find such an association²⁴. The Caucasian population analysis revealed the association of the presence of the long allele (9 repeats) in the gene *SLC6A3* (dopamine transporter) with an increase in the number of cigarettes smoked and the anxiety level²⁵. In contrast, in Polish populations, those individuals who did not harbor the long allele in *SLC6A3* had a higher risk of smoking before the age of 20, smoked a greater number of cigarettes daily, and had less abstinence compared to those carrying the short allele²⁶.

The serotonergic system is involved in nicotine consumption. The serotonin transporter maintains serotonin concentrations in the nerve synapse, and the gene encoding for the pertinent protein (solute carrier family 6 member 4, *SLC6A4*) is a study candidate for addiction, given that it is related to addiction and depression. The promoter region of the gene is related to transcriptional efficiency and two common alleles - a 44-bp insertion (allele L) or deletion (allele S). Allele S has been associated with a decrease in transcriptional activity compared to allele L. In male Chinese smokers, genotypes L/L and S/L were found with a higher frequency compared to non-smokers. Moreover, such genotypes were associated with cigarette smoking and nicotine dependence²⁷.

POLYMORPHISMS IN GENES RELATED TO NICOTINE ADDICTION IDENTIFIED BY GWAS

Using the GWAS strategy, genes involved in the addiction process not previously considered as functional candidates have been identified. Such is the case for neurexin-1 (*NRXN1*) and neurexin-3. The neurexin gene family encodes a group of cell-surface proteins that are mainly expressed in neurons; they

are necessary for neurotransmitter release and are a key factor in synapse genesis. Three SNPs in the intronic region associated with nicotine addiction were identified in African-American and Caucasian populations. Due to their location, these SNPs will likely affect the alternative splicing of the mRNA, which generates isoforms that affect the formation of neural circuits²⁸. Subsequently, rs1882296 in *NRXN1* was associated with smoking risk in a Mexican population when comparing smokers to non-smokers. Using an *in silico* analysis, the presence of the risk allele (C) was observed to generate a miRNA capable of silencing genes that encode different receptors of the GABAergic and glutamatergic pathways compared to the presence of the common allele (T)²⁹.

Vacuolar protein sorting 13 homolog A (*VPS13A*), homolog of *VPS13A* in *Saccharomyces cerevisiae* and brain-derived neurotrophic factor (*BDNF*) are examples of genes identified using the GWAS strategy. In *VPS13A*, SNP variants associated with a higher nicotine addiction risk have been found. In addition, variants of this gene have been observed to cause progressive neurodegeneration³⁰. In a meta-analysis, genetic markers located in a region close to the *BDNF* gene were identified to be associated with tobacco consumption. This gene encodes proteins of the neurotrophin family which regulate the plasticity and survival of dopaminergic and cholinergic neurons. It is likely that variations in *BDNF* can alter the rewarding effects of nicotine through the modulation of dopamine reward circuits. On the other hand, an association between rs3025343 of the gene dopamine beta-hydroxylase and the cessation of tobacco use was observed; the protein that encodes this gene is involved in dopamine metabolism³¹.

One of the genomic regions with a higher replication in nicotine addiction is the group of genes formed by *CHRNA5*, *CHRNA3*, and *CHRNB4*. In 2009, Stevens et al. compared casual smokers (< 5 CPD) with heavy smokers (≥ 30 CPD) and identified 13 SNPs associated with the group of heavy smokers³². In the same year, a study on nicotine dependence in Caucasian and African-American populations included individuals with a low score (0-1) in the Fagerström test as the control group and individuals with a high score (≥ 4) as the nicotine-dependent group³³. They found five SNPs with significant associations in *CHRNA5*, 11 in *CHRNA3*, and one in *CHRNB4*. In 2010, two SNPs

associated with the number of cigarettes consumed in Korean populations were reported in the same genetic region³⁴.

Variant rs16969968 in the gene *CHRNA5* is the most replicated in association with nicotine dependence. This polymorphism causes a change in the amino acid sequence of aspartic acid (allele G) by asparagine (allele A) in position 398 of the protein (D398N), causing a change in the charge of the second intracellular domain of subunit $\alpha 5$ ³⁵.

Stimulation with acetylcholine, nicotine, or varenicline under the presence of asparagine in subunit $\alpha 5$ in cellular assays causes the nicotine-cholinergic receptors (nAChR) to respond more slowly compared to the subunit containing aspartic acid in position 398 (398N)³⁶. The presence of allele A correlates with high expression levels in sputum and an increase in the risk of airway obstruction, even in non-smokers³⁷. *In vitro* assays in lung epithelial cells show silencing of the expression of *CHRNA5* with a decrease in the expression of adhesion molecules and an increase in cell migration capacity, contributing to airway damage³⁸.

The decrease in the function of subunit $\alpha 5$ is associated with a significant risk of nicotine dependence; those subjects with allele A (398N) require a higher amount of nicotine to activate the dopaminergic pathway³⁹. An example is a report in a European population, where individuals carrying the homozygous AA genotype are smokers consuming ≥ 20 CPD⁴⁰. This change affects brain signaling in subjects who consume tobacco, causing significant changes in the ventral striatum, amygdala, and hippocampus regions⁴¹. Other reports indicate that smokers carrying variant A of rs16969968 are more likely to have strong memories associated with cigarette smoking, suggesting that allele A has an important role in the memory process⁴². In 2010, the tobacco and genetics consortium performed a meta-analysis of 16 original studies on nicotine dependence with some phenotypes associated with tobacco consumption; among their results, *CHRNA5* polymorphisms were highlighted to be associated with cigarettes consumed per day, in particular, rs16969968³¹. Another important finding was allele A of rs1051730, which corresponds to an increase of 1 in the number of cigarettes smoked.

PHYSIOPATHOLOGY OF COPD

COPD is a broad term used to describe a group of lung disorders that share damage of the lung parenchyma (emphysema) and bronchial damage, which lead to the progressive obstruction of airflow. It is a multifactorial condition associated with exposure to inhaled toxic substances⁴³. COPD has a progressive nature, with an increased presence of signs and symptoms over time, although it is known that the individual impact is quite variable. Likewise, only a fraction of smokers develops COPD, which highlights its multi-genetic nature⁴⁴.

COPD is characterized by an abnormal chronic inflammatory response in the lung that causes irreversible structural changes. All smokers develop some level of lung inflammation, but in those developing COPD, the inflammation is more intense and harmful. Under normal conditions, humans can recover from the damage, and the healing process starts with an inflammatory response, important for the repair, and tissue remodeling processes. Inflammatory cells, mostly neutrophils, macrophages, and T CD8 lymphocytes, attract different mediators which remove the inhaled irritants. At the end of the process, the inflammation recedes. In contrast, in COPD, the airways remain continuously exposed to a great variety of toxic substances from the tobacco smoke, resulting in a continuous and amplified inflammatory reaction⁴⁵. Cellular infiltrates are observed in the walls of large and small airways from the trachea to the alveolar ducts, including alveolar walls and blood vessels⁴⁶.

Tobacco smoke exposes cells and tissues to high concentrations of oxidants and free radicals⁴⁷, such as oxygen and nitrogen reactive species, which result in oxidative stress due to an imbalance with the natural antioxidant processes^{47,48}. This oxidative stress directly contributes to the inflammatory process in the lung⁴⁶, causing damage and cell death⁴⁹. There is also an imbalance with proteases⁵⁰, where cells such as neutrophils and macrophages release important mediators called proteases that are significant in mediating protection mechanisms. The natural balance to avoid the exaggerated action of these proteases is altered with the oxidative potential of cigarette smoke. Thus, endogenous antiproteases such as $\alpha 1$ antitrypsin (AAT), antichymotrypsin,

and elafin oxidize and are inactivated, causing a protease-antiprotease imbalance. The excess of elastase not only causes emphysema but also promotes a higher production of mucins. Furthermore, tobacco smoke directly induces the death of lung cells (apoptosis)⁵¹. COPD damage starts in the respiratory epithelium, the portion most directly exposed to tobacco smoke and its cocktail of toxic chemicals⁵². Bronchial damage leads to chronic bronchitis, characterized by the presence of cough and chronic production of sputum. At the cellular level, there is a steep decrease in the ciliary apparatus in bronchial epithelial cells that predispose the individual to bacterial colonization, infections, and acute damage episodes called exacerbations⁵³. There is also hyperplasia of mucus cells at the epithelial and submucosal gland levels. Other changes contributing to the chronic airway obstructions and air trapping include the development of peribronchial fibrosis and bronchospasm. Moreover, there is a gradual decrease in lung elasticity due to the progressive destruction of elastic proteins due to excessive protease. This destruction of the alveolar walls (emphysema) causes a marked reduction in the surface area necessary for gas exchange to occur.

COPD AND GENETIC SUSCEPTIBILITY FACTORS

Among the environmental risk factors, the most clearly associated is cigarette smoke exposure. Although the dose-response relationship between cigarette smoke and lung function is well established, there is an important variability in the level of airflow obstruction that occurs as a response to cigarette smoke^{54,55}. The low percentage of the variation in lung function explained by cigarette smoke (approximately 15%), and the existence of early-onset cases with severely reduced lung function suggests variability in the individual genetic susceptibility to the effects of cigarette smoke^{55,56}. In addition, it must be considered that only between 10% and 20% of patients with COPD are or were smokers⁵⁷, indicating that there are other factors, including genetic, which confer susceptibility to develop the disease.

The deficiency of AAT (AATd) is considered a genetic risk factor, but it explains only < 1% of the cases⁵⁷. On the other hand, a large variety of genes have been proposed as possibly involved in the

development of COPD. These include α 1 antichymotrypsin (*SERPINA3*)⁵⁸⁻⁶⁰, certain extracellular matrix metalloproteinases (MMPs) such as *MMP1*⁶¹ and *MMP9*⁶² and their inhibitor (tissue inhibitor of metalloproteinases 2)⁶³, heme-oxygenase (*HMOX1*)⁶⁴, microsomal epoxide hydrolase (*EPHX1*)⁶⁵, glutathione S-transferase p1 (*GSTP1*)⁶⁶, and glutathione S-transferase m1⁶⁷, lung surfactant proteins b and d (*SFT-PB*)⁶⁸ and *SFTPD*⁶⁹, Vitamin D-binding protein (*GC*)⁷⁰, interleukin (IL) 13⁷¹, tumor necrosis factor α (*TNF- α*)⁷²⁻⁷⁴, C-C motif chemokine ligand 174, decorin, transforming growth factor β 1 (*TGF β 1*)⁷⁴, and the adrenergic receptor β 2 (*ADRB2*)⁷⁵ as some examples. The previously established associations of individual genes include biological pathways with diverse functions, including immune response, inflammation, xenobiotic metabolism, antioxidants, and protease/antiprotease systems⁷⁶.

Studies based on families have demonstrated an increase in the risk of lung function damage in the first-degree relatives of smoking or ex-smoking patients⁵⁵. Lung function studies in twins provide additional evidence on the role of genetic factors in determining lung function in the general population⁷⁷. Moreover, since the 1970s, several authors have reported an increase in the prevalence of airway obstruction among family members (first-degree relatives) of patients with COPD compared with the family members of control subjects⁷⁸. In addition, the existence of variability in lung function measurements has been described, with a high heritability rate⁷⁹ as well as severe and early-onset forms⁵⁵.

