

Role of Pharmacokinetics and Metabolism in Drug Discovery and Development

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I. Introduction

Drug research encompasses several diverse disciplines united by a common goal, namely the development of novel therapeutic agents. The search for new drugs can be divided functionally into two stages: discovery and development. The former consists of setting up a working hypothesis of the target enzyme or receptor for a particular disease, establishing suitable models (or surrogate markers) to test biological activities, and screening the new drug molecules for *in vitro* and/or *in vivo* biological activities. In the development stage, efforts are focused on evaluation of the toxicity and efficacy of new drug candidates. Recent surveys indicate that the average new chemical entity taken to market in the United States requires 10 to 15 years of research and costs more than \$300 million.

Once the target enzyme or receptor is identified, medicinal chemists use a variety of empirical and semiempirical structure-activity relationships to modify the chemical structure of a compound to maximize its *in vitro* activity. However, good *in vitro* activity cannot be extrapolated to good *in vivo* activity unless a drug has good bioavailability and a desirable duration of action. A growing awareness of the key roles that pharmacokinetics and drug metabolism play as determinants of *in vivo* drug action has led many drug companies to include examination of pharmacokinetics and drug metabolism properties as part of their screening processes in the selection of drug candidates. Consequently, industrial drug metabolism scientists have emerged from their traditional supportive role in drug development to provide valuable support in the drug discovery efforts.

To aid in a discovery program, accurate pharmacokinetic and metabolic data must be available almost as early as the results of the *in vitro* biological screening. Early pharmacokinetic and metabolic evaluation with rapid information feedback is crucial to obtain optimal pharmacokinetic and pharmacological properties. To be effective, the turnover rate needs to be at least three to five compounds per week for the support of each program. Due to time constraints and the availability of only small quantities of each compound in the discovery stage, studies are often limited to one or two animal species. Therefore, the selection of animal species and the experimental design of studies are important in providing a reliable prediction of drug absorption and elimination in humans. A good compound could be excluded on the basis of results from an inappropriate animal species or poor experimental design.

After a drug candidate is selected for further development, detailed information on the metabolic processes and pharmacokinetics of the new drug is required by regulatory agencies. The rationale for the regulatory requirement is best illustrated by the case of active metabolite formation. Many of the currently available psychotropic drugs form one or more metabolites that have their own biological activity (Baldessarini, 1990). Pharmacokinetically, the active metabolites may differ in distribution and clearance from that of the parent drug. Pharmacologically, the parent drug and its metabolites may act by similar mechanisms, different mechanisms, or even by antagonism. An understanding of the kinetics of active metabolite formation is important not only for predicting therapeutic outcome, but also for explaining the toxicity of specific drugs.

Conventionally, the metabolism of new drugs in humans is studied *in vivo* using radiotracer techniques as part of clinical absorption and disposition studies. However, this approach often occurs relatively late in the development stage. Ideally, the metabolism of new drugs should be studied *in vitro* before the initiation of clinical studies. Early information on *in vitro* metabolic processes in humans, such as the identification of the enzymes responsible for drug metabolism and sources of potential enzyme polymorphism, can be useful in the design of clinical studies, particularly those that examine drug-drug interactions. It is also desirable that the comparison of metabolism between animals and humans be performed in the early stage of the drug development process to provide information for the appropriate selection of animal species for toxicity studies before these toxicity studies begin.

The advance of *in vitro* enzyme systems used for drug metabolism studies (Wrighton and Stevens, 1992; Guilouzo et al., 1993; Berry et al., 1992; Rimmel and Burchell, 1993; Brendel et al., 1990; Chapman et al., 1993), together with the explosion of our knowledge of various drug-metabolizing enzymes including uridine-diphosphate-glucuronosyl-transferases (Cougletrie, 1992), cytochrome P-450s (Henderson and Wolf, 1992; Gonzalez and Nebert, 1990) and carboxylesterases (Wang, 1994; Hosokawa, 1990), allows us to obtain early information on the metabolic processes of new drug candidates well before the initial clinical studies. In addition, the advent of commercial liquid chromatography-mass spectrometry instrumentation and the development of high-field nuclear magnetic resonance as well as liquid chromatography-nuclear magnetic resonance techniques have fur-

ther strengthened our capability to study the metabolism of new drugs in the early drug discovery stage (Fenselau, 1992; Baillie and Davis, 1993). However, the role of drug metabolism scientists in drug discovery is more than just screening compounds *in vitro* and *in vivo*. It really entails a good understanding of the basic mechanisms of the events involved in absorption, distribution, metabolism and excretion; the interaction of chemicals with the drug-metabolizing enzymes, particularly cytochrome P-450; sources of pharmacokinetic and pharmacodynamic interindividual variability; and the consequences of metabolism on potential drug toxicities.

The purpose of this paper is to review the role of pharmacokinetics and drug metabolism in drug discovery and development from an industrial perspective. The intent is to provide a comprehensive, rather than exhaustive, overview of the pertinent literature in the field. Several excellent review articles on individual topics are available and the reader is referred to the most recent articles in the text. It is hoped that with a better understanding of the fate of the drugs, a balanced *in vitro/in vivo* approach and an intelligent application of sound principles in pharmacokinetics and enzymology, drug metabolism scientists can contribute significantly to the development of safe and more efficacious drugs.

II. Role of Pharmacokinetics and Metabolism in Drug Design

The history of the pharmaceutical industry shows that many important drugs have been discovered by a combination of fortuity and luck. This serendipity is best exemplified by the discovery of isoniazid. Isoniazid was first synthesized by Meyer and Mally (1912). Its antituberculosis properties were not found until 40 years later, when Robitzek et al. (1952) gave isoniazid to 92 "hopeless" patients with progressive caseous-pneumonic pulmonary tuberculosis that had failed to show improvement after any therapy. Furthermore, both indomethacin and ibuprofen compounds were developed as antirheumatic agents even without any knowledge of their mode of action (Shen, 1972; Adams et al., 1969, 1970). The mode of action was established several years after the drugs were on the market when Vane (1971) showed that these nonsteroidal anti-inflammatory drugs acted by inhibiting the synthesis of prostaglandins.

Another example of serendipity is the discovery of anxiolytics. Diazepam and chlordiazepoxide, the most widely used benzodiazepines, were found to have anxiolytic activity in 1958 and were marketed in 1960. Efforts to determine the mechanism of benzodiazepine action were initiated only after their introduction into the clinic. It was not until 1974 that convincing evidence from behavioral, electrophysiological, and biochemical experiments was accumulated to demonstrate that benzodiazepines act specifically at synapses in which γ -ami-

nobutyric acid (GABA)^b functions as a neurotransmitter (Baldessarini, 1990; Haefely et al., 1985; Williams and Olsen, 1989).

Over the past decades, through a better understanding of disease processes, mechanism-based drug design has evolved and produced drugs that interrupt specific biochemical pathways by targeting certain enzymes or receptors. This approach does not require a knowledge of the three-dimensional environment in which drugs act. Recent advances in molecular biology and protein chemistry have provided pure protein in sufficient quantities to allow structural studies to be carried out. Visualization of these structures by sophisticated computer graphics has made structure-based drug design feasible. These rational approaches of drug design have been successful historically in the fields of HIV protease inhibitors (Vacca et al., 1994), hepatic hydroxymethylglutaryl coenzyme A reductase inhibitors (Alberts et al., 1980) and angiotensin-converting enzyme (ACE) inhibitors (Patchett et al., 1980).

Today, the design of new drugs is still received by many medicinal chemists to mean maximization of the desired drug activity within certain structural limits. Sometimes, however, compounds that show very high activity *in vitro* may prove later to have no *in vivo* activity, or to be highly toxic in *in vivo* models. Lack of *in vivo* activity may be attributed to undesirable pharmacokinetic properties, and the toxicity may result from the formation of reactive metabolites. Therefore, rational drug design should also take both pharmacokinetic and metabolic information into consideration, and the information should be incorporated with molecular biochemical and pharmacological data to provide well-rounded drug design.

A. Metabolism and Drug Design

From toxicological and pharmacological points of view, it is desirable to design a "safer" drug that undergoes predictable metabolic inactivation or even under-

^b Abbreviations: 3-MC, 3-methylcholanthrene; 6-TGN, 6-thioguanine nucleotide; ACE, angiotensin-converting enzyme; AFB, aflatoxin B1; Ah, aromatic hydrocarbon; AUC, area under the curve; AZT, zidovudine; BBB, blood-brain barrier; CCK_B, cholecystokinin; cL, clearance; cL_H, hepatic clearance; cL_{int}, intrinsic clearance; CNS, central nervous system; CSF, cerebrospinal fluid; DMBA, 7,12-dimethylbenz[a]anthracene; DMBB, 5-(1,3-dimethylbutyl)-5-ethyl barbituric acid; EM, extensive metabolizer; f_p, fraction of unbound drug in plasma; f_t, free fraction in tissue; GABA, γ -aminobutyric acid; GSH, glutathione; K_i, dissociation constant of an inhibitor; K_{inact}, maximum inactivation rate constant; K_m, Michaelis constant; K_p, ratio of drug concentration in tissue to that in plasma after drug administration; L-dopa, levodopa; MPH, methylphenidate; NAT, N-acetyltransferase; NSAID, nonsteroidal anti-inflammatory agent; PEG, polyethylene glycol; PFDA, perfluorodecanoic acid; PM, poor metabolizers; PPAR, peroxisome proliferator-activated receptors; TPMT, thio methyltransferase; TPMT, thiopurine methyltransferase; UDPGT, uridine diphosphoglucose transferase; V_d, volume of distribution; V_i, velocity of an enzymic reaction in the presence of inhibitor extensive metabolizers; V_{max}, maximum velocity; V_o, velocity of an enzymic reaction in the absence of inhibitor.

goes no metabolism. Several approaches have been used for the design of safer drugs.

1. *Hard drugs.* The concept of nonmetabolizable drugs, or so-called hard drugs, was proposed by Ariëns (1972) and Ariëns and Simonis (1982). The hard drug design is quite attractive. Not only does it solve the problem of toxicity due to reactive intermediates or active metabolites, but the pharmacokinetics also are simplified because the drugs are excreted primarily through either the bile or kidney. If a drug is excreted mainly by the kidney, the differences in the elimination between animal species and humans will be dependent primarily on the renal function of the corresponding species giving highly predictable pharmacokinetic profiles using the allometric approach (Lin, 1995; Mordenti, 1985). A few successful examples of such hard drugs include bisphosphonates and certain ACE inhibitors.

Bisphosphonates are a unique class of drugs. As a class, they are characterized pharmacologically by their ability to inhibit bone resorption, whereas pharmacokinetically, they are classified by their similarity in absorption, distribution and elimination. In the clinic, these drugs are used in patients as antiosteolytic agents for the treatment of a broad range of bone disorders characterized by excessive bone resorption. These include hypercalcemia of malignancy, metastatic bone disease, Paget's disease, and osteoporosis.

The discovery of bisphosphonates was based on earlier studies of inorganic pyrophosphate by Fleisch and his coworkers (Fleisch et al., 1966, 1968, 1969; Fleisch and Russell, 1970). They found that pyrophosphate bound very strongly to calcium phosphate and inhibited not only the formation of calcium phosphate crystals, but also the crystal dissolution *in vitro*. However, pyrophosphate exhibited no effect on bone resorption *in vivo*. This was later explained by the observation that pyrophosphate is hydrolyzed before it reaches the site of bone resorption. These findings led to a search for analogs that would display the activities similar to pyrophosphate, but would also resist enzymatic hydrolysis. It was found that the bisphosphonates, characterized by a P-C-P bond rather than the P-O-P bond of pyrophosphate, fulfilled these criteria. As hard drugs, bisphosphonates are not metabolized in animals or humans, and the only route of elimination is renal excretion (Lin et al., 1991c; Lin, 1996a). In general, these compounds are very safe with no significant systemic toxicity (Fleisch, 1993).

Similarly, enalaprilat and lisinopril are considered hard drugs. These two ACE inhibitors undergo very limited metabolism and are exclusively excreted by the kidney (Ulm et al., 1982; Tocco et al., 1982; Lin et al., 1988). Unlike sulfhydryl-containing ACE inhibitors, such as captopril and its analogs, neither enalaprilat nor lisinopril exhibits significant side effects (Kelly and O'Malley, 1990). The most common side effects accompanying the clinical use of captopril are rashes and taste dysfunction (Atkinson and Robertson, 1979; Atkinson et

al., 1980). Similar side effects are observed with penicillamine, which is a sulfhydryl-containing heavy metal antagonist used extensively in the treatment of Wilson's disease (Levine, 1975; Suda et al., 1993). It is therefore speculated that captopril interacts with endogenous sulfhydryl-containing proteins to form disulfides that may act as haptens, resulting in immunological reactivity, which may be responsible for these side effects (Patchett et al., 1980). Enalaprilat and lisinopril were designed to avoid these undesirable side effects by removal of the sulfhydryl group (Patchett et al., 1980).

Due to their poor lipophilicity, the bisphosphonates, enalaprilat and lisinopril, are not metabolized *in vivo*. Ironically, the poor lipophilicity of these compounds results in poor oral absorption. For the bisphosphonate alendronate, the octanol/buffer partition coefficient is 0.0017 (Lin, 1996a). As a result of its poor lipophilicity, alendronate has very poor oral bioavailability in humans (<1%) (Lin, 1996a). To our knowledge, bisphosphonates are the only class of drugs being developed for oral dosage in spite of their poor bioavailability (Lin, 1996a). This is because the systemically available bisphosphonates are largely taken up by the target (bone) tissues, where their elimination is very slow (Lin, 1996a, 1992, 1993b). The half-life of alendronate in bone was estimated to be at least 10 years in humans.

Like bisphosphonates, both enalaprilat and lisinopril have low lipophilicity. The octanol-to-water partition coefficient is approximately 0.003 for both drugs (Ondetti, 1988). Interestingly, enalaprilat, a diacid compound with a net negative charge, is poorly absorbed (<10%), whereas lisinopril, a zwitterionic compound, has acceptable oral absorption (~30%) (Ulm et al., 1982; Tocco et al., 1982). Consequently, enalaprilat was developed as its ethyl ester prodrug (enalapril) to increase its bioavailability, whereas the prodrug approach was not employed for lisinopril.

Bisphosphonates and these two carboxyalkyldipeptide ACE inhibitors were not intentionally designed as hard drugs. The "hardness" came about only as a result of structural improvement. It so happens that the newer ACE inhibitors, such as benazepril, perindopril, and fosinopril, undergo significant metabolism (Kelly and O'Malley, 1990).

Although metabolically inert compounds are highly desirable candidates for drug design, the versatility of the drug-metabolizing enzymes presents quite a challenge to achieve this goal. For example, cytochrome P-450s are known to catalyze numerous oxidative reactions involving carbon, oxygen, nitrogen, and sulfur atoms in thousands of substrates with diverse structures. In addition, cytochrome P-450s are unique in that metabolic switchings can occur when the primary metabolic site of a compound is blocked. Thus, considering the broad substrate specificities and the versatilities of cytochrome P-450s and other drug-metabolizing enzymes,

designing drug candidates that are metabolically inert may not always be feasible.

2. *Soft drugs*. In contrast to the concept of hard drugs, Bodor (1984, 1982) and Bodor et al. (1980) have proposed the approach of soft drugs. A soft drug is pharmacologically active as such, and it undergoes a predictable and controllable metabolism to nontoxic and inactive metabolites. The main concept of soft drug design is to avoid oxidative metabolism as much as possible and to use hydrolytic enzymes to achieve predictable and controllable drug metabolism. Most oxidative reactions of drugs are mediated by hepatic cytochrome P-450 enzyme systems that are often affected by age, sex, disease, and environmental factors, resulting in complex biotransformation and pharmacokinetic variability (Hunt et al., 1992; Soons et al., 1992). In addition, P-450 oxidative reactions have the potential to form reactive intermediates and active metabolites that can mediate toxicity (Guengerich and Shimada, 1993). These undesirable effects attributed to oxidative metabolism may be circumvented to some extent by incorporating metabolic structural "softness."

Bodor and his colleagues (Bodor, 1984, 1982; Bodor et al., 1980) have designed soft quaternary-type drugs containing three structural components: an acidic group, an aldehyde, and a tertiary amine. Upon absorption, the soft quaternary drugs are hydrolyzed to three nontoxic components that are rapidly eliminated from the body.

Atracurium, a nondepolarizing muscle relaxant, can be considered a soft drug. This drug contains quaternary *N*-functions and ester groups. Atracurium is metabolized *in vivo* by two nonoxidative processes: a nonenzymatic metabolism by Hofmann-degradation to form a tertiary amine and an alkene, and hydrolysis of the ester groups by esterases (Mutschler and Derendorf, 1995; Hughes and Chapple, 1981).

Remifentanyl, a novel short-acting μ -opioid receptor agonist, may also be considered a soft drug. This drug is a methyl ester and is metabolized extensively by esterases to an inactive acid metabolite, GI-90291, of which over 90% is subsequently recovered in urine. To a much lesser extent, the drug also is metabolized by *N*-dealkylation to a second metabolite, GI-94219 (Feldman et al., 1991; Bürkle et al., 1996; Glass, 1995). The major metabolite GI-90291 is approximately 2000- to 4000-fold less potent compared with remifentanyl. Although both hard and soft drug designs are of academic interest, there are only a few successful examples in the drug market.

3. *Active metabolites*. For many years, the process of biotransformation was considered synonymous with the inactivation of pharmacologically active compounds. There is increasing evidence, however, that the metabolites of some drugs are pharmacologically active. Numerous examples of pharmacologically active metabolites being used as a source of new drug candidates exist

because these metabolites often are subject to phase II reactions and have better safety profiles.

Perhaps the best known example is acetaminophen, which is an *O*-deethylated metabolite of phenacetin. Acetaminophen shows superior analgesic activity when compared with phenacetin. The main advantage of acetaminophen over phenacetin is that it does not produce methemoglobinemia and hemolytic anemia (Flower et al., 1985). Phenacetin is converted to at least 1 dozen metabolites by *O*-deethylation, *N*-deacetylation, and hydroxylation processes. *N*-hydroxyphenatidine, a metabolite of phenacetin, has been shown to be responsible for the formation of methemoglobin and hemolysis of red blood cells (Jensen and Jollow, 1991). Conversely, acetaminophen primarily undergoes glucuronidation and sulfation exclusively and is quite safe clinically at the recommended dose. Similarly, the analgesic oxyphenbutazone is an active para-hydroxy metabolite of phenylbutazone. Similar to acetaminophen, this active metabolite also shows better analgesic activity than phenylbutazone and causes less gastric irritation (Flower et al., 1985).

Although pharmacologically active metabolites are generally formed by phase I oxidative reactions, phase II conjugation reactions also can produce biologically active metabolites. Morphine 6-glucuronide is more potent as a μ -opioid receptor agonist than morphine itself (Paul et al., 1989; Mulder, 1992). Recent clinical studies in cancer patients given morphine 6-glucuronide indicated that useful analgesic effects are achieved without the side effects of nausea and vomiting that are often associated with morphine (Osborne et al., 1992). These findings have led to the commercial marketing of morphine 6-glucuronide. Sulfation also produces biologically active metabolites. Minoxidil, a potent vasodilator, is a good example. Studies concerning the action of minoxidil revealed that the therapeutic activities were mediated by its sulfate conjugate (Bray and Quast, 1991).

In addition to the advantages that active metabolites may have in terms of efficacy with fewer unwanted side effects, active metabolites can also be preferred over the parent drugs for kinetic reasons. Many benzodiazepines form active metabolites with similar pharmacological properties. Oxazepam is the common active metabolite of chlordiazepoxide, halazepam, chlorazepate, and diazepam (Caccia and Garattini, 1990). Unlike other benzodiazepines, oxazepam undergoes only glucuronidation and has a shorter half-life than any of its precursors. This kinetic advantage has led to the marketing of oxazepam as a short-acting benzodiazepine in the treatment of sleeping disorders (Baldessarini, 1990).

B. Pharmacokinetics and Drug Design

Many of the failures of drug candidates in development programs are attributed to their undesirable pharmacokinetic properties, such as too long or too short $t_{1/2}$, poor absorption, and extensive first-pass metabolism. In

a survey, Prentis et al. (1988) reported that of 319 new drug candidates investigated in humans, 77 (40%) of the 198 candidates were withdrawn due to serious pharmacokinetic problems. This high failure rate illustrates the importance of pharmacokinetics in drug discovery and development.

To ensure the success of a drug's development, it is essential that a drug candidate has good bioavailability and a desirable $t_{1/2}$. Therefore, an accurate estimate of the pharmacokinetic data and a good understanding of the factors that affect the pharmacokinetics will guide drug design. This section includes a discussion of the chemically modifiable factors that influence drug absorption and disposition.

1. Absorption. Drug absorption is influenced by many biological and physicochemical factors. The two most important physicochemical factors that affect both the extent and the rate of absorption are lipophilicity and solubility (Leahy et al., 1989). The membrane of the gastrointestinal epithelial cells is composed of tightly packed phospholipids interspersed with proteins. Thus, the transcellular passage of drugs depends on their permeability characteristics to penetrate the lipid bilayer of the epithelial cell membrane, which is in turn dependent on the lipophilicity of the drugs. As in the example of bisphosphonates, drugs with poor lipophilicity will be poorly absorbed after oral administration (Lin, 1996a). The effect of lipophilicity on oral absorption is best exemplified by the classical study of barbiturates conducted by Schanker (1960). In this study, the absorption of these compounds increased with increasing lipophilicity as a result of increased membrane permeability. Similarly, Taylor et al. (1985) have shown that the absorption rates of a series of β -blockers in rat small intestine correlated well with their lipophilicity. However, it should be noted that although there is a correlation between lipophilicity and increased permeability, lipophilicity, in some cases, is not predictive of permeability because of external factors.

The oral bioavailability of a drug is defined as the fraction of an oral dose of the drug that reaches the systemic circulation. Because the entire blood supply of the upper gastrointestinal tract passes through the liver before reaching the systemic circulation, the drug may be metabolized by the liver and gut wall during the first passage of drug absorption. A drug with high metabolic clearance is always subject to an extensive first-pass effect, resulting in low bioavailability. The lipophilicity of a drug not only affects the membrane permeability, but the metabolic activity as well. In general, the higher the lipophilicity of a drug, the higher its permeability and the greater its metabolic clearance and thereby its first-pass metabolism (Seydel and Schaper, 1986; Toon and Rowland, 1983). The effects of lipophilicity on membrane permeability and first-pass metabolism appear to have opposing effects on the bioavailability. Thus, it is important to balance the effects of lipophilicity on mem-

brane permeability and first-pass metabolism to improve bioavailability. Also, it should be pointed out that there are many factors, in addition to lipophilicity, that can influence first-pass metabolism.

The influence of lipophilicity on the metabolic clearance of drugs is attributed mainly to the increased affinity of drugs for the enzymes. In vitro studies with rat liver microsomes by Martin and Hansch (1971) revealed that variations in maximum velocity (V_{max}) values for a series of compounds unrelated in chemical structure were small (only 3- to 5-fold), whereas the Michaelis constant (K_m) values varied by approximately 1000-fold. The K_m values were found to correlate significantly with their lipophilicity. The higher lipophilicity resulted in lower K_m values (higher enzyme affinities). Kinetic studies in dogs revealed that there was a positive correlation between metabolic clearance and lipophilicity for dihydropyridine calcium channel blockers in that the metabolic clearance increased with increasing lipophilicity (Humphrey, 1989).

The discovery of fluconazole (Richardson, 1993) is one of the examples of successfully applying the lipophilicity concept in drug design. Pfizer's initial efforts to find a novel antifungal agent resulted in tioconazole, which is clinically effective against fungal infections of the vagina and skin when administered topically. However, tioconazole shows poor efficacy when given intravenously or orally. Pharmacokinetic studies indicated that although this drug was absorbed reasonably well from the gastrointestinal lumen, it was subject to extensive first-pass metabolism, resulting in low oral bioavailability. In addition, the drug also was highly bound to plasma proteins, giving very low circulating levels of the unbound drug. Efforts were made to decrease the lipophilicity of this class of compounds to increase the metabolic stability and to decrease the protein binding. Efforts to decrease the lipophilicity included the replacement of the imidazole function with 12,4-triazole moiety to yield the bistriazole compound, UK-47,265. Although pharmacokinetic evaluation showed excellent absorption and kinetic profiles in several animal species after oral dosing, UK-47,265 exhibited hepatotoxicity in mice and dogs, which could be attributed to the 2,4-dichlorophenyl moiety. This finding led to the synthesis of a 2,4-difluorophenyl analog of UK-47,265, currently known as fluconazole. The introduction of fluorine into a molecule can alter both the metabolism and toxicity of a drug (Park and Kitteringham, 1994). In the case of fluconazole, the fluorine substitution was shown to reduce hepatotoxicity.

Solubility is also an important determinant in drug absorption; a drug must be reasonably soluble in the aqueous environment to be absorbed properly. The discovery of HIV protease inhibitors is an example that illustrates the concept of drug solubility. At Merck Research Laboratories (West Point, PA), starting from an initial peptide renin inhibitor (L-364,505), Vacca and his

coworkers (Vacca et al., 1994; Dorsey et al., 1994) successfully developed a novel hydroxyethylene dipeptide isostere series of highly potent and selective HIV protease inhibitors. However, like other HIV protease inhibitors that contain the hydroxyethylene or hydroxyethylamine transition state isosteres, the main drawback of Merck's initial inhibitors was that they were highly lipophilic and poorly soluble, resulting in poor bioavailability. Efforts were made to increase the solubility by incorporating a basic amine into the backbone of this series (table 1; fig. 1). The addition of pyridine to this series lead to the discovery of indinavir (MK-639, L-735,524). As shown in table 1, the aqueous solubility of indinavir is pH-dependent. The solubility of indinavir increased dramatically from 0.07 mg/mL at pH 7.4 to 60 mg/mL at pH 3.5 due to the protonation of the pyridine nitrogen ($\text{PK}_a = 3.7$). For this reason, indinavir sulfate is the clinical formulation, because it maintains the acidity of the gastrointestinal tract and dissolves more rapidly than free base. Indinavir sulfate is well absorbed after oral dosing and was approved recently for the treatment of AIDS.

A different approach to HIV protease inhibitor design was formulated by Abbott Laboratories (North Chicago, IL). The chemists successfully designed a series of C_2 symmetric inhibitors to match the C_2 symmetric active site of HIV protease. However, once again, the high lipophilicity and poor aqueous solubility limited these inhibitors for oral delivery. A77003, a C_2 symmetric inhibitor, was Abbott Laboratories' first HIV protease inhibitor to reach clinical trials for intravenous use (Kempf et al., 1991). Intravenous administration of A77003 was discontinued in phase I clinical trials because the large doses were required as a result of its short $t_{1/2}$ and the accompanying irritation and phlebitis at the injection site. A nonsymmetric analog, A80987, with improved aqueous solubility (pH 4, 122 $\mu\text{g}/\text{mL}$) had greater oral bioavailability and improved $t_{1/2}$ in animals (Kempf et al., 1995). Although A80987 gave reasonably good absorption in phase I clinical trials, the short $t_{1/2}$ limited the ability to maintain plasma levels in excess of the 95% effective concentration for viral replication. Intensive study of a series of A80987 analogs has yielded valuable insight into the relationship of chemical structure to antiviral activity, aqueous solubility, and hepatic metabolism. Application of these insights to drug design

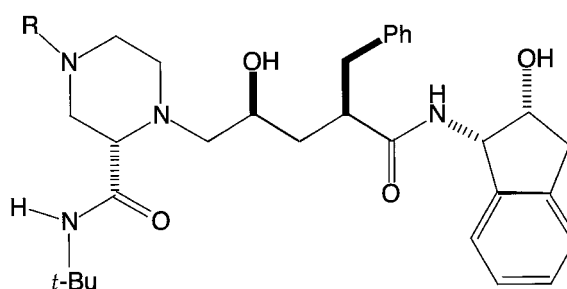


FIG. 1. Chemical structure of some HIV protease inhibitors. See table 1 for the substitutions (R). Reproduced with permission from Dorsey et al. (1994).

resulted in the discovery of ritonavir (Kempf et al., 1995).

In lieu of chemical modification, formulation approaches can be used sometimes to improve oral absorption of poorly soluble drugs. For further information, readers can reference a recent article reviewed by Aungst (1993), which discussed several formulation strategies of improving bioavailability of poorly soluble drugs. L-365,260 [a cholecystokinin (CCK_B) receptor antagonist] is a good example in which the formulation modification is applied. This drug has a very poor aqueous solubility of $<2 \mu\text{g}/\text{mL}$. When given orally as a suspension in 0.5% methylcellulose suspension, bioavailability was 14% for the rat and 9% for the dog (Lin et al., 1996c). The low bioavailability of L-365,260 was due mainly to its poor absorption as a result of its poor aqueous solubility, rather than extensive first-pass metabolism. In a separate study, by comparing the drug concentration in the systemic circulation during portal or femoral vein infusion of the drug, the hepatic first-pass metabolism was shown to be low, only 30% for the rat and 14% for the dog.

When L-365,260 was given orally as a solution in PEG 600 to rats and dogs, the bioavailability was increased 3- to 4-fold in rats and 8- to 9-fold in dogs (Lin et al., 1996c). With this information at hand, L-365,260 was dosed in capsules containing PEG 600 in the subsequent clinical studies. This formulation also gave good absorption of L-365,260 in humans. Although the underlying mechanism for the improved absorption is unknown, PEG 600 may have exerted a cosolubilizing effect to maintain a higher drug concentration in solution in the gastrointestinal tract.

TABLE 1
Effect of solubility of HIV protease inhibitors on drug absorption in dogs after a 5-mg/kg p.o. dose

| Compound | R | C_{max} (μM) ^a | Solubility (mg/ml) at pH 7.4 (or 3.5) | log P |
|-----------|--------------------------|--|---------------------------------------|-------|
| L-732,747 | benzyloxycarbonyl | <0.10 | <0.001 | 4.67 |
| L-735,482 | 8-quinolinylsulfonyl | <0.10 | <0.001 | 3.70 |
| L-738,891 | 2,4-difluorophenylmethyl | 0.73 ± 0.15 | 0.0012 | 3.69 |
| L-735,524 | 3-pyridylmethyl | 11.4 ± 2.3 | 0.07 (60) | 2.92 |

^a C_{max} , maximum drug concentration in plasma. See figure 1. Data excerpted from Dorsey et al. (1994).

2. *Prodrugs*. The prodrug concept was first proposed by Albert (1958). Since then, this approach has been widely used in drug design. Although there are many reasons to use prodrugs, improvement of oral absorption is by far the most common. Antibiotic prodrugs comprise the largest group of prodrugs developed to improve oral absorption (Wermuth, 1984). Pivampicillin, talampicillin, and bacampicillin are prodrugs of ampicillin, all resulting from the esterification of the polar carboxylate group to form lipophilic, enzymatically labile esters. The absorption of these prodrugs is nearly complete (98–99%), whereas that of ampicillin is <50% (Loo et al., 1974; Clayton et al., 1974; Bodin et al., 1975).

Enalapril, the most widely prescribed ACE inhibitor, is the ethyl ester prodrug of the active diacid, enalaprilat. Enalaprilat is poorly absorbed from the gastrointestinal tract (<10%), but absorption of the prodrug enalapril is greatly improved (60%). Hepatic metabolic hydrolysis is responsible for its conversion to the active diacid (Ulm et al., 1982; Tocco et al., 1982).

In addition to the simple approaches of ester and amide prodrug formation, more sophisticated manipulation of chemical entities can be used. For example, acyclovir, a potent antiherpes drug, is poorly and erratically absorbed after oral dosing due to its polarity. Although acyclovir possesses a derivatizable hydroxyl group in its structure, esterification of this hydroxy group did not improve the absorption. However, desoxyacyclovir (fig. 2), a prodrug of acyclovir that is activated by xanthine oxidase present in both the gut and liver, gives superior oral delivery of acyclovir over that of the parent drug or its esters (Rees et al., 1986; Krasny and Petty, 1987; Krenitsky et al., 1984). In vivo, phosphorylation of acyclovir is essential for antiviral activity. In normal mammalian cells, phosphorylation of this drug is extremely low, but in cells infected with the herpes simplex virus, there is an induction of a virus-coded thymidine kinase, which effectively catalyzes its phosphorylation (fig. 2). Thus, acyclovir is preferentially activated in virus-infected cells (Krenitsky and Elion, 1982). The example of acyclovir illustrates the point that medicinal chemists can use metabolic and kinetic information to design a better drug.

Sulindac (MK-231), a nonsteroidal anti-inflammatory agent (NSAID) is another interesting example in which

medicinal chemists applied metabolic and kinetic principles to design a prodrug. Sulindac is an indene analog of indomethacin. Replacement of the indole nucleus with indene reduces central nervous system (CNS) activity, and the addition of fluoro affords increased analgesic potency. Furthermore, the introduction of a methylsulfinyl (sulfoxide) group not only increases the aqueous solubility but also provides a center for metabolism in vivo (Shen and Winter, 1977). Sulindac, per se, is pharmacologically inactive; it is reversibly reduced to the sulfide metabolite, which is as potent as indomethacin. In vitro study with leukocytes showed a marked difference in the partition and permeation of sulindac and its active sulfide metabolite. The more hydrophilic sulindac tends to remain extracellular, whereas the more lipophilic sulfide accumulates inside the cell with cell/medium ratio of 50:1 (Duggan, 1981). The differential tissue distribution of sulindac and sulfide contributes to its patient tolerance, as well. Although most NSAIDs produce gastrointestinal lesions that are related to local depletion of prostaglandins, the gastrointestinal irritation is reduced by the oral administration of its inactive prodrug (sulindac) and the lack of enterohepatic recirculation of the active sulfide metabolite.

Another promising area for prodrugs is their application to site-specific drug delivery (Stella, 1989; Stella and Himmelstein, 1980). γ -Glutamyl dopa is an example of a site-specific prodrug of levodopa (L-dopa) (Wilk et al., 1978). L-dopa is a precursor of the neurotransmitter dopamine, which plays an important role in the CNS. Aside from its action as a neurotransmitter, dopamine also exerts receptor-mediated vasodilation in the kidney. Intraperitoneal injection of γ -glutamyl dopa into mice led to the selective generation of dopamine in the kidney as a consequence of the sequential actions of γ -glutamyl transpeptidase and L-aromatic amino acid decarboxylase, two enzymes that are highly concentrated in the kidney. The concentration of dopamine in the kidney after γ -glutamyl dopa administration was five times higher than that after administration of an equivalent dose of L-dopa (Wilk et al., 1978). Infusion of 10 nmol·g⁻¹·30 min of γ -glutamyl dopa to rats produced a 60% increase in renal plasma flow, whereas the same dose of L-dopa had no effect on renal plasma flow. The selective properties of γ -glutamyl dopa suggest that this

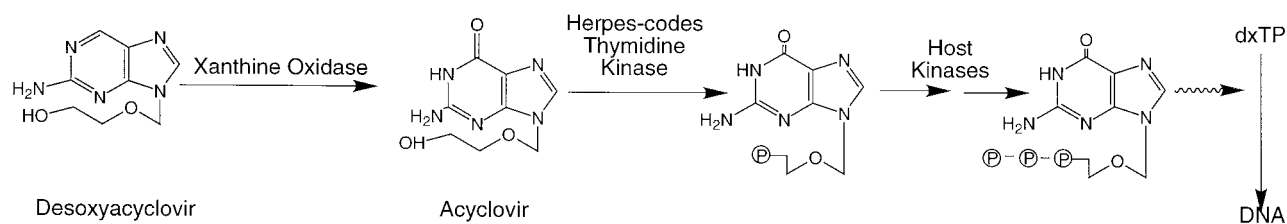


FIG. 2. Desoxyacyclovir, a prodrug of acyclovir that is activated by xanthine oxidase. The ultimate target enzyme of acyclovir is the viral DNA polymerase, which is inhibited by the triphosphate metabolite of acyclovir. Reproduced with permission from Krenitsky and Elion (1982).

prodrug would be beneficial in cases of impaired renal blood flow.

3. *Distribution.* The lipophilicity of a drug not only affects its absorption and metabolism but also its binding and distribution. Generally, the higher the lipophilicity of a drug, the stronger its binding to protein and the greater its distribution (Seydel and Schaper, 1986; Toon and Rowland, 1983). In studies with structure-related sulfonamides, Seydel et al. (1973) have shown that there was a strong positive correlation between plasma protein binding of the drugs and their lipophilicity. Watanabe and Kozaki (1978) found that the volume of distribution increased with increasing lipophilicity when administering 15 basic drugs to dogs.

Studies in the 1950s on the distribution of thiopental and polychlorinated insecticides (e.g., dichlorodiphenyltrichloroethane) have led to the misconception that highly lipophilic drugs tend to accumulate in adipose tissue. Recent studies by Bickel (1994) have shown that although the initial uptake of drugs into adipose tissue is related to their lipophilicity, the degree of adipose tissue storage does not correlate with their lipophilicity. Factors such as drug binding to plasma and tissue proteins also play a significant role in drug storage in adipose tissues.

The brain is different from other organs in several aspects. One of the most important features is that the brain is completely separated from the blood by the blood-brain barrier (BBB). All organs are perfused by capillaries lined with endothelial cells that have small pores to allow for the rapid movement of drugs into the organ interstitial fluid from the circulation. However, the capillary endothelium of the brain lacks these pores and, therefore, drugs must cross the BBB and enter the brain by simple diffusion. To design drugs for CNS activity, it is important to understand the factors that affect drug delivery to the site of action.

Because most drugs cross the BBB by passive diffusion, lipophilicity is an important determinant of brain penetration. Many reports show a correlation between lipophilicity and brain penetration of drugs (Pardridge, 1980; Rapoport, 1976). Ochs et al. (1985) found that the rate of brain uptake of drugs was dependent on their lipophilicity. There was a strong negative correlation between lipophilicity and the time of peak concentration in cerebrospinal fluid (CSF) postdose. The calculated lipophilicities (log D) of salicylic acid, antipyrine, and amitriptyline were -0.9 , 0.4 , and 3.0 , respectively, and the time required to reach the peak CSF concentration after intravenous administration to dogs was 200, 34, and 4 minutes, respectively (Ochs et al., 1985).

Although lipophilicity is an important factor affecting brain penetration, a linear relationship between lipophilicity and brain penetration can only be expected within a certain range. In a recent survey of 257 marketed drugs (fig. 3), the optimum log P value of lipophilicity was between 1 and 2 for the overall beneficial behavior

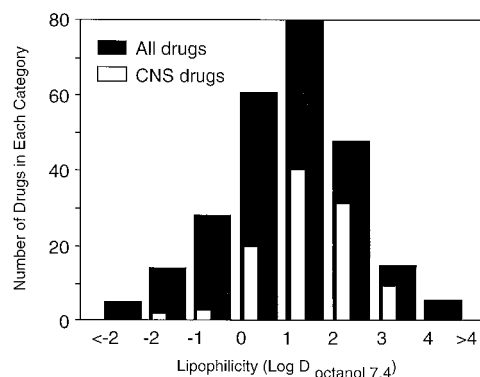


FIG. 3. Distribution of drugs with respect to their lipophilicity. Results of a survey of 257 marketed drugs based on available literature information. Reproduced with permission from Taylor & Francis (Jezequel, 1992).

of CNS drugs (Jezequel, 1992). Drugs with extremely high lipophilicity can be as poorly taken up by the brain as those with low lipophilicity. For example, L-365,260, a potent CCK_B receptor antagonist for the treatment of anxiety, is a very lipophilic drug with a log P value of 3.6. However, this drug displays poor BBB penetration (Lin and Lin, 1990).

