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# **Selection of internal reference genes for SYBR green qRT-PCR studies of rhesus monkey (*Macaca mulatta*) tissues**

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## Abstract

**Background** The rhesus monkey (*Macaca mulatta*) is a valuable and widely used model animal for biomedical research. However, quantitative analyses of rhesus gene expression profiles under diverse experimental conditions are limited by a shortage of suitable internal controls for the normalization of mRNA levels. In this study, we used a systematic approach for the selection of potential reference genes in the rhesus monkey and compared their suitability to that of the corresponding genes in humans.

**Results** Eight housekeeping genes (HKGs) (*GAPDH*, *SDHA*, *ACTB*, *RPL13A*, *RPL32*, *UBA52*, *PGK1Y*, and *YWHAZ*) from rhesus monkeys and humans were selected to test for normalization of expression levels in six different tissue types (brain, colon, kidney, liver, lung, and stomach). Their stability and suitability as reference genes were validated by *geNorm*, *NormFinder* and *BestKeeper* programs. Intriguingly, *RPL13A* and *RPL32* were selected as ideal reference genes only in rhesus monkeys.

**Conclusions** The results clearly indicated the necessity of using different reference genes for normalization of expression levels between rhesus monkeys and humans in various tissues.

## Background

The rhesus monkey (*Macaca mulatta*) has been one of the most valuable experimental model species for biomedical studies in various research fields including microbiology, neuroscience, and biochemistry. Recently, the genome sequence of the rhesus monkey was determined by the Rhesus Macaque Genome Sequencing and Analysis Consortium [1]. The consortium revealed that the average sequence similarity between humans and rhesus monkeys is 93%. Approximately twenty thousand genes were annotated or predicted by the human-based comparative analysis. In order to conduct biomedical research in areas such as infectious disease, cancer research, and drug development using various monkey tissues, quantitative gene expression analysis is essential.

Quantitative real-time PCR (qRT-PCR) is a recently developed and popularized molecular technique that can be used for gene expression studies that require highly sensitive and accurate quantification of mRNA levels from various samples [2]. These PCR-based analyses allow investigators to analyze a limited amount of material of interest while guaranteeing accurate quantification of target mRNAs. However, to achieve optimal results in qRT-PCR analysis, certain steps are indispensable including: (1) quality control of the material of interest and the primer sets, (2) determination of the PCR efficiency, and (3) selection of suitable internal control genes for normalization [3]. The availability of advanced biomaterials including sophisticated PCR instrumentation, kits for RNA isolation from various materials, and highly purified Taq polymerase, has overcome many of these difficult steps with the exception of identifying appropriate internal control genes [4]. Different expression profiling results from the same target genes can be obtained by using different HKGs as internal references [5]. Therefore, selection of proper HKGs is important for the accurate analysis and normalization of data generated by real-time qRT-PCR [2].

Before the systematic selection of reference genes, researchers interested in quantitative expression analysis have used empirically determined housekeeping genes such as *ACTB* and *GAPDH*. However, recent studies have provided solid evidence that transcription levels of housekeeping genes vary between cell types [6-11], developmental stages [12,13], and experimental conditions [14,15], individuals [16-20]. Therefore, stable reference gene selection procedures have to be undertaken before quantitative mRNA expression analysis can be conducted.

In this study, three different specific tools (*geNorm*, *Normfinder*, and *BestKeeper*) that can be used for the validation of the stability of selected HKGs (*GAPDH*, *SDHA*, *ACTB*, *RPL13A*, *RPL32*, *UBA52*, *PGK1Y*, and *YWHAZ*) were investigated in rhesus monkey and human tissues (brain, colon, kidney, liver, lung, stomach) using real-time qRT-PCR with SYBR green.

## Results and discussion

### Reference gene and qRT-PCR primer selection

For the selection of suitable reference genes from the human and rhesus monkey genomes, the eight most commonly used and available HKGs were selected: Hydroxymethylbilane synthase (*HMBS*), Beta-actin (*ACTB*), Glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*), Ribosomal protein 13 (*RPL13A*), Ribosomal protein 32 (*RPL32*), Tyrosine 3-monooxygenase (*YWHAZ*), Hypoxanthine phosphoribosyltransferase 1(*HPRT1*), and TATA box-binding protein (*TBP*). Rhesus monkey HKGs are identified by comparison of orthologous loci of the human HKGs. Summarized sequence information for the human and rhesus monkey genes is shown in Table 1. This gene information was also used to design qRT-PCR primers (Table 2).

### **Amplification efficiency**

The expression levels of HKGs were analyzed using the relative quantification (delta-Ct method) [21]. The amplification efficiencies and correlation coefficients ( $R^2$ ) of the eight HKGs were generated using the slopes of the standard curves obtained by serial dilutions. Standard curves with a ten-fold dilution series were used to calculate the amplification efficiency (additional files 1 and 2). Similar amplification efficiencies for the corresponding human and rhesus monkey reference genes are a prerequisite for accurate real-time RT-PCR quantification. The amplification efficiency was calculated by the formula:  $\text{efficiency (\%)} = (10^{(-1/\text{slope})} - 1) * 100$ . The efficiency range of the qRT-PCR amplifications for all of the genes tested was between 80% and 115%. Overall, similar efficiency levels were observed between human and rhesus samples (additional file 3). The amplification specificity for each qRT-PCR analysis was confirmed by melting curve analysis (additional files 4 and 5).

### **Data analysis**

We analyzed eight housekeeping genes for the selection of stable HKGs using three authorized programs (*geNORM*, *NormFinder*, and *BestKeeper*) [22, 23, 24].

