



Review

Sex Differences in Inflammation and Muscle Wasting in Aging and Disease

Chiara Della Peruta ¹, Biliana Lozanoska-Ochser ^{1,2}, Alessandra Renzini ¹ , Viviana Moresi ³ ,
Carles Sanchez Riera ¹, Marina Bouché ^{1,*} and Dario Coletti ^{1,4}

- ¹ Unit of Histology and Medical Embryology, Department of Anatomy, Histology, Forensic Medicine and Orthopedics, Sapienza University of Rome, 00161 Roma, Italy
² Department of Medicine and Surgery, LUM University, 70010 Bari, Italy
³ Institute of Nanotechnology (Nanotec), National Research Council (CNR), c/o Sapienza University of Rome, 00185 Roma, Italy
⁴ Biological Adaptation and Ageing (B2A), Institut de Biologie Paris-Seine, Sorbonne Université, CNRS UMR 8256, Inserm U1164, 75005 Paris, France
* Correspondence: marina.bouche@uniroma1.it

Abstract: Only in recent years, thanks to a precision medicine-based approach, have treatments tailored to the sex of each patient emerged in clinical trials. In this regard, both striated muscle tissues present significant differences between the two sexes, which may have important consequences for diagnosis and therapy in aging and chronic illness. In fact, preservation of muscle mass in disease conditions correlates with survival; however, sex should be considered when protocols for the maintenance of muscle mass are designed. One obvious difference is that men have more muscle than women. Moreover, the two sexes differ in inflammation parameters, particularly in response to infection and disease. Therefore, unsurprisingly, men and women respond differently to therapies. In this review, we present an up-to-date overview on what is known about sex differences in skeletal muscle physiology and dysfunction, such as disuse atrophy, age-related sarcopenia, and cachexia. In addition, we summarize sex differences in inflammation which may underly the aforementioned conditions because pro-inflammatory cytokines deeply affect muscle homeostasis. The comparison of these three conditions and their sex-related bases is interesting because different forms of muscle atrophy share common mechanisms; for instance, those responsible for protein dismantling are similar although differing in terms of kinetics, severity, and regulatory mechanisms. In pre-clinical research, exploring sexual dimorphism in disease conditions could highlight new efficacious treatments or recommend implementation of an existing one. Any protective factors discovered in one sex could be exploited to achieve lower morbidity, reduce the severity of the disease, or avoid mortality in the opposite sex. Thus, the understanding of sex-dependent responses to different forms of muscle atrophy and inflammation is of pivotal importance to design innovative, tailored, and efficient interventions.

Keywords: sarcopenia; aging; bed rest; microgravity; cachexia; inflammation; sex differences



Citation: Della Peruta, C.; Lozanoska-Ochser, B.; Renzini, A.; Moresi, V.; Sanchez Riera, C.; Bouché, M.; Coletti, D. Sex Differences in Inflammation and Muscle Wasting in Aging and Disease. *Int. J. Mol. Sci.* **2023**, *24*, 4651. <https://doi.org/10.3390/ijms24054651>

Academic Editors: Vincenzo Sorrentino and Stefano Perni

Received: 5 February 2023

Revised: 21 February 2023

Accepted: 23 February 2023

Published: 28 February 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

In medicine and in clinical practice, sex differences comprise sex-specific and sex-related diseases, i.e., disease states exclusively or prevalently occurring in people of one sex. Obvious examples of sex-related diseases are genetic diseases linked to sexual chromosomes [1–3]. In addition, an impressive list of pathologies includes diseases that display different outcomes in the two sexes, ranging from depression and epilepsy [4,5] to autoimmune diseases [6], and also ranging from myopathies [7] to organ failure or dysfunction [8–10]. Many illnesses are characterized by sex-specific differences in severity [11], natural history [12], or disease mechanisms [13].

