



# Serum calprotectin as a novel biomarker for severity of COVID-19 disease

Tezcan Kaya<sup>1</sup> · Selçuk Yaylacı<sup>1</sup> · Ahmet Nalbant<sup>1</sup> · İlhan Yıldırım<sup>2</sup> · Havva Kocayiğit<sup>3</sup> · Erdem Çokluk<sup>4</sup> · Mehmet Ramazan Şekeroğlu<sup>4</sup> · Mehmet Köroğlu<sup>5</sup> · Ertuğrul Güçlü<sup>6</sup>

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## Abstract

**Background** Some biomarkers have been reported to be related to the prognosis of the coronavirus disease 2019 (COVID-19). There are sparse data regarding the prognostic value of serum calprotectin in COVID-19 patients.

**Aims** This study aimed to investigate the relationship between serum calprotectin level and clinical severity of COVID-19 disease in hospitalized patients.

**Methods** This retrospective cross-sectional cohort study included 80 consecutive hospitalized patients with confirmed diagnosis of COVID-19. The study population was divided into two groups as patients hospitalized in the intensive care unit (ICU) and patients hospitalized but not in the ICU. The serum calprotectin levels, other laboratory, and clinical parameters were compared between groups.

**Results** The mean age of the patients was  $66.5 \pm 15.7$  years. Of the patients, 42 were in the ICU and 38 were not. Serum calprotectin level and acute-phase reactants such as C-reactive protein, procalcitonin, ferritin, fibrinogen, and white blood cell were significantly higher in ICU patients than in non-ICU patients. ROC curve analysis identified that serum calprotectin level was a predictor for ICU requirement with an area under the curve of 0.641 ( $p = 0.031$ ). Logistic regression analysis revealed that serum calprotectin was a significant determinant for whether or not patient required the ICU.

**Conclusions** These findings demonstrate that serum calprotectin level seems to be a useful biomarker that can predict the severity of COVID-19 disease. Serum calprotectin is a significant predictor of ICU requirement in patients with COVID-19.

**Keywords** Biomarkers · Calprotectin · Intensive care unit · Inflammation · SARS-CoV-2

## Introduction

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), classified as a novel coronavirus, causes coronavirus disease 2019 (COVID-19) in humans. This pandemic infection displays in a variety of ways, including asymptomatic, mild disease (such as fever, dry cough, weakness, muscle pain, diarrhea, nausea/vomiting, anosmia, and ageusia), and severe illness (such as pneumonia, acute respiratory distress syndrome (ARDS), disseminated intravascular coagulation, respiratory failure, shock, cytokine storm, multi-organ dysfunction, and death) [1, 2]. Some laboratory parameters have been reported to be related to the diagnosis, clinical course, and prognosis of the disease [3–5]. For example, lymphopenia, neutrophilia, anemia, thrombocytopenia, and an increase in neutrophil-to-lymphocyte ratio (NLR), C-reactive protein (CRP), procalcitonin, lactate dehydrogenase (LDH), prothrombin time (PT), and D-dimer have been reported to be significant [3–5].

✉ Tezcan Kaya  
tezcan@sakarya.edu.tr

<sup>1</sup> Department of Internal Medicine, Faculty of Medicine, Sakarya University, Sakarya, Turkey  
<sup>2</sup> Dr. Ergun Özdemir State Hospital, Giresun, Turkey  
<sup>3</sup> Department of Anesthesiology, Faculty of Medicine, Sakarya University, Sakarya, Turkey  
<sup>4</sup> Department of Biochemistry, Medical Faculty, Sakarya University, Sakarya, Turkey  
<sup>5</sup> Department of Microbiology, Faculty of Medicine, Sakarya University, Sakarya, Turkey  
<sup>6</sup> Department of Infectious Diseases, Faculty of Medicine, Sakarya University, Sakarya, Turkey

Calprotectin, a heterodimeric protein, is a member of the calcium-binding S100 protein family [6]. Calprotectin is located in the cytoplasm of neutrophils and monocytes, both of which play important roles in the inflammatory response in the human body [6]. It has been shown that serum calprotectin level increases secondary to inflammation in many inflammatory diseases [7–9]. It has also been reported that calprotectin can be used as a biomarker related to the activation, diagnosis, or prognosis of some diseases [7–12]. However, there are sparse data regarding the relationship between serum calprotectin and COVID-19 disease [13–16]. Therefore, this study aimed to investigate the relationship between serum calprotectin level and clinical severity of COVID-19 disease in hospitalized patients.