#### ***geNorm***

The normalization step must be conducted very carefully for accurate gene expression level analysis. However, the selection of stable HKGs for normalization is very complicated. *geNorm* is a useful tool for the selection of the most stable internal control genes using the principle that the expression ratio of two ideal internal control genes is identical in all tested samples [22].

Using the *geNorm* program, the eight HKGs selected for normalization were ranked according to their M values. The M value is the average pair-wise variation of a particular gene with all other control genes. The M values for *RPL13A*, *RPL32*, *TBP*, *HMBS*, *ACTB*, *YWHAZ*, *HPRT1*, and *GAPDH* in rhesus were lower than 1.5. *RPL32* and *RPL13A* displayed the most stability of all eight genes according to *geNorm* analysis. Both genes encode components of the 60S ribosomal subunit that is involved in protein biosynthesis (Fig.1a). In contrast to rhesus monkeys, human tissue analysis indicated that *GAPDH* and *HMBS* were the most stable genes (Fig. 1c). The M values of all tested human genes except for *HPRT1* were lower than 1.5. To investigate the optimal number of reference genes, it was determined whether the stepwise inclusion of less stable genes significantly affected the normalization factors, considering the term V (Fig. 1).

As the most stable reference genes, *RPL32* and *RPL13A* yielded a V value of 0.200 in the rhesus monkey (Fig. 1b). That value indicated that it would not be necessary to include additional reference genes in qRT-PCR of rhesus monkey tissues. However, in the case of humans, the addition of a third and fourth gene significantly affected the reliability of the selected HKGs (Fig. 1d).

The results of the *geNorm* tool indicated different results between humans and rhesus monkeys. *RPL32* and *RPL13A* in the rhesus monkey and *GAPDH*, *HMBS*, *ACTB* and *RPL32* in humans are the most stable HKGs for the accurate calculation of normalization factors.

### ***NormFinder***

*NormFinder* is another reference tool for identifying the ideal normalization genes among various candidate HKGs [23]. *NormFinder* ranks the various candidate reference genes according to their expression variation between inter and intra groups.

This analysis identified *HMBS* and *RPL32* as the most stable genes in rhesus monkeys and humans, respectively (Table 3). These rankings appear to be generally different from the results of *geNorm*. However, *RPL32*, *GAPDH* and *ACTB* in humans and *HMBS* and *RPL32* in rhesus monkeys are still ranked as the most stable reference genes. Although *geNorm* and *NormFinder* did not identify the exact same reference genes, *GAPDH* in humans and *HMBS* in rhesus monkeys are still considered candidate reference genes.

Intriguingly, human *HMBS* showed opposite results between *geNorm* and *Normfinder* program. In the *geNorm* software, the most important factor is gene expression stability (M). The principle of *geNorm* is that the expression ratio of two ideal internal control genes is identical in all tested samples [22]. Thus, the *geNorm* program did not consider the other HKGs. However, *Normfinder* program focuses on the two genes with the least intra-and inter-group expression variation. In other words, the *Normfinder* program considered other HKGs for the selection of good HKGs. For example, high rate of expression variation of *HMBS* among HKGs could result in the discordance between *Normfinder* and *geNorm* (Fig. 2).

### ***BestKeeper***

The BestKeeper program index was created using the geometric mean of each candidate gene's Ct values [24]. Initial analysis of the data, based on the inspection of raw Ct values, calculated variations for all the housekeeping genes in the six samples (SD[ $\pm$  Ct] and CV [%Ct]), and showed an overall stability in gene expression. Genes with SD values higher than 1 were considered inconsistent. *ACTB*, *HMBS*, *RPL13A*, *RPL32*, *TBP*, *YWHAZ* showed a SD value lower than 1 in the rhesus monkey. Among them, *RPL13A* (SD = 0.22) had the lowest Ct value variation. *GAPDH* had the highest



Ct value variation. In human, *HMBS* and *RPL32* exhibit SD values lower than 1. *RPL32* had the lowest Ct value variation and *HPRT1* had the highest Ct value variation (Table 4). In the case of *HMBS* and *RPL32*, *BestKeeper* also showed a different result compared to *Normfinder* and *geNorm* program, respectively. Because the variation of Ct value of *HMBS* and *RPL32* is ranked very low, the *BestKeeper* program indicated *HMBS* and *RPL32* as good candidate HKGs. However, compared to *BestKeeper*, *geNorm* is considered a pair-wise variation between the genes as the most important factor. Thus *geNorm* did not indicate one gene as a best reference gene but indicated a gene pair.

Based on the *BestKeeper* program, the *RPL13A* gene in the rhesus monkey and the *RPL32* gene in the human were selected as the best reference genes.

## **Validation**

To validate our results from the rhesus monkey, one unknown rhesus monkey was tested using same reference genes. And results were analyzed for the selection of stable HKGs using three authorized programs (*geNORM*, *NormFinder*, and *BestKeeper*) (additional files 6, 7 and 8). Although, slight differences were observed, overall patterns were coincided with our results of *RPL13A* and *RPL32* as ideal reference genes in rhesus monkeys.

## **Conclusions**

In this study, we investigated for the first time the most reliable HKGs for the normalization of real-time qRT-PCR data in rhesus monkey tissues (brain, colon, kidney, liver, lung, stomach) using three programs (*geNorm*, *Normfinder*, and *Bestkeeper*). The results from the rhesus monkey were compared to those of humans.

Our results indicated that the two most stable genes, *RPL13A* and *RPL32*, covering a broad expression range, could be used as reference genes for relative gene quantification and normalization purposes in gene profiling studies of the rhesus monkey as determined by three reference gene identification programs.

