Serologic Survey for Leptospirae in European Brown Bears (*Ursus arctos*) in Croatia

Zvonko Modrić¹ and Djuro Huber,² ¹ Department of Microbiology and Infectious Diseases, Veterinary Faculty of the University of Zagreb, Heinzelova 55, 41000 Zagreb, Republic of Croatia; ² Department of Biology, Veterinary Faculty of the University of Zagreb, Heinzelova 55, 41000 Zagreb, Republic of Croatia

ABSTRACT: From 1981 to 1991, sera of 42 European brown bears (Ursus arctos) from three areas in Croatia were tested for antibodies against 12 Leptospira interrogans serovars: grippotyphosa, sejroe, australis, pomona, canicola, icterohaemorrhagiae, tarassovi, saxkoebing, ballum, bataviae, poi, and hardjo. Diagnostic levels of antibody were found in 17 (40%) of 42 sera. Evidence of exposure to at least one of the serovars was found in seven of 14 free-ranging bears from the Lika region, four of 12 free-ranging bears from the Gorski Kotar region, zero of six orphaned cubs from the Gorski Kotar region, and six of 10 captive bears from the Zagreb Zoo. Based on the antibody titers, we implicated the following serovars: australis in five bears, sejroe in two bears, canicola in one bear, and icterohaemorrhagiae in one bear. There was a strong correlation between serovars implicated by this survey and serovars previously isolated from small mammals in Croatia.

Key words: European brown bear, Ursus arctos, leptospirosis, Leptospira interrogans, serovars, serology, survey, Croatia.

There are few reports of antibodies against leptospirae in bears. Isolation of leptospirae or the evidence of the disease in bears have not been reported. Kleinschmidt (cited in Kathe and Mochmann, 1967) found antibodies against Leptospira interrogans serovar icterohaemorrhagiae in a captive circus bear in Germany. Antibodies against serovar pomona (1:400) and serovar autumnalis (1:200) were found in a 2-yr-old female American black bear (Ursus americanus) and against serovar pomona (1:1,600) and serovar icterohaemorrhagiae (1:400) in a female cub from Pennsylvania (USA) (Matula et al., 1980). Two of 196 American black bears from Idaho (USA) had low antibody titers to serovar grippotyphosa (Binninger et al., 1980). In Gorski Kotar in Croatia, Karlović et al. (1985) found antibodies in three of ten European brown bears (U. arctos). One

each was positive to serovar grippotyphosa, saxkoebing or sejroe. Karlović et al. (1990) reported on 73 brown bear sera from Croatia; nine of 44 from Gorski Kotar, two of eight from Lika, and one of seven from Hrvatsko primorje were positive.

Approximately 400 brown bears are believed to inhabit Croatia (Huber and Morić, 1989). They are part of the Dinara Mountains population which is the largest in south-western Europe and is the source for reintroductions to other countries. Thus the knowledge about the health status of Croatian bears is an important management factor. Our objectives were to determine the prevalence of antibodies against leptospirae among bears in Croatia, the serovars with which the bears come in contact, possible sources of infection for bears, and any evidence of disease in bears.

Blood samples were taken from 22 male and 20 female European brown bears. Twenty-six (18 males and eight females) were sampled during capture for radiotagging in Croatia. Fourteen were captured in Lika (44°55'N, 15°39'E) and 12 in Gorski Kotar (45°27'N, 14°38'E). Age was estimated by counting cementum layers around the root of the extracted rudimentary first premolar tooth (Stonenberg and Jonkel, 1966). The mean age of the freeliving bears was 5.0 years (SD = 2.9, range 1 to 12 years). An additional six bears (one male and five females) were 1 to 3-mo-old orphaned cubs, originating from Gorski Kotar. The remaining seven bears (three males and four females) were from the Zagreb Zoo, with a mean age of 7.2 yr (SD = 4.3, range 0.4–14 years). Blood samples were taken from the femoral vein or artery during chemical immobilization with ketamine 11mg/kg hydrochloride (Ketalar,

