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# Serology describes a profile of declining malaria transmission in Farafenni, The Gambia

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## Abstract

**Background:** Malaria morbidity and mortality has declined in recent years in a number of settings. The ability to describe changes in malaria transmission associated with these declines is important in terms of assessing the potential effects of control interventions, and for monitoring and evaluation purposes.

**Methods:** Data from five cross-sectional surveys conducted in Farafenni and surrounding villages on the north bank of River Gambia between 1988 and 2011 were compiled. Antibody responses to MSP-1<sub>19</sub> were measured in samples from all surveys, data were normalized and expressed as seroprevalence and seroconversion rates (SCR) using different mathematical models.

**Results:** Results showed declines in serological metrics with seroprevalence in children aged one to 5 years dropping from 19 % (95 % CI 15–23 %) in 1988 to 1 % (0–2 %) in 2011 ( $p$  value for trend in proportions < 0.001) and the SCR dropping from 0.069 year<sup>-1</sup> (0.059–0.080) to 0.022 year<sup>-1</sup> (0.017–0.028;  $p = 0.004$ ). The serological data were consistent with previously described drops in both parasite prevalence in children aged 1–5 years (62 %, 57–66 %, in 1988 to 2 %, 0–4 %, in 2011;  $p < 0.001$ ), and all-cause under five mortality rates (37 per 1000 person-years, 34–41, in 1990 to 17, 15–19, in 2006;  $p = 0.059$ ).

**Conclusions:** This analysis shows accurate reconstruction of historical malaria transmission patterns in the Farafenni area using anti-malarial antibody responses. Demonstrating congruence between serological measures, and conventional clinical and parasitological measures suggests broader utility for serology in monitoring and evaluation of malaria transmission.

**Keywords:** *Plasmodium falciparum*, Malaria, Transmission, Serology, MSP-1<sub>19</sub>

## Background

Monitoring changes in malaria transmission levels due to natural fluctuations or as a result of control programmes is a key component of disease surveillance. There are a number of metrics available to assess malaria transmission ranging from passively collected morbidity and mortality data at health facilities to active detection of parasites in the mosquito and human host from field studies. The utility of different measures depends on

a variety of factors including the capacity of the health system to collect and analyse routine clinical data, local ecology of vectors and transmission intensity itself [1, 2].

Serological measures based on human anti-malarial antibodies can be used to assess malaria transmission intensity. Antibodies are produced during an infection with the *Plasmodium* parasite and are boosted upon subsequent infections. The presence of *Plasmodium*-specific antibody levels reflects exposure and, since many antibody responses are longer lived than infections in humans, they represent cumulative exposure over time. Therefore, anti-malarial antibody responses represent a population's history of malaria transmission [3]. Antibody responses can be expressed as age-adjusted continuous

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or binary variables to generate a seroconversion rate (SCR) which is analogous to the force of infection [3]. The SCR reflects the rate at which individuals become seropositive and can be estimated using simple reversible catalytic [3] or superinfection models [4]. The SCR has been used to describe malaria transmission intensity [3, 5], as well as to measure a change in malaria transmission [5–8] and to assess the effects of malaria control measures on transmission [7]. Serological measures are useful additional metrics to measure malaria transmission intensity, alongside parasite prevalence (parasite rate, PR) in humans and the entomological inoculation rate (EIR), especially at low transmission levels where other metrics lose their ability to discriminate [1, 9].

There are relatively few examples of repeated surveys to examine the validity of serological estimates of transmission over time. Historically, malaria transmission has been well-described in The Gambia with significant declines in malaria indices over the last 25 years [10–13]. Due to a longstanding collaboration between the London School of Hygiene and Tropical Medicine (LSHTM) and the Medical Research Council (MRC) in The Gambia, seroprevalence and PR data, obtained from samples collected in the villages around Farafenni in The Gambia, were combined. In Farafenni, on the north bank of the River Gambia, malaria was hyperendemic in 1988 with a PR over 60 % in children under 15 years of age [14, 15]. However, overall dry season microscopy PRs declined from 29 % in 1988 [14] to 11 % in 2008 [16]. In order to assess the utility of serological measures in describing this decline of malaria transmission, seroprevalence data from the Farafenni area between 1988 and 2011 have been compared with PR and previously described mortality rates.

## Methods

### Study area

Farafenni is situated on the north bank of the River Gambia, approximately 3 km from the border with Senegal (Fig. 1). Malaria transmission in the area is highly seasonal with nearly all transmission occurring during and after the rainy season [14], characterized by intense rains from June to October, with transmission continuing until December [10, 17]. *Plasmodium falciparum* is responsible for nearly all malaria infections in Farafenni [14, 18].