Only in recent years, thanks to a precision medicine approach, have treatments tailored to the sex of each patient emerged in clinical trials [14]. As an example, the treatment with the common immunosuppressant rapamycin in mice has sex-specific effects, such as extending the life-span in female mice more than in male mice, whereas the combination with the anti-hyperglycemic drug metformin levels these differences [15]. Indeed, finding sex differences in responses to disease or treatment may lead to implemented or totally new treatments [16–19]. In this review, we focus on sex differences in skeletal muscle. Indeed, significant differences between the two sexes concern both sexes' striated muscle tissues with important consequences for diagnosis and therapy [20,21]. However, the preference given to the musculature, which prominently characterizes sexual dimorphism, is based on the fact that the amount of lean mass is directly associated with survival in both healthy and disease conditions [22].

In this review we analyze the most significant papers reporting on sex differences in skeletal muscle physiological conditions as well as in three different pathological states characterized by marked sarcopenia and muscle dysfunction: disuse atrophy due to immobilization or microgravity [23], age-related sarcopenia [24], and muscle wasting in cachexia [25,26]. We also discuss sex differences in inflammation which may underly the conditions above; indeed, pro-inflammatory cytokines deeply affect muscle homeostasis. The rationale of comparing these three conditions is based on the fact that different forms of muscle atrophy share common mechanisms—for instance, those responsible for protein dismantling [27]—although differing in terms of kinetics, severity, and regulatory mechanisms [28,29]. Whether these differences can arise differently on a sex-related basis is of particular interest for a precision medicine-based approach.

2. Sex Differences in Muscle Homeostasis and Metabolism

Men have a remarkably different muscle phenotype compared to females, besides having greater muscle mass tout court. The major differences between the two sexes in muscle metabolism and homeostasis were extensively reviewed by Rosa-Caldwell and Greene [30]. In both rodents and humans, sex differences are observed in muscle fiber type, capillarity, and transcriptomes [31]. Indeed, glycolytic fibers are more abundant in men than in women [32], which has a direct consequence on the glucose metabolism [33] and respiratory capacity [34] of the musculature. This difference could account for the differential sensitivity to the diverse forms of muscle atrophy among sexes. Indeed, the fact that cachexia affects glycolytic fibers to a greater extent than oxidative ones [35], whereas disuse muscle atrophy affects predominantly oxidative fibers [36], is consistent with the fact that cachexia is more severe in men than in women [37] and that the opposite is observed in disuse muscle atrophy [38].

The mechanisms underpinning sex differences in fiber type composition remain to be determined: indeed, although the expression levels of several genes related to muscle fiber type phenotype (such as myosin heavy chain I, MyHC, and peroxisome proliferator-activated receptor delta, PPAR δ) are higher in women compared to men, there are no significant sex-based differences in the levels of the corresponding proteins [39]. However, higher mitochondria biogenesis and content was reported in female muscle compared to male muscle [40], which correlates with the higher number of oxidative fibers in females and with the prominent role of fat oxidation to produce adenosine triphosphate (ATP) [41]. Although it is recognized that women differ from men in their mitochondria features and activity, both in health and in disease [42], it is not clear how these differences may affect overall phenotypic and clinical outcomes [43]. Indeed, no differences in the respiration of gastrocnemius mitochondria between men and women have been observed [44]. Moreover, sex does not influence the expression of the creatine transporter or the content of creatine in the human skeletal muscle [45], which suggests that the major source of ATP for immediate use is equally available in the muscle tissue of both sexes.

Sex differences were also observed for lipid [46] and protein [47] metabolism and turnover. Different patterns of proteome regulation, including proteins involved in muscle

contraction and metabolism as well as in detoxification and antioxidant systems, were observed in rats between sexes [48]. In addition, human women have a higher protein turnover rate than men at all ages considered [49]. As expected, these differences in protein turnover are accounted for by hormones [50], which is reported in detail in this review. Nonetheless, the mechanisms underlying these differences between female and male muscles must be brought to light. Indeed, a major player in the balance of protein synthesis is mTOR (mammalian Target of Rapamycin), which, surprisingly, is similarly activated in the two sexes in response to well-known anabolic stimuli, such as exercise and food intake [51,52], with notable exceptions [53].

