

# Salicylic acid-induced abiotic stress tolerance and underlying mechanisms in plants

M. Iqbal R. Khan<sup>1</sup>, Mehar Fatma<sup>1</sup>, Tasir S. Per<sup>1</sup>, Naser A. Anjum<sup>2\*</sup> and Nafees A. Khan<sup>1</sup>

<sup>1</sup> Department of Botany, Aligarh Muslim University, Aligarh, India, <sup>2</sup> Centre for Environmental and Marine Studies, Department of Chemistry, University of Aveiro, Aveiro, Portugal

## OPEN ACCESS

### Edited by:

Zuhua He,  
Shanghai Institute for Biological  
Sciences – Chinese Academy  
of Sciences, China

### Reviewed by:

Vasileios Fotopoulos,  
Cyprus University of Technology,  
Cyprus  
Fang Chen,  
Yale University, USA

### \*Correspondence:

Naser A. Anjum,  
Centre for Environmental and Marine  
Studies, Department of Chemistry,  
University of Aveiro, 3810-193  
Aveiro, Portugal  
anjum@ua.pt

### Specialty section:

This article was submitted to  
Plant Physiology,  
a section of the journal  
Frontiers in Plant Science

**Received:** 23 January 2015

**Accepted:** 11 June 2015

**Published:** 30 June 2015

### Citation:

Khan MIR, Fatma M, Per TS,  
Anjum NA and Khan NA (2015)  
Salicylic acid-induced abiotic stress  
tolerance and underlying mechanisms  
in plants.  
Front. Plant Sci. 6:462.  
doi: 10.3389/fpls.2015.00462

Abiotic stresses (such as metals/metalloids, salinity, ozone, UV-B radiation, extreme temperatures, and drought) are among the most challenging threats to agricultural system and economic yield of crop plants. These stresses (in isolation and/or combination) induce numerous adverse effects in plants, impair biochemical/physiological and molecular processes, and eventually cause severe reductions in plant growth, development and overall productivity. Phytohormones have been recognized as a strong tool for sustainably alleviating adverse effects of abiotic stresses in crop plants. In particular, the significance of salicylic acid (SA) has been increasingly recognized in improved plant abiotic stress-tolerance via SA-mediated control of major plant-metabolic processes. However, the basic biochemical/physiological and molecular mechanisms that potentially underpin SA-induced plant-tolerance to major abiotic stresses remain least discussed. Based on recent reports, this paper: (a) overviews historical background and biosynthesis of SA under both optimal and stressful environments in plants; (b) critically appraises the role of SA in plants exposed to major abiotic stresses; (c) cross-talks potential mechanisms potentially governing SA-induced plant abiotic stress-tolerance; and finally (d) briefly highlights major aspects so far unexplored in the current context.

**Keywords:** abiotic stress, crop-loss, phytohormones, salicylic acid, stress tolerance mechanisms

## Introduction

### Abiotic Stresses and Salicylic Acid as a Major Stress Impact-Mitigation Tool in Plants

Abiotic stress has now been recognized as the biggest and potential threat for agricultural productivity all over the world. Nevertheless, anthropogenic activities in the developmental era have aggravated the degradation of agricultural system and its productivity due to major abiotic stresses such as such as metals/metalloids (hereafter termed as ‘metal/s’), salinity, ozone, UV-B radiation, extreme temperatures, nutrient (deficiency and excess), and drought (Khan and Khan, 2013; Anjum et al., 2014a). In fact, the imposed abiotic stresses can potentially influence almost all physiological, biochemical, and molecular processes in plants from the early stage of seed germination to maturity, and eventually cause severe losses in the economic yield of crop plants. It has been projected that abiotic stresses may adversely affect 70% yield of staple food crops (Kaur et al., 2008; Mantri et al., 2012). In one of the estimates of U.S. Environmental Action Group, the toxicity caused by varied metals has been one of the major concerns to the

agriculture system and health of more than 10 million people in eight countries (such as China, Dominican Republic, India, Kyrgyzstan, Peru, Russia, Ukraine, and Zambia; ENS, 2006). In addition, increasing salinization of arable land is expected to result in 30% land-loss by the end of 2028 and 50% by mid of the 21st century (Wang et al., 2003). Notably, the rise in CO<sub>2</sub> concentration by about 500–1000 ppm by the year 2100 has been projected to cause an increase in the mean temperature by approximately 3°C (Khan et al., 2013a). Hence, together with salinization of arable land, drought, nutrients-unavailability, metal toxicity, and climate change have been expected to significantly aggravate the problem (Anjum et al., 2014a).

Plant growth regulators play important roles in the regulation of plant developmental processes and signaling networks as they are involved either directly or indirectly in a wide range of biotic and abiotic stress responses and tolerance in plants (Khan et al., 2012a,b,c; Asgher et al., 2015). Salicylic acid (SA) is a phenolic compound involved in the regulation of growth and development of plants, and their responses to biotic and abiotic stress factors (Raskin, 1992; Khan et al., 2012a,b,c, 2013b; Miura and Tada, 2014). SA is involved in the regulation of important plant physiological processes such as photosynthesis, nitrogen metabolism, proline (Pro) metabolism, production of glycinebetaine (GB), antioxidant defense system, and plant-water relations under stress conditions and thereby provides protection in plants against abiotic stresses (Khan et al., 2010, 2012a,b,c, 2013b, 2014; Nazar et al., 2011; Miura and Tada, 2014). Apart from its involvement in the induction of defense-related genes and stress resistance in biotic stressed plants (Kumar, 2014), SA has been shown to improve plant tolerance to major abiotic stresses such as metal (Zhang et al., 2015), salinity (Khan et al., 2014; Nazar et al., 2015), osmotic (Alavi et al., 2014), drought (Fayez and Bazaid, 2014), and heat stress (Khan et al., 2013b). Exogenously sourced SA to stressed plants, either through seed soaking, adding to the nutrient solution, irrigating, or spraying was reported to induce major abiotic stress tolerance-mechanisms (Horváth et al., 2007; Khan et al., 2012a,b,c, 2013b, 2014; Anwar et al., 2013; Palma et al., 2013). SA influences plant functions in a dose dependent manner, where induced or inhibited plant functions can be possible with low and high SA concentrations, respectively. For example, in *Matricaria chamomilla*, 50 and 250 μM SA concentrations were reported to, respectively, promote and inhibit growth (Kováčik et al., 2009). In another instance, 0.1 and 0.5 mM SA promoted photosynthesis and growth of *Vigna radiata* but an inhibited growth was evidenced with 1.0 mM SA (Nazar et al., 2011). Besides the concentration of SA, the duration of the treatment, plant species, age, and treated plant organ can also influence the SA-effects in plants (Shi et al., 2009; Miura and Tada, 2014). Recent molecular studies have established that SA can regulate many aspects in plants at gene level, and thereby can improve plant-abiotic stress tolerance. SA was reported to induce several genes responsible for encoding chaperone, heat shock proteins (HSPs), antioxidants, and secondary metabolites [sinapyl alcohol dehydrogenase (SAD), cinnamyl alcohol dehydrogenase (CAD), and cytochrome P450; Jumali et al., 2011]. Additionally,

SA-involvement in mitogen-activated protein kinase (MAPK) regulation, and in the expression and activation of *non-expressor of pathogenesis-related genes 1 (NPR1)* has been evidenced (Chai et al., 2014). Nevertheless, the transcriptional reprogramming that occurs during the plant defense response against biotic and abiotic stress was reported to be modulated by SA, where the transcription of different sets of defense genes can be controlled in a spatio-temporal manner via SA-mediated mechanisms (Herrera-Vásquez et al., 2015).

In order to develop ideotypes for sustainable agriculture as well as to improve overall plant performance in the conditions of the changing climate and the increased severity of abiotic stresses, it would be imperative to exploit the information available on the involvement of SA in abiotic stress tolerance in plants. In the current effort, in addition to overviewing historical background and biosynthesis of SA in plants under both optimal and stressful environments, the role of SA in plants exposed to major abiotic stress conditions is critically discussed, potential mechanisms controlling SA-induced plant abiotic stress-tolerance are cross-talked, and major aspects so far unexplored in the current context are briefly highlighted.

## Salicylic Acid: Historical Background and Biosynthesis

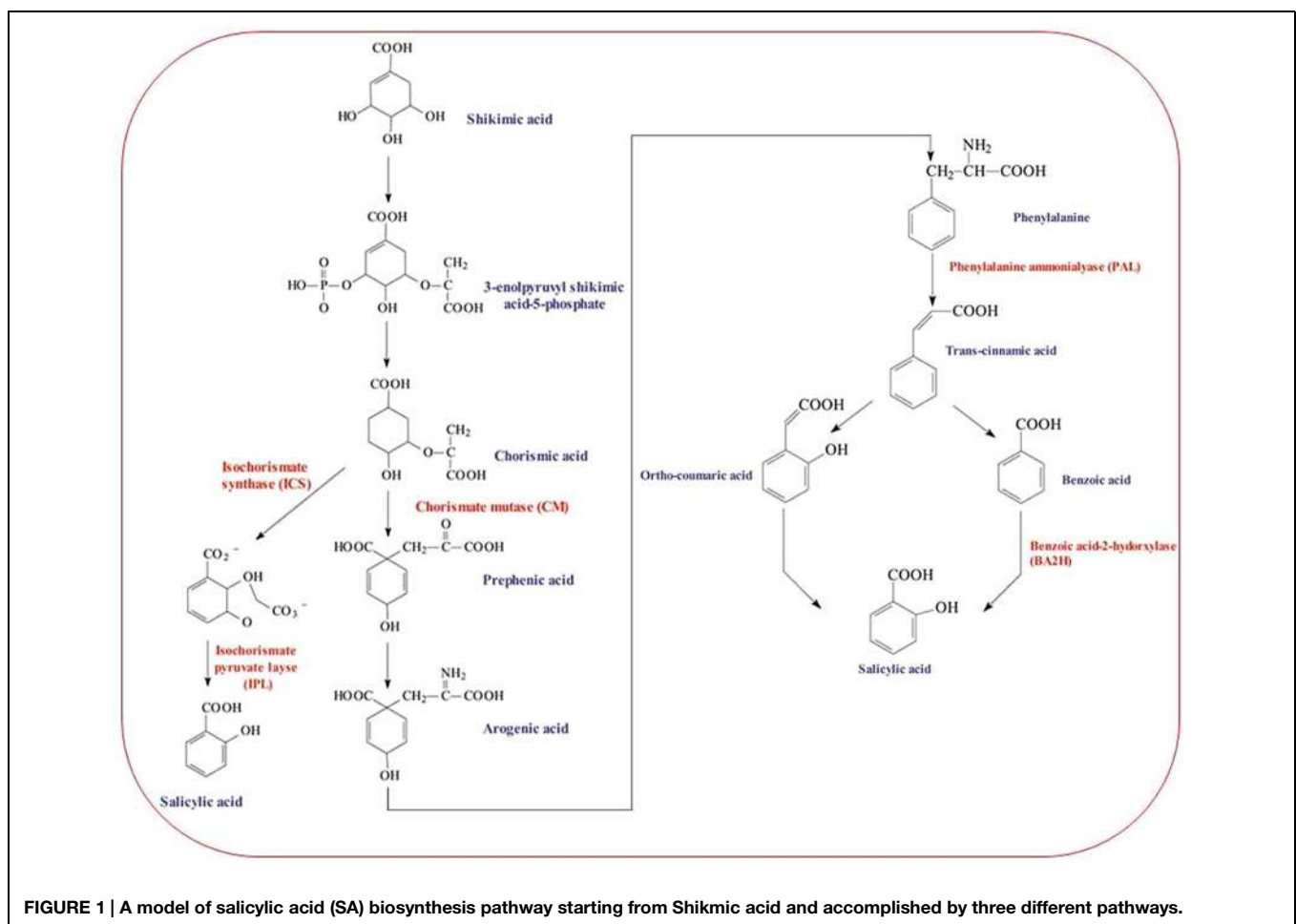
Centuries ago Americans, Indians, and Greeks used willow tree bark and leaves to cure aches and fevers, but it has been documented that Hippocrates prescribed the compound to relieve pain for women during child birth and fever was later recognized as SA. Ancient texts also indicate that Babylonians, Assyrians, and Chinese (Sharp, 1915) used willow (*Salix* sp.) bark and leaves for medicinal purposes. The importance of willow (*Salix* sp.) bark has also been shown in 1763 when the Reverend Edward Stone informed the Royal Society that it contained substances that relieved the symptoms of “ague” (probably malarial fever) effectively. Until 19th century the active principle of willow was not known, but later on the salicylates, methyl salicylate (MeSA), saligenin (alcohol of SA), and their glycosides were isolated from extracts of willow and other plants. The SA was isolated and purified from willow and meadowsweet in the first half of the 19th century (Leroux, 1830) and was chemically synthesized by the carboxylation of sodium phenoxide in 1860 (Kolbe and Lautemann, 1860). In 1928, Johann Buchner, a German scientist for the first time isolated SA from willow bark under the name “salicin” (the glucoside of salicyl alcohol), which was the major salicylate in willow bark (Weissmann, 1991). The name SA (Cleland and Ajami, 1974) is originated from the Latin word *Salix* and was given to the active ingredient of willow (*Salix* sp.) bark by Raffaele Piria in 1838. The first commercial production of synthetic SA began in Germany in 1874. Aspirin, a trade name for acetylsalicylic acid, was introduced by the Bayer Company in 1898 and rapidly became one of world’s best-selling drugs and it replaced the use of SA by producing less gastrointestinal irritation yet has similar medicinal properties. With the further advancement in scientific research in 20th century, uses of SA as a treatment for acne, psoriasis, warts, and

calluses became common. The use of SA as a skin softener, for removal of dead skin cells, dirt, oil and debris and cleaning of pores is widespread. In spite of the fact that the mode of medicinal action of salicylates is a subject of continual debate, they are being used to treat human diseases ranging from the common cold to heart attacks. As aspirin undergoes spontaneous hydrolysis to SA, the two compounds have similar effects in plants.

Salicylic acid is a seven carbon (C) containing, naturally occurring phenolic compound and endogenously synthesized signaling molecule in plants. The shikimic acid pathway and the malonic acid pathway are the two main pathways known to be involved in the synthesis of plant phenolics. The shikimic acid pathway takes part in the biosynthesis of most plant phenolic compounds. It simply converts simple carbohydrate precursors derived from glycolysis and pentose phosphate pathway to the aromatic amino acids including SA precursor, phenylalanine (Herrmann and Weaver, 1999). The most common pathway in plants for SA synthesis is phenylalanine pathway; however, SA biosynthesis may also be accomplished by isochorismate pathway (Kawano et al., 2004; Mustafa et al., 2009). SA is produced after a series of chemical reactions catalyzed by many enzymes. The hydroxylation of benzoic acid catalyzed by enzyme benzoic acid 2-hydroxylase synthesizes SA. Benzoic acid is synthesized

by cinnamic acid either via  $\beta$ -oxidation of fatty acids or a non-oxidative pathway (Verberne et al., 1999; Mustafa et al., 2009). Cinnamate 4-hydroxylase (C4H) catalyzes the second step in the pathway during the conversion of cinnamic acid to coumaric acid. Cinnamic acid is produced from phenylalanine by the action of enzyme phenylalanine ammonia-lyase (PAL). Cinnamic acid is hydroxylated to form coumaric acid followed by oxidation of the side chain, and further hydroxylated and form SA. In the third pathway, SA biosynthesis in plants has been reported from shikimic acid via chorismic acid and coumaric acid (Horváth et al., 2007; An and Mou, 2011; **Figure 1**).