In humans, *RPL32*, *HMBS* and *GAPDH* were found to be suitable reference genes. In the case of *RPL32*, two programs (*Bestkeeper* and *Normfinder*) identified this gene as a reliable reference gene. However, *geNorm* indicated that *RPL32* was an unsuitable reference gene by virtue of having a high M value. Likewise, the *GAPDH* gene was rejected by the *Bestkeeper* program as a result of having a high SD[ $\pm$  Ct] value. Finally, *HMBS* was ranked as the sixth best gene by the *Normfinder* program.

Intriguingly, the *GAPDH* and *ACTB* genes are the most widely used reference genes in human studies. However, *GAPDH* and *ACTB* are ranked as the least stable genes in the rhesus monkey. Genomic and environmental differences between humans and rhesus monkeys might result in the selection of different suitable HKGs in the six different tissues tested. Thus, different reference genes (*RPL32* and *RPL13A*) should be used for the analysis of rhesus monkey tissue than those used for human tissue analysis

## **Materials and Methods**

### **RNA samples**

Total RNA from rhesus macaque tissues (colon, liver, brain, lung, stomach, and kidney) and human total RNA (colon, liver, brain, lung, stomach, and kidney) were purchased from Clontech and Ambion, respectively. For the validation of our results for reference gene selection, corresponding unknown rhesus macaque tissues (colon, liver, brain, lung, stomach, and kidney) were provided by the National Primate

Research Center (NPRC) of Korea. Total RNA was quantified using a NanoDrop<sup>®</sup> ND-1000 UV-Vis Spectrophotometer.

### **Rhesus macaque and human tissue cDNA synthesis**

To eliminate DNA contamination from the total RNA samples, Turbo DNA-free<sup>TM</sup> (Ambion) was used. A no-RT control was also amplified to confirm the absence of DNA contamination. M-MLV (Moloney-Murine Leukemia Virus) reverse transcriptase with an annealing temperature of 42°C was used for the reverse transcription reaction with RNase inhibitor (Promega).

### **Candidate reference genes and primers for real-time quantitative RT-PCR**

Rhesus monkey reference genes were identified by comparison with human HKGs using the NCBI (National Center for Biotechnology Information) database and the UCSC (University of California Santa Cruz) genome browser (Table 1). Their sequences were used to design primers design using Primer3 software [25]. BLAST searches were performed to confirm the total gene specificity of the primer sequences, and the results showed the absence of multi-locus matching at the primer site. All primers except for *RPL32* spanned at least one intron to avoid inaccuracy due to genomic DNA contamination in RNA samples.

### **Real-time quantitative RT-PCR**

SYBR green qRT-PCR was performed on a Rotor Gene 3000 (Corbett Research). In each run, 1 ul of cDNA was used as template for amplification per reaction. The sample was added to 19 ul of reaction mixture containing 7 ul H<sub>2</sub>O, 10 ul QuantiTect<sup>®</sup> SYBR<sup>®</sup> Green PCR Master Mix (Qiagen) and 1 ul forward and reverse primers (Table2). Real-time qRT-PCR amplification of the HKGs was carried out for 50 cycles of 94°C for 10 sec, 58°C for 15 sec, and 72°C for 15 sec. The temperature

range for the analysis of melting curves was 55°C to 99°C over 30sec. Three independent experiments were performed.

### **Characterization of analysis programs**

The *geNorm* [22] program provides a measure of gene expression stability (M), which is the mean pair-wise variation between an individual gene and all other tested control genes. This method differs from model-based approaches by comparing genes based on the similarity of their expression profiles. Ct values were converted to scale expression quantities using the delta-Ct method and entered into *geNorm*, which then ranks the genes based on M values where the gene with the most stable expression has the lowest value. *NormFinder* [23] focuses on finding the two genes with the least intra- and inter-group expression variation. A *BestKeeper* [24] index was created using the geometric mean of each candidate gene's Ct values. This index was then compared to each individual candidate housekeeping gene by pair-wise correlation analyses, with each combination assigned a value for the Pearson correlation coefficient (r) and the probability (p).

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### **References**

1. Rhesus Macaque Genome Sequencing and Analysis Consortium: **Evolutionary and Biomedical Insights from the Rhesus Macaque Genome.** *Science* 2007, **316**:222
2. Vandesompele J, De Preter K, Pattyn F, Poppe B, Van Roy N, De Paepe A, Speleman F: **Accurate normalization of real-time quantitative RT-PCR**

- data by geometric averaging of multiple internal control genes.** *Genome Biol* 2002, **3**:34
3. Nolan T, Hands RE, Bustin SA: **Quantification of mRNA using real-time RT-PCR.** *Nat Protoc* 2006, **1**(3):1559-82
  4. Radonic A, Thulke S, Mackay IM, Landt O, Siegert W, Nitsche A: **Guideline to reference gene selection for quantitative realtime PCR.** *Biochem Biophys Res Commun* 2004, **313**:856-862
  5. Sellars MJ, Vuocolo T, Leeton LA, Coman GJ, Degnan BM, Preston NP: **Real-time RT-PCR quantification of Kuruma shrimp transcripts: a comparison of relative and absolute quantification procedures.** *J Biotechnol* 2007, **129**(3):391-9
  6. Schmittgen TD, Zakrajsek BA: **Effect of experimental treatment on housekeeping gene expression: validation by real-time, quantitative RT-PCR.** *J Biochem Biophys Methods* 2000, **46**:69-81
  7. Mogal A, Abdulkadir SA: **Effects of Histone Deacetylase Inhibitor (HDACi); Trichostatin-A (TSA) on the expression of housekeeping genes.** *Mol Cell Probe* 2006, **20**:81–86
  8. Li YP, Bang DD, Handberg KJ, Jorgensen PH, Zhang MF: **Evaluation of the suitability of six host genes as internal control in real-time RT-PCR assays in chicken embryo cell cultures infected with infectious bursal disease virus.** *Micro Biol* 2005, **110**:155–165
  9. Ohl F, Jung M, Radonić A, Sachs M, Loening SA, Jung K: **Identification and Validation of Suitable Endogenous Reference Genes for Gene Expression Studies of Human Bladder Cancer.** *J Urology* 2006, **175**:1915-1920