Bear iden- tifi-		Age (yr)	Mass (kg)	Date of													
titi- ca-	Sex				Leptospirae serovar ^h												
tion.					gri.	sej.	aus.	pom.	can.	ict.	tar.	sax.	bal.	bat.	poi.	har.	
LI	f°	1	39	Nov 1981	-	100	-	-	_	-	_	_	_	_	-	_	
L5	m	7	222	Sep 1984	-	-	100	-	-	-	-	-	-	-	100		
L8	m	1	20	Apr 1985	-	500	-	-	-	-	-	100	-		-	-	
L9	m	5	119	May 1986	-	-	100	-	-		-	-	-	-	-		
LII	m	5	86	May 1986	-	-	100	—	-	-	-	-	-	-	-	-	
L13	m	5	111	Apr 1988	-	100	100	-	-	-	-	100	-	-	100	100	
L14	m	3	59	Apr 1988	-	-	100	_	-		-	-	-	-	-	-	
G4	m	5	127	May 1989	-	100	100	-	-	-	-	-	-	-	-	-	
G8	m	4	120	Oct 1989	-	100	2,000	—	-	-	-	500	-	-	-	1,000	
G9	f	13	153	Oct 1989	-	-	100	_	-	-	-		-	-	-	-	
G11	f	2	43	Jun 1990	-	100	-	-	-	-	-	100	-	-	-	-	
Z2	f	3	47	Nov 1986	-	-	100	-	100	1,000	-	-	-	-	100	-	
Z5	f	4	120	Apr 1990	-	-	-	-	100	-	-	-	-	-	-	-	
Z10	f	8	103	Mar 1991	-		100	_	100	100	-	-	-	-			
Z11	f	14	138	May 1992	-	-	-	-		100	_		-	-	100		
Z13	m	8	100	May 1991	-		-	_	100	100	-	-	-	-	100	-	
Z14	f	7	97	Jun 1991	100	-	100	-	-	500	-	-	500	-		-	
Freq	ueno	ey of	havin	g the highest	titer	2	5		1	1							

TABLE 1. Reciprocal titers of *Leptospira interrogans* antibody in sera of European brown bears from Croatia and the frequency of highest titers. Only the 17 positive bears are listed.

· L = Lika; G = Gorski Kotar; Z = Zagreb Zoo.

¹ gri. = grippotyphosa, sej. = sejroe, aus. = australis, pom. = pomona, can. = canicola, ict. = icterohaemorrhagiae, tar. = tarassovi, sax. = saxkoebing, bal. = ballum, bat. = batavie, poi. = poi, har. = hardjo.

f = female; m = male.

Parke-Davis, Berlin, Germany) and 6 mg/kg xylazine hydrochloride (Rompun, Bayer, Leverkusen, Germany). After separation, serum was stored at -18 C.

The microscopic agglutination test (Gochenour et al., 1958) was performed for 12 L. interrogans serovars: grippotyphosa, se jroe, australis, pomona, canicola, icterohaemorrhagiae, tarassovi, saxkoebing, ballum, bataviae, poi, and hard jo. The initial serum dilution was 1:100; positive sera were further diluted to the end-point of 50% agglutination. Microagglutination titers of $\geq 1:100$ were considered as indicative of previous exposure. If sera were positive to two or more leptospirae serovars, the serovar with the highest antibody titer was interpreted to be the serovar that caused infection. However, occasionally cross-reactivity among serovars may result in a titer that may be the same or even higher (Modrić et al., 1985). For eight sera that had equal titers to more than one serovar, we did not ascribe one serovar as the source of infection. Chi-squared tests (Burington and May, 1958) were used for statistical comparisons; P < 0.05 was considered as a statistically significant difference.

We found antibodies against leptospirae in 17 (40%) of 42 European brown bear sera, with reciprocal titers ranging from 100 to 2,000 (Table 1). Antibodies against leptospirae were found in seven of 14 samples from Lika, in four of 12 from Gorski Kotar, and in six of 10 from the Zagreb Zoo. All six bear cubs found in Gorski Kotar were negative. Highest antibody titers were to serovar australis in five bears, se jroe in two, and canicola and icterohaemorrhagiae each in one serum. In eight sera we found the same antibody titer for two to five serovars (Table 1). No bears showed evidence of disease at the time of sampling.

The 40% antibody prevalence against leptospirae we found in Croatia is the

highest published for bears. Earlier Karlović et al. (1985, 1990) reported 15 (18%) of 83 bears positive. Their lower prevalence may be explained by their use of the paper strips with dried blood which is a less sensitive method. Comparison of our results with the earlier studies (Karlović et al., 1985, 1990) is also limited because their method did not permit ranking the frequency of titers.

The prevalence of antibodies in captive bears (six positive of 10) was higher than in free-living and free-born animals (11 positive of 32), but the difference was not statistically significant. There were no significant differences in antibody prevalence between the bears from Lika and Gorski Kotar, or between age and sex classes.

Antibody titers against serovar australis were highest in five bears (Table 1). In Croatia, serovar australis is a cause of leptospirosis in humans (Zaharija, 1955) and has been isolated from the kidney of a clinically healthy cat (Modrić, 1978). Borčić et al. (1982) isolated serovar australis from four species of small mammals (Apodemus agrarius, A. sylvaticus, A. flavicollis and Clethrionomys glareolus) in Croatia. Four other leptospirae serovars (grippotyphosa, pomona, sejroe, and bataviae) also were isolated from 15 small mammal species. Thus, small mammals may be potential sources of infection with serovar australis for other animals, including European brown bears.

Antibodies against serovar *sejroe* were highest in two bear sera. Borčić et al. (1982) also found evidence for infection of bears by small mammals.