Multiple abiotic stress factors have been evidenced to modulate major enzymes involved in plant-SA biosynthesis. Overproduction of SA via enhanced activity of SA biosynthetic pathway enzymes (mainly isochorismate synthase, ICS; PAL) in plants helps in their protection against environmental stresses. These enzymes are key regulators of SA functions and are known to be modulated by different abiotic and biotic stress factors. In *Arabidopsis*, ICS has been found to be involved in the biosynthesis of SA during plant defense process (Wildermuth et al., 2001). Synthesis of SA following exposure to ozone in *Arabidopsis* was also suggested to involve ICS (Ogawa et al., 2005). SA application can positively regulate *ICS1* and improve drought tolerance in



plants such as *Arabidopsis thaliana* (Hunter et al., 2013). In addition, mutant study on *Arabidopsis* (*ics1* and *ics2*) confirmed that in the absence of *ICS1*, *ICS2* can encode a functional ICS enzyme but in limited amounts (Garcion et al., 2008). Under salt stress, high expression of ICS and C4H enzymes can be correlated to the level of applied SA concentration. To this end, with 1.0 mM SA, induction of ICS and C4H was higher than that observed with 0.1 mM in *Carthamus tinctorius*. The isolated genes of ICS and C4H in *C. tinctorius* were considered significant for both salt stress tolerance and pathogen resistance (Dehghan et al., 2014). The involvement of enhanced PAL activity in heat and chilling tolerance was documented in *Musa* plants (Chen et al., 2008). Both Cd and Pb were reported to induce mRNA coding for PAL in *Glycine max* (Pawlak-Sprada et al., 2011). Isolation and profiling of PAL and CHS in salinity exposed *C. tinctorius* plants suggested that these two enzymes were highly responsive to 1.0 mM SA (vs. 0.1 mM SA; Dehghan et al., 2014). Recently, both water deficit and UV-B radiation were reported to cause SA accumulation as a result of increased activity of PAL and benzoic acid hydroxylase (BA2H; Bandurska and Cieslak, 2013).

## Salicylic Acid and Abiotic Stress-Tolerance in Plants

### Metals/Metalloids

Metal/metalloid-accrued stress has become a subject of great concern to sustainable agriculture and environmental system. Agricultural soils can receive metal pollutants from multiple pathways including industrial effluents, burning liquid and solid fuel, urban run-off, sewage waste disposal, agricultural toxic chemicals run-off, domestic garbage dump etc. Literature is full on the significance of SA in the minimization of metal-accrued stress-impacts. Exogenously applied SA was reported to improve growth and photosynthetic traits in several crop plants including lead [Pb—(0.05, 0.15, 0.25 mmol L<sup>-1</sup>)] exposed *Oryza sativa* (Chen et al., 2007), Cd (10, 15, and 25 μM)-exposed *Zea mays* (Krantev et al., 2008), and Cu (0.05, 0.10, 0.15, and 0.20 mM)-exposed *Phaseolus vulgaris* (Zengin, 2014). Applied SA was evidenced to modulate antioxidant system-components and significantly decrease membrane lipid peroxidation in to Cu-exposed *P. vulgaris* (Zengin, 2014) and Pb-exposed *O. sativa* (Chen et al., 2007). Recent evidences also suggested that SA is an important regulator of photosynthesis, photosystem II (PSII), photosynthetic pigments, and the activity of enzymes such as Rubisco and carbonic anhydrase under metal stress (Al-Whaibi et al., 2012; Noriega et al., 2012; Zhang et al., 2015). Recently, an increased tolerance of *Linum usitatissimum* to Cd was attributed to SA-mediated control of H<sub>2</sub>O<sub>2</sub> accumulation (Belkadhi et al., 2014). Ni-impact-mitigation role of SA can differ in plants exhibiting a differential Ni-accumulation. To this end, SA-biosynthetic pathway compounds and its derivative metabolites (phenylalanine, cinnamic acid, salicyloyl-glucose, and catechol) were elevated in the hyperaccumulator *Thlaspi goesingense* compared to the non-accumulators *A. thaliana* and *T. arvense* (Freeman et al., 2005). Further, the presence of

SA enhanced the activity of sulfur (S) assimilation pathway enzyme (serine acetyltransferase; SAT), glutathione (GSH) content and eventually increased Ni-resistance. Exogenous SA (3.0 mM) enhanced *OsWRKY45* gene expression and increased endogenous content of SA. It has been shown that increased endogenous SA level prevented membrane damage by lowering H<sub>2</sub>O<sub>2</sub> content in Cd-exposed *O. sativa* (Chao et al., 2010). Involvement of SA (0.5 mM) in the phytochelatin (PCs)-mediated protection of *Z. mays* was evidenced against Cd-toxicity (Szalai et al., 2013). SA can also significantly inhibit Fe-deficiency-caused chlorosis in plants (Kong et al., 2014).

### Salinity Stress

It has been estimated that about 45 million hectares of irrigated land have been damaged by salinity stress worldwide and considerable area of land affected by salinity is increasing day by day worldwide (Pitman and Lauchli, 2002; Munns and Tester, 2008). In fact, the loss of plant productivity due to salinity stress is a consequence of imbalance in cellular ionic and osmotic balances (Khan et al., 2012b). Major adverse effects of salinity stress include increased ion toxicity, osmotic stress, and nutrient-acquisition and homeostasis/deficiency, impaired stomatal conductance, increased cell-turgor loss, decreased reduction in leaf water potential, altered physiological/biochemical processes, and elevated ROS-caused oxidative stress (Munns and Tester, 2008; Nazar et al., 2011; Khan et al., 2014). The role of SA in strengthening salinity stress-tolerance mechanisms has been extensively evidenced in many crops including *Vicia faba* (Azooz, 2009), *Brassica juncea* (Nazar et al., 2011, 2015), *Medicago sativa* (Palma et al., 2013), and *V. radiata* (Khan et al., 2014).

Salicylic acid was reported to induce salinity tolerance and increased biomass of *Torreya grandis* as a result of enhanced chlorophyll content and the activity of antioxidant enzymes that eventually activated the photosynthetic process and alleviated oxidative stress (Li et al., 2014). SA-deficiency in plants was considered as a major reason of salinity-accrued increased damages and diminished activity of antioxidant enzymes in SA-deficient *NahG* transgenic of *Arabidopsis* lines (Cao et al., 2009). NaCl-accrued oxidative stress in *Hordeum vulgare* was minimized by SA (50 μM)-mediated decrease in cellular malondialdehyde (MDA, a marker of membrane-lipid peroxidation) and ROS (such as H<sub>2</sub>O<sub>2</sub>; Fayez and Bazaid, 2014; Khan et al., 2014). SA-priming can be an important strategy for enhancing major GSH-based H<sub>2</sub>O<sub>2</sub>-metabolizing enzyme such as GST. To this end, SA mitigated salinity stress-injury in *Solanum lycopersicum* by causing characteristic changes in the expression pattern of GST-gene family members such as *SIGSTT2*, *SIGSTT3*, *SIGSTF4* (Csiszár et al., 2014). Exogenously sourced SA (0.5 mM) was reported to improve salt tolerance in *Triticum aestivum* due to an enhanced transcript level of antioxidant genes; *GPX1*, *GPX2*, *DHAR*, *GR*, *GST1*, *GST2*, *MDHAR*, and *GS*, and an increased activity of ascorbate (AsA)-GSH pathway enzymes (Li et al., 2013). In another instance, SA-mediated restoration of membrane potential and prevention of salt-induced K<sup>+</sup>-loss via a GORK channel, and eventually improved salinity-tolerance were evinced in *A. thaliana* (Jayakannan et al., 2013).



## Ozone Stress

Ozone is among the major components of photochemical air pollution responsible for causing significant damage to both cultivated plants and forest trees (Wang et al., 2007). Ozone enters to mesophyll cells via stomata where it immediately interacts with water and other cellular components to generate phytotoxicity mainly by elevating the generation of ROS (such as  $O_2^-$ ,  $H_2O_2$ , OH, and  $^1O_2$ ), and triggering a series of signaling cascades and plant defense responses (Long and Naidu, 2002; Ashmore, 2005). Extensive reports are available on the key regulatory roles of SA in plant-ozone stress tolerances (Yalpani et al., 1994; Sharma et al., 1996; Yoshida et al., 2009; Khan et al., 2012a; Pál et al., 2014). SA can work as a signal molecule and promote molecular and physiological changes in ozone-exposed plants (Tamaoki, 2008). Moreover, SA is required to potentiate the antioxidant response, maintain cellular redox state, and activate processes against hypersensitive cell death and ozone-sensitivity (Rao and Davis, 1999). SA was reported to maintain plant growth, development and cellular redox system by activating the GSH-biosynthetic pathway (Yoshida et al., 2009).

In ozone-exposed *A. thaliana*, SA was involved in the accumulation of defense-related transcripts and induced resistance (Sharma et al., 1996). SA is also involved in signaling network integrating other phytohormones such as JA and/or ethylene in ozone-exposed plants (Rao et al., 2002). Notably, both SA and ethylene were evidenced by these authors to act in concert to regulate ozone-induced cell death in *A. thaliana*. In the same plant, ozone-mediated induced biosynthesis of JA or methyl jasmonate was reported to attenuate SA-dependent lesion-initiation that eventually was considered as a major factor for the decreased lesions caused by ozone (Rao et al., 2000). It has also been evidenced that ozone-accrued SA-accumulation can be promoted by ethylene-mediated regulation of the expression of the PAL and chorismate mutase (CM) genes in ozone-exposed *Nicotiana tabacum* (Ogawa et al., 2005). In another study, ozone-sensitivity in hybrid poplar was correlated with insensitivity to both SA and jasmonic acid, where these phytohormones were associated with the programmed cell death in lesion formation (Koch et al., 2000).

## UV-B Radiation

The effects of increased UV-B radiation (280–320 nm) on plant growth and development raised concerns on the need of protection mechanisms (Caldwell et al., 2007; Gill et al., 2015). Increased UV-B level can significantly diminish crop productivity by inhibiting PSII, electron transport systems, photosynthetic rate, photosynthetic pigments, nucleic acids, and biomass accumulation and partitioning (Sullivan and Teramura, 1989; Mohammed and Tarpley, 2009a,b, 2011). Exposure to UV-B radiation can decrease the expression and synthesis of key photosynthetic proteins such as chlorophyll a/b-binding proteins (Lhcb) and the D1 polypeptide of PSII (*psbA*; Jordan, 1996; Jordan et al., 1998). Significant induction in SA-accumulation was evidenced in UV radiation-exposed plants (Fodor et al., 1997; Horváth et al., 2002).

Exogenously applied SA was reported to modulate antioxidant levels, detoxify superoxide radicals, prevent oxidative damage,

and protect membranes and important metabolic enzymes (Mohammed and Tarpley, 2009a). SA-mediated activation of antioxidant enzymes was considered as a major factor for SA-mediated regulation of UV-induced oxidative stress in *Capsicum annuum* leaves (Mahdavian et al., 2007). Increased photosynthetic rate, pollen viability, leaf phenolic concentration and yield in UV-B stressed-*O. sativa* was also reported (Mohammed and Tarpley, 2013). SA can decrease UV-B caused chromosome aberration level in the meristematic root tip cells (Ranceliene and Vyšniauskienė, 2012). Exogenously applied SA was reported to significantly improve photosynthetic function and its related variables in UV-B exposed plants (Karioti et al., 2008; Li et al., 2014). Earlier, UV-C radiation was also reported to upregulate the transcription of the *SA induction deficient 2* gene coding for the SA biosynthetic isochorismate synthase 1 enzyme (Martinez et al., 2004). Additionally, SA-dependent pathway was reported to control the up-regulation of the pathogenesis-related (PR) proteins (*PR-1*, *PR-2*, and *PR-5*) in UV-B-exposed transgenic *NahG A. thaliana* plants (Surplus et al., 1998).

## Temperature Stress

In the current changing environmental scenario, both low (cold and chilling stress) and high (heat) temperatures have become a potential abiotic stress-threat to crop plants. Temperature stress affects many plant-physiological and biochemical processes and induces molecular mechanisms and gene expression to modulate plants responses (Larkindale and Knight, 2002; Khan et al., 2013a,b; Kazemi-Shahandashti et al., 2014; Siboza et al., 2014). SA-supplementation has been reported to differentially benefit several plant species exposed to low/chilling temperatures (Janda et al., 1999; Ding et al., 2002; Horváth et al., 2002; Kang et al., 2012; Kazemi-Shahandashti et al., 2014; Siboza et al., 2014) and high (He et al., 2002; Larkindale and Knight, 2002; Clarke et al., 2004; Shi et al., 2006; Wang and Li, 2006; Wang et al., 2010; Khan et al., 2013a,b). SA (0.5 mM) modulated antioxidant enzymes (such as ascorbate peroxidase, APX; superoxide dismutase, SOD; guaiacol peroxidase, GPOX; GSH reductase, GR) and improved chlorophyll fluorescence in *Z. mays* under low (2°C) temperature stress (Janda et al., 1999). Exogenously SA can inhibit the activity of isozymes (CAT-1 and CAT-2) of catalase (CAT) which in turn can mediate responses of *Z. mays* to low temperature stress (Horváth et al., 2002).

Salicylic acid (2.0 mM)-mediated increased synthesis of total phenolics and the activity of PAL were reported to improve chilling tolerance in cold-stored lemon fruit (*Citrus limon*; Siboza et al., 2014). Mutlu et al. (2013) reported that exogenously sourced SA results in cold tolerance by enhancing antioxidant enzymes, ice nucleation activity, and the patterns of apoplastic proteins in *H. vulgare* genotypes. In another report, SA significantly protected ultra-structures in *Musa acuminata* seedlings under chilling stress (Kang et al., 2012). Least reports are available in literature on the molecular mechanisms underlying SA-mediated improved plant tolerance to cold/chilling temperature. Increased chilling tolerance was evidenced in chilling-exposed *S. lycopersicum* fruit as a result of low concentrations of (0.01 mM) MeSA-mediated induction in the synthesis of some stress proteins, such as PR proteins

(Ding et al., 2002). The applied MeSA significantly increased accumulation of *PR-2b* and *PR-3a* mRNAs but slightly increased *PR-3b* mRNA accumulation (Ding et al., 2002).

Salicylic acid-mediated improved plant tolerance to heat stress has also been reported (He et al., 2002; Larkindale et al., 2005; Wang et al., 2010; Khan et al., 2013a,b). Larkindale and Knight (2002) reported that the transgenic *Arabidopsis* seedlings showing a bacterial SA-decomposing salicylate hydroxylase were less tolerant to heat stress. SA signaling played an important role in the acquisition of *Ocimum basilicum* to heat (Clarke et al., 2004). SA application increased antioxidant system in *Vitis vinifera* and induced activity of APX, GR, and monodehydro ascorbate (MDHA), increased redox ratios of AsA and GSH, and maintained Ca<sup>2+</sup>-homeostasis were reported in SA-supplemented and cold/heat treated *V. vinifera* (Wang and Li, 2006). SA (1.0 mM) decreased electrolyte leakage and oxidative stress, and improved maximum yield of PSII, Fv/Fm, and the quantum yield of the PSII electron transport in *Cucumis sativa* seedlings after both heat stress and recovery (Shi et al., 2006). Recently, Khan et al. (2013b) have shown that treatment of 0.5 mM SA can alleviate heat stress in *T. aestivum* by increasing Pro-production and restriction of the stress ethylene formation under heat stress. Notably, the details of the mechanisms of heat stress mitigation with the application of SA are not available and the area could be an open challenge at both physiological and molecular levels.