10. Fischer M, Skowron M, and Berthold F: **Reliable transcript quantification by real-time reverse transcriptase-polymerase chain reaction in primary neuroblastoma using normalization to averaged expression levels of the control genes *HPRT1* and *SDHA*.** *J Mol Diagn* 2005, **7**:1
11. Goidin D, Mamessier A, Staquet MJ, Schmitt D, Berthier-Vergnes O: **Ribosomal 18S RNA prevails over glyceraldehyde-3-phosphate dehydrogenase and beta-actin genes as internal standard for quantitative comparison of mRNA levels in invasive and noninvasive human melanoma cell subpopulations.** *Anal Biochem* 2001, **295**(1):17-21
- 12 . Goossens K, Van Poucke M, Van Soom A, Vandesompele J, Van Zeveren A, Peelman LJ: **Selection of reference genes for quantitative real-time PCR in bovine preimplantation embryos.** *BMC Dev Biol* 2005, **5**:2
13. Kuijk EW, du Puy L, van Tol HT, Haagsman HP, Colenbrander B, Roelen BA: **Validation of reference genes for quantitative RT-PCR studies in porcine oocytes and preimplantation embryos.** *BMC Dev Biol* 2007, **7**:58
14. Tricarico C, Pinzani P, Bianchi S, Paglierani M, Distante V, Pazzagli M, Bustin SA, Orlando C: **Quantitative real-time reverse transcription polymerase chain reaction: normalization to rRNA or single housekeeping genes is inappropriate for human tissue biopsies.** *Anal Biochem* 2002, **309**:293-300
15. Zhang X, Ding L, Sandford AJ: **Selection of reference genes for gene expression studies in human neutrophils by real-time PCR.** *BMC Mol Biol* 2005, **6**:4

16. Overbergh L, Kyama CM, Valckx D, Debrock S, Mwenda JM, Mathieu C, Hooghe TD: **Validation of real-time RT-PCR assays for mRNA quantification in baboons** *Cytokine* 2005, **31(6)**:454-458
17. Nygard AB, Jorgensen CB, Cirera S, Fredholm M: **Selection of reference genes for gene expression studies in pig tissues using SYBR green qPCR.** *BMC Mol Biol* 2007, **8**:67
18. Brinkhof B, Spee B, Rothuizen J, Penning LC: **Development and evaluation of canine reference genes for accurate quantification of gene expression.** *Anal Biochem* 2006, **356**:36-43
19. Spinsanti G, Panti C, Lazzeri E, Marsili L, Casini S, Frati F, Fossi CM: **Selection of reference genes for quantitative RT-PCR studies in striped dolphin (*Stenella coeruleoalba*) skin biopsies.** *BMC Mol Biol* 2006, **7**:32
20. Olsvik PA, Lie KK, Jordal AE, Nilsen TO, Hordvik I: **Evaluation of potential reference genes in real-time RT-PCR studies of Atlantic salmon.** *BMC Mol Biol* 2005, **6**:21
21. Brunner AM, Yakovlev IA, Strauss SH: **Validating internal controls for quantitative plant gene expression studies.** *BMC Plant Biol* 2004, **4**:14
22. geNorm Software [<http://medgen.ugent.be/~jvdesomp/genorm/>]
23. NormFinder Software [<http://www.mdl.dk/publicationsnormfinder.htm>]
24. BestKeeperSoftware[<http://www.genequantification.de/bestkeeper.htm>]
25. Rozen S, Skaletsky HJ: **Primer3 on the WWW for general users and for biologist programmers.** *Bioinformatics Methods and Protocols:Methods in Molecular Biology* 2000: 365-386 [[http://frodo.wi.mit.edu/cgi-bin/primer3/primer3\\_www.cgi](http://frodo.wi.mit.edu/cgi-bin/primer3/primer3_www.cgi)]. Totowa:Humana Press

26. Silver N, Best S, Jiang J, Thein SL: **Selection of housekeeping genes for gene expression studies in human reticulocytes using real-time PCR.** *BMC Mol Biol* 2006, **7**:33
27. Falk O, Monika J, Chuanliang X, Carsten S, Anja R, Mick B, Andreas N, Glen K, Stefan A L, Aleksandar R, Klaus J: **Gene expression studies in prostate cancer tissue: which reference gene should be selected for normalization?** *J Mol Med* 2005, **83**:1014–1024
28. Erkens T, Van Poucke M, Vandesompele J, Goossens K, Van Zeveren A, Peelman LJ: **Development of a new set of reference genes for normalization of real-time RT-PCR data of porcine backfat and longissimus dorsi muscle, and evaluation with PPARGC1A.** *BMC Biotechnol* 2006, **6**:41
29. Pfaffl MW, Tichopad A, Prgomet C & Neuvians TP: **Determination of stable housekeeping genes, differentially regulated target genes and sample integrity: bestkeeper–excel-based tool using pair-wise correlations.** *Biotechnol Lett* 2004, **26**:509–515

## Figure Legends

**Figure 1 - Selection of the most suitable reference genes for normalization in rhesus monkey (a and b) and human (c and d) samples using *geNorm* analysis**

Stepwise exclusion of the least stable genes by calculating the average expression stability (a and c) was conducted. The value of M was calculated for each gene, and the least stable gene with the highest M value was automatically excluded for the next round of calculations. The x-axis from left to right indicates the ranking of the genes



according to their expression stability. Determination of the optimal number of reference genes for normalization (b and d) was conducted. The software calculates the normalization factor from at least two genes at which the variable V defines the pair-wise variation between two sequential normalization factors.

