



University of Dundee

Sensors in the Stream

Rode, Michael; Wade, Andrew J.; Cohen, Matthew J.; Hensley, Robert T.; Bowes, Michael J.; Kirchner, James W.

Published in: **Environmental Science and Technology**

DOI:

10.1021/acs.est.6b02155

Publication date: 2016

Document Version Peer reviewed version

Link to publication in Discovery Research Portal

Citation for published version (APA):
Rode, M., Wade, A. J., Cohen, M. J., Hensley, R. T., Bowes, M. J., Kirchner, J. W., Arhonditsis, G. B., Jordan, P., Kronvang, B., Halliday, S. J., Skeffington, R. A., Rozemeijer, J. C., Aubert, A. H., Rinke, K., & Jomaa, S. (2016). Sensors in the Stream: The High-Frequency Wave of the Present. Environmental Science and Technology, 50(19), 10297-10307. https://doi.org/10.1021/acs.est.6b02155

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- · Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain.
 You may freely distribute the URL identifying the publication in the public portal.

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Download date: 26. Aug. 2022





Feature

Sensors in the stream: the high-frequency wave of the present

Michael Rode, Andrew J. Wade, Matthew J. Cohen, Robert T. Hensley, Michael J. Bowes, James W. Kirchner, George B Arhonditsis, Philip Jordan, Brian Kronvang, Sarah J. Halliday, Richard Skeffington, Joachim Rozemeijer, Alice H. Aubert, Karsten Rinke, and Seifeddine Jomaa

Environ. Sci. Technol., Just Accepted Manuscript • DOI: 10.1021/acs.est.6b02155 • Publication Date (Web): 29 Aug 2016

Downloaded from http://pubs.acs.org on August 30, 2016

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a free service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are accessible to all readers and citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.

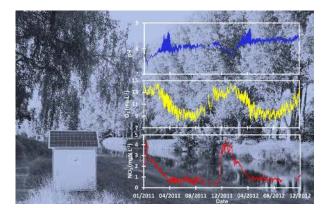
This document is the unedited author's version of a Submitted Work that was subsequently accepted for publication in Environmental Science & Technology, copyright © American Chemical Society after peer review. To access the final edited and published work, see http://pubs.acs.org/doi/abs/10.1021/acs.est.6b02155.



1	Sensors in the stream: the high-frequency wave of the present
2	Michael Rode ^{1,13*} , Andrew J. Wade ^{2,13} , Matthew J. Cohen ³ , Robert T. Hensley ⁴ , Michael J. Bowes ⁵ ,
3	James W. Kirchner ⁶ , George B. Arhonditsis ⁷ , Phil Jordan ⁸ , Brian Kronvang ⁹ , Sarah J. Halliday ² ,
4	Richard A. Skeffington ² , Joachim C. Rozemeijer ¹⁰ , Alice H. Aubert ¹¹ , Karsten Rinke ¹² , Seifeddine
5	Jomaa ¹
6 7	¹ Department of Aquatic Ecosystem Analysis and Management, Helmholtz Centre for Environmental Research-UFZ, Brueckstrasse 3a, D-39114 Magdeburg, Germany, *(michael.rode@ufz.de)
8 9	² Department of Geography and Environmental Sciences, University of Reading, Whiteknights, Reading, United Kingdom (a.j.wade@reading.ac.uk)
10	³ School of Natural Resources and Environment, University of Florida, Gainesville, Florida
11	⁴ School of Forest Resources and Conservation, University of Florida, Gainesville, Florida
12	⁵ Centre for Ecology and Hydrology, Wallingford, Oxon. OX10 8BB, United Kingdom
13 14	⁶ Department of Environmental Sciences, Swiss Federal Institute of Technology-ETH, Zürich, Switzerland; Swiss Federal Research Institute, WSL, Birmensdorf, Switzerland
15 16	⁷ Ecological Modelling Laboratory, Department of Physical and Environmental Sciences, University of Toronto, Toronto, Ontario, Canada
17	⁸ School of Environmental Sciences, Ulster University, Coleraine, N. Ireland, United Kingdom,
18 19	⁹ Department of Bioscience and DCE–National Centre of Environment and Energy, Aarhus University, Vejlsøvej 25, DK-8600 Silkeborg, Denmark
20	¹⁰ Deltares, P.O. Box 85467, NL-3508TA, Utrecht, The Netherlands
21 22	¹¹ Department of Environmental Social Sciences, Eawag, Überlandstrasse 133, 8600 Dübendorf, Switzerland
23 24	¹² Department of Lake Research, Helmholtz Centre for Environmental Research-UFZ, Brueckstrasse 3a, D-39114 Magdeburg, Germany
25	¹³ Both authors contributed equally to this work
26	
27	
28	
29	
30	
31	

Abstract

New scientific understanding is catalysed by novel technologies that enhance measurement precision, resolution or type, and that provide new tools to test and develop theory. Over the last 50 years, technology has transformed the hydrologic sciences by enabling direct measurements of watershed fluxes (evapotranspiration, streamflow) at time scales and spatial extents aligned with variation in physical drivers. High frequency water quality measurements, increasingly obtained by *in-situ* water quality sensors, are extending that transformation. Widely available sensors for some physical (temperature) and chemical (conductivity, dissolved oxygen) attributes have become integral to aquatic science, and emerging sensors for nutrients, dissolved CO₂, turbidity, algal pigments, and dissolved organic matter are now enabling observations of watersheds and streams at timescales commensurate with their fundamental hydrological, energetic, elemental, and biological drivers. Here we synthesize insights from emerging technologies across a suite of applications, and envision future advances, enabled by sensors, in our ability to understand, predict, and restore watershed and stream systems.



48 TOC Art

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

Recent progress in in-situ sensor monitoring

Just over a decade ago, Kirchner et al. ¹ envisioned the hydrologic sciences being transformed by the increased availability of stream chemistry measurements at time scales commensurate with hydrologic forcing, a theme echoed in the U.S. National Research Council's "Challenges and Opportunities in the Hydrologic Sciences" in 2012 (http://www.nap.edu/read/13293/chapter/1). At the same time, ecologists were recognizing the transformative potential of sensors that allow ecosystem processes to be measured at time and space scales that match relevant physical, chemical and biological drivers ²⁻⁴. The vision of Kirchner et al. 1 has been realised, in part, with significant progress in estimating solute residence times in watersheds 5, but it is the converging vision across hydrological, biogeochemical, biological and ecological disciplines that highlights the significant intellectual payoff from new sensor technologies in watershed and stream science. Now stream water chemistry data are available every hour, or even every minute across a broad range of analytes, and commensurate biological data are available at fortnightly to daily intervals for sustained periods greater than one year. These advances allow the study of multiple solutes at sub-daily intervals, not just single solute time series, and enable interpretations and hypothesis testing of ideas around river biogeochemistry, biology and ecology, in addition to catchment signals. These novel measurements have revealed complex temporal dynamics that were obscured by traditional sampling frequencies ⁶⁻⁸ and have enabled new insights into the inner-workings of watersheds and streams.

While automated collection and traditional laboratory processing of discrete samples have yielded enormously informative sub-daily data ⁸⁻⁹, the transformation of stream and watershed science will occur primarily in response to increasing availability of automated *in situ* sensors. Indeed, electrode-based measurements of pH, conductivity, temperature, and dissolved oxygen (DO) have been available for over half a century ¹⁰ and are now essential tools for stream and watershed studies; however, sensor technology has been extended through the development of other methods

such as optical, wet analytical chemical or flow cytometry techniques (a laser- or impedance-based,
biophysical technology employed in cell counting), recent advances in field deployment engineering
(anti-fouling, batteries, micropumps), and electronics (detectors, emitters) that have reduced costs.
This, in turn, has increased the number of sites at which in situ measurements are now made.
Among the solutes for which sensors are most widely available is nitrate. Early colorimetric based
sensors ¹¹ for nitrate were constrained by performance and reagent wastes, and have largely given
way to spectrophotometers ¹² enabling very high frequency (0.5 Hz, samples per second) sampling
that has proven enormously informative for understanding riverine dynamics $^{13-15}$. For other solutes,
wet analytical chemistry remains the most viable approach, with "lab-on-a-chip" sensors lowering
power requirements and reducing the interferences that are intrinsic in optical absorbance
measurements ^{16,17} . For example, measurements of orthophosphate using standard reagent-based
colorimetry has emerged as a robust field-deployable technology, permitting automated hourly
sampling and a host of attendant informative inferences enabled by this increase in temporal
resolution 6,18 . Other deployable optical sensors include fluorimeters that can measure chlorophyll-a
and other photosynthetic pigments, as well as fluorescent dissolved organic matter ¹⁹ ; while these
sensors have a long history in marine and estuarine settings, their use in streams and small
watersheds has revealed a variety of novel insights ²⁰ . Indeed, Fast repetition rate Fluorimetry (FrrF),
a technique which measures the variability of light emission from chlorophyll a, can be used to
measure photosynthetic rates in situ which reduce when algae are stressed due to the prevailing
environmental conditions (e.g. drought), and these measurements are supported by weekly
(imaging) flow cytometry (that can discriminate and assess abundance among phytoplankton and
phytobacterial functional groups) and environmental DNA techniques (that can characterise
microbial communities and detect invasive species) ^{21,22} . In short, the suite of widely used
parameters that hydrologists, geochemists, and stream ecologists consider relevant is almost
uniformly possible in real time and at high spatial or temporal resolution.