### Data collection and laboratory methods

Original datasets from surveys in 1988, 1990, 2003, 2008, and 2011, conducted as part of a longstanding collaboration between the LSHTM and the MRC in The Gambia, were obtained (Table 1). The location of the participating villages in the Farafenni area during these studies is shown in Fig. 1. A full description of the study

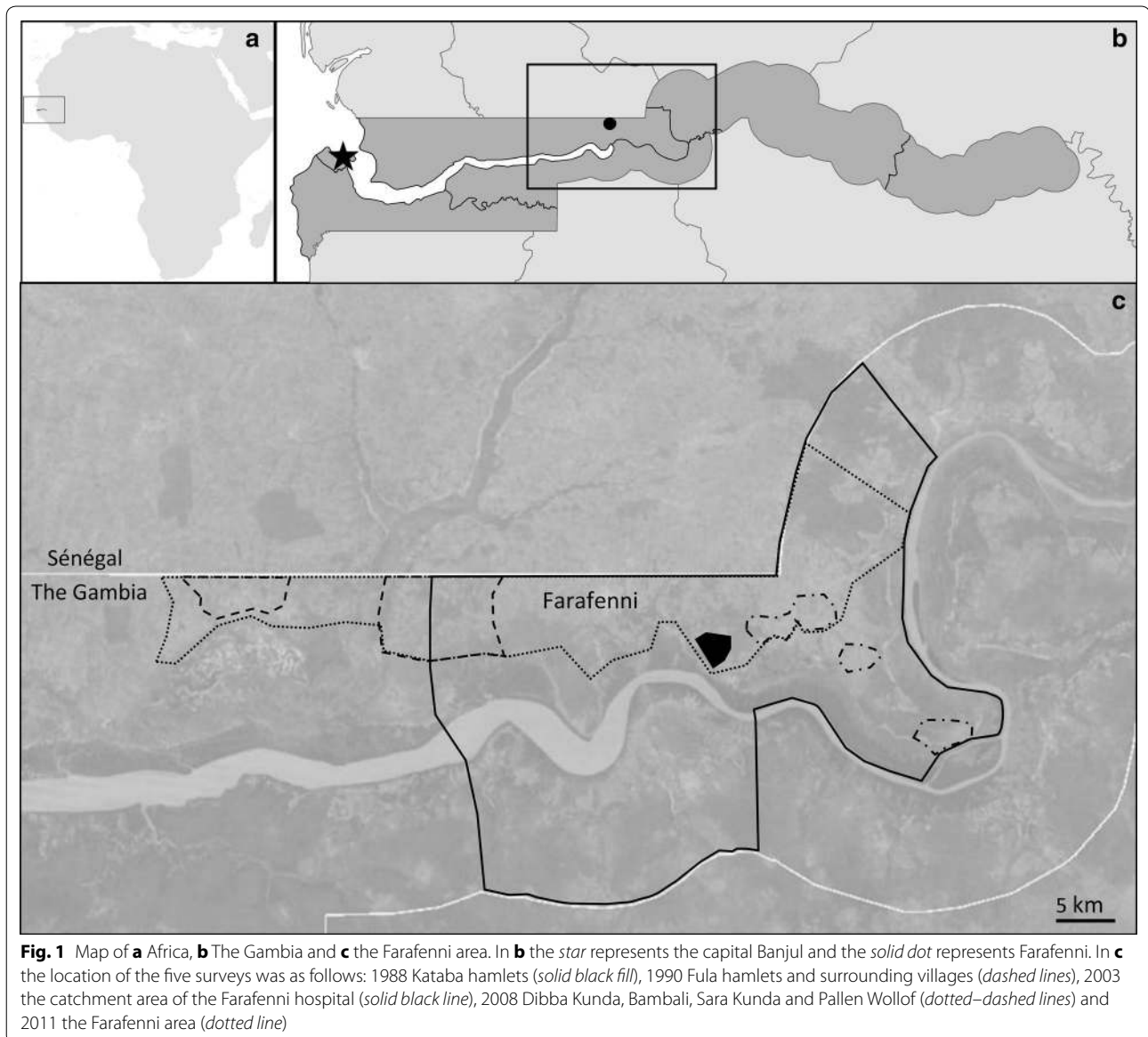
population, data collection and laboratory methods for each survey is detailed elsewhere [14, 16, 19–23].

The enzyme-linked immunosorbent assay (ELISA) protocol was the same in all surveys [24] and antibody responses to the 19 Kda fragment of *P. falciparum* merozoite surface protein 1 (MSP-1<sub>19</sub>) were recorded together with demographic data (e.g., age, sex) and parasite status. ELISAs were performed in The Gambia for all surveys, except 1990.

### Statistical analyses

All statistical analyses were conducted in Stata (version 13.1) and Prism (version 6.02). Infants under 1 year of age were excluded from each dataset to remove any influence of maternally derived antibodies [3]. A two-component Gaussian mixture model [25] was used to determine separate cut-off values for seropositivity for each dataset. SCRs (i.e., the rate at which the population becomes seropositive) and seroreversion rates (SRR; i.e., the rate at which the population reverts back to being seronegative) were calculated by fitting four different models. Firstly, a simple reversible catalytic model was fitted to age-adjusted seropositivity data using maximum likelihood [3]. Secondly, data were fitted to a superinfection model [4] which is comparable to the reversible catalytic model but allows for prolonged periods in the seropositivity state due to recurrent malaria exposure. More precisely, this model assumes that when an individual is re-infected while seropositive, the antibody response can be boosted and thus individuals can move between multiple seropositive states. Fixing and allowing SRR to vary was investigated for both of these models; the common SRR was estimated using maximum likelihood as described previously [3]. Furthermore, the presence of specific change points in transmission was investigated for each of the surveys separately as previously described [5, 7]. A likelihood ratio test was used to determine whether a model with a change point in transmission fit the data better than a model using one SCR. Finally, the change in the SCR was also investigated using an extension of the reversible catalytic model that allows multiple changes in the SCR, which was fitted to all surveys simultaneously. This novel approach can estimate the parameters using penalized maximum likelihood, by assuming that the log of the ratio of successive SCRs follows a normal distribution. This assumption results in smoothed estimates of the SCR over time, and so the model is referred to here as the 'smoothed model'. The reversible catalytic models are implemented as a Stata program called *revcat* (see Additional file 1) [26].