**Figure 2- Logarithmic histogram of the expression levels of 8 internal reference genes determined in 6 different tissues.**

Relative expression levels of rhesus monkey (A) and human (B) with six different tissues.

The respective bar and genes name are: actin beta (ACTB: light blue bar), Glyceraldehyde-3-phosphate dehydrogenase (GAPDH: light purple bar), Hydroxymethylbilane synthase (HMBS: light green bar), Hypoxanthine phosphoribosyltransferase 1 (HPRT1: green bar) Ribosomal protein L13A (RPL13A: dark purple bar), 60S ribosomal protein L32 (RPL32: pink bar), TATA box binding protein (TBP: dark blue bar), Tyrosine 3-monooxygenase (YWHAZ: sky blue bar).

## Tables

**Table 1 - GenBank accession numbers and functions of 8 reference genes in rhesus monkeys and humans**

Symbol	Gene Name	Function	Accession Number	
			Human	Rhesus
GAPDH	Glyceraldehyde-3-phosphate dehydrogenase	Glycolytic enzyme	<a href="#">NM_002046.3</a>	<a href="#">XM_001105471.1</a>
ACTB	Beta-actin	Cytoskeletal structural protein	<a href="#">NM_001101.2</a>	<a href="#">NM_001033084.1</a>
RPL13A	Ribosomal protein L13A	Member of ribosome protein	<a href="#">NM_012423.2</a>	<a href="#">XM_001093017.1</a>
RPL32	60S ribosomal protein L32	Member of ribosome protein	<a href="#">NM_001007074.1</a>	<a href="#">XM_001117016.1</a>
YWHAZ	Tyrosine 3-monooxygenase	Signal transduction by binding to phosphorylated serine residue on a variety of signaling molecules	<a href="#">NM_145690.1</a>	<a href="#">XM_001098275.1</a>
HMBS	hydroxymethylbilane synthase	Third enzyme of the heme biosynthetic pathway and catalyzes the head to tail condensation of four porphobilinogen molecules into the linear hydroxymethylbilane.	<a href="#">NM_000190.3</a>	<a href="#">XM_001101850.1</a>
TBP	TATA box binding protein	Transcription initiation from RNA polymerase II promoter	<a href="#">NM_003194.3</a>	<a href="#">XM_001085016.1</a>
HPRT1	Hypoxanthine phosphoribosyltransferase 1	Metabolic salvage of purines in mammals	<a href="#">NM_000194.2</a>	<a href="#">XM_001097691.1</a>

**Table 2 - Details of primers used for quantitative RT-PCR**

Gene	Forward Primer Sequence [5'→ 3']	Position in cDNA	Reverse Primer Sequence [5'→ 3']	Position in cDNA	Products Size
GAPDH	GAAATCCCATCACCATCTTCCAGG	4th Exon	GAGCCCCAGCCTTCTCCATG	5th Exon	120 bp
ACTB	CCTGGCACCCAGCACAAT	4th Exon	GGGCCGGACTCGTCATAC	5th Exon	144 bp
RPL13A	CCTGGAGGAGAAGAGGAAAGAGA	7th Exon	TTGAGGACCTCTGTGTATTGTCAA	8th Exon	126 bp
RPL32	CAACATTGGTTATGGAAGCAACA	3th Exon	TGACGTTGTGGACCAGGAAC	3th Exon	80 bp
YWHAZ	TCCTTTGCTTGCATCCCAC	6th Exon	AAGGCAGACAATGACAGACCA	6th Exon	132 bp
HMBS	ACCAAGGAGCTTGAACATGC	5th Exon	GAAAGACAACAGCATCATGAG	7th Exon	145 bp
TBP	TGCACAGGAGCCAAGAGTGAA	5, 6th Exon	CACATCACAGCTCCCCACCA	6th Exon	132 bp
HPRT1	AATTATGGACAGGACTGAACGTCTTGCT	2, 3th Exon	TCCAGCAGGTCAGCAAAGAATTTATAGC	3th Exon	117 bp

**Table 3 - Candidate reference genes for normalization and the best combination of two genes listed according to their expression stability calculated by the *NormFinder* program**

Ranking order	Gene name	Stability value of rhesus monkey	Gene name	Stability value of human
1	HMBS	0.349	RPL32	0.39
2	RPL32	0.4	GAPDH	0.594
3	RPL13A	0.434	ACTB	0.668
4	ACTB	0.589	TBP	0.705
5	HPRT1	0.649	YWHAZ	0.754
6	TBP	0.687	HMBS	0.936
7	YWHAZ	0.709	RPL13A	1.073
8	GAPDH	0.837	HPRT1	1.185