These new data sets have the ability to transform our understanding of a diverse range of fundamental aquatic processes, from watershed dynamics to nutrient spiralling to ecosystem response to disturbance. The potential for sensors to unravel ecosystem functioning and realize improved environmental management was illustrated by the recent commissioning of a "national nutrient sensor challenge" by the White House Office of Science and Technology Policy. This effort seeks to enable the next generation of long-term deployable, high accuracy, high precision *in situ* sensors, and to drive down costs to ensure broad adoption by academic, private and government scientists ²³.

Here we present four examples (inferring nutrient sources and transport, measuring *in situ* nutrient processing, detecting ecological effects, and temporal scaling of solute export), spanning continents and time scales, in which recent utilization of sensor technologies have advanced our understanding of stream and watershed systems. While rivers and their watersheds are our focus, the use of novel measurement technologies in other aquatic ecosystems such as lakes, estuaries and oceans has been equally transformative ²⁻⁴. In addition to the insights that have already been made, we highlight ongoing trends in sensor development and suggest areas in which sensors will enable new insights and allow tests of watershed and ecological theory.

Identifying nutrient sources and transport pathways in watersheds

Sub-daily monitoring of nutrient hydrochemistry has traditionally utilized automatic water-samplers, but these are expensive to run in terms of regular sample collection and subsequent laboratory analysis, and can have chemical and biological stability issues during sample storage ²⁴, which usually is in the range of days. Through the deployment of *in situ* sensor and colorimetric based auto-analyser technology, Bowes et al. ²⁵ measured hourly total reactive phosphorus (TRP) and nitrate concentrations and used these to characterise, on a storm-by-storm basis, the nutrient

source changes to a rural river in southern England over a two year period by analysing the hysteresis in the relationship between concentration and flow during storm events when the stream or river flow increases and then recedes. Differences in the hysteresis behavior between storms provide information on nutrient sources and pathways and the findings are summarized in Figure 1. In this case study of the River Enbourne in the UK, the results highlighted the importance of the acute mobilisation of sewage-derived phosphorus in bed sediment and the large diffuse phosphorus inputs entering the stream from manure applications during May storms, thereby helping to target

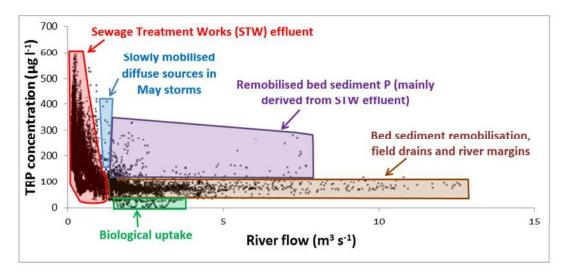


Figure 1: Phosphorus sources to the River Enborne (southern England) identified using two years of hourly total reactive phosphorus (TRP) and flow data. The clusters were derived from storm hysteresis analysis ²⁴. future remediation measures. Additional analysis showed clear double-peaked diel phosphorus and nitrate cycles during low flows, which pointed to chronic pollution related to the daily pattern of effluent discharges from sewage treatment works and septic tank systems ²⁶. Recently Mellander et al. ²⁷ also used 20 minute phosphorus concentration monitoring to identify that subsurface flows in bedrock cracks were the dominant phosphorus transport pathways in a karst landscape in County Mayo, Ireland, and Mellander et al. ²⁸ demonstrated that phosphorus could predominantly be transferred to streams via groundwater during winter in the south of Ireland using sub daily

colorimetric based auto-analyser measurements. Both these studies allowed a deeper

understanding of phosphorus transfer pathways and retention in the aquifer to be developed with advice on location of critical source areas for phosphorus loss in a Karst landscape resultant.

Furthermore, regular sub-daily measurements of dissolved organic matter (DOM) composition (which plays a central role in carbon dynamics and participates in the complexation of trace metals and the mobilization of pollutants) by *in situ* Chromophoric Dissolved Organic Matter (CDOM) fluorescence have revealed a complex short-term variability in DOM composition. This variability is a function of source, flow pathway and instream photochemical and biologically mediated processes ^{20,29}.

Together, these results highlight that sub-daily observations have high potential to accurately make source assignments and that watershed management can greatly benefit from high frequency measurements to identify site specific loss mechanisms and pathways and potential legacy issues ^{25,28}, such as groundwater nitrogen and stream bed phosphorus retention. The development of sensors for an increasing range of water quality constituents, with more widespread deployment, will lead to a greater ability to fingerprint chemical sources through seasons and individual storm events in the future.

Quantifying coupled nutrient processing and metabolism

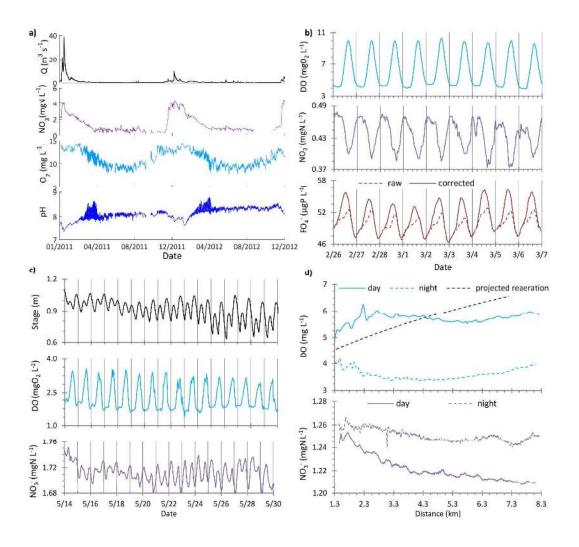


Figure 2. Continuous high frequency oxygen, nitrate and pH data reflect the seasonal pattern of primary production and assimilatory N uptake due to flow and light variability (panel a; adapted from Rode et al. ³²). The relative magnitude of diel variation in DO, NO₃ and PO₄ (panel b; adapted from Cohen et al. ¹⁸) is strongly correlated with autotroph stoichiometry. The assimilatory P uptake (PO₄ signal must be corrected for precipitation) notably appears temporally de-coupled from primary production and assimilatory N uptake by several hours. Comparison of day-night profiles can be used to partition assimilatory versus dissimilatory pathways (panel c ^{33,35}), while high resolution longitudinal profiling (panel d; adapted from Hensley et al. ³³⁾ has been used to identify spatial heterogeneity with N processing in a tidal river shown to be strongly influenced by residence time variation.

Drainage networks are not passive conduits, but are important for chemical retention and transformation. High frequency data have proven especially useful in quantifying in-stream nutrient

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

processing and understanding the stoichiometric coupling of autotrophic uptake across the periodic table. In addition to seasonal patterns (Fig. 2a), finely resolved time-series have revealed strong diel nitrate variability (Fig. 2b) similar to that observed for DO and interpreted as autotrophic N assimilation ^{14,15}. Actively measuring nutrient uptake rates (e.g., via isotope or nutrient dosing) is complex and expensive, limiting measurements to short (hours to days) periods ³⁰, typically under steady baseflow conditions, and with a significant bias towards small streams ³¹. In Florida's spring fed rivers, autotrophic nitrate uptake amounted to less than 20% of total net N retention¹⁸. In two central European streams, percentage daily autotrophic N uptake peaked at 47% (agricultural stream) and 75% (forest stream) of the daily N loading input to the stream network of the whole watershed³². There were different ranges of autotrophic areal rate of nutrient uptake (U, analogues to the mass of nutrient removed from water per unit area of streambed (m⁻²) per unit time (d)) with 30-160 mg N in the Florida rivers 14 and 0-270 mg N and 0-97 mg N in the central European agricultural and forest stream³², respectively. Dissimilatory pathways such as denitrification, which account for the balance of net retention, were also coupled with primary productivity through secondary relationships such as the availability of labile carbon ¹⁴. In a separate study, a more complex retention signal (Fig. 2c) arose in a tidal river, representing the convolution of diel assimilatory uptake and tidally varying denitrification based on residence time and benthic surface area ³³.