Antibodies against serovar *icterohae-morrhagiae* in the Zagreb Zoo bear may have been due to infection from rats (*Rat-tus norvegicus*), the main reservoir of this leptospirae serovar in Croatia (Zaharija and Perić, 1969). Antibodies against serovar *canicola* may reflect contact with dogs, the only documented reservoir of serovar *canicola* in Croatia (Zaharija, 1954). Free-living rats and mice and stray dogs are

practically the only mammals which can get in contact with caged bears in the Zagreb Zoo.

Bears are opportunistic feeders that forage on and below ground. Mammalian protein comprises only a small proportion of the European brown bear's diet (<1%)diet volume and 4% frequency of occurrence in scats), but the share of small mammals in protein is 75% (Cicnjak et al., 1987). We speculate that the leptospirae infections in bears depend on distribution of this agent among small mammals. The high prevalence of antibodies against leptospirae in bears in Croatia are evidence for the circulation of certain leptospire serovars in their population. The absence of clinical cases in our sample does not exclude the possibility of subclinical or past infections.

We thank the officials of the Plitvice Lakes and Risnjak National Parks, and of the Zagreb Zoo for giving us an opportunity to take the brown bear blood samples. Field work was in part sponsored by the National Geographic Society. Draft of the manuscript was improved by revision of Dr. Elizabeth Williams.

LITERATURE CITED

- BINNINGER, C. E., J. J. BEECHAM, L. A. THOMAS, AND L. D. WINWARD. 1980. A serological survey for selected infectious diseases of black bears in Idaho. Journal of Wildlife Diseases 16: 423–430.
- BORČIĆ, B., H. KOVAĆIĆ, Z. ŚEBEK, B. ALERAJ, AND N. TVRTKOVIĆ. 1982. Small terrestrial mammals as reservoirs of leptospires in the Sava Valley (Croatia). Folia Parasitologica 29: 177–182.
- BURINGTON, R. S., AND D. C. MAY. 1958. Handbook of probability and statistics with tables. Handbook Publishers, Inc., Sandusky, Ohio, p. 175.
- CICNJAK, L., D. HUBER, H. U. ROTH, R. L. RUFF, AND Z. VINOVRSKI. 1987. Food habits of brown bears in Plitvice Lakes National Park, Yugoslavia. International Conference on Bear Research and Management 7: 221–226.
- GOCHENOUR, W. S., C. A. GLEISER, AND N. K. WARD. 1958. Laboratory diagnosis of leptospirosis. Annals of the New York Academy of Sciences 70: 421-426.
- HUBER, D., AND S. MORIĆ. 1989. Štete od mrkih medvjeda u Jugoslaviji. *In* Zbornik radova 3. simpozijuma "Savremeni pravci uzgoja divljači," S.

Valentinčič (ed.). Savez veterinara i veterinarskih tehničara Jugoslavije, Ljubljana, Slovenija, pp. 197–202.

KARLOVIĆ, M., H. KOVAČIĆ, A. FRKOVIĆ, AND M. LACKOVIĆ. 1985. Prisutnost protutijela leptospira u krvi medvjeda s područja Gorskog Kotara. Veterinarski arhiv 55: 255–258.

, ____, AND ____. 1990. Proučavanje prisutnosti protutijela leptospira u krvi medvjeda (Ursus arctos L.). Veterinarska stanica 21: 331– 334.

- KATHE, J., AND H. MOCHMANN. 1967. Leptospiren und Leptospirosen. Teil II. VEB Gustav-Fischer Verlag, Jena, Germany, p. 668.
- MATULA, G. J., J. S. LINDZEY, AND H. ROTHENBACH-ER. 1980. Sex, age, and seasonal differences in the blood profile of black bears captured in northeastern Pennsylvania. International Conference on Bear Research and Management 4: 49–56.
- MODRIĆ, Z. 1978. Prirodna i eksperimentalna leptospiroza u mačke. Veterinarski arhiv 48: 147– 156.

- —, M. HERCEG, Ž. ŽUPANČIĆ, S. BAMBIR, V. HAHN, AND P. RAMADAN. 1985. Leptospiroza pasa u Zagrebu i okolici uzrokovana serološkim tipom *icterohaemorrhagiae*. Veterinarski arhiv 55: 93–102.
- STONENBERG, R. P., AND C. J. JONKEL. 1966. Age determination of black bears by cementum layers. The Journal of Wildlife Management 30: 411-414.
- ZAHARIJA, I. 1954. Sojevi L. canicola izolirani iz pasa u Zagrebu (Hrvatska). Veterinarski arhiv 24: 13–14.
- ——. 1955. Leptospira australis A (Veselići), izolirana u Hrvatskoj. Vojnosanitetski pregled 12: 535–536.
- ——, AND M. PERIĆ. 1969. Istraživanje leptospiroze u životinja u Hrvatskoj.—VII Štakor (*E. norvegicus*) kao kliconoša u gradovima Zadru, Šibeniku, Splitu, Karlovcu, Sisku i Varaždinu. Veterinarski arhiv 39: 71–73.

Received for publication 19 November 1992.