## Methods

### Design and patients

Consecutive hospitalized patients diagnosed with COVID-19 disease between November 1, 2020 and December 10, 2020 were included in this study, which was designed as a retrospective cross-sectional cohort study in a tertiary university hospital. COVID-19 diagnosis was made using the reverse transcription polymerase chain reaction test via nasopharyngeal and oropharyngeal swabs. Patients were excluded from this study if they were under the age of 18 years; pregnant; diagnosed with malignancy; had sepsis, septic shock, or proven bacterial infection; were using immunosuppressive drugs; or had chronic inflammatory disease or autoimmune disease.

Within the scope of this study, the demographic and clinical data of the patients were recorded from patient follow-up files. The study population was evaluated in two groups, as hospitalized in the intensive care unit (ICU) and hospitalized non-ICU. Criteria for ICU were respiratory failure, ARDS, or multiple organ failure. Demographic/clinical data, laboratory parameters, and serum calprotectin were compared between groups.

The study protocol was approved by the local University Ethics Committee (No. 71522473/050.01.04/471).

### Laboratory examinations

Venous blood samples were drawn from all included patients at the time of hospitalized in the ICU or non-ICU. EDTA tubes for complete blood count (CBC), tubes containing 3.8% Na-citrate for coagulation parameters, and anticoagulant-free biochemistry tubes were used for serum tests. CBC samples were studied immediately after the samples arrived at the biochemistry laboratory. Regarding the other tests, once the coagulation process of

the samples received in tubes without anticoagulants was completed, the venous blood samples were centrifuged at  $1500\times g$  for 10 min. The samples were closely monitored to determine whether any became hemolyzed or lipemic. Biochemistry parameters were studied from the serum. After the diagnosis of COVID-19 was made, sera obtained after centrifugation for the serum calprotectin study were stored at  $-80\text{ C}$  until further use. Prior to analysis for calprotectin, all samples were carefully brought to room temperature ( $15\text{--}18\text{ C}$ ) and homogenized before use.

Serum albumin, LDH, creatinine, calcium, alanine aminotransferase (ALT), aspartate aminotransferase (AST), and creatine kinase levels were measured by the spectrophotometric method, and CRP was measured by the immunoturbidimetric method via an Olympus AU5800 (Beckman Coulter, Inc. Brea, CA92821 USA) autoanalyzer. Ferritin level was measured with the chemiluminescence method in an ABBOTT ARCHITECT I 2000 SR device (Abbott Laboratories Abbott Park IL, 60064, USA); CBC parameters were measured using a CELL-DYN 3700 CD-3700SL device with laser measurement and LED Flow Cell method (Abbott Diagnostics Division, Abbott Laboratories Abbott Park IL, 60064, USA). PT and fibrinogen levels were measured by an optical method, D-dimer level was measured by a latex agglutination method in a Diagon CoagXL device (Budapest, Hungary), and procalcitonin level was measured by the immunoassay method using a Roche Cobas e 411 (Hitachi, 6544–01 Tokyo Japan). Serum calprotectin levels were measured by the sandwich ELISA method (Elabsience, Bioassay Technology Laboratory, Shanghai, China) (calprotectin ng/mL). In the precision study performed by the manufacturer, the within-run and between-run CV% of the kits were  $< 10\%$ .