P-glycoprotein, located on the apical surface of the endothelial cells of the brain capillaries toward the vascular lumen (Tew et al., 1993; Pardridge, 1991), is believed to be responsible for the poor BBB penetration of some highly lipophilic drugs. The poor BBB penetration of L-365,260 may be related to the efflux function of p-glycoprotein. Pretreatment of rats with quinine or verapamil, potent inhibitors of p-glycoprotein, resulted in a substantial increase in BBB penetration of L-365,260 by 3- to 5-fold (Lin et al., unpublished data). These results were consistent with the role of p-glycoprotein in excluding xenobiotics.

Factors other than lipophilicity also may play an important role in the transfer of drugs across the BBB. A strong negative correlation was found between the BBB permeability of steroid hormones and the total number of hydrogen bonds; the greater the hydrogen bond, the lower the permeability (Pardridge, 1980). Similarly, the hydrogen bond potential is a determinant of in vitro and in situ BBB permeability of peptides (Chikhale et al., 1994). It was concluded that a major impediment to BBB penetration of peptides was the energy required to break the water-peptide hydrogen bond.

L-663,581 is an investigational partial agonist of benzodiazepine receptors for potential application in the treatment of anxiety. Studies in rats, dogs, and monkeys have shown that the drug is eliminated mainly by biotransformation. Two metabolites, mono- and bis-hydroxy analogs, were demonstrated to be active in vitro. The potency of benzodiazepine receptor binding (K_i) is 3.7 nM for the parent drug, 3.3 nM for the mono-hydroxylated metabolite, and 1.2 nM for the bis-hydroxylated metabolite. Although the metabolites are as potent as, or

more potent than, the parent drug in vitro, they are inactive in rats in a conditioned emotional response model (Lin et al., 1994). The lack of in vivo activity of the metabolites cannot be explained by absorption and/or elimination kinetics. Brain uptake studies indicated that permeability of the BBB is high for L-663,581 but very poor for the metabolites (Lin et al., 1994). Because the metabolites have a reasonably good octanol/buffer partition coefficient ($\log P$ ranging from 0.7 to 1.2), and because two clinically used benzodiazepines, alprazolam and clobazam, have similar partition coefficients compared to the mono- and bis-hydroxylated metabolites (Arendt et al., 1983; Greenblatt et al., 1983), the poor penetration of the BBB of the metabolites may be due to their hydrogen bonding, rather than lipophilicity. According to Stein's assignment (1967), the mono-hydroxylated metabolite has two more hydrogen bonds, and the bis-hydrogenated analog has four more hydrogen bonds.

4. Plasma half-life. Most drugs are administered as a fixed dose given at regular intervals to achieve therapeutic objectives. Generally, the duration of drug action is reflected by its plasma $t_{1/2}$. Thus, the $t_{1/2}$ of drugs in plasma is one of the most important factors that determines the selection of a dosage regimen. Administration of drugs with a short $t_{1/2}$ requires frequent dosing and often results in a significant decrease in patient compliance. Because the $t_{1/2}$ of a drug is determined by its volume of distribution and elimination clearance, the prolongation of $t_{1/2}$ can be achieved by increasing the volume of distribution or decreasing the clearance. It appears to be easier to modify the chemical structure to slow a drug's clearance than to increase its volume of distribution.

Nifedipine, a calcium channel blocker widely used for the treatment of hypertension, has a short plasma $t_{1/2}$ (~2 h), resulting in a t.i.d. dosage regimen. Nifedipine also undergoes substantial first-pass metabolism and exhibits large interindividual variation in systemic concentrations (Kleinbloesem et al., 1984); these pharmacokinetic properties are not ideal for the chronic treatment of hypertension. Thus, a search was initiated for a backup drug with good oral bioavailability and duration of action that would allow a once-a-day dosage regimen.

The addition of an alkyl amide side-chain linked to the dihydropyridine 2-methyl group yielded amlodipine with a lower clearance, which has an improved oral bioavailability and plasma $t_{1/2}$ without loss of antihypertensive activity (Arrowsmith et al., 1986). Based on pharmacokinetic studies in dogs, amlodipine was chosen as a backup compound to meet the objectives of the program. Clinical studies proved that indeed amlodipine had good oral bioavailability (50–90%) and a prolonged plasma $t_{1/2}$ (30 h) (Humphrey, 1989).

Although chemical modification is preferred due to its ease, prolongation of the $t_{1/2}$ and a decrease in dosage frequency can be achieved by developing sustained-release dosage forms or coadministering inhibitors of

drug-metabolizing enzymes. Metoprolol, a β -blocker, has a relatively short $t_{1/2}$ (<3 h), so a once-a-day sustained-release tablet was developed. This sustained-release dosage form produced a more prolonged and uniform effect on the heart rate and systolic blood pressure than when given as a conventional tablet twice a day (Johnsson et al., 1980).

Sinemet and primaxin are examples of coadministration of enzyme inhibitors to prolong the duration of drug action. Sinemet (Merck Research Laboratories, West Point, PA), a combination product of L-dopa and carbidopa, is widely used for the treatment of Parkinson's disease. When L-dopa is given alone, >90% of the dose is decarboxylated peripherally and only 10% is available for CNS activity. To minimize the decarboxylation of L-dopa outside the CNS, carbidopa, a peripherally active decarboxylase inhibitor that cannot cross the BBB, is coadministered (Marsden, 1976). Primaxin (Merck Research Laboratories, West Point, PA), a combination of imipenem and cilastatin, is a widely used β -lactam antibiotic. Imipenem (MK-787) possesses a broad spectrum of action that comprises most of the gram-positive and gram-negative bacteria. In vivo imipenem is inactivated rapidly by a renal dipeptidase. This inactivation can be slowed by the combination of imipenem with the renal dipeptidase inhibitor, cilastatin (Kropp et al., 1980).

Although it is generally true that the duration of drug action is reflected by its plasma $t_{1/2}$, some drugs are given less frequently than their $t_{1/2}$. Despite its very short plasma $t_{1/2}$ in humans (≤ 1 h), omeprazole, a proton-pump inhibitor, is given once a day (Regårdh et al., 1985). This drug reduces gastric acid secretion through inhibition of the enzyme H^+, K^+ -ATPase located in the secretory canals of the parietal cells. Omeprazole is a weak base ($pK_a = 4.0$) and is rapidly and well absorbed from the alkaline environment of the small intestine. After entry of omeprazole into the parietal cells, the drug is converted to an intermediate (spiro compound) by protonation. The spiro intermediate subsequently forms the active metabolite, cyclic sulfenamide, which binds irreversibly to the enzyme H^+, K^+ -ATPase (Mutschler and Derendorf, 1995). Because formation of the spiro intermediate occurs only in an acidic medium, omeprazole accumulates pH-dependently in the parietal cells and inhibits acid secretion for a long duration.

5. Stereoselectivity. Although it has been long known that stereoisomers of a chiral drug often exhibit pronounced differences in their pharmacokinetic and pharmacodynamic properties both in quantitative and qualitative terms, more than 500 drugs are marketed currently as racemic mixtures without relevant pharmacokinetic and pharmacodynamic information for each individual stereoisomer. This neglect of stereochemistry in drug development was widespread until Ariëns' (1984) famous critical review of "sophisticated scientific nonsense" was published. It was Ariëns' review that

finally incited drug researchers to consider the importance of stereoselective differences.

MK-927, Merck's first carbonic anhydrase inhibitor to reach clinical trials for the treatment of glaucoma, is a racemic mixture. In 1986, due to the complexity of its stereoselective pharmacokinetic and pharmacodynamic properties (Lin et al., 1991b,d) and in consideration of Ariens' criticism, the development of MK-927 was terminated and replaced by its more active *S*-isomer, MK-417 (Lin et al., 1990a, 1991a). Subsequently, it was decided to develop dorzolamide (MK-507), which is the *S*-isomer of an MK-927 analog, because of its longer duration of action. Dorzolamide is now on the market for the treatment of ocular hypertension or open-angle glaucoma (Pfeiffer, 1994).

Although in principle it is preferable to synthesize and develop the more active single enantiomer, there are situations in which use of the racemate is justified based on pharmacodynamic or pharmacokinetic information, such as interconversion of stereoisomers or chiral inversion. Recently, Baillie and his coworkers (Zhang et al., 1994; Tang et al., 1994) have demonstrated an acid-catalyzed racemization of stiripentol in rats that takes place at low pH. After oral administration of either the *S*- or *R*-enantiomer, racemization of the drug in the stomach leads to a mixture of the *S*- and *R*-enantiomers before entering the gastrointestinal tract. Because both enantiomers are pharmacologically active (Shen et al., 1992), and because racemization occurs in the stomach, stiripentol is currently being developed as the racemic mixture. Similarly, the clinical use of racemic ibuprofen is justified by evidence of the unidirectional chiral inversion of the inactive *R*(-)-ibuprofen to its active *S*(+)-ibuprofen in vivo (Adams et al., 1976; Lee et al., 1985).

In some cases, enantiomers are purposely combined to optimize their therapeutic profiles. Indacrinone (MK-286) is a 9:1 mixture of the (+)- and (-)-isomers designed to optimize its uricosuric and diuretic activities (Blaine et al., 1982; Tobert et al., 1981). Both isomers are potent uricosuric agents, but the (-)-enantiomer is the more potent diuretic.

Sometimes, the chiral preference of subtypes of certain receptors in specific tissues can provide a basis for novel drug development. Two distinct subtypes of β -adrenergic receptors have been identified and characterized. In cardiac and pulmonary tissues, the β -adrenoceptors are predominantly of the β_1 -subtype, whereas in ocular tissues, they are mainly the β_2 -subtype (Weiner and Taylor, 1985). Timolol (MK-950), a nonselective β -adrenergic antagonist, contains a chiral center in the amino-hydroxypropoxy side-chain. Although both enantiomers inhibit adrenergic activity at β_1 - and β_2 -receptors, the ratio of *R*:*S* stereoselectivity is substantially greater for the β_1 -receptors than for the β_2 -receptors. The *R*:*S* ratios of the in vitro β -blockade by timolol in the guinea pig trachea and atria were approximately 1:80 to 1:90, whereas the *R*:*S* ratio of the aqueous humor reduc-

tion by timolol in the rabbit eye was approximately 1:3 (Share et al., 1984). Thus, timolol was prepared as the optically pure *S*-form for the treatment of hypertension (Keates and Stone, 1984), and the *R*-enantiomer was developed as a topical ocular hypotensive agent in the treatment of glaucoma to circumvent the unwanted cardiac and pulmonary effects (Weiner, 1985). The above examples illustrate that stereoselectivity can be used in novel drug design.

Despite the advances of molecular biology and protein chemistry, drug design is still not a precise science and usually requires an iterative process of reassessing structural changes to obtain optimal pharmacological and pharmacokinetic properties. The examples cited in this section are used to illustrate the principle that both pharmacokinetic and metabolic data can provide important information to guide drug design.

III. Role of Metabolism in Drug Toxicity

As part of drug development, the safety of a drug candidate has to be evaluated carefully before it can be approved. Due to ethical constraints on performing toxicity studies in humans, relevant safety assessments must be extensively studied in laboratory animals. One of the fundamental challenges drug metabolism scientists face in drug discovery and development is the extrapolation of risk assessment from animals to humans. This extrapolation is far from straightforward. As seen in the marked species differences in metabolism (Lin, 1995; Clark and Smith, 1984), drug-induced toxicity is often species-dependent, both in quantitative and qualitative terms. Some species of experimental animals have such unique mechanisms of developing toxicity that extrapolation of such toxicity assessments to the human situation would be fraudulent (Gregory, 1988; Green, 1991; Boobis, et al., 1990; Park and Kitteringham, 1990). Although there is no single method or model that can extrapolate the toxicity from animals to humans (Boxenbaum et al., 1988), the species differences in toxicity often can be explained by pharmacokinetic or pharmacodynamic effects of drugs. To make an accurate interpretation and a reasonable prediction of potential toxicity in humans, it is important to elucidate the underlying mechanisms responsible for the species differences in metabolism and pharmacokinetics.

A. Species Differences in Metabolism

From an evolutionary standpoint, all mammals are similar because they originate from a common ancestor, yet they have differentiated as a result of their dissimilar environmental adaptation. Biochemistry provides countless examples of similarities and differences between species, of which one of the most instructive is the structure of cytochrome P-450s. Cytochrome P-450s appear to have evolved from a single ancestral gene over a period of 1.36 billion years. To date, at least 14 P-450 gene families have been identified in mammals (Nelson

et al., 1996). Although all members of this superfamily possess highly conserved regions of amino acid sequence, there are considerable variations in the primary sequences across species. Table 2 shows the homology of nucleotide and amino acid sequences between humans and animal species (Kamatani, 1995). Even small changes in amino acid sequences can give rise to profound differences in substrate specificity (Lindberg and Negishi, 1989).

Similar to cytochrome P-450s, uridine diphosphoglucose transferases (UDPGTs) and carboxylesterases also show species similarities and differences. At least 10 rat UDPGTs and 8 human UDPGTs have been defined and characterized to date by cDNA cloning (Clarke and Burchell, 1994). Comparison of the amino acid sequences of all UDPGTs indicates that they share a common C-terminal domain, but the N-terminal half of these isoforms is quite variable. Examination of each of the UDPGT isoforms has revealed that their substrate specificities are different, although they still have overlapping substrate specificities.

Carboxylesterases, enzymes that are widely distributed in the tissues of mammals, hydrolyze drugs containing ester bonds or amide linkages and play an important role in drug metabolism, particularly for ester prodrugs. Hepatic microsomal carboxylesterases exist as multiple isozymes, and there are significant species differences in the activities of the carboxylesterase (Sato, 1987; Hosokawa et al., 1987). Hosokawa et al. (1990) have compared the amino acid sequences and substrate specificity of purified carboxylesterase from liver microsomes of mice, hamsters, rats, guinea pig, rabbits, and monkeys. Although high (80–95%) homology in amino acid sequences was shown, all carboxylesterases had a different N-terminal amino acid, and their substrate specificities were considerably different.

As a result of the species differences in the amino acid sequences of the isozymes, both the rate of drug metab-

olism and the metabolite pattern may differ between animal species. Similarly, the response of the enzymes to inducers, inhibitors, and hormones may vary between species due to their enzyme structural differences. This section will discuss the factors with respect to the species differences in drug metabolism.

1. *Oxidation and conjugation.* Indinavir (MK-639, L-735, 524), a potent HIV protease inhibitor, is subject to extensive metabolism in animals and humans. The major metabolic pathways of indinavir in humans are identified as (a) glucuronidation at the pyridine nitrogen to yield a quaternized ammonium conjugate, (b) pyridine N-oxidation, (c) para-hydroxylation of the phenylmethyl group, (d) 3'-hydroxylation of the indan, and (e) N-depyridomethylation (Chiba et al., 1996). All oxidative metabolites observed in humans also were formed in rats, dogs, and monkeys, whereas N-glucuronide was found only in monkey and human urine (Lin et al., 1996a). An additional metabolite, a cis-2'-3'-dihydroxyindan, was formed in monkeys, but not in other species. The intrinsic clearance (cL_{int}) (V_{max}/K_m) of the oxidative metabolism of indinavir was in the rank order: rat (157 mL/min/kg) \approx monkey (162 mL/min/kg) > dog (29 mL/min/kg) > human (17 mL/min/kg) (Lin et al., 1996a). Clearly, indinavir metabolism is qualitatively and quantitatively different among species.

The in vitro metabolism of losartan (MK-954; Dup 753), a potent nonpeptide angiotensin II receptor antagonist, has been studied with liver slices from rats, monkeys, and humans (Stearns et al., 1992). Metabolism of losartan also is qualitatively and quantitatively different among species. In the rat, the primary route of metabolism is oxidative, which leads to either monohydroxylated or oxidized (carboxylic acid) metabolites. In monkeys, glucuronidation of the tetrazole moiety predominates. The metabolism of losartan by human liver slices, however, is not dominated by a single metabolic pathway, as with rats and monkeys but is characterized

TABLE 2
Cytochrome P-450s: Homology of the nucleotide and deduced amino acid sequences between human and various animal species

| Isozyme | Human | Crab-eating monkey | Marmoset | Dog | Rabbit | Hamster | Rat | Mouse |
|---------|-------|--------------------|----------|-----|--------|---------|-----|-------|
| 1A1 | 100 | 95 | — | 84 | 81 | 79 | 81 | 82 |
| | 100 | 94 | — | 82 | 77 | 77 | 79 | 80 |
| 1A2 | 100 | — | 89 | 84 | 83 | 80 | 80 | 79 |
| | 100 | — | 86 | 82 | 78 | 73 | 75 | 73 |
| 2C | 100 | 96 | 89 | 73 | 81 | 79 | 79 | — |
| | 100 | 92 | 79 | 68 | 76 | 80 | 74 | — |
| 2D | 100 | — | 90 | — | — | 76 | 79 | 74 |
| | 100 | — | 90 | — | — | 73 | 78 | 70 |
| 2E1 | 100 | 96 | 93 | — | 81 | 80 | 80 | 79 |
| | 100 | 94 | 89 | — | 79 | 78 | 80 | 77 |
| 3A | 100 | 96 | 92 | 82 | 80 | 74 | 77 | 81 |
| | 100 | 93 | 87 | 77 | 74 | 68 | 71 | 76 |

Upper value, nucleotide sequence (% homology); lower value, deduced amino acid sequence (% homology); —, not known. In the 2C, 2D, and 3A gene subfamilies, the values indicate percentage of pairs that showed the highest identity among the multigene family. Data excerpted from Kamatani (1995).

by an approximately equal formation of both oxidized and glucuronidated metabolites. The investigators of this study suggest that the observed short duration of action of the drug in monkeys may be due to the low formation rate of the pharmacologically active carboxylic acid metabolite in this species. This carboxylic acid metabolite has a much longer $t_{1/2}$ than the parent drug in all species studied.

Stevens et al. (1993) recently compared phase I and phase II hepatic drug metabolism activities using human and monkey liver microsomes. Of the eight P-450-dependent activities measured, only *N*-nitrosodimethylamine *N*-demethylase activity was not significantly different in the two species. Coumarin 7-hydroxylase activity was higher in the humans than in the monkey. In contrast, erythromycin *N*-demethylase, benzphetamine *N*-demethylase, pentoxyresorufin *O*-dealkylase, ethoxycoumarin *O*-deethylase, and ethoxyresorufin *O*-deethylase activities were significantly greater in monkey microsomes than those from humans. Of the seven microsomal and cytosolic phase II activities measured, only 17α -ethynyl estradiol glucuronidation was significantly higher in the humans. These results clearly show that the metabolic capacities of the human and Rhesus monkey drug-metabolizing enzymes are quantitatively different.

The dihydropyridine calcium channel blockers are eliminated extensively by metabolism. The primary biotransformation route involves oxidation to their pyridine derivatives, a reaction that is known to be catalyzed by cytochrome P-450 (Bäärnhielm et al., 1984). In a recent review article, Smith (1993) compared the cl_{int} (metabolic clearance) of six dihydropyridines (amlodipine, nitrendipine, felodipine, nicardipine, nisoldipine, and nilvadipine) in rats, dogs, and humans. In all cases, the rat showed the highest cl_{int} when compared with dogs and humans. The overall ratio of cl_{int} of these compounds in dogs or rats to those in humans gives values of 1.4 for the dogs and 9 for the rats. For these drugs, the metabolism in humans is quantitatively similar to that in dogs, whereas rats show a much higher capacity for metabolism.

Drugs containing hydroxy groups are subject to both glucuronidation and sulfation reactions. The relative contribution of these two competing pathways depends on the nature of the drugs and animal species being studied. It is generally believed that glucuronidation predominates over sulfation in the rat, whereas in the dog and human, sulfation dominates (Rogers et al., 1987). Consistent with this general belief, xamoterol, a β_1 -adrenoceptor partial agonist, is extensively glucuronidated in the rat, whereas sulfation primarily occurs in the dog (Mulder et al., 1987; Groen et al., 1988). However, this is not the case for acetaminophen, which is predominately sulfated in the rat, but in humans, glucuronidation is quantitatively more important (Lin and Levy, 1986; Slattery and Levy, 1979).

Azidothymidine (AZT), an HIV reverse transcriptase inhibitor, is extensively metabolized in humans, but not in rats. Approximately 75% of an oral dose was recovered in human urine as the 5'-*O*-glucuronide, and 15% was recovered as unchanged drug (Blum et al., 1988). On the other hand, only 2% of an oral dose was recovered as AZT glucuronide in rat urine, whereas approximately 78% of the dose was excreted as unchanged drug (Good et al., 1986). Consistent with the *in vivo* data, *in vitro* studies confirmed that human liver UDPGT catalyzed the glucuronidation of 0.1 mM AZT 10- to 25-fold faster than did rat liver UDPGT (Resetar and Spector, 1989). Similarly, glucuronidation of some drugs, including quaternary amines, has been shown to occur only in human and primate species (Caldwell et al., 1989).

These examples clearly demonstrate that extrapolation of drug metabolism from animals to humans is very difficult, if not impossible, both in the qualitative and quantitative aspects. If drug-induced toxicity is related directly to systemic exposure to the drug and its metabolites, the species differences in the metabolism of the drug are perhaps the most important factors in explaining the observed species differences in toxic responses.

2. Induction. In the mid-1950s, Conney et al. (1956) showed that the treatment of animals with 3-methylcholanthrene (3-MC) increased the animals' ability to metabolize methylated aminoazo dyes. Remmer (1958) found that tolerance to barbiturate drugs was the result of the enhancement of their own metabolism by induction of cytochrome P-450. Although the phenomenon of induction has been known for over 4 decades, only in recent years, we began to uncover the mechanism involved in induction.

With the exception of the CYP1A1 isoform (Whitlock et al., 1996), many more studies are needed to explore the molecular mechanisms involved in CYP2B, 2E, 3A, and 4A induction. In the case of CYP1A1, inducing agents bind to the cytosolic polycyclic aromatic hydrocarbon (Ah) receptor and are translocated into the nucleus. The transcriptional process includes a sequence of events: ligand-dependent heterodimerization between the Ah receptor and Ah receptor nuclear translocator interaction of the heterodimer with a xenobiotic-responsive enhancer, transmission of the induction signal from the enhancer to the CYP1A1 promoter, and alterations in chromatin structure. This is followed by the subsequent transcription of the appropriate mRNA and translation of the corresponding proteins.

Although the fundamental mechanisms of CYP1A induction are qualitatively similar in different species, including mice, rats, rabbits, and humans (McDonnell et al., 1992), there are important quantitative differences in the effectiveness of inducer-receptor coupling. For example, the gastric acid-suppressing drug, omeprazole, is a CYP1A2 enzyme inducer in humans but has no such inductive effect in mice or rabbits (McDonnell et al., 1992; Diaz et al., 1990).

Important species differences also exist in the response of other inducible subfamilies of cytochrome P-450s. Phenobarbital induces predominately members of the CYP2B subfamily in rats, whereas in humans, it appears that the major form induced belongs to the CYP3A subfamily (Rice et al., 1992). Furthermore, members of the CYP3A subfamily in rats are inducible by the steroidal agent, pregnenolone-16 α -carbonitrile, but not by the antibiotic rifampin. The opposite is true in rabbits and humans (Strolin Benedetti and Dostert, 1994; Nebert and Gonzalez, 1990). Thus, drugs that do not induce P-450 enzymes in animals should not be assumed to not have enzyme-inducing capacity in humans, and vice versa. Despite the well-known species differences in the response to P-450 inducers, mice and rats have been routinely used in most pharmaceutical companies to assess the risk of potential drug induction in humans. This type of risk assessment may be of little direct relevance for certain drugs. More recently, however, both in vitro (human hepatocytes) and in vivo (probe drugs for certain human cytochrome P-450s) techniques have become available and have increasingly been used by investigators to evaluate the potential induction of human cytochrome P-450s by a variety of therapeutic agents.

Like a double-edged sword, induction of drug-metabolizing enzymes may lead to a decrease in toxicity through acceleration of detoxification, or to an increase in toxicity due to increased formation of reactive metabolites. Depending on the delicate balance between detoxification and activation, induction can be a beneficial or harmful response. The induction of CYP1A isoforms can reduce the carcinogenicity of certain compounds. For example, intraperitoneal injection of the CYP1A inducer β -naphthoflavone inhibited tumorigenesis in the lung and mammary glands of rodents treated with 7,12-dimethylbenzo[a]anthracene (DMBA), which is a highly carcinogenic compound (Wattenberg and Leong, 1968). In addition, 2,3,7,8-tetrachlorodibenzo-p-dioxin, a potent CYP1A inducer, dramatically reduced the initiation of skin tumors in mice caused by DMBA (Digiovanna et al., 1979). In contrast, CYP1A isoforms can activate some compounds, such as benzo[a]pyrene, to their ultimate carcinogenic forms (Gelboin, 1980), and induction of these isoforms increases the risk of carcinogenicity. Due to the complexity of the factors determining toxicity and carcinogenicity, the issue of whether induction is beneficial or harmful is still highly controversial (Ioannides and Parke, 1993; Beresford, 1993).

In addition to the induction of CYP1A isoforms, binding of an agent to the Ah receptor also leads to the induction of UDPGTs and glutathione (GSH)-S-transferases (Bock et al., 1990). The coinduction of phase I and phase II enzymes appears to decrease the risk due to P-450 induction alone. In vitro mutagenicity of benzo[a]pyrene and of benzo[a]pyrene-3,6-quinone was higher in the liver S9 fraction of 3-MC-treated rats than

with that of control rats when NADPH was the only added cofactor. The in vitro mutagenicity was decreased substantially by concomitant glucuronidation or GSH conjugation when UDPGA or GSH also was added to the system, and there was no significant difference in the in vitro mutagenicity between 3-MC-treated and control rats. (Bock et al., 1990). Thus, the protective effect appeared to be a result of coinduction of UDPGTs and GSH-S-transferases.

3. Inhibition. A drug interaction occurs when the disposition of one drug is altered by another. Because metabolism represents a major route of elimination for many drugs, inhibition of drug-metabolizing enzymes is one of the main reasons for drug interactions. Various mechanisms are known to underlie enzyme inhibition, including competition for the catalytic site of the enzyme, noncompetitive (allosteric) interaction with the enzyme, suicide destruction of the enzyme, and competition for cofactors. Among these, competitive inhibition is probably the most common. If enzyme inhibition occurs by the interaction of two substrates competing for the same enzyme, the competitive nature of the inhibition will depend on the K_m value of the substrate and the dissociation constant of an inhibitor (K_i) value of the inhibitor as well as their concentrations at the site of enzymes (Segel, 1975). Because there are quantitative differences in the K_m and K_i values between species, it is expected that the degree of enzyme inhibition would be species-dependent.

Isoforms of the CYP2D subfamily have been isolated from rats and humans and have been shown to have similar substrate specificities (Guengerich, 1987). Debrisoquine 4-hydroxylation is specifically catalyzed by these isoenzymes (Meyer et al., 1990b). The inhibition kinetics of debrisoquine 4-hydroxylase activity by quinidine and one of its diastereoisomers, quinine, have been compared in human and rat liver microsomes (Kobayashi et al., 1989). Both quinidine and quinine are potent competitive inhibitors of debrisoquine 4-hydroxylation. However, quinidine is a more potent inhibitor of this activity in humans than in rats, whereas the reverse is true for quinine. The K_i values of quinidine for debrisoquine 4-hydroxylation in humans and rats were 0.6 and 50 μM , whereas with quinine, the values were 13 and 1.7 μM , respectively. Similarly, furafylline exhibits species-dependent inhibition of phenacetin *O*-deethylase activity of liver microsomes (Sesardic et al. 1990). Furafylline, a mechanism-based inhibitor of CYP1A2, is more potent in inhibiting phenacetin *O*-deethylation in humans than in rats, despite the fact that phenacetin *O*-deethylation is catalyzed exclusively by CYP1A2 in both species.

The in vitro effects of α -naphthoflavone on aryl hydrocarbon hydroxylase activity (CYP1A subfamily) were studied in five animal species by Thorgeirsson et al. (1979). The activity of this enzyme was significantly inhibited by α -naphthoflavone in mice, rats, and ham-

sters in a concentration-dependent manner. In rabbits, the aryl hydrocarbon hydroxylase activity was stimulated, rather than inhibited, by this compound. Similarly, species-dependent stimulatory effects of flavonoids were also reported by Huang et al. (1981). Addition of 7,8-benzo[a]pyrene or flavone stimulated the hydroxylation of benzo[a]pyrene in liver microsomes from rabbits, hamsters, and humans by several-fold but had little or no effect on this activity in liver microsomes from rats or guinea pigs. The marked species differences in the inhibitory or stimulatory effects of flavonoids may be due to the involvement of species' different cytochrome P-450 isozymes, or to the structural differences at the active site of P-450 isozymes at which the flavonoids interact.

Recently, Shou et al. (1994) have shown that CYP3A4-catalyzed phenanthrene metabolism was stimulated by 7,8-benzoflavone. Kinetic studies with vaccinia virus coding CYP3A4 revealed that 7,8-benzoflavone increased the V_{\max} of phenanthrene metabolism without changing the K_m and that phenanthrene decreased the V_{\max} of 7,8-benzoflavone without increasing the K_m . With these data, these investigators speculate that both substrates are simultaneously bound to the enzyme in the active site that has access to the active oxygen and suggest that the increase in the V_{\max} of phenanthrene (or the decrease in the V_{\max} of 7,8-benzoflavone) indicates that there is competition for the active oxygen between the two substrates. Although the kinetic analyses support their speculation, the underlying mechanism of the stimulatory effect at the molecular level needs to be verified further.

In addition to the reversible competitive inhibitors, compounds can bind irreversibly to the enzyme via a reactive metabolic intermediate. Several drugs have been shown to be irreversible inhibitors or so-called mechanism-based suicide inhibitors (Murray, 1987). L-754,394, a potent HIV protease inhibitor, is a good example. Kinetic studies in rats, dogs, and monkeys have shown that the drug exhibits dose- and time-dependent pharmacokinetics (Lin et al., 1995). The apparent clearance decreased with increasing dose. However, the dose dependency cannot be explained by Michaelis-Menten kinetics. L-754,394 in plasma declined log-linearly with time, but with an apparent $t_{1/2}$ that increased with dose. Furthermore, the apparent clearance of L-754,394 decreased after chronic dosing. Subsequent *in vitro* microsomal studies revealed that the observed time- and dose-dependent kinetics of L-754,394 may be explained by mechanism-based enzyme inhibition of isozymes of the cytochrome CYP3A subfamily (Lin et al., 1995).

The magnitude of mechanism-based inhibition of cytochrome CYP3A isozymes by L-754,394 appeared to be species-dependent. For liver microsomal testosterone 6 β -hydroxylase, the potency of inhibition by L-754,394 was in the rank order human > monkey > dog > rat.

The values of maximum inactivation rate constant (K_{inact}) were 2.0, 0.25, 0.20, and 0.04 min^{-1} , respectively. Consistent with these results, *in vivo* kinetic studies indicated that L-754,394 inhibited the metabolism of indinavir (MK-0639), a drug known to be metabolized mainly by the isoforms of the CYP3A subfamily, more significantly in dogs than in rats (Lin et al., unpublished data). Because of this undesirable mechanism-based inhibition, the development of L-354,394 was terminated.

Drug inhibition is usually regarded as potentially dangerous, or at least undesirable. However, there are times when these interactions may be exploited. For example, ketoconazole is used with cyclosporin A to prolong elimination of the latter (Yee and McGuire, 1990; First et al., 1984). Ketoconazole, which is a potent antifungal agent, and cyclosporin A, which is a widely used immunosuppressive agent, are substrates for the same human cytochrome CYP3A4 (Combalbert et al., 1989). The idea is to use the relatively inexpensive ketoconazole to specifically inhibit cyclosporin A metabolism to minimize the cost of long-term therapy with this very expensive drug. Similarly, during World War II, when penicillins were very expensive, probenecid was coadministered to delay renal excretion of the antibiotics (Weiner and Mudge, 1985). Other successful examples of therapeutic inhibition are carbidopa and cilastatin (Marsden, 1976; Kropp et al., 1980). As mentioned earlier, carbidopa and cilastatin are used as inhibitors to slow the elimination of dopa and imipenem, respectively.

4. Sexual dimorphism. Sex-related differences in drug metabolism have been known for more than 60 years, but it was not until recently that the mechanisms for these differences were explored (Shapiro et al., 1995; Skett, 1989). Recent studies have shown that sexual dimorphism in rats, and possibly in other species, results from the differential expression of sex-dependent hepatic cytochrome P-450s. This differential expression, in turn, is largely influenced by steroid and pituitary hormone levels and profiles. Evidence has shown that the sexual dimorphic secretion pattern of growth hormone directly regulates the expression of certain hepatic cytochrome P-450s (Legrauerend et al., 1992b; Waxman, 1992; Kato and Yamazoe, 1990).

Such sex-related differences in the levels of cytochrome P-450 expression would be expected to give rise to profound differences in toxicological response because the susceptibility of a tissue to the toxic and/or carcinogenic effects of drugs often is determined by the rate of metabolic inactivation and/or activation by cytochrome P-450. For this reason, regulatory agencies require that equal numbers of males and females of each species be used in toxicity studies of drugs.

Although male rats generally exhibit distinctly higher activities than females, there are instances in which female rats have higher activities than males (Skett, 1989). This results from the fact that cytochrome P-450

can be expressed specifically or preferentially in either males or females. For example, CYP2C11 is expressed only in male rats, whereas CYP2C12 expression is limited to female rats. On the other hand, CYP2A2 and CYP3A2 are male-dominant, but CYP2A1 and CYP2C7 are female-dominant (Kobliakov et al., 1991; Bandiera, 1990; Legraverend et al., 1992a; Waxman et al., 1985, 1990).

The existence of sex-related differences in drug metabolism is not unique to the rat. Such differences have been seen in mice (Macleod et al., 1987), ferrets (Ioannides et al., 1977), dogs (Dogterom and Rothuizen, 1993), and humans (Hunt et al., 1992). However, the magnitude of the sexual differences in these species is invariably far more subtle than that found in rats. Sexual differences in drug metabolism are generally small and not detected easily in humans, due to the large interindividual variability in enzyme activities (Hunt et al., 1992).

Indinavir (MK-639, L-735,524), a potent HIV protease inhibitor, exhibits marked sex-related differences in clearance in rats and dogs, but not in monkeys. The clearance was 89 mL/min/kg for male rats and 41 mL/min/kg for female rats. In contrast to rats, female dogs cleared indinavir more rapidly than male dogs, with a clearance of 26 mL/min/kg for female dogs and 15 mL/min/kg for male dogs (Lin et al., 1996b). Consistent with the *in vivo* observations, hepatic microsomes from male rats had a substantially higher metabolizing activity toward indinavir than those from females, whereas liver microsomes from female dogs catalyzed the drug at a higher rate than those from male dogs. However, no sexual difference in indinavir metabolism was observed in monkey and human liver microsomes. The functional activity of CYP3A, measured by the formation of testosterone 6 β -hydroxylation, and immunoblot analysis of the level of CYP3A proteins strongly suggest that significant gender differences in the levels of CYP3A isoforms result in the observed sex-related differences in indinavir metabolism in rats and dogs (Lin et al., 1996b). This example demonstrates that the sexual dimorphism in drug metabolism can be species-dependent. The sexual dimorphism in indinavir metabolism is reversed in the rat and dog.

Reverse sexual dimorphism also has been observed in humans. The male has a higher unbound clearance of chlordiazepoxide than the female, whereas the reverse is true for diazepam and desmethyl diazepam (Wilson, 1984). The sex-related differences in drug disposition could be related to the phase of the menstrual cycle, sex hormones, and the use of oral contraceptives.

B. Species- and Tissue-Specific Toxicity

1. *Species-specific toxicity.* Toxic and carcinogenic responses for some drugs are evoked solely by the parent compounds, whereas for other drugs, the responses arise as a result of the formation of reactive toxic metabolites.

Sometimes, therefore, the *in vivo* monitoring of the parent drug alone may have very little relevance. Even if metabolites are monitored, the reactive toxic metabolites are often too labile and too small in quantity to be detected. Consequently, only the chemically stable and nonreactive metabolites are being monitored, yet their presence may be meaningless in predicting toxicity. This complicated situation can be illustrated by species differences observed in the pharmacokinetics and hepatotoxicity of acetaminophen. The elimination $t_{1/2}$ of acetaminophen was longer in rats than in mice by 2- to 3-fold. Acetaminophen caused hepatotoxicity at lower doses in mice (200–300 mg/kg) but evoked only a barely detectable hepatotoxicity in rats at considerably larger doses (>1500 mg/kg). Pretreatment of rats and mice with phenobarbital had little effect on the $t_{1/2}$ of acetaminophen in either of the two species but markedly increased the hepatotoxicity of acetaminophen in both species (Gillette, 1989). These species differences in toxicity depend mainly on the amount of reactive metabolite formed and the amount of GSH present in the liver. Thus, monitoring of acetaminophen and its sulfate and glucuronide conjugates in plasma is of little relevance in predicting hepatotoxicity. Clinically, acetaminophen is well tolerated within the therapeutic dose range; however, hepatotoxicity may occur after ingestion of a single high dose of 10 to 15 g (150–200 mg/kg) of acetaminophen.

Dichloromethane is a common industrial chemical that causes lung and liver cancer in mice after chronic inhalation exposure, but not in rats and hamsters under the same conditions (Burek et al., 1984). The compound is metabolized *in vivo* either by cytochrome P-450 to form carbon monoxide and carbon dioxide or by GSH-S-transferase to form GSH conjugates. Although the rate of oxidative metabolism is similar in rats, mice, and hamsters, there are marked species differences in the formation of GSH conjugates. The similar activity of cytochrome P-450 among these species results in very similar levels of carboxyhemoglobin in the blood of rats and mice. On the other hand, mice have substantially higher activity in GSH conjugation compared with rats and hamsters. Biochemical and toxicological studies suggest that the toxicity of dichloromethane is associated with the production of reactive metabolites derived from GSH conjugation via GSH-S-transferase (Green, 1990; Reitz et al., 1988, 1989).

Species-dependent toxicity also is observed with perfluorodecanoic acid (PFDA), a potent peroxisome proliferator. Treatment with PFDA resulted in pronounced hepatomegaly in the rat, but not in the guinea pig (Chinje et al., 1994). In a separate study, PFDA treatment caused a marked induction of lauric acid 12-hydroxylase activity in the rat, but not in the guinea pig, suggesting that hepatomegaly observed in rats may be associated with the induction of isozymes of the CYP4A subfamily mediated by peroxisome proliferator-acti-

vated receptors (PPAR) (Johnson et al., 1996). Another peroxisome proliferator, methylclofenapate, showed similar species differences in toxic response. This proliferator caused hepatomegaly in mice and rats, but not in guinea pigs (Bell et al., 1993). These data indicate that peroxisome proliferation is a species-dependent phenomenon most likely reflecting the differences in concentration of PPAR and the affinity of peroxisome proliferators to PPAR from different species.

D-limonene, a major component of orange oil, is an anticarcinogenic terpene. Studies in animals have shown that D-limonene reduces mammary tumorigenesis, although the underlying mechanism of this reduction is still unclear (Elson et al., 1988). Toxicity studies have shown that exposure to D-limonene causes nephrotoxicity only in male rats, but not in mice, guinea pigs, dogs, or monkeys (Webb et al., 1989; Hard and Whysner, 1994). Also, D-limonene causes no nephrotoxicity in female rats. This species- and sex-specific nephrotoxicity is characterized by an exacerbation of hyaline droplet accumulation. The mechanism underlying this accumulation of protein is due to the strong but reversible binding of D-limonene to α_{2u} -globulin, a specific protein only produced in male rats (Kanerva et al., 1987; Stornard et al., 1986). Because α_{2u} -globulin is not present in humans, it is concluded that D-limonene does not pose any nephrotoxic risk to humans.