Satellite cells (SC) are important players in muscle regeneration following acute or chronic injury [54–58]. In addition to fiber hypertrophy, SC contribute to muscle growth in early postnatal life and following muscle damage due to exercise [59,60]. Overall, men have more SC and show greater SC proliferation compared to women [61,62], which is likely linked to the different availability of humoral factors [63]. Interestingly, sex-based differences in SC content are specific to type II fibers without any correlation with fiber size [64]. It is not surprising, then, that skeletal muscle regeneration exhibits sex differences in mice [65].

Sexually dimorphic growth is attributed to the growth hormone (GH)/insulin-like growth factor 1 (IGF1) axis. In women, the expressions of growth factor receptor-bound 10 (GRB10), which is inhibitory for IGF-1 signaling, and activin receptor IIB (ActR-IIB), which mediates a pathway leading to muscle atrophy, are higher than in men [66]. The expression and activity of some myokines appear to be different among sexes. As an example, the brain-derived neurotrophic factor (BDNF), a muscle-generated myokine that controls metabolic reprogramming upon fasting in a similar manner as physical exercise, displays sexual dimorphism [67,68]. In addition, the effects of interleukin 6 (IL-6) and myostatin, whose expressions are influenced by fasting, are fiber type-dependent and sex-dependent [69]; IL-6 plays different roles in muscle metabolism in female and male mice [70], and the effects of myostatin on muscle tissue are dose-, sex-, and muscle type-dependent [71]. GH regulates the abundance of mature myostatin by acting not only via the activator of transcription 5B (STAT5B) but also via a non-STAT5B pathway to regulate myostatin mRNA expression [72]. This double signaling pathway could explain why, in response to GH, the intracellular signal transducer STAT5B is dispensable, as shown in STAT5B $-/-$ mice [73]. The expressions of other growth factors, such as FGFs, vary not only with the type of skeletal muscle fibers but also according to sex in mice [74], extending the paradigm of sex differences in the autocrine, paracrine, and endocrine control of muscle growth to other factors. All of these findings also show that humoral factors affect muscle mass in a complex and interdependent fashion.

Sex-specific involvement of the neurohypophyseal peptides oxytocin (OXT) and vasopressin (AVP) in human behavior is well-established [75]. Less known is the fact that these two hormones can also be considered myokines [76], as they have profound effects on muscle homeostasis and development [77–79]. An additional, major endocrine difference between men and women is the axis from the anterior pituitary gland—via gonadotrophs—to sex organs, leading to the production of estrogen and progesterone, which are both associated with muscle growth and health in humans [80–82]. The role of estrogens in sexual dimorphism was comprehensively reviewed by McMillin et al. [83]. Estrogens (produced by granulosa and Sertoli cells in female and male individuals, respectively) vary in their circulating concentrations during the menstrual cycle in humans or the estrous cycle in mice; therefore, their level and activity should be considered when dealing with women of reproductive age. A meta-analysis addressing the effects of estradiol-based hormone replacement therapy on muscle mass clearly indicates that estradiol is beneficial for muscle maintenance [84]. On the other hand, androgens are chiefly responsible for the male phenotype [85], and circulating testosterone is one of the major factors responsible for sex differences in athletic performance due to the well-known dose–response relationship between its levels and those of muscle mass and strength [86]. Sex hormones appear to

be responsible for greater fat oxidation in women during endurance exercise compared to men [87]. Recently, an interplay between female sex hormones and IL18 was reported with important, sex-specific consequences on glucose intolerance and insulin signaling [88].

Based on all of these findings, skeletal muscle growth, metabolism, and homeostasis are sexually dimorphic (Figure 1). This suggests that women and men suffer from sarcopenia to a very different extent, possibly with distinctive mechanisms of disease. In the following paragraphs, we will highlight the major sexually dimorphic features of muscle atrophy in various conditions.