Chi-squared was used to test differences between proportions from two surveys while the Wilcoxon–Mann–Whitney test was applied to continuous outcomes to



compare the medians of two independent surveys. The trend in previously published all-cause under five-year-old mortality rates per 1000 person-years (U5MR) for the Farafenni area and the SCR estimates over time were calculated with linear regression. The Cochrane-Armitage test for trend in proportions was used to test the trend in the PR and seroprevalence. PR, U5MR, SRR, as well as SCR estimates, are followed by 95 % confidence intervals in brackets.

## Results

The number of individuals sampled in each survey ranged from 488 in 2011 to 742 in 1988, Table 1. The 1988, 1990

and 2011 surveys were performed during the peak transmission season (October/November), while the 2003 survey was conducted during the start of the wet season (July) and the 2008 survey during the early dry season (January/February). The median age and age range were comparable in the 1990–2008 surveys. However, the median age was lower in the 1988 survey and the age range was lower in the 2011 survey, which only included children up to the age of 15 years. Seroprevalence to MSP-1<sub>19</sub> in 1–5 year olds decreased from 19 % (15–23 %) in 1988 to 1 % (0–2 %) in 2011 ( $p < 0.001$ ), while the PR in 1–5 year olds declined from 62 % (57–66 %) in 1988 to 2 % (0–4 %) in 2011 ( $p < 0.001$ ). When comparing the

**Table 1 General characteristics of the surveyed study population between 1988 and 2011 in Farafenni, the Gambia**

% (95 % CI)	1988	1990	2003	2008	2011
Timing of survey	November	November	July <sup>a</sup>	January/february	October
Season	Peak transmission	Peak transmission	Start wet season	Dry season	Peak transmission
Median age in years (range)	5 (1–41)	14 (1–90)	11 (1–83)	13 (1–80)	10 (1–15)
Parasite prevalence <sup>b</sup>					
1–5 year olds	62 (57–66)	17 (11–23)	39 <sup>c</sup> (31–46)	16 (9–22)	2 (0–4)
1–15 year olds	60 (56–64)	22 (18–26)	40 <sup>d</sup> (33–46)	18 (14–22)	3 (1–4)
All ages	53 (49–57)	16 (13–19)	34 (30–38)	13 (11–16)	3 (1–4)
Number sampled for serology	742	736	624	670	488
Seroprevalence <sup>e</sup>					
1–5 year olds	19 (15–23)	19 (12–25)	12 (7–17)	13 (7–19)	1 (0–2)
1–15 year olds	23 (20–26)	31 (27–36)	23 (18–27)	20 (16–24)	13 (10–16)
All ages	28 (25–31)	46 (42–50)	35 (31–39)	34 (30–38)	13 (10–16)
Reference	[14, 19]	[20, 21]	[22]	[16]	[23]

For 2011 'all ages' only consists of children up to the age of 15 years

CI confidence interval

<sup>a</sup> Survey for parasite prevalence performed in November

<sup>b</sup> Parasite prevalence for *P. falciparum* as measured by polymerase chain reaction in the 2011 survey and microscopy of blood films in all other surveys

<sup>c</sup> For age 6 months–5 years

<sup>d</sup> For age 6 months–15 years

<sup>e</sup> Seroprevalence for MSP-1<sub>19</sub>

1988 and 1990 surveys, overall seroprevalence was lower (28 vs 46 %,  $p < 0.001$ ) and human parasite carriage was higher (53 vs 16 %,  $p < 0.001$ ) in 1988, even though both surveys were performed in the same season and were only 2 years apart. A plausible explanation for this could be the younger population in 1988 compared to 1990 (median age 5 vs 14 years,  $p < 0.001$ ).

The reversible catalytic and superinfection models led to similar results with an overall decrease in the SCR between 1988 and 2011 of 67–68 % when the SRR was fixed and 74–76 % when the SRR was allowed to vary, Table 2. Since both models provided similar results, the reversible catalytic model with a fixed SRR was used for further analysis. Predicted age seroprevalence curves for MSP-1<sub>19</sub> for each survey are shown in Fig. 2. The SCR declined from 0.069 year<sup>-1</sup> (0.059–0.080) in 1988 to 0.022 year<sup>-1</sup> (0.017–0.028) in 2011 ( $p = 0.004$ ), Fig. 3. The SCR values suggest a drop in the estimated annual EIR from ~5 to ~0.5 infectious bites per person per year (ib/p/y) using the previously described relationship between SCR and EIR [3, 27]. The previously published U5MR in the Farafenni area [28] showed an overall decline of 54 %; from 37 per 1000 person-years (34–41) in 1990 to 17 (15–19) in 2006 ( $p = 0.059$ ; Fig. 3).