Recently, six new loci (in or close to *EFEMP1*, *BMP6*, *MIR129-2-HSD17B12*, *PRDM11*, *WVVOX*, and *KCNJ2*) have been associated with forced vital capacity (FVC) in a population of Caucasian descent, and two loci previously associated with spirometry measurements (*GSTCD* and *PTCH1*) related to FVC were identified using a GWAS meta-analysis, achieving the detection in human lung tissue of transcripts of all six genes recently involved⁸⁰. Such loci could be implicated in lung development mechanisms and in the pathogenesis of restrictive lung disease. Two new loci (*IL16/STARD5/TMC3* on chromosome 15 and *ME3* on chromosome 11) were previously identified and associated with the forced expiratory volume in the first second (FEV1) change rate that contains candidate genes with a biologically feasible involvement in lung function⁸¹.

POLYMORPHISMS IN GENES ASSOCIATED WITH COPD SECONDARY TO TOBACCO SMOKING

Polymorphisms found within the *SERPINA1* gene, encoding AAT, are among the genetic factors that are clearly associated with the development of COPD. It is a highly polymorphic gene, with > 100 variants of the protein described, of which 30 could have a pathological consequence⁸². Protein variants are classified according to their electrophoresis migration velocity in a magnetic field with different pH gradients. Pioneering researchers in this area designated these proteins as M (medium) for those with medium velocity, F (fast) for those with fast migration, and S (slow) for those with slow migration⁸³. When new variants were discovered, the anodic proteins were assigned the first letters of the alphabet, while the cathodic ones were assigned the last letters⁸². The normal genotype, present in > 90% of healthy individuals (94-96% in Caucasian populations), is called PiMM and is characterized by serum levels of approximately 150-350 mg/100 mL. Variants S (Glu264Val) and Z (Glu342Lys) constitute 95% of the mutations in patients with severe AAT deficiency; both are nonsense mutations in the *SERPINA1* gene (rs17580 and rs28929474, respectively). Individuals with genotypes SS, SZ, and ZZ express serum concentrations of the protein of 85%, 25%, and 15%, respectively, compared to the normal levels (genotype MM)⁸⁴.

Most studies conclude that genotype SZ is less important than ZZ because patients with genotype SZ develop emphysema at an older age compared with those carrying ZZ⁸². Alleles S and Z encode abnormal proteins that polymerize in the liver, and therefore, 80-90% of PiZ and 40-50% of PiS molecules are retained inside the hepatocyte-forming polymers, usually degraded by the proteasome. In clinical practice, the risk of having diseases related to AAT deficiency is limited to the phenotypes of ZZ (96%)⁸². The progressive decline of lung function in homozygous ZZ individuals is well known; regarding the medium deficiency genotypes (MZ and MS), it was determined that, compared to genotype MM, MZ is associated to a reduction in lung function in individuals with COPD. Genotypes SZ and ZZ are associated with airway obstructions and lung function reduction, particularly in smokers^{85,86}.

The prevalence of these polymorphisms is < 5.0% (0.2% for PiZ and 3.42% for PiS) in Mexican Mestizo population. However, the presence of the heterozygous variant PiS (AT) is associated with lower FEV1/FVC ratio compared with those individuals who do not have this polymorphism⁸⁷.

Other genes associated with COPD that contributes in small proportions to this pathology include those encoding the IL 6 receptor (*IL6R*) and glutathione S-transferase omega 2, associated with decreases in lung function and/or COPD⁷⁹.

Then, based on the known fact that lung function measurements are hereditary traits predicting morbidity and mortality related to COPD development⁸⁸, the cohort of the SpiroMeta consortium (European descent individuals) evaluated the association with lung function measurements (FEV1 and FEV1/FVC). The authors confirmed the formerly reported association between locus 4q31⁸⁹, previously associated with lung function, and COPD⁷⁹.

Interestingly, “protective” alleles have also been described in Hispanic populations in the USA⁹⁰, a finding that has been replicated in Mexican populations in the genes *IL6R* and *ADAM19*⁹¹.

GWAS IN THE STUDY OF COPD

Multiple studies have evaluated the individual involvement of genetic markers, mainly SNPs, in COPD susceptibility, while few include whole genome searches for new variants to find alternative or novel hypotheses for the underlying mechanisms of the pathology.

In the Framingham Heart Study, a GWAS was performed to identify genetic markers related to lung function measurements, identifying SNPs in two genes not previously described using “traditional” methods associated with lung function measurements. In this study, participants were grouped by phenotype according to the spirometry results. The authors suggested that, for SNPs with significant results, especially those located in genes with unknown biological function, a replica in another group of individuals is required⁷⁹. Pillai et al. reported, in 2009, a GWAS in COPD where they used the

multiple stage replication technique. Initially, in the study, 538,030 SNPs were analyzed in a group of cases and controls, obtaining a list of associations, from which the 100 SNPs (“top 100”) most strongly associated ($p = 1.6E-04$) were selected. These SNPs were characterized in an additional cohort of families, which they labeled replication Stage I. Based on the results from replicate I, the seven SNPs with the best association evidence in the joint analysis of both stages ($p < 1.0E-07$) were selected and analyzed in an additional case and control population, labeled replication Stage II. Finally, they performed a study for the validation of results in a cohort of individuals with COPD, resulting in six validated SNPs with a significant association to the disease. It is important to note that they found genes that had not been previously associated with COPD but that play a key role in other pathologies such as lung cancer. Such is the case of the *CHRNA3/5* cluster. Another gene highly associated with COPD is *HHIP*, about which little is known, but the authors suggest that, based on their results, it should be extensively studied⁹². Likewise, 70,978 SNPs were analyzed in a GWAS related to lung function measurements in the Framingham study cohort. Their results propose the *HHIP* gene as one of the possible regulators in COPD and suggest the importance of the gene encoding glycoporphin A (GYPA) in the disease due to its position in relation to *HHIP* and earlier findings of this protein in patients with COPD. Regarding the gene *CHRNA3/5*, they did not obtain significant results⁹³.

Recently, five loci associated with different lung function measurements were identified using GWAS associated with lung function measurements (FEV1 and FEV1/FVC)⁸⁹. In addition, the authors performed a meta-analysis with the strongest association signals from direct genotyping, a summary association of *in silico* data of the CHARGE consortium, and ultimately included data from the Health 2000 study, confirming the previously reported association of locus 4q31 as well as the associations with FEV1 and/or the FEV1/FVC ratio and common variants in five loci. The analysis of mRNA showed expression of *TNS1*, *GSTCD*, *AGER*, *HTR4*, and *THSD4* in human lung tissue. Finally, they suggest that these associations offer signals of the mechanisms that regulate lung function and indicate potential targets for therapeutic interventions in respiratory pathologies⁸⁹. In

general, 17 regions were observed to be associated independently with FEV1, and 23 were associated to the value of the FEV1/FVC ratio, including three regions (4q24 in *GSTCD*, 4q31 near *HHIP*, and 15q23 in *THSD4*). The SNP rs12504628 was found to be associated with both FEV1/FVC and FEV1 traits; it is located in an intergenic region close to *HHIP*, covering ~300 kb of the 4q31 region that was previously associated with lung function and COPD⁹³. The hedgehog gene family, of which *HHIP* is a member, encodes molecules involved in the regulation of lung morphogenesis, suggesting other overlying mechanisms for such associations⁹⁴.

In parallel, a study analyzing lung function measurements was published using a meta-analysis-like design and GWAS. This study included 20,890 participants of European descent (the same used in a study by Repapi et al.⁸⁹, defining eight loci associated with the FEV1/FVC ratio (*HHIP*, *GPR126*, *ADAM19*, *AGER-PPT2*, *FAM13A*, *PTCH1*, *PID1*, and *HTR4*) and one locus associated with FEV1 value (*INSTS12-GSTCD-NPNT*)^{94,95}. Several SNPs near the *HHIP* coding region were associated with the FEV1/FVC ratio in both the CHARGE and SpiroMeta consortia, confirming previous findings performed in the Framingham Heart Study⁷⁹.

THE CANDIDATE GENE STRATEGY IN COPD

On the other hand, association studies using the candidate gene strategy in cases and controls are frequently criticized due to the lack of population replicates^{96,97}, even finding contradictory results in some cases or the absence of association with previously identified genetic markers⁹⁸. In this regard, Hersh et al. performed a replication study in 2005 based on families and in parallel on cases and controls using the candidate gene strategy and individual genetic associations related to COPD development, where 29 polymorphisms were evaluated (24 SNPs, 1 insertion/deletion [indel], 3 STR, and 1 nonsense deletion) in 12 candidate genes previously reported on the literature. They found significant associations related to distinct qualitative and/or quantitative phenotypes with different SNPs, including *TNF α* (-308G > A), *SFTPB* (thr131ile), *HMOX1* (gt31), and *EPHX1* (his139arg)⁷⁶.

GENES RELATED TO INFLAMMATION AND THEIR ROLE IN COPD

Different mediators and cells have been involved in the pathology of COPD, including $TNF\alpha$, IL-8, and TGF- β . Cigarette smoke activates macrophages that release $TNF\alpha$, leukotriene B4, IL-8, and other neutrophil chemotactic factors, as well as antiproteases. This reaction increases the local inflammation, with a predominance of neutrophils and protease release⁹⁹. IL-8 is a chemokine that mediates the activation and migration of neutrophils from peripheral blood into the tissue. It plays a significant role at the onset of the amplification of the inflammatory response. A transversion (A→T) in position -351 and another in -251 of the *IL-8* promoter have been associated with COPD in different populations^{100,101} but are inversely associated with bronchial asthma^{102,103}.

An SNP was previously described in the promoter region within the *TNF* gene that was identified because it directly affects the transcriptional regulation of the gene¹⁰⁴. Several studies show certain relevance for the SNP -308G/A of *TNF* in Asian populations but not in Caucasian ones. For example, in Japanese and Taiwanese populations, an increase in the prevalence of COPD has been observed compared to their respective control groups⁷². However, these results have not been confirmed in other populations^{105,106}, and given the location of the *TNF* gene within the HLA region, it would be important to establish the existence of extended haplotypes that include the classic HLA genes. Recently, our group identified certain genetic variants associated with both the susceptibility and severity of COPD related to smoking¹⁰⁷ and smoke exposure due to the burning of biomass¹⁰⁸.

The expression of HLA-DR and CD80 in the cell surface of alveolar macrophages (AM) in patients with COPD is decreased compared to smokers with normal lung function and to non-smokers. In addition, these patients show an increased percentage of AM with low levels of expression of CD44 on the surface¹⁰⁹. Genes encoding HLA antigens, especially class II, are particularly interesting to us.