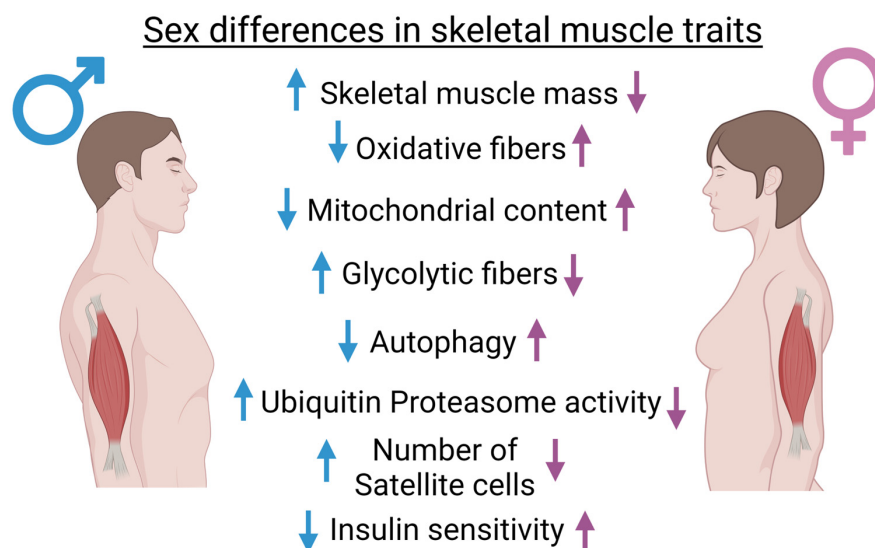


Figure 1. Major differences in muscle phenotypes of the two sexes in humans. Arrows pointing upwards mean stronger expression or higher number compared to the other sex, whereas arrows pointing downwards mean the opposite.

3. Sex Differences in Muscle Atrophy Associated with Disuse and Denervation

3.1. Major Differences in Clinical and Pre-Clinical Phenotypes

Muscle atrophy is associated with disuse, a condition due to prolonged bed rest or joint immobilization, resulting in the loss of skeletal muscle mass [43,89]. Similar to bed rest, the unloading condition due to microgravity, as in space flights, has multiple consequences, including a decrease in muscle mass [90]. Although disuse-induced muscle atrophy occurs in both men and women, many differences were observed between the sexes in both humans and animal models. Women suffer from greater muscle loss in intensive care units [38] and experience a higher risk of mortality compared to men [91]. Interestingly, a greater loss of knee extensor muscle strength (KES), despite a similar extent of atrophy, was observed in women compared to men following immobilization-induced disuse [92]. Conversely, following arm suspension, men displayed a significant decrease in the volume of flexor muscle that was not observed in women [93]. In another study, following hip fracture, men experienced a higher prevalence of sarcopenia than women [94]. Lastly, the mean thickness of the rectus femoris, although significantly different between male and female patients before surgery for femoral fractures, reached the same value in both sexes after a traction period of a few days [95]. Interestingly, patients of the two sexes may also differ in recovery capacity: men perform better than women after cast removal, as women require a more intense rehabilitation program [96]. During space flights, men and women show sex-specific adaptations with differences in immunity and metabolism, including compounds important for bone and muscle homeostasis and function [97].

Muscle atrophy is also associated with diseases such as osteoarthritis (OA), which is a frequent cause of disability due to lack of or poor joint mobility, ultimately resulting in disuse/reduced use of the muscle [98]. Sexual dimorphism was observed in OA; male patients display higher type IIa muscle fiber power and velocity compared to female patients. At

the molecular level, this can be due to the slower kinetics of myosin–actin cross-bridge in women compared to men [99]. In addition, the reduction of subsarcolemmal mitochondria observed in women with OA may also contribute to poorer muscle performance compared to men because mitochondrial fission and remodeling are involved in disuse muscle atrophy [100].