Drug-induced thyroid enlargement and tumors are seen primarily in rodents. For example, the sulfonamide, sulfamethoxazole, produced thyroid nodules in rats at a dose of 50 mg·kg·day for 1 year. In Rhesus monkeys, the drug caused no increase in thyroid weight and no morphological alterations even at 300 mg·kg·day administered for the same length of time (Swarm et al., 1973). This species-specific toxicity may be due to the fact that rodents lack thyroxin-binding globulin. As a consequence, the biological plasma $t_{1/2}$ of thyroxin in rats is approximately ten-fold shorter than in humans, i.e., 12 to 24 h versus 5 to 9 days (Döhler et al., 1979). Sulfonamides cause an increase in thyroxine elimination, which leads to a more rapid depletion of the hormone in the rat than in other species and produces a need for prompt regulatory responses by the hypophysis. This leads to the elevation of thyroid-stimulating hormone, which results in a chronic hyperplastic response in thyroid tissue.

Drugs and their metabolites are usually eliminated from the body via urine or bile, or both. The relative contribution of biliary and urinary excretion to the overall elimination of drugs depends on the physicochemical properties of the drug and the animal species. The biliary excretion of drugs varies widely among species. In general, the mouse, rat, and dog are good biliary excretors, whereas the rabbit, guinea pig, monkey, and human are relatively poor (Smith, 1971). Sometimes, biliary excretion of drugs may lead to unwanted adverse effects. Indomethacin, an anti-inflammatory agent, is widely

used and highly effective in the treatment of rheumatoid arthritis. However, at high doses, this drug may cause ulcerative lesions in the upper gastrointestinal tract. The biliary excretion of indomethacin appears to be an important factor in the development of intestinal lesions. The tendency for different species to develop intestinal lesions in response to indomethacin appears to correlate well with their respective biliary excretion of this compound (Duggan et al., 1975). Good biliary excretors, such as rats and dogs, appeared to be the most susceptible to indomethacin-induced intestinal lesions.

2. *Site-specific toxicity.* 4-*Ipomeanol*, a pulmonary toxin, is a naturally occurring fungal catabolite of a furanoterpenoid precursor produced by the moldy sweet potato *Ipomoea batatas*. It was discovered in the 1970s that 4-*ipomeanol* was the causative agent responsible for the outbreaks of lethal interstitial pneumonia in cattle. 4-*Ipomeanol* undergoes metabolic activation to a highly reactive metabolite that binds to nucleophilic tissue macromolecules (Boyd and Burka, 1978). In vivo and in vitro studies in several animal species revealed that the covalent binding occurs primarily in the lung, specifically in bronchiolar Clara cells (Boyd, 1977; Devereux et al., 1982). Because of its lung-specific toxicity, 4-*ipomeanol* was at one time considered to be a potential agent for the treatment of lung cancer. Although some human lung cancer cell lines, as well as a variety of human lung tumor biopsy specimens, are shown to be capable of activating 4-*ipomeanol* to a cytotoxic intermediate (Christian et al., 1989), the considerable toxicity of this compound hinders its clinical use in lung cancer therapy.

Although it is generally believed that most reactive intermediates and toxic metabolites of drugs are generated by oxidative reactions, an increasing number of examples suggest that phase II metabolism such as glucuronide, sulfate, and GSH conjugates may be related to drug-induced toxicity (Bock and Lilienblum, 1994; Miller and Surh, 1994; Monks and Lau, 1994). Glucuronides are capable of serving as transport vehicles for carcinogens that are responsible for site-specific toxicities in the urinary bladder or colon epithelium. Aromatic amines such as 2-naphthylamine and 4-aminobiphenyl are found in cigarette smoke and are considered to be a major factor in the incidence of urinary bladder cancer in humans (Mommsen and Aagaard, 1983). The *N*-hydroxy metabolite of 2-naphthylamine has been shown to be carcinogenic. Glucuronidation leads to the formation of *N*-hydroxy-*N*-glucuronide, which is more stable than the *N*-hydroxy metabolite and is excreted into the urinary bladder. In the urinary bladder, the *N*-hydroxy-*N*-glucuronide decomposes under the slightly acidic pH of urine to its protonated nitrenium ion, which readily reacts with DNA, thereby initiating bladder cancer (Kadlubar et al., 1981, 1977). A scheme of underlying mechanism for bladder-specific toxicity caused by 2-naphthylamine is illustrated in figure 4.

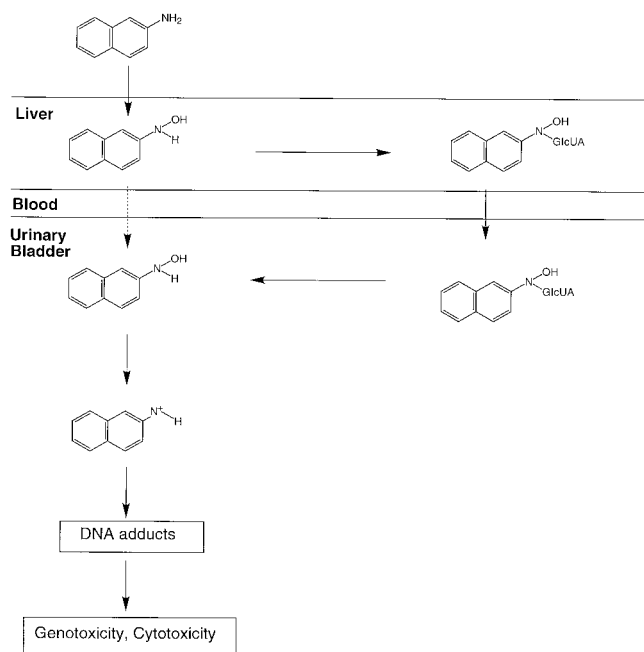


FIG. 4. Hypothesis for 2-naphthylamine-induced bladder cancer. Reprinted from Chem. Biol. Interact., Vol. 33, Kadlubar et al., Alteration of urinary levels of the carcinogen, *N*-hydroxy-2-naphthylamine, and its *N*-glucuronide in the rat by control of urinary pH, inhibition of metabolic sulfation, and changes in biliary excretion, pp. 129–147, 1981, with kind permission from Elsevier Science Ireland Ltd., Bay 15K, Shannon Industrial State, Co. Clare, Ireland.

Similar to the transportable glucuronides of arylamines involved in bladder carcinogenesis, glucuronides also play a major role in the colon cancer caused by heterocyclic arylamines. 2-Amino-1-methyl-6-phenylimidazo[4,5,*b*]pyridine is converted in the liver to the *N*-hydroxy-*N*-glucuronide metabolite, which is then excreted via the bile into the intestine where the corresponding carcinogenic hydroxylamine is liberated in the colon by bacterial β -glucuronidases (Alexander et al., 1991).

Although GSH conjugation is probably the most important detoxification function in all mammals, increasing evidence reveals that conjugation with GSH can result in toxicity (Monks and Lau, 1994, 1992). Kidneys possess very high levels of γ -glutamyl transpeptidase. The high renal activity of the enzyme plays a major role in the kidney-specific toxicity of some compounds. Some GSH conjugate-mediated toxicities involve the intermediary cleavage of the GSH moiety specifically by γ -glutamyl transpeptidase. The GSH conjugate of 2-bromohydroquinone is a good example of the GSH conjugate transporting form of a reactive metabolite. The nephrotoxicity of bromobenzene in rats probably occurs via its metabolism to 2-bromo-(di-glutathion-*S*-yl)hydroquinone. Although 2-bromo-(di-glutathion-*S*-yl)hydroquinone is formed in the liver, it travels in the blood stream to the kidney, where 2-bromo-(di-cystein-*S*-yl)hydroquinone, a nephrotoxic metabolite, is formed by renal γ -glutamyl transpeptidase (Monks and Lau, 1994).

Recently, a new mechanism of GSH conjugate-mediated toxicity has been proposed for toxic compounds, such as isothiocyanates and isocyanates (Baillie and Kassahun, 1994; Baillie and Slatter, 1991). Both of these classes are very electrophilic and react readily with GSH. The GSH conjugate is formed in one organ and can be transported to other organs, whereby the reactive moiety can be regenerated upon spontaneous decomposition of the GSH conjugate to GSH and the original reactive moiety. The reversible GSH conjugation may serve to extend the biological $t_{1/2}$ of reactive isocyanates and isothiocyanates and influence their tissue distribution. Unlike renal γ -glutamyl transpeptidase-mediated GSH conjugate toxicity, the toxicity caused by the reversible GSH conjugation is expected to be less site-specific.

C. Stereoselectivity and Toxicity

1. Stereoselective metabolism. As stated earlier, enantiomers must be considered as essentially different chemical compounds, because they usually differ greatly in pharmacokinetic and pharmacodynamic properties as a consequence of stereoselective interaction with biological macromolecules (Testa, 1988, 1989b; Ariens, 1990; Williams and Lee, 1985). During the last decade, there has been growing awareness of the stereoselectivity in pharmacokinetics and pharmacodynamics, and it has become a key issue in new drug development (Campbell, 1990; Testa and Trager, 1990).

As with the examples of stiripentol and timolol described previously (Zhang et al., 1994; Tang et al., 1994; Shen et al., 1992; Adams et al., 1976; Lee et al., 1985; Blaine et al., 1982; Tobert et al., 1981; Weiner and Taylor, 1985; Share et al., 1984; Keates and Stone, 1984), early evaluation of the stereoselective pharmacokinetics and pharmacodynamics of the enantiomers is essential to decide whether to develop a racemate or an individual enantiomer. However, at the early stage, such stereoselectivity studies can only be conducted in animals. Thus, species-related differences in stereoselectivity should be evaluated carefully before the data are extrapolated to humans.

Absorption from the gastrointestinal tract, distribution into tissues, and renal excretion are passive processes for most drugs in which the extent and rate are mainly governed by the physicochemical properties of the drug. Because the physicochemical properties of stereoisomers are similar, stereoselectivity is not expected for these processes unless an active transport system is involved. There are only a few examples of stereoselective drug absorption, tissue distribution and renal excretion, whereas the stereoselective plasma protein binding and metabolism of chiral drugs are well documented (Lee and Williams, 1990; Jamali et al., 1989).

Stereoselective plasma protein binding of drugs differs considerably among species. Plasma binding of MK-571, a potent leukotriene D₄ antagonist, has been stud-

ied in 12 mammalian species (Lin et al., 1990b). The binding of MK-571 enantiomers to plasma protein was extensive, stereoselective, and species-dependent. In some species, the *S*-enantiomer bound to a greater extent than the *R*-enantiomer. In others, the *R*-enantiomer bound more extensively, and in still other species, there was no stereoselectivity. For both enantiomers, the unbound fraction in plasma differed by a factor of 8 among the species studied. Consistent with these observations, the *R*-enantiomer of MK-571 bound to rat plasma to a greater extent than the *S*-enantiomer, whereas in dog and monkey plasma, the reverse was true (Tocco et al., 1990). The elimination clearance of the enantiomers was related to the stereoselective plasma protein binding, with the greater unbound fraction being cleared more rapidly.

Such stereoselectivity among species also has been seen in metabolism. The stereoselective metabolism of mephenytoin was studied *in vitro* using livers from different animal species and humans (Yasumori et al., 1993a). The rates of microsomal 4'-hydroxylation were 2 to 6 times higher with the *R*-mephenytoin than *S*-enantiomer in rabbits, dogs, and rats, whereas the rates of microsomal 4'-hydroxylation were 5 to 15 times higher with the *S*-mephenytoin than *R*-enantiomer in monkeys and humans. Reconstituted enzyme systems and immunoinhibition experiments revealed that stereoselective involvement of CYP2C and CYP3A isoforms is the major factor in the species differences in the stereoselective metabolism of mephenytoin (Yasumori et al., 1993a). Propranolol also shows species-dependent stereoselective metabolism. The *S*-propranolol had a higher cl_{int} than the *R*-enantiomer in dogs, whereas the *R*-propranolol had a higher cl_{int} than the *S*-enantiomer in humans (Silber et al., 1982).

Species differences in stereoselectivity have been seen in phase II metabolism reactions, such as glucuronidation. Using immobilized microsomal protein from rabbit, monkey, and human liver, El Mouelhi et al. (1987) have shown that the glucuronidation of three racemic 2-arylpropionic acids, naproxen, ibuprofen, and benoxaprofen, was stereoselective and species-dependent. Similarly, species-dependent stereoselective glucuronidation of oxazepam has been shown among rabbits, dogs, monkeys, miniature pigs, and humans (Sisenwine et al., 1982). The ratios of *S/R* oxazepam glucuronides in the urinary excretion was 2.0 for rabbits, 3.0 for dogs, 0.55 for monkeys, 1.2 for pigs, and 3.4 for humans. In addition, species differences in stereoselective hydrolysis have been reported. Methylphenidate (MPH), a methyl ester, is used as a racemic mixture in the treatment of children with attention deficit disorder, and its *D*-MPH is pharmacologically more active. The plasma esterases of rats, cattle, and rabbits appeared to hydrolyze *L*-MPH faster than the *d*-enantiomer, whereas the plasma of humans, dogs, and horses hydrolyzed *d*-MPH faster than *L*-MPH (Srinivas et al., 1991).

In addition to the quantitative species differences in the magnitude of stereoselective metabolism, qualitative species differences in stereoselective metabolism also occur. The metabolism of disopyramide, a quinidine-like antiarrhythmic agent containing a chiral center, is an example. Cook et al. (1982) have shown that arylhydroxylation is the major metabolic pathway of racemic disopyramide in rats, whereas *N*-dealkylation is the only pathway in dogs and that the *S*-disopyramide is cleared more rapidly than the *R*-enantiomer in both rats and dogs. These results indicate that different metabolic pathways, presumably by different isoforms of cytochrome P-450, are responsible for the stereoselective metabolism between animals. Similarly, the involvement of different enzyme systems in stereoselective metabolism also has been reported for other drugs. Sulindac, an NSAID, contains a chiral sulfoxide moiety and is dosed as a racemate. Sulindac is reversibly reduced to the achiral, pharmacologically active sulfide metabolite. The oxidation of sulfide back to sulindac is stereoselective, forming two enantiomers, sulindac A and sulindac B. *In vitro* studies with human liver microsomes indicated that sulindac A is formed by flavin-containing monooxygenases, and the formation of sulindac B is catalyzed by cytochrome P-450 enzymes (Hamman et al., 1994).

Considerable interspecies variability exists with respect to the process of chiral inversion. Flurbiprofen, a 2-arylpropionic acid NSAID, is a racemate and is marketed as such. Like other 2-arylpropionic acid NSAIDs, the anti-inflammatory activity of flurbiprofen is believed to reside in the *S*-enantiomer only. A unique characteristic of the metabolism of this class of drugs is the unidirectional inversion of the *R*- to the *S*-enantiomer (Wechter et al., 1974; Hutt and Caldwell, 1983), a process that is species-dependent. The extent of chiral inversion of flurbiprofen is complete in the guinea pig (100%), incomplete in the dog (40%), and very low in the rat and gerbil ($\leq 5\%$) (Manzel-Soglowek et al., 1992). Similar to the rodent species, the inversion of flurbiprofen is also insignificant in humans (Jamali et al., 1988). Species-dependent chiral inversion was also observed for another 2-arylpropionic acid NSAID, ketoprofen. The extent of inversion was high in the rat (Foster and Jamali, 1988), low in the rabbit (Abas and Meffin, 1987), and very small in humans (Jamali et al., 1989).

2. Stereoselective toxicity. Drugs exert wanted and unwanted pharmacological effects that are determined, on the one hand, pharmacodynamically by their interaction with the particular enzyme or receptor, and on the other hand, pharmacokinetically by their access to their site of action. Because there are frequently large pharmacodynamic and pharmacokinetic differences between enantiomers, it is not surprising that enantiomers may result in stereoselective toxicity.

Most anticancer drugs are cytotoxic due to their chemical reactivity. For this class of drugs, toxicity is simply

an extension of the therapeutic action. Not surprisingly, the major problem with the currently used anticancer drugs is their toxicity toward noncancerous cells. To reduce this undesirable toxicity of antineoplastics, various approaches have been taken to improve their therapeutic indices. One approach has been to exploit the stereoselective toxicity of some chiral antitumor agents. Cyclophosphamide, for example, contains a chiral center at the phosphorus atom and is used clinically as its racemic form. Cox et al. (Cox et al., 1976a,b) reported that the (-)-enantiomer of cyclophosphamide had twice the therapeutic index (LD_{50}/ID_{90}) of the (+)-enantiomer against the ADJ/PC6 cell turnover in mice. In clinical applications, however, there was no significant therapeutic advantage gained by using the single enantiomer.

In the past, barbiturates were used extensively as hypnotics. These compounds are rarely used today because of the numerous adverse reactions that have been associated with their use. One of the untoward effects of barbiturates is their excitatory aftereffects. The excitation phenomena range from mild tremors to conclusive seizures. 5-(1,3-dimethylbutyl)-5-ethyl barbituric acid (DMBB) has been used extensively in the investigation of the mechanism of the excitatory effects associated with barbiturate administration. The *S*-(+)-DMBB isomer induced extensive seizures, whereas the *R*-(-)-isomer induced preanesthetic excitation without seizures (Downes et al., 1970; Downes and Williams, 1969). The LD_{50} of the *S*-(+)-DMBB in mice was 3 mg/kg i.v., whereas that of the *R*-(-)-isomer was 72 mg/kg i.v., indicating stereoselective toxicity in that species.

The hypnotic drug thalidomide was taken off the market in Europe after it was tragically found to cause a rare birth defect known as phocomelia. The tragedy led to the passage of the Harris-Kefauver Amendment to the Federal Pure Food and Drug Act in the United States in 1962 to ensure that approved drugs have proof of safety and efficacy (Blaschke et al., 1985). Thalidomide contains a chiral center, and both enantiomers are equally sedating; thus, during its use it was supplied as the racemate. In studies with mice and rats, Blaschke and his coworkers (Blaschke et al., 1979; Blaschke, 1980) found that the *S*-enantiomer of thalidomide was teratogenic, whereas the *R*-isomer was not teratogenic. After intraperitoneal administration of the *S*-enantiomer (200 mg·kg·day) to pregnant animals, the percentage of fetuses born deformed was approximately 30% in mice and 50% in rats. However, no deformed fetuses were found when the *R*-isomer was given intraperitoneally at the same dose to a similar population of animals. By contrast, both enantiomers of thalidomide appeared to be equally teratogenic when administered to rabbits, and the racemate appeared to be even more teratogenic (Fabro et al., 1967; Simonyi, 1984). The percentage of deformed fetuses born from rabbits which were given these treatments was approximately 40% for the racemate, but only 16 to 17% for either *S*- or *R*-enantiomer

when given at an equal daily dose (150 mg·kg·day) to pregnant rabbits. Clearly, the stereoselective toxicity of thalidomide is species-dependent. A sad thought is that if the underlying mechanism of the species-dependent stereoselective toxicity was carefully explored, then the thalidomide tragedy could have been avoided.

Due to the limits in available technology in the 1960s and 1970s, studies carried out with thalidomide as a racemate and each of its enantiomers could not be clearly elucidated regarding the mechanisms that resulted in the stereoselective toxicities. With advances in knowledge of molecular biology and stereochemistry, the underlying mechanisms of many stereoselective toxicities are beginning to be understood. For instance, it is now known that stereoselective bioactivation plays a very important role in the carcinogenesis of environmental pollutants (Testa, 1989a; Trager and Testa, 1985; Trager, 1989).

One of the best-documented examples illustrating stereoselective bioactivation is the biotransformation of benzo[*a*]pyrene by CYP1A1 (Thakker et al., 1988; Vermeulen, 1989; Jerina et al., 1979). Initial oxidation of benzo[*a*]pyrene by the CYP1A1 results in the selective formation of the 7*R*,8*S*-arene oxide, which, upon hydrolysis by epoxide hydrolase, is converted to 7*R*,8*S*-dihydrodiol-benzo[*a*]pyrene. This compound is then converted in a highly stereoselective reaction by the same cytochrome P-450 isozyme to the diastereomeric (+)benzo[*a*]pyrene 7*R*,8*S*-diol-9*S*,10*R*-epoxide-2 (>80%) (fig. 5). For the diastereomeric pairs of bay-region diol epoxides

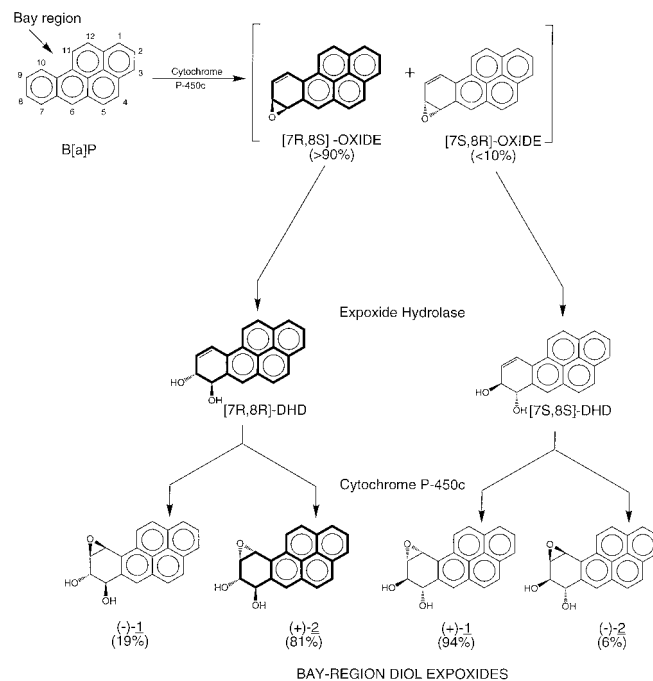


FIG. 5. Regio- and stereoselectivity of CYP1A and epoxide hydrolase in the formation of bay-region diol epoxides of benzo[*a*]pyrene. Reprinted from Thakker et al. (1988).

of benzo[a]pyrene, only diol epoxide-2 diastereomers show substantial carcinogenic activity, indicating that stereochemical factors play an important role in the carcinogenicity of benzo[a]pyrene. It has been proposed that the hydroxy groups in the bay region are predominantly axial in diol epoxide-1 and equatorial in diol epoxide-2 and that their absolute configuration is directly related to their carcinogenicity (Thakker et al., 1988).

Similar to benzo[a]pyrene, aflatoxin B1 (AFB) is a potent carcinogen that undergoes stereoselective bioactivation by cytochrome P-450 isozymes to form AFB exo-epoxide plus small amounts of the AFB endo-epoxide (Baertschi et al., 1989; Raney et al., 1992). The exo-epoxide reacts readily with DNA to give high adduct yields, but the endo-epoxide is nonreactive (Baertschi et al., 1988; Iyer et al., 1994).

IV. Role of Pharmacokinetics and Metabolism in Drug Development

In recent years, there has been a large expansion in both the range and use of in vitro systems to study absorption and metabolism. Due to the simplicity of in vitro systems, they are very useful in studying the factors influencing drug absorption and metabolism. A trickier task is to use these in vitro systems to predict quantitatively in vivo drug absorption and metabolism. The difficulty in extrapolating in vitro to in vivo lies in the complexity of the whole body with its greater number of interdependent events. Therefore, it is important to carefully set up the in vitro experimental conditions that simulate the in vivo situations. In addition, a good understanding of pharmacokinetic principles will help the in vitro/in vivo extrapolation.

A. In Vitro Studies of Drug Metabolism

1. *Determination of metabolic pathways.* In drug development, early information on human metabolism of a new drug is critical in predicting potential clinical drug-drug interactions and in selecting the appropriate animal species for toxicity studies. For human risk assessment, it is required by regulatory agencies to demonstrate that the systemic exposure of an unchanged drug and its major metabolites in animal species used in the toxicity study exceed that expected in humans to ensure a safety margin. It is important, therefore, to select animal species that have metabolite profiles similar to humans. However, the in vivo human drug metabolism normally is not carried out until the later stages of drug development, which is often too late for animal selection. Fortunately, the increased availability of human tissues and the advances in bioanalytical and biochemical technologies have provided opportunities for in vitro studies of human metabolism at the early stage of drug development before the toxicity studies (Wrighton et al., 1993; Chiu, 1993; Rodrigues, 1994; Powis, 1989).

The metabolite profile of a drug obtained in vitro generally reflects the in vivo metabolite pattern, although limited to qualitative aspects. From physiological and biochemical points of view, precision-cut liver slices are especially useful to obtain the complete in vitro metabolite profile of a drug, because this system retains the physiological conditions of enzymes and cofactors of both phase I and phase II reactions and, therefore, better simulates the in vivo situation (Dogterom, 1993).

As mentioned earlier, the major metabolic pathways of indinavir (MK-639, L-735,524) in humans have been identified as: (a) glucuronidation at the pyridine nitrogen to yield a quaternary ammonium conjugate, (b) pyridine *N*-oxidation, (c) *para*-hydroxylation of the phenylmethyl group, (d) 3'-hydroxylation of the indan and (e) *N*-depyridomethylation. The metabolite profile of indinavir obtained from human liver slices accurately reflected the in vivo human metabolite pattern (Balani et al., 1995). Although all of the oxidative metabolites of indinavir also were formed by human liver microsomes, the *N*-glucuronide was not detected when indinavir was incubated with native or Triton X-100-treated (Sigma, St. Louis, MO) human liver microsomes in the presence of 10 mM UDPGA (Lin et al., 1996a). The reason for the inability of human liver microsomes to form the *N*-glucuronide is not clear. Nevertheless, these results suggest that the liver slice is a better in vitro model to study the metabolic pathways of drugs.

Although liver slices are valuable in identifying metabolic pathways, their use in obtaining kinetic parameters may be limited. Houston and his coworkers (Worboys et al., 1996) have shown that the values of cL_{int} (V_{max}/K_m) of a series of drugs in slices are consistently less than those in hepatocytes by a factor ranging from 2 to 20. These results strongly suggest that a distribution equilibrium is not achieved between all the cells within the slice and the incubation media, due to the slice thickness ($\sim 260 \mu\text{m}$).

Isolated and cultured hepatocytes also are used often as in vitro models for identifying metabolic pathways of drugs. In vitro metabolism of ketotifen, an antiasthmatic drug, by cultured rat, rabbit, and human hepatocytes was consistent with the in vivo metabolic pathways, namely oxidation in rat hepatocytes, oxidation, glucuronidation, and sulfation in rabbit hepatocytes, and reduction and glucuronidation in human hepatocytes (Le Bigot et al., 1987). However, the results obtained from hepatocytes should also be interpreted with caution when quantitative comparison is the objective, because many enzyme activities decline spontaneously during hepatocyte isolation or culture. The metabolism of biphenyl has been compared by isolated hepatocytes and liver slices from rats, dogs, and humans (Powis et al., 1989). Human and dog, but not rat, isolated hepatocytes had decreased drug-metabolizing activities of oxidation and conjugation reactions of biphenyl as com-

pared with liver slices. Furthermore, it has been reported that substantial loss of cytochrome P-450 content was observed during the first 24 h of culture (Padgham and Paine, 1993; Padgham et al., 1992).

Another important consideration is the choice of drug concentrations for in vitro studies. The major metabolic pathway may be shifted, depending on the drug concentration used. The clinical studies indicated that *N*-demethylation is the major metabolic pathway of diazepam in humans (Bertilsson et al., 1989). However, in vitro studies in human liver microsomes showed that 3-hydroxylation was the major metabolic pathway of diazepam metabolism when a high (100 μM) drug concentration was incubated (Inaba et al., 1988). This in vitro and in vivo discrepancy could be due to the differences in the substrate concentration used. Indeed, the major metabolic pathway of diazepam is *N*-demethylation in human liver microsomes when an in vivo relevant substrate concentration (2–4 μM) is used (Yasumori et al., 1993b).

2. Identification of drug-metabolizing enzymes. Over the last 10 years, a great deal of information on human cytochrome P-450s and phase II drug-metabolizing enzymes at the molecular level has become available (Gonzalez et al., 1991; Nelson et al., 1993, 1996; Burchell et al., 1991). This information, with the availability of antibodies and probe substrates, has made it possible to determine which isozyme(s) is/are responsible for a specific reaction of a drug in vitro and in vivo.

To identify which cytochrome P-450 isozymes are responsible for metabolizing drugs in humans, several in vitro approaches have been developed, including (a) use of selective inhibitors with microsomes, (b) demonstration of catalytic activity in cDNA-based vector systems, (c) metabolic correlation of an activity with markers for known enzymes, (d) immunoinhibition of catalytic activity in microsomes, and (e) catalytic activity of purified enzyme isoforms (Tuengerich and Shimada, 1993). Each approach has its advantages and disadvantages, and a combination of approaches is usually required to accurately identify which cytochrome P-450 isozyme is responsible for metabolizing a drug.

Metabolism of drugs is usually very complex, involving several pathways and various enzyme systems. In some cases, all the metabolic reactions of a drug are catalyzed by a single isozyme, whereas, in other cases, a single metabolic reaction may involve multiple isozymes or different enzyme systems. The oxidative metabolic reactions of indinavir (MK-639, L-735,524) are all catalyzed by a single isozyme, CYP3A4, in human liver microsomes (Chiba et al., 1996). Similarly, CYP3A4 catalyzes both the *N*-dealkylation and *C*-hydroxylation of the antihistamine drug terfenadine in humans (Yun et al., 1993). In contrast, two isozymes, CYP1A2 and CYP3A4, are involved in imipramine *N*-demethylation in human liver microsomes (Lemoine et al., 1993). The *S*-oxidation of 10-(*N,N*-dimethylaminoalkyl)phenothiazines in human liver microsomes is catalyzed by several cyto-

chrome P-450s, including CYP2A6, 2C8, and 2D6 (Cashman et al., 1993). The complexity of metabolism results from the multiplicity of enzyme systems.

The stereoselective metabolism of drugs may result from the involvement of different isoforms. Warfarin, an oral anticoagulant, is marketed as a racemic mixture consisting of equal amounts of *R*- and *S*-warfarin, and its metabolism is stereoselective. Humans metabolize *S*-warfarin almost entirely to form *S*-7-hydroxywarfarin and a smaller amount of *S*-6-hydroxywarfarin. On the other hand, *R*-warfarin is converted mainly to *R*-6-hydroxywarfarin and some 7-hydroxywarfarin (Lewis et al., 1974). In vitro studies with human liver microsomes indicate that both 6- and 7-hydroxylation of *S*-warfarin are catalyzed exclusively by CYP2C9, whereas 6- and 7-hydroxylation of *R*-warfarin is mediated mainly by CYP1A2 and CYP2C19 (Kunze et al., 1996). Similarly, the 4'-hydroxylation of *R*- and *S*-mephenytoin is stereoselective and catalyzed by different isoforms. The rate of microsomal 4'-hydroxylation was 2 to 3 times higher with *R*-mephenytoin than its *S*-enantiomer in rats. Reconstituted systems and immunoinhibition studies suggest that 4'-hydroxylation of *S*-mephenytoin in rats is catalyzed by CYP2C11, whereas 4'-hydroxylation of *R*-mephenytoin is metabolized by CYP3A1/2 (Yasumori et al., 1993a). The stereoselective metabolism of mephenytoin is species-dependent. In contrast to rats, 4'-hydroxylation of *S*-mephenytoin is catalyzed preferentially in human liver microsomes, and this reaction is mediated exclusively by CYP2C9 (Yasumori et al., 1993a).

Like the cytochrome P-450s, multiple UDPGT isoforms can be involved in the glucuronidation of drugs. In vitro evidence has shown that at least two forms of human liver UDPGTs catalyze morphine 3-glucuronidation (Miners et al., 1988). Studies of the effects of different enzyme inducers, such as phenobarbital, 3-MC, and β -naphthoflavone, in rats suggest that multiple forms of UDPGTs are involved in the glucuronidation of diflunisal (Lin et al., 1987a).

Recently, Stearns et al. (1995) have demonstrated that losartan (MK-954) is converted to its active carboxylic acid metabolite (L-158,641) via the aldehyde intermediate (L-158,610) in human liver microsomes. In an atmosphere of $^{18}\text{O}_2$, losartan and L-158,610 were converted to the active metabolite L-158,641 in a reaction that was both NADPH- and oxygen-dependent. The investigators have also shown that CYP2C9 and 3A4 are the major enzymes responsible for each of the two-step oxidative reactions of the formation of the active metabolite (Stearns et al., 1995). However, published in the same issue of *Drug Metabolism and Disposition*, Yun et al. (1995) concluded that only CYP3A4 is involved in the biotransformation of losartan to its active metabolite. Although the reason for this discrepancy is not clear at the present time, differences in experimental conditions between the studies may have led to different conclusions. Furthermore, because different human liver mi-

crossosomal preparations were often used by different laboratories, differences in intrinsic properties of the cytochrome P-450 population (such as the presence of allelic variants) also may contribute to different results regarding specific cytochrome P-450 involvement.

Because several cytochrome P-450 isoforms with distinct K_m values can contribute to the metabolism of a single drug, selection of the substrate concentration in enzyme identification is important. For example, a CYP2C antibody shows no inhibition of diazepam *N*-demethylation at a substrate concentration of 200 μM but inhibits over 80% at a substrate concentration of 20 μM (Kato and Yamazoe, 1994). This clearly illustrates the importance of using clinically relevant substrate concentrations for *in vitro* studies, either in determining metabolic profiles or identifying drug-metabolizing enzymes. The selection of concentrations of isozyme-selective inhibitors also is important. A recent study by Newton et al. (1995) has shown that the specificity of isozyme-selective inhibitors of cytochrome P-450 is concentration-dependent. Quinidine, a selective inhibitor of CYP2D6, exhibited maximum inhibitory effect on CYP2D6-catalyzed bufuralol 1'-hydroxylation activity at 5 to 10 μM . At higher (>20 μM) concentrations, quinidine also inhibited CYP3A4-mediated testosterone 6 β -hydroxylation activity. The concentration-dependent selectivity of inhibitors also has been reported by other investigators (Guengerich, 1986; Ward and Back, 1993). Judicious selection of inhibitor concentration is of importance when determining the contribution of a specific cytochrome P-450 isoform(s) to a given reaction.

3. Drug-drug interaction. Concomitant administration of several drugs is common and, indeed, is often the situation in hospitalized patients. Whenever two or more drugs are administered over similar or overlapping time periods, the possibility for drug interactions exists. Although drug interactions can be explained by pharmacokinetic or pharmacodynamic effects, in many cases, the interactions have a pharmacokinetic, rather than pharmacodynamic, basis.

Interaction by mutual competitive inhibition between drugs is almost inevitable, because metabolism represents a major route of drug elimination and because many drugs can compete for the same enzyme system. The risk of clinical consequences from drug-drug interactions is higher with some drugs than with others. Patients receiving anticoagulant, antidepressant, or cardiovascular drugs are, due to the narrow therapeutic index of these drugs, at much greater risk than patients receiving other kinds of drugs (May et al., 1977). Most of the interactions are predictable and manageable, usually by appropriate dosage adjustment, whereas a few are potentially life-threatening. Coadministration of terfenadine, an antihistamine agent, and ketoconazole led to fatal ventricular arrhythmias in some patients (Monahan et al., 1990). Studies by Honig and his colleagues (1993, 1992) revealed that terfenadine is metab-

olized extensively by CYP3A4 isozymes, and ketoconazole, a potent CYP3A4 inhibitor, inhibited the metabolism of terfenadine, resulting in elevation of terfenadine, which, in turn, caused the prolongation of the QT interval.

Drug-drug interaction studies have become an important aspect of the development process of new drug candidates because of potential adverse effects. Because studies of all possible interactions are neither practicable nor economic, careful selection of a limited number of drug combinations to be studied is essential. Principally, the selection of drug interaction studies is usually based on two main criteria: the likelihood of coadministration and the therapeutic index (Tucker, 1992). Even with these criteria, clinical studies to assess drug interactions with new drug candidates are still very costly and time-consuming. With the advanced *in vitro* technologies in drug metabolism available today, an alternative approach is to use *in vitro* systems. These systems are valuable aids as screening tools to predict drug-drug interactions (Peck et al., 1993; Wrighton and Ring, 1994). Increasing evidence has demonstrated that *in vitro* interaction studies can accurately reflect the *in vivo* situation (Wrighton and Ring, 1994; Riesenman, 1995; Shen, 1995).

Many pharmaceutical companies now use *in vitro* techniques to predict the potential drug interactions of new drug candidates. However, several factors need to be considered when the *in vitro* approach is employed. It is essential to accurately identify those enzyme systems involved in metabolizing particular drugs and to evaluate the relative contribution of the metabolic pathways being inhibited to overall elimination of the drug. A significant drug-drug interaction occurs only when drugs compete for the same enzyme system and when the metabolic reaction is a major elimination pathway.

Another important factor is the use of clinically relevant concentrations of inhibitor and substrate. For competitive inhibition, the velocity of an enzymic reaction in the absence (V_o) and presence (V_i) of inhibitor can be expressed as:

$$V_o = \frac{V_{\max} \cdot S}{K_m + S} \quad [1]$$

$$V_i = \frac{V_{\max} \cdot S}{K_m \left(1 + \frac{I}{K_i} \right) + S}, \quad [2]$$

where V_{\max} is the maximum velocity, K_m is the Michaelis constant of the substrate, K_i is the inhibition constant of the inhibitor, and S and I are the substrate and inhibitor concentrations, respectively. By rearrangement of equations [1] and [2], the percent of inhibition

can be described as:

$$\frac{V_o - V_i}{V_o} (\%) = \frac{\frac{I}{K_i}}{1 + \frac{I}{K_i} + \frac{S}{K_m}} \times 100. \quad [3]$$

As shown in equation [3], the percent of inhibition is dependent on both the ratio of $[I/K_i]$ and $[S/K_m]$. Thus, an understanding of the relationship between substrate and inhibitor concentrations is critical to the design and interpretation of in vitro inhibition studies. Although the metabolic reactions of most drugs in their clinical dose range follow linear kinetics and the ratio $[S/K_m]$ can be neglected, there are several drugs, such as antiviral and anticancer drugs, for which the $[S/K_m]$ ratio can be very high in relation to the $[I/K_i]$ ratio.

As in the case of the terfenadine-ketoconazole interaction, an understanding of the mechanisms involved in drug interactions also is essential to provide a rational basis for interpreting and preventing adverse effects. Warfarin, an oral anticoagulant, exists in enantiomeric forms, in which the *S*-enantiomer of warfarin is much more potent than the *R*-enantiomer. As noted earlier, the more potent *S*-warfarin in humans is eliminated almost entirely as *S*-7-hydroxywarfarin, whereas *R*-warfarin is metabolized mainly as *R*-6-hydroxywarfarin (Lewis et al., 1974). Furthermore, these two hydroxylation reactions are mediated by different cytochrome P-450 isoforms (Kunze et al., 1996). Coadministration of enoxacin, a quinoline-azaquinoline antibiotic, resulted in a decrease in the clearance of *R*-warfarin but not in the clearance of *S*-warfarin. The decreased clearance of *R*-warfarin was found to be a consequence of inhibition by enoxacin on the (*R*)-6-hydroxywarfarin metabolic pathway. As expected, enoxacin did not affect the hypoprothrombinemic response produced by warfarin, because this antibiotic had no effect on *S*-warfarin elimination (Toon et al., 1987). Similarly, cimetidine inhibited human metabolism of *R*-warfarin while having little effect on that of *S*-warfarin (Somogyi and Gugler, 1982). Further studies in healthy subjects indicated that treatment with cimetidine resulted in a significant decrease in the formation of *R*-6- and *R*-7-hydroxywarfarin but had no effect on the formation of *S*-6- and *S*-7-hydroxywarfarin (Niopas et al., 1991). Because only *R*-warfarin metabolism is inhibited by cimetidine, and because *R*-warfarin is much less active, it is expected that cimetidine has little effect on the anticoagulant activity of warfarin.