While early insights into coupled nutrient processing have focused on nitrate dynamics, sensors for other solutes have proven equally valuable. Cohen et al. ¹⁸ used an ortho-phosphate sensor, along with optical nitrate, and electrode based DO and specific conductance sensors. The high frequency signals enabled identification and deconvolution of geochemical P-retention pathways that created overlapping diel P signals. The data also revealed clear coupling of N and C assimilation, and that while also coupled, P uptake was not synchronous with the timing of N and C assimilation. This asynchronous N and P assimilation may represent timing differences in protein and ribosome production in aquatic plants. It has been suggested that temporal nutrient coupling occurs

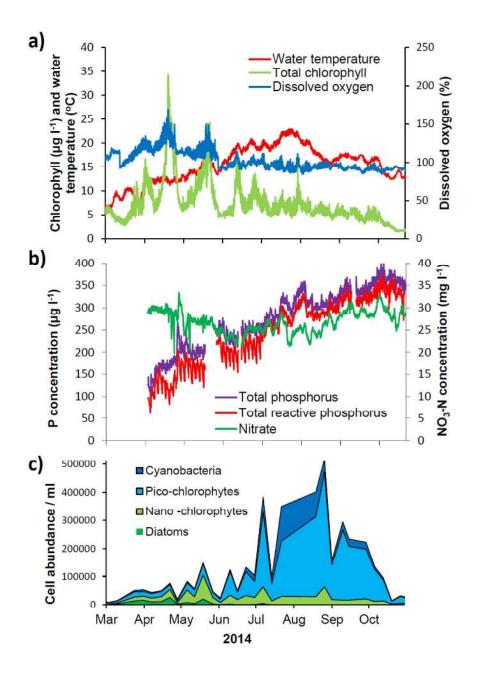
Environmental Science & Technology

only when an input nutrient is limiting and therefore the identification of such temporal coupling, through *in situ* high frequency monitoring, is a useful indicator of ecosystem limitation status ³⁴. Diel concentration variation for biologically active trace metals (e.g., Ba, Fe, Mn, and U) has similarly been observed in spring systems in Florida, USA, and suggests that aquatic plant metabolism controls diel and seasonal cycling of metals ³⁵. As sensors emerge for measuring other solutes such as other nutrients (e.g., Si, Fe, Mg), the organic nutrient forms (e.g. DON, DOP), measurements of total concentrations which include the particulate fraction and therefore all the total nutrient potentially available, and organic pollutants, their dynamics can be compared to metabolic, thermal, flow and photolytic forcing, which will enable a rich new arena for understanding aquatic systems.

Sensors can also be applied using an alternative Lagrangian approach ³⁶. Reach-scale nutrient processing rates such as areal rate of nutrient uptake (U) and uptake length (average distance a nutrient molecule, typically nitrate, moves downstream in dissolved form before being assimilated by the biota) have been estimated from longitudinal changes in stream solute concentrations (Fig. 2d), revealing a large degree of spatial heterogeneity in nutrient uptake which appears related to changes in river morphology³³. Furthermore, estimates of reach-scale metabolism and nutrient processing show diel variations along a continuous gradient from headwaters to mouth, as envisioned in the River Continuum Concept ^{37,38}, and highlight how river regulation disrupts the continuum, for example, in terms of stream metabolism through increased total dissolved N uptake below dams ^{38,39}.

These early insights suggest that the emergence of sensor-derived high-frequency time series for multiple solutes will better allow us to observe the stoichiometric coupling of metabolic and geochemical processes and thereby test stoichiometric theory. It will also enable a deeper understanding of variation in retention rates and pathways with flow, temperature and other abiotic drivers across watersheds spanning a gradient of size and geochemical and physical features.

Separating the effects of multiple processes on aquatic ecology



223

224

225

226

Figure 3: Combined physical (a), chemical (b) and biological monitoring (a, c) of the River Thames at Goring, UK (Unpublished Data, supplied by Centre for Ecology & Hydrology, Wallingford, UK, and the UK Environment Agency). The time axis is the same in the three panels.

The effects of multiple pressures on freshwater ecosystems are difficult to separate because of the multiple, interrelated abiotic-biotic interactions. New biological monitoring techniques have

Environmental Science & Technology

been developed that allow the high-frequency characterisation of river plankton composition and
function for the first time. Fluorimeters now reliably measure total chlorophyll and other
photosynthetic pigments at sub-hourly frequencies to estimate phytoplankton concentrations
(Figure 3a). FrrFs are able to monitor changes in the photosynthetic stress of both chlorophyte and
cyanobacterial communities at sub-hourly intervals 40 . These <i>in situ</i> techniques can be further
supplemented by in situ flow cytometry (Figure 3c), which provides a rapid and simple methodology
to characterise, at high-frequency, the river phytoplankton community by quantifying the cell
concentrations of diatoms, chlorophytes, cryptophytes and different classes of cyanobacteria ⁴¹ . By
combining these new biological data with physico-chemical data of the same high temporal
resolution, it has been demonstrated that it is water temperature, flow and light conditions that are
controlling the onset and magnitude of phytoplankton blooms in the River Thames, rather than
increases in nutrient concentration ⁴² . However, phosphorus and silicon may ultimately terminate
large phytoplankton blooms due to nutrient depletion and limitation ⁴² . Such insights are only
possible through long-term, sub-daily biogeochemical observations that are able to capture the
conditions at the precise time points where chlorophyll concentrations begin to increase or
decrease. In the coming years, high frequency next generation DNA sequencing will provide an even
greater understanding of river microbiological dynamics and their biotic-abiotic interactions ²² .

Quantifying water quality across multiple time scales

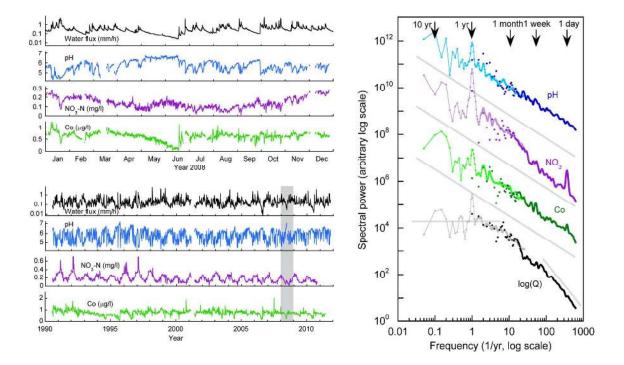


Figure 4. Water quality time series in Upper Hafren streamwater, Plynlimon, Wales, at 7-hour intervals for one year (left upper panel) and weekly intervals for 21 years (left lower panel) (Unpublished Data). Three water quality parameters are shown: pH (an indicator of acid-base status), nitrate (a nutrient that exhibits both diurnal and seasonal cycles) and cobalt (Co). The shaded band in the lower panels shows the time interval covered by the upper panels. The right panel shows power spectra of weekly and 7-hourly time series (light and dark points, respectively), calculated using the methods of Kirchner and Neal ⁸.

Detecting water quality trends requires an understanding of water quality fluctuations over many time scales, including those that are invisible in typical weekly or monthly sampling ⁴³. At several small research watersheds, broad suites of chemical parameters have been measured at daily or higher frequency ^{8,9,44}, facilitating chemical dynamic characterization on shorter timescales as well as those captured by typical monitoring programs (weekly or monthly) (Figure 4 – left panels).