Taken together, these studies suggest that women are more susceptible to disuse muscle atrophy than men and display functional alterations different from men upon atrophying conditions. However, the results can be inconsistent or even entirely different depending on the conditions. For instance, cast immobilization (limited to a few muscles of one arm) in a subject capable to move and use other muscles is not comparable with almost total immobilization due to bed rest for a patient of the same or the opposite sex. A more correct view is probably that features other than sex (muscle type, immobilization length and extent, etc.) interact with sex to trigger muscle atrophy upon immobilization or to unload in various ways and to different extents.

It is worth noting that denervation [18,29,101] achieved by various means differs from casting [102], hindlimb suspension [103], or tenotomy [104] inasmuch as muscle atrophy occurs in the absence of the neurotrophic affects deriving from innervation (i.e., the maintenance of neuromuscular junctions). Nonetheless, we report here the few studies on sex differences in this condition due to its clinical relevance. By exploiting a novel murine model of mild spinal muscular atrophy, Kothari and coworkers demonstrated that men are slightly more susceptible than women to neuromuscular junction (NMJ) transmission defects and muscle fiber atrophy [105]; similarly, sex differences were observed in a mouse model of amyotrophic lateral sclerosis [106] and in humans with milder types of spinal muscular atrophy [107]. In xenopus, denervation induces muscle fiber atrophy in the muscles of the larynx, whereas androgen treatment induces muscle fiber hypertrophy; no sex differences were observed in fiber size modification due to innervation or androgen treatment but in the control of the number of muscle fibers [108]. Consistently, crush-induced nerve injury negatively affected the isometric contractile capacity of muscle EDL in mice regardless of sex [109]. These interesting, albeit sparse, findings are relevant because, taken together, they suggest that men could be more heavily affected than women following nerve resection or damages of motor neurons, whereas muscle atrophy is aggravated in women in innervated, unloaded muscles. Because age-related sarcopenia is partially due to a progressive and selective denervation of the fast-twitch fibers, denervation will be further discussed in Section 4, which is dedicated to aging.

3.2. Molecular Mechanisms and Sex Differences in Disuse Muscle Atrophy

To address the molecular mechanisms underlying disuse-induced atrophy, several animal models are available, which were reviewed by Musacchia [110]. Disuse muscle atrophy generally encompasses categories such as tenotomy, unloading, immobilization, and denervation. However, all of them are fundamentally unique. Rotator cuff tenotomy-induced muscle atrophy is sex-specific (exacerbated in male mice) and regulated by autophagy independently of Nuclear factor- κ B (NF- κ B) [104], which we and others have shown controls muscle wasting in other conditions [111,112]. In rats subjected to hindlimb unloading, there is a greater reduction in soleus muscle mass and fiber cross-sectional area (CSA) in women than in men due to a different activation of the FoxO3a/ubiquitin-proteasome pathway [113]. These results were confirmed in mice: upregulation of ubiquitin-ligases expression was observed in women, but not in men, as early as 24–48 h after hindlimb unloading together with the upregulation of Deptor and Redd1, two inhibitors of mTOR Complex 1 (mTORC1) [43]. In a model of hindlimb unloading, damage to mitochondrial functions were also investigated [114,115]: whereas mitochondrial degeneration was evident in male mice before the onset of muscle atrophy, the opposite occurred in women despite massive ROS production followed by degradative pathways and mitophagy [116]. Thus, oxidative stress may play a pivotal role in disuse-induced muscle atrophy [117].