The two examples above illustrate the importance of an understanding of the mechanisms for interpreting drug interactions and predicting their clinical consequences. Another good example is omeprazole-diazepam interactions. Omeprazole is a proton pump blocker used to treat peptic ulcers and reflux esophagitis. This drug is

metabolized mainly by CYP2C19 (Andersson et al., 1990). Diazepam, an antianxiety agent, also is metabolized predominantly by CYP2C19 (Andersson et al., 1990). The CYP2C19 isoform is known to be polymorphic; approximately 2 to 6% of Caucasians or 14 to 22% of Asians are found to be poor metabolizers (Wilkinson et al., 1992; Kalow and Bertilsson, 1994). Coadministration of omeprazole resulted in a significant decrease in plasma clearance of diazepam in extensive metabolizers (EMs) but had no effect on diazepam clearance in poor metabolizers (PMs) (Andersson et al., 1990). Because both omeprazole and diazepam are metabolized mainly by the same enzyme, CYP2C19, this explains why the two drugs interact in EMs but not in PMs. In PMs, there is no enzyme for which diazepam and omeprazole could compete. Similarly, coadministration of a quinidine CYP2D6 inhibitor has been shown to increase plasma concentrations of encainide (CYP2D6 substrate) in EMs but had little effect on the plasma concentrations in PMs (Turgeon et al., 1990).

Although it is easy to determine in vitro drug-drug interaction, the accurate interpretation and extrapolation of in vitro interaction data also require a good understanding of pharmacokinetic principles. If the elimination of a drug is mainly by the liver, the total clearance is approximately equal to the hepatic clearance (cL_H) that can be expressed as (Lin, 1995; Wilkinson, 1987):

$$cL_H = Q_h \cdot E = Q_h \cdot \left(\frac{f_p \cdot cL_{int}}{Q_h + f_p \cdot cL_{int}} \right), \quad [4]$$

where Q_h is the hepatic blood flow, E is the hepatic extraction, f_p is the free fraction in plasma, and cL_{int} , the intrinsic clearance, is a measure of the drug-metabolizing activity (V_{max}/K_m).

Kinetically, drugs can be classified by whether their hepatic clearance is "enzyme-limited" or "flow-limited" with an intermediate class (Wilkinson and Shand, 1975). When the cL_{int} of a drug is very small relative to the hepatic blood flow ($Q_h \gg f_p \cdot cL_{int}$), then the hepatic clearance is low, and the cL_H is directly related to f_p and cL_{int} as shown in equation [5]:

$$cL_H \cong f_p \cdot cL_{int}. \quad [5]$$

Thus, a decrease in the cL_{int} caused by metabolism-based drug interaction will result in an almost proportional decrease in the clearance of "low-clearance" drugs. On the other hand, if the cL_{int} is so high that $f_p \cdot cL_{int} \gg Q_h$, then the hepatic clearance is limited by the hepatic blood flow as shown in equation [6]:

$$cL_H \cong Q_h. \quad [6]$$

Thus, a decrease in the cL_{int} caused by drug interaction has little effect on the hepatic clearance of "high-clearance" drugs.

Because the hepatic first-pass effect reflects the hepatic cL_{int} , hepatic bioavailability (F) can be expressed as:

$$F = 1 - E = \frac{Q_h}{Q_h + f_p \cdot cL_{int}}, \quad [7]$$

and the area under the curve (AUC) after oral dosing can be described as:

$$AUC_{po} = \frac{F \cdot \text{dose}}{cL_H} = \frac{\text{dose}}{f_p \cdot cL_{int}}. \quad [8]$$

As shown in equation [8], a decrease in the cL_{int} caused by metabolism-based drug-drug interaction will yield an almost proportional increase in the AUC after oral dosing, regardless of whether it is a low- or high-clearance drug. In contrast, after intravenous administration, a decrease in the cL_{int} only affects the clearance and AUC of low-clearance drugs (fig. 6).

Indinavir (L-375,524, MK-639) is a high-clearance drug that is cleared rapidly with a clearance of 80 to 90 mL/min/kg in rats and 15 to 17 mL/min/kg in AIDS patients. These values are greater than rat hepatic blood flow (60–70 mL/min/kg) or close to human hepatic blood flow (20 mL/min/kg). In vitro studies with rat and human liver microsomes indicate that ketoconazole competitively inhibited the metabolism of indinavir with a K_i value of approximately 2.5 μM . Pretreatment of rats

with ketoconazole (25 mg/kg p.o.) had little inhibitory effect on the clearance of indinavir and its AUC after intravenous administration of indinavir. The clearance decreased from 87 mL/min/kg in control rats to 83 mL/min/kg in ketoconazole-pretreated rats. However, ketoconazole significantly increased the bioavailability of indinavir and its AUC after oral dosing. The bioavailability increased from approximately 20% in control rats to 89% in ketoconazole-pretreated rats (Lin, 1996b). Similarly, coadministration of ketoconazole (6 mg/kg p.o.) increased the AUC of indinavir in AIDS patients by approximately 62% after oral administration (McCrea et al., 1996).

On the other hand, ketoconazole is a low-clearance drug with a plasma clearance of 6 to 7 mL/min/kg in rats. In vitro studies with rat liver microsomes revealed that indinavir also competitively inhibited the metabolism of ketoconazole with a K_i value of 4.5 μM . As expected, pretreatment of rats with indinavir (20 mg/kg p.o.) significantly increased the AUCs of ketoconazole by two-fold after both intravenous and oral administration of ketoconazole (Lin, 1996b).

4. Prediction of in vivo metabolic clearance. One of the main objectives of in vitro metabolism studies is the quantitative prediction of in vivo drug metabolism from in vitro data. The prediction of metabolic clearance from in vitro systems is difficult and highly controversial. Some scientists believe that in vitro/in vivo extrapolation is possible, whereas others are less optimistic and believe that it is extremely difficult, if not impossible, to predict in vivo metabolism from in vitro metabolism data, especially in quantitative terms. Each group can cite examples from literature in support of their views (Sugiyama et al., 1989; Pang and Chiba, 1994; Houston, 1994; Gillette, 1984). Despite the difficulty of extrapolating in vitro data, we believe that quantitative in vitro metabolic data can be extrapolated reasonably well to in vivo situations with the application of appropriate pharmacokinetic principles.

There are many examples of good quantitative correlation between in vitro and in vivo drug metabolism. Ethoxybenzamide, an antipyretic agent, is exclusively metabolized to salicylamide by rat liver microsomes. The in vitro V_{max} and K_m values (3.46 $\mu\text{mol}/\text{min}/\text{kg}$ and 0.378 mM) are in good agreement with those obtained in vivo by application of a two-compartment model (3.77 $\mu\text{mol}/\text{min}/\text{kg}$ and 0.192 mM) (Lin et al., 1978). Indinavir (MK-639, L-735,524), a potent HIV protease inhibitor, exhibited marked species differences in hepatic clearance. This drug was metabolized mainly by isoforms of the CYP3A subfamily to form oxidative metabolites in all species examined (Lin et al., 1996a). The in vitro hepatic clearance of indinavir estimated from in vitro V_{max}/K_m values using liver microsomes from rats, dogs, and monkeys was in good agreement with the corresponding in vivo hepatic clearance values. The in vitro hepatic clearance of indinavir was 31, 25, and

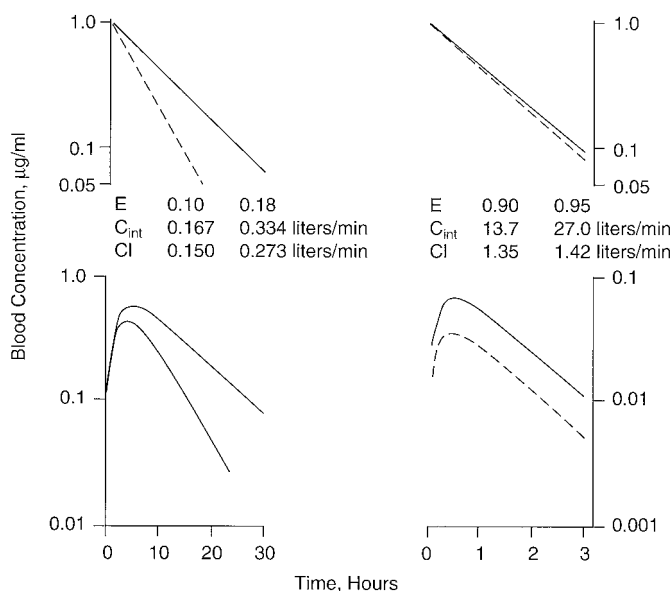


FIG. 6. Effect of cL_{int} on the disposition of low-clearance drugs (left panels) and high-clearance drugs (right panels) after intravenous (upper panels) and oral (lower panels) administration of equal doses of two totally metabolized drugs. Dotted line indicates normal situation; solid line indicates decreased cL_{int} . Reproduced with permission from Wilkinson (1987). Each reprint must be printed or stamped on the first page with authorship, title or article, name of journal, volume, Copyright date, The American Society for Pharmacology and Experimental Therapeutics. Reproduced by permission. Further reproduction is prohibited.

7.8 mL/min/kg for rats, monkeys, and dogs, respectively, and the corresponding *in vivo* hepatic clearance was 43, 36, and 11 mL/min/kg (Lin et al., 1996a). Chiba et al. (1990) have successfully predicted the steady-state concentration of imipramine and its active metabolite, desipramine, in rats using the V_{\max} and K_m values obtained from *in vitro* microsomal studies. Felodipine, a calcium channel blocker, is primarily metabolized to its pyridine analog in rats, dogs, and humans. The hepatic clearance of this drug obtained from *in vitro* studies with hepatic microsomes was 16 L/h for rat, 39 L/h for dog, and 259 L/h for humans and agreed reasonably well with those observed *in vivo*; the corresponding values were 6.2 L/h, 88 L/h, and 321 L/h (Bäärnhielm et al., 1986). Similarly, a good *in vitro* and *in vivo* correlation of the clearance of cytarabine hydrochloride has been reported by Dedrick et al. (1972). Furthermore, Iwatsubo et al. (1996) successfully predicted the *in vivo* clearance and bioavailability of YM796, a CNS drug for the treatment of Alzheimer's disease, using a recombinant system of human CYP3A4 together with the information of the content of this isoform in human liver microsomes (Shimada et al., 1994). Thus, these examples clearly show that *in vitro* to *in vivo* extrapolation is indeed possible if appropriate pharmacokinetic principles are employed.

However, the literature review revealed that in some cases, *in vitro* metabolic data failed to predict *in vivo* clearance. Sources of inaccuracy in predicting the *in vivo* metabolic clearance may include the nature and design of *in vitro* experiments, presence of extrahepatic metabolism, and active transport in liver. Unfortunately, the reason for the lack of *in vitro/in vivo* correlation has rarely been examined.

B. *In Vitro* Studies of Drug Absorption

Good absorption is one of the most important criteria in selecting new drug candidates for development. In the discovery stage, drug absorption studies can be performed only in laboratory animals and/or *in vitro* systems in an effort to characterize the absorptive process both qualitatively and quantitatively. Therefore, one must ask whether the *in vitro* models are useful in predicting drug absorption in humans or whether animal absorption data can be extrapolated to humans.

1. *Extrapolation of in vitro absorption data.* Numerous *in vitro* techniques have been developed for the study of drug absorption. These techniques include the use of everted intestinal sacs, everted intestinal rings, isolated brush border and basolateral membrane, and Ussing diffusion cells (Osiecka et al., 1985; Weiser, 1973; Windmueller and Spaeth, 1975; Grass and Sweetana, 1988). The limitations associated with these techniques often restrict their usefulness in the study of drug absorption. In 1989, Hidalgo and Borchardt introduced the Caco-2 cell monolayer model into the research field of drug absorption. During the last few years, the use of Caco-2 cells in the study of drug absorption has increased dra-

matically. The Caco-2 cell line is derived from a human colorectal carcinoma. It spontaneously differentiates into monolayers of polarized enterocytes under conventional cell culture conditions. After 2 to 3 weeks in cell culture, the monolayers have well-developed junctional complexes. Recently, a new cell line 2/4/A1, isolated from rat fetal intestinal epithelial cells, was used in studying drug absorption (Milovic et al., 1996).

Drugs pass through the intestinal lumen into the blood stream via two routes: (a) transcellularly, in which the drugs are transported actively or passively into and through epithelial cells into the blood circulation; and (b) paracellularly, in which drugs reach the blood circulation via the tight junctions between the epithelial cells. Because the surface area of the epithelial cell membrane is >1000-fold larger than the paracellular surface area (Pappenheimer, 1987), it is reasonably assumed that absorption of drugs via transcellular transport is always much better than that via paracellular transport. To date, most studies with Caco-2 cells are used to characterize whether a drug is actively or passively transported across the intestinal epithelium and to provide new insight into the regulation of drug transport. Bisphosphonates are poorly absorbed from the gastrointestinal lumen, and the bioavailability was approximately 0.7% for alendronate, 0.3% for pamidronate, and 1 to 2% for clodronate (Lin, 1996a). The poor absorption of bisphosphonates is speculated to be attributed to their very poor lipophilicity, preventing transcellular transport across the epithelial membrane, and therefore, the drugs must be absorbed via the paracellular route. Recently, *in vitro* studies with the Caco-2 cells have proven that two bisphosphonates (pamidronate and tiludronate) indeed are transported paracellularly (Boulenc et al., 1993; Twiss et al., 1994). Although it is believed that the paracellular permeability of hydrophilic compounds is inversely related to their molecular size (Chadwick et al., 1977), *in vitro* studies with the Caco-2 cell model show that, in addition to molecular size, flexibility of the drug's geometric structure is also an important factor in determining the permeation through the paracellular pathway (Artursson et al., 1993). Furthermore, the Caco-2 monolayer model was used to illustrate the influence of lipophilicity on the epithelial permeability of a series of β -blockers with similar $PK_{a,s}$ and molecular weights but different lipophilicities (Artursson, 1990). The Caco-2 cell model also has been employed by Conradi et al. (1991, 1992) to show that the permeability of peptides through the intestinal epithelial membrane is governed by hydrogen bond potential rather than lipophilicity. These examples demonstrate the usefulness of Caco-2 cells for determining the factors that influence drug absorption.

Although most studies of the Caco-2 cells are of a mechanistic nature, attempts have been made to predict quantitatively drug absorption in humans. In their classic study, Artursson and Karlsson (1991) have corre-

lated the epithelial permeability of 20 structurally unrelated drugs in Caco-2 monolayers with the extent of drug absorption in humans after oral administration. They concluded that drugs with complete (100%) absorption were found to have high permeability coefficients ($P_{app} \geq 1 \times 10^{-6}$ cm/s) in the Caco-2 cells, whereas poorly absorbed drugs had low permeability coefficients ($P_{app} < 1 \times 10^{-7}$ cm/s). However, in a similar correlation study, Rubas et al. (1993) reported that compounds with complete absorption in humans had Caco-2 permeability coefficients $>7 \times 10^{-5}$ cm/s, whereas compounds with poor absorption had permeability coefficients $<1 \times 10^{-5}$ cm/s. These values were approximately 70 to 100 times greater than those obtained by Artursson and Karlsson (1991). In a more recent study, Stewart et al. (1995) claimed that compounds completely absorbed in humans had Caco-2 permeability coefficients $>3 \times 10^{-5}$ cm/s. The reasons for this discrepancy in the reported permeability values between laboratories are not clear. Such results indicate that although the Caco-2 cell line is a useful model in ranking the permeabilities of drugs, it cannot be used quantitatively in predicting human absorption in vivo. Gan et al. (1993) reported that the Caco-2 permeability coefficient of ranitidine was 1.0×10^{-7} cm/s. If solely based on this in vitro value, one might predict poor absorption of ranitidine and throw away a billion-dollar drug. Actually, ranitidine has a good (50–70%) bioavailability in humans (Lin, 1991). Similarly, a poor Caco-2 permeability coefficient ($<1.0 \times 10^{-7}$ cm/s) was obtained for cimetidine, which is absorbed well in humans (personal communication with J. Hochman).

Attempts also have been made to compare in vitro and in vivo drug permeability. The permeabilities of a series of drugs were investigated in Caco-2 cells and in the human jejunum in situ using a double balloon technique (Lennernäs et al., 1996). Although the rank order of the permeability of these drugs was similar between the Caco-2 monolayers and the human jejunum, the permeability values of all drugs were much greater in the human jejunum than in the Caco-2 monolayers. The permeabilities of the drugs with complete absorption differed 2- to 4-fold between in vitro and in situ models, whereas the permeabilities of drugs with poor absorption differed as much as 30- to 80-fold. Thus, the permeability measured by Caco-2 cells can only be used for qualitative comparison, but not for quantitative purposes. Nevertheless, because drug transport studies in Caco-2 monolayers are easy to perform and require only small quantities of drugs, the Caco-2 cell monolayers can be used for screening of drug absorption (by ranking the permeability) at the early stages of drug discovery. Recently, Caco-2 monolayers were used to screen the permeability of a synthetic peptide library containing 375,000 compounds (Stevenson et al., 1995). Because the properties of Caco-2 monolayers can be varied with time in culture (Wilson et al., 1990), the passage number

(Walter and Kissel, 1995), and the cell culture medium (Jumarie and Malo, 1991), it is therefore important to include a reference drug for comparison purposes when screening the permeability of drugs.

2. *Extrapolation of animal absorption data.* In addition to the drug's permeability, many other factors, such as gastric and intestinal transit time and hepatic and intestinal metabolism, can influence the rate and extent of absorption. Because the in vitro models cannot provide quantitative prediction of drug absorption in humans, alternatively one can use animal absorption to predict human drug absorption. A rough estimate of human drug absorption from animal data is possible if species differences in the magnitude of first-pass metabolism can be assessed accurately. This is based on the assumption that the membrane permeability of drugs is similar across species. Membrane permeability is characterized as the relative magnitude of the interaction of the drug with the aqueous environment and lipophilic interior of the membrane, and is a function of the lipophilicity, molecular size, and PK_a of drugs (Ho et al., 1983). Because the nature of the biomembrane of the intestinal epithelial cells is similar across species, and because the main absorptive process (simple diffusion) is basically an interaction between the drug and the biomembrane (Wilson et al., 1989; Jackson, 1987), the permeability of a drug across the wall of the gastrointestinal tract is expected to be similar among species. There are numerous examples that support species similarity in the epithelial permeability. Amidon et al. (1988) successfully predicted the fraction of dose absorbed from the gastrointestinal tract in humans, using rat intestinal membrane permeability for a series of structurally unrelated compounds. Similarly, a good correlation between drug absorption rate constants in the human Caco-2 model and in a rat intestinal in situ model was obtained for a series of β -blocking agents (Artursson, 1990). In addition, the permeabilities of drugs that are transported by paracellular transport, due to their inability to cross the epithelial membranes, have been demonstrated to be similar among species. The paracellular permeability of a series of hydrophilic compounds obtained from human Caco-2 cells were quantitatively in good agreement with those from rat colon (Artursson et al., 1993).

Another key factor controlling drug absorption is first-pass metabolism. The oral bioavailability of a drug is defined as the fraction of an oral dose of the drug that reaches the systemic circulation. Because the entire blood supply of the upper gastrointestinal tract passes through the liver before reaching the systemic circulation, the drug may be metabolized by the liver and gut wall. Kinetically, the oral bioavailability (F) of a drug can be described as:

$$F = f_{abs} \cdot (1 - f_g) \cdot (1 - f_h), \quad [9]$$

where f_{abs} is the fraction of drug absorbed from the gastrointestinal lumen, and f_g and f_h are the fractions of drug metabolized by the gut wall and liver during the first passage of drug absorption (Lin, 1995). The f_{abs} of a drug is expected to be similar among species because it is determined mainly by its permeability. On the other hand, the f_g and f_h of a drug could be substantially different from one species to another.

Marked interspecies differences in the bioavailability of indinavir (MK-639, L-735,524) were observed when the drug was given orally as a solution in 0.05 M citric acid. The bioavailability varied from 72% in the dog to 19% in the monkey and 24% in the rat (Lin et al., 1996a). The low bioavailability observed in rats and monkeys was due to extensive hepatic first-pass metabolism. By comparing the drug concentration in the systemic circulation during portal or femoral vein infusion, hepatic first-pass extraction was estimated to be approximately 68% in rats. On the other hand, in situ studies with rat isolated intestinal loop preparation showed that intestinal first-pass metabolism was minimal (<8%). Consistent with in vivo and in situ studies, in vitro intestinal and hepatic first-pass extraction (f_g and f_h) were estimated to be 5 and 55%, respectively, for the rat using the intestinal and liver microsomal V_{max}/K_m data. Although in vivo hepatic first-pass extraction was not determined for the dog and monkey, the in vitro values were estimated to be 17% and 65%, respectively, using dog and monkey liver microsomes (Lin et al., 1996a). Taking the hepatic first-pass metabolism into account, the extent (f_{abs}) of indinavir absorbed from the gastrointestinal lumen was quite similar among species (~55 to 80%). Thus, observed species differences in the bioavailability of indinavir were due mainly to the differences in magnitude of hepatic first-pass metabolism. Using human intestinal and hepatic microsomes, the intestinal and hepatic first-pass metabolism of indinavir in humans were estimated to be 5 and 26%, respectively (Chiba et al., 1997). With the extent of absorption (55–80%) obtained from animal studies, we predicted the bioavailability of indinavir in patients would be 40 to 60%. As predicted, when clinical data became available, the bioavailability of indinavir was found to be approximately 60% (Yeh et al., unpublished data).

Another example that shows species similarity in drug absorption is L-365,260, a potent CCK_B receptor antagonist for the treatment of anxiety. The bioavailability of L-365,260 was 14% for the rat and 9% for the dog when given orally as a suspension in 0.5% methylcellulose (Lin et al., 1996c). The limited bioavailability was attributed mainly to poor absorption as a result of its low aqueous solubility (<2 $\mu\text{g}/\text{mL}$), because the hepatic first-pass metabolism was low and estimated to be 30% for the rat and 14% for the dog (Lin et al., 1996c).

When L-365,260 was given as a solution in PEG 600, the bioavailability increased to 50% in the rat and 70% in the dog. Taking hepatic first-pass metabolism into

consideration, the extent (f_{abs}) of L-365,260 absorbed from the gastrointestinal lumen was similar between rats and dogs (~70–80%). With this information at hand, L-365,260 was dosed in capsules containing PEG 600 in the subsequent clinical studies. As expected, the formulation gave good absorption of L-365,260 in healthy volunteers. The C_{max} and AUC were, respectively, 2.3 $\mu\text{g}/\text{mL}$ and 450 $\mu\text{g}\cdot\text{min}/\text{mL}$ for the dogs and 0.5 $\mu\text{g}/\text{mL}$ and 148 $\mu\text{g}\cdot\text{min}/\text{mL}$ for normal human subjects when the same dose (50 mg) of L-365,260 in polyethylene glycol (PEG) capsules was given orally to dogs (12 kg) and normal volunteers (70 kg) (Lin et al., unpublished data). The C_{max} and AUC values were comparable in dogs and humans when compared on a weight-normalized dose basis.

The examples of indinavir and L-365,260 suggest that drug absorption in humans can be extrapolated reasonably well from animal data when information on first-pass metabolism is also available. Indeed, Clark and Smith (1984) have reported in a survey that the fractions (f_{abs}) of dose absorbed from the gastrointestinal lumen for a large variety of drugs are remarkably consistent between animal species and humans. The bioavailability, however, differs substantially among species, presumably as a result of species differences in the magnitude of first-pass metabolism.

C. In Vitro Studies of Protein Binding

A basic tenet of biochemical pharmacology is that the intensity and duration of drug action is mediated via the time course of unbound drug concentrations at the site of action. Although direct measurement of unbound drug concentrations at the site of action is seldom possible, the unbound drug concentrations in plasma often bear a proportional relation, such that unbound drug concentrations in plasma can be used in lieu of site unbound concentrations. This assumption implies that drugs bind reversibly to plasma and tissue protein and that equilibrium of unbound drug occurs readily between plasma and tissues. Several reports are available to support this tenet that the unbound drug concentration correlates with pharmacological response and toxicity better than the total drug concentration (Yacobi and Levy, 1975; Yacobi et al., 1976; Mungall et al., 1984; Booker and Darcey, 1973; Rimmer et al., 1984; Huang and Øie, 1982; Øie and Chiang, 1991).

1. *In vitro/in vivo protein binding.* There are numerous in vitro methods for the determination of protein binding, including equilibrium dialysis, dynamic dialysis, ultrafiltration, ultracentrifugation, exclusion chromatography, and circular dichroism. The advantages and disadvantages of each method have been discussed, and the reliability of these methods was compared (Kurz et al., 1977; Kurz, 1986). It is concluded that equilibrium dialysis and ultrafiltration are most likely to provide both an accurate and precise assessment of plasma protein binding. Koike et al. (1985) compared an ultrafil-

tration technique with an equilibrium dialysis method for measuring the unbound phenytoin fraction in plasma in 36 patients with normal renal function and 6 uremic patients. The unbound concentrations of phenytoin determined by the ultrafiltration and equilibrium dialysis were essentially identical in both normal and uremic plasma obtained from patients under treatment.

Because the binding of drugs to plasma proteins is an important factor in determining their pharmacokinetics and pharmacological effects, plasma protein binding is routinely determined *in vitro* for drugs in discovery and development. The question is whether the *in vitro* binding data accurately reflects the *in vivo* binding.

The ratio of CSF drug concentration to plasma drug concentration has been used to determine *in vivo* drug binding. CSF is a very low-protein fluid, and therefore, drug in CSF is considered to be almost unbound. Chou and Levy (1981) demonstrated that the *in vitro* free fraction of phenytoin serum (0.155) obtained by equilibrium dialysis was essentially identical with the *in vivo* CSF:serum drug ratio (0.183). Similarly, Bertilsson et al. (1979) showed that the CSF:plasma ratio of demethylchlorimpiramine (0.026) was similar to the *in vitro* free fraction of the drug determined by ultrafiltration (0.035). These results suggest that *in vitro* plasma protein binding may accurately reflect *in vivo* binding. However, the CSF:plasma concentration ratio can only be viewed as an *in vivo* free fraction if there is no active transport involved in brain penetration. Enprofylline and theophylline have virtually identical *in vitro* free fractions in plasma (0.53 and 0.51, respectively) (Tegner et al., 1983). However, in a clinical study, the CSF:plasma ratios averaged 0.095 with enprofylline and 0.36 with theophylline (Laursen et al., 1989). The lower CSF levels of enprofylline than theophylline may be explained by the active transport of enprofylline, but not of theophylline, from CSF to blood.

Recently, microdialysis has been developed for measuring the unbound drug concentration in biological fluid. The use of microdialysis to determine the plasma protein binding of drugs was evaluated by comparing with ultrafiltration and equilibrium dialysis. Values of the free fraction of several drugs determined *in vitro* by microdialysis agreed very well with those by ultrafiltration and equilibrium dialysis (Herrera et al., 1990; Ekblom et al., 1992). The development of microdialysis technique provides the potential use of direct measurement of *in vivo* plasma protein binding. Recently, we used microdialysis to assess the *in vivo* plasma protein binding of warfarin, salicylate, and acetaminophen under steady-state conditions in conscious rats. Microdialysis probes were implanted in a jugular vein and continuously perfused with saline. The *in vivo* free fraction measured by microdialysis was 0.041 for warfarin, 0.185 for salicylate, and 0.76 for acetaminophen. These values correlated very well with the corresponding *in vitro* values determined by ultrafiltration (0.048, 0.192, and

0.62) (Wong and Lin, unpublished data). Similarly, microdialysis was performed *in vivo* to determine the plasma protein of the nonindolic melatonin analog S 20098 in rats under steady-state conditions, yielding similar free fraction values (0.26) to those obtained *in vitro* (0.24) (Quellec et al., 1994).

In view of the evidence presented above, it appears that the *in vitro* binding data determined by ultrafiltration and equilibrium dialysis accurately reflect the *in vivo* binding situations. However, care still must be exercised in determination of *in vitro* binding when the goal is to represent the *in vivo* situation. For example, in some cases, the metabolite of a drug may also bind to the plasma proteins and thus may be in competition with the parent drug for binding sites. Therefore, an *ex vivo* experiment in which plasma is taken from a species that has already received the drug may better reflect the *in vivo* binding situations. Dorzolamide (MK-507) is a good example. This drug is a potent carbonic anhydrase inhibitor used for the treatment of glaucoma. Carbonic anhydrase predominantly localized in red blood cells, accounting for >90% of the enzyme in the body. After administration of dorzolamide to the rat, a substantial fraction of the drug was converted to the *N*-demethylated metabolite, which is also a potent carbonic anhydrase inhibitor. Both dorzolamide and its *N*-demethylated metabolite bind extensively to erythrocytes. The rat erythrocyte:plasma concentration ratio of dorzolamide was approximately 200 when the drug was added to blood *in vitro* to yield a concentration of 30 μM . However, the ratio of dorzolamide was <10 at the same drug concentration when the ratio was determined *ex vivo* with the blood obtained from the rats that received a 25-mg/kg *i.v.* dose (Wong et al., 1996). The discrepancy between *in vitro* and *ex vivo* erythrocyte:plasma ratio is attributed mainly to the competitive binding interaction between dorzolamide and its *N*-demethylated metabolite. Similarly, competition in plasma protein binding between parent compound and its metabolite has been reported for sulfamethazine and its *N*-acetyl metabolite (du Souich and Babini, 1986).

2. *Plasma and tissue protein binding.* It is generally believed that only the unbound drug can diffuse across membranes that restrict distribution of a drug from the vascular compartment to the tissues and vice versa. Therefore, drug protein binding in plasma and tissues can affect the distribution of drugs in the body. Kinetically, the simplest quantitative expression relating the volume of distribution (V_d) to plasma and tissue binding (Lin, 1995) is given as:

$$V_d = V_p + \sum V_t \frac{f_p}{f_t}, \quad [10]$$

where V_p is the plasma volume, V_t is the tissue volume, and f_p and f_t are the fraction of unbound drug in plasma

and tissue, respectively. From this relationship, it is seen that the V_d increases when f_p is increased and decreases when f_t is increased.

Rearrangement of equation [10] yields:

$$V_f = \frac{V_d}{f_p} = \frac{V_p}{f_p} + \sum \frac{V_t}{f_t}, \quad [11]$$

where V_f is defined as the volume of distribution of unbound drugs. From this equation, it is clear that a change in f_t has a greater effect than f_p on V_f , because ΣV_t is much greater than V_p .

Although it is easy to determine the plasma protein binding of drugs, the study of tissue binding is hampered by methodological problems. Several methods have been developed for the study of tissue binding. These include perfused intact organs, tissue slices, or tissue homogenates. In principle, these methods allow the direct determination of tissue binding but require removal of tissues from the body, which limits their applicability. Furthermore, the necessary handling of tissues, such as of tissue slices and homogenization, may alter binding properties. The technical difficulties associated with determinations of drug binding to tissues are reflected by the very limited amount of published information on that subject (Fichtl et al., 1991).

Despite the technical difficulties, attempts have been made to extrapolate the in vitro tissue binding to that of in vivo. Assuming that the unbound drug concentration in tissues and plasma is equal at distribution equilibrium, the ratio (K_p) of drug concentration in tissue to that in plasma after drug administration is equivalent to the ratio of free fraction in plasma (f_p) to the free fraction in tissue (f_t). By applying this principle, Lin et al. (1982) showed a good agreement between the in vitro K_p values of ethoxybenzamide obtained from tissue homogenate binding and those from in vivo study of ethoxybenzamide in nine tissues of rats. Schuhmann et al. (1987) determined the K_p values for 11 drugs in muscle, liver, lungs, and kidneys of rabbits after constant rate infusion. For muscle tissue, a good agreement between the in vivo- and in vitro-calculated K_p values of the 11 drugs was observed, whereas in the other tissues (liver and lung), the in vivo and in vitro K_p values of some drugs were not in agreement. For example, the in vivo K_p values of quinidine, imipramine, and buphenine were 10- to 20-fold greater than the corresponding in vitro K_p values calculated from in vitro binding data. Similarly, major discrepancies between in vitro and in vivo K_p values for other drugs also were reported by other investigators (Igari et al., 1982; Harashima et al., 1984). These results suggest that in vivo binding of drugs to tissues may not be predicted readily by simple in vitro methods, because distribution of drugs in tissues may involve active uptake and secretion or metabolism processes.

As shown in equation [12], a drug's $t_{1/2}$ is directly related to its V_d . Therefore, it is very useful if one can predict the V_d in humans before its initial clinical studies. Unfortunately, it is difficult to predict the V_d on the basis of in vitro binding data, because the V_d is determined by both its plasma and tissue binding as indicated in equation [10] and because it is difficult to assess tissue binding. Alternatively, it is hoped that V_d of drugs in humans can be extrapolated from data of animals.

Fichtl et al. (1991) reported that there were striking species differences in plasma protein binding and V_d of propranolol. The values for the V_d varied by >20-fold, being lowest in monkeys and highest in rabbits. However, when the V_d was corrected for the f_p , the volume of distribution of unbound propranolol, V_f , was virtually the same for all species. Consistent with this, Sawada et al. (1984a) reported that the V_f s of 10 basic drugs were quite similar among species including humans. Based on these results, Fichtl et al. (1991) proposed that the V_f of drugs should be similar in humans and other species. Therefore, with knowledge of the V_f from laboratory animals and of f_p from human plasma protein determined in vitro, one can predict the V_d ($V_f \times f_p$) in humans before the initial clinical studies. Unfortunately, this approach is not valid for all drugs. Boxenbaum (1982) compared the pharmacokinetic parameters for 12 benzodiazapines in dogs and humans. Eight of the 12 benzodiazapines had quite different V_f values between the dog and human, the differences being as much as seven-fold for lorazepam. The large species differences in the V_f values also were reported for β -lactam antibiotics (Sawada et al., 1984b). Thus, the species similarity in the V_f of propranolol observed by Fichtl et al. (1991) could be fortuitous. In conclusion, these results suggest that the V_d of drugs in humans cannot be extrapolated from animal data.

3. Protein binding displacement interactions. Like metabolism-based competitive interactions, binding displacement interaction occurs when drugs compete for a common binding site of plasma proteins. Substantial drug displacement occurs when the displacing agent occupies a significant portion of the binding. Human plasma contains over 60 proteins. Albumin is the major component of plasma proteins responsible for the binding of most drugs in plasma. The concentration of albumin in normal subjects is approximately 650 μM (Lin et al., 1987b). The concentration of α_1 -acid glycoprotein can vary considerably in several physiological and pathological conditions. In healthy subjects, the concentrations of α_1 -acid glycoprotein ranged from 10 to 30 μM (Kremer et al., 1988). Although it is generally believed that basic drugs bind mainly to α_1 -acid glycoprotein and acid drugs bind to albumin, it has been shown that acid drugs bind to α_1 -acid glycoprotein, and basic drugs bind to albumin as well (Urien et al., 1986; Israili and El-Attar, 1983). Because of the high albumin concentration, a relatively high concentration of inhibitors (displacers) would be

required to displace the drug binding from the binding sites. Thus, *in vitro* studies designed to assess the possibility of *in vivo* binding displacement must use undiluted plasma and clinically relevant drug concentrations.

The use of supratherapeutic drug concentrations or unusually low protein concentrations may produce binding displacement *in vitro*, but not *in vivo*. Zini et al. (1979) showed that indomethacin markedly decreased warfarin binding to human serum albumin *in vitro* at an indomethacin concentration of 100 μM . However, Vesell et al. (1975) found no clinically significant displacement interaction between indomethacin and warfarin *in vivo* where the indomethacin concentration ranged from 0.08 to 1.0 μM . Bupivacaine caused a 109% increase in the free fraction of mepivacaine in a solution of α_1 -acid glycoprotein, but only a 9% increase in the free fraction of mepivacaine in plasma containing the same α_1 -acid glycoprotein concentration (Hartrick et al., 1984). Both bupivacaine and mepivacaine are highly bound to high-affinity and low-capacity α_1 -acid glycoprotein and low-affinity and high-capacity albumin in plasma.

Similar to metabolism-based drug interaction, the interpretation and extrapolation of *in vitro* displacement interaction data requires a good understanding of pharmacokinetic principles. Rowland and Aarons (Rowland, 1980; Aarons and Rowland, 1981) have reviewed the theoretical and clinically relevant issues regarding drug displacement interactions. Depending on whether it is a low- or high-clearance drug, displacement interaction will cause different alterations in pharmacokinetics. As shown in equation [10], changes in the free fraction (f_p) in plasma caused by displacement binding will affect drug distribution. As seen in equations [6] and [10], an increase in the f_p of high-clearance drugs caused by binding displacement interaction will have little change in the clearance (cL), but will lead to an increase in the volume of distribution (V_d); hence, the elimination $t_{1/2}$ will increase. The $t_{1/2}$ is related to both the cL and V_d as follows:

$$t_{1/2} = \frac{0.693 \times V_d}{cL}. \quad [12]$$

For low-clearance drugs, both cL and V_d will increase with an increase in f_p as shown in equations [5] and [10]. Although the changes in cL and V_d may not exactly balance, the $t_{1/2}$ will be affected to a much smaller degree compared with that of highly cleared drugs.

Because only unbound drug is responsible for pharmacological effect, it is important to make a clear distinction of the effects of displacement interaction on unbound and total drug concentrations in plasma. The simplest way of considering the effect of protein binding on the unbound and total drug concentration profiles is to examine the AUC. For low-clearance drugs, the AUC

of unbound and total drug after intravenous dosing can be expressed as:

$$AUC_{\text{total}} = \frac{\text{dose}}{cL} = \frac{\text{dose}}{f_p \cdot cL_{\text{int}}} \quad [13]$$

and

$$AUC_{\text{unbound}} = AUC_{\text{total}} \cdot f_p = \frac{\text{dose}}{cL_{\text{int}}}. \quad [14]$$

On the other hand, the AUC of unbound and total drug of high-clearance drugs after intravenous administration can be expressed as:

$$AUC_{\text{total}} = \frac{\text{dose}}{cL} = \frac{\text{dose}}{Q_h} \quad [15]$$

and

$$AUC_{\text{unbound}} = AUC_{\text{total}} \cdot f_p = \frac{\text{dose} \cdot f_p}{Q_h}. \quad [16]$$

From equations [13] and [14], it is evident that the AUC of unbound drug for low-clearance drugs is independent of any change in f_p if cL_{int} is unaffected by displacement interaction, whereas an increase in the f_p caused by binding displacement interactions will result in a decrease in the AUC of total drug. On the other hand, exactly the opposite situation occurs with a high-clearance drug in which the clearance and, hence, total drug concentration is unaffected by changes in plasma protein binding, whereas the unbound drug concentration increases as a result of increased f_p as shown in equations [15] and [16]. Figure 7 depicts the effects of displacement from protein binding sites on the steady-state unbound and total drug concentrations of low- and high-clearance drugs during intravenous infusion (Aarons, 1986).

After oral administration, the AUC of unbound and total drug, regardless of whether it is a high- or low-clearance drug, can be expressed as equation [17], which

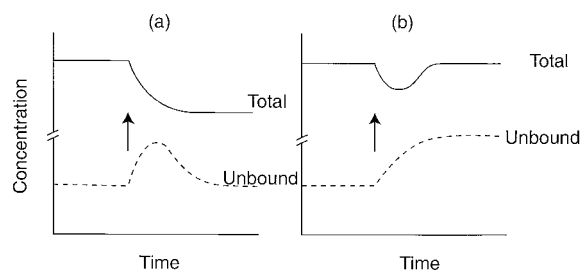


FIG. 7. The effect of displacing a low-clearance drug (a) or high-clearance drug (b), given chronically, from plasma protein binding sites. Displacement is produced by infusing a drug that displaces the first drug, starting from the arrowed point. Reproduced with permission from Aarons (1986).

is similar to equation [8]:

$$\text{AUC}_{\text{total}} = \frac{F \cdot \text{dose}}{cL} = \frac{\text{dose}}{f_p \cdot cL_{\text{int}}} \quad [17]$$

and

$$\text{AUC}_{\text{unbound}} = \text{AUC}_{\text{total}} \cdot f_p = \frac{\text{dose}}{cL_{\text{int}}} \quad [18]$$

From equations [17] and [18], the AUC of unbound drug after oral dosing is insensitive to the changes in the f_p , whereas the AUC of total drug will decrease when the f_p increases as a result of displacement interactions.

