Roles of Long Non-Coding RNAs on Tumorigenesis and Glioma Development

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Jong Heon Kim Cancer Cell and Molecular Biology Branch, Research Institute, National Cancer Center, 323 Ilsan-ro, Ilsandong-gu, Goyang 410-769, Korea **Tel:** +82-31-920-2204 **Fax:** +82-31-920-2206 **E-mail:** jhkim@ncc.re.kr *These two authors contributed equally to this work. More than 98% of eukaryotic transcriptomes are composed of non-coding RNAs with no functional protein-coding capacity. Those transcripts also include tens of thousands of long non-coding RNAs (IncRNAs) which are emerging as key elements of cellular homeostasis, essentially tumorigenesis steps. However, we are only beginning to understand the nature and extent of the involvement of IncRNAs on tumorigenesis. Here, we highlight recent progresses that have identified a myriad of molecular functions on tumorigenesis for several IncRNAs including metastasis-associated lung adenocarcinoma transcript 1 (MALAT1), prostate cancer associated non-coding RNA 1 (PRNCR1), prostate cancer gene expression marker 1 (PCGEM1), *H19*, and homeobox transcript antisense intergenic RNA (HO-TAIR), and several new IncRNAs for glioma development. Potential therapeutic approaches for the IncRNAs in various human diseases are also discussed.

Key Words Non-coding RNA; IncRNA; Tumorigenesis; Glioma.

INTRODUCTION

The global human genome project was launched in the 1990's and researchers expect to understand information of the whole human genome, especially genes which encode functional proteins. However, they recognized that the human genome contains only about 20000 protein-coding genes [1,2]. It is not surprising that the numbers of protein-coding genes are pretty similar in number to those of *Caenorhabditis elegans*. Because the protein-coding genes could not properly explain the human gene expression complexity in view of physiological and evolutional aspects, investigators have suspected that the potential non-protein coding but transcriptionally active genes (>98% of transcribed genes) may be fully accountable for gene expression complexity of human [3-5].

Recently, investigators categorized the non-coding RNAs (ncRNAs) as short ncRNA, mid-size ncRNA, and long non-

coding RNA (lncRNA) by their lengths, and sometimes ncRNAs are subdivided by function, loci, and post-transcriptional modification. Among these, most onco-molecular biologists are deeply interested in the short ncRNAs [e.g., microRNAs (miRNAs)] [6] and very recently lncRNAs for their research projects as an aspect of gene expression regulation [7-11]. This review, we essentially highlight and introduce our recent understanding of roles of lncRNAs in tumorigenesis, including glioma development.

LNCRNAS: MODE OF ACTION

Recent reports showed that lncRNAs have multifunctional roles on modulating embryonic pluripotency, differentiation, development, and various diseases, essentially in cancers [7-11]. Therefore, dysregulation of lncRNAs has been shown to be associated with a broad range of defects on those physiological phenomena. lncRNAs may be classified according to their mode of action and functions in cells such as, 1) mediators on signaling pathway, 2) serving as molecular decoys, 3) work as molecular guides for the ribonucleoprotein complexes to cer-

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tain specific chromatin site, and also have 4) scaffold function for the proper complex formation (Fig. 1) [11].

IncRNAs: mediators on signaling pathway

In most cases, transcription of lncRNAs is temporally regulated and the expression has tissue specificity. Moreover, their expression is modulated in response to the internal and external stimuli [12]. Therefore, lncRNAs may serve as molecular signaling mediators which modulate a certain set of gene expression temporally and even spatially (Fig. 1A).

IncRNAs: molecular decoys

lncRNAs may serve as molecular decoys which take proteins or RNAs away from a specific location. Sometimes decoy lncRNAs can serve as "sponge" to proteins (e.g., transcription factors and chromatin modifiers) and small ncRNAs (e.g., miRNAs) (Fig. 1B) [13]. This event can lead to overall transcriptome change of cells.

IncRNAs: work as molecular guides

lncRNAs can serve as the molecular guides by locating certain ribonucleoprotein complexes to a specific target site on the chromatin (Fig. 1C) [14]. The gene expression can be altered either in neighboring genes (in *cis*) or distantly located ones (in *trans*).