Spectral analysis decomposes a time-series into a spectrum of cycles of different wavelengths with the power spectrum defined by the contribution of each frequency, f, to the time series. The combination of high-frequency and long-term chemical analysis has demonstrated that,

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

on time scales from hours to decades, the power spectrum of mulitple solute time-series, in this case pH (or H $^+$), NO $^-$ 3 and Co, can be characterised as $1/f^\alpha$ noise, where α , the scaling exponent, is approximately equal to 1 (known as "pink noise") 8 . This is shown in Figure 4 (right panel), where the water quality parameters have power-law slopes of -1 (parallel to the long-grey reference lines), indicating that spectral power is inversely proportional to frequency (the 1/f pink noise – which is sometimes referred to as "fractal" noise to emphasise that the scaling exponent, α , can be a non-integer - it does not mean there is self-similarity in the power spectrum). The result implies that three ions do not simply flush through the catchment with the water but that the catchment has a long chemical memory. The stream discharge spectrum, by contrast, has a power-law slope near 0 at low frequencies and -2 at high frequencies (as indicated by the shorter grey reference lines).

Whilst the idea of catchment solute storage is not new, analysis of rainfall and stream water power spectra, for the same solute, allows a transfer function to be derived which can be used to quantify the travel time distribution which is very useful in understanding solute retention in different geographical settings. In addition, the 1/f behaviour of the chemical time series has important implications⁴⁵. Such time series are "non-self-averaging"; they do not converge to stable averages when sampled for longer periods, because their fluctuations do not average out over time. This non-self-averaging behaviour implies that even purely random time series can exhibit spurious trends, on all time scales, which appear to be statistically significant when evaluated by conventional statistics ^{9,46}. Even more disconcertingly, collecting more data makes this problem worse; non-selfaveraging time series exhibit more spuriously "significant" trends (not fewer, as one would expect) when sampled for longer periods, or at higher frequency. Thus, environmental trends should be analysed with more sophisticated statistical methods that are not confounded by the multi-scale correlations that characterize these time series. A recent example of such analysis includes the application of Dynamic Harmonic Regression to use this non-stationary technique to explore stream water nitrate dynamics across decadal to sub-daily timescales and to derive the main cause and effect links at long-term, seasonal and diel time-scales ^{47,48}.

Additional advances

Where a particular water constituent cannot yet be measured directly with an *in situ* sensor, it may still be possible to construct a high frequency time-series for that constituent if there is a strong relationship with a water quality parameter that can be readily measured, such as turbidity, dissolved oxygen, temperature and pH. In this way, such sensor measurements have recently been used as proxies for a range of water quality parameters, including total suspended solids ^{49,50}, alkalinity⁴⁹, total nitrogen⁴⁹, total phosphorus ^{49,50}, sodium⁴⁹, chloride⁴⁹, fluoride⁴⁹, sulfate⁴⁹, fecal coliform bacteria⁴⁹, fluoranthene and mercury⁵¹, polycyclic aromatic hydrocarbons⁵², , and when coupled with discharge measurements allow flux estimation. For example, high resolution *in situ* measurements of turbidity and fluorescence were used to estimate total mercury transport between the San Francisco estuary and an adjacent tidal wetland ⁵³. High frequency water quality measurements can also be used to reconstruct concentration patterns in combination with other commonly available continuous data, such as precipitation or discharge. In this way, Rozemeijer et al. ⁵⁴ reduced the bias of total phosphorus load calculations by up to 63% using 20 events sampled at 15 minute intervals.

Measurements of lake, reservoir, wetland and estuarine diel dynamics help identify internal processing of nutrients and metals. High frequency monitoring in lakes and reservoirs using autonomous vertical profiling systems is increasingly exploited for safeguarding high water quality (e.g., for drinking water abstraction ⁵⁵). Such systems detect river intrusions that may quickly reach water abstraction infrastructure ^{56,57}. High frequency measurements also allow new insights into lake metabolism and help constrain biogeochemical budgets or to differentiate the importance of internal versus external factors ⁵⁸. It has been recently shown that monitoring external watershed loading, as well as within lake chemistry, at high frequency, enables separation of carbon accumulation due to internal phytoplankton dynamics versus external inputs of organic carbon from

runoff events⁵⁹. High frequency oxygen measurements revealed that external seasonal forcing plays a key role in determining the extent to which a lake ecosystem is a seasonal carbon sink or source to the atmosphere ⁶⁰. Further experimental uses of high-frequency sensors extends to monitoring tracers ⁶, changes in artificial environments (e.g., benthic chambers) and in process-control systems, for example, to control iron dosing to co-precipitate phosphorus at Wessex Water's Keynsham Sewage Treatment Works in the UK (http://www.worldpumps.com/view/316/control-of-chemical-dosing-in-wastewater-treatment/).

For parameters with a strong diel variation, such as DO, the value obtained, and thus the classification of the European Union Water Framework Directive (WFD), which commits European Union member states to achieve good qualitative and quantitative status of all water bodies, can depend markedly on the frequency of sampling. In lowland UK river-systems, monthly sampling for a year can result in the same water body being assigned to three or four of the WFD classes with 95% confidence, due to random sampling effects, although the specific effect on WFD classification depends on the closeness of the range of measured concentrations to the class boundaries. Where water body status is estimated using parameters, such as water temperature, that are assessed using extreme percentiles in a distribution of measurements, such as the 98th percentile for water temperature as done in the UK, then monthly sampling does not capture the full variance observed and causes an inaccurate estimate of the true value ⁶¹.

Implications for environmental modelling

From a modelling perspective, the emerging evidence for excessive nutrient contribution of short-term events puts into question the ubiquitous applications of the data-driven models, such as the water quality balance model SPARROW in North America ^{62,63}. Underestimating nutrient export by a factor of two or three and missing the timing of greatest nutrient delivery into a waterbody

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

impedes efforts to delineate watershed "hot-spots" or time periods with increased likelihood of violations of water quality targets ⁶⁴. The advent of high resolution data offers a new perspective on process-based model parameterization and our capacity to accommodate threshold-type of behaviours when locating critical source areas of non-point source pollution ⁶⁵. In this regard, Wellen et al. ⁶⁶ presented a Bayesian hierarchical framework which postulated that the watershed response to precipitation occurs in distinct states, depending, for example, on precipitation and catchment storage. The proposed calibration framework enabled the identification of extreme states and the characterization of different watershed behaviours and improved model performance by allowing parameter values to vary between low and high flow conditions. In addition, estimates of instream assimilation and denitrification help to constrain catchment nitrogen delivery and transport models³², and sub-daily chemistry data coupled with weekly biological monitoring are providing the basis on which to develop a process-based description of aquatic biotic-abiotic interactions, thus enabling an enhanced understanding compared to using ecological indicators alone. Furthermore, development of intelligent water body-specific, cost efficient monitoring schemes combining modelling tools with high frequency monitoring would also help to optimize monitoring schemes and make these technologies accessible for large scale water management.

355

356

357

358

359

360

361

362

363

Limitations of current in situ technologies

In case of conventional *in situ* chemical and biological measurements there are major issues related to calibration (requiring stable reagents and standards) and supporting infrastructure (e.g., of pumped flow systems) and frequency of servicing intervals which in turn affect the scalability of *in situ* deployment. *In situ* optical sensors, such as those for nitrate, require cleaning to remove biofilm. They can also suffer from interferences due to turbidity and from co-absorbing species like humic acids. However, performing multi-parameter sensing, such as monitoring turbidity and nitrate simultaneously, enables the robustness of the nitrate measurements to be assessed. Given the costs,

service requirements, the risks of theft and vandalism, and instrument power requirements, which has decreased recently, there is a need for a cost-benefit analysis to assess the utility of *in situ* sensors for widespread operational and regulatory monitoring. Furthermore, "big" data streams from *in situ* measurements pose a challenge to environmental scientists because traditional approaches to data quality assurance and quality control are no longer practical when confronted with the demands of real-time processing. Despite routine maintenance and calibration of sensors, there is a pressing need for the development of automated tools and standards for quality assurance and quality control of sensor data ⁶⁷.

Future directions

The use of high frequency sensors has moved beyond the realm of purely academic research⁶⁸ and these sensors are now employed by numerous national, state, and municipal level environmental authorities. In the United States there are over 500 stations with continuous DO sensors, and over 100 stations with continuous nitrate sensors (http://waterdata.usgs.gov/nwis). There are similar levels of deployment in other developed nations.