When the SRR was allowed to vary across surveys it was between 1.7 and 1.9 times as high in the surveys in 1988 and 1990 in comparison with the more recent

surveys in 2003 and 2008 (Table 2). Since only individuals under 15 years old were included in 2011, the SRR for that year approximated to zero. The SRRs reflect the time for a seropositive individual to become seronegative, which ranged from ~14–26 years in the earlier surveys (superinfection model and reversible catalytic model, respectively) to ~27–46 years in the more recent surveys.

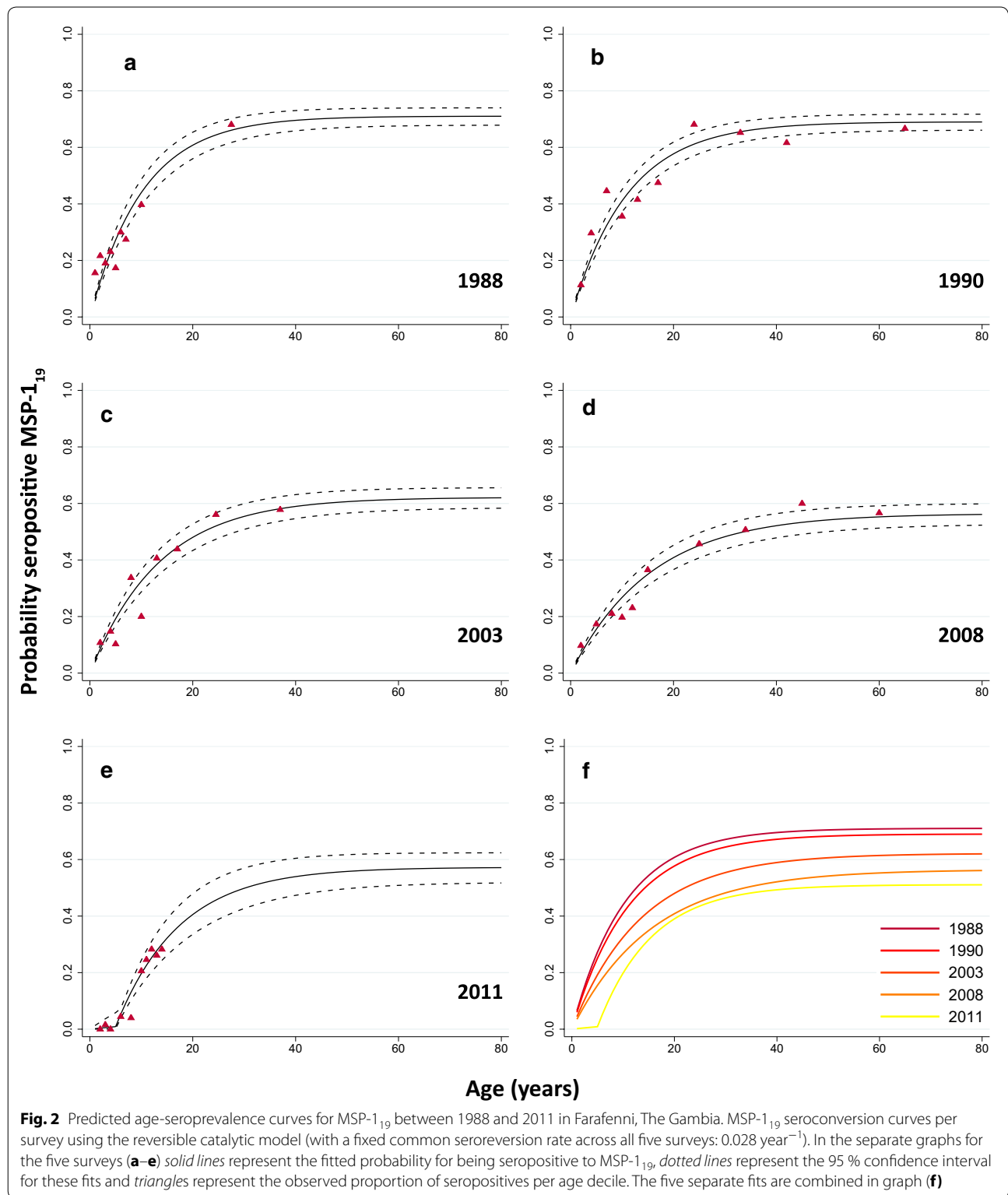
Analysis per survey—while fixing the SRR to the previously determined common SRR (0.028 year<sup>-1</sup>)—identified a significant change in transmission intensity only in the 2011 survey data and indicated that the change occurred in 2006 (5 years previously; range 4–8;  $p < 0.001$ ). A corresponding change point was not seen in the 2008 survey ( $p = 0.179$ ). SCR estimates for the 2011 surveys were as follows: 0.002 year<sup>-1</sup> (0.000–0.013) up to the age of 5 and 0.053 year<sup>-1</sup> (0.040–0.070) for those aged five and over (Fig. 2e). The SCR before the change point reflects most recent transmission and suggests an even more pronounced decline in the SCR of 97 % since 1988. However, since this survey only included children, it was decided to use the 'average' SCR for 2011 in the analyses described above.