## Drought Stress

Drought has been considered as one of the most acute abiotic stresses presently affecting agriculture. Drought stress can significantly reduce photosynthesis and stomatal conductance, inhibit photosynthetic pigments synthesis and ultimately lead to reduction in growth of plants (reviewed by Hasanuzzaman et al., 2013). SA (500 μM)-supplementation to drought stressed *H. vulgare* resulted in increased net CO<sub>2</sub> assimilation rate due to increased stomatal conductance and eventually in increased plant dry mass (Habibi, 2012). Exogenously applied SA can modulate important enzymatic (including monodehydroascorbate reductase, MDHAR; dehydroascorbate reductase, DHAR; GR; GSH peroxidase, GPX) and non-enzymatic (including GSH) components of AsA-GSH pathway, and also glyoxalase system (Gly I and Gly II) and decrease oxidative stress in drought-exposed plants (Alam et al., 2013). Foliar application of SA (1.0 μM) strengthened antioxidant defense system in drought-tolerant *Z. mays* cultivar to a great extent (vs. drought-sensitive cultivar; Saruhan et al., 2012). Low membrane lipid peroxidation but increased plant height and dry mass, and less wilting of leaves were reported in drought-exposed and SA (0.5 mM)-supplemented *T. aestivum* Kang et al. (2012). Recently, SA-biosynthetic enzymes (such as CS and ICS) were not correlated with the SA level, but ortho-hydroxy-cinnamic (oHCA) was correlated with SA biosynthesis and played crucial role in drought tolerance in *O. sativa* (Pál et al., 2014).

The application of 5.0 μM SA induced the expression of genes in *Mitragyna speciosa* (Jumali et al., 2011). Among the analyzed 292 expressed sequence tags (ESTs) randomly, the most of the genes that encode chaperone, HSPs, antioxidants and secondary

metabolite biosynthesis, such as SAD, CAD, and Cytochrome P450 (CYP) responded to SA treatment. The authors correlated the gene-expression-responses to SA with signaling pathway in plants under drought stress condition. SA (0.5 mM)-mediated significantly improved growth in drought-exposed *T. aestivum* seedlings was argued as a result of SA-mediated enhancements in the transcription of *GST1*, *GST2*, *GR*, *MDHAR* genes (Kang et al., 2013). SA-accumulating mutants (*cpr5* and *acd6*) exhibited stomatal closure and improved drought tolerance in *A. thaliana* by SA-mediated induced expression of PR genes (*PR1*, *PR2*, and *PR5*; Liu et al., 2013). *SIZ1*-mediated endogenous SA-accumulation was reported to play an important role in stomatal closure and drought tolerance in *A. thaliana* (Miura et al., 2013). Potential involvement of SA in the 76 identified proteins was reported in drought-exposed *T. aestivum* (Kang et al., 2012). These identified proteins were advocated to perform major physiological functions such as photosynthesis, carbohydrate metabolism, protein metabolism, stress and defense, energy production, signal transduction, and toxin metabolism. Some of the recent studies have shown that SA played significant role at different concentration by regulating many metabolic mechanisms (Table 1).

## Potential Mechanisms Underlying SA-Mediated Plant Stress-Tolerance

In order to enlighten the potential mechanisms underlying SA-mediated improved abiotic stress tolerance in plants, this section briefly appraises recent reports available on the interaction of SA with major osmolytes, mineral nutrients, secondary metabolites, and other phytohormones, and SA-involvement in ROS-signaling and the modulation of antioxidants.

### Interaction of SA with Osmolytes

To counteract the adverse effects of abiotic stresses-induced excess ROS production, plants have developed mechanisms facilitating their adaptation to osmotic and ionic stresses. Nevertheless, to maintain osmotic balance plants have well developed protective mechanism termed as osmoregulation mediated by osmolytes such as GB, Pro, soluble sugars, amines etc. These compounds do not interfere with the other plant metabolic processes and contribute to the turgor-maintenance in stressed plants (Misra and Saxena, 2009).

Glycinebetaine is considered as an effective compatible solute for osmotic adjustment and protection against osmotic stress (Munns, 2005), salt stress (Khan et al., 2014), and also to stresses caused by heat (Wang et al., 2010) and metals (Bharwana et al., 2014). The accumulation of GB in stressed plants adjusts cell osmotic balance, stabilizes membrane integrity, prevents the dissociation of polypeptides from the PSII complex, protects Rubisco activity, and also detoxifies toxic ions (Ashraf and Foolad, 2007). Notably, SA and its analog aspirin can induce GB-accumulation in the range of 0.5–2.5 mM in plants at high levels of NaCl, drought, and cold stresses (Jagendorf and Takabe, 2001). The effectiveness of both salicylate and aspirin in the induction of GB-accumulation is a part of systematic

**TABLE 1 | Summary of representative studies on salicylic acid (SA)-mediated control of major abiotic stress-impacts in plants.**

Plant name	Applied SA concentration	Parameters studied	Response	Reference
<b>Salt stress</b>				
<i>Vigna radiata</i>	0.5 mM	Glycinebetaine (GB) production, net photosynthesis, plant dry mass	+	Khan et al. (2014)
<i>Torrey grandis</i>	0.5 mM	Chlorophyll content, net CO <sub>2</sub> assimilation rates, proline content	+	Li et al. (2014)
<i>Glycine max</i>	0.5 mM	Na <sup>+</sup> /K <sup>+</sup> ratio	–	Ardebili et al. (2014)
<i>G. max</i>	0.5 mM	Superoxide dismutase activity, ascorbate content	+	Ardebili et al. (2014)
<i>Hordeum vulgare</i>	0.05 mM	MDA content, Na <sup>+</sup> /K <sup>+</sup> ratio	–	Fayez and Bazaid (2014)
	10 <sup>-4</sup> mM	Content and activity of Rubisco, Rubisco activase	+	Lee et al. (2014)
<b>Cadmium stress</b>				
<i>Cucumis melo</i>	0.1 mM	Photosynthetic capacity, PSII photochemistry efficiency, water use efficiency	+	Zhang et al. (2015)
<i>Brassica juncea</i>	1.0 mM	Mineral nutrients content	+	Ahmad et al. (2011)
<i>G. max</i>	120 mM	Superoxide dismutase activity, chlorophyll, GSH content, heme-oxygenase-1 activity, relative protein amount	+	Noriega et al. (2012)
<i>Poa pratensis</i>	0.5 mM	Nutrient elements content (K, Ca, Mg, and Fe)	+	Guo et al. (2013)
<i>P. pratensis</i>	0.5 mM	Cd uptake	–	Guo et al. (2013)
<i>Ricinus Communis</i>	0.5 mM	Gas exchange, chlorophyll content	–	Liu et al. (2011)
<b>Nickel stress</b>				
<i>Catharanthus roseus</i>	10 <sup>-5</sup> M	Content of alkaloids vincristine and vinblastine	+	Idrees et al. (2013)
<b>Chilling stress</b>				
<i>Punica granatum</i>	1 and 2 mM	Total phenolics	+	Mirdehghan and Ghotbi (2014)
<i>Citrus limon</i>	2 mM	Total phenolics, activity of phenylalanine ammonia-lyase (PAL)	+	Siboza et al. (2014)
<i>Musa acuminata</i>	0.5 mM	Ultrastructure of chloroplast of mesophyll cells, mitochondria of mesophyll cells	+	Kang et al. (2007)
<b>Cold stress</b>				
<i>H. vulgare</i>	0.1 mM	Apoplasmic antioxidative enzymes, ice nucleation activity, pattern of apoplasmic proteins	+	Mutlu et al. (2013)
<b>Heat stress</b>				
<i>Triticum aestivum</i>	0.5 mM	Proline content, glutamyl kinase activity, gas exchange, water potential	+	Khan et al. (2013b)
<b>Drought stress</b>				
<i>Zea mays</i>	0.001 mM	Leaf rolling degree, water potential, dry weight	+	Saruhan et al. (2012)
<i>Simarouba glauca</i>	0.05 mM	Polyphenol, alkaloids, flavonoid content	+	Awate and Gaikwad (2014)
<b>UV-B stress</b>				
<i>G. max</i>	0.5 mM	Water use efficiency, Fv/Fm	+	Li et al. (2014)
<i>G. max</i>	0.5 mM	Flavonoid content	–	Li et al. (2014)
<i>Oryza sativa</i>	12.9 g ha <sup>-1</sup>	PS II activity, crop grain characteristics, total phenolics	+	Mohammed and Tarpley (2013)

+, – symbols indicate increase or decrease, respectively.

acquired resistance and could be an important part of the reason for its induction in plants under NaCl-, drought-, and cold stresses-plants. The induction of GB might have other effects as well, such as activation of protein kinase when aroused by hyperosmotic stress (Hoyos and Zhang, 2000). SA-mediated increase in GB-level can improve overall plant growth (Misra and Misra, 2012). The increase in GB content led to the increase in biomass of *Rauwolfia serpentina*. Recently, Khan et al. (2014) have shown that alleviation of salinity-inhibited photosynthesis and growth by SA involves GB in *V. radiata*. It was shown that SA (at 0.5 mM) induced GB-accumulation through increased methionine content and suppressed excess ethylene formation under salinity stress. Similar effect was observed by these authors with the application of SA-analog, 2, 6, dichloro-isonicotinic acid on GB accumulation and

alleviation of salinity-induced adverse effects on photosynthesis and growth.

The accumulation of another major osmolyte Pro is one of the adaptive mechanisms that plants operate for survival especially under salinity/osmotic stress conditions. Pro detoxifies excess ROS, adjusts cellular osmotic balance, protects biological membranes, and stabilizes enzymes/proteins (Iqbal et al., 2014). Researches have well documented that SA is involved in increasing Pro metabolism under abiotic stresses (Misra and Saxena, 2009; Khan et al., 2013b). SA (at 0.5 mM) significantly induced activity of Pro biosynthesis enzymes (such as pyrroline-5-carboxylate reductase and  $\gamma$ -glutamyl kinase) under salinity stress along with the increased Pro content. This increase in Pro metabolism was attributed to salinity stress tolerance in *Lens esculenta* (Misra and Saxena, 2009). Misra and Misra (2012)

have shown that up-regulation of Pro biosynthesis enzymes (such as pyrroline-5-carboxylate reductase and  $\gamma$ -glutamyl kinase) and down-regulation of Pro oxidase activity were responsible for increased Pro level. In turn, the increased Pro level was advocated to the maintenance of the cell turgor in *R. serpentina* under salinity stress. SA-treatment (0.5 mM) alleviated heat stress in *T. aestivum* by increasing Pro-production as a result of SA-mediated increase in  $\gamma$ -glutamyl kinase and decrease in Pro oxidase activity (Khan et al., 2013b). Increased Pro production was argued to improve nitrogen assimilation and to alleviate the heat stress-impact on photosynthesis. In contrast, exogenous Pro-application was evidenced earlier to induce SA production (mediated by NDR1-dependent signaling pathway) and was shown to modulate calcium (Ca)-mediated oxidative burst defense response in plants (Chen et al., 2011).

Accumulation of soluble sugars and sugar alcohol mannitol has also been reported to contribute in plant stress tolerance as osmoprotectants (Murakeözy et al., 2003; Cheng et al., 2009). SA was reported to inhibit valine and sucrose uptake in a concentration-dependent manner (10–200  $\mu$ M; Bourbouloux et al., 1998). Improved plant health can also be achieved with increased contents of polysaccharides and soluble sugars, respectively, with 100  $\mu$ mol L<sup>-1</sup> (Yuan et al., 2014), and 0.5 and 1.0 mM SA (Luo et al., 2014).

### Interaction of SA with Mineral Nutrients

Mineral nutrition is a basic requirement for proper growth and development and survival under different environmental stress conditions. Studies have shown that mineral nutrient status in plants plays a critical role in the alleviation of abiotic stress (Iqbal et al., 2011; Nazar et al., 2011, 2015). SA can significantly modulate the uptake and metabolism of important mineral elements and thereby improve growth and development in abiotic stressed plants (Alpaslan and Gunes, 2001; Gunes et al., 2007; Chen et al., 2011; Khokon et al., 2011; Wang et al., 2011; Tufail et al., 2013; Nazar et al., 2015). In addition, the protective role of SA in membrane integrity and regulation of ions including nutrients uptake has also been reported (Alpaslan and Gunes, 2001; Gunes et al., 2007).

Salicylic acid can be involved in the regulation of uptake of several plant-beneficial elements such as Mn, Ca, Cu, Fe, P, and Zn and thereby minimize oxidative stress under Pb stress (Wang et al., 2011). SA-mediated changes in photosynthesis were attributed to the nutrients content of N, P, K, and Ca in *B. juncea* cultivars differing in salt tolerance (Syed et al., 2011). In particular, 0.5 mM SA increased photosynthesis under salt stress by decreasing cellular Na<sup>+</sup> and Cl<sup>-</sup> ions, and increasing the content of nutrients. Earlier, strongly inhibited Na<sup>+</sup> and Cl<sup>-</sup> accumulation but stimulated N, Mg, Fe, Mn, and Cu concentrations were reported in SA-supplemented and salinity stressed *Z. mays* (Gunes et al., 2007). In another work, SA (0.5 mM)-mediated maintenance of higher K<sup>+</sup>/Na<sup>+</sup> and Ca<sup>2+</sup>/Na<sup>+</sup> ratios was considered as a major factor underlying SA-assisted improved growth, gas exchange, yield, and salinity tolerance in *Z. mays* (Tufail et al., 2013). Recently, Nazar et al. (2015) demonstrated that SA can improve salinity tolerance in

*B. juncea* by upregulating the assimilation of S. Exogenously sourced SA (seed-soaked or soil-incorporated) can improve overall plant growth under B-toxicity and salinity stress by stimulating the accumulation of mineral elements including K, Mg, Mn, N, and P (Gunes et al., 2005).

Calcium is one of the important nutrient elements known for various structural roles under both optimal and stressful conditions in plants (White and Broadley, 2003). Literature supports a close link between SA and Ca (and Ca-signaling) in stressed plants (Liu and Zhu, 1997; Kawano et al., 1998; Chen et al., 2001, 2011; Yang and Poovaiah, 2002; Du et al., 2009). Interaction between SA and Ca<sup>2+</sup> signaling might be involved in defense mechanisms induced by stress and maintenance of K<sup>+</sup>/Na<sup>-</sup> ion selectivity (Liu and Zhu, 1997; Chen et al., 2001). SA and Ca (alone or in combination) can markedly improve salinity tolerance in plants via increasing Pro level (Al-Whaibi et al., 2012). Involvement of Pro in the induction of Ca-mediated oxidative burst and SA signaling has also been reported (Chen et al., 2011). A range of abiotic/environmental stresses (such as high temperature, UV-B stress, or salt stress) can induce the expression of Genes encoding calmodulin-binding protein (Ca-containing protein). On the other hand, SA was reported to induce calmodulin-binding proteins in abiotic stressed *A. thaliana* (Yang and Poovaiah, 2002). Ca-dependent protein kinases (CDPKs) have been shown to be involved in abiotic stress responses and may also be induced by SA (Chung et al., 2004; Leclercq et al., 2005). Kawano et al. (1998) reported that SA-induced a rapid and transient generation of superoxide anion followed by a transient increase in cytosolic free Ca ion concentration in *N. tabacum*. Khokon et al. (2011) reported that SA failed to induce Ca-Cyt oscillations in guard cells; whereas, K<sup>+</sup> channel activity was suppressed by SA and lead to the stomatal closure.