4. Sex Differences in Aging-Associated Sarcopenia

4.1. Major Differences in Clinical and Pre-Clinical Phenotypes

Age-related sarcopenia is a condition characterized by a reduction in muscle mass, strength, and function with increasing age, with a relevant burden on global health and the management of elderly people [118]. The definition of sarcopenia evolved over the last 25 years thanks to discussion groups, such as EWGSOP, giving rising importance to the functional deficit, which is characteristic of sarcopenic muscle, in the diagnosis and management of sarcopenia [119–122]. Currently, recommendations exist for the treatment of sarcopenia, which include exercise and nutritional supplementation, e.g., vitamin D [25,123]; nonetheless, sex differences remain a neglected aspect for both primary (age-related) and secondary (disease-related) sarcopenia [118]. Indeed, sex differences can influence how men and women respond to aging, as discussed by Anderson et al. [124]. The risk factors for the development of age-related sarcopenia are different for men and women, and they were identified by Hwang and Park [125]. Both men and women manifest loss of skeletal muscle mass and function with increasing age, but men have a greater loss than women, even though this gross difference can be partly explained by the greater initial muscle mass that men have compared to women [126]. However, a different study showed that the quadriceps muscle cross-sectional area decreases with age, especially in women [127]. When assessing age-related strength loss, the abrupt age-related decline measured (KES) occurs earlier in women than men, whereas the corresponding isometric strength loss is similar between sexes [128]. Indeed, the differences in KES are accounted for by sex differences in the kinetics of the muscles contributing to this measurement, i.e., the rectus femoris, quadriceps, etc. [129]. Consistently, single fibers show sex-dependent alterations in size and a decrease in intermyofibrillar mitochondrial size with age, primarily in women [34]. Consistently, the typical slowing of myosin cross-bridge kinetics is particularly evident in elderly women, and this may account for the increased disability and contractile dysfunction of skeletal muscle [130]. Aging is also associated with progressive denervation, a phenomenon that can be reversed by exercise [131]. The effects of aging on the regulation of muscle contraction by neurons were studied [132], but, to our knowledge, most studies have not examined denervation in a sex-stratified manner or addressed the sex-dependent mechanisms underlying this phenomenon.

The lower appendicular mass of the skeletal muscle is associated with the increased risk of falls observed among elderly women compared to men [133,134], suggesting that differences in sarcopenia between the two sexes account for additional issues associated with aging, such as risk of morbidities and incidents. Certainly, frailty as a clinical condition, defined as an increased susceptibility to unfavorable health outcomes [135], contributes to aging-associated sarcopenia. Indeed, in the elderly, frailty represents the link between a healthy status and a poor outcome, including death, in people of the same chronological age. Some conflicting data were collected in the last 20 years regarding the sex differences in frailty [136], mainly because of the lack of a consensus in its definition and assessment or due to discrepancies in the study samples' characteristics or ethnicities. However, by using phenotypic and accumulated deficits as a frailty index, two systematic reviews found the prevalence of frailty to be higher in older women than men [137,138], which was also confirmed in a recent meta-analysis [136]. These conclusions are in alignment with, and may contribute to, an overall aging-associated sarcopenia that is particularly evident in elderly women compared to men.

4.2. Possible Mechanisms Accounting Sex Differences in Aging-Associated Sarcopenia

During aging, several factors underpinning muscle quality come into play, including muscle composition, aerobic capacity and metabolism, fatty infiltration, insulin resistance, fibrosis, and neural activation [139]. Looking for mechanisms responsible for sarcopenia in a sex-dependent fashion, it was proposed that a decrease in IGF1 contributes to the development of sarcopenia only in women [140]. In rats, soleus and extensor digitorum longus (EDL) muscle to body weight ratios steadily decrease with age in men but not in

women up to 26 months of age; these sex-dependent differences were associated with differences in the regulation of IGF-1 downstream effectors, such as protein kinase B (Akt), mTOR, and p70s6k, in the slow-twitch soleus and with the regulation of AMP activated protein kinase (AMPK), Eukaryotic translation initiation factor 4E-binding protein 1 (4EBP1), p70s6k, and rpS6 in the fast-twitch EDL [141].