Because a significant change in the unbound AUC of drugs after oral dosing is not expected, and because most drugs are given orally, the displacement interactions rarely have significant clinical effects (Mackichan, 1984, 1989; Sellers, 1979). When changes in binding are associated with clinical effects, it has almost always been found that this is the result of a change in the cL_{int} caused by a mechanism quite independent of plasma protein binding as indicated in equation [18]. Warfarin-phenylbutazone interaction is a good example. When concomitantly administered with warfarin, phenylbutazone caused profound potentiation of a hypoprothrombinemic response (Sellers, 1986). Although phenylbutazone is known to displace warfarin from plasma proteins, it is clear from equation [18] that the hypoprothrombinemic effect was not caused by binding displacement of phenylbutazone, because the unbound concentration of warfarin should not be changed. Later, it was found that phenylbutazone stereoselectively inhibited the metabolism of *S*-warfarin (Lewis et al., 1974; O'Reilly et al., 1980). Thus, the metabolism inhibition, rather than binding displacement, causes the serious hemorrhagic complications of warfarin-phenylbutazone interaction. Similarly, although sulfaphenazole is known to displace tolbutamide from plasma proteins, the inhibitory effect of sulfaphenazole on the metabolism of tolbutamide is responsible for the serious hypoglycemic reactions (Christensen et al., 1963).

Whereas the unbound concentration after oral dosing is unaffected by displacement interaction, the transient increase in the unbound drug concentration occurring immediately after introduction of the displacing drug sometimes may be of clinical significance (Levy, 1976). Øie and Levy (1979a,b) reported that rapid intravenous infusion of salicylic acid or sulfisoxazole resulted in a transitory increase of unbound bilirubin concentration in rats. This suggests that the fatal kernicterus seen in the newborn after administration of sulfonamides may be due to a transitory increase in unbound bilirubin in the brain. In addition, the displacement interactions will be of clinical significance for high-clearance drugs after intravenous dosing. As shown in figure 7, a substantial increase in the unbound concentration may occur.

V. Interindividual Variability: A Critical Issue in Drug Development

From the market point of view, it is desirable that the dosage can be generalized to provide drugs for the treatment of a large number of patients. In reality, the generalization may work for most patients, but not for all. The standard dosage regimen of a drug may prove to be therapeutically effective in most patients, ineffective in some patients, and toxic in others. Variability in drug response becomes an important problem in drug therapy for drugs that have a narrow therapeutic window. Warfarin is a good example. There is a wide range of daily dose requirements (<2 mg–>11 mg) of warfarin needed to produce a similar prothrombin time (Koch-Weser, 1975). Variability in drug response can be broadly divided into pharmacokinetic and pharmacodynamic bases. Sources of pharmacokinetic variability include genetics, disease, age, and environmental factors (Breimer, 1983).

A. Pharmacokinetic Variability

The patient's exposure to drug is a crucial determinant of the drug's actions, and therefore its efficacy and safety. The term "drug exposure" is defined as the time course of the concentration of the drug and its active metabolites in plasma. The time course of drug concentration is governed by absorption, distribution, metabolism, and excretion. All these processes can contribute to pharmacokinetic variability.

1. *Variability in absorption.* Variation in absorption is one of the major sources of pharmacokinetic variability. An impression prevails that the degree of variability in the amount of drug reaching the systemic blood circulation is minimized if a drug has high bioavailability, whereas the risk of greater variation in the amount taken up is increased if a drug has low bioavailability. However, all too often, the degree of variability in absorption is similar for drugs of high and low bioavailability. The causes of absorption variability include pharmaceutical formulation, gastrointestinal physiology, and first-pass metabolism.

Being absorbed primarily from the upper part of the small intestine, oral absorption of drugs is often affected by the gastric emptying time and small intestinal motility, which vary considerably between individuals (Meyer, 1987; Weisbrodt, 1987). Usually, rapid gastric emptying results in rapid drug absorption. Changes in gastric emptying normally affect the rate of absorption but do not affect the amount of drug absorbed unless the drug is chemically unstable in the stomach or associated with saturable first-pass metabolism (Nimmo, 1976).

Dietary factors are also important sources of absorption variability that can be accounted for. The influence of food on the absorption of drugs is largely unpredictable. Food may enhance or reduce the absorption of some drugs while having no effect on others, depending not

only on the composition and volume of the meal or the drink, but also on the physicochemical properties of drugs. For example, absorption of the lipophilic drugs griseofulvin and sulfamethoxydiazine increased considerably when given with a high-fat meal (Crouse, 1961; Kaumeier, 1979). Amoxicillin, a poorly soluble antibiotic, was absorbed to a greater extent when swallowed with 250 mL water (Welling et al., 1977). In addition, dietary factors have been shown to alter drug-metabolizing enzyme activity, leading to changes in first-pass metabolism and bioavailability. Both charcoal-broiled beef and a high-protein, low-carbohydrate diet cause an increase in theophylline and antipyrine metabolism (Kappas et al., 1978, 1976). Certain vegetables, including brussels sprouts, cabbage, broccoli, and cauliflower, contain chemicals that induce drug-metabolizing enzyme activities (Pantuck, 1979). Because the diet is so different among patients, it is conceivable that the effects of food account for a substantial part of the absorption variability. Ironically, most clinical studies designed to address the question as to whether food intake affects drug absorption were conducted in healthy volunteers with or without a more or less standardized meal. Thus, such information may not be meaningful, sometimes even misleading.

The problem of absorption variability is complicated further by diseases. Hepatic disease may influence the oral bioavailability of drugs highly metabolized by the liver. The bioavailability of propranolol was increased significantly from 35% in normal subjects to 54% in cirrhotic patients, and the steady-state unbound propranolol concentration increased from 7.5 ng/mL to 22 ng/mL (Wood et al., 1978). The increased bioavailability was due mainly to a decrease in hepatic first-pass metabolism.

2. Variability in binding. As discussed earlier, plasma protein binding is an important determinant of the drug's disposition and actions. The f_p varies widely among drugs, and often (for highly bound drugs) among individuals. Differences in binding among drugs arise primarily from differences in their affinities for binding proteins, whereas differences in binding among individuals are due mainly to qualitative or quantitative differences in binding proteins. Nevertheless, interindividual variability in drug binding is generally less as compared with that in other pharmacokinetic processes such as absorption and metabolism (Yacobi and Levy, 1977; Barth et al., 1976).

α_1 -Acid glycoprotein is a major determinant for the binding of basic drugs in plasma (Piafsky and Borgå, 1977; Piafsky et al., 1978). Several inflammatory states (infections, rheumatic disorders, and surgical injury) and pathological conditions (myocardial infarction, malignancies, and nephritis) elevate the plasma concentration of α_1 -acid glycoprotein (Abramson, 1982; Freilich and Giardini, 1984). Furthermore, α_1 -acid glycoprotein is known to be inducible. Treatment with phenobarbital

resulted in a substantial increase in plasma concentration of α_1 -acid glycoprotein (Abramson, 1991). Because there is a strong correlation between the binding of basic drugs and the plasma levels of α_1 -acid glycoprotein (Lunde et al., 1986; Sjöqvist and Koike, 1986), an elevation of this protein will increase the binding of basic drugs.

In contrast to the elevation of α_1 -acid glycoprotein, hypoalbuminemia is always associated with a large variety of pathological conditions, including liver cirrhosis, renal failure, nephrotic syndrome, chronic inflammation, malignancies, and sepsis (Gugler and Jensen, 1986). In hypoalbuminemia, the binding of acidic drugs is reduced, and the decrease is related to a decrease in the plasma albumin concentration. Although normal subjects have a plasma albumin concentration of at least 35 mg/mL, plasma albumin concentrations can be as low as 10 mg/mL in patients with nephrotic syndrome.

In addition to the quantitative changes in plasma protein concentrations, qualitative structural changes of plasma proteins also alter the binding of drugs. High doses of acetylsalicylic acid can acetylate serum albumin and modify its binding sites (Hawkins et al., 1968). Cyanate, spontaneously formed from urea, carbamylates lysine residues on the albumin molecules and decreases the binding of acidic drugs in uremic patients (Erill et al., 1980). Furthermore, in uremic patients, retained endogenous acids that are highly protein bound can displace the binding of drugs from proteins. Collier et al. (1986) have identified one of these acids, 3-carboxy-4-methyl-5-propyl-2-furanpropanoic acid, as a potent displacer of drug binding. From these data, it is clear that disease states also are the main sources of binding variability.

Genetically determined variations in amino acid sequences of serum albumin and α_1 -acid glycoprotein also can contribute to binding variability. To date, more than 30 apparently different genetic variants of human serum albumin have been identified. Only approximately half of these variants have been absolutely characterized by peptide mapping and sequence determination (Eap and Baumann, 1991). Kragh-Hansen et al. (1990) have compared the binding affinities (association constants) of warfarin, salicylate, and diazepam to five variants of human serum albumin with known mutations. The association constants of all three drugs to albumin Canterbury (313 Lys→Asn) and to albumin Parkland (365 Asp→His) decreased substantially by a factor of 4- to 10-fold, whereas the binding affinity to albumin Verona (570 Glu→Lys) was unchanged. These results suggest that the region 313-365 seems to exert important effects on the binding of drugs, whereas the mutation 570 near the C-terminus does not affect drug binding.

Three main variants of α_1 -acid glycoprotein, namely ORM1 F1, ORM1 S, and ORM2 A, have been fully characterized (Eap and Baumann, 1991). Among the three variants, ORM2 A is the most important variant associ-

ated with the binding of basic drugs. Eap et al. (1990) have determined the *in vitro* binding of *d*-methadone, *L*-methadone and *dl*-methadone in plasma samples from 45 healthy subjects. The concentrations of α_1 -acid glycoprotein variants also were measured. Using multiple stepwise regression analysis, significant correlations were obtained between the binding of methadone and the total α_1 -acid glycoprotein or ORM2 A concentrations, but only a weak correlation between the binding and ORM1 S concentrations, and no correlation between the binding and ORM1 F1 concentrations were found. The frequencies for the three phenotypes, i.e., ORM1 F1/ORM2 A, ORM1 F1/ORM1 S/ORM2 A, and ORM1 S/ORM2 A were found to be 33.7, 50.5, and 15.2%, respectively, in a Swiss population (Eap et al., 1988). These results suggest genetically determined variations in α_1 -acid glycoprotein could be a major source of variability in the binding of basic drugs.

3. Variability in excretion. Although metabolism is the major route of elimination for most drugs, some drugs are excreted mainly as unchanged drug via the kidneys and liver. Both biliary and renal excretion correlate to their function. Ceftazidime, a cephalosporin antibiotic, is excreted mainly by the kidneys. The total clearance of ceftazidime correlated linearly with creatinine renal clearance in patients with varying degrees of renal function (Van Dalen et al., 1986). Similarly, a strong correlation should exist between the clearance and hepatic function if a drug is excreted mainly by the liver. The biliary excretion of indocyanine green correlated well with hepatic function in cirrhotic patients (Kawasaki et al., 1985).

Many endogenous organic acids are accumulated in the plasma of patients with renal dysfunction. These endogenous organic acids may inhibit the transport of certain drugs in the liver. The hepatic uptake and biliary excretion of bromosulfophthalein and dibromosulfophthalein are decreased in rats with acute renal failure (Silberstein et al., 1988). These data demonstrate that variations in hepatic and renal function, particularly in patients with hepatic and renal disorders, contribute significantly to pharmacokinetic variability.

Reabsorption is one of the important factors governing renal clearance of drugs. Lipophilic drugs tend to be extensively reabsorbed, whereas hydrophilic drugs do not. Urine flow and pH have a substantial effect on the renal clearance of a drug that is mostly reabsorbed. An increase in the urine flow will result in a decrease in reabsorption, leading to an increase in renal clearance. The renal clearance of theophylline increases with increasing urine flow rate (Tan-Liu et al., 1982). Similarly, the renal clearance of phenobarbital is also dependent on the urine flow rate (Linton et al., 1967).

Unlike plasma that has a narrow pH range of 7.3 to 7.5, urine pH ranges from 4.5 to 8.5. Thus, the urine pH is an additional factor that influences the reabsorption of drugs that are weak acids and bases. The renal ex-

cretion of salicylic acid is markedly pH-dependent. Renal excretion of salicylate increases more than ten-fold as the urinary pH increases from 5 to 8 (Macpherson et al., 1955). In contrast, the renal clearance of quinidine has been shown to diminish with increasing urinary pH (Gerhardt et al., 1969). Drugs that show pH-sensitive reabsorption also generally show flow-rate dependence. Clearly, variations in urine flow and pH also contribute significantly to excretion variability.

B. Pharmacogenetics of Drug Metabolism

All enzymes involved in the metabolism of drugs are regulated by genes and gene products. Because of evolutionary and environmental factors, there is a remarkable degree of genetic variability built into the population. Thus, the genetic factor represents an important source of interindividual variation in drug metabolism. Mutations in the gene for a drug-metabolizing enzyme result in enzyme variants with higher, lower, or no activity or may lead to a total absence of the enzyme. Therefore, it is not unusual to find a ten-fold or as much as a 50-fold difference in the rate of drug metabolism among patients.

With the technological breakthroughs in molecular biology, significant progress has been made in understanding the role of genetic polymorphisms in drug metabolism. The major polymorphisms that have clinical implications are those related to the oxidation of drugs by CYP2D6 and CYP2C19 (Meyer et al., 1990b, 1992; Meyer, 1994; Wilkinson et al., 1989; Broly and Meyer, 1993; Alvan et al., 1990), acetylation by *N*-acetyltransferase (Evans, 1992), and *S*-methylation by thiopurine methyltransferase (Weinshiboum, 1992; Creveling and Thakker, 1994). Individuals who inherit an impaired ability to catalyze one or more of these enzymatic reactions may be at an increased risk of concentration-related adverse effects and toxicity.

1. Polymorphism in drug oxidation. CYP2D6 polymorphism is perhaps the most studied genetic polymorphism in drug metabolism. Since its discovery in 1977 (Mahgoub et al., 1977), hundreds of studies have been carried out to investigate the nature of CYP2D6 polymorphism, the mode of inheritance, and the consequences of the deficient trait on drug disposition and pharmacological effects. This polymorphism divides the populations into two phenotypes: EM and PM. Approximately 5 to 10% of individuals in Caucasian populations are the PM phenotype, compared with only 1 to 2% of individuals in Asian populations. To date, more than 50 drugs, including antidepressants, antipsychotics, and cardiovascular drugs, are known to be catalyzed primarily by CYP2D6 (Parkinson, 1996).

Clinical studies have demonstrated that the PMs of CYP2D6-mediated drugs represent a high-risk group with a propensity to develop adverse effects. The disposition of haloperidol, a potent neuroleptic, was studied in a panel of six EMs and six PMs of debrisoquine (Llerena

et al., 1992). The PMs that received 4 mg of haloperidol developed neurological side effects, whereas at the same dose, the EMs experienced only mild side effects, such as tiredness, difficulty concentrating, and some restlessness. The PMs eliminated haloperidol significantly slower than the EMs, and the high plasma concentrations of haloperidol might, therefore, be associated with the side effects observed in the PMs. Similarly, an increased risk of side effects also was observed in the PMs of debrisoquine when taking other neuroleptics, such as perphenazine (Dahl-Puustinen et al., 1989) and thioridazine (Meyer et al., 1990a). Both drugs also are metabolized by CYP2D6.

Similarly, propafenone, a class I antiarrhythmic agent, is metabolized by CYP2D6. The relationship between debrisoquine phenotype and pharmacokinetics and pharmacodynamics of propafenone was studied in 28 patients (22 EMs and 6 PMs) with chronic ventricular arrhythmias (Siddoway et al., 1987). Steady-state concentrations of propafenone in plasma were found to be significantly higher in PMs than EMs. These higher concentrations were associated with a greater incidence of CNS side effects in the PMs (67%), relative to the EMs (14%).

The effects of CYP2D6 polymorphism on pharmacological responses can be quite complex, depending on whether the parent drug or metabolites, or both, are pharmacologically active. Encanide, a class I antiarrhythmic, is a good example. CYP2D6 *O*-demethylates encanide to a metabolite that is 6 to 10 times more potent than the parent drug in blocking sodium channels. In both PMs and EMs, standard doses of this drug tend to produce similar therapeutic responses, because relatively high parent drug concentrations in the former are matched by relatively high active metabolite concentrations in the latter (Buchert and Woosley, 1992). Similarly, both propafenone and its 5-hydroxy propafenone metabolite are pharmacologically active. The metabolism of propafenone to 5-hydroxy propafenone is grossly impaired in the PMs, resulting in very low or no levels of this active metabolite. However, as with encanide, there were no significant differences between EMs and PMs in an effective propafenone dosage or frequency of antiarrhythmic response (Siddoway et al., 1987). This again can be explained by the compensatory effect of the active metabolite of 5-hydroxy propafenone, present in the plasma of EMs but not in that of PMs.

Codeine is metabolized extensively by glucuronidation; the *O*-demethylation of codeine to morphine is a minor pathway that is mediated by CYP2D6 (Chen et al., 1988). As only a small fraction of the drug is metabolized by the *O*-demethylated pathway, PMs are not expected to have an altered disposition of codeine relative to EMs. As anticipated, plasma concentrations of codeine were similar in PMs and EMs, but measurable concentrations of morphine, its more analgesic *O*-demethylation product, were only detected in EMs (Sindrup

et al., 1991). Consequently, codeine increased the pain thresholds to copper vapor laser stimuli in EMs, but not in PMs, affirming the functional importance of the codeine-morphine biotransformation for codeine analgesia.

CYP2C19 also exhibits genetic polymorphism in drug metabolism. The incidence of the PM phenotype in populations of different racial origin varies; approximately 2 to 6% of individuals in the Caucasian populations are the PM phenotype, as are 14 to 22% in the Asian populations (Wilkinson et al., 1992; Kalow and Bertilsson, 1994). Although it is expected that PMs will have higher plasma concentrations of drugs metabolized by CYP2C19 than EMs and experience an increase in adverse effects, the clinical implications of CYP2C19 polymorphism have not been thoroughly characterized. Contrary to CYP2D6, CYP2C19 has been studied far less, which is reflected by the much shorter list of known drugs characterized by CYP2C19 than by CYP2D6 (Parkinson, 1996).

Diazepam is demethylated by CYP2C19 in humans (Anderson et al., 1990). The disposition of diazepam has been studied in 13 Caucasians of the EM phenotype and 3 Caucasians of the PM phenotype (Bertilsson et al., 1989). The plasma clearance of diazepam in the EMs was more than 2 times that in the PMs (11.0 and 5.0 mL/min, respectively), whereas the $t_{1/2}$ in the EMs was shorter than that in the PMs (59 and 128 h, respectively). The difference in the plasma clearance appeared to be related to formation of the desmethyl metabolite.

Omeprazole, a proton pump inhibitor, is metabolized (by CYP2C19) by hydroxylation and oxidation of the sulfoxide group to a sulfone (Anderson et al., 1990). The metabolism of omeprazole has been studied in the EMs and PMs of *S*-mephenytoin selected from phenotyped healthy Swedes and Chinese (Andersson et al., 1992). The plasma concentrations of omeprazole and its metabolites were determined after a single oral dose (20 mg). The AUC of omeprazole was substantially higher in PMs than in EMs in both Swedes (11.1 and 0.94 $\mu\text{M}\cdot\text{h}$) and Chinese (13.3 and 2.6 $\mu\text{M}\cdot\text{h}$). Although the AUC was not different between Swedish and Chinese PMs, there was a significant interethnic difference in EMs. The fact that the AUCs in Chinese EMs were 3 times higher than those of the Swedish EMs might be due to the higher proportion of heterozygotes in the Chinese.

From a genetic point of view, the different enzyme polymorphisms in drug metabolism are inherited independently. However, an inherited deficiency of different drug-metabolizing enzymes could occur simultaneously on the basis of probability. A population study of mephenytoin hydroxylation and debrisoquine hydroxylation was carried out in 221 unrelated normal volunteers (Küpfer and Preisig, 1984). Twelve (5%) of them exhibited defective hydroxylation of mephenytoin, and 23 (10%) could be identified as PMs of debrisoquine. Among these 35 subjects, 3 (1 female and 2 males) displayed

simultaneously both defects of mephenytoin and debrisoquine hydroxylation.

Propranolol is hydroxylated by CYP2D6 and *N*-dealkylated by CYP2C19. The relative contributions of these two isoforms to propranolol metabolism have been studied in a panel of phenotyped normal volunteers (Ward et al., 1989). Six subjects were EMs of both mephenytoin and debrisoquine. Four subjects were PMs of debrisoquine but rapid metabolizers of mephenytoin. Five subjects were PMs of mephenytoin but rapid metabolizers of debrisoquine, and one subject had a deficiency for both debrisoquine and mephenytoin. PMs of either mephenytoin or debrisoquine had a similar disposition of propranolol to that of EMs, whereas the subject with both mutations had a $t_{1/2}$ 2 times longer than the other subjects'.

In view of the examples presented above, it is clear that genetic polymorphism in drug metabolism could lead to clinically significant differences in pharmacokinetics and pharmacological responses of some patients and therefore might result in adverse effects or therapeutic failure. Thus, drugs metabolized by enzymes exhibiting genetic polymorphism are considered to be undesirable. However, the development of a drug sometimes is prematurely terminated based solely on the fact that its metabolism is polymorphic. To avoid premature termination, the clinical relevance of genetic polymorphism must be assessed carefully. Pharmacokinetic differences between phenotypes are most relevant for drugs with narrow therapeutic indices. For compounds with a variability of plasma concentrations outside the therapeutic range that is not associated with adverse effects, polymorphic metabolism will be of less or little concern. Propranolol is a typical example. Despite the critical involvement of CYP2D6 and CYP2C19 polymorphism in the metabolism of propranolol, this drug is quite safe clinically. Another important factor in determining the go/no-go decision is the overall benefit-to-risk ratio. If the benefit of a drug is significantly greater than its risk, and dosage can be titrated by direct clinical monitoring, then polymorphic metabolism is of less consequence.

2. *N*-Acetylation polymorphism. Acetylation is an important route of elimination for a large number of hydrazine and arylamine drugs (Weber et al., 1990). The *N*-acetyltransferase (NAT) polymorphism in humans was discovered as a result of studying the rate of isoniazid elimination in tuberculous patients in 1960 (Evans et al., 1960). The patients could be classified as slow and rapid acetylators based on their plasma concentrations of isoniazid. In addition to isoniazid, sulfamethazine, hydralazine, procainamide, dapsone, and nitrazepam also are polymorphically acetylated (Evans, 1992, 1989). The proportions of rapid and slow acetylators vary considerably between ethnic groups. For example, the percentage of slow acetylators in Egyptians and Mideasterners is 80 to 90%, whereas in Asian populations, it is

only 10 to 20%, with European and North American Caucasians having an intermediate value of 40 to 70% (Evans, 1989). On the other hand, other *N*-acetylated compounds, such as *p*-aminobenzoic acid and *p*-aminosalicylic acid, were unable to distinguish rapid and slow acetylators in vivo and in vitro (Evans, 1989). These compounds are, therefore, classified as monomorphic substrates.

Although the acetylation polymorphism was suspected for nearly 40 years, the molecular mechanics underlying this polymorphism were not known until recently. Meyer and his colleagues (Blum et al., 1990; Grant et al., 1991) have successfully cloned three human genes: NAT1, NAT2, and a related pseudogene, NATP. The discovery of two separate genes encoding NAT1 and NAT2 resolved the old question on monomorphic and polymorphic substrates. NAT2 has a high affinity for polymorphic substrates, whereas NAT1 has a high affinity for monomorphic substrates. Mutations of the NAT2 gene result in slow acetylation. The most common acetylator allele in Caucasians clearly is that with three mutations at positions 341, 481, and 803 (NAT2-B), followed by that with two mutations at positions 282 and 590 (NAT2-C) and that with two mutations at positions 282 and 287 (NAT2-D). These three alleles account for >95% of mutant alleles in Caucasian slow acetylators (Meyer et al., 1993; Lin et al., 1993a).

In general, slow acetylators are more susceptible to adverse effects than are rapid acetylators, because the *N*-acetylated drugs are not cleared from the body as well in slow acetylators. On the contrary, therapeutic effects may be suboptimal in rapid acetylators because of the rapid elimination of drugs. In a study of 744 pulmonary tuberculosis patients, there was a tendency for cavity closure and sputum conversion to occur significantly earlier in slow acetylators (Harris, 1961). However, the slow acetylators were more susceptible to hepatotoxicity (Mitchell et al., 1976). Furthermore, slow acetylators are more prone to develop systemic lupus erythematosus and rheumatoid arthritis (Lawson et al., 1979; Reindenberg and Martin, 1974).

Recently, the association of the acetylation morphism with an increased risk to develop certain cancers, e.g. bladder cancer or colorectal, has received much attention (Evans, 1992; Bock, 1992). It has been shown that the relative risk of developing bladder cancer in slow acetylators is 2 to 3 times that in rapid acetylators (Hassen et al., 1985). Consistent with this, the incidence of bladder cancer is low (6.3/100,000) in Japan, which has a low frequency of slow acetylator phenotype, approximately 11%, compared with the situation in the United States, where the incidence and frequency are 25.8/100,000 and 58%, respectively (Schultz, 1988). Similarly, the Japanese population exhibits a very low incidence of colorectal cancer (Connor et al., 1986). These data suggest that the *N*-acetylation phenotype is prob-

ably an important factor contributing to the multifactorial etiology of certain cancers.

Unlike the polymorphism of drug oxidation, neither slow nor rapid acetylation phenotype is rare in all ethnic groups. For example, most populations in Europe and North America have 40 to 70% slow acetylators and 30 to 60% rapid acetylators. Therefore, an important point to consider is the impact of polymorphic acetylation on the development of new drugs. In clinical trials, sufficient numbers of people should be studied to ensure that both the slow and rapid acetylation phenotypes are adequately represented. In some instances, it might be of value to phenotype patients to adjust dose regimens.

3. *S*-Methylation polymorphism. *S*-Methylation is an important metabolic pathway of many sulfhydryl drugs. Two enzymes, thio methyltransferase (TMT) and thio-purine methyltransferase (TPMT), are involved in the *S*-methylation. TPMT is a cytoplasmic enzyme that preferentially catalyzes the *S*-methylation of aromatic and heterocyclic sulfhydryl drugs, such as 6-mercaptopurine and azathioprine, whereas TMT is a membrane-bound enzyme and preferentially catalyzes the *S*-methylation of aliphatic sulfhydryl drugs, such as captopril and *D*-penicillamine (Weinshiboum, 1992; Creveling and Thakker, 1994).

Both TPMT and TMT are genetically polymorphic. In a study of 298 subjects, 88.6% had high erythrocyte TPMT activities, 11.1% had intermediate activities, and 0.3% had undetectable activity (Weinshiboum and Sladek, 1980). Although the TPMT activities in the red blood cells do not play a significant role in the *S*-methylation, the regulation of TPMT activity in the red blood cells reflects those in other tissues such as the kidney and liver (Woodson et al., 1982; Szumlanski et al., 1988). A significant correlation was found between myelosuppression in patients who were being treated with 6-mercaptopurine and azathioprine and low TPMT activities in their erythrocytes (Lennard et al., 1987, 1989). The patients with low TPMT activities had high blood levels of 6-thioguanine nucleotide (6-TGN) that may be incorporated into DNA. Both 6-mercaptopurine and its pro-drug azathioprine are catalyzed competitively by *S*-methylation and the metabolic pathway, leading to the formation of 6-TGN. Because of compensatory effects, the patients with low TPMT activities will have higher 6-TGN levels and be more susceptible to the risk of developing thiopurine-induced bone marrow suppression.

TMT also exhibits genetic polymorphism. The genetic frequencies for low and high activities were estimated to be approximately 60 and 40%, respectively (Price et al., 1989). It is believed that the genetic variability is related to interindividual differences in the *S*-methylation of aliphatic sulfhydryl drugs, such as captopril and *D*-penicillamine. Unlike TPMT, the clinical implications of TMT polymorphism have not been thoroughly characterized yet.

4. *Atypical butyrylcholinesterase.* Patients with genetic variants of butyrylcholinesterase exhibit prolonged paralysis after standard doses of neuromuscle blockers, such as succinylcholine, suxamethonium, and mivacurium, as a result of impaired ester hydrolysis (Lockridge, 1992; Bevan, 1993; Goudsouzian et al., 1993). The genetic variant most frequently found in patients who responded abnormally to the neuromuscular blockers is atypical butyrylcholinesterase, which occurs in homozygous form in 1 of 3500 Caucasians (Lockridge, 1992). By definition, the genetic allele that regulates the butyrylcholinesterase activities is not a common polymorphism but is a rare genetic variant. Several human enzymes may be involved in hydrolysis of ester drugs, including arylesterase and acetylcholinesterase. Genetic variants are known not only for butyrylcholinesterase, but also for arylesterase (La Du, 1992). No genetic variants are known for human acetylcholinesterase.

Although problems with the neuromuscular blockers are rare (<1% of patients), the prolonged muscle paralysis can be serious. The patients may be unable to breathe and have to be maintained on mechanical ventilators. Because butyrylcholinesterase is present in plasma and because the *in vitro* test procedures using dibucaine are relatively simple (Kalow and Genest, 1957), patients should be screened for their butyrylcholinesterase activity before being given the muscle relaxants. So far, no drug-induced toxicity was found to be related to the genetic variants of arylesterase.

As described above, genetic polymorphism in drug metabolism is undesirable and can at times be problematic. However, it should be emphasized that even if a large proportion of the metabolism of a compound is subject to genetic polymorphism, this should not influence its development as a drug. Careful evaluation of clinical relevance of the polymorphic metabolism has to be taken into consideration in making the go/no-go decisions.

VI. Conclusions

Drug research is an extremely complicated endeavor. It encompasses several diverse disciplines united by a common goal, namely the development of novel therapeutic agents. As described in this paper, pharmacokinetics and drug metabolism play an important role as determinants of *in vivo* drug action. Ideally, the process of rational drug design should provide a delicate balance between the chemistry, pharmacology, and pharmacokinetics of the drug. The discoveries of HIV protease inhibitors, indinavir and ritonavir, and the antifungal agent fluconazole are good examples of successfully incorporating pharmacokinetic and metabolic information into drug design.

Due to ethical constraints, relevant pharmacokinetic and metabolism studies must be carried out extensively in laboratory animals or *in vitro* systems before first drug administration in humans. Although these studies

provide useful information about absorption, distribution, metabolism, and excretion of the drug, extrapolation from *in vitro* and animal data to humans must be done cautiously. Marked species differences occur in the enzymatic systems involved in drug metabolism, whereas greater similarities are seen in physiological characteristics among different species. Therefore, it is of great importance that the underlying mechanisms responsible for these similarities and differences be examined carefully and weighted appropriately to ensure a reliable prediction from animal data to humans.

REFERENCES

- AARONS, L.: Kinetics of drug-drug interactions. *In* Pharmacokinetics: Theory and Methodology, ed. by M. Rowland and G. Tucker, pp. 163–186, Pergamon Press, New York, 1986.
- AARONS, L. J., AND ROWLAND, M.: Kinetics of drug displacement interactions. *J. Pharmacokinet. Biopharm.* **9**: 181–190, 1981.
- ABAS, A., AND MEFFIN, P. J.: Enantioselective disposition of 2-arylpropionic acid nonsteroidal anti-inflammatory drugs: IV—ketoprofen disposition. *J. Pharmacol. Exp. Ther.* **240**: 637–641, 1987.
- ABRAMSON, F. P.: Methadone plasma protein binding: alteration in cancer and displacement from α_1 -acid glycoprotein. *Clin. Pharmacol. Ther.* **32**: 652–658, 1982.
- ABRAMSON, F. P.: Species differences in the induction of α_1 -acid glycoprotein by drugs. *In* Plasma Binding of Drugs and Its Consequences, ed. by F. Belpaire, M. Bogaert, J. P. Tillement, and R. Verbeek, pp. 57–68, Academia Press, Ghent, Belgium, 1991.
- ADAMS, S. S., BOUGH, R. G., CLIFFE, E. E., DICKINSON, W., LESSEL, B., MCCULLOUGH, K. F., MILLS, R. F. N., NICHOLSON, J. S., AND WILLIAMS, G. A. H.: Some aspects of the pharmacology, metabolism and toxicology of ibuprofen. *Rheumatol. Phys. Med.* **10**(suppl.): 9–26, 1970.
- ADAMS, S. S., BRESLOFF, P., AND MANSON, C. G.: Pharmacological differences between the optical isomers of ibuprofen: evidence for metabolic inversion of the (–) isomer. *J. Pharm. Pharmacol.* **28**: 256–257, 1976.
- ADAMS, S. S., MCCULLOUGH, K. F., AND NICHOLSON, J. S.: The pharmacological properties of ibuprofen, an anti-inflammatory analgesic and antipyretic agent. *Arch. Int. Pharmacodyn. Ther.* **178**: 115–129, 1969.
- ALBERT, A.: Chemical aspects of selective toxicity. *Nature (Lond.)* **182**: 421–426, 1958.
- ALBERTS, A. W., CHEN, J., KURON, G., HUNT, V., HUFF, J., HOFFMAN, C., ROTHROCK, J., LOPEZ, M., JOSHUA, H., HARRIS, E., PATCHETT, A. A., MONAGHAN, R., CURRIE, S., STAPLEY, E., ALBERS-SCHONBERG, G., HENSENS, O., HIRSHFIELD, J., HOOGSTEEN, K., LIESCH, J., AND SPRINGER, J.: Mevinolin: a highly potent competitive inhibitor of hydroxymethylglutaryl-coenzyme A reductase and a cholesterol-lowering agent. *Proc. Natl. Acad. Sci. USA* **77**: 3957–3961, 1980.
- ALEXANDER, J., WALLIN, H., ROSSLAND, O. J., SOLBERG, K. E., HOLME, J. A., BECHER, G., ANDERSON, R., AND GRIVAS, S.: Formation of a glutathione conjugate and a semistable transportable glucuronide conjugate of N2-oxidized species of 2-amino-1-methyl-6-phenylimidazo[4,5-b]pyridine PhIP in rat liver. *Carcinogenesis* **12**: 2239–2245, 1991.
- ALVAN, G., BECHTEL, P., ISELIUS, L., AND GUNDELT-REMY, U.: Hydroxylation polymorphisms of debrisoquine and mephenytoin in European populations. *Eur. J. Clin. Pharmacol.* **39**: 533–537, 1990.
- AMIDON, G. L., SINKO, P. J., AND FLEISHER, D.: Estimating human oral fraction dose absorbed: a correlation using rat intestinal membrane permeability for passive and carrier-mediated compounds. *Pharm. Res.* **5**: 651–654, 1988.
- ANDERSSON, T., CEDERBERG, C., EDVARDSSON, G., HEGGELUND, A., AND LUNDBORG, P.: Effect of omeprazole treatment on diazepam plasma levels in slow versus normal rapid metabolizers of omeprazole. *Clin. Pharmacol. Ther.* **47**: 79–85, 1990.
- ANDERSSON, T., REGÄRDH, C. G., LOU, Y. C., ZHANG, Y., DAHL, M., AND BERTILSSON, L.: Polymorphic hydroxylation of *S*-mephenytoin and omeprazole metabolism in Caucasian and Chinese subjects. *Pharmacogenetics* **2**: 25–31, 1992.
- ARENDE, R. M., GREENBLATT, D. J., DEJONE, R. H., AND SELLERS, E. M.: Benzodiazepine entry into CSF and brain: kinetic, dynamic and *in vitro* correlations. *Clin. Pharmacol. Ther.* **33**: 239 (Abstract), 1983.
- ARIËNS, E. J.: Excursions in the field of SAR: a consideration of the past, the present and the future. *In* Biological Activity and Chemical Structure, ed. by J. A. Keeverling Buisman, pp. 1–35, Elsevier, Amsterdam, 1972.
- ARIËNS, E. J.: Stereochemistry, a basis for sophisticated nonsense in pharmacokinetics and clinical pharmacology. *Eur. J. Clin. Pharmacol.* **26**: 663–668, 1984.
- ARIËNS, E. J.: Racemic therapeutics: problems all along the line. *In* Chirality in Drug Design and Synthesis, ed. by C. Brown, pp. 29–43, Academic Press, New York, 1990.
- ARIËNS, E. J., AND SIMONIS, A. M.: Optimization of pharmacokinetics: an essential aspect of drug development—by “metabolic stabilization”. *In* Strategy in Drug Research, ed. by J. A. Keeverling Buisman, pp. 165–178, Elsevier, Amsterdam, 1982.
- ARROWSMITH, J. E., CAMPBELL, S. F., CROSS, P. E., STUBBS, J. K., BURGESS, R. A., GARDINER, D. G., AND BLACKBURN, K. J.: Long acting dihydropyridine calcium antagonists: I—2-Alkoxymethyl derivatives incorporating basic substituents. *J. Med. Chem.* **29**: 1696–1702, 1986.
- ARTURSSON, P.: Epithelial transport of drugs: I—a model for studying the transport of drugs (β -blocking agents) over an intestinal epithelial cell line (Caco-2). *J. Pharm. Sci.* **79**: 476–482, 1990.
- ARTURSSON, P., AND KARLSSON, J.: Correlation between drug absorption in humans and apparent drug permeability coefficients in human intestinal epithelial (Caco-2) cells. *Biochem. Biophys. Res. Commun.* **175**: 880–885, 1991.
- ARTURSSON, P., UNGELL, A.-L., AND LÖFROTH, J. E.: Selective paracellular permeability in two models of intestinal absorption: cultured monolayers of human intestinal epithelial cells and rat intestinal segments. *Pharm. Res.* **10**: 1123–1129, 1993.
- ATKINSON, A. B., BROWN, J. J., LEVER, A. F., McAREAVEY, D., ROBERTSON, J. I. S., BEHAN, P. O., MELVILLE, I. D., AND WEIR, A. I.: Neurological dysfunction in the two patients receiving captopril and cimetidine. *Lancet* **ii**: 36–37, 1980.
- ATKINSON, A. B., AND ROBERTSON, J. I. S.: Captopril in the treatment of clinical hypertension and cardiac failure. *Lancet* **ii**: 836–839, 1979.
- AUNST, B. J.: Novel formulation strategies for improving oral bioavailability of drug with poor membrane permeation or presystemic metabolism. *J. Pharm. Sci.* **82**: 979–987, 1993.
- BÄÄRNHJELM, C., DAHLBÄCK, H., AND SKANBERG, I.: *In vivo* pharmacokinetics of felodipine predicted from *in vitro* studies in rat, dog and man. *Acta Pharmacol. Toxicol.* **59**: 113–122, 1986.
- BÄÄRNHJELM, C., SKANBERG, I., AND BORG, K. O.: Cytochrome P-450-dependent oxidation of felodipine: a 1,4-dihydropyridine—to the corresponding pyridine. *Xenobiotica* **14**: 719–726, 1984.
- BAERTSCHI, S. W., RANEY, K. D., SHIMADA, T., HARRIS, T. M., AND GUENGERICH, F. P.: Comparison of rates of enzymatic oxidation of aflatoxin B1, aflatoxin G1 and sterigmatocystin and activation of the epoxides in forming guanyl-N7 adducts and inducing different gene responses. *Chem. Res. Toxicol.* **2**: 114–122, 1989.
- BAERTSCHI, S. W., RANEY, K. D., STONE, M. P. AND HARRIS, T. M.: Preparation of the 8,9-epoxide of the mycotoxin aflatoxin B1: the ultimate carcinogenic species. *J. Am. Chem. Soc.* **110**: 7929–7931, 1988.
- BAILLIE, T. A., AND DAVIS, M. R.: Mass spectrometry in the analysis of glutathione conjugates. *Biol. Mass Spectrom.* **22**: 319–325, 1993.
- BAILLIE, T. A., AND KASSAHUN, K.: Reversibility in glutathione-conjugate formation. *Adv. Pharmacol.* **27**: 163–181, 1994.
- BAILLIE, T. A., AND SLATTER, J. G.: Glutathione: a vehicle for the transport of chemically reactive metabolites *in vivo*. *Acc. Chem. Res.* **24**: 264–270, 1991.
- BALANI, S. K., ARISON, B. H., MATHAI, L., KAUFFMAN, L. R., MILLER, R. R., STEARNS, R. A., CHEN, I.-W., AND LIN, J. H.: Metabolites of L-735,524, a potent HIV-1 protease inhibitor, in human urine. *Drug Metab. Dispos.* **23**: 266–270, 1995.
- BALDESSARINI, R. J.: Drugs and the treatment of psychiatric disorders. *In* Goodman and Gilman's The Pharmacological Basis of Therapeutics, ed. by G. Gilman, L. S. Goodman, T. W. Rall, and I. Murad, pp. 383–435, Macmillan Publishing Co., New York, 1990.
- BANDIERA, S.: Expression and catalysis of sex-specific cytochrome P-450 isozymes in rat liver. *Can. J. Physiol. Pharmacol.* **68**: 762–768, 1990.
- BARTH, N., ALAN, G., BORGÅ, O., AND SjöQVIST, F.: Two-fold interindividual variation in plasma protein binding of phenytoin in patients with epilepsy. *Clin. Pharmacokinet.* **1**: 441–452, 1976.
- BELL, D. R., PLANT, N. J., RIDER, C. G., NA, L., BROWN, S., ATEITALLA, I., ACHARYA, S. K., DAVIES, M. H., ELIAS, E., AND JENKINS, R.: Species-specific induction of cytochrome P-450 4A RNAs: PCR cloning of partial guinea-pig, human and mouse CYP 4A cDNAs. *Biochem. J.* **294**: 173–180, 1993.
- BERESFORD, A. P.: CYP 1A1: friend or foe? *Drug Metab. Rev.* **25**: 503–517, 1993.
- BERRY, M. N., HALLS, H. J., AND GRIVELL, M. B.: Techniques for pharmacological and toxicological studies with isolated hepatocyte suspensions. *Life Sci.* **51**: 1–16, 1992.
- BERTILSSON, L., BRAITHWAITE, T., TYBRING, G., GARLE, M., AND BORGÅ, O.: Techniques for plasma protein binding of demethylchlorimipramine. *Clin. Pharmacol. Ther.* **26**: 265–271, 1979.
- BERTILSSON, L., HENTHORN, T. K., SANZ, E., TYBRING, G., SAWE, J., AND VILLÉN, T.: Importance of genetic factors in the regulation of diazepam metabolism: relationship to *S*-mephenytoin, but not debrisoquin hydroxylation phenotype. *Clin. Pharmacol. Ther.* **45**: 348–355, 1989.
- BEVAN, D. R.: Prolonged mivacurium-induced neuromuscular block. *Anesth. Analg.* **77**: 4–6, 1993.
- BICKEL, M. H.: Factors affecting the storage of drugs and other xenobiotics in adipose tissue. *In* Advances in Drug Research, ed. by B. Testa, and U. Meyer, pp. 56–86, Academic Press, New York, 1994.
- BLAINE E. H., FANELLI, G. M., JR., IRVIN, J. D., TOBERT, J. A., AND DAVIES, R. O.: Enantiomers of indacrinone: a new approach to producing an isouricemic diuretic. *Clin. Exp. Hypertens. [A]* **4**: 161–176, 1982.