The knowledge gained from new sensor technology has and will continue to stimulate further advancement. Already microfluidic sensors for measuring nutrients based on colormetric techniques have advantages of small size and limited reagent and power requirements ^{16,17}, though further improvements of these devices are necessary to increase robustness and reduce maintenance during permanent deployment. There is still a clear need for further development of new types of sensors, particularly for chemical and organism-based measurements of freshwater ecosystems. Increasing the number of analytes to include redox sensitive elements, micro-nutrients and pesticides would be highly beneficial for more complete environmental assessment. The exciting

prospect of micro-scale inductively coupled plasma spectrometers would allow the measurement of a wide-array of elements in water⁶⁹.

The inferences drawn from the examples we present above are broadly applicable, as suggested by their geographic range and variation in temporal scales, and move beyond findings that can be obtained from single experiments. Real time sensor deployment for measuring water quality properties continuously from multi-parameter probes offers new prospects to develop sensor networks for whole river networks, watersheds, and lakes. High frequency measurements will expand from the water column to hot spots of biogeochemical transformation and ecological significance, such as the interfaces between aquatic and terrestrial sites (e.g., hyporheic and riparian zones, wetlands, and river-estuarine transition zones). This would significantly increase our understanding of the interaction between sources, uptake (e.g., primary production) and retention (e.g., denitrification) in whole river networks. The co-location of isotope and dissolved anions and cations measurements will also enable enhanced understanding of pollutant storage and transfer and integration of hydrological and water quality models through better characterisation of water and ion transit times⁷⁰.

Current use of real time sensors is still restricted to fundamental aquatic attributes such as DO, pH value, SRP and NO₃ ⁷¹, but the field of sensor development is rapidly advancing and we see great potential for developing observational data sets that can substantially improve our ability to understand and predict the causes and consequences of environmental changes of aquatic ecosystems. Furthermore, such high temporal resolution data streams can be complemented by additional data types like satellite products for a synoptic survey of water quality of wetlands, large rivers, and lakes to create new scope for validating ecosystem models across multiple scales ⁷². The utility of *in situ* sensor measurements has already begun to transform routine monitoring in the U.S., with federal (e.g., U.S. Geological Survey), and state agencies (e.g., St Johns River Water Management District in Florida) investing heavily in the structural and personnel capacity to deploy

and interpret high-resolution solute time series. In Germany, routine high resolution sensor deployments are not restricted to highly sensitive water bodies like drinking water reservoirs and the gain of scientific transport process understanding⁶⁵; German state water authorities (e.g., state environmental agency of Hesse and Baden Wurttemberg) increasingly use high frequency monitoring to quantify matter fluxes (especially at the outlet of large rivers) and for early warning systems for drinking water river bank infiltration facilities (e.g. at the Rhein river). There is also potential to use high frequency monitoring more widely to measure intermittent discharges from Combined Sewer Overflows.

Automated sensors that collect novel data, or even traditional data at novel time scales, can enable analyses that inspire new paradigms in aquatic ecology ⁴. The susceptibility of an ecosystem to changing drivers or random events depends on the characteristics of critical thresholds, such as in ecosystem metabolism⁷³ or in the physical drivers of change, such as flow, light and temperature. Fundamental progress in ecology requires better understanding of thresholds and the rate of anthropogenic induced change in aquatic ecosystems. Emerging technology such as FrrF, "lab-on-a-chip", and DNA technology for observing time series data at high temporal resolution will make a growing contribution to this field ⁷⁴. High frequency measurements gained by automated sensors will increase our opportunities to better determine the severity of extreme events in terms of water quality and freshwater ecological impacts, and identify the most important variables for assessing the links to environmental change ⁷⁵ at different spatial scales and for different aquatic ecosystem types⁴¹. Based on this, we will be in a stronger position to spot early warning signals of critical transitions of watershed biogeochemistry and aquatic ecosystems, and identify and evaluate management options to help mitigate adverse water quality and ecological impacts⁷⁶.

References

- 436 (1) Kirchner, J.W.; Feng, X.H.; Neal, C. et al. The fine structure of water-quality dynamics: the (high-
- frequency) wave of the future. *Hydrol. Process.* **2004**, 18:1353-1359.
- 438 (2) Porter, J.; Arzberger, P.; Braun, H. W.; Bryant, P.; Gage, S.; Hansen, T.; Hanson, P.; Lin, C. C.; Lin,
- 439 F. P.; Kratz, T.; Michener, W.; Shapiro, S.; Williams, T., Wireless sensor networks for ecology.
- 440 *Bioscience* **2005**, 55, (7), 561-572.
- 441 (3) Johnson, K. S.; Needoba, J. A.; Riser, S. C.; Showers, W. J., Chemical sensor networks for the
- 442 aguatic environment. *Chemical Reviews* **2007**, 107, (2), 623-640.
- 443 (4) Porter, J. H.; Nagy, E.; Kratz, T. K.; Hanson, P.; Collins, S. L.; Arzberger, P., New Eyes on the
- World: Advanced Sensors for Ecology. *Bioscience* 2009, 59, (5), 385-397.
- 445 (5) Godsey, S. E.; Aas, W.; Clair, T. A.; de Wit, H. A.; Fernandez, I. J.; Kahl, J. S.; Malcolm, I. A.; Neal,
- 446 C.; Neal, M.; Nelson, S. J.; Norton, S. A.; Palucis, M. C.; Skjelkvale, B. L.; Soulsby, C.; Tetzlaff, D.;
- 447 Kirchner, J. W., Generality of fractal 1/f scaling in catchment tracer time series, and its implications
- for catchment travel time distributions. *Hydrol. Process.* **2010**, 24, (12), 1660-1671.
- 449 (6) Cassidy, R.; Jordan, P., Limitations of instantaneous water quality sampling in surface-water
- 450 catchments: Comparison with near-continuous phosphorus time-series data. J. Hydrol. 2011, 405, (1-
- 451 2), 182-193.
- 452 (7) Wade, A. J.; Palmer-Felgate, E. J.; Halliday, S. J.; Skeffington, R. A.; Loewenthal, M.; Jarvie, H. P.;
- Bowes, M. J.; Greenway, G. M.; Haswell, S. J.; Bell, I. M.; Joly, E.; Fallatah, A.; Neal, C.; Williams, R. J.;
- 454 Gozzard, E.; Newman, J. R., Hydrochemical processes in lowland rivers: insights from in situ, high-
- 455 resolution monitoring. *Hydrol. Earth Syst. Sci.* **2012**, 16, (11), 4323-4342.
- 456 (8) Neal, C.; Reynolds, B.; Kirchner, J. W.; Rowland, P.; Norris, D.; Sleep, D.; Lawlor, A.; Woods, C.;
- 457 Thacker, S.; Guyatt, H.; Vincent, C.; Lehto, K.; Grant, S.; Williams, J.; Neal, M.; Wickham, H.; Harman,
- 458 S.; Armstrong, L., High-frequency precipitation and stream water quality time series from Plynlimon,
- 459 Wales: an openly accessible data resource spanning the periodic table. *Hydrol. Process.* **2013**, 27,
- 460 (17), 2531-2539.
- 461 (9) Kirchner, J. W.; Neal, C., Universal fractal scaling in stream chemistry and its implications for
- 462 solute transport and water quality trend detection. Proc. Natl. Acad. Sci. U. S. A. 2013, 110, (30),
- 463 12213-12218.
- 464 (10) Carrit, D. E.; Kanwisher, J. W., An electrode system for measuring dissolved oxygen. Analytical
- 465 Chemistry **1959**, 31, 5-9.
- 466 (11) Jannasch, H. W.; Johnson, K. S.; Sakamoto, C. M., Submersible, osmotically pumped analyzers
- 467 for continuous determination of nitrate in situ. Analytical Chemistry 1994, 66, (20), 3352-3361.
- 468 (12) Johnson, K. S.; Coletti, L. J., In situ ultraviolet spectrophotometry for high resolution and long-
- 469 term monitoring of nitrate, bromide and bisulfide in the ocean. Deep-Sea Research Part I-
- 470 Oceanographic Research Papers **2002**, 49, (7), 1291-1305.
- 471 (13) Pellerin, B. A.; Downing, B. D.; Kendall, C.; Dahlgren, R. A.; Kraus, T. E. C.; Saraceno, J.; Spencer,
- 472 R. G. M.; Bergamaschi, B. A., Assessing the sources and magnitude of diurnal nitrate variability in the
- San Joaquin River (California) with an in situ optical nitrate sensor and dual nitrate isotopes.
- 474 Freshwater Biol. **2009**, 54, (2), 376-387.
- 475 (14) Heffernan, J. B.; Cohen, M. J.; Frazer, T. K.; Thomas, R. G.; Rayfield, T. J.; Gulley, J.; Martin, J. B.;
- 476 Delfino, J. J.; Graham, W. D., Hydrologic and biotic influences on nitrate removal in a subtropical
- 477 spring-fed river. *Limnol. Oceanogr.* **2010**, 55, (1), 249-263.
- 478 (15) Heffernan, J. B.; Cohen, M. J., Direct and indirect coupling of primary production and diel nitrate
- dynamics in a subtropical spring-fed river. Limnol. Oceanogr. 2010, 55, (2), 677-688.