IncRNAs: scaffold function

lncRNAs can support the assembly of protein complexes which link the factors together to generate brand new functions (Fig. 1D). Some lncRNAs possess distinct protein-binding domains that combine each molecule together. This event may have impact on action of transcription or repression.

LNCRNAS ON TUMORIGENESIS

Generally the lncRNAs designate ncRNAs which is more than 200 nt long [7-11]. 1) Usually those modulate DNA methylation which is closely related with genome imprinting (e.g.,



Fig. 1. Schematic diagram of IncRNA action mechanisms. A: Mediators on signaling pathway: IncRNAs can serve as molecular signaling mediators which modulate certain set of gene expression in conjunction with specific transcription factors or chromatin modifiers. B: Molecular decoys: IncRNAs can serve as the molecular decoy which takes away proteins or RNAs from the specific location. C: Work as molecular guides: IncRNAs can serve as the molecular guides by locating certain ribonucleoprotein complexes to specific target site on chromatin. D: Scaffold function: IncRNAs can support the assembly of protein complexes which link the factors together to generate brand new functions. IncRNA: long non-coding RNA.

X-chromosome inactivation by lncRNA XIST) [15]. 2) Several large intergenic non-coding RNAs (lincRNAs) which locate intergenic (gene to gene) region induced by tumor suppressor p53 in response to DNA damage (e.g., lincRNA-p21) [16]. 3) The ultra-conserved region (UCR) is extremely conserved DNA sequences among species which is more than 200 nt long [17,18]. From this region the lncRNA T-UCR is transcribed actively. Although the exact physiological roles of T-UCR have not been reported yet, however investigators assumed that T-UCRs have existed from early evolution stage, and have special roles for interacting with miRNA or different distribution in tissues.

In most cases lncRNAs are involved in transcriptional regulation [19]. The lncRNAs can be classified by their action mechanisms, 1) acting in *cis*; the lncRNA transcription affects the surrounding coding gene expression, 2) modulating histone modification [14,20], and 3) acting in *trans*; XIST induces X-chromosome inactivation of female [15].

Recent reports showed that the dysfunction of lncRNAs is closely associated with tumor formation, proliferation, invasion, and metastasis [3,8,9,11]. In this review, we highlight several essential lncRNAs which are closely related with tumorigeneis. As the lncRNAs 1) metastasis-associated lung adenocarcinoma transcript 1 (MALAT1) and 2) prostate cancer associated non-coding RNA 1 (PRNCR1), prostate cancer gene expression marker 1 (PCGEM1), and as the lincRNA 3) *H19* and 4) homeobox transcript antisense intergenic RNA (HOTAIR) are discussed.

IncRNA MALAT1

The lncRNA MALAT1 is a 7-kb long, spliced non-coding RNA, also known as non-coding nuclear-enriched abundant transcript 2 (NEAT2), which is highly conserved amongst mammals and dominantly expressed in the nucleus [21]. Interestingly, a conserved tRNA-like sequence at the 3' end is cleaved off and processed to generate a short tRNA-like ncRNA, MALAT1-associated small cytoplasmic RNA (mascRNA) [22]. Moreover, MALAT1 modulates the speckle association of a subset of pre-mRNA splicing factors [23-25].

MALAT1 RNA is usually overexpressed in cancer tissues, and overexpression of MALAT1 is associated with cell hyperproliferation and metastasis [21,26]. Some reports showed that MALAT1 regulates the expression of metastasis-associated genes and MALAT1 regulates expression of cell cycle genes



Fig. 2. Potential therapeutic approaches for targeting IncRNAs. Several methods, including small interfering RNAs (siRNAs), antisense oligonucleotides (ASOs) and ribozymes or deoxyribozymes, can be used to block the function of IncRNAs. A: Synthetic double-stranded short RNA can be delivered to cells and the antisense strand of the siRNA duplex loads on to the RNA-induced silencing complex (RISC) and degrades the targeted IncRNA. B: ASOs are single-stranded, chemically modified DNA oligomers (less than 25 nt in length) that are designed to be complementary to a target IncRNA. ASOs form a heteroduplex with the target IncRNA, and RNase H recognizes the IncRNA-DNA heteroduplex and cleaves the RNA strand. C: However, ribozymes or deoxyribozymes do not dependent on the RISC, which mediates siRNA-induced degradation, or on RNase H. IncRNA: long non-coding RNA.

and performs key roles in G_1/S and mitotic progression [27].