ASSOCIATION BETWEEN HLA ALLELES AND COPD

Despite the contribution of allelic variation of the HLA system to several diseases, mainly autoimmune and

inflammatory, the role of HLA alleles in COPD has not been extensively studied; only a limited number of studies have described this association¹¹⁰. Among the few reports on the association of this disorder with the HLA system, the 1990 study by Sugiyama et al. typified antigens HLA-A, HLA-B, and HLA-C using serological methods in patients with diffuse panbronchiolitis. This is a form of COPD with unknown etiology, which is clinically characterized by chronic cough, sputum, and dyspnea, physiologically by the limitations of airflow, and histologically by inflammatory lesions around the bronchioles, with an increase in the frequency of HLA-B54 antigen¹¹¹. Although the number of patients in this study was relatively small, it is important to mention that HLA-B54 is a serotype mainly found in East Asians. Subsequently, Keicho et al. discovered, in 1998, a positive association between PBD and alleles HLA-A11, HLA-B54, and HLA-Cw1, while HLA-A33 and HLA-B44 increased their frequencies in the control groups, suggesting a protective association for these alleles in Japanese populations¹¹². In addition, the findings for A11 were replicated in Korean populations, but those for B54 and Cw1 were not; in contrast, HLA-B55 is increased, and HLA-B62 and HLA-Cw4 are weakly associated¹¹³. Regarding Caucasian populations, the oldest record in the literature is the one from Kauffmann et al., who in 1983 demonstrated that HLA-B7 is a genetic risk factor for developing COPD; the finding is worthy of mentioning given that HLA-B7 is predominant in Caucasian populations¹¹⁴. Kasuga et al., in 2005, performed a replication study where the tentative association between HLA-B7 and COPD described by Kauffmann was analyzed using a case-control design in smokers. They studied the contribution of HLA-B7 to the decay of lung function, which was measured by FEV1, but they did not find significant differences in HLA-B7 frequencies between COPD patients and patients without airway obstruction nor an association between HLA-B7 and lung function decline. These findings allow us to conclude that HLA-B7 does not contribute to COPD development and is not involved in lung function variations¹¹⁰.

GENES OF EXTRACELLULAR MMPs AND COPD

MMPs are a family of at least 20 proteolytic enzymes that have an essential role in tissue remodeling. Most

MMP genes contain hundreds of polymorphisms¹¹⁵, some of which are associated with COPD. At least 20 polymorphisms in *MMP1* gene have been associated with COPD¹¹⁶, mainly in regulatory regions (promoter, 3'UTR, 5'UTR, and introns)¹¹⁷. Some genetic studies associating MMPs with COPD include genes such as *MMP1*, *MMP2*, *MMP3*, *MMP9*, and *MMP12*.

The insertion/deletion polymorphism 1G-1607/2G in the promoter region of *MMP1*, which increases transcription due to the introduction of a new binding site for the ETS-1 transcription factor¹¹⁸, has been evaluated in relation to lung function decline, with controversial results^{119,120}. In Russian population, the homozygous genotype 2G was associated with risk in COPD patients¹²¹, a finding that was not replicated elsewhere^{121,123}. In addition, rs1799750 in *MMP1* was identified as associated with the mainly apical densitometric distribution of emphysema¹²⁴, while the carriers of allele T of rs470358 are associated with the transference of gases in patients with COPD secondary to AATd¹²⁵. Finally, polymorphisms in *MMP1* have also been associated with the predisposition to occupational chronic bronchitis¹²⁶.

In *MMP2*, the homozygous genotype T of the polymorphism rs243865 (C-1306T), located in the intron, shows a significant association with the FEV1 decrease in Caucasian populations¹²⁷, while in Tunisian populations, it was correlated with disease severity¹²⁸. In Mexican populations, rs243864 and rs11646643 were associated with the risk of COPD, and in addition, MMP-2 serum levels were lower¹²⁹.

The identification of MMP-3 in COPD is recent. At the genetic level, it has been described that, in patients with AATd, those carrying allele rs678815/G in *MMP3* shows a lower transference of gases¹²⁵. Genotype 6A6A of polymorphism -1171 5A > 6A (rs35068180) was associated with a significantly high risk of COPD in Russian population¹²³ and with COPD lung cancer in Polish population¹³⁰. Polymorphisms rs3025058 and rs678815 have been proposed as susceptibility markers for the development of lung cancer in individuals with COPD¹³¹.

Another well-explored gene in this metalloproteinase family is *MMP9*. Polymorphism rs13925 (C-1562T), located in the gene's promoter, is associated with COPD development in Japanese⁶² and Chinese

populations^{132,133} and was recently confirmed in a meta-analysis including Caucasian and Asian populations¹³⁴. In Russian populations, allele T and the homozygous genotype TT are associated with increased severity and younger age of disease onset¹²². In Northern European populations, rs3918242 was associated with centrilobular emphysema¹³⁴. In Tunisian populations, a significant correlation was identified between polymorphism 279 R/Q (836G > A) and disease severity. Likewise, Q homozygotes (genotype AA) were associated with a drop in FEV1 and FVC in patients with COPD compared to individuals with genotypes AG and GG. In contrast, the activity of MMP-9 improved in individuals carrying allele R (G) compared to those homozygous for variant Q (A)¹³⁵. Our group used SNP tagging and determined that the rs3918253 of *MMP9* associated with COPD risk increased in serum levels in a group of patients with stable COPD¹²⁹.

Polymorphism rs2276109 (-82A/G) of gene *MMP12* was associated with an increase in the promoter activity and has an effect in the *cis* element of transcription factor AP-1¹³⁶. It is also associated with a positive effect on lung function in children with asthma and adult smokers. This allele is further associated with a reduced risk of COPD in adult smokers¹³⁷. Although only one tendency was identified in Polish populations¹³⁰, in Bulgaria, patients carrying genotypes with at least one copy of allele G could have a lower risk of COPD¹³⁸.

Recently, it has been proposed that COPD has its origin in early childhood, by demonstrating associations between several important genes related to COPD and early transient symptoms (wheezing and decrease in lung function). The associated genetic variants include rs2276109 in *MMP12*¹³⁷. In Indian populations, this polymorphism showed an association in additive and dominant models, while the infrequent allele (G) showed a significant positive association with lung function in different genetic models. In contrast, haplotypes carrying allele A showed a negative correlation with lung function¹³⁹. Genetic associations have also been described for SNPs in or near *MMP12* with different lung emphysema patterns quantified by automated tomography¹⁴⁰. This polymorphism has been demonstrated to be associated with the severity of lung disease¹⁴¹. Recently, with the advent of exome analysis, the associations of SNPs

rs17368582 and rs2276109 could be replicated¹⁴². Polymorphism Asp357Ser (rs652438) was associated with the decline rate in lung function in Caucasian smokers⁶¹ and with COPD in Chinese populations¹⁴³. The common variant (Ser) in codon 357 is associated with clinical manifestations that are consistent with a more aggressive matrix degradation in other tissues; it is associated with an increased severity of the disease in patients with COPD¹³⁷.

GENETIC CONSIDERATIONS IN COPD EXACERBATIONS

Hospitalizations are an important part of patient care and are directly related to the deterioration of the health status¹⁴⁴, mortality increase¹⁴⁵, and a substantial rise in health-care costs¹⁴⁶. Hospital readmissions are common and occur in an alarming 60% of patients in the 1st year after the last exacerbation^{147,148}.

At the molecular level, a reduction in the expression of HLA-DR on the surface of peripheral-blood mononuclear cells has been described, as well as a higher frequency of allele QBP 5.12 in the promoter of the gene *HLA-DQB1* in patients under exacerbation¹⁴⁹.

Infectious exacerbations are common in patients with COPD, where cells and molecules of the immune response are involved at different levels. The protein mannose-binding lectin (MBL) is a key component in the defense of the host against an infection¹⁵⁰. It is a pattern recognition receptor that binds to carbohydrates on the microorganism's surface, which, in turn, activates the complement system^{151,152}. The protein MBL is encoded by the gene *MBL2*; patients carrying allele D in codon 54 of exon 1 of *MBL2* (rs1800450 G/A), which at the protein level consists of a substitution of glycine (G) by aspartate (D), have more risk of hospitalization due to infectious exacerbation¹⁵³. In recurrent exacerbations, the mortality and frequency of infectious exacerbations were significantly higher in patients with deficient *MBL* genotypes than in those with non-deficient *MBL* genotypes¹⁵⁴.

Lung surfactant is a multimolecule complex formed by phospholipids and cholesterol (90% in total) and surfactant proteins (SPs) (10%); multiple proteins of its composition have been associated with respiratory pathology. Among them, those known as lung SPs can

be found. They are composed of the high molecular weight hydrophilic proteins SP-A and SP-D; the extraordinarily lipophilic, low-molecular weight SP-B; and the SP-C, which are vital for the biophysical properties of surfactant phospholipids¹⁵⁵. SP-D has immunomodulatory functions¹⁵⁶; by keeping a potentially aggressive inflammatory reaction under control, SP-D decreases the production of proteases and oxidants¹⁵⁷. The association between rs3024791 in *SFTPB* (SP-B) and COPD exacerbation was confirmed using logistic regression models. Negative binomial regression models demonstrated the association of multiple SNPs (rs2118177, rs2304566, rs1130866, and rs3024791) with exacerbation rates¹⁵⁸.

Sialic acid-binding immunoglobulin-like lectins (Siglecs) are a group of cell-surface transmembrane receptors expressed on immune cells that regulate the immune equilibrium in inflammatory diseases¹⁵⁹. Homozygotes for the null allele in *SIGLEC14* are associated with a reduced risk of exacerbation in a population of Japanese patients¹⁶⁰, while allele G of rs2075803 and haplotype GA (rs2075803 G/rs2258983 A) in *SIGLEC9* were associated with a higher frequency of exacerbations and with the emphysema level¹⁶¹.

Using site-directed mutagenesis and recombinant expression studies, it was found that several polymorphisms of the gene *ADRB2* alter the function of *ADRB2*. Of these, polymorphisms in positions 16 (Arg16Gly) and 27 (Gln27Glu) are prevalent, are functionally relevant, and have been widely studied from the clinical and pharmacogenetic perspective.

The homozygous genotype Arg16Arg predisposes patients to a clinically more severe episode¹⁶², greater dyspnea, more symptoms, and frequent exacerbations¹⁶³. Patients with genotype Arg16Arg in *ADRB2* showed better results during exacerbation in response to salmeterol than genotypes Gly16Gly and Arg16Gly, which suggests a potential differential effect of genotype Arg16Gly in the response to the treatment with long-acting β -agonists¹⁶⁴.

Haplotype Gly16/Glu27 can affect the severity of the obstructive ventilatory alteration but not the immediate response to salbutamol during the acute exacerbation¹⁶⁵. On the other hand, two independent pharmacogenetic studies in patients with

moderate-to-severe COPD showed that the therapeutic response and the tolerance to long-term treatment with formoterol (alone or in combination with budesonide) were not modified by the genotype of Arg16Gly¹⁶⁶. Another receptor involved in the pharmacological response is the muscarinic cholinergic receptor 2 (*CHRM2*). Recently, a significant association of polymorphism rs1824024 in *CHRM2* with disease severity was identified, with lower values in lung function tests, frequent exacerbations, and a poor response to anticholinergic drugs¹⁶⁷.

Other agents associated with COPD exacerbations include biological pathways such as the metabolism of xenobiotics, glucocorticoids, and inflammation. *GSTP1* (rs1695), superoxide dismutase 2 (*SOD2*) (rs4880), and *NFE2L2* (rs1806649) were associated with a tendency toward a higher risk of hospital admissions during the periods with high PM10 levels¹⁵⁴.

Individuals with allele C for rs4588 in the *GC* gene showed a higher exacerbation frequency, higher susceptibility to COPD, emphysema, and a tendency toward a rapid decrease of airflow obstruction¹⁶⁸. The minor allele of SNP rs2227744G > A in the protease-activated receptor 1 (*F2R*) was associated with protection against frequent exacerbations¹⁶⁹, while genotype DD of the gene *ACE* was associated with a lower risk in males¹⁷⁰.