### Statistical analysis

Data were analyzed using SPSS 20.0 for Windows software (SPSS Inc., Chicago, IL, USA). Descriptive analyses of the variables were expressed as mean  $\pm$  SD in normal distributions, and parameters with abnormal distribution were expressed as medians of the 25th–75th percentile. Categorical data were expressed as proportions. The Kolmogorov–Smirnov test was used to control the data distribution. The chi-square and the Student's *t* test were used for categorical and continuous variables, respectively. For continuous variables, differences between the two groups were evaluated using the Student's *t* test when data were normally distributed and the Mann–Whitney *U* test when the assumption of normality was not met. Receiver operating curve (ROC) analysis was used to calculate for serum

**Table 1** Demographic and clinical characteristics of inpatients with COVID-19

Parameters	Patients in ICU (n=38)	Patients in non-ICU (n=42)	P value
Age (years)	70.8 ± 12.8	62.5 ± 17.2	0.034
Age ≥ 65 years	29	20	0.008
Gender (male/female)	24/14	19/23	0.083
Diabetes mellitus	9	17	0.086
Hypertension	19	23	0.420
Coronary artery disease	13	7	0.060
Renal failure	5	3	0.301
Chronic respiratory disease	4	2	0.291

ICU intensive care unit

calprotectin levels the required cut-off values to distinguish ICU patients with maximum sensitivity and specificity. Binary logistic regression analysis was performed to determine predictive value of calprotectin for ICU patients. *P* values < 0.05 were considered statistically significant.

## Results

Of the 80 consecutive COVID-19 patients included in the study, 42 were followed in the ICU and 38 in the non-ICU. Forty-three of the patients were male, 37 were female, and the mean age was 66.5 ± 15.7 years. Comorbid chronic diseases and demographic characteristics of the study population are shown in Table 1. The average age of the ICU patients was significantly higher than those in the non-ICU, and most patients in the ICU were geriatric (age ≥ 65 years). Serum calprotectin level and acute-phase reactants such as CRP, procalcitonin, ferritin, fibrinogen, and white blood cells (WBCs) were significantly higher in ICU patients than in non-ICU patients ( $p=0.031$ ,  $p<0.001$ ,  $p<0.001$ ,  $p=0.009$ ,  $p=0.001$  respectively) (Table 2). Serum calprotectin levels of COVID-19 patients in the ICU and non-ICU are shown in Fig. 1 ( $p=0.031$ ). Mechanical ventilation requirement and mortality rate of 42 patients followed in ICU were 61.9% (26 patients) and 57.1% (24 patients) respectively.

An ROC curve was used to verify the optimum cut-off point for serum calprotectin level for predicting whether or not a patient required the ICU. The area under the ROC curve (AUC) was 0.641 (95% CI: 0.518–0.763,  $p=0.031$ ) for calprotectin (Fig. 2). The recommended cut-off value for calprotectin was 45.65 with a sensitivity of 50% and a specificity of 73.2%. Binary logistic regression analysis was performed to determine the ability of the serum calprotectin level to predict the patient required the ICU. The analysis revealed that serum calprotectin was a significant determinant for patients requiring the ICU (OR 1.029, 95% CI 1.004–1.005,  $p=0.024$ ).

**Table 2** Patients' serum calprotectin and other laboratory parameters

Variable	Patients in ICU (n=38)	Patients in non-ICU (n=42)	P value
White blood cells (K/ $\mu$ L)	8.4 (6–11.3)	5.1 (4.2–8)	0.001
Neutrophil (K/ $\mu$ L)	6.3 (4.4–9.2)	3.5 (2.3–4.9)	<0.001
Lymphocytes (K/ $\mu$ L)	0.7 (0.5–1.2)	1.3 (0.9–1.9)	<0.001
Hemoglobin (g/dL)	12.2 ± 1.7	12.2 ± 1.5	0.969
Platelets (K/ $\mu$ L)	187 (145–241)	190 (133–240)	0.278
D-Dimer ( $\mu$ gFEU/L)	1360 (789–2082)	516 (278–895)	<0.001
Fibrinogen (mg/dL)	405 (346–472)	320 (294–380)	0.009
Prothrombin time (s)	13.3 (12.3–14.5)	12 (11–13.7)	0.003
NLR	7 (4.9–11.3)	2.7(1.6–4)	<0.001
Calprotectin (ng/mL)	44.8 (34.5–61.5)	37.9 (30.3–46.2)	0.031
Ferritin ( $\mu$ g/L)	617 (416–1469)	268(132–623)	<0.001
Creatinine (mg/dL)	0.9 (0.8–1.1)	0.73 (0.6–0.97)	0.040
AST (U/L)	44 (28–56)	31 (22.7–41)	0.008
ALT (U/L)	26 (15–36.2)	22.5 (15–37.7)	0.828
Albumin (gr/dL)	2.9 ± 0.4	3.4 ± 0.4	<0.001
CRP (mg/L)	120 (73–178)	26.5 (7.8–90.2)	<0.001
Procalcitonin (ng/ml)	0.26 (0.09–1.03)	0.08 (0.04–0.20)	<0.001
LDH (U/L)	431 (367–536)	259 (195–329)	<0.001