It also known that MALAT1 controls cell cycle progression by modulating the oncogenic transcription factor *B-MYB* (*Mybl2*) [28]. Tripathi et al. [27] showed that specific deletion of MALAT1 in cells leads to activation of p53 and its target genes. MALAT1-depleted cells show cell cycle defects which are sensitive to the p53 levels, indicating that p53 is a main downstream mediator for the MALAT1 activity. In MALAT1depleted cells, replication of the S-phase is reduced, and rather cell population of G₁ and G₂/M increased. Also, expression of genes associated with cell cycle arrest (e.g., tumor suppressor p53, cdk inhibitor p21) also increases. In addition, reduced expression of oncogenic transcription factor *B-MYB* which is related with G₂/M progression, was observed in MALAT1-depleted cells [27].

IncRNAs PRNCR1 and PCGEM1

Two lncRNAs, PRNCR1 (also known as PCAT8) and PC-GEM1, are usually overexpressed in aggressive prostate cancers and have a close relationship with castration resistance and proliferation of cancer cells [29-31]. PRNCR1 and PC-GEM1, specifically interact with androgen receptor (AR) and strongly enhance androgen receptor-mediated gene activation in both ligand-dependent and -independent manner [14].

After the interaction between PRNCR1 and AR, acetylation occurs at the C-terminal of AR protein, which accelerates the association of DOT1L (disruptor of telomeric silencing 1 like histone H3 methyltransferase) to the PRNCR1-AR complex. Consequently, DOT1L mediates N-terminal acetylation of AR protein which enhances recruitment of lncRNA PCGEM1. In the castration-resistant prostate cancer cells, expression for short hairpin RNA targeting these two lncRNAs strongly suppressed proliferation of cancer cells and xenograft tumor growth in mice [14]. Taken together, the data indicate that lncRNAs are potentially essential for the castration resistance in prostate cancers. However, the specific mechanistic roles of these two lncRNAs in castration resistance during prostate cancer development are still under investigation.

lincRNA H19

The lincRNA *H19* gene encodes around 2.3-kb long transcript which does not contain any known open reading frames [32-34]. It is normally transcribed by RNA polymerase II likewise general mRNA transcripts and transported to the cytoplasm after sequential capping-polyadenylation-splicing steps. Furthermore, *H19* lincRNA is one of the most highly expressed in the placenta and evolutionarily conserved at the nucleotide level between human and rodent [34]. Interestingly, *H19* is the imprinted gene which is only transcribed from the maternally inherited allele; the paternal *H19* allele dose not expressed. *H19* gene is located on downstream of growth-promoting insulin-like growth factor 2 (*igf2*) and *H19* and *igf2* share common imprinting mechanism. Recent reports show that *H19* is closely associated with tumorigenesis and fetal growth syndrome. However, the exact physiological functions of *H19* are largely unknown yet [35-40].

Intriguingly, recent reports showed that *H19* serves as miR-NA precursor. miR-675 is located in the first exon of *H19*, and expressed through the excision from full *H19* lincRNA transcript in the placenta of gestational time point which shows to stop normal growth [41]. The overexpression of miR-675 in embryonic cell and extra-embryonic cell caused decrease of cell proliferation. The target gene of miR-675 (ex. *lgf1r*) is derepressed in *H19* null placenta. Furthermore, processing of miR-675 from *H19* is known to be regulated by stress-response RNA-binding protein HuR. HuR inhibits miR-675 processing by binding to *H19* transcript. Release of miR-675 from *H19* transcript inhibits cell proliferation rapidly in response to cellular stress or oncogenic signals [34]. miR-675 processing mechanism from *H19* seems to be associated with molecular pathology in fetal growth and tumorigenesis.

lincRNA HOTAIR

HOTAIR usually shows overexpressed patterns in early and metastatic breast cancer cells [20]. HOTAIR regulates the gene expression by interacting with polycomb repressive complex 2 (PRC2) and lysine-specific demethylase 1A. Together with these two enzymes, HOTAIR can control methylation and demethylation status of histones [20]. PRC2 seems to be more important related with cancer because cancer cells which contain lysine-27 methylated histone H3 showed similar gene expression of embryonic fibroblasts. Generally, PRC2 expression induces metastasis of cancer cells. However metastasis can be suppressed when PRC2 is too overexpressed.