The smoothed model estimating a separate SCR for each two-year period between 1988 and 2012 produced similar results to the fixed reversible catalytic model as shown in Fig. 3. The SCR declined from 0.067 year<sup>-1</sup> (0.059–0.077) in 1988 to 0.025 year<sup>-1</sup> (0.017–0.035) in 2012; reflecting an overall decrease of 63 %

**Table 2 Seroconversion and seroreversion rate estimates for MSP-1<sub>19</sub> between 1988 and 2011 in Farafenni, the Gambia**

Year	Reversible catalytic model						Superinfection model					
	Varied SRR (average SRR 0.024 year <sup>-1</sup> ; time to become seronegative 42 years)		Fixed SRR (common SRR 0.028 year <sup>-1</sup> ; time to become seronegative 36 years)		Varied SRR (average SRR 0.043 year <sup>-1</sup> ; time to become seronegative 23 years)		Fixed SRR (common SRR 0.049 year <sup>-1</sup> ; time to become seronegative 20 years)					
	SRR (95% CI)	SCR (95% CI)	% Decrease in SCR since previous survey	% Decrease in SCR since 1988 survey	SCR (95% CI)	% Decrease in SCR since previous survey	SCR (95% CI)	% Decrease in SCR since previous survey	SCR (95% CI)	% Decrease in SCR since previous survey	SCR (95% CI)	% Decrease in SCR since previous survey
1988	0.039 (0.017–0.090)	0.073 (0.059–0.091)	4	4	0.069 (0.059–0.080)	10	0.070 (0.035–0.137)	0	0.078 (0.062–0.099)	0	0.072 (0.062–0.083)	11
1990	0.037 (0.023–0.059)	0.070 (0.055–0.088)	4	10	0.062 (0.055–0.071)	10	0.071 (0.047–0.106)	0	0.078 (0.060–0.100)	0	0.064 (0.046–0.091)	11
2003	0.022 (0.011–0.045)	0.043 (0.034–0.055)	39	26	0.046 (0.040–0.053)	33	0.038 (0.020–0.073)	41	0.046 (0.035–0.060)	41	0.050 (0.036–0.072)	22
2008	0.022 (0.011–0.041)	0.033 (0.026–0.043)	23	22	0.036 (0.031–0.042)	48	0.036 (0.020–0.064)	22	0.036 (0.027–0.047)	54	0.041 (0.029–0.059)	18
2011	~0 (NA)	0.019 (0.015–0.024)	42	39	0.022 (0.017–0.028)	68	~0 (NA)	47	0.019 (0.015–0.024)	76	0.024 (0.016–0.037)	42

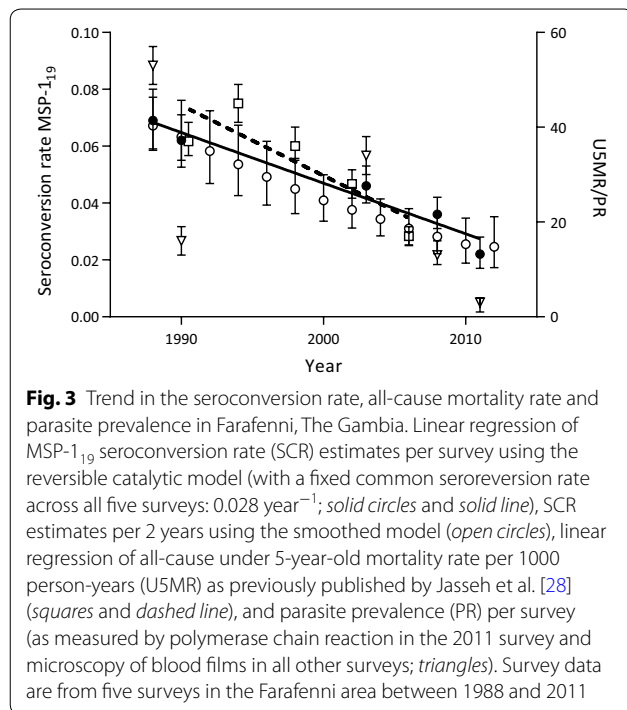
CI confidence interval; NA not applicable. Seroconversion rate (SCR) and seroreversion rate (SRR) estimates according to the reversible catalytic and superinfection model for five surveys between 1988 and 2011, while fixing the SRR and allowing the SCR to vary



(46–75 %). The common SRR in the smoothed model was 0.033 year<sup>-1</sup> (0.026–0.043), equating to an antibody decay time of 30 years.

### Discussion

This study describes the retrospective analysis of antibody responses to the MSP-1<sub>19</sub> antigen of *P. falciparum*



malaria in samples collected over a 23-year period in villages in the Farafenni area, The Gambia. Serological outcomes showed a clear relation to documented changes in malaria transmission with a decline in the seroprevalence and the SCR in line with a decline in parasite prevalence and all-cause mortality rates in children under 5 years old (U5MR).

The data describe very similar decreases over the time period in parasite prevalence (from 62 to 2–97 % decrease) and seroprevalence (from 19 to 1–95 % decrease) in children under 5 years old, while other studies from this area [23, 25] and elsewhere [29] have typically observed serological measures to be consistently higher than infection measures. Although similar prevalence of parasite carriage by PCR and seropositivity has been previously described in the Farafenni area [16], the reasons for the results in the current study are unclear. It may be due to the fact that responses to only one malaria antigen were assessed and, therefore, the data represent a minimal estimate of seroprevalence. All age seroprevalence predictably is higher than infection measures and operationally it may be logistically more attractive to focus on the whole population rather than specific age groups to generate these data. Further analysis is underway to examine parasite and serological metrics over a much wider geographical range.