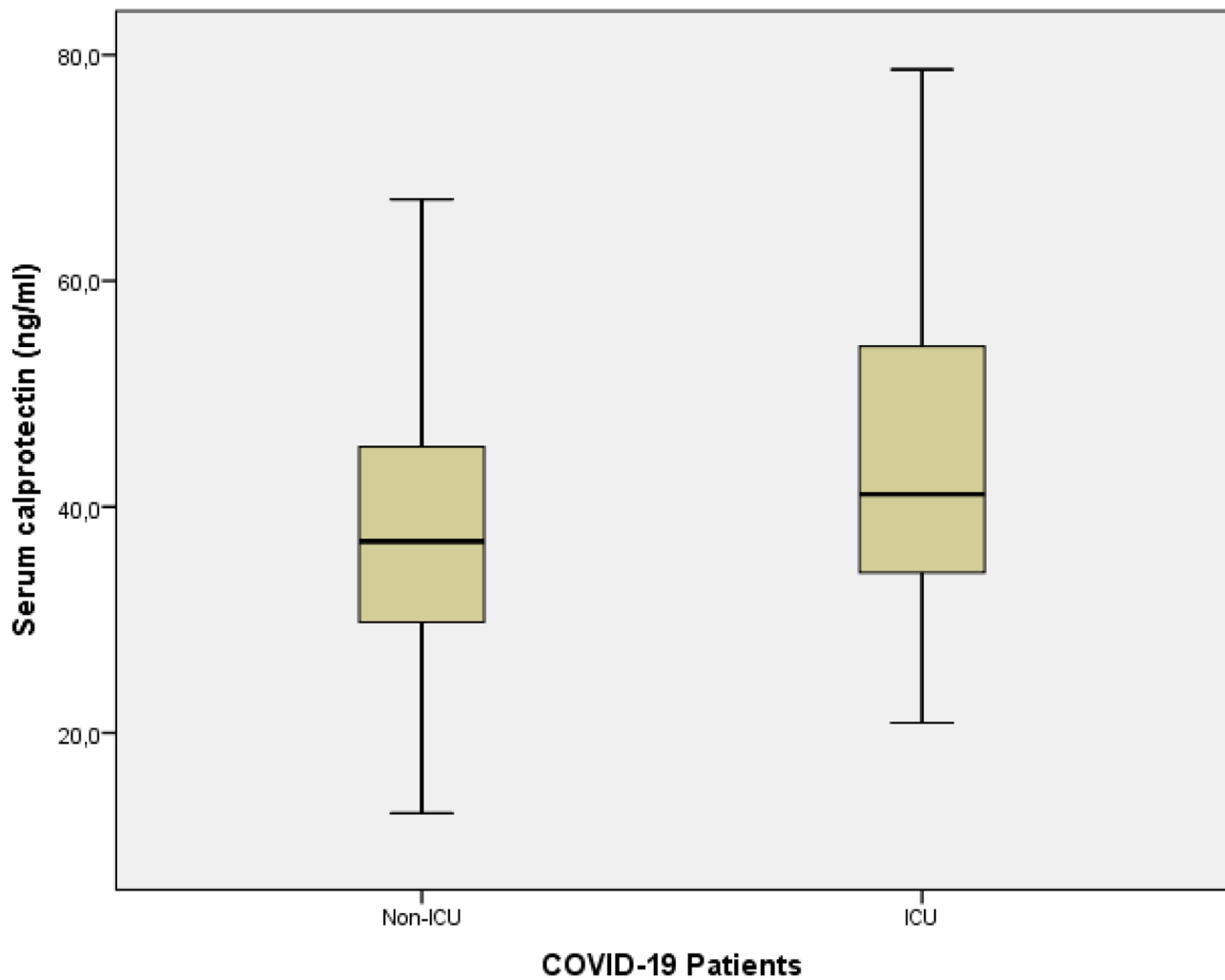
Data are presented as mean ± standard deviation for continuous variables with normal distribution, median and 25th and 75th percentiles (P25–P75) for variables with non-normal distribution