GENETIC POLYMORPHISMS RELATED TO NICOTINE ADDICTION AND COPD

Polymorphisms associated with nicotine addiction and COPD have been identified. Among genes with such pleiotropic effects are those encoding nAChR, specifically subunits $\alpha 5$ and $\alpha 3$ (*CHRNA5-CHRNA3*)^{171,172}. These genes not only encode subunits that can be part of the neuronal nAChR but also form part of the muscle receptors.

The nicotine that enters the body of the smoker activates these receptors in the brain, unleashing the signaling for the release of neurotransmitters, with dopamine being the most important in the addiction process. This increase in dopamine in the brain acts as a positive reinforcer, causing the constant need to smoke cigarettes^{173,174}. It has been observed *in vitro* that nicotine reversibly induces the

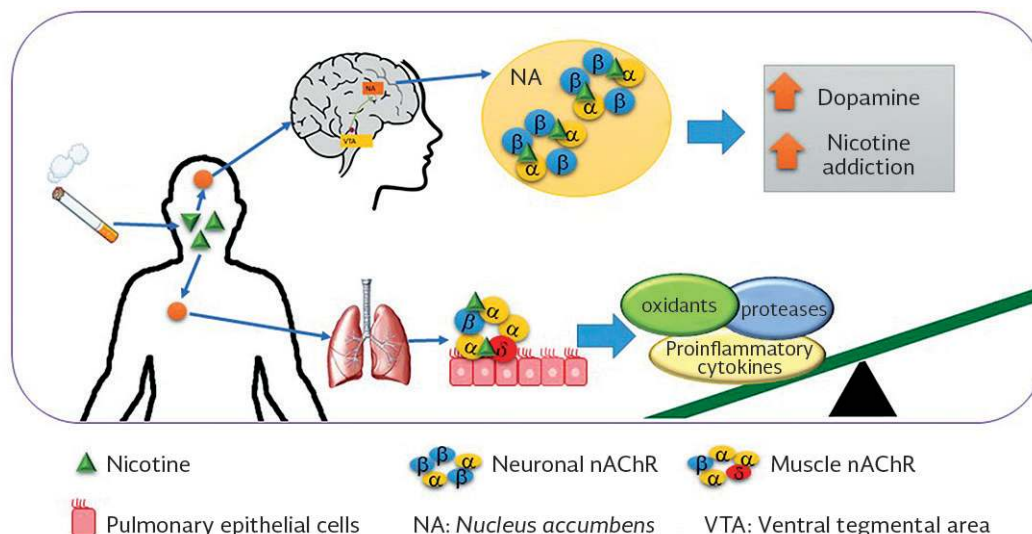
expression of *CHRNA5*, which could be contributing to the nicotine addiction mechanism¹⁷⁵. The non-neuronal nAChR that contain subunits formed by proteins encoded by *CHRNA3* and *CHRNA5* are expressed in airway cells (bronchial epithelial cells, neutrophil macrophages, monocytes, and lymphocytes) and are also activated when nicotine enters the lungs, causing the release of proteases, pro-inflammatory cytokines, and oxidants that are responsible for lung remodeling, which leads to lung damage after chronic exposure¹⁷⁶ (Fig. 2).

SNP rs16969968 in *CHRNA5* is among the SNPs localized in genes encoding nAChR with the highest evidence of association with nicotine addiction, CPD, and COPD. A meta-analysis including 34 studies (case-control studies, families, and cohorts) reported that allele A of this SNP was associated ($p = 5.96E-31$) with a higher risk of cigarette smoking in European populations when comparing heavy (CPD > 20) versus casual smokers (CPD \leq 10)¹⁷⁷. This polymorphism was also associated with airway obstruction independent of cigarette smoking in seven European population cohorts³⁷, and when Asian populations are analyzed, the association with COPD remains when the risk allele (A) is present.

Other interesting polymorphisms in the cluster of genes in *CHRNA3* are rs6495309 (T/C) and rs1051730 (C/T). The latter was evaluated in smokers and non-smokers in Denmark; in smokers, the presence of genotype TT was associated with the risk of suffering from COPD in more severe stages (GOLD III-IV)¹⁷⁸. In a Dutch cohort of smoking individuals, it was reported that carriers of allele T (rs1051730) showed an increased risk of presenting bronchial obstruction and emphysema¹⁷⁹. It has been reported that carrying a copy of allele T increases smoking by 1.2 CPD, suggesting that this region of chromosome 15 has an important function in nicotine levels in smokers¹⁸⁰. On the other hand, for rs6495309 in populations from Southeast China, the presence of allele C was reported to be a risk factor for developing COPD and leads to a higher annual decrease in FEV1. Later, cellular assays suggested that the presence of this allele increases the transcriptional activity of *CHRNA3*¹⁸¹.

In a Russian population that included patients with COPD and clinically healthy individuals (26%

Figure 2. Proposal of the dual participation of nicotinic cholinergic receptors in nicotine addiction and chronic obstructive pulmonary disease.



non-smokers), it was found that the haplotype formed by alleles AAG in *CHRNA3/A5* (rs16969968-rs1051730-r6495309) was present at a higher frequency in patients with COPD than in the control group¹⁸². In Mestizo Mexican populations, we have identified, using multi-stage association studies, polymorphisms in *CHRNA5* (rs16969968 and rs17408276) that are associated with cigarette smoking²⁹. A later analysis, including smokers with and without COPD, showed that patients with COPD and of Caucasian descent had a haplotype formed by 12 SNPs in *CHRNA5/A3* (AGGAAGAGGGCA) that includes rs16969968 and rs17408276. This group of genetic variants was strongly associated with the risk of suffering from COPD; the same behavior is not reported in those individuals of mostly Amerindian ancestral descent¹⁸³.

AGING AND COPD

With increasing age, even in healthy individuals, lung function declines and the respiratory system also experiences structural changes that include pulmonary remodeling, limited tissue regeneration, and an enhanced susceptibility to pulmonary diseases¹⁸⁴. Age-related alterations in the respiratory tract originate from cumulative damage or oxidative stress occurring throughout life. The impaired DNA repair capacity

leads to accumulation of oxidative damaged DNA and cell senescence¹⁸⁵. There is increasing clinical and cellular evidence for the concept that accelerated aging can serve as an underlying mechanism of COPD¹⁸⁶. An accelerated cell senescence in alveolar Type II epithelial cells has been observed in COPD patients¹⁸⁷. Moreover, telomere length dysfunction, decreased expression of anti-aging molecules such as sirtuins, increased expression of senescence-associated molecules including p16 and p21, and β -galactosidase activity have been observed in lungs of patients with COPD¹⁸⁷⁻¹⁹⁰.

Telomere shortening, one of the proposed nine hallmarks of aging¹⁹¹, has been implicated in a variety of lung diseases such as idiopathic pulmonary fibrosis, COPD, emphysema, and lung cancer¹⁹²⁻¹⁹⁴. Telomeres are the protective structures at the end of chromosomes that consist of stretches of repetitive hexanucleotides (5'-TTAGGG-3'). They protect the end of chromosomes from erosion or fusion by DNA repair processes. Telomeres become progressively shorter as cells divide, due to an event that is known as the end-replication process¹⁹⁵. The enzyme telomerase is capable of maintaining telomere regeneration, but its activity is limited mainly to embryonic and adult stem cells. Telomeres in regular somatic cells are therefore reduced with each replication until critical telomere length is reached leading to cell cycle arrest¹⁸⁵.

Many studies have tried to shed some light on the interaction between telomere length and COPD. Shortened telomeres have been described in current and former smokers in comparison with non-smokers¹⁹⁶. Several studies have documented shorter telomeres in circulating leukocytes in patients with COPD than age-matched controls¹⁹⁷⁻¹⁹⁹. Interestingly, shortened telomeres measured in peripheral leukocytes of patients with COPD have been related to all-cause and cancer mortality²⁰⁰ supporting the concept that telomere length could serve as a biomarker of disease progression. Recently, the first longitudinal study on telomere dynamics and COPD has shown an accelerated telomere shortening in patients compared to age-matched smoker controls over a 3-year period of follow-up²⁰¹.

However, what do we know about telomere attrition and its relationship with lung function in COPD? The results from different studies are controversial. A study performed over a large cohort of patients with COPD has reported a weak correlation between telomere length and the lung function expressed by the FEV₁²⁰². Others have shown that leukocyte telomere length was the only marker between a panel of aging markers that were associated with lung function (FEV₁)²⁰³. However, the absence of a relationship between telomere shortening and lung function has also been described^{197,199}. A meta-analysis performed over seven studies has found just a modest association between telomere length and FEV₁ decline²⁰⁴. In the same line, a recent study that analyzed other parameters (inspiratory capacity/ total lung capacity, K_{CO}, and PaO₂) besides FEV₁ could not find any association between them or the change in lung function over time and telomere length²⁰¹. Of note, if present, the association of telomere attrition with lung function would not be so strong. Comparisons between different studies are difficult because of many factors that may influence the results, for example, the cohort sample size, diverse stages of disease severity, the proper correction for confounders, or the technique used to measure telomere length.

To expand our knowledge over the role of telomere biology in COPD and to explore about causality, studies on large cohorts of patients with telomere length measurements at multiple time-points are needed. Unraveling the molecular mechanisms that link premature aging with COPD would allow the

development of promising antiaging therapies with impact on disease progression and outcomes in COPD.

SOCIAL IMPACT OF COPD AND ITS RELATIONSHIP WITH GENETICS

Signs and symptoms such as dyspnea, persistent cough, wheezing, sputum, poor tolerance to exercise, weight loss, and fatigue are typical indicators of COPD²⁰⁵, in addition to anxiety and depression^{148,206,207}. Depression as a response to hypoxia in patients with COPD can be influenced by genetic factors, with an autosomal dominant inheritance model²⁰⁸. On the other hand, the severity of depression is higher in patients with COPD than in smokers without COPD. Genotype Val/Val at position 9 of the signal peptide of the SOD2 enzyme is associated with more severe depression, trait anxiety, and state anxiety compared to patients with genotypes Val/Ala and Ala/Ala²⁰⁹. In Japanese populations, rs3794808 in SLC6A4 was correlated with the Hospital Anxiety and Depression Scale score²¹⁰.

CONCLUSION

There are numerous studies that provide evidence of the involvement of a genetic component that contributes to the risk of developing nicotine addiction and COPD. In addition to genetic variability as a component in the susceptibility in its simplistic way, this contribution could be associated to clinical parameters. These include severity, age of onset, increased tobacco intake (as CPD, heavy consumption, high addiction, etc.), the decline of lung function, frequency of exacerbations, alterations in protein levels driving to chronic inflammatory, and/or oxidant processes, often accompanied by histopathological changes mediated by cellular mechanisms easily altered by tobacco smoke and consequent epigenetic changes. This multifactorial nature of nicotine addiction and COPD requires coordinated research from multiple disciplines, moving forward to precision medicine.

REFERENCES

1. World Health Organization. WHO Report on the Global Tobacco Epidemic, 2013: enforcing Bans on Tobacco Advertising, Promotion and Sponsorship. Geneva: World Health Organization; 2013.