By contrast, sex-related differences in the serum levels of the other major regulator of muscle mass, myostatin, with aging is unclear, and further investigations are needed. In men, serum levels of myostatin slightly increase with age up to around 57 years and then decrease [142], and low serum levels of myostatin were associated with low skeletal muscle mass in older adult men, but not in women. According to these findings, serum levels of myostatin cannot be used to diagnose sarcopenia or to monitor how sarcopenic muscles respond to treatments [143]. On the other hand, a different study showed that serum concentrations of myostatin and myostatin-interacting proteins do not differ between young and sarcopenic elderly men [144]. In addition, a strong negative association between circulating myostatin, follistatin, and muscle power in women but not in men was described [145]. The decrease of sex hormones that occurs with increasing age was also proposed to be responsible for sarcopenia. Indeed, the loss of skeletal muscle associated with the perimenopausal stage may be potentially related to increased levels in FSH [146]. In parallel, the deficit of hormones, such as testosterone and 17 β estradiol, associated with aging would be the cause of the altered activation of SC, which are critical for muscle repair and regeneration processes [147]. Malnutrition also plays an important role in muscle homeostasis, and because it is often associated with aging [148], it might be responsible for age-related sarcopenia [149]. Malnutrition leads to an increased risk of sarcopenia in women [140]. In addition, low levels of vitamin D are associated with muscle loss in elderly Chinese individuals [150] and lower appendicular skeletal muscle mass index scores in Korean women, for whom it is also associated with a greater proportion of hypovitaminosis [151] that, again, highlights the importance of vitamin D balance to counteract sarcopenia associated with aging.

5. Sex Differences in Cancer Cachexia

5.1. Major Differences in Clinical and Pre-Clinical Phenotypes

Cachexia is a wasting syndrome associated with chronic illnesses, including cancer, and characterized by weight loss and skeletal muscle wasting [152]. The consensus definition of cancer cachexia [153] boosted the recognition of its clinical relevance [154]. The prevalence of cachexia is very high (50–80%) in advanced malignant cancer [155]. Due to severe muscle wasting, cancer patients experience weakness and fatigue, which significantly lower their quality of life [26]. The onset of cachexia has a predictive value of poor survival and response to therapy [156], and it affects 20% of cancer patients [157].

Although the mechanisms of cachexia receive increasing attention, sex differences in this syndrome are far less appreciated. Biological differences between men and women may account for different responses to cachexia at multiple levels: susceptibility, progression, and response to treatment [158]. The diagnostic and prognostic assessment of cachexia relies on both the body mass index (BMI) and the rate of ongoing weight loss [153,159]. The fact that men and women have different BMI immediately suggests that the susceptibility to cachexia and its severity are different between the two sexes. Moreover, men and women differ in the relative amount of fiber types, with women generally having mitochondria-enriched, more oxidative muscles. This fact results in an intrinsic higher respiratory capacity in mitochondria from women with respect to men [42,160] as well as differences in the metabolism of malonyl-CoA [161], which may account for the sex differences in cancer cachexia. Two studies on hundreds of cancer patients revealed that men showed muscle wasting two times more frequently than women [162,163]. Quite consistently, sexual dimorphism was observed in cachexia, including different decreases of muscle fiber cross-sectional area, expressions of *atrogenes* (*Foxo*, *Ub-ligases*, etc.), or expressions of genes responsible for muscle growth (*AKT1*, *MSTN*, etc.), apoptosis (*CASP9*), and inflammation

(TNF and STAT3) [164]. All of these findings result in a greater reduction of force in men than women [37]. Among patients with lymphoma, both progression-free survival and overall survival were decreased in men with sarcopenia and not significantly affected in sarcopenic women [165], confirming the importance of muscle wasting for prognosis.

5.2. Sex Differences in the Mechanisms Leading to Cachexia

Consistent with the clinical observations described previously, the mechanisms underlying cachexia appear to be different for the two sexes. As a caveat, it is worth noting that, although they confirm the existence of sex differences, animal models do not always mirror the prevalent human condition in cancer cachexia. Indeed, in a tumor-bearing mouse model, female mice developed body and limb muscle weight loss at early stages of cachexia but maintained their protein amounts and specific force, whereas the opposite was observed in male mice [166]. Alterations of mitochondria were widely reported in cachexia, suggesting a new avenue of investigation [167,168]. Nonetheless, no studies so far have been dedicated to identifying sex differences regarding mitochondria's role in cachexia.

Similarly, the role of microRNAs in cachexia is a growing field of investigation [169]; however, the characterization of their differential modulation in the two sexes during cachexia is still missing today.