**Table 4 - Results from Bestkeeper descriptive statistical analysis**

	GAPDH		ACTB		RPL13A		RPL32		YWHAZ		HMBS		TBP		HPRT1	
n	6		6		6		6		6		6		6		6	
	human	rhesus monkey	human	rhesus monkey	human	rhesus monkey	human	rhesus monkey	human	rhesus monkey	human	rhesus monkey	human	rhesus monkey	human	rhesus monkey
geo Mean [CP]	17.01	15.79	15.03	16.31	19.31	16.83	22.43	25.36	19.30	20.71	27.06	29.87	19.65	17.53	22.89	23.47
ar Mean [CP]	17.04	15.86	15.08	16.31	19.36	16.83	22.44	25.37	19.34	20.72	27.08	29.88	19.71	17.54	22.98	23.50
min [CP]	15.84	13.97	13.25	15.75	17.43	16.53	21.12	24.44	17.62	19.87	26.19	28.26	17.74	17.10	20.44	21.87
max [CP]	18.08	17.85	16.65	16.92	21.71	17.28	23.16	25.81	20.22	21.42	28.76	30.95	22.63	18.15	25.61	24.71
std dev [ $\pm$ CP]	1.02	1.34	1.04	0.30	1.04	0.22	0.63	0.35	1.10	0.54	0.91	0.72	1.31	0.30	1.95	1.04
CV [% CP]	6.00	8.45	6.92	1.85	5.37	1.33	2.79	1.39	5.71	2.60	3.35	2.41	6.62	1.73	8.49	4.41

## **Additional files**

### **Additional file 1 – Standard curves for calculation of PCR efficiency and quantification according to reference genes in the rhesus monkey.**

Amplification of 10-fold serial dilutions of the plasmid standard ranging from  $10^0$  to  $10^5$  copies per reaction was carried out in duplicate.

### **Additional file 2 – Standard curves for calculation of PCR efficiency and quantification according to reference genes in humans.**

Amplification of 10-fold serial dilutions of the plasmid standard ranging from  $10^0$  to  $10^5$  copies per reaction was carried out in duplicate.

### **Additional file 3 – Efficiencies calculated from the slopes of dilution curves**

### **Additional file 4 – Melting curve analysis**

Melting curve analysis of 8 different primer sets for reference genes in the rhesus monkey.