- 480 (16) Beaton, A. D.; Cardwell, C. L.; Thomas, R. S.; Sieben, V. J.; Legiret, F. E.; Waugh, E. M.; Statham,
- 481 P. J.; Mowlem, M. C.; Morgan, H., Lab-on-Chip Measurement of Nitrate and Nitrite for In Situ
- 482 Analysis of Natural Waters. *Environ. Sci. Technol.* **2012**, 46, (17), 9548-9556.
- 483 (17) Nightingale, A. M.; Beaton, A. D.; Mowlem, M. C., Trends in microfluidic systems for in situ
- chemical analysis of natural waters. Sensors and Actuators B-Chemical 2015, 221, 1398-1405.
- 485 (18) Cohen, M. J.; Kurz, M. J.; Heffernan, J. B.; Martin, J. B.; Douglass, R. L.; Foster, C. R.; Thomas, R.
- 486 G., Diel phosphorus variation and the stoichiometry of ecosystem metabolism in a large spring-fed
- 487 river. *Ecological Monographs* **2013**, 83, (2), 155-176.
- 488 (19) Lorenzen, C. J., A method for the continuous measurement of in vivo chlorophyll
- concentrations. *Deep-Sea Research Part I-Oceanographic Research Papers* **1966**, 13, 223-227.
- 490 (20) Pellerin, B. A.; Saraceno, J. F.; Shanley, J. B.; Sebestyen, S. D.; Aiken, G. R.; Wollheim, W. M.;
- 491 Bergamaschi, B. A., Taking the pulse of snowmelt: in situ sensors reveal seasonal, event and diurnal
- 492 patterns of nitrate and dissolved organic matter variability in an upland forest stream.
- 493 *Biogeochemistry* **2012**, 108, (1-3), 183-198.
- 494 (21) Robinson, J. P.; Roederer, M., Flow cytometry strikes gold. *Science* **2015**, 350, (6262), 739-740.
- 495 (22) Read, D. S.; Gweon, H. S.; Bowes, M. J.; Newbold, L. K.; Field, D.; Bailey, M. J.; Griffiths, R. I.,
- 496 Catchment-scale biogeography of riverine bacterioplankton. *Isme Journal* **2015**, 9, (2), 516-526.
- 497 (23) Alliance for Coastal Technologies Nutrient Sensor Challenge. http://www.act-us.info/nutrients-
- 498 challenge/ (07 February 2016),
- 499 (24) Harmel, R. D.; Cooper, R. J.; Slade, R. M.; Haney, R. L.; Arnold, J. G., Cumulative uncertainty in
- 500 measured streamflow and water quality data for small watersheds. Trans. ASABE 2006, 49, 689-701.
- 501 (25) Bowes, M. J.; Jarvie, H. P.; Halliday, S. J.; Skeffington, R. A.; Wade, A. J.; Loewenthal, M.;
- 502 Gozzard, E.; Newman, J. R.; Palmer-Felgate, E. J., Characterising phosphorus and nitrate inputs to a
- rural river using high-frequency concentration-flow relationships. Sci. Tot. Environ. 2015, 511, 608-
- 504 620.
- 505 (26) Halliday, S. J.; Skeffington, R. A.; Bowes, M. J.; Gozzard, E.; Newman, J. R.; Loewenthal, M.;
- 506 Palmer-Felgate, E. J.; Jarvie, H. P.; Wade, A. J., The Water Quality of the River Enborne, UK:
- 507 Observations from High-Frequency Monitoring in a Rural, Lowland River System. Water 2014, 6, (1),
- 508 150-180
- 509 (27) Mellander, P.-E.; Jordan, P.; Melland, A. R.; Murphy, P. N. C.; Wall, D. P.; Mechan, S.; Meehan,
- 510 R.; Kelly, C.; Shine, O.; Shortle, G., Quantification of Phosphorus Transport from a Karstic Agricultural
- 511 Watershed to Emerging Spring Water. Environ. Sci. & Technol. 2013, 47, (12), 6111-6119.
- 512 (28) Mellander, P. E.; Jordan, P.; Shore, M.; McDonald, N. T.; Wall, D. P.; Shortle, G.; Daly, K.,
- 513 Identifying contrasting influences and surface water signals for specific groundwater phosphorus
- 514 vulnerability. *Sci. Tot. Environ.* **2016**, 541, 292-302.
- 515 (29) Spencer, R. G. M.; Pellerin, B. A.; Bergamaschi, B. A.; Downing, B. D.; Kraus, T. E. C.; Smart, D. R.;
- 516 Dahgren, R. A.; Hernes, P. J., Diurnal variability in riverine dissolved organic matter composition
- determined by in situ optical measurement in the San Joaquin River (California, USA). *Hydrol*.
- 518 *Process.* **2007**, 21, (23), 3181-3189.
- 519 (30) Ensign, S. H.; Doyle, M. W., Nutrient spiraling in streams and river networks. J. Geophys. Res.-
- 520 Biogeosci. 2006, 111, (G4).
- 521 (31) Hall, R. O.; Baker, M. A.; Rosi-Marshall, E. J.; Tank, J. L.; Newbold, J. D., Solute-specific scaling of
- 522 inorganic nitrogen and phosphorus uptake in streams. Biogeosciences 2013, 10, (11), 7323-7331.