### Involvement of SA in ROS-Signaling and the Modulation of Antioxidants

The generation and scavenging of varied ROS such as O<sub>2</sub><sup>-</sup>, H<sub>2</sub>O<sub>2</sub>, and •OH are usual in the normal aerobic metabolism in plants. Notably, important signal transducing roles and triggering and/or orchestrating plant responses to varied stress factors can be possible with the minimal levels of ROS. However, (abiotic) stresses cause imbalance between generation and scavenging of ROS, and eventually lead to a physiological condition known as oxidative stress. Thus, a number of consequences such as oxidative modification of vital biomolecules, cell death, and the arrest of plant growth and development can be possible under uncontrolled oxidative stress (Gill and Tuteja, 2010; Anjum et al., 2012).

Apoplastic-ROS have been found as regulators of cell death through their interplay with several other signaling pathways including SA-mediated signaling pathways (Overmyer et al., 2003). Both endogenous and exogenous SA was evidenced to play roles in antioxidant metabolism and have a tight control over cellular ROS (Kang et al., 2014; Khan et al., 2014). However, the coordination of SA-dependent and independent signaling components with ROS-signaling provided an appropriate defense response. SA can act as a signal for the development of the



systemic acquired resistance (Shirasu et al., 1997), and can also induce the activation of a protein kinase (Mikolajczyk et al., 2000). SA was evidenced as a major signaling molecule required for agonizing ozone-induced GSH-based defense gene expression in *A. thaliana* (Rao and Davis, 1999). *A. thaliana* plants were able to recognize the response of ROS–SA interaction via an antagonistic action of SA and SA-signaling on apoplastic ROS-signaling (Xu and Brosché, 2014). SA-signaling except from the well-documented role in the regulation of defense responses was also evidenced to be involved in the regulation of light-acclimation processes and thereby influencing photosynthetic function (Mullineaux and Karpinski, 2002; Gawroński et al., 2013).

Involvement of SA in the modulation of antioxidant metabolism has been widely reported to control plant-tolerance to major abiotic stresses including ozone, UV-B, heat, metal, and osmotic stress (Wang et al., 2010; Nazar et al., 2011; Khan et al., 2012a,b,c, 2013a,b, 2014). SA-pretreatment was evidenced to alleviate the adverse effects of salinity stress on photosynthesis and growth in *V. radiata* through enhancing the activities of antioxidant enzymes including SOD, CAT, GPX, APX, and GR (Khan et al., 2014). Activities of H<sub>2</sub>O<sub>2</sub>-metabolizing enzymes (such as CAT, POD, and APX) and superoxide-dismutating enzymes (SOD) were also modulated with SA in plants exposed to drought (Saruhan et al., 2012) and cold (Mutlu et al., 2013). SA-application (at 0.5 mM) increased activity of enzymes of AsA–GSH pathway resulted in the increased tolerance of *B. juncea* to salinity stress (Nazar et al., 2015). AsA and GSH, as redox active compounds have been extensively reported to maintain a homeostatic balance of the cellular redox state, and are involved in protective mechanisms against both abiotic and biotic stresses (Anjum et al., 2010, 2014b; Noctor et al., 2012; Khan et al., 2015a). In Cd-exposed *T. aestivum* varieties, SA-signaling was correlated with GSH-related mechanisms (Kovács et al., 2014). Exogenous SA (0.5 mM) significantly improved salinity (250 mM NaCl) tolerance in *T. aestivum* by markedly increasing the pools of AsA and GSH (Li et al., 2013).

Salicylic acid-mediated differential regulation of the transcript levels of the genes encoding AsA–GSH cycle enzymes GPX (*GPX1*), phospholipid hydroperoxide GPX (*GPX2*), and DHAR (*DHAR*), GR (*GR*), GST (*GST1* and *GST2*), MDHAR (*MDHAR*), and GSH synthetase (*GS*) was advocated as a major mechanisms underlying previous role of SA in salinity tolerance via AsA and GSH (Li et al., 2013). Recently, in *S. lycopersicum*, SA-priming exhibited a concentration-dependent modulation of GST-supergene family, where SA-mitigated the salinity stress-injury and caused characteristic changes in the expression pattern of *SIGSTs* only at 10<sup>-4</sup> M concentration (Csizsár et al., 2014). In particular, *SIGSTF4* displayed a significant up-regulation in the leaves, while the abundance of *SIGSTL3*, *SIGSTT2*, and *SIGSTZ2* transcripts were enhanced in the roots of plants primed with high SA-concentration. SA-mediated induction in SOD and GSH-based H<sub>2</sub>O<sub>2</sub>-metabolizing enzymes namely GPX and GST was argued to improve in ozone tolerance in plants (Abarca et al., 2001; Milla et al., 2003). SA (500 μM) improved Cd-tolerance and photosynthetic capacity in hemp (*Cannabis sativa*) by enhancing both SOD and POD (Shi et al., 2009). In another instance the

applied SA-induced SOD activity accompanied an increase in shoot Ca<sup>2+</sup> (a second messenger) and caused a transient increase in H<sub>2</sub>O<sub>2</sub> which in turn was argued to induce antioxidant enzymes and eventually to decrease in cellular ROS (Arfan, 2009).

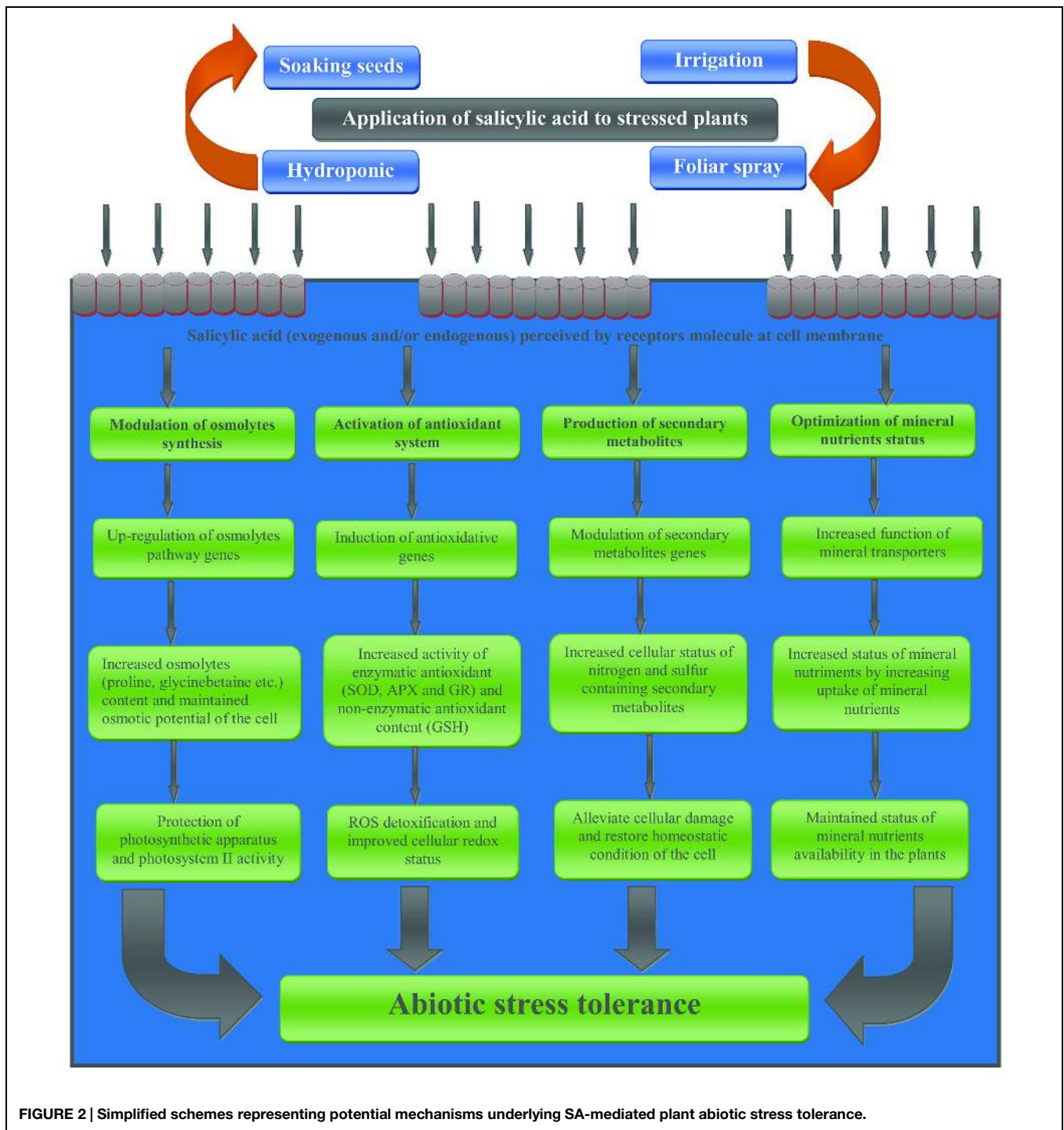
## Interaction of SA with Major Secondary Metabolites

Secondary metabolites, substances biosynthetically derived from primary metabolites, produced by plants as defense chemicals and are not involved in plant metabolic activity. Examples of secondary metabolites include including terpenes, phenolics, and compounds with N (alkaloids, cyanogenic glucosides, non-protein amino acids) and S (GSH, glucosinolates, phytoalexins, thionins, defensins, and allinin; Irchhaiya et al., 2015). Secondary metabolites have also been established as beneficial for plants tolerance under biotic and abiotic stress (Ramakrishna and Ravishankar, 2011).

Extensive reports are available on the direct and indirect involvement of SA in the induction of the secondary metabolite synthesis in plants (Kiddle et al., 1994; Ali et al., 2007; Idrees et al., 2013). SA-mediated induction of glucosinolates production was reported in *Brassica napus* (Kiddle et al., 1994). Exogenous application of acetyl-SA to tumor lines of *Catharanthus roseus* elevated therein the production of important secondary metabolites such as alkaloids, phenolics, furanocoumarins, and anthocyanins (Godoy-Hernández and Loyola-Vargas, 1997). Recently, in the same plant SA-mediated restoration of Ni-induced inhibition was attributed to SA-assisted improved the content of alkaloids such as vincristine and vinblastine (Idrees et al., 2013). Role of exogenously applied SA was reported in UV-B exposed *T. aestivum*, where SA increased the accumulation of anthocyanin and tocopherol, and also modulated the expression of *PR* proteins (Horváth et al., 2007). Elicitation in the PAL activity and subsequent increased vanillin production were reported in SA-supplemented in *C. chinense* (Rodas-Junco et al., 2013). Foliar application of SA at the rate of 50 mg L<sup>-1</sup> induced the level of secondary metabolites such coumarins, sterols, xanthoproteins, cardiac glycosides and saponins, and ameliorated water stress in *Simarouba glauca* (Awate and Gaikwad, 2014). Earlier, SA-mediated improved thermo-tolerance in *V. vinifera* was advocated as a result of SA-induced accumulation of *PAL* mRNA, the synthesis of new *PAL* protein, and significant accumulation of phenolics (Wen et al., 2008). A schematic presentation of SA-mediated metabolisms responsible for abiotic stress tolerance is shown in **Figure 2**.

## Interaction of SA with Other Hormones

Many factors are involved in a complex process of plant growth and development. SA can regulate various aspects of plant responses under both stressful and optimal environments through signaling cross-talks with other phytohormones (Horváth et al., 2007; Asensi-Fabado and Munné-Bosch, 2011; Khan et al., 2012a,b, 2013b, 2014). The interaction between SA and other phytohormones such as auxin (Iglesias et al., 2011; Khan et al., 2012a), cytokinin (Peleg et al., 2011; Khan et al., 2012a), gibberellins (Alonso-Ramírez et al., 2009; Khan et al., 2012a), abscisic acid (Szepesi et al., 2009; Khan et al., 2012a),



ethylene (Khan et al., 2012a, 2013b, 2014), nitric oxide (NO; Khan et al., 2012a; Kong et al., 2014), and brassinosteroids (Divi et al., 2010; Khan et al., 2012a) has been established under both normal and stressed conditions.

The outcomes of the SA-interaction with other phytohormones can be either from synergistic or antagonistic relation therein under optimal and stressful conditions. SA-mediated repression of auxin signaling was demonstrated by

Iglesias et al. (2011). The authors showed that the expression of the *PR1*, a SA-induced gene in *tir1 afb2* (auxin perception small family of F-box proteins) mutant plants plays a significant role in auxin-signaling in stressed plants. Furthermore, in these mutants, *PR-1* was induced to a great extent as compared with SA-treated wild-type plants, where the negative effect of SA on auxin signaling indicates the occurrence of an antagonism between auxin and SA-signaling pathways (Iglesias et al., 2011).

SA triggered the accumulation of ABA under both normal and salinity stressed plants which in turn helped in the osmotic adaptation and improved photosynthetic pigments and growth attributes in *S. lycopersicum* (Szepesi et al., 2009). ABA may alter the SA-related abiotic (cold) stress response in plants. To this end, in chilling-exposed *Z. mays*, ABA treatment induced changes in the endogenous SA and the content of oHCA, and suggested that SA related stress responses may overlap with ABA-induced cold-acclimation (Szalai et al., 2011).

The increased ethylene production under environmental stress referred as stress ethylene is believed to induce oxidative stress (Khan and Khan, 2014; Khan et al., 2015b). Reports suggest an antagonism between SA and ethylene. Under stressful conditions, the application of SA could inhibit ethylene biosynthesis by restricting the conversion of 1-aminocyclopropane carboxylic acid (ACC) to ethylene (Leslie and Romani, 1986). Exogenous SA was reported to alleviate heat stress by increasing Pro-metabolism and restricting ethylene formation in heat-stressed plants to an optimal range by inhibiting activity of ACC synthase (ACS; Khan et al., 2013b). Increased methionine and GB-accumulation in *V. radiata* plants was concomitant with improved photosynthesis and growth, and the suppression of ethylene formation as a result of SA (0.5 mM)-mediated inhibition in ACS activity under salinity stress (Khan et al., 2014). Salinity stress was evidenced to induce cell death mainly by ethylene-induced ROS-production; however, ROS generated by SA was not controlled by ethylene in *Lycopersicon esculentum* cell suspension (Poór et al., 2013). In contrast, Ghanta et al. (2014) suggested a synergistic cross-talk between ethylene and SA for combating environmental stress-impact in plants. In a similar study, an increased ethylene formation was observed with SA-addition in the MS medium; whereas, it was decreased in regenerants exposed to AgNO<sub>3</sub> in *Prunus persica* rootstock (Molassiotis et al., 2005). In contrast, the accumulation of polyamines under salinity stress was prevented by SA-application along with the increased ACC content (Palma et al., 2013). Further, SA-induced inhibition in the lipid peroxidation and polyamines in nodules of *M. sativa* can be an indication of SA-mediated activation of ethylene-dependent hypersensitive response (Palma et al., 2013).