LNCRNAS ON GLIOMA DEVELOPMENT

A glioma is a type of tumor that arises from glial cells mostly in the human brain. High-grade gliomas usually have a tendency to infiltrate into the extracellular matrix of the brain and this trait makes it difficult to perform surgery and radio-therapy [42]. Therefore the understanding of the molecular mechanism of infiltrative phenotype of gliomas and identification of key regulator(s) for invasion are essential for the efficient treatment of this hardly curable disease [43,44].

Recent reports showed that several lncRNAs have close relationship with glioma development. Some lncRNAs that may contribute to brain development and certain specific differentially expressed lncRNAs may play an important role in the pathogenesis of glioblastoma multiforme [45]. The most highly upregulated lncRNA is Colorectal Neoplasia Differentially Expressed (CRNDE) [46]. The importance of CRNDE and its roles in specialized processes such as brain function have not been addressed yet.

Another lncRNA, maternally expressed gene 3 (MEG3), has been found that it markedly decreased in glioma tissues compared with adjacent normal tissues [47]. Moreover, overexpression of the lncRNA MEG3 in human glioma cell lines inhibits cell proliferation and promotes cell apoptosis. Therefore MEG3 might have inhibitory role in glioma development and can serve as potential drug in anti-glioma therapy.

The lincRNA *H19* and its derivative miR-675 were positively correlated with glioma grades. Moreover, *H19*-derived miR-675 regulated cadherin 13 which is the directly target of miR-675, thereby modulating glioma cell invasion [48]. The potential oncogenic role of lincRNA *H19*/miR-675 may serve as development of anti-glioma therapy.

THERAPEUTIC APPROACHES FOR TARGETING LNCRNAS

Various therapeutic approaches for the lncRNAs have been developed and several pharmaceutical companies are also actively developing lncRNA-targeting therapeutics [49,50]. First, the small interfering RNAs (siRNAs) against the specific lncRNA can be used for the strategies to regulate lncRNA function (Fig. 2A). In most cases, predominant localization of lncRNAs is in the nucleus, and thus siRNAs may be less accessible to lncRNAs than mRNAs. However, successful knockdown of lncRNAs have been reported by many researchers irrespective of their subcellular localization.

Other strategies [e.g., antisense oligonucleotides (ASOs) as well as ribozymes or deoxyribozymes] can be adapted to directly target lncRNAs when an overall secondary structure or the nucleotide sequence is not favorable for the optimized design of siRNAs (Fig. 2B). Antisense oligonucleotides have advantages over siRNAs including independence of RNA-induced silencing complex (RISC) machinery, high specificity, and low off-target effects. Recent reports showed that MALAT1 function on lung cancer cells in mouse successfully inhibited by using ASOs [22,23,27].

Ribozymes or deoxyribozymes (e.g., hammerhead ribozyme) bind to a target sequence complementarily and promote the cleavage of the flanking RNA region (Fig. 2C). These may be useful for the targeting of lncRNAs that are not favorable for optimal siRNA design [49,50].

CONCLUSION

Recent global analysis showed that cancer transcriptome is

more complex than previously expected. Dysregulated expression of lncRNAs, including protein-coding genes and miRNAs have potential pervasive roles as the driver of human cancers and development and progression of the cancers [3,8,11]. The epigenomic reprogramming by lncRNAs can be applicable to many other human diseases characterized by aberrant lncRNA expression [51]. Therefore, in the context of cancer cells, ectopic expression or specific knockdown of lncRNAs such as PRNCR1, PCGEM1, and HOTAIR seems to re-impose that chromatin state, thereby enabling gene expressions are more favorable or unfavorable to the mobilization and matrix invasion of cancer cells.

As the non-epigenomic regulation, such as regulation of alternative splicing (e.g., MALAT1) [23] and generation of miR-NA precursor (e.g., lincRNA *H19*) [41] lncRNAs can modulate gene expression more favorable to tumor development.

Therefore, understanding the precise molecular mechanisms of lncRNAs to the various biological processes will be a critical step in exploring new strategies in future cancer therapy.

Conflicts of Interest.

The authors have no financial conflicts of interest.

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