- 523 (32) Rode, M.; Halbedel S. (née Angelstein), Anis M.R., Borchardt D., Weitere M., Continuous In-
- 524 Stream Assimilatory Nitrate Uptake from High Frequency Sensor Measurements. Environ. Sci.
- 525 Technol., 50 (11), pp 5685–5694
- 526 (33) Hensley, R. T.; Cohen, M. J.; Korhnak, L. V., Inferring nitrogen removal in large rivers from high-
- resolution longitudinal profiling. Limnol. and Oceanogr. 2014, 59, (4), 1152-1170.
- 528 (34) Appling, A.P.; Heffernan, J.B., Nutrient limitation and physiology mediate the fine-scale
- 529 (de)coupling of biogeochemical cycles. American Naturalist 2014, 184: 384-406.
- 530 (35) Kurz, M. J.; de Montety, V.; Martin, J. B.; Cohen, M. J.; Foster, C. R., Controls on diel metal cycles
- 531 in a biologically productive carbonate-dominated river. Chemical Geology 2013, 358, 61-74.
- 532 (36) Doyle, M. W.; Ensign, S. H., Alternative Reference Frames in River System Science. *Bioscience*
- **2009**, 59, (6), 499-510.
- 534 (37) Vannote, R. L.; Minshall, G. W.; Cummins, K. W.; Sedell, J. R.; Cushing, C. E., River Continuum
- 535 Concept. Canadian Journal of Fisheries and Aquatic Sciences 1980, 37, (1), 130-137.
- 536 (38) Ward, J. V.; Standford, J. A., The serial discontinuity concept of lotic ecosystems. In: Dynamics of
- Lotic Ecosystems, Fontaine, T. D.; Bartell, S. M., Eds. Ann Arbor Science: Ann Arbor, 1983; pp 29-42.
- 538 (39) von Schiller, D.; Aristi, I.; Ponsati, L.; Arroita, M.; Acuna, V.; Elosegi, A.; Sabater, S., Regulation
- 539 causes nitrogen cycling discontinuities in Mediterranean rivers. Sci. Tot. Environ. 2016, 540, 168-177.
- 540 (40) Twiss, M.; MacLeod, I., Phytoplankton community assessment in eight Lake Ontario tributaries
- made using fluorimetric methods. Aquatic Ecosystem Health & Management 2008, 11, (4), 422-431.
- 542 (41) Read, D. S.; Bowes, M. J.; Newbold, L. K.; Whiteley, A. S., Weekly flow cytometric analysis of
- riverine phytoplankton to determine seasonal bloom dynamics. Environ. Sci.-Process. & Impacts
- **2014**, 16, (3), 594-603.
- 545 (42) Bowes, M. J.; Loewenthal, M.; Read, D. S.; Hutchins, M. G.; Prudhomme, C.; Armstrong, L. K.;
- Harman, S. A.; Wickham, H. D.; Gozzard, E.; and Carvalho, L., Identifying multiple stressor controls on
- 547 phytoplankton dynamics in the River Thames (UK) using high-frequency water quality data, Sci. Tot.
- 548 Environ. **2016**, 569–570, 1489-1499.
- 549 (43) Halliday, S. J.; Wade, A. J.; Skeffington, R. A.; Neal, C.; Reynolds, B.; Rowland, P.; Neal, M.;
- Norris, D., An analysis of long-term trends, seasonality and short-term dynamics in water quality
- data from Plynlimon, Wales. Sci. Tot. Environ. 2012, 434, 186-200.
- 552 (44) Aubert, A. H.; Kirchner, J. W.; Gascuel-Odoux, C.; Faucheux, M.; Gruau, G.; Merot, P., Fractal
- Water Quality Fluctuations Spanning the Periodic Table in an Intensively Farmed Watershed.
- 554 Environ. Sci. Technol. **2014**, 48, (2), 930-937.
- 555 (45) Godsey, S. E.; Aas, W.; Clair, T. A.; de Wit, H. A.; Fernandez, I. J.; Kahl, J. S.; Malcolm, I. A.;
- Neal, C.; Neal, M.; Nelson, S. J.; Norton, S. A.; Palucis, M. C.; Skjelkvåle, B. L.; Soulsby, C.; Tetzlaff, D.;
- 557 Kirchner, J. W., Generality of fractal 1/f scaling in catchment tracer time series, and its implications
- for catchment travel time distributions. Hydrol. Process. 2010, 24, 1660–1671.
- 559 (46) Gisiger, T., Scale invariance in biology: coincidence or footprint of a universal mechanism?
- 560 *Biological Reviews* **2001**, 76, (2), 161-209.
- 561 (47) Halliday, S. J.; Skeffington, R. A.; Wade, A. J.; Neal, C.; Reynolds, B.; Norris, D.; Kirchner, J. W.,
- 562 Upland streamwater nitrate dynamics across decadal to sub-daily timescales: a case study of
- 563 Plynlimon, Wales. *Biogeosciences* **2013**, 10, (12), 8013-8038.
- 564 (48) Dupas, R., Jomaa, S., Musolff, S., Borchardt, D., Rode, M., Disentangling the influence of
- 565 hydroclimatic patterns and agricultural management on river nitrate dynamics from sub-hourly to
- decadal time scales. Sci. Total Environ. 2016, doi:10.1016/j.scitotenv.2016.07.053

- 567 (49) Horsburgh, J. S.; Jones, A. S.; Stevens, D. K.; Tarboton, D. G.; Mesner, N. O., A sensor network for
- 568 high frequency estimation of water quality constituent fluxes using surrogates. Environ. Model.
- 569 *Software* **2010**, 25, (9), 1031-1044.
- 570 (50) Jones, A. S.; Stevens, D. K.; Horsburgh, J. S.; Mesner, N. O., Surrogate Measures for Providing
- 571 High Frequency Estimates of Total Suspended Solids and Total Phosphorus Concentrations. J. Am.
- 572 *Water Resour. Ass.* **2011**, 47, (2), 239-253.
- 573 (51) Kirchner, J. W.; Austin, C. M.; Myers, A.; Whyte, D. C., Quantifying Remediation Effectiveness
- 574 under Variable External Forcing Using Contaminant Rating Curves. Environ. Sci. Technol. 2011, 45,
- 575 (18), 7874-7881.
- 576 (52) Rugner, H.; Schwientek, M.; Beckingham, B.; Kuch, B.; Grathwohl, P., Turbidity as a proxy for
- 577 total suspended solids (TSS) and particle facilitated pollutant transport in catchments. Environ. Earth
- 578 *Sci.* **2013**, 69, (2), 373-380.
- 579 (53) Bergamaschi, B. A.; Fleck, J. A.; Downing, B. D.; Boss, E.; Pellerin, B. A.; Ganju, N. K.;
- 580 Schoellhamer, D. H.; Byington, A. A.; Heim, W. A.; Stephenson, M.; Fujii, R., Mercury Dynamics in a
- 581 San Francisco Estuary Tidal Wetland: Assessing Dynamics Using In Situ Measurements. Estuaries and
- 582 *Coasts* **2012**, 35, (4), 1036-1048.
- 583 (54) Rozemeijer, J. C.; Van der Velde, Y.; Van Geer, F. C.; De Rooij, G. H.; Torfs, P.; Broers, H. P.,
- Improving Load Estimates for NO3 and P in Surface Waters by Characterizing the Concentration
- 585 Response to Rainfall Events. *Environ. Sci. Technol.* **2010**, 44, (16), 6305-6312.
- 586 (55) Effler, S. W.; O'Donnell, S. M.; Prestigiacomo, A. R.; O'Donnell, D. M.; Matthews, D. A.; Owens,
- 587 E. M.; Effler, A. J. P., Tributary Plunging in an Urban Lake (Onondaga Lake): Drivers, Signatures, and
- 588 Implications. J. Am. Water Res. Ass. 2009, 45, (5), 1127-1141.
- 589 (56) Chung, S. W.; Hipsey, M. R.; Imberger, J., Modelling the propagation of turbid density inflows
- 590 into a stratified lake: Daecheong Reservoir, Korea. Environ. Model. Software 2009, 24, (12), 1467-
- 591 1482.
- 592 (57) Bertone, E.; Stewart, R. A.; Zhang, H.; O'Halloran, K., Intelligent data mining of vertical profiler
- 593 readings to predict manganese concentrations in water reservoirs. Journal of Water Supply Research
- 594 and Technology-Aqua 2014, 63, (7), 541-552.
- 595 (58) Powers, S. M.; Tank, J. L.; Robertson, D. M., Control of nitrogen and phosphorus transport by
- reservoirs in agricultural landscapes. *Biogeochemistry* **2015**, 124, (1-3), 417-439.
- 597 (59) Rinke, K.; Kuehn, B.; Bocaniov, S.; Wendt-Potthoff, K.; Buettner, O.; Tittel, J.; Schultze, M.;
- 598 Herzsprung, P.; Roenicke, H.; Rink, K.; Rinke, K.; Dietze, M.; Matthes, M.; Paul, L.; Friese, K.,
- 599 Reservoirs as sentinels of catchments: the Rappbode Reservoir Observatory (Harz Mountains,
- 600 Germany). Environ. Earth Sci. 2013, 69, (2), 523-536.
- 601 (60) Tsai, J.-W.; Kratz, T. K.; Hanson, P. C.; Wu, J.-T.; Chang, W. Y. B.; Arzberger, P. W.; Lin, B.-S.;
- 602 Lin, F.-P.; Chou, H.-M. & Chiu, C.-Y., Seasonal dynamics, typhoons and the regulation of lake
- 603 metabolism in a subtropical humic lake. Freshwater Biol., 2008, 53, 1929-1941.
- 604 (61) Skeffington, R. A.; Halliday, S. J.; Wade, A. J.; Bowes, M. J.; Loewenthal, M., Using high-
- 605 frequency water quality data to assess sampling strategies for the EU Water Framework Directive.
- 606 *Hydrol. Earth Syst. Sci.* **2015**, 19, (5), 2491-2504.
- 607 (62) Pellerin, B. A.; Bergamaschi, B. A.; Gilliom, R. J.; Crawford, C. G.; Saraceno, J.; Frederick, C. P.;
- 608 Downing, B. D.; Murphy, J. C., Mississippi River Nitrate Loads from High Frequency Sensor
- 609 Measurements and Regression-Based Load Estimation. Environ. Sci. Technol. 2014, 48, (21), 12612-
- 610 12619.