Nitric oxide, a major ubiquitous signal in plant systems, plays significant roles in a wide range of responses to environmental and endogenous cues (Freschi, 2013). SA is among the major phytohormones to which NO interacts, plays role as a second messenger, and controls stomatal movement in higher plants (Hao et al., 2010). NO has been advocated to act as downstream of SA-signaling in the reduction of induced oxidative damage in osmotic stressed *T. aestivum* seedlings (Naser Alavi et al., 2014). SA can also stimulate the synthesis of NO via enhancing the activity of NO-synthesizing enzymes (Zottini et al., 2007). Under Ni stress, addition of SA or SNP partially reduced the toxic effects of Ni. However, Ni-stressed plants supplemented with SA+NO exhibited an improved growth and photosynthetic pigments in canola (*B. napus*; vs. Ni-treated plants; Kazemi et al., 2010). Recently, the combined application of SA and NO benefitted *Arachis* seedlings (in terms of high Fe-uptake and less leaf interveinal chlorosis) maximally compared to exogenous

individual SA and SNP under Fe-deficiency (Kong et al., 2014). The stress tolerance conferring ability of brassinosteroid in plants was advocated to occur in part in its interactions with other stress hormones such as SA (Divi et al., 2010). It was observed that *NPRI* gene, a master regulator of SA-mediated defense genes is a critical component of 24-epibrassinolide-mediated increase in thermo- and salinity tolerance in *A. thaliana* (Divi et al., 2010). There occur antagonistic interactions between SA and JA at the levels of MAPKs signaling and biosynthesis (Khan et al., 2012a,b). The antagonistic relationship between SA and JA can also modulate the expression of PR protein genes, where induction and inhibition of PR genes can be possible with SA and JA, respectively, (Thaler et al., 1999; Wang et al., 2001).

## Conclusion and Future Prospects

Abiotic stress has been recognized as a major threat to the agricultural system. To cope up the adverse effects of abiotic stress, plants induce several physiological processes and molecular mechanisms. Investigations have shown SA as a strong and potential tool in reducing or alleviating the adverse effects of abiotic stress in plants. Application of SA has been shown to be beneficial for plants either in optimal or stress environments. SA can regulate various plant metabolic processes and modulate the production of varied osmolytes and secondary metabolites, and also maintain plant-nutrient status hence, to protect plants under abiotic stress conditions. Literature appraised herein confirmed the focus of the studies related with SA in abiotic stressed plants mainly on unveiling various physiological/biochemical processes.

There are still a few of interesting questions awaiting further investigation. For example, *NPRI*, 3, and 4 are discovered as the SA receptor in plants. However, it is required to perform exhaustive molecular studies cross-talking *NPRI*, *NPR3*, and *NPR4*, and their cumulative potential role in SA-perception in orchestrating defense gene expression in abiotic stressed plants. Unveiling intricacies of the complete SA-signaling in abiotic stresses, and the relationship between distinct facets of SA in plant immunity and abiotic stresses responses would be fascinating and rewarding. Meanwhile, more genomics and proteomics studies are expected to broadly reveal SA-responsive genes and proteins upon stresses. Since SA influences the plant functions in a dose dependent manner and a high SA-concentration doesn't profit stress tolerance, abiotic stress-regulated SA-catabolism maybe a field worth further investigation. Molecular dissection is also required to unravel insights into the SA-mediated control of other the production and/or signaling of specific plant hormones/metabolites, and their feed-back influence in the machinery responsible for controlling SA-endogenous levels. Overall, finer details of SA-mediated defense networks/plant-immunity as well as further insights into the cross-talk of SA with other defense signaling pathways in abiotic stressed plants can be uncovered through adopting an integrated approach incorporating genetics, molecular biology, biochemistry, genomics, bioinformatics techniques, and computational biology.



## Acknowledgments

Financial support to NAK laboratory by the University Grants Commission (UGC) and Department of Biotechnology-BUILDER (DBT-BUILDER), New Delhi (India) is gratefully

acknowledged. NAA is grateful for funds to the Portuguese Foundation for Science and Technology (FCT) (SFRH/BPD/84671/2012), and the Aveiro University Research Institute/CESAM, Portugal (UID/AMB/50017/2013).

## References

- Abarca, D., Martín, M., and Sabater, B. (2001). Differential leaf stress responses in young and senescent plants. *Physiol. Plant.* 113, 409–415. doi: 10.1034/j.1399-3054.2001.1130315.x
- Ahmad, P., Nabi, G., and Ashraf, M. (2011). Cadmium-induced oxidative damage in mustard [*Brassica juncea* (L.) Czern. & Coss.] plants can be alleviated by salicylic acid. *South Afr. J. Bot.* 77, 36–44. doi: 10.1016/j.sajb.2010.05.003
- Alam, M. M., Hasanuzzaman, M., Nahar, K., and Fujita, M. (2013). Exogenous salicylic acid ameliorates short-term drought stress in mustard (*Brassica juncea* L.) seedlings by up-regulating the antioxidant defense and glyoxalase system. *Aust. J. Crop Sci.* 7, 1053–1063.
- Alavi, S. M. N., Arvin, M. J., and Kalantari, K. M. (2014). Salicylic acid and nitric oxide alleviate osmotic stress in wheat (*Triticum aestivum* L.) seedlings. *J. Plant Interac.* 9, 683–688. doi: 10.1080/17429145.2014.900120
- Ali, M. B., Hahn, E. J., and Paek, K. Y. (2007). Methyl jasmonate and salicylic acid induced oxidative stress and accumulation of phenolics in *Panax ginseng* bioreactor root suspension cultures. *Molecules* 12, 607–621. doi: 10.3390/12030607
- Alonso-Ramírez, A., Rodríguez, D., Reyes, D., Jiménez, J. A., Nicolás, G., López-Climent, M., et al. (2009). Cross-talk between gibberellins and salicylic acid in early stress responses in *Arabidopsis thaliana* seeds. *Plant Signal. Behav.* 4, 750–751. doi: 10.4161/psb.4.8.9175
- Alpaslan, M., and Gunes, A. (2001). Interactive effect of boron and salinity stress on the growth, membrane permeability and mineral composition of tomato and cucumber plants. *Plant Soil.* 236, 123–128. doi: 10.1023/A:1011931831273
- Al-Wahaibi, M. H., Siddiqui, M. H., and Basalah, M. O. (2012). Salicylic acid and calcium-induced protection of wheat against salinity. *Protoplasma* 249, 769–778. doi: 10.1007/s00709-011-0322-1
- An, C., and Mou, Z. (2011). Salicylic acid and its function in plant immunity. *J. Integ. Plant Biol.* 53, 412–428. doi: 10.1111/j.1744-7909.2011.01043.x
- Anjum, N. A., Gill, S. S., and Gill, R. (2014a). *Plant Adaptation to Environmental Change: Significance of Amino Acids and their Derivatives*, 1st Edn. Wallingford: CAB. doi: 10.1079/9781780642734.0000
- Anjum, N. A., Aref, I. M., Duarte, A. C., Pereira, E., Ahmad, I., and Iqbal, M. (2014b). Glutathione and proline can coordinately make plants withstand the joint attack of metal(loid) and salinity stresses. *Front. Plant Sci.* 5:662. doi: 10.3389/fpls.2014.00662
- Anjum, N. A., Umar, S., and Ahmad, A. (2012). *Oxidative Stress in Plants: Causes, Consequences and Tolerance*. New Delhi: IK International Publishing House.
- Anjum, N. A., Umar, S., and Chan, M. T. (2010). *Ascorbate-Glutathione Pathway And Stress Tolerance In Plants*. Springer: Dordrecht. doi: 10.1007/978-90-481-9404-9
- Anwar, S., Iqbal, M., Raza, S. H., and Iqbal, N. (2013). Efficacy of seed preconditioning with salicylic and ascorbic acid in increasing vigor of rice (*Oryza sativa* L.) seedling. *Pak. J. Bot.* 45, 157–162.
- Ardebili, N. O., Saadatmand, S., Niknam, V., and Khavari-Nejad, R. A. (2014). The alleviating effects of selenium and salicylic acid in salinity exposed soybean. *Acta Physiol. Plant* 36, 3199–3205. doi: 10.1007/s11738-014-1686-6
- Arfan, M. (2009). Exogenous application of salicylic acid through rooting medium modulates ion accumulation and antioxidant activity in spring wheat under salt stress. *Intl. J. Agri. Biol.* 11, 437–442.
- Asensi-Fabado, M. A., and Munné-Bosch, S. (2011). The aba3-1 mutant of *Arabidopsis thaliana* withstands moderate doses of salt stress by modulating leaf growth and salicylic acid levels. *J. Plant Growth Regul.* 30, 456–466. doi: 10.1007/s00344-011-9208-x
- Asgher, M., Khan, M. I. R., Anjum, N. A., and Khan, N. A. (2015). Minimizing toxicity of cadmium in plants—role of plant growth regulators. *Protoplasma* 252, 399–413. doi: 10.1007/s00709-014-0710-4
- Ashmore, M. R. (2005). Assessing the future global impacts of ozone on vegetation. *Plant Cell Environ.* 28, 949–964. doi: 10.1111/j.1365-3040.2005.01341.x
- Ashraf, M., and Foolad, M. (2007). Roles of glycine betaine and proline in improving plant abiotic stress resistance. *Environ. Exp. Bot.* 59, 206–216. doi: 10.1016/j.envexpbot.2005.12.006
- Awate, P. D., and Gaikwad, D. K. (2014). Influence of growth regulators on secondary metabolites of medicinally important oil yielding plant *Simarouba glauca* DC. under water stress conditions. *J. Stress Physiol. Biochem.* 10, 222–229.
- Azooz, M. M. (2009). Salt stress mitigation by seed priming with salicylic acid in two faba bean genotypes differing in salt tolerance. *Intl. J. Agric. Biol.* 11, 343–350.
- Bandurska, H., and Cieslak, M. (2013). The interactive effect of water deficit and UV-B radiation on salicylic acid accumulation in barley roots and leaves. *Environ. Exp. Bot.* 94, 9–18. doi: 10.1016/j.envexpbot.2012.03.001
- Belkadi, A., De Haro, A., Obregon, S., Chaïbi, W., and Djebali, W. (2014). Positive effects of salicylic acid pretreatment on the composition of flax plastidial membrane lipids under cadmium stress. *Environ. Sci. Pollu. Res.* 22, 1457–1467. doi: 10.1007/s11356-014-3475-6
- Bharwana, S. A., Ali, S., Farooq, M. A., Ali, B., Iqbal, N., Abbas, F., et al. (2014). Hydrogen sulfide ameliorates lead-induced morphological, photosynthetic, oxidative damages and biochemical changes in cotton. *Environ. Sci. Pollu. Res.* 21, 717–731. doi: 10.1007/s11356-013-1920-6
- Bourbouloux, A., Raymond, P., and Delrot, S. (1998). Effects of salicylic acid on sugar and amino acid uptake. *J. Exp. Bot.* 49, 239–247. doi: 10.1093/jxb/49.3.19.239
- Caldwell, M. M., Bornman, J. F., Ballaré, C. L., Flint, S. D., and Kulandaivelu, G. (2007). Terrestrial ecosystems, increased solar ultraviolet radiation, and interactions with other climate change factors. *Photochem. Photobiol. Sci.* 6, 252–266. doi: 10.1039/b700019g
- Cao, Y., Zhang, Z. W., Xue, L. W., Du, J. B., Shang, J., Xu, F., et al. (2009). Lack of salicylic acid in *Arabidopsis* protects plants against moderate salt stress. *Z. Naturforsch. C* 64, 231–238. doi: 10.1515/znc-2009-3-414
- Chai, J., Liu, J., Zhou, J., and Xing, D. (2014). Mitogen-activated protein kinase 6 regulates NPR1 gene expression and activation during leaf senescence induced by salicylic acid. *J. Exp. Bot.* 65, 6513–6528. doi: 10.1093/jxb/eru369
- Chao, Y. Y., Chen, C. Y., Huang, W. D., and Kao, C. H. (2010). Salicylic acid-mediated hydrogen peroxide accumulation and protection against Cd toxicity in rice leaves. *Plant Soil.* 329, 327–337. doi: 10.1007/s11104-009-0161-4
- Chen, H. J., Hou, W. C., Kuć, J., and Lin, Y. H. (2001). Ca<sup>2+</sup>-dependent and Ca<sup>2+</sup>-independent excretion modes of salicylic acid in tobacco cell suspension culture. *J. Exp. Bot.* 52, 1219–1226. doi: 10.1093/jxb/52.359.1219
- Chen, J. Y., He, L. H., Jiang, Y. M., Wang, Y., Joyce, D. C., Ji, Z. L., et al. (2008). Role of phenylalanine ammonia-lyase in heat pretreatment-induced chilling tolerance in banana fruit. *Physiol. Plant.* 132, 318–328. doi: 10.1111/j.1399-3054.2007.01013.x
- Chen, J., Zhang, Y., Wang, C., Lü, W., Jin, J. B., and Hua, X. (2011). Proline induces calcium-mediated oxidative burst and salicylic acid signaling. *Amino Acids* 40, 1473–1484. doi: 10.1007/s00726-010-0757-2
- Chen, J., Zhu, C., Li, L. P., and Sun, Z. Y. (2007). Effects of exogenous salicylic acid on growth and H<sub>2</sub>O<sub>2</sub>-metabolizing enzymes in rice seedlings under lead stress. *J. Environ. Sci.* 19, 44–49. doi: 10.1016/S1001-0742(07)60007-2
- Cheng, Y. J., Yang, S. H., and Hsu, C. S. (2009). Synthesis of conjugated polymers for organic solar cell applications. *Chem. Rev.* 109, 5868–5923. doi: 10.1021/cr900182s
- Chung, E., Park, J. M., Oh, S. K., Joung, Y. H., Lee, S., and Choi, D. (2004). Molecular and biochemical characterization of the *Capsicum annuum* calcium-dependent protein kinase 3 (CaCDPK3) gene induced by abiotic and biotic stresses. *Planta* 220, 286–295. doi: 10.1007/s00425-004-1372-9