- BLASCHKE, T. F.: Chromatographic resolution of racemates. *Angew. Chem. Int. Ed. Engl.* **19**: 13–24, 1980.
- BLASCHKE, T. F., KRAFT, H. P., FICKENTSCHER, K., AND KÖHLER, F.: Chromatographische Racemattrennung von thalidomid und teratogene Wirkung der enantiomere. *Drug Res.* **29**: 1640–1642, 1979.
- BLASCHKE, T. F., NIES, A. S., AND MAMELOK, R. D.: Principles of therapeutics. In Goodman and Gilman's The Pharmacological Basis of Therapeutics, ed. by A. F. Gilman, L. S. Goodman, T. W. Rall, and F. Murad, 7th ed., pp. 49–65, Macmillan Publishing Co., New York, 1985.
- BLUM, M., GRANT, D. M., MCBRIDE, W., HEIM, M. AND MEYER, U. A.: Human arylamine N-acetyltransferase genes: isolation, chromosomal localization and function expression. *DNA Cell Biol.* **9**: 193–203, 1990.
- BLUM, M. R., LIAO, S. H. T., GOOD, S. S., AND DEMIRANDA, P.: Pharmacokinetics and bioavailability of zidovudine in man. *Am. J. Med.* **85**: 189–196, 1988.
- BOCK, K. W.: Metabolic polymorphisms affecting activation of toxic and mutagenic arylamines. *Trends Pharmacol. Sci.* **13**: 223–226, 1992.
- BOCK, K. W., AND LILJENBLUM, W.: Roles of uridine diphosphate glucuronosyltransferase in chemical carcinogenesis. In *Conjugation-Deconjugation Reactions in Drug Metabolism and Toxicity*, ed. by F. C. Kauffman, pp. 391–428, Springer-Verlag, Berlin, Germany, 1994.
- BOCK, K. W., LIPP, H.-P., AND BOCK-HENNIG, B. S.: Induction of drug-metabolizing enzymes by xenobiotics. *Xenobiotica* **20**: 1101–1111, 1990.
- BODIN, N. O., EKSTRÖM, B., FORSGREN, U., JULAR, L. P., MAGNI, L., RAMSEY, C. H., AND SJÖBERG, B.: An orally well-absorbed derivative of ampicillin. *Antimicrob. Agents Chemother.* **8**: 518–525, 1975.
- BODOR, N.: Soft drugs: strategies for design of safer drugs. In *Strategy in Drug Research*, ed. by J. A. Keverling Buisman, pp. 137–164, Elsevier, Amsterdam, 1982.
- BODOR, N.: Novel approaches to the design of safer drugs: soft drugs and site-specific chemical delivery systems. In *Advances in Drug Research*, ed. by B. Testa, pp. 255–331, Academic Press, New York, 1984.
- BODOR, N., WOODS, R., RAPER, C., KEARNEY, P., AND KAMINSKI, J.: Soft drugs: 3—a new class of anticholinergic agents. *J. Med. Chem.* **23**: 474–480, 1980.
- BOOBIS, A. R., SESARDIC, D., MURRAY, B. P., EDWARDS, R. J., SINGLETON, A. M., RICH, K. J., MURRAY, S., DE LA TORRE, R., SEGURA, J., PELKONEN, O., PASANEN, M., KOBAYASHI, S., ZHI-GUANG, T., AND DAVIES, D. S.: Species variation in the response of the cytochrome P-450-dependent monooxygenase system to inducers and inhibitors. *Xenobiotica* **20**: 1139–1161, 1990.
- BOOKER, H. E., AND DARCEY, B.: Serum concentrations of free diphenylhydantoin and their relationships to clinical intoxication. *Epilepsia* **14**: 177–184, 1973.
- BOULENC, X., MARTI, E., JOYEUX, H., ROQUES, C., BERGER, Y., AND FABRE, G.: Importance of paracellular pathway for the transport of a new bisphosphonate using the human Caco-2 monolayers model. *Biochem. Pharmacol.* **46**: 1591–1600, 1993.
- BOXENBAUM, H.: Comparative pharmacokinetics of benzodiazepines in dog and man. *J. Pharmacokin. Biopharm.* **10**: 411–426, 1982.
- BOXENBAUM, H., NEAFSEY, P. J., AND FOURNIER, D. J.: Hormesis, Gompertz functions and risk assessment. *Drug Metab. Rev.* **9**: 195–229, 1988.
- BOYD, M. R.: Evidence for the Clara cell as a site of cytochrome P-450-dependent mixed-function oxidase activity in lung. *Nature (Lond.)* **269**: 713–717, 1977.
- BOYD, M. R., AND BURKA, L. T.: In vivo studies on the relationship between target organ alkylation and pulmonary toxicity of a chemically reactive metabolite of 4-ipomeanol. *J. Pharmacol. Exp. Ther.* **207**: 687–697, 1978.
- BRAY, K., AND QUAST, U.: Some degree of overlap exists between the K⁺-channels opened by cromakalim and those opened by minoxidil sulphate in rat isolated aorta. *Naunyn-Schmeideberg Arch. Pharmacol.* **344**: 351–359, 1991.
- BREIMER, D. D.: Interindividual variations in drug disposition: clinical implications and methods of investigation. *Clin. Pharmacokin.* **8**: 371–377, 1983.
- BRENDEL, K., FISHER, R. L., KRUMDIECK, C. L., AND GANDOLFI, A. J.: Precision-cut rat liver slices in dynamic organ culture for structure-toxicity studies. *J. Am. Coll. Toxicol.* **9**: 621–627, 1990.
- BROLY, F., AND MEYER, U. A.: Debrisoquine oxidation polymorphism: phenotypic consequences of a 3-base-pair deletion in exon 5 of the CYP2D6 gene. *Pharmacogenetics* **3**: 123–130, 1993.
- BUCHERT, E., AND WOOSLEY, R. L.: Clinical implications of variable antiarrhythmic drug metabolism. *Pharmacogenetics* **2**: 2–11, 1992.
- BURCHELL, B., NEBERT, D. W., NELSON, D. R., BOCK, K. W., IYANAGI, T., JANSEN, P. L. M., LANCET, D., MULDER, G. J., CHOWDHURY, J. R., SIEST, G., TEPHLY, T. R., AND MACKENZIE, P. I.: The UDP-glucuronosyltransferase gene superfamily: suggested nomenclature based on evolutionary divergence. *DNA Cell Biol.* **10**: 487–494, 1991.
- BUREK, D. J., NITSCHKE, K. D., BELL, T. J., WACKERLE, D. L., CHILDS, R. C., BEYER, J. E., DITTENBER, D. A., RAMPY, L. W., AND MCKENNA, M. J.: Methylene chloride: a two year inhalation toxicity and oncogenicity study in rats and hamsters. *Fundam. Appl. Toxicol.* **4**: 30–47, 1984.
- BÜRKE, H., DUNBAR, S., AND VAN AKEN, H.: Remifentanyl: a novel, short-acting, μ -opioid. *Anesth. Analg.* **83**: 646–651, 1996.
- CACCIA, S., AND GARATTINI, S.: Formation of active metabolites of psychotropic drugs: an updated review of their significance. *Clin. Pharmacokin.* **18**: 434–459, 1990.
- CALDWELL, J., WEIL, A., AND TANAKA, Y.: Species differences in xenobiotic conjugation. In *Xenobiotic Metabolism and Disposition*, ed. by R. Kato, R. W. Estabrook, and M. N. Cayen, pp. 217–224, Taylor & Francis, London, UK, 1989.
- CAMPBELL, D. B.: Stereoselectivity in clinical pharmacokinetics and drug development. *Eur. J. Drug Metab. Pharmacokin.* **15**: 109–125, 1990.
- CASHMAN, J. R., YOUNG, Z., YANG, L., AND WRIGHTON, S. A.: Stereo- and regioselective N- and S-oxidation of tertiary amines and sulfides in the presence of adult human liver microsomes. *Drug Metab. Dispos.* **21**: 492–501, 1993.
- CHADWICK, V. S., PHILLIPS, S. F., AND HOFMANN, A. F.: Measurements of intestinal permeability using low molecular weight polyethylene glycols (PEG 400): II—application to normal and abnormal permeability states in man and animals. *Gastroenterology* **73**: 247–251, 1977.
- CHAPMAN, D. E., CHRISTENSEN, T. A., MICHENER, S. R., AND POWIS, G.: Xenobiotic metabolism studies with human liver. In *Human Drug Metabolism: From Molecular Biology to Man*, ed. by E. H. Jeffery, pp. 53–63, CRC Press, Boca Raton, 1993.
- CHEN, Z. R., SOMOGYI, A. A., AND BOCHNER, F.: Polymorphic O-demethylation of codeine. *Lancet* **ii**: 914–915, 1988.
- CHIBA, M., FUJITA, S., AND SUZUKI, T.: Pharmacokinetic correlation between in vitro hepatic microsomal enzyme kinetics and in vivo metabolism of imipramine and desipramine in rats. *J. Pharm. Sci.* **79**: 281–287, 1990.
- CHIBA, M., HENSLEIGH, M., AND LIN, J. H.: Hepatic and intestinal metabolism of indinavir, a potent HIV protease inhibitor, in rat and human microsomes. *Biochem. Pharmacol.* **53**: 1187–1195, 1997.
- CHIBA, M., HENSLEIGH, M., NISHIME, J. A., BALANI, S. K., AND LIN, J. H.: Role of cytochrome P-450 3A4 in human metabolism of MK-639, a potent human immunodeficiency virus protease inhibitor. *Drug Metab. Dispos.* **24**: 307–314, 1996.
- CHIKHALE, E. G., NG, K.-Y., BURTON, P. S., AND BORCHARDT, R. T.: Hydrogen bonding potential as a determinant of the in vitro and in situ blood-brain barrier permeability of peptides. *Pharm. Res.* **11**: 412–419, 1994.
- CHINJE, E., KENTISH, P., JARNOT, B., GEORGE, M., AND GIBSON, G.: Induction of the CYP 4A subfamily by perfluorodecanoic acid: the rat and the guinea pig as susceptible and non-susceptible species. *Toxicol. Lett.* **71**: 69–75, 1994.
- CHIU, S.-H. L.: The use of in vitro metabolism studies in the understanding of new drugs. *J. Pharmacol. Toxicol. Methods* **29**: 77–83, 1993.
- CHOU, R. C., AND LEVY, G.: Effect of heparin or salicylate infusion on serum protein binding and on concentrations of phenytoin in serum, brain and cerebrospinal fluid of rats. *J. Pharmacol. Exp. Ther.* **210**: 42–48, 1981.
- CHRISTENSEN, L. K., HANSEN, J. M., AND KRISTENSEN, M.: Sulfaphenazole-induced hypoglycemic attacks in tolbutamide-treated diabetics. *Lancet* **ii**: 1298–1301, 1963.
- CHRISTIAN, M. C., WITTES, R. E., LEYLAND-JONES, B., MCLEMORE, T. L., SMITH, A. C., GRIESHABER, C. K., CHABNER, B. A., AND BOYD, M. R.: 4-Ipomeanol: a novel investigational new drug for lung cancer. *J. Natl. Cancer Inst.* **81**: 1155–1159, 1989.
- CLARK, B., AND SMITH, D. A.: Pharmacokinetics and toxicity testing. *Crit. Rev. Toxicol.* **12**: 343–385, 1984.
- CLARKE, D. J., AND BURCHELL, B.: The uridine diphosphate glucuronosyltransferase multigene family: function and regulation. In *Conjugation-Deconjugation Reactions in Drug Metabolism and Toxicity*, ed. by F. C. Kauffman, pp. 3–43, Springer-Verlag, Berlin, Germany, 1994.
- CLAYTON, J. P., COLE, M., ELSON, S. W., AND FERRES, H.: BRL 8988 (talampicillin): a well absorbed oral form of apicillin. *Antimicrob. Agents Chemother.* **5**: 670–671, 1974.
- COLLIER, R., LINDUP, W. E., LIEBICH, H. M., AND SPITTELER, G.: Inhibitory effect of the uremic metabolite 3-carboxy-4-methyl-5-propyl-2-furanpropionic acid on plasma protein binding. *Br. J. Pharmacol.* **21**: 610P–611P, 1986.
- COMBALBERT, J., FABRE, I., FABRE, G., DALET, I., AND DERANCOURT, J.: Metabolism of cyclosporin A: IV—purification and identification of the rifampicin-inducible human liver cytochrome P-450 (Cyclosporin A oxidase) as a product of P-450 IIIA gene subfamily. *Drug Metab. Dispos.* **17**: 197–207, 1989.
- CONNEY, A. H., MILLER, E. C., AND MILLER, J. A.: The metabolism of methylated aminoazo dyes: V—evidence for induction of enzyme systems in the rat by 3-methyl-cholanthrene. *Cancer Res.* **16**: 450–459, 1956.
- CONNOR, A., ALTORKI, N., AND MOOSSA, A. R.: Tumors of colon, rectum and anus: clinical features and surgical treatment. In *Comprehensive Textbook of Oncology*, ed. by A. R. Moossa, M. C. Robson, and S. C. Schimpff, pp. 1063–1079, Williams and Wilkins, Baltimore, 1986.
- CONRADI, R. A., HILGERS, A. R., HO, N. F. H., AND BURTON, P. S.: The influence of peptide structure on transport across Caco-2 cells. *Pharm. Res.* **8**: 1453–1460, 1991.
- CONRADI, R. A., HILGERS, A. R., HO, N. F. H., AND BURTON, P. S.: The influence of peptide structure on transport across Caco-2 cells: II—peptide bond modification which results in improved permeability. *Pharm. Res.* **9**: 435–439, 1992.
- COOK, C. S., KARIM, A., AND SOLLMAN, P.: Stereoselectivity in the metabolism of disopyramide enantiomers in rat and dog. *Drug Metab. Dispos.* **10**: 116–121, 1982.
- COUGLETRE, M. W. H.: Role of molecular biology in the structural and functional characterization of the UDP-glucuronosyltransferases. In *Progress in Drug Metabolism*, ed. by G. G. Gibson, pp. 35–72, Taylor & Francis, London, UK, 1992.

- COX, P. J., FARMER, P. B., FOSTER, A. B., GILBY, E. D., AND JARMAN, M.: The use of deuterated analogs in qualitative and quantitative investigations of the metabolism of cyclophosphamide (NSC-26271). *Cancer Treat. Rep.* **6**: 483-491, 1976a.
- COX, P. J., FARMER, P. B., JARMAN, M., JONES, M., STEC, W. J., AND KINAS, R.: Observations on the differential metabolism and biological activity of the optical isomers of cyclophosphamide. *Biochem. Pharmacol.* **25**: 993-996, 1976b.
- CREVELING, C. R., AND THAKKER, D. R.: O-, N- and S-methyltransferase. In *Conjugation-Deconjugation Reactions in Drug Metabolism and Toxicity*, ed. by F. C. Kauffman, pp. 189-216, Springer-Verlag, Berlin, Germany, 1994.
- CROUSE, R. G.: Human pharmacology of griseofulvin: the effect of fat intake on gastrointestinal absorption. *J. Invest. Dermatol.* **37**: 529-532, 1961.
- DAHL-PUUSTINEN, M.-L., LIDEN, A., ALM, C., NORDIN, C., AND BERTILSSON, L.: Disposition of perphenazine is related to polymorphic debrisoquine hydroxylation in human beings. *Clin. Pharmacol. Ther.* **46**: 78-81, 1989.
- DEDRICK, R. L., FORRESTER, D. D., AND HO, D. H. W.: In vitro-in vivo correlation of drug metabolism: deamination of 1- β -D-arabino-furano-sylcytosine. *Biochem. Pharmacol.* **21**: 1-16, 1972.
- DEVEREUX, T. R., JONES, K. G., BEND, J. R., FOUTS, Y. R., STATHAM, C. N., AND BOYD, M. R.: In vitro metabolic activation of pulmonary toxin 4-ipomeanol, in nonciliated bronchiolar epithelia (Clara) and alveolar type II cells isolated from rabbit lung. *J. Pharmacol. Exp. Ther.* **220**: 223-227, 1982.
- DAZ, D., FABRE, I., DAUJAT, M., SAINT AUBERT, B., BORIES, P., MICHEL, H., AND MAUREL, P.: Omeprazole is an aryl hydrocarbon-like inducer of human hepatic cytochrome P-450. *Gastroenterology* **99**: 737-747, 1990.
- DI GIOVANNA, J., BERRY, D. L., JUCHAU, M. R., AND SLAGA, T. J.: 2,3,7,8-tetrachlorodibenzo-p-dioxin: potent anticarcinogenic activity in CD-1 mice. *Biochem. Biophys. Res. Commun.* **86**: 577-584, 1979.
- DOGTEROM, P.: Development of a simple incubation system for metabolism studies with precision-cut liver slices. *Drug Metab. Dispos.* **21**: 699-704, 1993.
- DOGTEROM, P., AND ROTHUIZEN, J.: A species comparison of tolbutamide metabolism in precision-cut liver slices from rats and dogs: qualitative and quantitative sex differences. *Drug Metab. Dispos.* **21**: 705-709, 1993.
- DÖHLER, K. D., WONG, C. C., AND VON ZUR MÜHLEN, A.: The rat as model for the study of drug effects on thyroid function: consideration of methodological problems. *Pharmacol. Ther.* **5**: 305-318, 1979.
- DORSEY, B. D., LEVIN, R. B., MCDANIEL, S. L., VACCA, J. P., GUARE, J. P., DARKE, P. L., ZUGAY, J. A., EMINI, E. A., SCHLEIF, W. A., QUINTERO, J. C., LIN, J. H., CHEN, I.-W., HOLLOWAY, M. K., FITZGERALD, P. M. D., AXEL, M. G., OSTOVIC, D., ANDERSON, P. S., AND HUFF, J. R.: L735 524: the design of a potent and orally bioavailable HIV protease inhibitor. *J. Med. Chem.* **37**: 3443-3451, 1994.
- DOWNES, H., PERRY, R. S., OSTLUND, R. E., AND KARLER, R.: A study of the excitatory effects of barbiturates. *J. Pharmacol. Exp. Ther.* **175**: 692-699, 1970.
- DOWNES, H., AND WILLIAMS, J. K.: Effects of a convulsant barbiturate on the spinal monosynaptic pathway. *J. Pharmacol. Exp. Ther.* **168**: 283-289, 1969.
- DUGGAN, D. E.: Sulindac: therapeutic implications of the prodrug/pharmacophore equilibrium. *Drug Metab. Rev.* **12**: 325-337, 1981.
- DUGGAN, D. E., HOOKE, K. F., NOLL, R. M., AND KWAN, K. C.: Enterohepatic circulation of indomethacin and its role in intestinal irritation. *Biochem. Pharmacol.* **24**: 1749-1754, 1975.
- DU SOUCH, P., AND BABINI, R.: Drug-plasma protein binding and apparent volume of distribution: a complex relationship. In *Drug-Protein Binding*, ed. by M. M. Reidenberg, and S. Erill, pp. 70-92, Praeger, New York, 1986.
- EAP, C. B., AND BAUMANN, P.: Pharmacogenetics of drug binding to albumin and α_1 -acid glycoprotein. In *Plasma Binding of Drugs and Its Consequences*, ed. by F. Belpaire, M. Bogaert, J. P. Tillement, and R. Verbeek, pp. 69-82, Academia Press, Ghent, Belgium, 1991.
- EAP, C. B., CUENDET, C., AND BAUMANN, P.: Orosomucoid (α_1 -acid glycoprotein) phenotyping by use of immobilized pH gradients with 8 M urea and immunoblotting: a new variant encountered in a population study. *Hum. Genet.* **80**: 183-185, 1988.
- EAP, C. B., CUENDET, C., AND BAUMANN, P.: Binding of *d*-methadone, *L*-methadone and *dl*-methadone to proteins in plasma of healthy volunteers: role of the variants of α_1 -acid glycoprotein. *Clin. Pharmacol. Ther.* **47**: 338-346, 1990.
- EKBLOM, M., HAMMARLUND-UDENAES, M., LUNDGRIST, T., AND SJOBERG, P.: Potential use of microdialysis in pharmacokinetics: a protein binding study. *Pharm. Res.* **9**: 155-158, 1992.
- EL MOUËLHI, M., RUELIUS, H. W., FENSELAU, C., AND DULIK, D. M.: Species-dependent enantioselective glucuronidation of three 2-arylpropionic acids: naproxen, ibuprofen and benoxaprofen. *Drug Metab. Dispos.* **15**: 767-772, 1987.
- ELSON, C. E., MALTZMAN, T. H., BOSTON, J. L., TANNER, M. A., AND GOULD, M. N.: Anticarcinogenic activity of *d*-limonene during the initiation and promotion/progression stages of DMBA-induced rat mammary carcinogenesis. *Carcinogenesis* **9**: 331-332, 1988.
- ERILL, S. R., CALVO, R., AND CARLOS, R.: Plasma protein carbamylation and decreased acidic drug protein binding in uremia. *Clin. Pharmacol. Ther.* **27**: 612-618, 1980.
- EVANS, D. A. P.: *N*-Acetyltransferase. *Pharmacol. Ther.* **42**: 157-234, 1989.
- EVANS, D. A. P.: *N*-acetyltransferase. In *Pharmacogenetics of Drug Metabolism*, ed. by W. Kalow, pp. 95-178, Pergamon Press, New York, 1992.
- EVANS, D. A. P., MANLEY, K. A., AND MCKUSICK, V. A.: Genetic control of isoniazid metabolism in man. *Br. Med. J.* **2**: 485-491, 1960.
- FABRO, S., SMITH, R. L., AND WILLIAMS, R. T.: Toxicity and teratogenicity of optical isomers of thalidomide. (Comment) *Nature (Lond.)* **215**: 296, 1967.
- FELDMAN, P. L., JAMES, M. K., BRACKEEN, M. F., BILOTTA, J. M., SCHUSTER, S. V., LAHEY, A. P., LUTZ, M. W., JOHNSON, M. R., AND LEIGHTON, H. F.: Design, synthesis and pharmacological evaluation of ultrashort-to-long-acting opioid analgetics. *J. Med. Chem.* **34**: 2202-2208, 1991.
- FENSELAU, C.: Tandem mass spectrometry: the competitive edge for pharmacology. *Annu. Rev. Pharmacol. Toxicol.* **32**: 555-578, 1992.
- FICHTL, B., NIECIECKI, A. V., AND WALTER, K.: Tissue binding versus plasma binding of drugs: general principles and pharmacokinetic consequences. In *Advances in Drug Research*, ed. by B. Testa, vol. 20, pp. 117-166, Academic Press, New York, 1991.
- FIRST, M. R., WEISKITTEL, P., ALEXANDER, J. W., SCHROEDER, T. J., AND MYRE, S. A.: Concomitant administration of cyclosporin and ketoconazole in renal transplant recipients. *Lancet* **ii**: 1198-1201, 1984.
- FLEISCH, H.: Adverse events. In *Bisphosphonates in Bone Disease: From the Laboratory to the Patients*, ed. by H. Fleisch, pp. 126-132, Stämpfli Co., Ltd., Berne, Switzerland, 1993.
- FLEISCH, H., AND RUSSELL, R. G. G.: Pyrophosphate and polyphosphate. In *Pharmacology of the Endocrine System and Related Drugs*, ed. by H. Hellar, and B. Pickering, pp. 61-100, Pergamon Press, New York, 1970.
- FLEISCH, H., RUSSELL, R. G. G., BISAZ, S., CASEY, P. A., AND MÜHLBAUER, R. C.: The influence of pyrophosphate analogues (diphosphonates) on the precipitation and dissolution of calcium phosphate in vitro and in vivo. *Calcif. Tissue Res.* **2**(suppl.): 10-10A, 1968.
- FLEISCH, H., RUSSELL, R. G. G., AND FRANCIS, M. D.: Diphosphonates inhibit hydroxyapatite dissolution in vitro and bone resorption in tissue culture and in vivo. *Science (Wash. DC)* **165**: 1262-1264, 1969.
- FLEISCH, H., RUSSELL, R. G. G., AND STRAUMANN, F.: Effect of pyrophosphate on hydroxyapatite and its implications in calcium homeostasis. *Nature (Lond.)* **212**: 901-903, 1966.
- FLOWER, R. J., MONCADA, S., AND VANE, J. R.: Analgesic-antipyretics and anti-inflammatory agents: drugs employed in the treatment of gout. In *Goodman and Gilman's The Pharmacological Basis of Therapeutics*, ed. by A. G. Gilman, L. S. Goodman, T. W. Rall, and F. Murad, pp. 674-715, Macmillan Publisher, Inc., New York, 1985.
- FOSTER, R. T., AND JAMALI, F.: Stereoselective pharmacokinetics of ketoprofen in the rat: influence of route of administration. *Drug Metab. Dispos.* **16**: 623-626, 1988.
- FREILICH, D. J., AND GIARDINI, E.-G. V.: Imipramine binding to α_1 -acid glycoprotein in normal subjects and cardiac patients. *Clin. Pharmacol. Ther.* **35**: 670-674, 1984.
- GAN, L.-S., HSYU, P.-H., PRITCHARD, J. F., AND THAKKER, D.: Mechanism of intestinal absorption ranitidine and Ondansetron: transport across Caco-2 cell monolayers. *Pharm. Res.* **10**: 1722-1725, 1993.
- GELBOIN, H. V.: Benzo(a)pyrene metabolism, activation and carcinogenesis: role of mixed function oxidases and related enzymes. *Pharmacol. Rev.* **60**: 1107-1166, 1980.
- GERHARDT, R. E., KNOUSS, R. F., THYRUM, P. T., LUCHI, R. J., AND MORRIS, J. J.: Quinidine excretion in aciduria and alkaluria. *Ann. Intern. Med.* **71**: 927-933, 1969.
- GILLETTE, J. R.: Problems in correlating in vitro and in vivo studies of drug metabolism. In *Pharmacokinetics: A Modern View*, ed. by L. Z. Benet, G. Levy, and B. L. Ferraiolo, pp. 235-252, Plenum Press, New York, 1984.
- GILLETTE, J. R.: Problems in extrapolations from animals to man. In *Xenobiotic Metabolism and Disposition*, ed. by R. Kato, R. W. Estabrook, and M. N. Cayen, pp. 209-216, Taylor & Francis, London, UK, 1989.
- GLASS, P. S. A.: Remifentanyl: a new opioid. *J. Clin. Anesth.* **7**: 558-563, 1995.
- GONZALEZ, F. J., CRESPI, C. L., AND GELBOIN, H. V.: cDNA-expressed human cytochrome P-450s: a new age of molecular toxicology and human risk assessment. *Mutat. Res.* **247**: 113-127, 1991.
- GONZALEZ, F. J., AND NEBERT, D. W.: Evolution of the P450 gene superfamily: animal-plant "warfare", molecular drive and human genetic differences in drug metabolism. *Trends Genet.* **6**: 182-187, 1990.
- GOOD, S. S., DURACK, D. T., AND DEMIRANDA, P.: Biotransformation in various species and in humans of 3'-azido-3'-deoxythymidine, a potential agent for the treatment of AIDS. *Fed. Proc.* **45**: 44-52, 1986.
- GOUDSOUZIAN, N. G., D'HOLLANDER, A. A., AND VIBY-MOGENSEN, J.: Prolonged neuromuscular block from mivacurium in two patients with cholinesterase deficiency. *Anesth. Analg.* **77**: 183-185, 1993.
- GRANT, D. M., BLUM, M., BEER, M., AND MEYER, U. A.: Monomorphic and polymorphic human arylamine *N*-acetyltransferases: a comparison of liver isozymes and expressed products of two cloned genes. *Mol. Pharmacol.* **39**: 184-191, 1991.
- GRASS, G. M., AND SWEETANA, S. A.: In vitro measurement of gastrointestinal tissue permeability using a new diffusion cell. *Pharm. Res.* **5**: 372-376, 1988.
- GREEN, T.: Changes in metabolism during toxicity tests. *Xenobiotica* **20**: 1233-1240, 1990.
- GREEN, T.: Species differences in carcinogenicity: the role of metabolism and pharmacokinetics in risk assessment. *Ann. Ist. Super. Sanita* **27**: 595-600, 1991.

- GREENBLATT, D. J., ARENDT, R. M., ABERNATHY, D. R., GILES, H. G., SELLERS, E. M., AND SHADER, R. I.: In vitro quantitation of benzodiazepine lipophilicity: relation to in vivo distribution. *Br. J. Anaesth.* **55**: 985-1089, 1983.
- GREGORY, A. R.: Species comparisons in evaluating carcinogenicity in humans. *Regul. Toxicol. Pharmacol.* **8**: 160-190, 1988.
- GROEN, K., WARRANDER, A., MILES, G. S., BOOTH, B. S., AND MULDER, G. J.: Sulphation and glucuronidation of Xamoterol in the dog: dose dependence and site of sulphation. *Xenobiotica* **18**: 511-518, 1988.
- GUENGERICH, F. P.: Oxidation of quinidine by human liver cytochrome O-450. *Mol. Pharmacol.* **30**: 287-295, 1986.
- GUENGERICH, F. P.: Mammalian Cytochrome P-450, pp. 1-201, vol. 1, CRC Press, Boca Raton, FL, 1987.
- GUENGERICH, F. P., AND SHIMADA, T.: Human cytochrome P-450 enzymes and chemical carcinogenesis. In *Human Metabolism: From Molecular Biology to Man*, ed. by E. H. Jeffery, pp. 5-12, CRC Press, Boca Raton, FL, 1993.
- GUGLER, R., AND JENSEN, J. C.: Drug-protein binding in liver disease and in patients with hypoalbuminemia. In *Drug-Protein Binding*, ed. by M. M. Reidenberg, and S. Erill, pp. 189-200, Praeger, New York, 1986.
- GUILLOUZO, A., MOREL, F., FARDEL, O., AND MEUNIER, B.: Use of human hepatocyte cultures for drug metabolism studies. *Toxicology* **82**: 209-219, 1993.
- HAEFELY, W., KYBURZ, E., GERECKE, M., AND MÖHLER, H.: Recent advances in the molecular pharmacology of benzodiazepine receptors and in the structure-activity relationships of their agonists and antagonists. In *Advances in Drug Research*, ed. by B. Testa, pp. 166-299, Academic Press, New York, 1985.
- HAMMAN, M. A., WRIGHTON, S. A., AND HALL, S. D.: Sulindac metabolism by human liver microsomes. *International Society for the Study of Xenobiotics Proceedings* **6**: 251, 1994.
- HARASHIMA, H., SUGIYAMA, Y., SAWADA, Y., IGA, T., AND HANANO, M.: Comparison between in vivo and in vitro tissue-to-plasma unbound concentration ratios (K_p, p) of quinidine in rats. *J. Pharm. Pharmacol.* **36**: 340-342, 1984.
- HARD, G. C., AND WHYSNER, J.: Risk assessment of *d*-limonene: an example of male rat-specific renal tumorigens. *Crit. Rev. Toxicol.* **24**: 231-254, 1994.
- HARRIS, H. W.: High dose isoniazid compared with standard-dose isoniazid with PAS in the treatment of previously untreated cavitory pulmonary tuberculosis. In *Transactions of the 20th Conference of Chemotherapy and Tuberculosis*, pp. 39-68, U. S. Veterans Administration Army Navy, Washington, DC, 1961.
- HARTRICK, C. T., DIRKES, W. E., COYLE, D. E., RAJ, P. R., AND DENSON, D. D.: Influence of bupivacaine on mepivacaine protein binding. *Clin. Pharmacol. Ther.* **36**: 546-550, 1984.
- HASSEN, H. P., AGARWAL, D. P., GOEDDE, H. W., BUCHER, H., HULAND, H., BRACHMANN, W., AND OVENBECK, R.: Association of *N*-acetyltransferase polymorphism and environmental factors with bladder carcinogens. *Eur. Urol.* **11**: 263-266, 1985.
- HAWKINS, D., PINKARD, R. N., AND FARR, R. S.: Acetylation of human serum albumin by acetylsalicylic acid. *Science (Wash. DC)* **160**: 780-781, 1968.
- HENDERSON, C. J., AND WOLF, C. R.: Molecular analysis of cytochrome P450s in the CYP2 gene family. In *Progress in Drug Metabolism*, ed. by G. G. Gibson, pp. 73-139, Taylor & Francis, London, UK, 1992.
- HERRERA, A. M., SCOTT, D. O., AND LUNTE, C. E.: Microdialysis sampling for determination of plasma protein binding of drugs. *Pharm. Res.* **7**: 1077-1081, 1990.
- HIDALGO, I. J., RAUB, T. J., AND BORCHARDT, R. T.: Characterization of the human colon carcinoma cell line (Caco-2) as a model system for intestinal epithelial permeability. *Gastroenterology* **96**: 736-749, 1989.
- HO, N. F. H., MERKLE, H. P., AND HIGUCHI, W. I.: Quantitative, mechanistic and physiologically realistic approach to the biopharmaceutical design of oral drug delivery system. *Drug Dev. Ind. Pharm.* **9**: 1111-1184, 1983.
- HONIG, P. K., WOOSLEY, R. L., ZAMANI, K., CONNER, D. P., AND CANTILENA, L. R.: Changes in the pharmacokinetics and electrocardiographic pharmacodynamics of terfenadine with concomitant administration of erythromycin. *Clin. Pharmacol. Ther.* **52**: 231-238, 1992.
- HONIG, P. K., WORTHAM, D. C., ZAMANI, K., CONNER, D. P., MULLIN, J. C., AND CANTILENA, L. R.: Terfenadine-ketoconazole interaction: pharmacokinetic and electrocardiographic consequences. *JAMA* **269**: 1535-1539, 1993.
- HOSOKAWA, M., MAKI, T., AND SATOH, T.: Multiplicity and regulation of liver microsomal carboxylesterase in rats. *Mol. Pharmacol.* **31**: 579-584, 1987.
- HOSOKAWA, M., MAKI, T., AND SATOH, T.: Characterization of molecular species of liver microsomal carboxylesterases of several animal species and human. *Arch. Biochem. Biophys.* **277**: 219-227, 1990.
- HOUSTON, J. B.: Utility of in vitro drug metabolism data in predicting in vivo metabolic clearance. *Biochem. Pharmacol.* **47**: 1469-1479, 1994.
- HUANG, J. D., AND ØIE, S.: Effect of altered disopyramide binding its pharmacologic response in rabbits. *J. Pharmacol. Exp. Ther.* **223**: 469-471, 1982.
- HUANG, M.-T., CHANG, R. L., FORTNER, J. G., AND CONNEY, A. H.: Studies on the mechanism of activation of microsomal benz[*a*]pyrene hydroxylation by flavonoids. *J. Biol. Chem.* **256**: 6829-6836, 1981.
- HUGHES, R., AND CHAPPLE, D. J.: The pharmacology of atracurium: a new competitive neuromuscular blocking agent. *Br. J. Anaesth.* **53**: 31-44, 1981.
- HUMPHREY, M. J.: Pharmacokinetic studies in the selection of new drugs: a case history on dihydropyridine calcium channel blockers. In *Xenobiotic Metabolism and Disposition*, ed. by R. (Kato, R. W. Estahrook, and M. N. Gayen, pp. 245-253, Taylor & Francis, New York, 1989.
- HUNT, C. M., WESTERKAM, W. R., AND STAVE, G. M.: Effect of age and gender on the activity of human hepatic CYP3A. *Biochem. Pharmacol.* **44**: 275-283, 1992.
- HUTT, A. J., AND CALDWELL, J.: The metabolic chiral inversion of 2-arylpropionic acids: a novel route with pharmacological consequences. *J. Pharm. Pharmacol.* **35**: 693-704, 1983.
- IGARI, Y., SUGIYAMA, Y., AWAZU, S., AND HANANO, M.: Comparative physiologically based pharmacokinetics of hexobarbital, phenobarbital and thiopental in the rat. *J. Pharmacokin. Biopharm.* **10**: 53-75, 1982.
- INABA, T., TAIT, A., NAKANO, M., MAHON, W. A., AND KALOW, W.: Metabolism of diazepam in vitro by human liver: independent variability of *N*-demethylation and C3-hydroxylation. *Drug Metab. Dispos.* **16**: 605-608, 1988.
- IOANNIDES, C., AND PARKE, D. V.: Induction of cytochrome P-4501 as an indicator of potential chemical carcinogenesis. *Drug Metab. Rev.* **25**: 485-501, 1993.
- IOANNIDES, C., SWEATMAN, B., RICHARDS, R., AND PARKE, D. V.: Drug metabolism in the ferret: effects of age, sex and strain. *Gen. Pharmacol.* **8**: 243-249, 1977.
- ISRAILI, Z. H., AND EL-ATTAR, H.: Binding of some acidic drugs to α_1 -acid glycoprotein. *Clin. Pharmacol. Ther.* **33**: 255-258, 1983.
- IWATSUBO, T., HIROTA, N., OOIE, T., SUZUKI, K., SHIMADA, N., CHIBA, K., ISHIZAK, T., GREEN, C. E., TYSON, C. A., AND SUGIYAMA, Y.: Prediction of in vivo drug metabolism in the human liver from in vitro metabolism data. *Pharmacol. Ther.*, in press, 1997.
- IYER, R. S., COLES, B. F., RANEY, K. D., THIER, R., GUENGERICH, F. P., AND HARRIS, T. M.: DNA adduction by the potent carcinogen aflatoxin B1: mechanistic studies. *J. Am. Chem. Soc.* **116**: 1603-1609, 1994.
- JACKSON, M. J.: Drug transport across gastrointestinal epithelia. In *Physiology of the Gastrointestinal Tract*, 2nd ed., ed. by L. R. Jackson, pp. 1597-1621, Raven Press, New York, 1987.
- JAMALI, F., BERRY, B. W., TEHRANI, M. R., AND RUSSELL, S. A.: Stereoselective pharmacokinetics of flurbiprofen in humans and rats. *J. Pharm. Sci.* **77**: 666-669, 1988.
- JAMALI, F., MEHVAR, R., AND PASUTTO, F. M.: Enantioselective aspects of drug action and disposition: therapeutic pitfalls. *J. Pharm. Sci.* **78**: 695-715, 1989.
- JENSEN, C. B., AND JOLLOU, D. J.: The role of *N*-hydroxyphenetidine in phenacetin-induced hemolytic anemia. *Toxicol. Appl. Pharmacol.* **111**: 1-12, 1991.
- JERINA, D. M., YAGI, H., THAKKER, D. R., KARLE, J. M., BUENING, M., CHANG, R. L., LEVIN, W., AND CONNEY, A. H.: Stereoselective metabolic activation of polycyclic aromatic hydrocarbons. In *Advances in Pharmacology and Therapeutics*, ed. by Y. Cohen, vol. 9, pp. 53-62, Pergamon Press, New York, 1979.
- JEZEQUEL, S. G.: Central nervous system penetration of drugs: importance of physicochemical properties. In *Progress in Drug Metabolism*, ed. by G. G. Gibson, vol. 13, pp. 141-178, Taylor & Francis, London, UK, 1992.
- JOHNSON, E. J., PALMER, C. N. A., GRIFFIN, K. J., AND HSU, M. H.: Role of the peroxisome proliferator-activated receptor in cytochrome P450 4A gene regulation. *FASEB J.* **10**: 1241-1248, 1996.
- JOHNSON, G., JORDÖ, L., LUNDBORG, P., REGÄRDH, C.-G., AND RÖNN, O.: Plasma levels and pharmacological effects of metoprolol administered as controlled release (Durules) and ordinary tablets in healthy volunteers. *Int. J. Clin. Pharmacol. Ther. Toxicol.* **18**: 292-297, 1980.
- JUMARIE, C., AND MALO, C.: Caco-2 cells culture in serum-free medium as a model for the study of enterocytic differentiation in vitro. *J. Cell. Physiol.* **149**: 24-33, 1991.
- KADLUBAR, F. F., MILLER, J. A., AND MILLER, E. C.: Hepatic microsomal *N*-glucuronidation and nucleic acid binding of *N*-hydroxy arylamines in relation to urinary bladder carcinogenesis. *Cancer Res.* **37**: 805-814, 1977.
- KADLUBAR, F. F., UNRUH, L. E., FLAMMANG, T. J., SPARKS, D., MITCHUM, R. K., AND MULDER, G. J.: Alteration of urinary levels of the carcinogen, *N*-hydroxy-2-naphthylamine, and its *N*-glucuronide in the rat by control of urinary pH, inhibition of metabolic sulfation, and changes in biliary excretion. *Chem. Biol. Interact.* **33**: 129-147, 1981.
- KALOW, W., AND BERTILSSON, L.: Interethnic factors affecting drug response. In *Advances in Drug Research*, ed. by B. Testa and U. A. Meyer, pp. 1-53, Academic Press, New York, 1994.
- KALOW, W., AND GENEST, K.: A method for the detection of atypical forms of human serum cholinesterase: determination of dibucaine numbers. *Can. J. Biochem. Physiol.* **35**: 339-346, 1957.
- KAMATAKI, T.: Molecular toxicology of cytochrome P-450: focusing on interspecies homology. *Yakugaku Zasshi* **115**: 370-377, 1995.
- KANERVA, R. L., RIDDER, G. M., LEFFEVER, F. R., AND ALDEN, C. L.: Comparison of short-term renal effects due to oral administration of decalin or *d*-limonene in young adult male Fischer 344 rats. *Food Chem. Toxicol.* **25**: 345-354, 1987.
- KAPPAS, A., ALVARES, A. P., ANDERSON, K. E., PANTUCK, E. J., PANTUCK, C. B., CHANG, R., AND CONNEY, A. H.: Effect of charcoal-broiled beef on antipyrine and theophylline metabolism. *Clin. Pharmacol. Ther.* **23**: 445-450, 1978.
- KAPPAS, A., ANDERSON, K. E., CONNEY, A. H., AND ALVARES, A. P.: Influence of dietary protein and carbohydrate on antipyrine and theophylline metabolism in man. *Clin. Pharmacol. Ther.* **20**: 643-653, 1976.
- KATO, R., AND YAMAZOE, Y.: Sex-dependent regulation of cytochrome P-450 expression. In *Principles, Mechanisms and Biological Consequences of In-*