2. Menezes AM, Perez-Padilla R, Jardim JR, et al. Chronic obstructive pulmonary disease in five latin American cities (the PLATINO study): A prevalence study. *Lancet*. 2005;366:1875-81.
3. Minakata Y, Sugiura H, Yamagata T, et al. Prevalence of COPD in primary care clinics: Correlation with non-respiratory diseases. *Intern Med*. 2008;47:77-82.
4. Mannino DM, Homa DM, Akinbami LJ, Ford ES, Redd SC. Chronic obstructive pulmonary disease surveillance-United states, 1971-2000. *Respir Care*. 2002;47:1184-99.
5. Reynales-Shigematsu LM, Rodríguez-Bolaños R de los Á, Jiménez JA, Juárez-Márquez SA, Castro-Ríos A, Hernández-Ávila M. Costos de la atención médica atribuibles al consumo de tabaco en el Instituto Mexicano del Seguro Social. *Salud Publica Mex. Instituto Nacional de Salud Pública*. 2002;48:s48-64. Available from: http://www.scielosp.org/scielo.php?script=sci_arttext&pid=S0036-36342006000700007&lng=es&nrm=iso&tng=es. [Last accessed on 2018 Mar 2].
6. Secretaría de Salubridad y Asistencia. Instituto Nacional de Salud Pública (México), Centro Nacional de Información y Documentación en Salud (México). *Salud Pública de México. Mexico: Salud Pública de México*. [Secretaría de Salubridad y Asistencia]; 2004. p. 169-85. Available from: http://www.scielo.org.mx/scielo.php?script=sci_arttext&pid=S0036-36342004000200012. [Last accessed on 2018 Mar 23].
7. Tønnesen P. Smoking cessation and COPD. *Eur Respir Rev*. 2013; 22:37-43.
8. Benowitz NL, Hukkanen J, Jacob P 3rd. Nicotine chemistry, metabolism, kinetics and biomarkers. *Handb Exp Pharmacol*. 2009; 192:29-60.
9. Li MD. The genetics of nicotine dependence. *Curr Psychiatry Rep*. 2006;8:158-64.
10. World Health Organization. WHO Report on the Global Tobacco Epidemic 2008 : the MPOWER Package. World Health Organization; 2008. p. 342.
11. U.S. Department of Health and Human Services. The Health Consequences of Smoking: A Report of the Surgeon General. Atlanta, GA: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention, National Center for Chronic Disease Prevention and Health Promotion, Office on Smoking and Health, 2004.
12. Swayampakala K, Thrasher J, Carpenter MJ, et al. Level of cigarette consumption and quit behavior in a population of low-intensity smokers-Longitudinal results from the international tobacco control (ITC) survey in Mexico. *Addict Behav*. 2013; 38:1958-65.
13. Stillman F, Yang G, Figueiredo V, Hernandez-Avila M, Samet J. Building capacity for tobacco control research and policy. *Tob Control*. 2006;15 Suppl 1:i18-23.
14. Regalado-Pineda J, Rodríguez-Ajenjo CJ. La función de la oficina nacional para el control del tabaco en México. *Salud Publica Mex*. 2008;50:355-65. Available from: <http://www.medigraphic.com/cgi-bin/new/resumen.cgi?IDARTICULO=19202>. [Last accessed on 2018 Mar 15].
15. Encuesta Global de Tabaquismo en Adultos. México; 2015, 2017. Available from: <http://www.who.int/tobacco/surveillance/survey/gats/mex-report-2015-spanish.pdf>. [Last accessed on 2018 Mar 15].
16. Secretaría de Salud. Prevención, diagnóstico y tratamiento del consumo de tabaco y humo ajeno, en el primer nivel de atención. 2009;13. Available from: <http://www.cenetec.salud.gob.mx/interior/gpc.html>. 2009. [Last accessed on 2018 Mar 15].
17. Ponciano-Rodríguez G. The urgent need to change the current medical approach on tobacco cessation in latin America. *Salud Publica Mex*. 2010;52 Suppl 2:S366-72.
18. Stevens G, Dias RH, Thomas KJ, et al. Characterizing the epidemiological transition in Mexico: National and subnational burden of diseases, injuries, and risk factors. *PLoS Med* 2008; 5:e125.
19. Boardman JD, Blalock CL, Pampel FC. Trends in the genetic influences on smoking. *J Health Soc Behav*. 2010;51:108-23.
20. Kim DK, Hersh CP, et al. Epidemiology, radiology, and genetics of nicotine dependence in COPD. *Respir Res*. 2011;12:9.
21. Ingebrigtsen T, Thomsen SF, Vestbo J, et al. Genetic influences on chronic obstructive pulmonary disease-a twin study. *Respir Med*. 2010;104:1890-5.
22. Hersh CP, Hokanson JE, Lynch DA, et al. Family history is a risk factor for COPD. *Chest*. 2011;140:343-50.
23. Jimenez-Sanchez G, Silva-Zolezzi I, Hidalgo A, March S. Genomic medicine in Mexico: Initial steps and the road ahead. *Genome Res*. 2008;18:1191-8.
24. Shields PG, Lerman C, Audrain J, et al. Dopamine D4 receptors and the risk of cigarette smoking in African-Americans and caucasians. *Cancer Epidemiol Biomarkers Prev*. 1998;7:453-8.
25. Perkins KA, Lerman C, Grotenthaler A, et al. Dopamine and opioid gene variants are associated with increased smoking reward and reinforcement owing to negative mood. *Behav Pharmacol*. 2008;19:641-9.
26. Sieminska A, Buczkowski K, Jassem E, Niedoszytko M, Tkacz E. Influences of polymorphic variants of DRD2 and SLC6A3 genes, and their combinations on smoking in polish population. *BMC Med Genet*. 2009;10:92.
27. Chu SL, Xiao D, Wang C, Jing H. Association between 5-hydroxytryptamine transporter gene-linked polymorphic region and smoking behavior in chinese males. *Chin Med J (Engl)*. 2009; 122:1365-8.
28. Nussbaum J, Xu Q, Payne TJ, et al. Significant association of the neurexin-1 gene (NRXN1) with nicotine dependence in european-and African-American smokers. *Hum Mol Genet*. 2008; 17:1569-77.
29. Pérez-Rubio G, Pérez-Rodríguez ME, Fernández-López JC, et al. SNPs in NRXN1 and CHRNA5 are associated to smoking and regulation of GABAergic and glutamatergic pathways. *Pharmacogenomics*. 2016;17:1145-58.
30. Bierut LJ, Madden PA, Breslau N, et al. Novel genes identified in a high-density genome wide association study for nicotine dependence. *Hum Mol Genet*. 2007;16:24-35.
31. Tobacco and Genetics Consortium. Genome-wide meta-analyses identify multiple loci associated with smoking behavior. *Nat Genet*. 2010;42:441-7.
32. Stevens VL, Bierut LJ, Talbot JT, et al. Nicotinic receptor gene variants influence susceptibility to heavy smoking. *Cancer Epidemiol Biomarkers Prev*. 2008;17:3517-25.
33. Saccone NL, Wang JC, Breslau N, et al. The CHRNA5-CHRNA3-CHRNA4 nicotinic receptor subunit gene cluster affects risk for nicotine dependence in African-Americans and in European-Americans. *Cancer Res*. 2009;69:6848-56.
34. Li MD, Yoon D, Lee JY, et al. Associations of variants in CHRNA5/A3/B4 gene cluster with smoking behaviors in a Korean population. *PLoS One*. 2010;5:e12183.
35. Saccone SF, Hinrichs AL, Saccone NL, et al. Cholinergic nicotinic receptor genes implicated in a nicotine dependence association study targeting 348 candidate genes with 3713 SNPs. *Hum Mol Genet*. 2007;16:36-49.
36. Tamminen A, Herder P, Li P, et al. Impact of human D398N single nucleotide polymorphism on intracellular calcium response mediated by $\alpha 3\beta 4\alpha 5$ nicotinic acetylcholine receptors. *Neuropharmacology*. 2012;63:1002-11.
37. Wilk JB, Shrine NR, Loehr LR, et al. Genome-wide association studies identify CHRNA5/3 and HTR4 in the development of airflow obstruction. *Am J Respir Crit Care Med*. 2012;186: 622-32.
38. Kraus AM, Hautefeuille AH, et al. CHRNA5 as negative regulator of nicotine signaling in normal and cancer bronchial cells: Effects on motility, migration and p63 expression. *Carcinogenesis*. 2011;32:1388-95.
39. Bierut LJ, Stitzel JA, Wang JC, et al. Variants in nicotinic receptors and risk for nicotine dependence. *Am J Psychiatry*. 2008; 165:1163-71.
40. Lips EH, Gaborieau V, McKay JD, et al. Association between a 15q25 gene variant, smoking quantity and tobacco-related cancers among 17 000 individuals. *Int J Epidemiol*. 2010; 39:563-77.
41. Hong LE, Hodgkinson CA, Yang Y, et al. A genetically modulated, intrinsic cingulate circuit supports human nicotine addiction. *Proc Natl Acad Sci USA*. 2010;107:13509-14.
42. Janes AC, Smoller JW, David SP, et al. Association between CHRNA5 genetic variation at rs16969968 and brain reactivity to smoking images in nicotine dependent women. *Drug Alcohol Depend*. 2012;120:7-13.
43. From the Global Strategy for the Diagnosis, Management and Prevention of COPD, Global Initiative for Chronic Obstructive Lung Disease (GOLD); 2017. Available from: <http://www.goldcopd.org>. [Last accessed on 2018 April 11].
44. Higgins MW, Keller JB, Landis JR, et al. Risk of chronic obstructive pulmonary disease. Collaborative assessment of the validity of the tecumseh index of risk. *Am Rev Respir Dis*. 1984; 130:380-5.
45. Weathington NM, van Houwelingen AH, Noerager BD, et al. A novel peptide CXCR ligand derived from extracellular matrix degradation during airway inflammation. *Nat Med*. 2006; 12:317-23.