ICU intensive care unit, NLR neutrophil–lymphocyte ratio, AST aspartate aminotransferase, ALT alanine aminotransferase, CRP C-reactive protein, LDH lactate dehydrogenase

## Discussion

The current study revealed that serum calprotectin levels of COVID-19 patients in the ICU are significantly higher than those of COVID-19 patients in the non-ICU. In addition, the present study showed that serum calprotectin value for hospitalized COVID-19 patients is a significant predictor for requiring the ICU. Serum calprotectin level appears to be a useful biomarker as an indicator of COVID-19 disease severity. Calprotectin may be useful for distinguishing moderate or severe COVID-19 patients.

COVID-19 disease can cause several different clinical situations. Conditions such as pneumonia, ARDS,



**Fig. 1** Serum calprotectin levels of COVID-19 patients in intensive care unit (ICU) and non-ICU ( $p=0.031$ , assessed by Mann–Whitney  $U$  test)

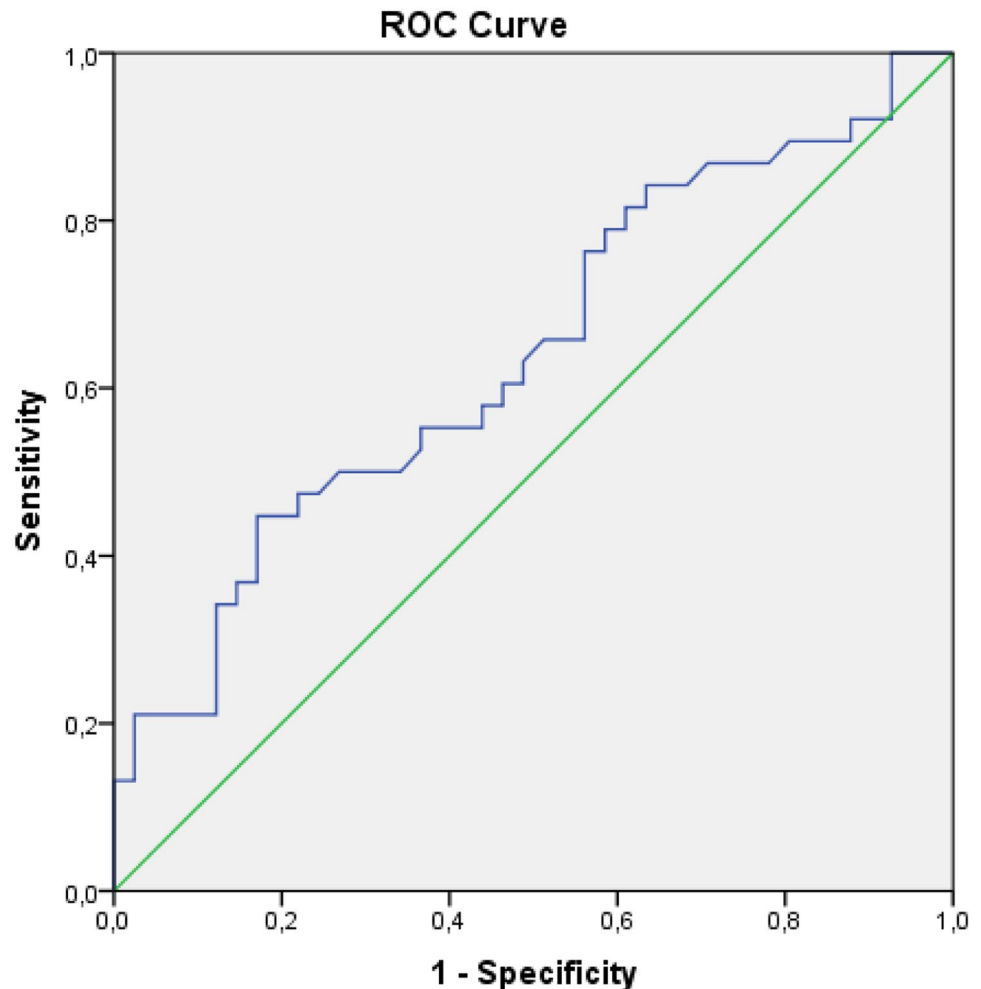
disseminated intravascular coagulation, respiratory failure, shock, cytokine storm, and multi-organ dysfunction can be seen in severe COVID-19 disease [1, 2, 17]. In these clinical situations, patients generally require follow-up and treatment in the ICU [17, 18]. Severe disease also increases morbidity and mortality rates [17, 18]. Studies have reported that some biomarkers can distinguish mild/moderate and severe disease in COVID-19 patients at an early stage [4–6, 19]. In a meta-analysis, Henry et al. reported that WBC count, lymphocyte count, platelet count, IL-6, and serum ferritin are markers for progression to critical illness [19].