- 611 (63) Alexander, R. B.; Smith, R. A.; Schwarz, G. E.; Boyer, E. W.; Nolan, J. V.; Brakebill, J. W.,
- 612 Differences in phosphorus and nitrogen delivery to the gulf of Mexico from the Mississippi river
- 613 basin. Environ. Sci. Technol. **2008**, 42, (3), 822-830.
- 614 (64) Wellen, C.; Arhonditsis, G. B.; Labencki, T.; Boyd, D., Application of the SPARROW model in
- watersheds with limited information: a Bayesian assessment of the model uncertainty and the value
- of additional monitoring. *Hydrol. Processes* **2014**, 28, (3), 1260-1283.
- 617 (65) Hesser, F. B.; Franko, U.; Rode, M., Spatially Distributed Lateral Nitrate Transport at the
- 618 Catchment Scale. Journal of Environmental Quality **2010**, 39, (1), 193-203.
- 619 (66) Wellen, C.; Arhonditsis, G. B.; Long, T.; Boyd, D., Quantifying the uncertainty of nonpoint source
- 620 attribution in distributed water quality models: A Bayesian assessment of SWAT's sediment export
- 621 predictions. J. Hydrol. **2014**, 519, 3353-3368.
- 622 (67) Campbell, J. L.; Rustad, L. E.; Porter, J. H.; Taylor, J. R.; Dereszynski, E. W.; Shanley, J. B.; Gries,
- 623 C.; Henshaw, D. L.; Martin, M. E.; Sheldon, W. M.; Boose, E. R., Quantity is Nothing without Quality:
- 624 Automated QA/QC for Streaming Environmental Sensor Data. Bioscience 2013, 63, (7), 574-585.
- 625 (68) von der Geest KB, Hyvonen J, Laurila T. Real-time determination of metal concentrations in
- 626 liquid flows using microplasma emission spectroscopy. IEEE Photonics Global Conference (PGC),
- 627 Singapore, DEC 13-16, **2012.**
- 628 (69) Hrachowitz M, Benettin P, van Breukelen BM, Fovet O, Howden NJK, Ruiz L, van der Velde Y,
- Wade AJ. Transit times the link between hydrology and water quality at the catchment scale.
- 630 WIREs Water. 2016, Doi: 10.1002/wat2.1155.
- 631 (70) Zacharias, S.; Bogena, H.; Samaniego, L.; Mauder, M.; Fuss, R.; Putz, T.; Frenzel, M.; Schwank,
- M.; Baessler, C.; Butterbach-Bahl, K.; Bens, O.; Borg, E.; Brauer, A.; Dietrich, P.; Hajnsek, I.; Helle, G.;
- 633 Kiese, R.; Kunstmann, H.; Klotz, S.; Munch, J. C.; Papen, H.; Priesack, E.; Schmid, H. P.; Steinbrecher,
- 634 R.; Rosenbaum, U.; Teutsch, G.; Vereecken, H., A Network of Terrestrial Environmental
- 635 Observatories in Germany. *Vadose Zone J.* **2011**, 10, (3), 955-973.
- 636 (71) Carpenter, S. R.; Cole, J. J.; Pace, M. L.; Batt, R.; Brock, W. A.; Cline, T.; Coloso, J.; Hodgson, J. R.;
- 637 Kitchell, J. F.; Seekell, D. A.; Smith, L.; Weidel, B., Early Warnings of Regime Shifts: A Whole-
- 638 Ecosystem Experiment. *Science* **2011**, 332, (6033), 1079-1082.
- 639 (72) Hipsey, M. R.; Hamilton, D. P.; Hanson, P. C.; Carey, C. C.; Coletti, J. Z.; Read, J. S.; Ibelings, B. W.;
- 640 Valesini, F. J.; Brookes, J. D., Predicting the resilience and recovery of aquatic systems: A framework
- for model evolution within environmental observatories. Water Resour. Res. 2015, 51, (9), 7023-
- 642 7043.
- 643 (73) Scheffer, M.; Bascompte, J.; Brock, W. A.; Brovkin, V.; Carpenter, S. R.; Dakos, V.; Held, H.; van
- Nes, E. H.; Rietkerk, M.; Sugihara, G., Early-warning signals for critical transitions. *Nature* **2009**, 461,
- 645 (7260), 53-59.
- 646 (74) Batt, R. D.; Carpenter, S. R.; Cole, J. J.; Pace, M. L.; Johnson, R. A., Changes in ecosystem
- 647 resilience detected in automated measures of ecosystem metabolism during a whole-lake
- 648 manipulation. Proc. Natl. Acad. Sci. U. S. A. **2013**, 110, (43), 17398-17403.
- 649 (75) Michalak AM. 2016. Study role of climate change in extreme threats to water quality. *Nature*
- 650 535, 349-350.
- 651 (76) Halliday, S. J., Skeffington, R. A., Wade, A. J., Bowes, M. J., Reed, D. S., Jarvie, H. P. and
- 652 Loewenthal, M. Riparian shading controls instream spring phytoplankton and benthic diatom
- 653 growth. Environmental Science: Process and Impacts 2016, 18 (6), pp. 677-689. ISSN 2050-7895 doi:
- 654 10.1039/C6EM00179C

Biography

656

657 Dr. Michael Rode is Deputy Head of the Department of Aquatic Ecosystem Analysis at Helmholtz 658 Centre for Environmental Research-UFZ in Magdeburg; he leads a research team on new water 659 quality monitoring techniques and modelling of matter fluxes at watershed scale. Andrew J Wade is 660 a Professor of Hydrology in the Department of Geography and Environmental Science at the 661 University of Reading. His research focuses on catchment hydrology, water quality and the links to 662 ecology. Dr. Cohen is a professor in the School of Forest Resources and Conservation at the 663 University of Florida in Gainesville, Florida. He works on a variety of watershed and aquatic systems 664 research. Dr. Hensley is a post-doctoral associate in the Ecohydrology Lab at the University of 665 Florida. He is part of a research team using in-situ sensors to better understand riverine metabolism 666 and nutrient spiraling. Dr. Bowes leads the Water Quality Processes Group at the Centre for Ecology 667 & Hydrology. His research investigates the sources and fates of phosphorus and nitrogen in river 668 catchments, and how they impact on river ecology. James Kirchner is the Professor of the Physics of 669 Environmental Systems at ETH Zurich and the former Director of the Swiss Federal Research Institute 670 WSL; his group conducts research at the interfaces between hydrology, geomorphology, and 671 watershed geochemistry. Professor Arhonditsis is the Chair of the Department of Physical and 672 Environmental Sciences at the University of Toronto. He leads the Ecological Modelling Laboratory 673 that has developed integrated biogeochemical models to guide the policy making process in several 674 impaired systems in North America over the past decade. Phil Jordan is Professor of Catchment 675 Science, Ulster University, Northern Ireland. His research focuses on the dynamics and fate of 676 nutrients and sediment in catchments with an emphasis on the capture and analysis of high 677 resolution water quality data. Professor Brian Kronvang is Head of Section of Catchment Science and 678 Environmental Management at Department of Bioscience, Aarhus University, Dr. Sarah Halliday 679 works as a researcher at the Department of Geography and Environmental Science, University of 680 Reading, UK; her research focuses on the new insights into catchment biogeochemical processing 681 attainable from high-frequency hydrochemical data. Dr. Richard Skeffington is Professor of 682 Geography at the University of Reading, with research interests in water quality monitoring and 683 modelling and understanding biogeochemical processes. Dr. Joachim Rozemeijer coordinates the 684 Environmental Monitoring and Modeling research program at Deltares, The Netherlands; he is 685 scientific researcher and advisor in the Groundwater and Soil Quality department and holds a PhD in 686 Dynamics in Groundwater and Surface water Quality. Dr. Aubert was a post-doctoral fellow at Justus 687 Liebig University in Giessen working on high-frequency nitrate time series analysis; now she is a 688 fellow at Eawag (the Swiss federal institute of aquatic science and technology) in Dübendorf studying 689 how water quality can be considered as part of structured decision making. Karsten Rinke is heading 690 the Department of Lake Research at the Helmholtz Centre for Environmental Research-UFZ in 691 Magdeburg, Germany; he is a limnologist working on lake modelling and the application of online 692 water quality monitoring in lake and reservoir management. Dr. Seifeddine Jomaa is a research 693 scientist at the Helmholtz Centre for Environmental Research-UFZ in Magdeburg. He leads a 694 research effort on hydrological water quality monitoring and modelling at the watershed scale.