- duction, ed. by K. Ruckpaul and H. Rein, pp. 82–112, Taylor & Francis, New York, 1990.
- KATO, R., AND YAMAZOE, Y.: The importance of substrate concentration in determining cytochrome P-450 therapeutically relevant *in vivo*. *Pharmacogenetics* **4**: 359–362, 1994.
- KAUMEIER, S.: The effect of the composition of food on the absorption of sulfamerazine. *Int. J. Clin. Pharmacol. Biopharm.* **17**: 260–263, 1979.
- KAWASAKI, S., SUGIYAMA, Y., IGA, T., HANANO, M., SANJO, K., BEPPU, T., AND IDEZUKI, Y.: Pharmacokinetic study on the hepatic uptake of indocyanine green in cirrhotic patients. *Am. J. Gastroenterol.* **80**: 801–806, 1985.
- KEATES, E. U., AND STONE, R.: The effect of d-timolol on intraocular pressure in patients with ocular hypertension. *Am. J. Ophthalmol.* **98**: 73–78, 1984.
- KELLY, J. G., AND O'MALLEY, K.: Clinical pharmacokinetics of the newer ACE inhibitors: a review. *Clin. Pharmacokinet.* **19**: 177–196, 1990.
- KEMPF, D. J., MARCH, K. C., DENISSEN, J. F., McDONALD, E., VASAVANONDA, S., FLENTGE, C. A., GREEN, B. E., FINO, L., PARK, C. H., LONG, X-P., WIDEBURG, N. E., SALDIVAR, A., RUIZ, L., KATI, W. M., SHAM, H. L., ROBINS, T., STEWART, K. D., HSU, A., PLATTNER, J. J., LEONARD, J. M., AND NORBECK, D. W.: ABT-538 is a potent inhibitor of human immunodeficiency virus protease and has high oral bioavailability in humans. *Proc. Natl. Acad. Sci. USA* **92**: 2484–2488, 1995.
- KEMPF, D. J., MARSH, K. C., PAUL, D. A., KNIGGE, M. K., NORBECK, D. W., KOHLBRENNER, W. E., CODACOV, L., VASAVANONDA, S., BRYANT, P., WANG, X. C., WIDEBURG, N. E., CLEMENT, J. J., PLATTNER, J. J., AND ERICKSON, J.: Antiviral and pharmacokinetic properties of C₂ symmetric inhibitors of the human immunodeficiency virus type 1 protease. *Antimicrob. Agents Chemother.* **35**: 2209–2214, 1991.
- KLEINBLOESEM, C. H., VAN BRUMMELEN, P., FABER, H., DANHOF, M., VERMEULEN, N. Y., AND BREIMER, D. D.: Variability in nifedipine pharmacokinetics and dynamics: a new oxidation polymorphism in man. *Biochem. Pharmacol.* **33**: 3721–3724, 1984.
- KOBAYASHI, S., MURRAY, S., WATSON, D., SESARDIC, D., DAVIES, D. S., AND BOOBIS, A. R.: The specificity of inhibition of debrisoquine 4-hydroxylase activity in quinidine and quinine in the rat is the reverse of that in man. *Biochem. Pharmacol.* **38**: 2795–2799, 1989.
- KOBLIAKOV, V., POPOVA, N., AND ROSSI, L.: Regulation of the expression of the sex-specific isoforms of cytochrome P-450 in rat liver. *Eur. J. Biochem.* **195**: 585–591, 1991.
- KOCH-WESER, J.: The serum level approaches to individualization of drug dosage. *Eur. J. Clin. Pharmacol.* **9**: 1–8, 1975.
- KOIKE, Y., MAGNUSSON, A., STEINER, E., RANE, A., AND SJOGVIST, F.: Ultrafiltration compared with equilibrium dialysis in determination of unbound phenytoin in plasma. *Ther. Drug Monit.* **7**: 461–465, 1985.
- KRAGH-HANSEN, U., BRENNAN, S. O., GALLIANO, M., AND SUGITA, O.: Binding of warfarin, salicylate and diazepam to genetic variants of human serum albumin with known mutations. *Mol. Pharmacol.* **37**: 238–242, 1990.
- KRASNY, H. C., AND PETTY, B. G.: Metabolism of desciclovir, a prodrug of acyclovir, in humans after multiple oral dosing. *J. Clin. Pharmacol.* **27**: 74–77, 1987.
- KREMER, J. M. H., WILTING, J., AND JANSSEN, L. H. M.: Drug binding to α_1 -acid glycoprotein in health and disease. *Pharmacol. Res.* **40**: 1–47, 1988.
- KRENTSKY, T. A., AND ELION, G. B.: Enzymes as tools and targets in drug research. *In Strategy in Drug Research*, ed. by J. A. Keverling Buisman, pp. 65–87, Elsevier, Amsterdam, 1982.
- KRENTSKY, T. A., HALL, W. W., DE MIRANDA, P., BEAUCHAMP, L. M., SCHAEFFER, M. J., AND WHITEMAN, P. D.: 6-Deoxyacyclovir: a xanthine oxidase-activated prodrug of acyclovir. *Proc. Natl. Acad. Sci. USA* **81**: 3209–3213, 1984.
- KROPP, H., SUNDELDF, J. G., KAHAN, J. S., KAHAN, F. M., AND BIRNBAUM, J.: MK-0787 (*N*-formimidoyl thienamycin): evaluation of *in vitro* and *in vivo* activities. *Antimicrob. Agents Chemother.* **17**: 993–1000, 1980.
- KUNZE, K. L., WIENKERS, L. C., THUMMEL, K. E., AND TRAGER, W. F.: Warfarin-fluconazole: I—Inhibition of the human cytochrome P-450-dependent metabolism of warfarin by fluconazole: *in vitro* studies. *Drug Metab. Dispos.* **24**: 414–421, 1996.
- KÜPPER, A., AND PREISIG, R.: Pharmacogenetics of mephenytoin: a new drug hydroxylation polymorphism in man. *Eur. J. Clin. Pharmacol.* **26**: 753–759, 1984.
- KURZ, H.: Methodological problems in drug-binding studies. *In Drug-protein Binding*, ed. by M. M. Reidenberg and S. Erill, pp. 70–92, Fraeger Publishers, New York, 1986.
- KURZ, H., TRUNK, H., AND WEITZ, B.: Evaluation of methods to determine protein-binding of drugs: equilibrium dialysis, ultrafiltration, ultracentrifugation and gel filtration. *Arzneim-Forsch./Drug Res.* **27**: 1373–1380, 1977.
- LA DU, B. N.: Human serum paraoxonase/arylesterase. *In Pharmacogenetics and Drug Metabolism*, ed. by W. Kalow, pp. 51–91, Pergamon Press, New York, 1992.
- LAURSEN, L. C., BORGA, O., KROHN, L., AND WEEKE, B.: Distribution of enprofylline and theophylline between plasma and cerebrospinal fluid. *Ther. Drug Monit.* **11**: 162–164, 1989.
- LAWSON, D. H., HENRY, D. A., LOWE, J., REAVEY, P., RENNIE, J. A. N., AND SOLOMAN, A.: Acetylator phenotype in spontaneous SLE and rheumatoid arthritis. *Ann. Rheum. Dis.* **38**: 171–173, 1979.
- LEAHY, D. E., LYNCH, J., AND TAYLOR, D. D.: Mechanisms of absorption of small molecules. *In Novel Drug Delivery*, ed. by L. F. Prescott and W. S. Nimmo, pp. 33–44, John Wiley & Sons, New York, 1989.
- LE BIGOT, J. F., BEGUE, J. M., KIECHEL, J. R., AND GUILLOUZO, A.: Species differences in metabolism of ketotifen in rat, rabbit and human: demonstration of similar pathways *in vivo* and in cultured hepatocytes. *Life Sci.* **40**: 883–890, 1987.
- LEE, E. J. D., AND WILLIAMS, K. M.: Chirality: clinical pharmacokinetic and pharmacodynamic considerations. *Clin. Pharmacokinet.* **18**: 339–345, 1990.
- LEE, E. J. D., WILLIAMS, K. M., DAY, R. O., GRAHAM, G. G., AND CHAMPION, G. D.: Stereoselective disposition of ibuprofen enantiomers in man. *Br. J. Clin. Pharmacol.* **19**: 669–674, 1985.
- LEGRAVEREND, C., MODE, A., WELLS, T., ROBINSON, I., AND GUSTAFSSON, J.-A.: Hepatic steroid hydroxylating enzymes are controlled by the sexually dimorphic pattern of growth hormone secretion in normal and dwarf rats. *FASEB J.* **6**: 711–718, 1992a.
- LEGRAVEREND, C., MODE, A., WESTIN, S., STRÖM, A., EGUCHI, H., ZAPHIROPOULOS, P., AND GUSTAFSSON, J.-A.: Transcriptional regulation of rat P-450 2C gene subfamily members by the sexually dimorphic pattern of growth hormone secretion. *Mol. Endocrin.* **6**: 259–266, 1992b.
- LEMOINE, A., GAUTIER, J. C., AZOULAY, D., KIFFEL, L., BELLOC, C., GUENGERICH, F. P., MAUREL, P., BEAUNE, P., AND LEREUX, J. P.: Major pathway of imipramine metabolism is catalyzed by cytochrome P-450 1A2 and P-450 3A4 in human liver microsomes. *Mol. Pharmacol.* **43**: 827–832, 1993.
- LENNARD, L., VAN LOON, J., LILLEYMAN, J. S., AND WEINSHILBOUM, R. M.: Thiopurine pharmacogenetics in leukemia: correlation of erythrocyte thiopurine methyltransferase activity and 6-thioguanine nucleotide concentrations. *Clin. Pharmacol. Ther.* **41**: 18–25, 1987.
- LENNARD, L., VAN LOON, J., AND WEINSHILBOUM, R. M.: Pharmacogenetics of acute azathioprine toxicity: relationship to thiopurine methyltransferase genetic polymorphism. *Clin. Pharmacol. Ther.* **46**: 149–154, 1989.
- LENNERNÄS, H., PALM, K., FAGERHOLM, U., AND ARTURSSON, P.: Correlation between paracellular and transcellular drug permeability in the human jejunum and Caco-2 monolayers. *Int. J. Pharm.* **127**: 103–107, 1996.
- LEVINE, W. G.: Heavy-metal antagonists. *In The Pharmacological Basis of Therapeutics*, ed. by L. S. Goodman and A. Gilman, pp. 912–923, Macmillan Publishing Co., Inc., New York, 1975.
- LEVY, G.: Effect of plasma protein binding of drugs on duration and intensity of pharmacological activity. *J. Pharm. Sci.* **65**: 1264–1265, 1976.
- LEWIS, R. J., TRAGER, W. F., CHAN, K. K., BRECKENRIDGE, A., ORME, M., ROWLAND, M., AND SCHARY, W.: Warfarin: stereochemical aspects of its metabolism and its interactions with phenylbutazone. *J. Clin. Invest.* **53**: 1607–1617, 1974.
- LIN, H. J., HAN, C.-Y., LIN, B. K., AND HARDY, S.: Slow acetylator mutations in the human polymorphic *N*-acetyltransferase gene in 786 Asians, blacks, Hispanics and whites: application to metabolic epidemiology. *Am. J. Hum. Genet.* **52**: 827–834, 1993a.
- LIN, J. H.: Pharmacokinetic and pharmacodynamic properties of histamine H₂-receptor antagonists: relationship between intrinsic potency and effective plasma concentrations. *Clin. Pharmacokinet.* **20**: 218–236, 1991.
- LIN, J. H.: Species similarities and differences in pharmacokinetics. *Drug Metab. Dispos.* **23**: 1008–1021, 1995.
- LIN, J. H.: Bisphosphonates: a review of their pharmacokinetic properties. *Bone* **18**: 75–85, 1996a.
- LIN, J. H.: *In vitro* and *in vivo* drug interaction with indinavir. *In Symposium on Recent Advances in Drug-Drug Interactions*, Washington, DC, December 10–11, 1996b.
- LIN, J. H., CHEN, I.-W., AND DELUNA, F. A.: Dose-dependent pharmacokinetics of MK-417, a potent carbonic anhydrase inhibitor, in experimental polycythemic and anemic rats. *Pharm. Res.* **8**: 608–614, 1991a.
- LIN, J. H., CHEN, I.-W., AND DELUNA, F. A.: Effects of dose, sex, and age on the disposition of alendronate, a potent antiosteolytic bisphosphonate, in rats. *Drug Metab. Dispos.* **20**: 473–478, 1992.
- LIN, J. H., CHEN, I.-W., AND DELUNA, F. A.: Uptake of alendronate by the bone tissues in hypocalcemic and hypercalcemic rats. *Drug Metab. Dispos.* **21**: 800–804, 1993b.
- LIN, J. H., CHEN, I.-W., AND LIN, T.-H.: Species-dependent stereopharmacokinetics of MK-927, a potent carbonic anhydrase inhibitor. *Drug Metab. Dispos.* **19**: 816–822, 1991b.
- LIN, J. H., CHEN, I.-W., AND LIN, T.-H.: Blood-brain barrier permeability and *in vivo* activity of partial agonists of benzodiazepine receptor: a study of L-663,581 and its metabolites in rats. *J. Pharmacol. Exp. Ther.* **271**: 1197–1202, 1994.
- LIN, J. H., CHEN, I.-W., ULM, E. H., AND DUGGAN, D. E.: Differential effects of phenobarbital on ester and ether glucuronidation of diflunisal in rats. *J. Pharmacol. Exp. Ther.* **242**: 1013–1018, 1987a.
- LIN, J. H., CHEN, I.-W., ULM, E. H., AND DUGGAN, D. E.: Differential renal handling of angiotensin converting enzyme inhibitors enalaprilat and lisinopril in rats. *Drug Metab. Dispos.* **16**: 392–396, 1988.
- LIN, J. H., CHEN, I.-W., ULM, E. H., GEHRET, J. R., AND DUGGAN, D. E.: Dose-dependent pharmacokinetics of MK-417, a potent carbonic anhydrase inhibitor, in rabbits following single and multiple doses. *Drug Metab. Dispos.* **18**: 836–841, 1990a.
- LIN, J. H., CHIBA, M., BALANI, S. K., CHEN, I.-W., KWEL, G. Y.-S., VASTAG, K. J., AND NISHIME, J. A.: Species differences in the pharmacokinetics and metab-

- olism of indinavir, a potent HIV protease inhibitor. *Drug Metab. Dispos.* **24**: 1111–1120, 1996a.
- LIN, J. H., CHIBA, M., CHEN, I-W., NISHIME, J. A., AND VASTAG, K. J.: Sex-dependent pharmacokinetics of indinavir: in vivo and in vitro evidence. *Drug Metab. Dispos.* **24**: 1298–1306, 1996b.
- LIN, J. H., CHIBA, M., CHEN, I-W., VASTAG, K. J., NISHIME, J. A., DORSEY, B. D., MICHAELSON, S., AND MCDANIEL, S. L.: Time- and dose-dependent pharmacokinetics of L-754,394, an HIV protease inhibitor, in rats, dogs and monkeys. *J. Pharmacol. Exp. Ther.* **274**: 264–269, 1995.
- LIN, J. H., COCCHETTO, D. M., AND DUGGAN, D. E.: Protein binding as a primary determinant of the clinical pharmacokinetic properties of non-steroidal anti-inflammatory drugs. *Clin. Pharmacokinet.* **12**: 402–432, 1987b.
- LIN, J. H., DELUNA, F. A., ULM, E. H., AND TOCCO, D. J.: Species-dependent enantioselective plasma protein binding of MK-571, a potent leukotriene D₄ antagonist. *Drug Metab. Dispos.* **18**: 484–487, 1990b.
- LIN, J. H., DUGGAN, D. E., CHEN, I-W., AND ELLSWORTH, R. L.: Physiological disposition of alendronate, a potent anti-osteolytic bisphosphonate, in laboratory animals. *Drug Metab. Dispos.* **19**: 926–932, 1991c.
- LIN, J. H., HAYASHI, M., AWAZU, S., AND HANANO, M.: Correlation between in vitro and in vivo drug metabolism rate: oxidation of ethoxybenzamide in rat. *J. Pharmacokinet. Biopharm.* **6**: 327–337, 1978.
- LIN, J. H., AND LEVY, G.: Effect of prevention of inorganic sulfate depletion on the pharmacokinetics of acetaminophen in rats. *J. Pharmacol. Exp. Ther.* **239**: 94–98, 1986.
- LIN, T. H., AND LIN, J. H.: Effects of protein binding and experimental disease states on brain uptake of benzodiazepines in rats. *J. Pharmacol. Exp. Ther.* **253**: 45–50, 1990.
- LIN, J. H., STOREY, D. E., CHEN, I-W., AND XU, X.: Improved oral absorption of L-365,260, a poorly soluble drug. *Biopharm. Drug Dispos.* **17**: 1–16, 1996c.
- LIN, J. H., SUGIYAMA, Y., AWAZU, S., AND HANANO, M.: In vitro and in vivo evaluation of tissue to plasma partition coefficient for physiological pharmacokinetic model. *J. Pharmacokinet. Biopharm.* **10**: 637–647, 1982.
- LIN, J. H., ULM, E. H., AND LOS, L. E.: Dose-dependent stereopharmacokinetics of 5,6-dihydro-4H-4(isobutylamino)thieno(2,3, B)thiopyran-2-sulfonamide-7,7-dioxide (MK-927), a potent carbonic anhydrase inhibitor, in rats. *Drug Metab. Dispos.* **19**: 233–238, 1991d.
- LINDBERG, L. P., AND NEGISHI, M.: Alternation of mouse cytochrome P450c₁ substrate specificity by mutation of a single amino-acid residue. *Nature (Lond.)* **339**: 632–634, 1989.
- LINTON, A. L., LUKE, R. G., AND BRIGGS, M. D.: Methods of forced diuresis and its application in barbiturate poisoning. *Lancet* **ii**: 377–380, 1967.
- LLERENA, A. L., ALM, C., DAHL, M-J., EKQUIST, B., AND BERTILSSON, L.: Haloperidol disposition is dependent on debrisoquine oxidation phenotype. *Ther. Drug Monit.* **14**: 92–97, 1992.
- LOCKRIDGE, O.: Genetic variants of human serum buthrylcholinesterase influence the metabolism of muscle relaxant succinylcholine. In *Pharmacogenetics of Drug Metabolism*, ed. by W. Kalow, pp. 15–50, Pergamon Press, New York, 1992.
- LOO, J. C. K., FOLTZ, E. L., WALLICK, H., AND KWAN, K. C.: Pharmacokinetics of pivampicillin and ampicillin in man. *Clin. Pharmacol. Ther.* **16**: 35–43, 1974.
- LUNDE, P. K. M., PIKE, E., AND BREDESEN, J. E.: Inflammation and α_1 -acid glycoprotein: effect on drug binding. In *Drug-Protein Binding*, ed. by M. M. Reidenberg and S. Erill, pp. 201–219, Praeger, New York, 1986.
- MACKICHAN, J. J.: Pharmacokinetic consequences of drug displacement from blood and tissue proteins. *Clin. Pharmacokinet.* **9**: 32–41, 1984.
- MACKICHAN, J. J.: Protein binding drug displacement interactions: fact or fiction. *Clin. Pharmacokinet.* **16**: 65–73, 1989.
- MACLEOD, J. N., SOREENSEN, M. P., AND SHAPIRO, B. H.: Strain independent elevation of hepatic monooxygenase enzyme in female mice. *Xenobiotics* **17**: 1095–1102, 1987.
- MACPHERSON, C. R., MILNE, M. D., AND EVANS, D. M.: The excretion of salicylate. *Br. J. Pharmacol.* **10**: 484–489, 1955.
- MAHGOUN, A., IDLE, J. R., DRING, L. G., LANCASTER, R., AND SMITH, R. L.: Polymorphic hydroxylation of debrisoquine in man. *Lancet* **ii**: 584–586, 1977.
- MANZEL-SOGLOWEK, S., GEISSLINGER, G., BECK, W. S., AND BRUNE, K.: Variability of inversion of (R)-flurbiprofen in different species. *J. Pharm. Sci.* **81**: 888–891, 1992.
- MARSDEN, C. D.: Advances in the management of Parkinson's disease. *Scott. Med. J.* **21**: 139–148, 1976.
- MARTIN, Y. C., AND HANSCH, C.: Influence of hydrophobic character on the relative rate of oxidation of drugs by rat liver microsomes. *J. Med. Chem.* **14**: 777–779, 1971.
- MAY, F. E., STEWART, R. B., AND CLUFF, L. E.: Drug interactions and multiple drug administration. *Clin. Pharmacol. Ther.* **22**: 323–327, 1977.
- MCCREA, J., WOOLF, E., STERRETT, A., MATTHEWS, C., DEUTSCH, P., YEH, K. C., WALDMAN, S., AND BJORNSSON, T.: Effects of ketoconazole and other P-450 inhibitors on the pharmacokinetics of indinavir. (Abstract) *Pharm. Res.* **13**(suppl.): S485, 1996.
- MCDONNELL, W. M., SCHEIMAN, J. M., AND TRABER, P. G.: Induction of cytochrome P450 IA genes (CYP 1A) by omeprazole in the human alimentary tract. *Gastroenterology* **103**: 1509–1516, 1992.
- MEYER, H., AND MALLY, J.: über hydrazinderivate der pyridin-carbonsäuren. *Mk. Chem.* **33**: 393–414, 1912.
- MEYER, J. H.: Motility of stomach and gastroduodenal junction. In *Physiology of the Gastrointestinal Tract*, ed. by L. R. Johnson, pp. 613–629, Raven Press, New York, 1987.
- MEYER, J. W., WOGGON, B., BAUMANN, P., AND MEYER, U. A.: Slow sulphoxidation of thioridazine in a poor metabolizer of debrisoquine type. *Eur. J. Clin. Pharmacol.* **39**: 613–614, 1990a.
- MEYER, U. A.: The molecular basis of genetic polymorphisms of drug metabolism. *J. Pharm. Pharmacol.* **46**(suppl. 1): 409–415, 1994.
- MEYER, U. A., BLUM, M., GRANT, D. M., HEIM, M., BROLY, F., HOFFMANN, F., PROBST, M., AND GARCIA-AGUNDEZ, J.: Acetylation pharmacogenetics. In *Human Drug Metabolism: From Molecular Biology to Man*, ed. by E. H. Jefferey, pp. 117–124, CRC Press, Boca Raton, FL, 1993.
- MEYER, U. A., SKODA, R. C., ZANGER, U. M., HEIM, M., AND BROLY, F.: The genetic polymorphism of debrisoquine/sparteine metabolism: molecular mechanisms. In *Pharmacogenetics of Drug Metabolism*, ed. by W. Kallow, pp. 609–623, Pergamon Press, New York, 1992.
- MEYER, U. A., ZANGER, U. M., GRANT, D., AND BLUM, M.: Genetic polymorphisms of drug metabolism. *Adv. Drug Res.* **19**: 197–241, 1990b.
- MILLER, J. A., AND SURH, Y-J.: Sulfonation in chemical carcinogenesis. In *Conjugation-Deconjugation Reactions in Drug Metabolism and Toxicity*, ed. by F. C. Kauffman, pp. 429–457, Springer-Verlag, Berlin, Germany, 1994.
- MILOVIC, V., OLSSON, S., OCKLIND, G., HOCHMAN, J., PAUL, E. C. A., AND ARTURSSON, P.: A new intestinal epithelial cell culture model (2/4A1) for studies of paracellular drug transport. (Abstract) *Pharm. Res.* **13**(suppl.): 358S, 1996.
- MINERS, J. O., LILLYWHITE, K. J., AND BIRKETT, D. J.: In vitro evidence for the involvement of at least two forms of human liver UDP-glucuronosyltransferase in morphine 3-glucuronidation. *Biochem. Pharmacol.* **37**: 2839–2845, 1988.
- MITCHELL, J. R., ZIMMERMAN, H. J., ISHAK, K. G., THORGERSSON, U. P., TIMBRELL, J. A., SNOODGRASS, W. R., AND NELSON, S. D.: Isoniazid liver injury: clinical spectrum, pathology and possible pathogenesis. *Ann. Intern. Med.* **84**: 181–192, 1976.
- MOMMSEN, S., AND AAGAARD, J.: Tobacco as a risk factor in bladder cancer. *Carcinogenesis* **4**: 335–338, 1983.
- MONAHAN, B. P., FERGUSON, C. L., KILLEAVY, E. S., LLOYD, B. K., TROY, J., AND CANTILENA, L. R.: Torsades de points occurring in association with terfenadine use. *JAMA* **264**: 2788–2790, 1990.
- MONKS, T. J., AND LAU, S. S.: Toxicology of quinone-thioethers. *CRC Crit. Rev. Toxicol.* **22**: 243–270, 1992.
- MONKS, T. J., AND LAU, S. S.: Glutathione conjugation as a mechanism for the transport of reactive metabolites. *Adv. Pharmacol.* **27**: 183–210, 1994.
- MONKS, T. J., AND LAU, S. S.: Glutathione conjugate-mediated toxicities. In *Conjugation-Deconjugation Reactions in Drug Metabolism and Toxicity*, ed. by F. C. Kauffman, pp. 459–509, Springer-Verlag, Berlin, Germany, 1994.
- MORDENTI, J.: Pharmacokinetic scale-up: accurate prediction of human pharmacokinetic profiles from animal data. *J. Pharm. Sci.* **74**: 1097–1099, 1985.
- MULDER, G. J.: Pharmacological effects of drug conjugates: is morphine 6-glucuronide an exception? *Trends Pharmacol. Sci.* **13**: 302–304, 1992.
- MULDER, G. J., GROEN, K., TIJDENS, R. B., AND MEIJER, D. K. F.: Pharmacokinetics of Xamoterol glucuronidation in the rat in vivo and in liver perfusion. *Xenobiotica* **17**: 85–92, 1987.
- MUNGALL, D., LUDDEN, T. M., MARSHALL, J., AND HAWKINS, D.: Relationships between steady-state warfarin concentrations and anticoagulant effect. *Clin. Pharmacokinet.* **9**(suppl. 1): 99–100, 1984.
- MURRAY, M.: Mechanisms of the inhibition of cytochrome P-450-mediated drug oxidation by therapeutic agents. *Drug Metab. Rev.* **18**: 55–81, 1987.
- MUTSCHLER, E., AND DERENDORF, H.: Muscle relaxants. In *Drug Actions: Basic Principles and Therapeutic Aspects*, ed. by E. Mutschler and H. Derendorf, pp. 195–203, CRC Press, Boca Raton, FL, 1995.
- MUTSCHLER, E., AND DERENDORF, H.: Ulcer therapy. In *Drug Actions: Basic Principles and Therapeutic Aspects*, ed. by E. Mutschler and H. Derendorf, pp. 424–429, CRC Press, Boca Raton, FL, 1995.
- NEBERT, D. W., AND GONZALEZ, F. J.: The P450 gene superfamily. In *Principles, Mechanisms and Biological Consequences of Induction*, ed. by K. Ruckpaul and H. Rein, pp. 35–61, Taylor & Francis, London, UK, 1990.
- NELSON, D. R., KAMATAKI, T., WAXMAN, D. J., GUENGERICH, F. P., ESTABROOK, R. W., FEYEREISEN, R., GONZALEZ, F. J., COON, M. J., GUNSALES, I. C., GOTOH, O., OKUDA, K., AND NEBERT, D. W.: The P-450 superfamily: update on new sequences, gene mapping, accession numbers, early trivial names of enzymes, and nomenclature. *DNA Cell Biol.* **12**: 1–51, 1993.
- NELSON, D. R., KOYMANS, L., KAMATAKI, T., STEGEMAN, J. J., FEYEREISEN, R., WAXMAN, D. J., WATERMAN, M. R., GOTOH, O., COON, M. J., ESTABROOK, R. W., GUNSALES, I. C., AND NEBERT, D. W.: P450 superfamily, update on new sequences, gene mapping, accession numbers and nomenclature. *Pharmacogenetics* **6**: 1–42, 1996.
- NEWTON, D. J., WANG, R. W., AND LU, A. Y. H.: Cytochrome P-450 inhibitors: evaluation of specificities in the in vitro metabolism of therapeutic agents by human liver microsomes. *Drug Metab. Dispos.* **23**: 154–158, 1995.
- NIMMO, W. S.: Drugs, disease and gastric emptying. *Clin. Pharmacokinet.* **1**: 189–203, 1976.
- NIOPAS, I., TOON, S., AND ROWLAND, M.: Further insight into the stereoselective interaction between warfarin and cimetidine in man. *Br. J. Clin. Pharmacol.* **32**: 508–511, 1991.
- OCHS, H. R., GREENBLATT, D. J., ABERDNETHY, D. R., ARENDT, R. M., GERLOFF,