Analysis of age-related seroprevalence data collected using both the reversible catalytic [3] and the

superinfection model [4] gave similar estimates for seroconversion irrespective of whether the SRR was fixed or allowed to vary across surveys [3]. Estimates from different models suggest a decline in seroconversion of approximately 70 % between 1988 and 2011, equating to an estimated ten-fold reduction in EIR to less than 1 ib/p/year. This is consistent with reports of declines in transmission both in The Gambia [12, 13], including Farafenni [10, 13], based on a variety of metrics (PR and U5MR), and similar declines between 1990 and 2012 reported by Trape et al. in nearby Dielmo, Senegal [30]. The reported decline in malaria transmission in Farafenni is likely related to scale-up of national intervention policies during this period (free malaria diagnosis in 1998, the distribution of free bed nets in 2000, intermittent preventive treatment in pregnant women in 2002, and the authorization of DDT for indoor residual spraying in 2007) [11]. Whilst caution should be observed with any extrapolation, data from the linear trend suggest a predicted current SCR for 2015 of 0.020 year<sup>-1</sup> (0.010–0.030) which correlates with an estimated EIR of ~0.4 ib/p/year.

Previous serological analyses of samples from Bioko Island, Equatorial Guinea [7] and southern Vanuatu [6] have shown a step-change in malaria transmission representing a distinct drop in exposure to infection after the introduction of malaria control methods. This is a unique facet of serological measures in that, when integrated with age, they allow examination of historical changes in exposure to infection. In the current analysis the absence of a step phenomenon in the smoothed model, and only in the 2011 survey when surveys were analysed separately, indicates that transmission reduction has been gradual rather than sudden. This may reflect the steady increase in intervention delivery, coverage and uptake as illustrated in the country profile in the WHO World Malaria Report 2014 [11].

The absence of a step-change (before the 2011 survey) may also be due to the sample sizes being too small to detect this phenomenon [31]. A recent serological analysis of a longitudinal sample set from Asembo Bay in Kenya similarly did not find a distinct drop in transmission associated with the widespread distribution of nets in the 1990s despite significant reductions in morbidity and mortality at the time. This was most likely due to a combination of limited sample size and a gradual decrease in transmission [32]. In the current analysis the change point in 2006 (detected in the 2011 survey) corresponds with previously reported minimal MSP-1<sub>19</sub> responses under the age of ten in 2009 by Ceesay et al. [13]. It suggests an even more pronounced decline in the SCR of 97 % since 1988. However, the 2011 data presented here included children only up to the age of

15 years and it would have been interesting to analyse the full serological profile across all ages to confirm this trend.

Interestingly, when the SRR was allowed to vary across surveys, it was nearly two-fold higher in the older surveys (1988 and 1990) compared to the more recent surveys (2003 and 2008), suggesting slightly paradoxically that the average decay time for antibodies increased while malaria transmission decreased. This increase in the half-life of anti-malarial antibodies over time might be explained by the fact that fewer children are infected in recent surveys and thus older individuals with more established antibody responses remain. It has previously been observed that anti-malarial antibody titres are relatively stable (consisting of long-lived plasma cells after repeated exposure) in adults but very variable (consisting of short-lived plasma cells) in children [33].

As previously described by Bosomprah *et al.* [4] the superinfection and reversible catalytic models gave very similar estimations of the SCR, yet the time for anti-malarial antibodies to decay was approximately 1.8 times faster in the superinfection model. This gave more realistic estimations of the time to decay (approximately 20 years) in comparison with the previously described minimum of 10 years [3]. Although both findings, the increased half-life of anti-malarial antibodies following declining transmission as well as the overall decreased half-life in the superinfection model, seem valid, further research is still needed to assess antibody decay rates in relation to age.

The reversible catalytic model was extended to a model that allowed fitting all surveys simultaneously and to allow multiple SCRs, with a smoothing parameter to prevent over-fitting. This model is particularly useful when the trend in the SCR, with a possible change point, is analysed from data collected from the same population over an extended period of time, and in the current study these estimates overlapped well with the reversible catalytic model with a fixed SRR. This will allow more robust analyses of data with multiple survey points with a predictive element for SCR, though this model would benefit from testing against other datasets from areas of different endemicity and transmission patterns. However, the data suggest that the parsimonious reversible catalytic model (with the exploration of a possible change point) is robust when compared to more involved models and is appropriate for studies such as a single cross-sectional survey. At high transmission levels, the superinfection model would provide additional information. Ideally, a population with a broad age range should be used in all of these models, since this increases the accuracy of seroconversion estimates [31].

There are a number of caveats to be taken into account in interpreting these data. Firstly the serological assays for these surveys were conducted at different times and by different groups. However, the same ELISA protocol was used including the same recombinant protein as target antigen. Secondly, the five surveys were conducted in the same broad geographical area but in different villages and there is likely to be heterogeneity of transmission between the villages. The small sample sizes do not allow village specific estimates and so only overall SCRs are presented to represent population level exposure on a larger scale in a similar manner to PR. Additionally, surveys were conducted at different times of the year and this may have influenced serological profiles. However, as described above, some anti-malarial antibody responses have estimated half-lives of at least 10 years [3] and are therefore likely to be stable over shorter periods of time. This suggests that the seasonal timing of the survey should not affect the measurement of exposure, as shown in 2008/2009 in The Gambia [34], in contrast to point prevalence estimates of PR data. The 2011 survey included children only up to the age of 15 years and this may result in an underestimate of the overall SCR. However, this should not influence the observed trend in declining transmission significantly. Whilst these surveys were not originally conceived for longitudinal analyses the use of the same ELISA protocol and target protein construct, together with the correlation between the decrease in serological profile and decreases in the PR and U5MR over nearly 25 years suggests that these data are representative of the underlying disease dynamics in the study area.