- Clarke, S. M., Mur, L. A., Wood, J. E., and Scott, I. M. (2004). Salicylic acid dependent signaling promotes basal thermotolerance but is not essential for acquired thermotolerance in *Arabidopsis thaliana*. *Plant J.* 38, 432–447. doi: 10.1111/j.1365-3113X.2004.02054.x
- Cleland, F. C., and Ajami, A. (1974). Identification of the flower-inducing factor isolated from aphid honeydew as being salicylic acid. *Plant Physiol.* 54, 904–906. doi: 10.1104/pp.54.6.904
- Csiszár, J., Horváth, E., Váry, Z., Gallé, Á., Bela, K., Brunner, S., et al. (2014). Glutathione transferase supergene family in tomato: salt stress-regulated expression of representative genes from distinct GST classes in plants primed with salicylic acid. *Plant Physiol. Biochem.* 78, 15–26. doi: 10.1016/j.plaphy.2014.02.010
- Dehghan, S., Sadeghi, M., Poppel, A., and Fischer, R. (2014). Differential inductions of phenylalanine ammonia-lyase and chalcone synthase during wounding, salicylic acid treatment, and salinity stress in safflower, *Carthamus tinctorius*. *Bioscienc. Rep.* 34, 3. doi: 10.1042/BSR20140026
- Ding, C. K., Wang, C., Gross, K. C., and Smith, D. L. (2002). Jasmonate and salicylate induce the expression of pathogenesis-related-protein genes and increase resistance to chilling injury in tomato fruit. *Planta* 214, 895–901. doi: 10.1007/s00425-001-0698-9
- Divi, U. K., Rahman, T., and Krishna, P. (2010). Brassinosteroid-mediated stress tolerance in *Arabidopsis* shows interactions with abscisic acid, ethylene and salicylic acid pathways. *BMC Plant Biol.* 10:151. doi: 10.1186/1471-2229-10-151
- Du, L., Ali, G. S., Simons, K. A., Hou, J., Yang, T., Reddy, A. S., et al. (2009). Ca<sup>2+</sup>/calmodulin regulates salicylic-acid-mediated plant immunity. *Nature* 457, 1154–1158. doi: 10.1038/nature07612
- ENS. (2006). *Environment News Service: Report List World's 10 Worst Pollution Spots*. Available at: [http://www.google.pt/url?sa=t&rct=j&q=&esrc=s&source=web&cd=2&cad=rja&uact=8&ved=0CCGQFjAB&url=http%3A%2F%2Fwww.chem.unep.ch%2Fpb\\_and\\_cd%2FDocuments%2FNews%2F10%2520most%2520polluted%2520cities.pdf&ei=1zZgVb2iOKKR7AaMoIAo&usq=AFQjC NHOAyar\\_69TBPA143KXTt-cAvkP3g&sig2=\\_ITZpa2YQnsF0sl0Xm7ew&bv m=bv.93990622,d.ZGU](http://www.google.pt/url?sa=t&rct=j&q=&esrc=s&source=web&cd=2&cad=rja&uact=8&ved=0CCGQFjAB&url=http%3A%2F%2Fwww.chem.unep.ch%2Fpb_and_cd%2FDocuments%2FNews%2F10%2520most%2520polluted%2520cities.pdf&ei=1zZgVb2iOKKR7AaMoIAo&usq=AFQjC NHOAyar_69TBPA143KXTt-cAvkP3g&sig2=_ITZpa2YQnsF0sl0Xm7ew&bv m=bv.93990622,d.ZGU)
- Fayez, K. A., and Bazaid, S. A. (2014). Improving drought and salinity tolerance in barley by application of salicylic acid and potassium nitrate. *J. Saudi Soc. Agri. Sci.* 13, 45–55. doi: 10.1016/j.jssas.2013.01.001
- Fodor, J., Gullner, G., Adam, A. L., Barna, B., Komives, T., and Kiraly, Z. (1997). Local and systemic responses of antioxidants to tobacco mosaic virus infection and to salicylic acid in tobacco. *Plant Physiol.* 114, 1443–1451.
- Freeman, J. L., Garcia, D., Kim, D., Hopf, A., and Salt, D. E. (2005). Constitutively elevated salicylic acid signals glutathione-mediated nickel tolerance in *Thlaspi* nickel hyperaccumulators. *Plant Physiol.* 137, 1082–1091. doi: 10.1104/pp.104.055293
- Freschi, L. (2013). Nitric oxide and phytohormone interactions: current status and perspectives. *Front. Plant Sci.* 4:398. doi: 10.3389/fpls.2013.00398
- Garcion, C., Lohmann, A., Lamodièrre, E., Catinot, J., Buchala, A., Doermann, P., et al. (2008). Characterization and biological function of the ISOCHORISMATE SYNTHASE2 gene of *Arabidopsis*. *Plant Physiol.* 147, 1279–1287. doi: 10.1104/pp.108.119420
- Gawroński, P., Górecka, M., Bederska, M., Rusaczonok, A., Ślesak, I., Kruk, J., et al. (2013). Isochorismate synthase 1 is required for thylakoid organization, optimal plastoquinone redox status, and state transitions in *Arabidopsis thaliana*. *J. Exp. Bot.* 64, 3669–3679. doi: 10.1093/jxb/ert03
- Ghanta, S., Datta, R., Bhattacharyya, D., Sinha, R., Kumar, D., Hazra, S., et al. (2014). Multistep involvement of glutathione with salicylic acid and ethylene to combat environmental stress. *J. Plant Physiol.* 171, 940–950. doi: 10.1016/j.jplph.2014.03.002
- Gill, S. S., Anjum, N. A., Gill, R., and Tuteja, N. (2015). DNA damage and repair in plants under ultraviolet and ionizing radiations. *Sci. World J.* 2015, 250158. doi: 10.1155/2015/250158
- Gill, S. S., and Tuteja, N. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiol. Biochem.* 48, 909–930. doi: 10.1016/j.plaphy.2010.08.016
- Godoy-Hernández, G., and Loyola-Vargas, V. M. (1997). Effect of acetylsalicylic acid on secondary metabolism of *Catharanthus roseus* tumor suspension cultures. *Plant Cell Rep.* 16, 287–290. doi: 10.1007/BF01088282
- Gunes, A., Inal, A., Alpaslan, M., Cicek, N., Guneria, E., Eraslan, F., et al. (2005). Effects of exogenously applied salicylic acid on the induction of multiple stress tolerance and mineral nutrition in maize (*Zea mays* L.). *Arch. Agron. Soil Sci.* 51, 687–695. doi: 10.1080/03650340500336075
- Gunes, A., Inal, A., Alpaslan, M., Eraslan, F., Bagci, E. G., and Cicek, N. (2007). Salicylic acid induced changes on some physiological parameters symptomatic for oxidative stress and mineral nutrition in maize (*Zea mays* L.) grown under salinity. *J. Plant Physiol.* 164, 728–736. doi: 10.1016/j.jplph.2005.12.009
- Guo, Q., Meng, L., Mao, P. C., Jia, Y. Q., and Shi, Y. J. (2013). Role of exogenous salicylic acid in alleviating cadmium-induced toxicity in Kentucky bluegrass. *Biochem. Syst. Ecol.* 50, 269–276. doi: 10.1016/j.bse.2013.05.002
- Habibi, G. (2012). Exogenous salicylic acid alleviates oxidative damage of barley plants under drought stress. *Acta Biol. Szeged.* 56, 57–63.
- Hao, F., Zhao, S., Dong, H., Zhang, H., Sun, L., and Miao, C. (2010). Nial and Nia2 are involved in exogenous salicylic acid-induced nitric oxide generation and stomatal closure in *Arabidopsis*. *J. Integ. Plant Biol.* 52, 298–307. doi: 10.1111/j.1744-7909.2010.00920.x
- Hasanuzzaman, M., Nahar, K., Gill, S. S., and Fujita, M. (2013). “Drought stress responses in plants, oxidative stress, and antioxidant defense,” in *Climate Change and Plant Abiotic Stress Tolerance*, eds N. Tuteja and S. S. Gill (Weinheim: Wiley-VCH Verlag GmbH & Co. KGaA), 209–250. doi: 10.1002/9783527675265.ch09
- He, Y. L., Liu, Y. L., Chen, Q., and Bian, A. H. (2002). Thermotolerance related to antioxidant induced by salicylic acid and heat hardening in tall fescue seedlings. *J. Plant Physiol. Mol.* 28, 89–95.
- Herrera-Vásquez, A., Salinas, P., and Holoigue, L. (2015). Salicylic acid and reactive oxygen species interplay in the transcriptional control of defense genes expression. *Front. Plant Sci.* 6:171. doi: 10.3389/fpls.2015.00171
- Herrmann, K. M., and Weaver, L. M. (1999). The shikimate pathway. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 50, 473–503. doi: 10.1146/annurev.arplant.50.1.473
- Horváth, E., Janda, T., Szalai, G., and Páldi, E. (2002). *In vitro* salicylic acid inhibition of catalase activity in maize: differences between the isozymes and a possible role in the induction of chilling tolerance. *Plant Sci.* 163, 112–1135. doi: 10.1016/S0168-9452(02)00324-2
- Horváth, E., Pál, M., Szalai, G., Páldi, E., and Janda, T. (2007). Exogenous 4-hydroxybenzoic acid and salicylic acid modulate the effect of short-term drought and freezing stress on wheat plants. *Biol. Plant.* 51, 480–487. doi: 10.1007/s10535-007-0101-1
- Hoyos, M. E., and Zhang, S. (2000). Calcium-independent activation of salicylic acid-induced protein kinase and a 40-kilodalton protein kinase by hyperosmotic stress. *Plant Physiol.* 122, 1355–1364. doi: 10.1104/pp.122.4.1355
- Hunter, L. J., Westwood, J. H., Heath, G., Macaulay, K., Smith, A. G., Macfarlane, S. A., et al. (2013). Regulation of RNA-dependent RNA polymerase 1 and isochorismate synthase gene expression in *Arabidopsis*. *PLoS ONE* 8:e66530. doi: 10.1371/journal.pone.0066530
- Idrees, M., Naeem, M., Aftab, T., and Khan, M. (2013). Salicylic acid restrains nickel toxicity, improves antioxidant defence system and enhances the production of anticancer alkaloids in *Catharanthus roseus* (L.). *J. Haz. Mat.* 252, 367–374. doi: 10.1016/j.jhazmat.2013.03.005
- Iglesias, M. J., Terrile, M. C., and Casalongué, C. A. (2011). Auxin and salicylic acid signalling counteract during the adaptive response to stress. *Plant Signal. Behav.* 6, 452–454. doi: 10.4161/psb.6.3.14676
- Iqbal, N., Nazar, R., Khan, M. I. R., Masood, A., and Khan, N. A. (2011). Role of gibberellins in regulation of source-sink relations under optimal and limiting environmental conditions. *Curr. Sci.* 100, 998–1007.
- Iqbal, N., Umar, S., Khan, N. A., and Khan, M. I. R. (2014). A new perspective of phytohormones in salinity tolerance: regulation of proline metabolism. *Environ. Exp. Bot.* 100, 34–42. doi: 10.1016/j.envexpbot.2013.12.006
- Irchhaiya, R., Kumar, A., Yadav, A., Gupta, N., Kumar, S., Gupta, N., et al. (2015). Metabolites in plants and its classification. *World J. Pharm. Pharmaceut. Sci.* 4, 287–305.
- Jagendorf, A. T., and Takabe, T. (2001). Inducers of glycinebetaine synthesis in barley. *Plant Physiol.* 127, 1827–1835. doi: 10.1104/pp.010392
- Janda, T., Szalai, G., Tari, I., and Páldi, E. (1999). Hydroponic treatment with salicylic acid decreases the effects of chilling injury in maize (*Zea mays* L.) plants. *Planta* 208, 175–180. doi: 10.1007/s004250050547
- Jayakannan, M., Bose, J., Babourina, O., Rengel, Z., and Shabala, S. (2013). Salicylic acid improves salinity tolerance in *Arabidopsis* by restoring membrane potential