46. Saetta M. Airway inflammation in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 1999;160:S17-20.
47. Jones JG, Minty BD, Lawler P, et al. Increased alveolar epithelial permeability in cigarette smokers. *Lancet.* 1980;1:66-8.
48. Tsuchiya M, Asada A, Kasahara E, et al. Smoking a single cigarette rapidly reduces combined concentrations of nitrate and nitrite and concentrations of antioxidants in plasma. *Circulation.* 2002;105:1155-7.
49. Eiserich JP, van der Vliet A, Handelman GJ, Halliwell B, Cross CE. Dietary antioxidants and cigarette smoke-induced biomolecular damage: A complex interaction. *Am J Clin Nutr.* 1995;62:1490S-1500S.
50. Rangasamy T, Cho CY, Thimmulappa RK, et al. Genetic ablation of *nrf2* enhances susceptibility to cigarette smoke-induced emphysema in mice. *J Clin Invest.* 2004;114:1248-59.
51. Shoshani T, Faerman A, Mett I, et al. Identification of a novel hypoxia-inducible factor 1-responsive gene, RTP801, involved in apoptosis. *Mol Cell Biol.* 2002;22:2283-93.
52. Yoshida T, Tuder RM. Pathobiology of cigarette smoke-induced chronic obstructive pulmonary disease. *Physiol Rev.* 2007;87:1047-82.
53. Tetley TD. Inflammatory cells and chronic obstructive pulmonary disease. *Curr Drug Targets Inflamm Allergy.* 2005;4:607-18.
54. Burrows B, Knudson RJ, Cline MG, Lebowitz MD. Quantitative relationships between cigarette smoking and ventilatory function. *Am Rev Respir Dis.* 1977;115:195-205.
55. Silverman EK, Chapman HA, Drazen JM, et al. Genetic epidemiology of severe, early-onset chronic obstructive pulmonary disease. Risk to relatives for airflow obstruction and chronic bronchitis. *Am J Respir Crit Care Med.* 1998;157:1770-8.
56. Silverman EK, Speizer FE. Risk factors for the development of chronic obstructive pulmonary disease. *Med Clin North Am.* 1996;80:501-22.
57. Sandford AJ, Weir TD, Spinelli JJ, Paré PD. Z and S mutations of the alpha1-antitrypsin gene and the risk of chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol.* 1999;20:287-91.
58. Ishii T, Matsuse T, Teramoto S, Matsui H, Hosoi T, Fukuchi Y, et al. Association between alpha-1-antichymotrypsin polymorphism and susceptibility to chronic obstructive pulmonary disease. *Eur J Clin Invest.* 2000;30:543-8.
59. Poller W, Faber JP, Weidinger S, et al. A leucine-to-proline substitution causes a defective alpha 1-antichymotrypsin allele associated with familial obstructive lung disease. *Genomics.* 1993;17:740-3.
60. Poller W, Faber JP, Scholz S, et al. Mis-sense mutation of alpha 1-antichymotrypsin gene associated with chronic lung disease. *Lancet.* 1992;339:1538.
61. Joos L, He JQ, Shepherdson MB, et al. The role of matrix metalloproteinase polymorphisms in the rate of decline in lung function. *Hum Mol Genet.* 2002;11:569-76.
62. Minematsu N, Nakamura H, Tateno H, Nakajima T, Yamaguchi K. Genetic polymorphism in matrix metalloproteinase-9 and pulmonary emphysema. *Biochem Biophys Res Commun.* 2001;289:116-9.
63. Hirano K, Sakamoto T, Uchida Y, et al. Tissue inhibitor of metalloproteinases-2 gene polymorphisms in chronic obstructive pulmonary disease. *Eur Respir J.* 2001;18:748-52.
64. Yamada N, Yamaya M, Okinaga S, et al. Protective effects of heme oxygenase-1 against oxidant-induced injury in the cultured human tracheal epithelium. *Am J Respir Cell Mol Biol.* 1999;21:428-35.
65. Smith CA, Harrison DJ. Association between polymorphism in gene for microsomal epoxide hydrolase and susceptibility to emphysema. *Lancet.* 1997;350:630-3.
66. Ishii T, Matsuse T, Teramoto S, et al. Glutathione S-transferase P1 (GSTP1) polymorphism in patients with chronic obstructive pulmonary disease. *Thorax.* 1999;54:693-6.
67. Harrison DJ, Cantlay AM, Rae F, Lamb D, Smith CA. Frequency of glutathione S-transferase M1 deletion in smokers with emphysema and lung cancer. *Hum Exp Toxicol.* 1997;16:356-60.
68. Guo X, Lin HM, Lin Z, et al. Surfactant protein gene A, B, and D marker alleles in chronic obstructive pulmonary disease of a Mexican population. *Eur Respir J.* 2001;18:482-90.
69. Seifart C, Plagens A, Brödjje D, et al. Surfactant protein B intron 4 variation in German patients with COPD and acute respiratory failure. *Dis Markers.* 2002;18:129-36.
70. Ito I, Nagai S, Hoshino Y, et al. Risk and severity of COPD is associated with the group-specific component of serum globulin 1F allele. *Chest.* 2004;125:63-70.
71. van der Pouw Kraan TC, Küçükaycan M, Bakker AM, et al. Chronic obstructive pulmonary disease is associated with the -1055 IL-13 promoter polymorphism. *Genes Immun.* 2002;3:436-9. Available from: <http://www.nature.com/articles/6363896>. [cited on 2018 Feb 28].
72. Huang SL, Su CH, Chang SC. Tumor necrosis factor-alpha gene polymorphism in chronic bronchitis. *Am J Respir Crit Care Med.* 1997;156:1436-9.
73. Sakao S, Tatsumi K, Igari H, et al. Association of tumor necrosis factor alpha gene promoter polymorphism with the presence of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 2001;163:420-2.
74. Vernooij JH, Küçükaycan M, Jacobs JA, et al. Local and systemic inflammation in patients with chronic obstructive pulmonary disease: Soluble tumor necrosis factor receptors are increased in sputum. *Am J Respir Crit Care Med.* 2002;166:1218-24.
75. Brøgger J, Steen VM, Eiken HG, Gulsvik A, Bakke P. Genetic association between COPD and polymorphisms in TNF, ADRB2 and EPHX1. *Eur Respir J.* 2006;27:682-8.
76. Hersh CP, Demeo DL, Lange C, et al. Attempted replication of reported chronic obstructive pulmonary disease candidate gene associations. *Am J Respir Cell Mol Biol.* 2005;33:71-8.
77. Redline S, Tishler PV, Rosner B, et al. Genotypic and phenotypic similarities in pulmonary function among family members of adult monozygotic and dizygotic twins. *Am J Epidemiol.* 1989;129:827-36.
78. Kueppers F, Miller RD, Gordon H, Hepper NG, Offord K. Familial prevalence of chronic obstructive pulmonary disease in a matched pair study. *Am J Med.* 1977;63:336-42.
79. Wilk JB, Walter RE, Laramie JM, Gottlieb DJ, O'Connor GT. Framingham heart study genome-wide association: Results for pulmonary function measures. *BMC Med Genet.* 2007;8 Suppl 1:S8.
80. Loth DW, Artigas MS, Gharib SA, et al. Genome-wide association analysis identifies six new loci associated with forced vital capacity. *Nat Genet.* 2014;46:669-77.
81. Tang W, Kowgier M, Loth DW, et al. Large-scale genome-wide association studies and meta-analyses of longitudinal change in adult lung function. *PLoS One.* 2014;9:e100776.
82. American Thoracic Society, European Respiratory Society. American thoracic society/European respiratory society statement: Standards for the diagnosis and management of individuals with alpha-1 antitrypsin deficiency. *Am J Respir Crit Care Med.* 2003;168:818-900.
83. Vidal R, Blanco I, Casas F, et al. Diagnóstico y tratamiento del déficit de alfa-1-antitripsina. *Arch Bronconeumol.* 2006;42:645-59.
84. Fregonese L, Stolk J, Frants RR, Veldhuisen B. Alpha-1 antitrypsin null mutations and severity of emphysema. *Respir Med.* 2008;102:876-84.
85. Dahl M, Nordestgaard BG, Lange P, Vestbo J, Tybjaerg-Hansen A. Molecular diagnosis of intermediate and severe alpha(1)-antitrypsin deficiency: MZ individuals with chronic obstructive pulmonary disease may have lower lung function than MM individuals. *Clin Chem.* 2001;47:56-62.
86. Dahl M, Tybjaerg-Hansen A, Lange P, Vestbo J, Nordestgaard BG. Change in lung function and morbidity from chronic obstructive pulmonary disease in alpha1-antitrypsin MZ heterozygotes: A longitudinal study of the general population. *Ann Intern Med.* 2002;136:270-9.
87. Pérez-Rubio G, Jiménez-Valverde LO, Ramírez-Venegas A, et al. Prevalence of alpha-1 antitrypsin high-risk variants in Mexican mestizo population and their association with lung function values. *Arch Bronconeumol.* 2015;51:80-5.
88. Schünemann HJ, Dorn J, Grant BJ, Winkelstein W Jr, Trevisan M. Pulmonary function is a long-term predictor of mortality in the general population: 29-year follow-up of the buffalo health study. *Chest* 2000;118:656-64.
89. Repapi E, Sayers I, Wain LV, et al. Genome-wide association study identifies five loci associated with lung function. *Nat Genet.* 2010;42:36-44.
90. Young RP, Hopkins RJ. A review of the hispanic paradox: Time to spill the beans? *Eur Respir Rev.* 2014;23:439-49.
91. Pérez-Rubio G, Silva-Zolezzi I, Fernández-López JC, et al. Genetic variants in IL6R and ADAM19 are associated with COPD severity in a Mexican mestizo population. *COPD.* 2016;13:610-5.
92. Pillai SG, Ge D, Zhu G, et al. A genome-wide association study in chronic obstructive pulmonary disease (COPD): Identification of two major susceptibility loci. *PLoS Genet.* 2009;5:e1000421.
93. Wilk JB, Chen TH, Gottlieb DJ, et al. A genome-wide association study of pulmonary function measures in the framingham heart study. *PLoS Genet.* 2009;5:e1000429.