## Conclusion

The rationale for this work is that in areas with low malaria transmission conventional measures of malaria transmission, such as the PR and the EIR, may have little discriminative power unless sampling of humans and mosquitoes is intense [1, 9, 35]. Serological metrics have promise as adjunct measures of transmission [6, 7] but for wider use and better understanding of the implications of these metrics, studies comparing serological outcomes with conventional biological and health measures (disease incidence, mortality, etc.) are required. This report documents a decline in anti-malarial specific antibodies in Farafenni over a time period of 23 years. This decline mirrors the decrease in other malarial indices and of the U5MR in the Farafenni area [10, 12, 13] thereby confirming the suitability and validity of serology to monitor long-term changes in transmission. Anti-malarial antibody decay times in relation to age should be further explored, in order to obtain a more



## comprehensive insight into the immuno-epidemiology of (decreasing) malaria transmission.

### Additional file

**Additional file 1.** Reverse catalytic models with a constant rate of sero-conversion and multiple seroconversion rates.

### Authors' contributions

DC, PM, GT, UD, SC, SA, ER, and BG participated in the design and coordination of the separate surveys. CP, MA, JC, and PC carried out the ELISA assays. LH, JG, NS, and JC performed the statistical analyses. LH, CD, JG, JC, and ER drafted the manuscript and interpreted findings. All authors read and approved the final manuscript.

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### Competing interests

The authors declare that they have no competing interests.

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### References

- Tusting LS, Bousema T, Smith DL, Drakeley C. Measuring changes in *Plasmodium falciparum* transmission: precision, accuracy and costs of metrics. *Adv Parasitol*. 2014;84:151–208.
- Smith DL, Drakeley CJ, Chiyaka C, Hay SI. A quantitative analysis of transmission efficiency versus intensity for malaria. *Nat Commun*. 2010;1:108.
- Drakeley CJ, Corran PH, Coleman PG, Tongren JE, McDonald SLR, Carneiro I, et al. Estimating medium- and long-term trends in malaria transmission by using serological markers of malaria exposure. *Proc Natl Acad Sci USA*. 2005;102:5108–13.
- Bosomprah S. A mathematical model of seropositivity to malaria antigen, allowing seropositivity to be prolonged by exposure. *Malar J*. 2014;13:12.
- Stewart L, Gosling R, Griffin J, Gesase S, Campo J, Hashim R, et al. Rapid assessment of malaria transmission using age-specific sero-conversion rates. *PLoS One*. 2009;4:e6083.
- Cook J, Reid H, Iavro J, Kuwahata M, Taleo G, Clements A, et al. Using serological measures to monitor changes in malaria transmission in Vanuatu. *Malar J*. 2010;9:169.
- Cook J, Kleinschmidt I, Schwabe C, Nseng G, Bousema T, Corran PH, et al. Serological markers suggest heterogeneity of effectiveness of malaria control interventions on Bioko Island, equatorial Guinea. *PLoS One*. 2011;6:e25137.
- Cook J, Speybroeck N, Sochanta T, Somony H, Sokny M, Claes F, et al. Sero-epidemiological evaluation of changes in *Plasmodium falciparum* and *Plasmodium vivax* transmission patterns over the rainy season in Cambodia. *Malar J*. 2012;11:86.
- Bousema T, Okell L, Felger I, Drakeley C. Asymptomatic malaria infections: detectability, transmissibility and public health relevance. *Nat Rev Microbiol*. 2014;12:833–40.
- Ndungwa RP, Ramroth H, Müller O, Jasseh M, Sié A, Kouyaté B, et al. Comparison of all-cause and malaria-specific mortality from two West African countries with different malaria transmission patterns. *Malar J*. 2008;7:15.
- WHO. Gambia country profile. Geneva: World Health Organization; 2014.
- Ceesay SJ, Casals-Pascual C, Erskine J, Anya SE, Duah NO, Fulford AJC, et al. Changes in malaria indices between 1999 and 2007 in The Gambia: a retrospective analysis. *Lancet*. 2008;372:1545–54.
- Ceesay SJ, Casals-Pascual C, Nwakanma DC, Walther M, Gomez-Escobar N, Fulford AJC, et al. Continued decline of malaria in The Gambia with implications for elimination. *PLoS One*. 2010;5:e12242.
- Riley EM, Allen SJ, Bennett S, Thomas PJ, O'Donnell A, Lindsay SW, et al. Recognition of dominant T cell-stimulating epitopes from the circumsporozoite protein of *Plasmodium falciparum* and relationship to malaria morbidity in Gambian children. *Trans R Soc Trop Med Hyg*. 1990;84:648–57.
- Corran PH, O'Donnell RA, Todd J, Uthaipibull C, Holder AA, Crabb BS, et al. The fine specificity, but not the invasion inhibitory activity, of 19-kilodalton merozoite surface protein 1-specific antibodies is associated with resistance to malarial parasitemia in a cross-sectional survey in The Gambia. *Infect Immun*. 2004;72:6185–9.
- Satoguina J, Walther B, Drakeley C, Nwakanma D, Oriero EC, Correa S, et al. Comparison of surveillance methods applied to a situation of low malaria prevalence at rural sites in The Gambia and Guinea Bissau. *Malar J*. 2009;8:274.
- Ord R, Alexander N, Dunyo S, Hallett R, Jawara M, Targett G, et al. Seasonal carriage of pfprt and pfmdr1 alleles in Gambian *Plasmodium falciparum* imply reduced fitness of chloroquine-resistant parasites. *J Infect Dis*. 2007;196:1613–9.
- Kirby MJ, Ameh D, Green C, Jawara M, Milligan PJ, Bottomley C, et al. Efficacy of two different house screening interventions against exposure to malaria and anaemia in children in The Gambia: a randomized controlled trial. *Lancet*. 2009;374:998–1009.
- Riley EM, Jakobsen PH, Allen SJ, Wheeler JG, Bennett S, Jepsen S, et al. Immune response to soluble exoantigens of *Plasmodium falciparum* may contribute to both pathogenesis and protection in clinical malaria: evidence from a longitudinal, prospective study of semi-immune African children. *Eur J Immunol*. 1991;21:1019–25.
- Lulat A A study of humoral and cell-mediated immune responses to *Plasmodium falciparum* malaria gametocyte antigens in a rural Gambian population. PhD Thesis. London: London School of Hygiene and Tropical Medicine; 1993.
- Drakeley CJ, Akim NI, Sauerwein RW, Greenwood BM, Targett GA. Estimates of the infectious reservoir of *Plasmodium falciparum* malaria in The Gambia and in Tanzania. *Trans R Soc Trop Med Hyg*. 2000;94:472–6.
- Ismaili J The role of ethnicity and immune responses to *Plasmodium falciparum* Antigens (MSP1-19, AMA1 and CSP) on malaria risk in a seasonal transmission area in The Gambia. MSc Thesis. London: London School of Hygiene and Tropical Medicine; 2007.
- Ceesay SJ, Koivogui L, Nahum A, Taal MA, Okebe J, Affara M, et al. Malaria prevalence among young infants in different transmission settings. *Africa. Emerg Infect Dis*. 2015;21:1114–21.
- Corran PH, Cook J, Lynch C, Leendertse H, Manjurano A, Griffin J, et al. Dried blood spots as a source of anti-malarial antibodies for epidemiological studies. *Malar J*. 2008;7:195.
- Okebe J, Affara M, Correa S, Muhammad AK, Nwakanma D, Drakeley C, et al. School-Based countrywide seroprevalence survey reveals spatial heterogeneity in malaria transmission in the Gambia. *PLoS One*. 2014;9:e110926.
- Griffin J. REVCAT Stata module to fit reversible catalytic models. Boston: Boston College Department of Economics; 2015 [Statistical Software Components].
- Corran P, Coleman P, Riley E, Drakeley C. Serology: a robust indicator of malaria transmission intensity? *Trends Parasitol*. 2007;23:575–82.