In another systematic review and meta-analysis, Ghahramani et al. revealed that compared with a non-severe group, a group of patients with severe COVID-19 disease had significant decreases in lymphocytes, monocytes, eosinophils, hemoglobin, platelets, and albumin and significant increases in neutrophils, CRP, procalcitonin, LDH, fibrinogen, PT, AST, D-dimer, and NLR [5]. In the current study, WBC,

neutrophils, lymphocytes, D-dimer, fibrinogen, PT, NLR, ferritin, AST, albumin, CRP, procalcitonin, and LDH values were significantly different in ICU patients. The results of our study are consistent with these two prior meta-analyses [5, 19].

Calprotectin is a protein that is especially secreted by neutrophils secondary to inflammation. It has recently been reported that calprotectin can be used as a biomarker of inflammation for assessing the activity of some inflammatory disorders [7–12]. However, there are few published studies regarding the relationship between serum calprotectin and COVID-19 severity. In their study including 121 COVID-19 patients (40 ICU, 81 non-ICU), Chen et al. reported that ICU-admission COVID-19 patients had significantly elevated levels of serum calprotectin (S100A8/A9) compared with non-ICU patients, and that patients with fatal outcomes had significantly more calprotectin than those who lived [20]. In addition, according to that study, a significant elevation

**Fig. 2** Receiver-operating characteristic (ROC) analysis of calprotectin as indicator to predict COVID19 patients for intensive care unit (AUC=0.641; 95%CI, 0.518–0.763)



of serum calprotectin was associated with high mortality in COVID-19 patients [20].

In their study with 94 COVID-19 patients, Shi et al. reported that the serum calprotectin levels of patients requiring mechanical ventilation were higher than those who did not need intubation [14]. In another study of 66 COVID-19 patients, Romualdo et al. revealed that the serum calprotectin levels of patients who died were higher than of those who lived [16].

The present study has some limitations. First, there was relatively small number of patients, and the patients were from a single center. A multi-center study involving more patients and countries would be more beneficial. Second, in this study, serum calprotectin was measured only once at the time of admission to the inpatient service or ICU. Patient follow-up could be better with daily or every other day serum calprotectin measurement. Finally, serum calprotectin measurement is not routinely performed in most hospitals, and this assay may be relatively expensive compared with other acute phase reactants.

Similar to the results of the very recent studies reported above, in the present study, serum calprotectin was significantly higher in ICU patients with severe disease than in non-ICU patients with moderate disease. In addition, calprotectin served as a significant predictor for ICU requirement in COVID-19 patients. In conclusion, these findings suggest that serum calprotectin may be a novel biomarker in the assessment of poor outcome in COVID-19 patients.

**Author contribution** Conceptualization: TK, SY, AN. Methodology: TK, IY, HK. Formal analysis and investigation: EÇ, MRŞ, İY. Writing—original draft preparation: TK, SY, AN. Writing—review and editing: TK, MK, EG. Resources: İY, MK, EÇ. Supervision: MRŞ, MK, EG.

## Declarations

**Ethical approval** The study protocol was approved by the Ethics Committee of Sakarya University Faculty of Medicine, Sakarya, Turkey (No: 71522473 / 050.01.04 / 471).

**Conflict of interest** The authors declare that they have no conflict of interest.

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