- and preventing salt-induced  $K^+$  loss via a GORK channel. *J. Exp. Bot.* 64, 2255–2268. doi: 10.1093/jxb/ert085
- Jordan, B. R. (1996). The effects of ultraviolet-B radiation on plants: a molecular perspective. *Adv. Bot. Res.* 22, 97–162. doi: 10.1016/s0065-2296(08)60057-9
- Jordan, B. R., James, P. E., and Soheila, A. H. (1998). Factors affecting UV-B-induced changes in *Arabidopsis thaliana* L. gene expression: the role of development, protective pigments and the chloroplast signal. *Plant Cell Physiol.* 39, 769–778.
- Jumali, S. S., Said, I. M., Ismail, I., and Zainal, Z. (2011). Genes induced by high concentration of salicylic acid in '*Mitragyna speciosa*'. *Aust. J. Crop Sci.* 5, 296–303.
- Kang, G., Li, G., and Guo, T. (2014). Molecular mechanism of salicylic acid-induced abiotic stress tolerance in higher plants. *Acta Physiol. Plant.* 36, 2287–2297. doi: 10.1007/s11738-014-1603-z
- Kang, G. Z., Li, G. Z., Liu, G. Q., Xu, W., Peng, X. Q., Wang, C. Y., et al. (2013). Exogenous salicylic acid enhances wheat drought tolerance by influence on the expression of genes related to ascorbate-glutathione cycle. *Biol. Plant.* 57, 718–724. doi: 10.1007/s10535-013-0335-z
- Kang, G., Li, G., Xu, W., Peng, X., Han, Q., Zhu, Y., et al. (2012). Proteomics reveals the effects of salicylic acid on growth and tolerance to subsequent drought stress in wheat. *J. Proteome Res.* 11, 6066–6079. doi: 10.1021/pr300728y
- Kang, G. Z., Wang, Z. X., Xia, K. F., and Sun, G. C. (2007). Protection of ultrastructure in chilling-stressed banana leaves by salicylic acid. *J. Zhejiang Univ. Sci. B* 8, 277–282. doi: 10.1631/jzus.2007.B0277
- Karioti, A., Kitsaki, C. K., Zygouraki, S., Ziobora, M., Djeddia, S., Skaltsaa, H., et al. (2008). Occurrence of flavonoids in *Ophrys* (Orchidaceae) flower parts. *Flora* 203, 602–609. doi: 10.1016/j.flora.2007.09.009
- Kaur, G., Kumar, S., Nayyar, H., and Upadhyaya, H. D. (2008). Cold stress injury during the pod-filling phase in Chickpea (*Cicer arietinum* L.): effects on quantitative and qualitative components of seeds. *J. Agron. Crop Sci.* 194, 457–464.
- Kawano, T., Furuichi, T., and Muto, S. (2004). Controlled salicylic acid levels and corresponding signaling mechanisms in plants. *Plant Biotech.* 21, 319–335. doi: 10.5511/plantbiotechnology.21.319
- Kawano, T., Sahashi, N., Takahashi, K., Uozumi, N., and Muto, S. (1998). Salicylic acid induces extracellular superoxide generation followed by an increase in cytosolic calcium ion in tobacco suspension culture: the earliest events in salicylic acid signal transduction. *Plant Cell Physiol.* 39, 721–730.
- Kazemi, N., Khavari-Nejad, R. A., Fahimi, H., Saadatmand, S., and Nejad-Sattari, T. (2010). Effects of exogenous salicylic acid and nitric oxide on lipid peroxidation and antioxidant enzyme activities in leaves of *Brassica napus* L. under nickel stress. *Sci. Hortic.* 126, 402–407. doi: 10.1016/j.scienta.2010.07.037
- Kazemi-Shahandashti, S. S., Maali-Amiri, R., Zeinali, H., Khazaei, M., Talei, A., and Ramezani, S. S. (2014). Effect of short-term cold stress on oxidative damage and transcript accumulation of defense-related genes in chickpea seedlings. *J. Plant Physiol.* 171, 1106–1116. doi: 10.1016/j.jplph.2014.03.020
- Khan, M. I. R., Asgher, M., and Khan, N. A. (2013a). Rising temperature in the changing environment: a serious threat to plants. *Climate Change Environ. Sustain.* 1, 25–36. doi: 10.5958/j.2320-6411.1.1.004
- Khan, M. I. R., Iqbal, N., Masood, A., Per, T. S., and Khan, N. A. (2013b). Salicylic acid alleviates adverse effects of heat stress on photosynthesis through changes in proline production and ethylene formation. *Plant Signal. Behav.* 8:e26374. doi: 10.4161/psb.26374
- Khan, M. I. R., Asgher, M., and Khan, N. A. (2014). Alleviation of salt-induced photosynthesis and growth inhibition by salicylic acid involves glycinebetaine and ethylene in mungbean (*Vigna radiata* L.). *Plant Physiol. Biochem.* 80, 67–74. doi: 10.1016/j.plaphy.2014.03.026
- Khan, M. I. R., Iqbal, N., Masood, A., Mobin, M., Anjum, N. A., and Khan, N. A. (2015a). Modulation and significance of nitrogen and sulfur metabolism in cadmium challenged plants. *Plant Growth Regul.* doi: 10.1007/s10725-015-0071-9
- Khan, M. I. R., Nazir, F., Asgher, M., Per, T. S., and Khan, N. A. (2015b). Selenium and sulfur influence ethylene formation and alleviate cadmium-induced oxidative stress by improving proline and glutathione production in wheat. *J. Plant Physiol.* 173, 9–18. doi: 10.1016/j.jplph.2014.09.011
- Khan, M. I. R., and Khan, N. A. (2013). Salicylic acid and jasmonates: approaches in abiotic stress. *J. Plant Biochem. Physiol.* 1:e113. doi: 10.4172/2329-9029.1000e113
- Khan, M. I. R., and Khan, N. A. (2014). Ethylene reverses photosynthetic inhibition by nickel and zinc in mustard through changes in PSII activity, photosynthetic nitrogen use efficiency, and antioxidant metabolism. *Protoplasma* 251, 1007–1019. doi: 10.1007/s00709-014-0610-7
- Khan, N. A., Nazar, R., Iqbal, N., and Anjum, N. A. (2012a). *Phytohormones and Abiotic Stress Tolerance in Plants*. Berlin: Springer. doi: 10.1007/978-3-642-25829-9
- Khan, M. I. R., Syeed, S., Nazar, R., and Anjum, N. A. (2012b). "An insight into the role of salicylic acid and jasmonic acid in salt stress tolerance," in *Phytohormones and Abiotic Stress Tolerance in Plants*, eds N. A. Khan, R. Nazar, N. Iqbal, and N. A. Anjum (Berlin: Springer), 277–300. doi: 10.1007/978-3-642-25829-9\_12
- Khan, M. I. R., Iqbal, N., Masood, A., and Khan, N. A. (2012c). Variation in salt tolerance of wheat cultivars: role of glycinebetaine and ethylene. *Pedosphere* 22, 746–754. doi: 10.1016/S1002-0160(12)60060-5
- Khan, N., Syeed, S., Masood, A., Nazar, R., and Iqbal, N. (2010). Application of salicylic acid increases contents of nutrients and antioxidative metabolism in mungbean and alleviates adverse effects of salinity stress. *Intl. J. Plant. Biol.* 1:e1. doi: 10.4081/pb.2010.e1
- Khokon, M., Okuma, E. I. J. I., Hossain, M. A., Munemasa, S., Uraji, M., Nakamura, Y., et al. (2011). Involvement of extracellular oxidative burst in salicylic acid-induced stomatal closure in *Arabidopsis*. *Plant Cell Environ.* 34, 434–443. doi: 10.1111/j.1365-3040.2010.02253.x
- Kiddle, G. A., Doughty, K. J., and Wallsgrove, R. M. (1994). Salicylic acid-induced accumulation of glucosinolates in oilseed rape (*Brassica napus* L.) leaves. *J. Exp. Bot.* 45, 1343–1346. doi: 10.1093/jxb/45.9.1343
- Koch, J. R., Creelman, R. A., Eshita, S. M., Seskar, M., Mullet, J. E., Davis, K. R., et al. (2000). Ozone sensitivity in hybrid poplar correlates with insensitivity to both salicylic acid and jasmonic acid. The role of programmed cell death in lesion formation. *Plant Physiol.* 123, 487–496. doi: 10.1104/pp.123.2.487
- Kolbe, H., and Lautemann, E. (1860). Über die constitution und basicität der salicylsäure. *Justus Liebigs Ann. Chem.* 115, 157–206. doi: 10.1002/jlac.18601150207
- Kong, J., Dong, Y., Xu, L., Liu, S., and Bai, X. (2014). Effects of foliar application of salicylic acid and nitric oxide in alleviating iron deficiency induced chlorosis of *Arachis hypogaea* L. *Bot. Stud.* 55:9. doi: 10.1186/1999-3110-55-9
- Kováčik, J., Grúz, J., Baėkor, M., Srnađ, M., and Repcák, M. (2009). Salicylic acid-induced changes to growth and phenolic metabolism in *Matricaria chamomilla* plants. *Plant Cell Rep.* 28, 135–143. doi: 10.1007/s00299-008-0627-5
- Kovács, V., Gondor, O. K., Szalai, G., Darkó, E., Majláth, I., Janda, T., et al. (2014). Synthesis and role of salicylic acid in wheat varieties with different levels of cadmium tolerance. *J. Hazard. Mater.* 280, 12–19. doi: 10.1016/j.jhazmat.2014.07.048
- Krantev, A., Yordanova, R., Janda, T., Szalai, G., and Popova, L. (2008). Treatment with salicylic acid decreases the effect of cadmium on photosynthesis in maize plants. *J. Plant Physiol.* 165, 920–931. doi: 10.1016/j.jplph.2006.11.014
- Kumar, D. (2014). Salicylic acid signaling in disease resistance. *Plant Sci.* 228, 127–124. doi: 10.1016/j.plantsci.2014.04.014
- Larkindale, J., Hall, J. D., Knight, M. R., and Verling, E. (2005). Heat stress phenotypes of *Arabidopsis* mutants implicate multiple signaling pathways in the acquisition of thermotolerance. *Plant Physiol.* 138, 882–897. doi: 10.1104/pp.105.062257
- Larkindale, J., and Knight, M. R. (2002). Protection against heat stress induced oxidative damage in *Arabidopsis* involves calcium, abscisic acid, ethylene, and salicylic acid. *Plant Physiol.* 128, 682–695. doi: 10.1104/pp.010320
- Leclercq, J., Ranty, B., Sanchez-Ballesta, M. T., Li, Z., Jauneau, A., Pech, J.-C., et al. (2005). Molecular and biochemical characterization of LeCRK1, a ripening-associated tomato CDPK-related kinase. *J. Exp. Bot.* 56, 25–35.
- Lee, S. Y., Damodaran, P. N., and Roh, K. S. (2014). Influence of salicylic acid on rubisco and rubisco activase in tobacco plant grown under sodium chloride *in vitro*. *Saudi J. Biol. Sci.* 21, 417–426. doi: 10.1016/j.sjbs.2014.04.002
- Leroux, H. (1830). Discovery of salicine. *J. Chim. Med.* 6, 340–432.

- Leslie, C. A., and Romani, R. J. (1986). Salicylic acid: a new inhibitor of ethylene biosynthesis. *Plant Cell Rep.* 5, 144–146. doi: 10.1007/BF00269255
- Li, G., Peng, X., Wei, L., and Kang, G. (2013). Salicylic acid increases the contents of glutathione and ascorbate and temporally regulates the related gene expression in salt-stressed wheat seedlings. *Gene* 529, 321–325. doi: 10.1016/j.gene.2013.07.093
- Li, T., Hu, Y., Du, X., Tang, H., Shen, C., and Wu, J. (2014). Salicylic acid alleviates the adverse effects of salt stress in *Torreya grandis* cv. merrillii seedlings by activating photosynthesis and enhancing antioxidant systems. *PLoS ONE* 9:e109492. doi: 10.1371/journal.pone.0109492
- Liu, C., Guo, J., Cui, Y., Lü, T., Zhang, X., and Shi, G. (2011). Effects of cadmium and salicylic acid on growth, spectral reflectance and photosynthesis of castor bean seedlings. *Plant Soil* 344, 131–141. doi: 10.1007/s11104-011-0733-y
- Liu, J., and Zhu, K. (1997). An *Arabidopsis* mutant that requires increased calcium for potassium nutrition and salt tolerance. *Proc. Natl. Acad. Sci. U.S.A.* 94, 14960–14964. doi: 10.1073/pnas.94.26.14960
- Liu, P., Xu, Z. S., Pan-Pan, L., Hu, D., Chen, M., Li, L. C., et al. (2013). A wheat PI4K gene whose product possesses threonine autophosphorylation activity confers tolerance to drought and salt in *Arabidopsis*. *J. Exp. Bot.* 64, 2915–2927. doi: 10.1093/jxb/ert133
- Long, S. P., and Naidu, S. L. (2002). “Effects of oxidants at the biochemical, cell and physiological levels, with particular reference to ozone,” in *Air Pollution and Plant Life*, eds J. N. B. Bell and M. Treshow (West Sussex: John Wiley & Sons, Ltd.), 69–88.
- Luo, Y. L., Su, Z. L., Bi, T. J., Cui, X. L., and Lan, Q. Y. (2014). Salicylic acid improves chilling tolerance by affecting antioxidant enzymes and osmoregulators in *sacha inchi* (*Plukenetia volubilis*). *Braz. J. Bot.* 37, 357–363. doi: 10.1007/s40415-014-0067-0
- Mahdavian, K., Kalantari, K. M., and Ghorbanli, M. (2007). The effect of different concentrations of salicylic acid on protective enzyme activities of pepper (*Capsicum annum* L.) plants. *Pak. J. Biol. Sci.* 10, 3162–3165. doi: 10.3923/pjbs.2007.3162.3165
- Mantri, N., Patade, V., Penna, S., and Ford, R. (2012). “Abiotic stress responses in plants: present and future,” in *Abiotic Stress Responses in Plants*, eds P. Ahmad and M. N. V. Prasad (New York, NY: Springer), 1–19. doi: 10.1007/978-1-4614-0634-1\_1
- Martinez, C., Pons, E., Prats, G., and Leon, J. (2004). Salicylic acid regulates flowering time and links defence responses and reproductive development. *Plant J.* 37, 209–217. doi: 10.1046/j.1365-313X.2003.01954.x
- Mikolajczyk, M., Awotunde, O. S., Muszynska, G., Klessig, D. F., and Dobrowolska, G. (2000). Osmotic stress induces rapid activation of a salicylic acid-induced protein kinase and a homolog of protein kinase ASK1 in tobacco cell. *Plant Cell* 12, 165–178. doi: 10.1105/tpc.12.1.165
- Milla, M. A. R., Maurer, A., Huete, A. R., and Gustafson, J. P. (2003). Glutathione peroxidase genes in *Arabidopsis* are ubiquitous and regulated by abiotic stresses through diverse signaling pathways. *Plant J.* 36, 602–615. doi: 10.1046/j.1365-313X.2003.01901.x
- Mirdehghan, S. H., and Ghotbi, F. (2014). Effects of salicylic acid, jasmonic acid, and calcium chloride on reducing chilling injury of pomegranate (*Punica granatum* L.) fruit. *J. Agric. Sci. Technol.* 16, 163–173.
- Misra, N., and Misra, R. (2012). Salicylic acid changes plant growth parameters and proline metabolism in *Rauwolfia serpentina* leaves grown under salinity stress. *Am. Eurasian J. Agri. Environ. Sci.* 12, 1601–1609.
- Misra, N., and Saxena, P. (2009). Effect of salicylic acid on proline metabolism in lentil grown under salinity stress. *Plant Sci.* 177, 181–189. doi: 10.1016/j.plantsci.2009.05.007
- Miura, K., Okamoto, H., Okuma, E., Shiba, H., Kamada, H., Hasegawa, P. M., et al. (2013). SIZ1 deficiency causes reduced stomatal aperture and enhanced drought tolerance via controlling salicylic acid-induced accumulation of reactive oxygen species in *Arabidopsis*. *Plant J.* 73, 91–104. doi: 10.1111/tpj.12014
- Miura, K., and Tada, Y. (2014). Regulation of water, salinity, and cold stress responses by salicylic acid. *Front. Plant Sci.* 5:4. doi: 10.3389/fpls.2014.00004
- Mohammed, A. R., and Tarpley, L. (2009a). Impact of high nighttime temperature on respiration, membrane stability, antioxidant capacity, and yield of rice plants. *Crop Sci.* 49, 313–322. doi: 10.2135/cropsci2008.03.0161
- Mohammed, A. R., and Tarpley, L. (2009b). Effects of elevated ultraviolet-B radiation on productive tillers, spikelet sterility and grain characteristics of southern US rice (*Oryza sativa* L.) cultivars. *J. Agro. Crop Sci.* 195, 292–300. doi: 10.1111/j.1439-037X.2009.00377.x
- Mohammed, A. R., and Tarpley, L. (2011). Morphological and physiological responses of nine southern U.S. rice cultivars differing in their tolerance to enhanced ultraviolet-B radiation. *Environ. Exp. Bot.* 70, 174–184. doi: 10.1016/j.envexpbot.2010.09.001
- Mohammed, A. R., and Tarpley, L. (2013). Effects of enhanced ultraviolet-B (UV-B) radiation and antioxidative-type plant growth regulators on Rice (*Oryza sativa* L.) leaf photosynthetic rate, photochemistry and physiology. *J. Agric. Sci.* 5, 115–128. doi: 10.5539/jas.v5n5p115
- Molassiotis, A., Diamantidis, G., Therios, I., and Dimassi, K. (2005). Effects of salicylic acid on ethylene induction and antioxidant activity in peach rootstock regenerants. *Biol. Plant.* 49, 609–612. doi: 10.1007/s10535-005-0057-y
- Mullineaux, P., and Karpinski, S. (2002). Signal transduction in response to excess light: getting out of the chloroplast. *Curr. Opin. Plant Biol.* 5, 43–48. doi: 10.1016/S1369-5266(01)00226-6
- Munns, R. (2005). Genes and salt tolerance: bringing them together. *New Phytol.* 167, 645–663. doi: 10.1111/j.1469-8137.2005.01487.x
- Munns, R., and Tester, M. (2008). Mechanism of salinity tolerance. *Annu. Rev. Plant Biol.* 59, 651–681. doi: 10.1146/annurev.arplant.59.032607.092911
- Murakeözy, É. P., Nagy, Z., Duházé, C., Bouchereau, A., and Tuba, Z. (2003). Seasonal changes in the levels of compatible osmolytes in three halophytic species of inland saline vegetation in Hungary. *J. Plant Physiol.* 160, 395–401. doi: 10.1078/0176-1617-00790
- Mustafa, N. R., Kim, H. K., Choi, Y. H., Erkelens, C., Lefeber, A. W., Spijksma, G., et al. (2009). Biosynthesis of salicylic acid in fungus elicited *Catharanthus roseus* cells. *Phytochemistry* 70, 532–539. doi: 10.1016/j.phytochem.2009.01.009
- Mutlu, S., Karadağoğlu, Ö., Atici, Ö., and Nalbantoğlu, B. (2013). Protective role of salicylic acid applied before cold stress on antioxidative system and protein patterns in barley apoplast. *Biol. Plant.* 57, 507–513. doi: 10.1007/s10535-013-0322-4
- Naser Alavi, S. M., Arvin, M. J., and Kalantari, K. M. (2014). Salicylic acid and nitric oxide alleviate osmotic stress in wheat (*Triticum aestivum* L.) seedlings. *J. Plant Interac.* 9, 683–688. doi: 10.1080/17429145.2014.900120
- Nazar, R., Iqbal, N., Syeed, S., and Khan, N. A. (2011). Salicylic acid alleviates decreases in photosynthesis under salt stress by enhancing nitrogen and sulfur assimilation and antioxidant metabolism differentially in two mungbean cultivars. *J. Plant Physiol.* 168, 807–815. doi: 10.1016/j.jplph.2010.11.001
- Nazar, R., Umar, S., and Khan, N. A. (2015). Exogenous salicylic acid improves photosynthesis and growth through increase in ascorbate-glutathione metabolism and S assimilation in mustard under salt stress. *Plant Signal. Behav.* 10:e1003751. doi: 10.1080/15592324.2014.1003751
- Noctor, G., Mhamdi, A., Chaouch, S., Han, Y., Neukermans, J., Marquez-Garcia, B., et al. (2012). Glutathione in plants: an integrated overview. *Plant Cell Environ.* 35, 454–484. doi: 10.1111/j.1365-3040.2011.02400.x
- Noriega, G., Caggiano, E., Lecube, M. L., Cruz, D. S., Batlle, A., Tomaro, M., et al. (2012). The role of salicylic acid in the prevention of oxidative stress elicited by cadmium in soybean plants. *Biometals* 25, 1155–1165. doi: 10.1007/s10534-012-9577-z
- Ogawa, D., Nakajima, N., Sano, T., Tamaoki, M., Aono, M., Kubo, A., et al. (2005). Salicylic acid accumulation under O<sub>3</sub> exposure is regulated by ethylene in tobacco plants. *Plant Cell Physiol.* 46, 1062–1072. doi: 10.1093/pcp/pci118
- Overmyer, K., Brosché, M., and Kangasjärvi, J. (2003). Reactive oxygen species and hormonal control of cell death. *Trends Plant Sci.* 8, 335–342. doi: 10.1016/S1360-1385(03)00135-3
- Pál, M., Kovács, V., Szalai, G., Soós, V., Ma, X., Liu, H., et al. (2014). Salicylic acid and abiotic stress responses in rice. *J. Agro. Crop Sci.* 200, 1–11. doi: 10.1111/jac.12037
- Palma, F., López-Gómez, M., Tejera, N. A., and Lluch, C. (2013). Salicylic acid improves the salinity tolerance of *Medicago sativa* in symbiosis with *Sinorhizobium meliloti* by preventing nitrogen fixation inhibition. *Plant Sci.* 208, 75–82. doi: 10.1016/j.plantsci.2013.03.015
- Pawlak-Sprada, S., Arasimowicz-Jelonek, M., Podgórska, M., and Deckert, J. (2011). Activation of phenylpropanoid pathway in legume plants exposed to heavy metals. Part I. Effects of cadmium and lead on phenylalanine ammonia-lyase gene expression, enzyme activity and lignin content. *Acta Biochim. Pol.* 58, 211–216.