- J., EICHELKRAUT, W., AND HAHN, N.: Cerebrospinal fluid uptake and peripheral distribution of centrally acting drugs: relation to lipid solubility. *J. Pharm. Pharmacol.* **37**: 428–431, 1985.
- ØIE, S., AND CHIANG, J.: Drug binding and pharmacologic activity of α_1 -adrenergic antagonists. *In* Plasma binding of drugs and its consequences, ed. by P. Belpaire, M. Bogaert, J. P. Tillement, and R. Verbeeck, pp. 159–169, Academia Press, Ghent, Belgium, 1991.
- ØIE, S., AND LEVY, G.: Effect of salicylic acid on pharmacokinetics of free and plasma protein bound bilirubin in experimental unconjugated hyperbilirubinemia. *J. Pharm. Sci.* **68**: 1–6, 1979a.
- ØIE, S., AND LEVY, G.: Effect of sulfisoxazole on pharmacokinetics of free and plasma protein bound bilirubin in experimental unconjugated hyperbilirubinemia. *J. Pharm. Sci.* **68**: 6–8, 1979b.
- ONDETTI, M. A.: Structural relationships of angiotensin-converting enzyme inhibitors to pharmacologic activity. *Circulation* **77**(suppl.): 174–178, 1988.
- O'REILLY, R. A., TRAGER, W. F., MOTLEY, C. H., AND HOWALD, W. M.: Stereoselective interaction of phenylbutazone with [$^{12}\text{C}/^{13}\text{C}$] warfarin pseudoracemates in man. *J. Clin. Invest.* **65**: 746–753, 1980.
- OSBORNE, R., THOMPSON, P., JOEL, S., TREW, D., PATEL, N., AND SLEVIN, M.: The analgesic activity of morphine 6-glucuronide. *Br. J. Clin. Pharmacol.* **34**: 130–138, 1992.
- OSIECKA, I., PORTER, P. A., BORCHARDT, R. T., FIX, J. A., AND GARDNER, C. R.: In vitro drug absorption models: I—brush border membrane vesicles, isolated mucosal cells and everted intestinal rings: characterization and salicylate accumulation. *Pharm. Res.* **2**: 284–292, 1985.
- PADGHAM, C. R. W., AND PAINE, A. J.: Altered expression of cytochrome P-450 mRNA's, and potentially of other transcripts encoding key hepatic functions, are triggered during the isolation of rat hepatocytes. *Biochem. J.* **289**: 621–624, 1993.
- PADGHAM, C. R. W., PAINE, A. J., PHILLIPS, I. R., AND SHEPPARD, E. A.: Maintenance of total cytochrome P-450 content in rat hepatocyte culture and the abundance of CYP1A2 and CYP2B1/2 in RNAs. *Biochem. J.* **285**: 929–932, 1992.
- PANG, K. S., AND CHIBA, M.: Metabolism: scaling-up from in vitro to organ and whole body. *In* Handbook of Experimental Pharmacology: Pharmacokinetics of Drugs, ed. by P. G. Welling and L. P. Balant, vol. 110, pp. 101–187, Springer-Verlag, Berlin, Germany, 1994.
- PANTUCK, E. J.: Stimulatory effect of brussel sprouts and cabbage in human drug metabolism. *Clin. Pharmacol. Ther.* **25**: 88–92, 1979.
- PAPPENHEIMER, J. R.: Contribution of solvent drag through intercellular junctions to absorption of nutrients by the small intestine of the rat. *J. Membr. Biol.* **100**: 123–136, 1987.
- PARDRIDGE, W. M.: Transport of protein-bound hormones into tissues in vivo. *Endocr. Rev.* **2**: 103–123, 1980.
- PARDRIDGE, W. M.: Development of brain-specific transport vectors: a molecular biological response. *In* Peptide Drug Delivery to the Brain, ed. by W. M. Pardridge, pp. 280–302, Raven Press, New York, 1991.
- PARK, B. K., AND KITTINGHAM, N. R.: Effects of fluorine substitution on drug metabolism: pharmacological and toxicological implications. *Drug Metab. Rev.* **26**: 625–643, 1994.
- PARK, B. K., AND KITTINGHAM, N. R.: Assessment of enzyme induction and enzyme inhibition in humans: toxicological implications. *Xenobiotics* **20**: 1171–1185, 1990.
- PARKINSON, A.: An overview of current cytochrome P-450 technology for assessing the safety and efficacy of new materials. *Toxicol. Pathol.* **24**: 45–57, 1996.
- PATCHETT, A. A., HARRIS, E., TRISTRAM, E. W., WYVRATT, M. J., WU, M. T., TAUB, D., PETERSON, E. R., IKELER, T. J., TEN BROEKE, J., PAYNE, L. G., ONDEYKA, D. L., THORSETT, E. D., GRENLEE, W. J., LOHR, N. S., HOTTSSOMMER, R. D., JOSHUA, H., RUYLE, W. V., ROTHROCK, J. W., ASTER, S. D., MAYCOCKER, A. L., ROBINSON, F. M., HIRSCHMANN, R., SWEET, C. S., ULM, E. H., GROSS, D. M., VASSIL, T. C., AND STONE, C. A.: A new class of angiotensin-converting enzyme inhibitors. *Nature (Lond.)* **288**: 280–283, 1980.
- PAUL, D., STANDIFER, K. M., INTURRISI, C. E., AND PASTERNAK, G. W.: Pharmacological characterization of morphine 6-glucuronide, a very potent morphine metabolite. *J. Pharmacol. Exp. Ther.* **251**: 477–483, 1989.
- PECK, C. C., TEMPLE, R., AND COLLINS, J. M.: Understanding consequences of concurrent therapies. *JAMA* **269**: 1550–1552, 1993.
- PFEIFFER, N.: The potential for topical carbonic anhydrase inhibitors in glaucoma therapy. *Curr. Opin. in Ophthalmology* **5**: 20–25, 1994.
- PIAFSKY, K., AND BORGÅ, O.: Plasma protein binding of basic drugs: II—importance of α_1 -acid glycoprotein for interindividual variation. *Clin. Pharmacol. Ther.* **22**: 545–549, 1977.
- PIAFSKY, K., BORGÅ, O., ODAR-CEDERLÖF, I., JOHANSSON, C., AND SJÖQVIST, F.: Increased plasma protein binding of propranolol and chlorpromazine mediated by disease-induced elevations of plasma α_1 -acid glycoprotein. *N. Engl. J. Med.* **299**: 1435–1439, 1978.
- POWIS, G.: The use of human liver for foreign compound metabolism and toxicity studies. *Drug Metab. Rev.* **20**: 379–394, 1989.
- POWIS, G., MELDER, D. C., AND WILKE, T. J.: Human and dog, but not rat, isolated hepatocytes have decreased foreign compound-metabolizing activities compared to liver slices. *Drug Metab. Dispos.* **17**: 526–531, 1989.
- PRENTIS, R. A., LIS, Y., AND WALKER, S. R.: Pharmaceutical innovation by seven UK-owned pharmaceutical companies (1964–1985). *Br. J. Clin. Pharmacol.* **25**: 387–396, 1988.
- PRICE, R. A., KEITH, R. A., SPIELMAN, R. S., AND WEINSHILBOUM, R. M.: Major gene polymorphism for human erythrocyte (RBC) thio methyltransferase (TMT). *Genet. Epidemiol.* **6**: 651–662, 1989.
- QUELLEC, A. L., DUPIN, S., TUFENKJI, A. E., GENISSEL, P., AND HOUIN, G.: Microdialysis: an alternative for in vitro and in vivo protein binding studies. *Pharm. Res.* **11**: 835–838, 1994.
- RANEY, K. D., COLES, B. F., GUENGERICH, F. P., AND HARRIS, T. M.: The endo-8,9-epoxide of alfatoxin B1: a new metabolite. *Chem. Res. Toxicol.* **5**: 333–335, 1992.
- RAPOPORT, S. I.: Transport in cells and tissues. *In* Blood-Brain Barrier in Physiology and Medicine, ed. by S. I. Rapoport, pp. 153–176, Raven Press, New York, 1976.
- REES, P. J., SELBY, P., PRENTICE, H. G., WHITEMAN, P. D., AND GRANT, D. M.: A prodrug of acyclovir with increased bioavailability. *J. Antimicrob. Chemother.* **18**(suppl. B): 215–222, 1986.
- REGÅRDH, C. G., GABRIELSSON, M., HOFFMAN, K. J., LÖFBERG, I., AND SKÅNBERG, I.: Pharmacokinetics and metabolism of omeprazole in animals and man: an overview. *Scand. J. Gastroenterol.* **20**(suppl. 108): 79–94, 1985.
- REINDENBERG, M. M., AND MARTIN, J. H.: The acetylator phenotype of patients with systemic lupus erythematosus. *Drug Metab. Dispos.* **3**: 71–73, 1974.
- REITZ, R. H., MANDRALA, A. L., AND GUENGERICH, F. P.: In vitro metabolism of methylene chloride in humans and animal tissues: use in physiologically based pharmacokinetic models. *Toxicol. Appl. Pharmacol.* **97**: 230–246, 1989.
- REITZ, R. H., MANDRALA, A. L., PARK, C. N., ANDERSON, M. E., AND GUENGERICH, F. P.: Incorporation of in vitro enzyme data into the physiologically based pharmacokinetic model for methylene chloride: implications for risk assessment. *Toxicol. Lett.* **43**: 97–116, 1988.
- REMMELE, R. P., AND BURCHELL, B.: Validation and use of cloned, expressed human drug-metabolizing enzymes in heterologous cells for analysis of drug metabolism and drug-drug interactions. *Biochem. Pharmacol.* **46**: 559–566, 1993.
- REMMER, H.: Die Beschleunigung des Evipanabbaues unter der Wirkung Von Barbituraten. *Naturwissenschaften* **8**: 189–191, 1958.
- RESETER, A., AND SPECTOR, T.: Glucuronidation of 3'-azido-3'-deoxythymidine: human and rat enzyme specificity. *Biochem. Pharmacol.* **38**: 1389–1393, 1989.
- RICE, J. M., DIWAN, B. A., WARD, J. M., NIMES, R. W., AND LUBET, R. A.: Phenobarbital and related compounds: approaches to interspecies extrapolation. *Prog. Clin. Biol. Res.* **374**: 231–249, 1992.
- RICHARDSON, K.: The discovery of fluconazole. *DN&P* **6**: 299–303, 1993.
- RIESENMAN, C.: Antidepressant drug interactions and the cytochrome P-450 system: a critical appraisal. *Pharmacotherapy* **15**: 84S–99S, 1995.
- RIMMER, E. M., BUSS, D. C., ROUTLEDGE, P. A., AND RICHENS, A.: Should we routinely measure free plasma phenytoin concentration? *Br. J. Clin. Pharmacol.* **17**: 99–102, 1984.
- ROBITZEK, E. H., SELIKOFF, I. J., AND ORNSTEIN, G. G.: Chemotherapy of human tuberculosis with hydrazine derivatives of isonicotinic acid. *Q. Bull. Sea View Hosp. N. Y.* **13**: 27–31, 1952.
- RODRIGUES, A. D.: Use of in vitro human metabolism studies in drug development: an industrial perspective. *Biochem. Pharmacol.* **48**: 2147–2156, 1994.
- ROGERS, S. M., BACK, D. J., AND ORME, L. E.: Intestinal metabolism of ethinylloestradiol and paracetamol in vitro: studies using Ussing chambers. *Br. J. Clin. Pharmacol.* **23**: 727–734, 1987.
- ROWLAND, M.: Plasma protein binding and therapeutic drug monitoring. *Ther. Drug Monit.* **2**: 29–37, 1980.
- RUBAS, W., JEZYK, N., AND GRASS, G. M.: Comparison of permeability characteristics of a human colonic epithelial (Caco-2) cell line to colon of rabbit, monkey, and dog intestine and human drug absorption. *Pharm. Res.* **10**: 113–118, 1993.
- SATOH, T.: Role of carboxylesterases in xenobiotic metabolism. *In* Review in Biochemical Toxicology, ed. by E. Hodgson, J. R. Bend, and R. M. Philot, vol. 8, pp. 155–181, Elsevier, Amsterdam, 1987.
- SAWADA, Y., HANANO, M., SUGIYAMA, Y., HARASHIMA, H., AND IGA, T.: Prediction of the volumes of distribution of basic drugs in humans based on data from animals. *J. Pharmacokin. Biopharm.* **12**: 587–596, 1984a.
- SAWADA, Y., HANANO, M., SUGIYAMA, Y., AND IGA, T.: Prediction of the disposition of β -lactam antibiotics from pharmacokinetic parameters in animals. *J. Pharmacokin. Biopharm.* **12**: 241–261, 1984b.
- SCHANKER, L. S.: On the mechanism of absorption from the gastrointestinal tract. *J. Med. Pharm. Chem.* **2**: 343–346, 1960.
- SCHUHMAN, G., FICHTL, B., AND KURZ, H.: Prediction of drug distribution in vivo on the basis of in vitro binding data. *Biopharm. Drug Dispos.* **8**: 73–86, 1987.
- SCHULTZ, P. A.: The role of genetic factors in bladder cancer. *Cancer Detect. Prev.* **11**: 379–388, 1988.
- SEGEL, I. H.: Simple inhibition systems: competitive inhibition. *In* Enzyme Kinetics: Behavior and Analysis of Rapid Equilibrium and Steady-State Enzyme Systems, ed. by I. H. Segel, pp. 100–112, John Wiley & Sons, New York, 1975.
- SELLERS, E. M.: Plasma protein displacement interactions are rarely of clinical significance. *Pharmacology* **18**: 225–227, 1979.
- SELLERS, E. M.: Drug displacement interactions: a case study of the phenylbutazone-warfarin interaction. *In* Drug-Protein Binding, ed. by M. M. Reidenberg and S. Erill, pp. 257–273, Praeger, New York, 1986.

- SESARDIC, D., BOOBIS, A. R., MURRAY, P., MURRAY, S., SEGURA, J., DE LA TORRE, R., AND DAVIES, D. S.: Furfurylline is a potent and selective inhibitor of cytochrome P450 1A2 in man. *Br. J. Clin. Pharmacol.* **29**: 651-663, 1990.
- SEYDEL, J. K., MILLER, G., AND DOUKAS, P. H.: Structure-activity correlations of sulfonamides in cell-free systems compared to correlations obtained in whole cell systems and in vivo. *In Medicinal Chemistry*, ed. by P. Pratesi, pp. 139-151, Butterworths, London, UK, 1973.
- SEYDEL, J. K., AND SCHAPER, K. J.: Quantitative structure-pharmacokinetic relationships and drug design. *In Pharmacokinetics: Theory and Methodology*, ed. by M. Rowland and G. Tucker, pp. 311-366, Pergamon Press, New York, 1986.
- SHAPIRO, B. H., AGRAWAL, A. K., AND PAMPORI, N. A.: Gender differences in drug metabolism regulated by growth hormone. *Int. J. Biochem. Cell Biol.* **27**: 9-20, 1995.
- SHARE, N. N., LOTTI, V. J., AND GAUTHERON, P.: R-enantiomer of timolol: a potential selective ocular antihypertensive agent. *Graefes Arch. Clin. Exp. Ophthalmol.* **221**: 234-238, 1984.
- SHEN, D. D., LEVY, R. H., SAVITCH, J. L., BODDY, A. V., LEPAGE, F., AND TOMBRET, F.: Comparative anticonvulsant potency and pharmacokinetics of the (+)- and (-)-enantiomers of stiripentol. *Epilepsy Res.* **12**: 29-36, 1992.
- SHEN, T. Y.: Perspectives in non-steroidal anti-inflammatory agents. *Angew. Chem. Int. Ed. Engl.* **11**: 460-472, 1972.
- SHEN, T. Y., AND WINTER, C. A.: Chemical and biological studies on indomethacin, sulindac and their analogs. *Adv. Drug Res.* **12**: 90-245, 1977.
- SHEN, W. W.: Cytochrome P-450 monooxygenase and interactions of psychotropic drugs: a five-year update. *Int. J. Psychiatry Med.* **25**: 277-290, 1995.
- SHIMADA, T., YAMAZAKI, H., MIMURA, M., INUI, Y., AND GUENGERICH, F. P.: Interindividual variations in human liver cytochrome P-450 enzymes involved in oxidation of drugs, carcinogens and toxic chemicals: studies with liver microsomes of 30 Japanese and 30 Caucasians. *J. Pharmacol. Exp. Ther.* **270**: 414-422, 1994.
- SHOU, M., GROGAN, J. O., MANCEWICZ, J. A., KRAUSZ, K. W., GONZALEZ, F. J., GELBOIN, H. V., AND KORZEKWA, K. R.: Activation of CYP3A4: evidence for simultaneous binding of two substrates in a cytochrome P450 active site. *Biochemistry* **33**: 6450-6455, 1994.
- SIDDOWAY, L. A., THOMPSON, K. A., MCALLISTER, C. B., WANG, T., WILKINSON, G. R., RODEN, D. M., AND WOOSLEY, R. L.: Polymorphism of propafenone metabolism and disposition in man: clinical and pharmacokinetic consequences. *Circulation* **75**: 785-791, 1987.
- SILBER, B., HOLFORD, N. H. G., AND RIEGELMAN, S.: Stereoselective disposition and glucuronidation of propranolol in humans. *J. Pharm. Sci.* **71**: 699-704, 1982.
- SILBERSTEIN, D. J., BOWMER, C. J., AND YATES, M. S.: Dibromosulphophthalein: its pharmacokinetics and binding to hepatic cytosol proteins in rats with acute renal failure. *Br. J. Pharmacol.* **95**: 343-352, 1988.
- SIMONYI, M.: On chiral drug action. *Med. Res. Rev.* **4**: 359-413, 1984.
- SINDRUP, S. H., BRØSEN, K., BJERRING, P., ARENDT-NIELSEN, L., LARSEN, U., ANGELO, H. R., AND GRAM, L. F.: Codeine increases pain thresholds to copper vapor stimuli in extensive but no poor metabolizers of Sparteine. *Clin. Pharmacol. Ther.* **49**: 686-693, 1991.
- SISENWINE, S. F., TIO, C. O., HADLEY, F. V., LIU, A. L., KIMMEL, H. B., AND RUELIUS, H. W.: Species-related differences in the stereoselective glucuronidation of oxazepam. *Drug Metab. Dispos.* **10**: 605-608, 1982.
- SJÖQVIST, F., AND KOIKE, Y.: Interindividual differences in drug-protein binding. *In Drug-Protein Binding*, ed. by M. M. Reidenberg and S. Erill, pp. 141-152, Praeger, New York, 1986.
- SKETT, P.: Biochemical basis of sex differences in drug metabolism. *Pharmacol. Ther.* **38**: 269-304, 1989.
- SLATTERY, J. T., AND LEVY, G.: Acetaminophen kinetics in acutely poisoned patients. *Clin. Pharmacol. Ther.* **25**: 185-195, 1979.
- SMITH, D. A.: Species differences in metabolism and pharmacokinetics: are we close to an understanding? *Drug Metab. Rev.* **23**: 355-373, 1993.
- SMITH, R. L.: Excretion of drugs in bile. *In Handbook of Experimental Pharmacology: Concepts in Biochemical Pharmacology*, ed. by B. B. Brodie and J. R. Gillette, pp. 354-389, vol. 28, Springer-Verlag, Berlin, Germany, 1971.
- SOMOGYI, A., AND GUGLER, R.: Drug interactions with cimetidine. *Clin. Pharmacokinet.* **7**: 23-41, 1982.
- SOONS, P. A., SCHELLENS, J. H. M., AND BREIMER, D. D.: Variability in pharmacokinetics and metabolism of nifedipine and other dihydropyridine calcium entry blockers. *In Pharmacogenetics of Drug Metabolism*, ed. by E. Kalow, pp. 769-790, Pergamon Press, New York, 1992.
- SRINIVAS, N. R., HUBBARD, J. W., MCKAY, C., HAWES, E. M., AND MIDHA, K. K.: In vitro hydrolysis of RR, SS-three-methylphenidate by blood esterases: differential and enantioselective interspecies variability. *Chirality* **3**: 99-103, 1991.
- STEARNS, R. A., CHAKRAVARTY, P. K., CHEN, R., AND CHIU, S. H. L.: Biotransformation of losartan to its active carboxylic acid metabolite in human liver microsomes: role of cytochrome P-4502C and 3A subfamily members. *Drug Metab. Dispos.* **23**: 207-215, 1995.
- STEARNS, R. A., MILLER, R. R., DOSS, G. A., CHAKRAVARTY, P. K., ROSEGAY, A., GATTO, G. J., AND CHIU, S. H. L.: The metabolism of Dup 753, a nonpeptide angiotensin II receptor antagonist by rat, monkey and human liver slices. *Drug Metab. Dispos.* **20**: 281-287, 1992.
- STEIN, W. D.: The molecular basis of diffusion across cell membranes. *In The Movement of Molecules Across Cell Membrane*, ed. by W. D. Stein, pp. 65-125, Academic Press, New York, 1967.
- STELLA, V. J.: Prodrugs and site-specific drug delivery. *In Xenobiotic Metabolism and Disposition*, ed. by R. Kato, R. W. Estabrook, and M. N. Cayen, pp. 109-116, Taylor & Francis, New York, 1989.
- STELLA, V. J., AND HIMMELSTEIN, K. J.: Prodrugs and site-specific drug delivery. *J. Med. Chem.* **23**: 1275-1282, 1980.
- STEVENS, J. C., SHIPLEY, L. A., CASHMAN, J. R., VANDENBRANDEN, M., AND WRIGHTON, S. A.: Comparison of human and Rhesus monkey in vitro phase I and phase II hepatic drug metabolism activities. *Drug Metab. Dispos.* **21**: 753-760, 1993.
- STEVENSON, C. L., AUGUSTIJNS, P. F., AND HENDREN, R. W.: Permeability screen for synthetic peptide combinatorial libraries using Caco-2 cell monolayers and LC/MS/MS. *Pharm. Res.* **12**: 3-94, 1995.
- STEWART, B. H., CHAN, O. H., LU, R. H., REYNER, E. L., SCHMID, H. L., HAMILTON, H. W., STEINBAUGH, B. A., AND TAYLOR, M. D.: Comparison of intestinal permeabilities determined in multiple in vitro and in situ models: relationship to absorption in humans. *Pharm. Res.* **12**: 693-699, 1995.
- STONARD, M. D., PHILLIPS, P. G. N., FOSTER, J. R., SIMPSON, M. G., AND LOCK, E. A.: α 2u-Globulin: measurement in rat kidney and relationship to hyaline droplets. *Clin. Chim. Acta.* **160**: 197-203, 1986.
- STROLIN BENEDETTI, M., AND DOSTERT, P.: Induction and autoinduction properties of rifamycin derivatives: a review of animal and human studies. *Environ. Health Perspect.* **102**(suppl. 9): 101-105, 1994.
- SUDA, H., OKAMOTO, M., AND FUKUMOTO, M.: Delayed-type skin allergic reaction in guinea pigs induced by anti-rheumatic compounds with sulfhydryl groups. *Immunopharmacol. Immunotoxicol.* **15**: 387-396, 1993.
- SUGIYAMA, Y., SAWADA, Y., IGA, T., AND HANANO, M.: Reconstruction of in vivo metabolism from in vitro data. *In Xenobiotic Metabolism and Disposition*, ed. by R. Kato, R. W. Estabrook, and M. N. Cayen, pp. 225-235, Taylor & Francis, London, 1989.
- SWARM, R. L., ROBERTS, G. K. S., LEVY, A. C., AND HINES, L. R.: Observations on the thyroid gland in rats following the administration of sulfamethoxazole and trimethoprim. *Toxicol. Appl. Pharmacol.* **24**: 351-363, 1973.
- SZUMLANSKI, C. L., SCOTT, M. C., AND WEINSHILBOUM, R. M.: Thiopurine methyltransferase pharmacogenetics: human liver enzyme activity. *Clin. Pharmacol. Ther.* **43**: 134-139, 1988.
- TANG, C., ZHANG, K., LEPAGE, F., LEVY, R. H., AND BAILLIE, T. A.: Metabolic chiral inversion of stiripentol in the rat: II—Influence of route of administration. *Drug Metab. Dispos.* **22**: 554-560, 1994.
- TAN-LIU, D. D., WILLIAMS, R. L., AND RIEGELMAN, S.: Nonlinear theophylline elimination. *Clin. Pharmacol. Ther.* **31**: 358-369, 1982.
- TAYLOR, D. C., DOWNALL, R., AND BURKE, W.: The absorption of β -adrenoceptor antagonists in rat in situ small intestine: the effect of lipophilicity. *J. Pharm. Pharmacol.* **37**: 280-283, 1985.
- TEGNER, K., BORGA, O., AND SVENSSON, I.: Protein binding of enprofylline. *Eur. J. Clin. Pharmacol.* **25**: 703-708, 1983.
- TESTA, B.: Substrate and product stereoselectivity in monooxygenase-mediated drug activation and inactivation. *Biochem. Pharmacol.* **37**: 85-92, 1988.
- TESTA, B.: Conceptual and mechanistic overview of stereoselective drug metabolism. *In Xenobiotic Metabolism and Disposition*, ed. by R. Kato, R. W. Estabrook, and M. N. Cayen, pp. 153-160, Taylor & Francis, London, UK, 1989a.
- TESTA, B.: Mechanisms of chiral recognition in xenobiotic metabolism and drug-receptor interactions. *Chirality* **1**: 7-9, 1989b.
- TESTA, B., AND TRAGER, W. F.: Racemates versus enantiomers in drug development: dogmatism or pragmatism? *Chirality* **2**: 129-133, 1990.
- TEW, K. D., HOUGHTON, P. J., AND HOUGHTON, J. A.: Modulation of p-glycoprotein-mediated multidrug resistance. *In Preclinical and Clinical Modulation of Anticancer Drugs*, ed. by K. D. Tew, P. J. Houghton, and J. A. Houghton, pp. 125-196, CRC Press, Boca Raton, FL, 1993.
- THAKKER, D. R., LEVIN, W., WOOD, A. W., CONNEY, A. H., YAGI, H., AND JERINA, D. M.: Stereoselective biotransformation of polycyclic aromatic hydrocarbons to ultimate carcinogens. *In Drug Stereochemistry: Analytical Methods and Pharmacology*, ed. by I. W. Wainer and D. E. Drayer, pp. 271-296, Marcel Dekker, Inc., New York, 1988.
- THORGEIRSSON, S. S., ATLAS, S. A., BOOBIS, A. R., AND FELTON, J. S.: Species differences in the substrate specificity of hepatic cytochrome P-448 from polycyclic hydrocarbon-treated animals. *Biochem. Pharmacol.* **28**: 217-226, 1979.
- TOBERT, J. A., CHIRILLO, V. J., AND HITZENBERGER, G.: Enhancement of uricosuric properties of indacrinone by manipulation of the enantiomers in man. *Clin. Pharmacol. Ther.* **29**: 344-350, 1981.
- TOCCO, D. J., DELUNA, F. A., DUNCAN, A. E. W., HSIEH, J. H., AND LIN, J. H.: Interspecies differences in stereoselective protein binding and clearance of MK-571. *Drug Metab. Dispos.* **18**: 388-392, 1990.
- TOCCO, D. J., DELUNA, F. A., DUNCAN, A. E. W., VASSIL, T. C., AND ULM, E. H.: The physiological disposition and metabolism of enalapril maleate in laboratory animals. *Drug Metab. Dispos.* **10**: 15-19, 1982.
- TOON, S., HOPKINS, K. J., GARSTANG, F. M., AARONS, L., SEDMAN, A., AND ROWLAND, M.: Enoxacin-warfarin interaction: pharmacokinetics and stereochemical aspects. *Clin. Pharmacol. Ther.* **44**: 32-41, 1987.
- TOON, S., AND ROWLAND, M.: Structure-pharmacokinetic relationships among the barbiturates in the rat. *J. Pharmacol. Exp. Ther.* **225**: 752-763, 1983.

- TRAGER, W. F.: Stereochemistry of P-450 catalyzed reactions. *In* Xenobiotic Metabolism and Disposition, ed. by R. Kato, R. W. Estabrook, and M. N. Cayen, pp. 171-177, Taylor & Francis, London, UK, 1989.
- TRAGER, W. F., AND TESTA, B.: Stereoselective drug disposition. *In* Drug Metabolism and Disposition, ed. by G. R. Wilkinson and D. M. Rawlins, pp. 35-61, MTP Press, Lancaster, England, 1985.
- TUCKER, G. T.: The rational selection of drug interaction studies: implications of recent advances in drug metabolism. *Int. J. Clin. Pharmacol. Ther. Toxicol.* **30**: 550-553, 1992.
- TURGEON, J., PAVLOU, H. N., WONG, W., FUNCK-BRENTANO, C., AND RODEN, D. M.: Genetically determined steady-state interaction between encaïnide and quinidine in patients with arrhythmias. *J. Pharmacol. Exp. Ther.* **255**: 642-649, 1990.
- TWISS, I. M., DE LA WATER, R., DEN HARTIGH, J., SPARIDANS, R., RAM-KOOPMANSCHAP, W., BRILL, H., WJDEVELD, M., AND VERMEIJ, P.: Cytotoxic effects of pamidronate on monolayers of human intestinal epithelial (Caco-2) cells and its epithelial transport. *J. Pharm. Sci.* **83**: 699-703, 1994.
- ULM, E. H., HICHENS, M., GOMEZ, H. J., TILL, A. E., HAND, E., VASSIL, T. C., BIOLLAZ, J., BRUNNER, H. R., AND SCHELLING, J. L.: Enalapril maleate and a lysine analogue (MK-521) disposition in man. *Br. J. Clin. Pharmacol.* **14**: 357-362, 1982.
- URIEN, S., ALBENGRES, E., PINQUIER, J. L., AND TILLEMENT, J. P.: Role of α_1 -acid glycoprotein, albumin, and nonesterified fatty acids in serum binding of apazone and warfarin. *Clin. Pharmacol. Ther.* **39**: 683-689, 1986.
- VACCA, J. P., DORSEY, B. D., SCHLEIF, W. A., LEVIN, R. B., MCDANIEL, S. L., DARKE, P. L., ZUGAY, J., QUINTERO, J. C., BLAHY, O. M., ROTH, E., SARDANA, V. V., SCHLABACH, A. J., GRAHAM, P. I., CONDRÁ, J. H., GOTLIB, L., HOLLOWAY, M. K., LIN, J. H., CHEN, I-W., VASTAG, K., OSTOVIC, D., ANDERSON, P. S., EMINI, E. A., AND HUFF, J. R.: L-735,524: an orally bioavailable human immunodeficiency virus type-1 protease inhibitor. *Proc. Natl. Acad. Sci. USA* **91**: 4096-4100, 1994.
- VAN DALEN, R., VREE, T. B., BAARS, A. M., AND TERMOND, E.: Dosage adjustment for ceftazidime in patients with impaired renal function. *Eur. J. Clin. Pharmacol.* **30**: 597-605, 1986.
- VANE, J. R.: Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nature (Lond)* **231**: 232-235, 1971.
- VERMEULAN, N. P. E.: Stereoselective biotransformation and its toxicological implications. *In* Xenobiotic Metabolism and Disposition, ed. by R. Kato, R. W. Estabrook, and M. N. Cayen, pp. 193-206, Taylor & Francis, London, UK, 1989.
- VESELL, E. S., PASSANANTI, G. T., AND JOHNSON, A. Q.: Failure of indomethacin and warfarin to interact in normal human volunteers. *J. Clin. Pharmacol.* **15**: 486-495, 1975.
- WALTER, E., AND KISSEL, T.: Heterogeneity in the human intestinal cell line Caco-2 leads to differences in transepithelial transport. *Eur. J. Pharm. Sci.* **3**: 215-230, 1995.
- WANG, C. Y.: Microsomal amidases and carboxylesterases. *In* Conjugation-Deconjugation Reactions in Drug Metabolism and Toxicity, ed. by F. C. Kauffman, pp. 161-187, Springer-Verlag, Berlin, Germany, 1994.
- WARD, S., AND BACK, D. J.: Metabolism of gestodene in human liver cytosol and microsomes in vitro. *J. Steroid. Biochem. Mol. Biol.* **46**: 235-243, 1993.
- WARD, S. A., WALLE, T., WALLE, K., WILKINSON, G. R., AND BRANCH, R. A.: Propranolol's metabolism is determined by both mephenytoin and debrisoquine hydroxylase activities. *Clin. Pharmacol. Ther.* **45**: 72-79, 1989.
- WATANABE, J., AND KOZAKI, A.: Relationships between partition coefficients and apparent volume of distribution for basic drugs. *Chem. Pharm. Bull.* **26**: 3463-3470, 1978.
- WATTENBERG, L. W., AND LEONG, J. L.: Inhibition of the carcinogenic action of 7,12-dimethylbenz(a)anthracene by β -naphthoflavone. *Proc. Soc. Exp. Biol. Med.* **128**: 940-943, 1968.
- WAXMAN, D. J.: Regulation of liver specific steroid metabolizing cytochromes P-450: cholesterol 7 α -hydroxylase, bile acid 6 β -hydroxylase and growth hormone-responsive steroid hormone hydroxylase. *J. Steroid Biochem. Mol. Biol.* **43**: 1055-1072, 1992.
- WAXMAN, D. J., DANNAN, G. A., AND GUENGERICH, F. P.: Regulation of rat hepatic cytochrome P-450: age-dependent expression, hormonal imprinting and xenobiotic inducibility of sex-specific isoenzymes. *Biochemistry* **24**: 4409-4417, 1985.
- WAXMAN, D. J., RAM, P. A., NOTANI, G., LEBLANC, G. A., ALBERTA, J. A., MORRISSEY, J. J., AND SUNDSETH, S. S.: Pituitary regulation of the male-specific steroid 6 β -hydroxylase P-450 2a (gene product IIIA2) in adult rat liver: suppressive influence of growth hormone and thyroxine acting at a pretranslational level. *Mol. Endocrinol.* **4**: 447-454, 1990.
- WEBB, D. W., RIDDER, G. M., AND ALDEN, C. L.: Acute and subchronic nephrotoxicity of *d*-limonene in Fischer 344 rats. *Food Chem. Toxicol.* **27**: 639-649, 1989.
- WEBER, W. W., LEVY, G. N., AND HEIN, D. W.: Acetylation. *In* Conjugation Reactions in Drug Metabolism, ed. by G. J. Mulder, pp. 163-191, Taylor & Francis, New York, 1990.
- WECHTER, W. J., LOUGHEAD, D. G., REISCHER, R. J., VAN GIESSEN, G. J., AND KAISER, D. G.: Enzymatic inversion at saturated carbon: nature and mechanism of the inversion of R(-)-p-isobutyl-hydratropic acid. *Biochem. Biophys. Res. Commun.* **61**: 833-837, 1974.
- WEINER, I. M., AND MUDGE, G. H.: Inhibitors of tubular transport of organic compounds. *In* Goodman and Gilman's The Pharmacological Basis of Therapeutics, ed. by A. G. Gillman, L. S. Goodman, T. W. Rall, and F. Murad, 7th ed., pp. 920-925, Macmillan Publishing Company, New York, 1985.
- WEINER, N.: Drugs that inhibit adrenergic nerves and block adrenergic receptors. *In* Goodman and Gilman's The Pharmacological Basis of Therapeutics, ed. by A. Gilman, L. S. Goodman, T. W. Rall, and F. Murad, 7th ed., pp. 181-214, Macmillan Publishing Company, New York, 1985.
- WEINER, N., AND TAYLOR, P.: Neurohumoral transmission: the autonomic and somatic motor nervous systems. *In* Goodman and Gilman's The Pharmacological Basis of Therapeutics, ed. by A. G. Gilman, L. S. Goodman, T. W. Rall, and F. Murad, 7th ed., pp. 66-99, Macmillan Publishing Company, New York, 1985.
- WEINSHILBOUM, R. M.: Methyltransferase pharmacogenetics. *In* Pharmacogenetics of Drug Metabolism, ed. by W. Kalow, pp. 179-194, Pergamon Press, New York, 1992.
- WEINSHILBOUM, R. M., AND SLADEK, S. L.: Mercaptopurine pharmacogenetics: monogenetic inheritance of erythrocyte thiopurine methyltransferase activity. *Am. J. Hum. Genet.* **32**: 651-662, 1980.
- WEISBRODT, N. W.: Motility of the small intestine. *In* Physiology of Gastrointestinal Tract, ed. by L. R. Johnson, pp. 631-663, Raven Press, New York, 1987.
- WEISER, M. M.: Intestinal epithelial cell surface membrane glycoprotein synthesis: I—an indicator of cellular differentiation. *J. Biol. Chem.* **248**: 2536-2541, 1973.
- WELLING, P. G., HUANG, H., KOCH, P. A., CRAIG, W. A., AND MADSEN, P. O.: Bioavailability of ampicillin and amoxicillin in fasted and non-fasted subjects. *J. Pharm. Sci.* **66**: 549-552, 1977.
- WERMUTH, C. G.: Designing prodrugs and bioprecursors. *In* Drug Design: Fact or Fantasy?, ed. by G. Jolles and K. R. H. Woolbridge, pp. 47-72, Academic Press, New York, 1984.
- WHITLOCK, J. P., OKINO, S. T., DOUG, L., KO, H. P., CLARKE-KATZENBERG, R., MA, Q., AND LI, H.: Induction of cytochrome P4501A1: a model for analyzing mammalian gene transcription. *FASEB J.* **10**: 809-818, 1996.
- WILK, S., MIZOGUCHI, H., AND ORLOWSKI, M.: γ -Glutamyl dopa: a kidney-specific dopamine precursor. *J. Pharmacol. Exp. Ther.* **206**: 227-232, 1978.
- WILKINSON, G. R.: Clearance approaches in pharmacology. *Pharmacol. Rev.* **39**: 1-47, 1987.
- WILKINSON, G. R., GUENGERICH, F. P., AND BRANCH, R. A.: Genetic polymorphism of S-mephenytoin hydroxylation. *Pharmacol. Ther.* **43**: 53-76, 1989.
- WILKINSON, G. R., GUENGERICH, F. P., AND BRANCH, R. A.: Genetic polymorphism of S-mephenytoin hydroxylation. *In* Pharmacogenetics of Drug Metabolism, ed. by W. Kalow, pp. 657-685, Pergamon Press, New York, 1992.
- WILKINSON, G. R., AND SHAND, D. G.: A physiological approach to hepatic drug clearance. *Clin. Pharmacol. Ther.* **18**: 377-390, 1975.
- WILLIAMS, K., AND LEE, E.: Importance of drug enantiomers in clinical pharmacology. *Drugs* **30**: 333-354, 1985.
- WILLIAMS, M., AND OLSEN, R. W.: Benzodiazepine receptors and tissue function. *In* Receptor Pharmacology and Function, ed. by M. William, R. A. Glennon, and P. B. Timmermans, pp. 385-413, Marcel Dekker, New York, 1989.
- WILSON, C. G., WASHINGTON, C., AND WASHINGTON, N.: Overview of epithelial barriers and drug transport. *In* Physiological Pharmaceutics: Biological Barriers to Drug Absorption, ed. by C. G. Wilson and N. Washington, pp. 11-20, Ellis Horwood Limited, Chichester, UK, 1989.
- WILSON, G., HASSAN, I. F., DIX, C. J., WILLIAMSON, I., SHAH, R., MACKAY, M., AND ARTURSSON, P.: Transport and permeability properties of human Caco-2 cells: an in vitro model of the intestinal epithelial cell barrier. *J. Controlled Release* **11**: 25-40, 1990.
- WILSON, K.: Sex-related differences in drug disposition in man. *Clin. Pharmacokinet.* **9**: 189-202, 1984.
- WINDMUELLER, H. G., AND SPAETH, A. E.: Intestinal metabolism of glutamine and glutamate from the lumen as compared to glutamine from blood. *Arch. Biochem. Biophys.* **171**: 662-672, 1975.
- WONG, B. K., BRUHIN, P. J., BARRISH, A., AND LIN, J. H.: Nonlinear dorzolamide pharmacokinetics in rats: concentration dependent erythrocyte distribution and drug-metabolite displacement interaction. *Drug Metab. Dispos.* **24**: 659-663, 1996.
- WOOD, A. J. J., KORNSHAUSER, D. M., WILKINSON, G. R., SHAND, D. G., AND BRANCH, R. A.: The influence of cirrhosis on steady-state blood concentration of unbound propranolol after oral administration. *Clin. Pharmacokinet.* **3**: 478-487, 1978.
- WOODSON, L. C., DUNNETTE, J. H., AND WEINSHILBOUM, R. M.: Pharmacogenetics of human thiopurine methyltransferase: kidney-erythrocyte correlation and immunotitration studies. *J. Pharmacol. Exp. Ther.* **222**: 174-181, 1982.
- WORBOYS, P. D., BRADBURY, A., AND HOUSTON, B.: Kinetics of drug metabolism in rat liver slices: II—comparison of clearance by liver slices and freshly isolated hepatocytes. *Drug Metab. Dispos.* **24**: 676-681, 1996.
- WRIGHTON, S. A., AND RING, B. J.: Inhibition of human CYP3A catalyzed 1'-hydroxy midazolam formation by ketoconazole, nifedipine, erythromycin, cimetidine and nizatidine. *Pharm. Res.* **11**: 921-924, 1994.
- WRIGHTON, S. A., AND STEVENS, J. C.: The human hepatic cytochrome P450 involved in drug metabolism. *CRC Crit. Rev. Toxicol.* **22**: 1-21, 1992.
- WRIGHTON, S. A., VANDENBRANDEN, M., STEVENS, J. C., SHIPLEY, L. A., RING, B. J., RETTIE, A. E., AND CASHMAN, J. R.: In vitro methods for assessing

- human hepatic drug metabolism: their use in drug development. *Drug Metab. Rev.* **25**: 453-484, 1993.
- YACOBI, A., AND LEVY, G.: Effect of plasma protein binding on the anticoagulant action of warfarin in rats. *Res. Commun. Chem. Pathol. Pharmacol.* **12**: 405-408, 1975.
- YACOBI, A., AND LEVY, G.: Frequency distribution of free warfarin and free phenytoin fraction values in serum of healthy human adults. *Clin. Pharmacol. Ther.* **21**: 283-286, 1977.
- YACOBI, A., UDALL, J. A., AND LEVY, G.: Serum protein binding as a determinant of warfarin body clearance and anticoagulant effect. *Clin. Pharmacol. Ther.* **19**: 552-558, 1976.
- YASUMORI, T., CHEN, L., NAGATA, K., YAMAZOE, Y., AND KATO, R.: Species differences in stereoselective metabolism of mephenytoin by cytochrome P-450 (CYP 2C and CYP 3A). *J. Pharmacol. Exp. Ther.* **264**: 89-94, 1993a.
- YASUMORI, T., NAGATA, K., YANG, S. K., CHEN, L.-S., Murayama, N., Yamazoe, Y., and Kato, R.: Cytochrome P-450 mediated metabolism of diazepam in human and rat: involvement of human CYP 2C in *N*-demethylation in the substrate concentration-dependent manner. *Pharmacogenetics* **3**: 291-301, 1993b.
- YEE, G. C., AND MCGUIRE, T. R.: Pharmacokinetic drug interactions with cyclosporin: part I. *Clin. Pharmacokinet.* **19**: 319-332, 1990.
- YUN, C. H., LEE, H. S., LEE, H., RHO, J. K., JEONG, H. G., AND GRUENGERICH, F. P.: Oxidation of the angiotensin II receptor antagonist losartan (DuP 753) in human liver microsomes: role of cytochrome P-4503A(4) in formation of the active metabolite Exp3174. *Drug Metab. Dispos.* **23**: 285-289, 1995.
- YUN, C. H., OKERHOLM, R. A., AND GRUENGERICH, F. P.: Oxidation of the antihistaminic drug terfenadine in human liver microsomes: role of cytochrome P-450 3A(4) in *N*-dealkylation and *C*-hydroxylation. *Drug Metab. Dispos.* **21**: 403-409, 1993.
- ZHANG, K., TANG, C., RASHED, M., CUI, D., TOMBRET, F., BOTTE, H., LEPAGE, F., LEVY, R. H., AND BAILLIE, T. A.: Metabolic chiral inversion of stiripentol in the rat: I—mechanistic studies. *Drug Metab. Dispos.* **22**: 544-553, 1994.
- ZINI, R., D'ATHIS, P., BARRE, J., AND TILLEMENT, J. P.: Binding of indomethacin to human serum albumin: its non-displacement by various agents, influence of free fatty acid and the unexpected effect of indomethacin on warfarin binding. *Biochem. Pharmacol.* **28**: 2661-2665, 1979.

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