28. Jasseh M, Webb EL, Jaffar S, Howie S, Townend J, Smith PG, et al. Reaching millennium development goal 4—The Gambia. *Trop Med Int Health*. 2011;16:1314–25.
29. Stresman GH, Stevenson JC, Ngwu N, Marube E, Owaga C, Drakeley C, et al. High Levels of asymptomatic and subpatent *Plasmodium falciparum* parasite carriage at health facilities in an area of heterogeneous malaria transmission intensity in the Kenyan Highlands. *Am J Trop Med Hyg*. 2014;91:1101–8.
30. Trape J-F, Tall A, Sokhna C, Ly AB, Diagne N, Ndiath O, et al. The rise and fall of malaria in a west African rural community, Dielmo, Senegal, from 1990 to 2012: a 22 year longitudinal study. *Lancet Infect Dis*. 2014;14:476–88.
31. Sepúlveda N, Drakeley C. Sample size determination for estimating antibody seroconversion rate under stable malaria transmission intensity. *Malar J*. 2015;14:141.
32. Wong J, Hamel MJ, Drakeley CJ, Kariuki S, Shi YP, Lal AA, et al. Serological markers for monitoring historical changes in malaria transmission intensity in a highly endemic region of Western Kenya, 1994–2009. *Malar J*. 2014;13:451.
33. White MT, Griffin JT, Akpogheneta O, Conway DJ, Koram KA, Riley EM, et al. Dynamics of the antibody response to *Plasmodium falciparum* infection in African children. *J Infect Dis*. 2014;210:1115–22.
34. Oduro AR, Conway DJ, Schellenberg D, Satoguina J, Greenwood BM, Bojang KA. Seroepidemiological and parasitological evaluation of the heterogeneity of malaria infection in the Gambia. *Malar J*. 2013;12:222.
35. Kilama M, Smith DL, Hutchinson R, Kigozi R, Yeka A, Lavoy G, et al. Estimating the annual entomological inoculation rate for *Plasmodium falciparum* transmitted by *Anopheles gambiae* s.l. using three sampling methods in three sites in Uganda. *Malar J*. 2014;13:111.

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