- Peleg, Z., Reguera, M., Tumimbang, E., Walia, H., and Blumwald, E. (2011). Cytokinin-mediated source/sink modifications improve drought tolerance and increase grain yield in rice under water-stress. *Plant Biotech. J.* 9, 747–758. doi: 10.1111/j.1467-7652.2010.00584.x
- Pitman, M. G., and Lauchli, A. (2002). “Global impact of salinity and agricultural ecosystem,” in *Salinity: Environment-Plants-Molecules*, eds A. Lauchli and U. Luttge (Dordrecht: Kluwer Academic Publishers), 3–20.
- Poór, P., Kovács, J., Szopkó, D., and Tari, I. (2013). Ethylene signaling in salt stress- and salicylic acid-induced programmed cell death in tomato suspension cells. *Protoplasma* 250, 273–284. doi: 10.1007/s00709-012-0408-4
- Ramakrishna, A., and Ravishankar, G. A. (2011). Influence of abiotic stress signals on secondary metabolites in plants. *Plant Signal. Behav.* 6, 1720–1731. doi: 10.4161/psb.6.11.17613
- Ranceliene, V., and Vyšniauskienė, R. (2012). Modification of UV-B radiation effect on *Crepis capillaris* by antioxidant and environmental conditions. *Emirates J. Food Agric.* 24, 614–620. doi: 10.9755/ejfa.v24i6.614620
- Rao, M. V., and Davis, K. R. (1999). Ozone-induced cell death occurs via two distinct mechanisms in *Arabidopsis*: the role of salicylic acid. *Plant J.* 17, 603–614. doi: 10.1046/j.1365-313X.1999.00400.x
- Rao, M. V., Lee, H. I., Creelman, R. A., Mullet, J. E., and Davis, K. R. (2000). Jasmonate perception desensitizes O<sub>3</sub>-induced salicylic acid biosynthesis and programmed cell death in *Arabidopsis*. *Plant Cell* 12, 1633–1646. doi: 10.1105/tpc.12.9.1633
- Rao, M. V., Lee, H. I., and Davis, K. R. (2002). Ozone-induced ethylene production is dependent on salicylic acid, and both salicylic acid and ethylene act in concert to regulate ozone-induced cell death. *Plant J.* 32, 447–456. doi: 10.1046/j.1365-313X.2002.01434.x
- Raskin, I. (1992). Role of salicylic acid in plants. *Annu. Rev. Plant Biol.* 43, 439–463. doi: 10.1146/annurev.pp.43.060192.002255
- Rodas-Junco, B. A., Cab-Guillen, Y., Muñoz-Sanchez, J. A., Vázquez-Flota, F., Monforte-González, M., and Hernández-Sotomayor, S. M. (2013). Salicylic acid induces vanillin synthesis through the phospholipid signaling pathway in *Capsicum chinense* cell cultures. *Plant Signal. Behav.* 8:e26752. doi: 10.4161/psb.26752
- Saruhan, N., Sağlam, A., and Kadioglu, A. (2012). Salicylic acid pretreatment induces drought tolerance and delays leaf rolling by inducing antioxidant systems in maize genotypes. *Acta Physiol. Plant.* 34, 97–106. doi: 10.1007/s11738-011-0808-7
- Sharma, Y. K., Leon, J., Raskin, I., and Davis, K. R. (1996). Ozone-induced responses in *Arabidopsis thaliana*: the role of salicylic acid in the accumulation of defense-related transcripts and induced resistance. *Proc. Natl. Acad. Sci. U.S.A.* 93, 5099–5104. doi: 10.1073/pnas.93.10.5099
- Sharp, G. (1915). The history of the salicylic compounds and of salicin. *Pharma J.* 94, 857.
- Shi, G. R., Cai, Q. S., Liu, Q. Q., and Wu, L. (2009). Salicylic acid-mediated alleviation of cadmium toxicity in hemp plants in relation to cadmium uptake, photosynthesis, and antioxidant enzymes. *Acta Physiol. Plant.* 31, 969–977. doi: 10.1007/s11738-009-0312-5
- Shi, Q., Bao, Z., Zhu, Z., Ying, Q., and Qian, Q. (2006). Effects of different treatments of salicylic acid on heat tolerance, chlorophyll fluorescence, and antioxidant enzyme activity in seedlings of *Cucumis sativa* L. *Plant Growth Regul.* 48, 127–135. doi: 10.1007/s10725-005-5482-6
- Shirasu, K., Nakajima, H., Rajasekhar, V. K., Dixon, R. A., and Lamb, C. (1997). Salicylic acid potentiates an agonist-dependent gain control that amplifies pathogen signals in the activation of defence mechanisms. *Plant Cell* 9, 261–270. doi: 10.1105/tpc.9.2.261
- Siboza, X. I., Bertling, I., and Odindo, A. O. (2014). Salicylic acid and methyl jasmonate improve chilling tolerance in cold-stored lemon fruit (*Citrus limon*). *J. Plant Physiol.* 171, 1722–1731. doi: 10.1016/j.jplph.2014.05.012
- Sullivan, J. H., and Teramura, A. H. (1989). The effect of UV-B radiation on loblolly pine. I. Growth, photosynthesis and pigment production in greenhouse grown seedlings. *Physiol. Plant.* 77, 202–207. doi: 10.1111/j.1399-3054.1989.tb04970.x
- Surplus, S. L., Jordan, B. R., Murphy, A. M., Carr, J. P., Thomas, B., and Mackerness, S. A.-H. (1998). Ultraviolet-B-induced responses in *Arabidopsis thaliana*: role of salicylic acid and reactive oxygen species in the regulation of transcripts encoding photosynthetic and acidic pathogenesis-related proteins. *Plant Cell Environ.* 21, 685–694. doi: 10.1046/j.1365-3040.1998.00325.x
- Syed, S., Anjum, N. A., Nazar, R., Iqbal, N., Masood, A., and Khan, N. A. (2011). Salicylic acid-mediated changes in photosynthesis, nutrients content and antioxidant metabolism in two mustard (*Brassica juncea* L.) cultivars differing in salt tolerance. *Acta Physiol. Plant.* 33, 877–886. doi: 10.1007/s11738-010-0614-7
- Szalai, G., Krantev, A., Yordanova, R., Popova, L. P., and Janda, T. (2013). Influence of salicylic acid on phytochelatin synthesis in *Zea mays* during Cd stress. *Turk. J. Bot.* 37, 708–714.
- Szalai, G., Pál, M., and Janda, T. (2011). Abscisic acid may alter the salicylic acid-related abiotic stress response in maize. *Acta Biol. Szeged.* 55, 155–157.
- Szepesi, Á., Csiszár, J., Gémes, K., Horváth, E., Horváth, F., Simon, M. L., et al. (2009). Salicylic acid improves acclimation to salt stress by stimulating abscisic aldehyde oxidase activity and abscisic acid accumulation, and increases Na<sup>+</sup> content in leaves without toxicity symptoms in *Solanum lycopersicum* L. *J. Plant Physiol.* 166, 914–925. doi: 10.1016/j.jplph.2008.11.012
- Tamaoki, M. (2008). The role of phytohormone signaling in ozone-induced cell death in plants. *Plant Signal. Behav.* 3, 166–174. doi: 10.4161/psb.3.3.5538
- Thaler, J. S., Fidantsef, A. L., Duffey, S. S., and Bostock, R. M. (1999). Trade-offs in plant defense against pathogens and herbivores: a field demonstration of chemical elicitors of induced resistance. *J. Chem. Ecol.* 25, 1597–1609. doi: 10.1023/A:1020840900595
- Tufail, A., Arfan, M., Gurmani, A. R., Khan, A., and Bano, A. (2013). Salicylic acid induced salinity tolerance in maize (*Zea mays*). *Pak. J. Bot.* 45, 75–82.
- Verberne, M. C., Budi Muljono, R. A., and Verpoorte, R. (1999). “Salicylic acid biosynthesis,” in *Biochemistry and Molecular Biology of Plant Hormones*, eds P. P. J. Hooykaas, M. A. Hall, and K. R. Libbenga (Amsterdam: Elsevier Science BV), 295–314. doi: 10.1016/S0167-7306(08)60493-7
- Wang, C., Zhang, S., Wang, P., Hou, J., Qian, J., Ao, Y., et al. (2011). Salicylic acid involved in the regulation of nutrient elements uptake and oxidative stress in *Vallisneria natans* (Lour.) Hara under Pb stress. *Chemosphere* 84, 136–142. doi: 10.1016/j.chemosphere.2011.02.026
- Wang, L. J., Fan, L., Loescher, W., Duan, W., Liu, G. J., Cheng, J. S., et al. (2010). Salicylic acid alleviates decreases in photosynthesis under heat stress and accelerates recovery in grapevine leaves. *BMC Plant Biol.* 10:34. doi: 10.1186/1471-2229-10-34
- Wang, L. J., and Li, S. H. (2006). Salicylic acid-induced heat or cold tolerance in relation to Ca<sup>2+</sup> homeostasis and antioxidant systems in young grape plants. *Plant Sci.* 170, 685–694. doi: 10.1016/j.plantsci.2005.09.005
- Wang, W., Vinocur, B., and Altman, A. (2003). Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance. *Planta* 218, 1–14. doi: 10.1007/s00425-003-1105-5
- Wang, X., Manning, W., Feng, Z., and Zhu, Y. (2007). Ground-level ozone in China: distribution and effects on crop yields. *Environ. Pollut.* 147, 394–400. doi: 10.1016/j.envpol.2006.05.006
- Wang, Y., Mopper, S., and Hasenstein, K. H. (2001). Effects of salinity on endogenous ABA, IAA, JA, and SA in *Iris hexagona*. *J. Chem. Ecol.* 27, 327–342. doi: 10.1023/A:1005632506230
- Weissmann, G. (1991). Aspirin. *Sci. Am.* 264, 84–90. doi: 10.1038/scientificamerican0191-84
- Wen, P. F., Chen, J. Y., Wan, S. B., Kong, W. F., Zhang, P., Wang, W., et al. (2008). Salicylic acid activates phenylalanine ammonia-lyase in grape berry in response to high temperature stress. *Plant Growth Regul.* 55, 1–10. doi: 10.1007/s10725-007-9250-7
- White, P. J., and Broadley, M. R. (2003). Calcium in plants. *Ann. Bot.* 92, 487–511. doi: 10.1093/aob/mcgl64
- Wildermuth, M. C., Dewdney, J., Wu, G., and Ausubel, F. M. (2001). Isochorismate synthase is required to synthesize salicylic acid for plant defence. *Nature* 414, 562–565. doi: 10.1038/35107108
- Xu, E., and Brosché, M. (2014). Salicylic acid signaling inhibits apoplastic reactive oxygen species signaling. *BMC Plant Biol.* 14:155. doi: 10.1186/1471-2229-14-155
- Yalpani, N., Enyedi, A. J., León, J., and Raskin, I. (1994). Ultraviolet light and ozone stimulate accumulation of salicylic acid, pathogenesis-related proteins and virus resistance in tobacco. *Planta* 193, 372–376. doi: 10.1007/BF00201815
- Yang, T., and Poovaiah, B. W. (2002). A calmodulin-binding/CGCG box DNA-binding protein family involved in multiple signaling pathways in plants. *J. Biol. Chem.* 277, 45049–45058. doi: 10.1074/jbc.M207941200



- Yoshida, S., Tamaoki, M., Ioki, M., Ogawa, D., Sato, Y., Aono, M., et al. (2009). Ethylene and salicylic acid control glutathione biosynthesis in ozone-exposed *Arabidopsis thaliana*. *Physiol. Plant.* 136, 284–298. doi: 10.1111/j.1399-3054.2009.01220.x
- Yuan, Z., Cong, G., and Zhang, J. (2014). Effects of exogenous salicylic acid on polysaccharides production of *Dendrobium officinale*. *South Afr. J. Bot.* 95, 78–84. doi: 10.1016/j.sajb.2014.08.007
- Zengin, F. (2014). Exogenous treatment with salicylic acid alleviating copper toxicity in bean seedlings. *Proc. Natl. Acad. Sci. India Sec. B Biol. Sci.* 84, 749–755. doi: 10.1007/s40011-013-0285-4
- Zhang, Y., Xu, S., Yang, S., and Chen, Y. (2015). Salicylic acid alleviates cadmium-induced inhibition of growth and photosynthesis through upregulating antioxidant defense system in two melon cultivars (*Cucumis melo* L.). *Protoplasma* 252, 911–924. doi: 10.1007/s00709-014-0732-y
- Zottini, M., Costa, A., De Michele, R., Ruzzene, M., Carimi, F., and Lo Schiavo, F. (2007). Salicylic acid activates nitric oxide synthesis in *Arabidopsis*. *J. Exp. Bot.* 58, 1397–1405. doi: 10.1093/jxb/erm001

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Khan, Fatma, Per, Anjum and Khan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.