### **Additional file 5 – Melting curve analysis**

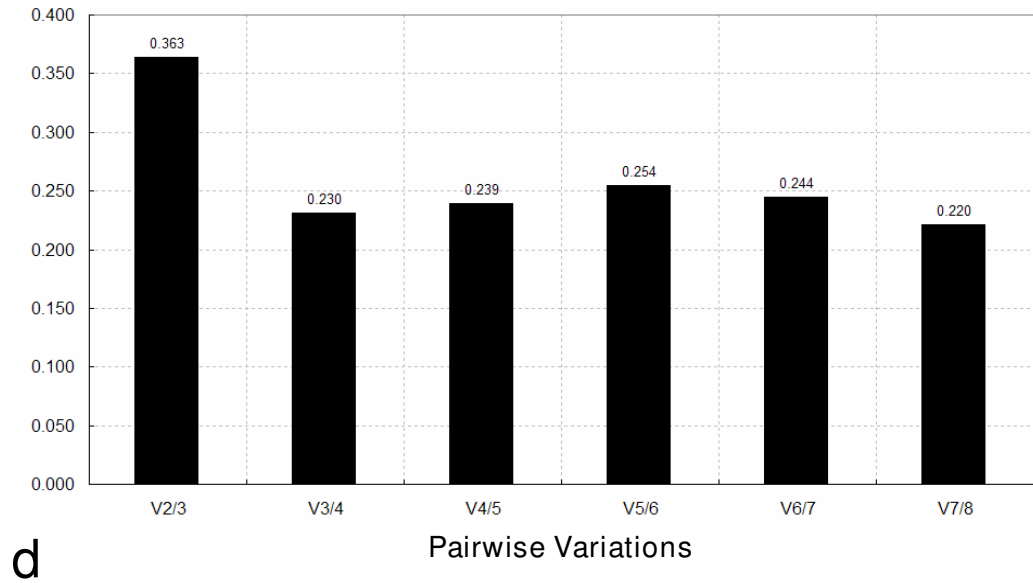
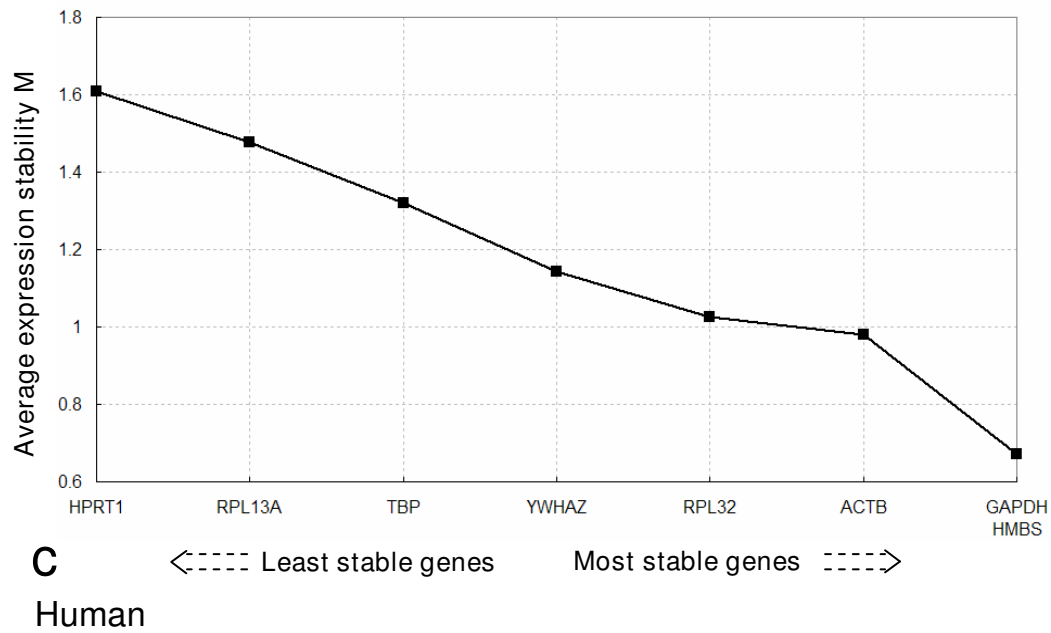
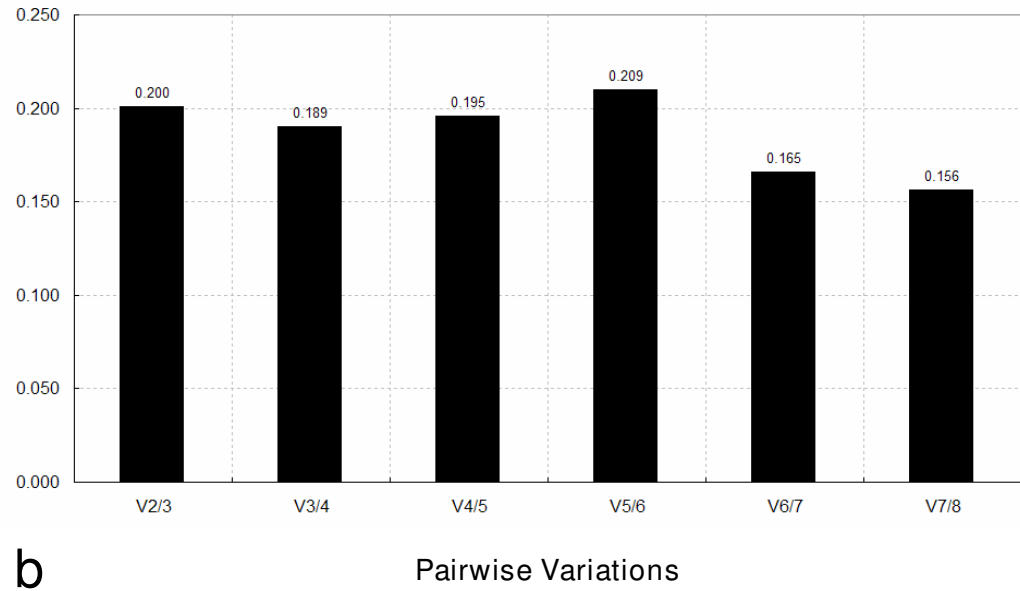
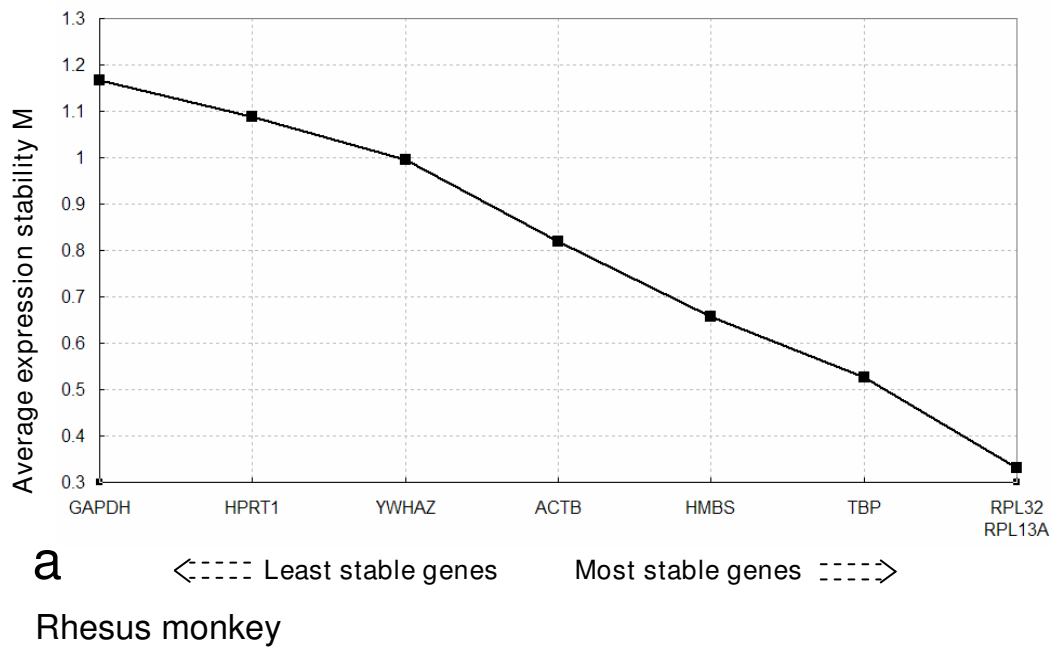
Melting curve analysis of 8 different primer sets for reference genes in humans.