94. Miller LA, Wert SE, Clark JC, et al. Role of sonic hedgehog in patterning of tracheal-bronchial cartilage and the peripheral lung. *Dev Dyn*. 2004;231:57-71.
95. Hancock DB, Eijgelsheim M, Wilk JB, et al. Meta-analyses of genome-wide association studies identify multiple loci associated with pulmonary function. *Nat Genet*. 2010;42:45-52.
96. Editorial: freely Associating. *Nat Genet*. 1999;22:1-2. Available from: http://www.nature.com/articles/ng0599_1. [cited on 2018 Mar 1].
97. Hirschhorn JN, Altshuler D. Once and again-issues surrounding replication in genetic association studies. *J Clin Endocrinol Metab*. 2002;87:4438-41.
98. Ioannidis JP, Ntzani EE, Trikalinos TA, Contopoulos-Ioannidis DG. Replication validity of genetic association studies. *Nat Genet*. 2001;29:306-9.
99. Seifart C, Plagens A. Genetics of chronic obstructive pulmonary disease. *Int J Chron Obstruct Pulmon Dis*. 2007;2:541-50.
100. Matheson MC, Ellis JA, Raven J, Walters EH, Abramson MJ. Association of IL8, CXCR2 and TNF-alpha polymorphisms and airway disease. *J Hum Genet*. 2006;51:196-203.
101. Shen M, Vermeulen R, Chapman RS, et al. A report of cytokine polymorphisms and COPD risk in xuan wei, China. *Int J Hyg Environ Health*. 2008;211:352-6.
102. Heinzmann A, Ahlert I, Kurz T, Berner R, Deichmann KA. Association study suggests opposite effects of polymorphisms within IL8 on bronchial asthma and respiratory syncytial virus bronchiolitis. *J Allergy Clin Immunol*. 2004;114:671-6.
103. Shen L, Fahey JV, Hussey SB, et al. Synergy between IL-8 and GM-CSF in reproductive tract epithelial cell secretions promotes enhanced neutrophil chemotaxis. *Cell Immunol*. 2004;230:23-32.
104. Wilson AG, Symons JA, McDowell TL, McDevitt HO, Duff GW. Effects of a polymorphism in the human tumor necrosis factor alpha promoter on transcriptional activation. *Proc Natl Acad Sci U S A*. 1997;94:3195-9.
105. Gane JM, Stockley RA, Sapey E. The rs361525 polymorphism does not increase production of tumor necrosis factor alpha by monocytes from alpha-1 antitrypsin deficient subjects with chronic obstructive pulmonary disease-a pilot study. *J Negat Results Biomed*. 2015;14:20.
106. Seifart C, Dempfle A, Plagens A, et al. TNF-alpha-, TNF-beta-, IL-6-, and IL-10-promoter polymorphisms in patients with chronic obstructive pulmonary disease. *Tissue Antigens*. 2005;65:93-100.
107. Reséndiz-Hernández JM, Sansores RH, Hernández-Zenteno Rde J, et al. Identification of genetic variants in the TNF promoter associated with COPD secondary to tobacco smoking and its severity. *Int J Chron Obstruct Pulmon Dis*. 2015;10:1241-51.
108. Reséndiz-Hernández JM, Ambrocio-Ortiz E, Pérez-Rubio G, et al. TNF promoter polymorphisms are associated with genetic susceptibility in COPD secondary to tobacco smoking and biomass burning. *Int J Chron Obstruct Pulmon Dis*. 2018;13:627-37.
109. Pons AR, Noguera A, Blanquer D, et al. Phenotypic characterisation of alveolar macrophages and peripheral blood monocytes in COPD. *Eur Respir J*. 2005;25:647-52.
110. Kasuga I, Ruan J, Connett JE, Anthonisen NR, Sandford AJ. Lack of association of human leukocyte antigen-B7 with COPD and rate of decline in lung function. *Respir Med*. 2005;99:1528-33.
111. Sugiyama Y, Kudoh S, Maeda H, Suzuki H, Takaku F. Analysis of HLA antigens in patients with diffuse panbronchiolitis. *Am Rev Respir Dis*. 1990;141:1459-62.
112. Keicho N, Tokunaga K, Nakata K, et al. Contribution of HLA genes to genetic predisposition in diffuse panbronchiolitis. *Am J Respir Crit Care Med*. 1998;158:846-50.
113. Park MH, Kim YW, Yoon HI, et al. Association of HLA class I antigens with diffuse panbronchiolitis in Korean patients. *Am J Respir Crit Care Med*. 1999;159:526-9.
114. Kauffmann F, Kleisbauer JP, Cambon-De-Mouzon A, et al. Genetic markers in chronic air-flow limitation. A genetic epidemiologic study. *Am Rev Respir Dis*. 1983;127:263-9.
115. Vlaykova T, Dimov D. Polymorphisms of matrix metalloproteinases (MMP) in COPD. *Biotechnol Biotechnol Equip*. 2012;26 Suppl 1:111-9. Available from: <http://www.tandfonline.com/doi/abs/10.5504/50YRTIMB.2011.0021>. [Last cited on 2018 Mar 21].
116. Haq I, Chappell S, Johnson SR, et al. Association of MMP-2 polymorphisms with severe and very severe COPD: a case control study of MMPs-1, 9 and 12 in a European population. *BMC Med Genet*. 2010;11:7.
117. Rutter JL, Mitchell TI, Buttice G, et al. A single nucleotide polymorphism in the matrix metalloproteinase-1 promoter creates an ets binding site and augments transcription. *Cancer Res*. 1998;58:5321-5.
118. Affara M, Dunmore BJ, Sanders DA, et al. MMP1 bimodal expression and differential response to inflammatory mediators is linked to promoter polymorphisms. *BMC Genomics*. 2011;12:43.
119. Wallace AM, Mercer BA, He J, et al. Functional characterization of the matrix metalloproteinase-1 cigarette smoke-responsive region and association with the lung health study. *Respir Res*. 2012;13:79.
120. van Diemen CC, Postma DS, Aulchenko YS, et al. Novel strategy to identify genetic risk factors for COPD severity: a genetic isolate. *Eur Respir J*. 2010;35:768-75.
121. Ianbaeva DG, Korytina GF, Viktorova TV. Complex search for antiprotease-protease enzyme gene polymorphisms in patients with chronic obstructive pulmonary diseases. *Mol Biol (Mosk)*. 2004;38:973-9.
122. Korytina GF, Akhmadishina LZ, Ianbaeva DG, Viktorova TV. Polymorphism in promoter regions of matrix metalloproteinases (MMP1, MMP9, and MMP12) in chronic obstructive pulmonary disease patients. *Genetika*. 2008;44:242-9.
123. Korytina GF, Tselousova OS, Akhmadishina LZ, et al. Association of the MMP3, MMP9, ADAM33 and TIMP3 genes polymorphic markers with development and progression of chronic obstructive pulmonary disease. *Mol Biol (Mosk)*. 2012;46:487-99.
124. DeMeo DL, Hersh CP, Hoffman EA, et al. Genetic determinants of emphysema distribution in the national emphysema treatment trial. *Am J Respir Crit Care Med*. 2007;176:42-8.
125. McAloon CJ, Wood AM, Gough SC, Stockley RA. Matrix metalloprotease polymorphisms are associated with gas transfer in alpha 1 antitrypsin deficiency. *Thorax*. 2009;64:323-30.
126. Akhmadishina LZ, Korytina GF, Kochetova OV, Viktorova EV, Viktorova TV. Analysis of polymorphisms of genes associated with immune response and tissue remodeling in occupational chronic bronchitis. *Russ J Genet*. 2014;50:1208-17. Available from: <http://www.link.springer.com/10.1134/S1022795414110027>. [Last cited on 2018 Mar 22].
127. van Diemen CC, Postma DS, Siedlinski M, et al. Genetic variation in TIMP1 but not MMPs predict excess FEV1 decline in two general population-based cohorts. *Respir Res*. 2011;12:57.
128. Bchir S, Nasr HB, Anes AB, et al. MMP-2 (-1306 C/T) polymorphism affects serum matrix metalloproteinase (MMP)-2 levels and correlates with chronic obstructive pulmonary disease severity: a case-control study of MMP-1 and-2 in a tunisian population. *Mol Diagn Ther*. 2016;20:579-90.
129. Hernández-Montoya J, Pérez-Ramos J, Montaña M, et al. Genetic polymorphisms of matrix metalloproteinases and protein levels in chronic obstructive pulmonary disease in a Mexican population. *Biomark Med*. 2015;9:979-88.
130. Grudny J, Kołakowski J, Kruszewski M, et al. Association of genetic dependences between lung cancer and chronic obstructive pulmonary disease. *Pneumonol Alergol Pol*. 2013;81:308-18.
131. Brzóška K, Bartłomiejczyk T, Sochanowicz B, et al. Matrix metalloproteinase 3 polymorphisms as a potential marker of enhanced susceptibility to lung cancer in chronic obstructive pulmonary disease subjects. *Ann Agric Environ Med*. 2014;21:546-51.
132. Xu L, Bian W, Gu XH, Shen C. Genetic polymorphism in matrix metalloproteinase-9 and transforming growth factor-β1 and susceptibility to combined pulmonary fibrosis and emphysema in a Chinese population. *Kaohsiung J Med Sci*. 2017;33:124-9.
133. Zhou M, Huang SG, Wan HY, et al. Genetic polymorphism in matrix metalloproteinase-9 and the susceptibility to chronic obstructive pulmonary disease in han population of South China. *Chin Med J (Engl)*. 2004;117:1481-4.
134. Kukkonen MK, Tiili E, Vehmas T, et al. Association of genes of protease-antiprotease balance pathway to lung function and emphysema subtypes. *BMC Pulm Med*. 2013;13:36.
135. Bchir S, Nasr HB, Hakim IR, et al. Matrix metalloproteinase-9 (279R/Q) polymorphism is associated with clinical severity and airflow limitation in tunisian patients with chronic obstructive pulmonary disease. *Mol Diagn Ther*. 2015;19:375-87.
136. Dollery CM, McEwan JR, Henney AM, et al. Matrix metalloproteinases and cardiovascular disease. *Circ Res*. 1995;77:863-8.
137. Hunninghake GM, Cho MH, Tesfaigzi Y, et al. MMP12, lung function, and COPD in high-risk populations. *N Engl J Med*. 2009;361:2599-608.
138. Tacheva T, Dimov D, Aleksandrova E, et al. The G allele of MMP12-82 A>G promoter polymorphism as a protective factor for COPD in Bulgarian population. *Arch Physiol Biochem*. 2017;123:371-6.