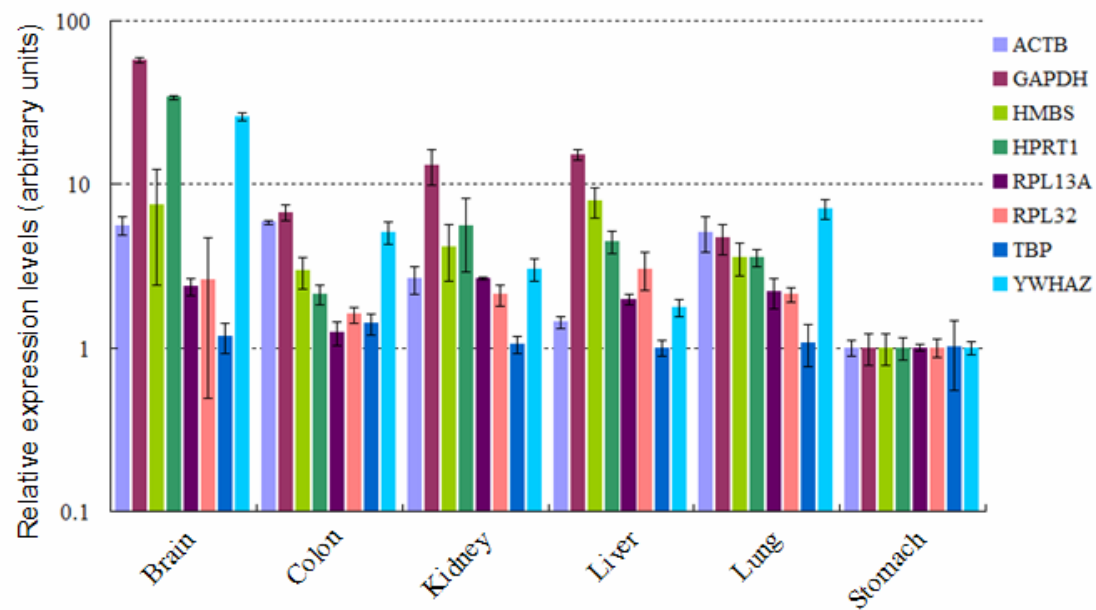
### **Additional file 6 – Selection of the most suitable reference genes for normalization using *geNorm* analysis using the unknown rhesus monkey samples for validation**

### **Additional file 7 – Candidate reference genes for normalization and the best combination of two genes listed according to their expression stability calculated by the NormFinder program using the unknown rhesus monkey samples for validation**

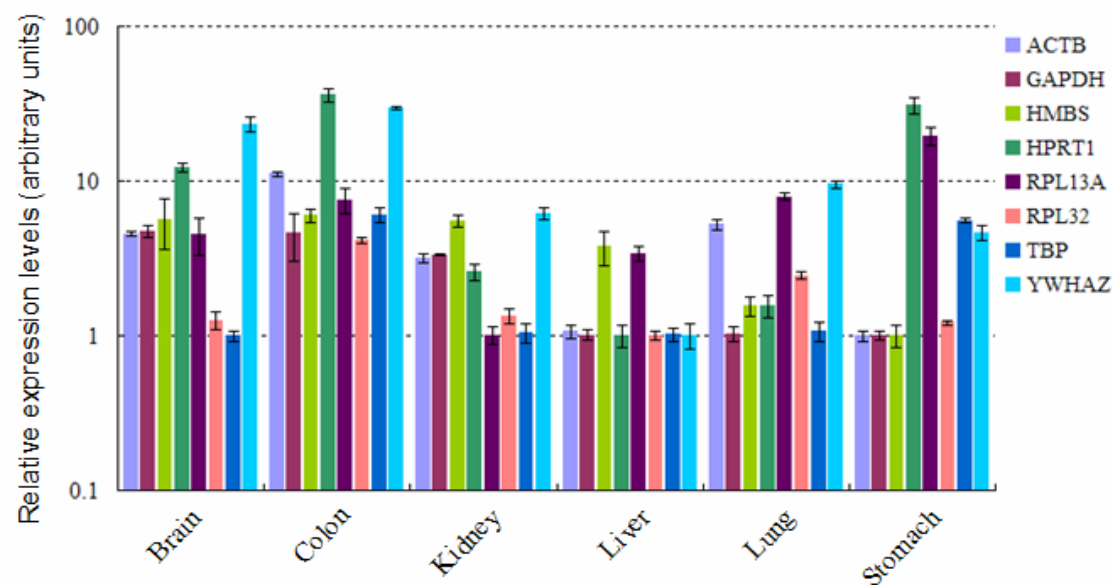
### **Additional file 8 – Results from Bestkeeper descriptive statistical analysis using the unknown rhesus monkey samples for validation**

Figure 1





### A Rhesus



### B Human

### **Additional files provided with this submission:**

Additional file 1: additional file 1 (rhesus standard curve).pdf, 65K

<http://www.biomedcentral.com/imedia/1848329960213806/supp1.pdf>

Additional file 2: additional file 2 (human standard curve).pdf, 54K

<http://www.biomedcentral.com/imedia/8951537421380638/supp2.pdf>

Additional file 3: additional file 3 (amplification efficiency table)).xls, 15K

<http://www.biomedcentral.com/imedia/2140456704213806/supp3.xls>

Additional file 4: additional file 4 (rhesus melting curve ).pdf, 155K

<http://www.biomedcentral.com/imedia/8569087572138069/supp4.pdf>

Additional file 5: additional file 5 (human melting curves ).pdf, 176K

<http://www.biomedcentral.com/imedia/1656426682138064/supp5.pdf>

Additional file 6: additional file 6.pdf, 22K

<http://www.biomedcentral.com/imedia/1725394145213806/supp6.pdf>

Additional file 7: additional file 7(normfinder).xls, 18K

<http://www.biomedcentral.com/imedia/1739325042213807/supp7.xls>

Additional file 8: additional file 8(bestkeeper).xls, 17K

<http://www.biomedcentral.com/imedia/1857462551213806/supp8.xls>