139. Arja C, Ravuri RR, Pulamaghatta VN, et al. Genetic determinants of chronic obstructive pulmonary disease in South Indian male smokers. *PLoS One*. 2014;9:e89957.
140. Castaldi PJ, Cho MH, San José Estépar R, et al. Genome-wide association identifies regulatory loci associated with distinct local histogram emphysema patterns. *Am J Respir Crit Care Med*. 2014;190:399-409.
141. Trojanek JB, Cobos-Correa A, Diemer S, et al. Airway mucus obstruction triggers macrophage activation and matrix metalloproteinase 12-dependent emphysema. *Am J Respir Cell Mol Biol*. 2014;51:709-20.
142. Jackson VE, Ntalla I, Sayers I, et al. Exome-wide analysis of rare coding variation identifies novel associations with COPD and airflow limitation in MOCOS3, IFIT3 and SERPINA12. *Thorax*. 2016;71:501-9.
143. Zhang RB, He QY, Yang RH, Lu BB, Liu YJ. Study on matrix metalloproteinase 1, 9, 12 polymorphisms and susceptibility to chronic obstructive pulmonary disease among han nationality in Northern China. *Zhonghua Liu Xing Bing Xue Za Zhi*. 2005;26:907-10.
144. Seemungal TA, Donaldson GC, Paul EA, et al. Effect of exacerbation on quality of life in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 1998;157:1418-22.
145. Connors AF Jr., Dawson NV, Thomas C, et al. Outcomes following acute exacerbation of severe chronic obstructive lung disease. The SUPPORT investigators (Study to understand prognoses and preferences for outcomes and risks of treatments) *Am J Respir Crit Care Med*. 1996;154:959-67.
146. Andersson F, Borg S, Jansson SA, et al. The costs of exacerbations in chronic obstructive pulmonary disease (COPD). *Respir Med*. 2002;96:700-8.
147. Garcia-Aymerich J, Farrero E, Féliz MA, et al. Risk factors of readmission to hospital for a COPD exacerbation: a prospective study. *Thorax*. 2003;58:100-5.
148. Gudmundsson G, Gislason T, Janson C, et al. Risk factors for rehospitalisation in COPD: role of health status, anxiety and depression. *Eur Respir J*. 2005;26:414-9.
149. Recalde H, Cuccia M, Oggionni T, et al. Lymphocyte expression of human leukocyte antigen class II molecules in patients with chronic obstructive pulmonary disease. *Monaldi Arch Chest Dis*. 1999;54:384-9.
150. Medzhitov R, Janeway C Jr. Innate immunity. *N Engl J Med*. 2000;343:338-44.
151. Neth O, Jack DL, Dodds AW, et al. Mannose-binding lectin binds to a range of clinically relevant microorganisms and promotes complement deposition. *Infect Immun*. 2000;68:688-93.
152. Turner MW. Mannose-binding lectin (MBL) in health and disease. *Immunobiology*. 1998;199:327-39.
153. Yang IA, Seeney SL, Wolter JM, et al. Mannose-binding lectin gene polymorphism predicts hospital admissions for COPD infections. *Genes Immun*. 2003;4:269-74.
154. Lin CL, Siu LK, Lin JC, et al. Mannose-binding lectin gene polymorphism contributes to recurrence of infective exacerbation in patients with COPD. *Chest*. 2011;139:43-51.
155. Wright JR. Immunoregulatory functions of surfactant proteins. *Nat Rev Immunol*. 2005;5:58-68.
156. Borron PJ, Mostaghel EA, Doyle C, et al. Pulmonary surfactant proteins A and D directly suppress CD3+/CD4+ cell function: evidence for two shared mechanisms. *J Immunol*. 2002;169:5844-50.
157. Vandivier RW, Ogden CA, Fadok VA, et al. Role of surfactant proteins A, D, and C1q in the clearance of apoptotic cells *in vivo* and *in vitro*: calreticulin and CD91 as a common collectin receptor complex. *J Immunol*. 2002;169:3978-86.
158. Foreman MG, DeMeo DL, Hersh CP, et al. Polymorphic variation in surfactant protein B is associated with COPD exacerbations. *Eur Respir J*. 2008;32:938-44.
159. Liu YC, Yu MM, Chai YF, Shou ST. Sialic acids in the immune response during sepsis. *Front Immunol*. 2017;8:1601.
160. Angata T, Ishii T, Motegi T, et al. Loss of siglec-14 reduces the risk of chronic obstructive pulmonary disease exacerbation. *Cell Mol Life Sci*. 2013;70:3199-210.
161. Ishii T, Angata T, Wan ES, et al. Influence of SIGLEC9 polymorphisms on COPD phenotypes including exacerbation frequency. *Respirology*. 2017;22:684-90.
162. Emeryk-Maksymiuk J, Emeryk A, Krawczyk P, Wojas-Krawczyk K, Milanowski J. Beta-2-adrenoreceptor polymorphism at position 16 determines the clinical severity of chronic obstructive pulmonary disease. *Pulm Pharmacol Ther*. 2017;43:1-5.
163. Hussein MH, Sobhy KE, Sabry IM, El Serafi AT, Toraih EA. Beta2-adrenergic receptor gene haplotypes and bronchodilator response in Egyptian patients with chronic obstructive pulmonary disease. *Adv Med Sci*. 2017;62:193-201.
164. Rabe KF, Fabbri LM, Israel E, et al. Effect of ADRB2 polymorphisms on the efficacy of salmeterol and tiotropium in preventing COPD exacerbations: a prespecified substudy of the POET-COPD trial. *Lancet Respir Med*. 2014;2:44-53.
165. Mokry M, Joppa P, Slaba E, et al. Beta2-adrenergic receptor haplotype and bronchodilator response to salbutamol in patients with acute exacerbations of COPD. *Med Sci Monit*. 2008;14:CR392-8.
166. Bleecker ER, Meyers DA, Bailey WC, et al. ADRB2 polymorphisms and budesonide/formoterol responses in COPD. *Chest*. 2012;142:320-8.
167. Cherubini E, Esposito MC, Scozzi D, et al. Genetic polymorphism of CHRM2 in COPD: clinical significance and therapeutic implications. *J Cell Physiol*. 2016;231:1745-51.
168. Ishii T, Motegi T, Kamio K, Gemma A, Kida K. Association of group component genetic variations in COPD and COPD exacerbation in a Japanese population. *Respirology*. 2014;19:590-5.
169. Platé M, Lawson PJ, Hill MR, et al. Impact of a functional polymorphism in the PAR-1 gene promoter in COPD and COPD exacerbations. *Am J Physiol Lung Cell Mol Physiol*. 2014;307:L311-6.
170. Mlak R, Homa-Mlak I, Powrózek T, et al. Impact of I/D polymorphism of ACE gene on risk of development and course of chronic obstructive pulmonary disease. *Arch Med Sci*. 2016;12:279-87.
171. Zhang J, Peng S, Cheng H, et al. Genetic pleiotropy between nicotine dependence and respiratory outcomes. *Sci Rep*. 2017;7:16907.
172. Reynolds PR, Allison CH, Willnauer CP. TTF-1 regulates $\alpha 5$ nicotinic acetylcholine receptor (nAChR) subunits in proximal and distal lung epithelium. *Respir Res*. 2010;11:175.
173. Benowitz NL. Neurobiology of nicotine addiction: implications for smoking cessation treatment. *Am J Med*. 2008;121:S3-10.
174. Klinke ME, Jónsdóttir H. Smoking addiction in chronic obstructive pulmonary disease: integrating neurobiology and phenomenology through a review of the literature. *Chron Respir Dis*. 2014;11:229-36.
175. Lam DC, Luo SY, Fu KH, et al. Nicotinic acetylcholine receptor expression in human airway correlates with lung function. *Am J Physiol Lung Cell Mol Physiol*. 2016;310:L232-9.
176. Gwilt CR, Donnelly LE, Rogers DF. The non-neuronal cholinergic system in the airways: an unappreciated regulatory role in pulmonary inflammation? *Pharmacol Ther*. 2007;115:208-22.
177. Saccone NL, Culverhouse RC, Schwantes-An TH, et al. Multiple independent loci at chromosome 15q25.1 affect smoking quantity: a meta-analysis and comparison with lung cancer and COPD. *PLoS Genet*. 2010;6:e1001053.
178. Kaur-Knudsen D, Nordestgaard BG, Bojesen SE. CHRNA3 genotype, nicotine dependence, lung function and disease in the general population. *Eur Respir J*. 2012;40:1538-44.
179. Lambrechts D, Buysschaert I, Zanen P, et al. The 15q24/25 susceptibility variant for lung cancer and chronic obstructive pulmonary disease is associated with emphysema. *Am J Respir Crit Care Med*. 2010;181:486-93.
180. Keskitalo K, Broms U, Heliövaara M, et al. Association of serum cotinine level with a cluster of three nicotinic acetylcholine receptor genes (CHRNA3/CHRNA5/CHRNA4) on chromosome 15. *Hum Mol Genet*. 2009;18:4007-12.
181. Yang L, Qiu F, Lu X, et al. Functional polymorphisms of CHRNA3 predict risks of chronic obstructive pulmonary disease and lung cancer in chinese. *PLoS One*. 2012;7:e46071.
182. Korytina GF, Akhmadishina LZ, Viktorova EV, Kochetova OV, Viktorova TV. *IREB2*, *CHRNA5*, *CHRNA3*, *FAM13A* and hedgehog interacting protein genes polymorphisms and risk of chronic obstructive pulmonary disease in tatar population from Russia. *Indian J Med Res*. 2016;144:865-76.
183. Pérez-Rubio G, Falfán-Valencia R, Sánchez-Romero C, et al. Susceptibilidad genética a EPOC se modificó por la contribución ancestral (Amerindia-Europea) en mestizos mexicanos. *Neumol Cir Torax*. 2017;76:143.
184. Miller MR. Structural and physiological age-associated changes in aging lungs. *Semin Respir Crit Care Med*. 2010;31:521-7.
185. Campisi J. Aging, cellular senescence, and cancer. *Annu Rev Physiol*. 2013;75:685-705.
186. Ito K, Barnes PJ. COPD as a disease of accelerated lung aging. *Chest*. 2009;135:173-80.
187. Tsuji T, Aoshiba K, Nagai A. Alveolar cell senescence in patients with pulmonary emphysema. *Am J Respir Crit Care Med*. 2006;174:886-93.

188. Tsuji T, Aoshiba K, Nagai A. Alveolar cell senescence exacerbates pulmonary inflammation in patients with chronic obstructive pulmonary disease. *Respiration*. 2010;80:59-70.
189. Rajendrasozhan S, Yang SR, Kinnula VL, Rahman I. SIRT1, an antiinflammatory and antiaging protein, is decreased in lungs of patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2008;177:861-70.
190. Amsellem V, Gary-Bobo G, Marcos E, et al. Telomere dysfunction causes sustained inflammation in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2011;184:1358-66.
191. López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell*. 2013;153:1194-217.
192. Chilosi M, Carloni A, Rossi A, Poletti V. Premature lung aging and cellular senescence in the pathogenesis of idiopathic pulmonary fibrosis and COPD/emphysema. *Transl Res*. 2013;162:156-73.
193. Alder JK, Guo N, Kembou F, et al. Telomere length is a determinant of emphysema susceptibility. *Am J Respir Crit Care Med*. 2011;184:904-12.
194. Shen M, Cawthon R, Rothman N, et al. A prospective study of telomere length measured by monochrome multiplex quantitative PCR and risk of lung cancer. *Lung Cancer*. 2011;73:133-7.
195. Houben JM, Moonen HJ, van Schooten FJ, Hageman GJ. Telomere length assessment: biomarker of chronic oxidative stress? *Free Radic Biol Med*. 2008;44:235-46.
196. Morlá M, Busquets X, Pons J, Sauleda J, MacNee W, Agustí AG. Telomere shortening in smokers with and without COPD. *Eur Respir J*. 2006;27:525-8.
197. Savale L, Chaouat A, Bastuji-Garin S, et al. Shortened telomeres in circulating leukocytes of patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2009;179:566-71.
198. Mui TS, Man JM, McElhaney JE, et al. Telomere length and chronic obstructive pulmonary disease: evidence of accelerated aging. *J Am Geriatr Soc*. 2009;57:2372-4.
199. Houben JM, Mercken EM, Ketelslegers HB, et al. Telomere shortening in chronic obstructive pulmonary disease. *Respir Med*. 2009;103:230-6.
200. Lee J, Sandford A, Man P, Sin DD. Is the aging process accelerated in chronic obstructive pulmonary disease? *Curr Opin Pulm Med*. 2011;17:90-7.
201. Córdoba-Lanús E, Cazorla-Rivero S, Espinoza-Jiménez A, et al. Telomere shortening and accelerated aging in COPD: findings from the BODE cohort. *Respir Res*. 2017;18:59.
202. Rode L, Bojesen SE, Weischer M, Vestbo J, Nordestgaard BG. Short telomere length, lung function and chronic obstructive pulmonary disease in 46,396 individuals. *Thorax*. 2013;68:429-35.
203. Rutten EP, Gopal P, Wouters EF, et al. Various mechanistic pathways representing the aging process are altered in COPD. *Chest*. 2016;149:53-61.
204. Albrecht E, Sillanpää E, Karrasch S, et al. Telomere length in circulating leukocytes is associated with lung function and disease. *Eur Respir J*. 2014;43:983-92.
205. Kara M, Mirici A. Loneliness, depression, and social support of Turkish patients with chronic obstructive pulmonary disease and their spouses. *J Nurs Scholarsh*. 2004;36:331-6.
206. Dahlén I, Janson C. Anxiety and depression are related to the outcome of emergency treatment in patients with obstructive pulmonary disease. *Chest*. 2002;122:1633-7.
207. van Ede L, Yzermans CJ, Brouwer HJ. Prevalence of depression in patients with chronic obstructive pulmonary disease: a systematic review. *Thorax*. 1999;54:688-92.
208. Ji R, He Q, Zhang R, Gao Z, Ding D. A genetic study of the depressive respiratory responses to hypoxia in chronic obstructive pulmonary disease patients with type II respiratory failure. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi*. 2000;17:173-7.
209. Pietras T, Witusik A, Panek M, et al. Anxiety, depression and polymorphism of the gene encoding superoxide dismutase in patients with chronic obstructive pulmonary disease. *Pol Merkuri Lekarski*. 2010;29:165-8.
210. Ishii T, Wakabayashi R, Kurosaki H, Gemma A, Kida K. Association of serotonin transporter gene variation with smoking, chronic obstructive pulmonary disease, and its depressive symptoms. *J Hum Genet*. 2011;56:41-6.