More significant progress was done on sexual dimorphism related to humoral factors as triggers of muscle atrophy in cachexia. The ligands of the activin receptor IIB (ActR-IIB), such as myostatin, activin, and other members of the TGF β superfamily, were identified as major players in muscle wasting and proposed as therapeutic targets [170]. In pancreatic ductal adenocarcinoma patients, activin is a preferential driver of muscle wasting in men [171]. Altered levels of GDF15 associated with aging in humans—higher in older men than in age-matching women [172]—were proposed as causative of both sarcopenia and the low physical performance of the muscle [173,174]. Therefore, GDF15 is now heavily investigated in cachexia because blocking GDF15 signaling may have the potential to counteract cachexia [175]. However, to the best of our knowledge, the impact of sex on GDF15's effects have not been carefully investigated yet. Whereas IL-6 levels inversely correlate with BMI in cancer patients [176], the samples were not stratified according to the sex of the patient. However, in animal models, female animals are more resistant to high levels of pro-inflammatory cytokines, such as IL-6 [177], which is probably due to a reduced catabolic response in muscle tissue [178]; in addition, a sex-dependent genetic predisposition to produce high levels of IL-6 exists due to polymorphism in the promoter of this gene [179]. The role of sex hormones was addressed in animal models of cancer, revealing that cachexia is associated with the cessation of estrous cycling [180]. The expression of estrogen receptors in muscle cells is not clear due to conflicting results [83], and additional research is required to fully elucidate the cellular and molecular mechanisms underlying 17- β estradiol-mediated effects. However, the effort will be rewarding because 17- β estradiol deficiency is shared by several conditions of skeletal muscle wasting, such as disuse, injury, cachexia, and sarcopenia, and any progress in this query will lead to applications for multiple conditions.

6. Sex Differences in Immune Responses and Inflammation

6.1. Major Differences in the Inflammatory Response

There is now ample evidence that sex is an important determinant of the immune response in the context of inflammation in various disease settings, including infection, autoimmunity, and cancer, and that sex differences strongly influence disease symptoms' severity and mortality. Existing epidemiological data reveal a critical role for sex differences in the immune response against viral, self, and tumor antigens, with women generally showing more robust innate and adaptive immune responses [181–183]. These differences are largely driven by differences in sex chromosome gene expression and in circulating levels of sex hormones including estrogens, progesterone, and androgens [181–185]. Estrogen and progesterone receptors are expressed by most immune cells, and 17- β estradiol

boosts both cell-mediated and humoral immune responses [184,185], whereas progesterone has anti-inflammatory effects [186]. By contrast, androgens generally dampen the immune response [181,183]. Moreover, a number of genes on the X chromosome code for immune response-related proteins such as Toll-like receptors (TLRs) (in particular TLR7 and TLR8), interleukin 2 receptors (IL2R), and transcriptional factors (such as FOXP3), which regulate the immune response, and therefore, they contribute towards sex differences in the development of inflammatory diseases [187].

Circulating levels of estrogen were associated with more severe symptoms in a mouse model of systemic lupus erythematosus (SLE), and the removal of estrogen improves disease prognosis [188]. On the other hand, lower serum levels of androgens in elderly men is associated with an increased incidence of rheumatoid arthritis (RA) [189]. Although elevated innate and adaptive immunity in women may drive the progression of autoimmune diseases, such as Systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA), it is advantageous in anti-tumor responses.

6.2. Inflammation throughout Different Conditions of Muscle Atrophy

Aging is typically associated with a moderately, albeit relevant, increased level of inflammation, even though it is not clear whether the so-called “inflammaging” is a cause or an effect of aging [190]. Some chronic conditions that present as age-associated comorbidities can definitely accelerate aging due to increased inflammation mediated by immune dysfunction [191]. Bed rest induces a small rise in pro-inflammatory cytokines, which can reach a statistically significant increase for specific ones, such as IL-6 [192,193]. Microgravity determines aging-like phenomena mediated by chronic low-grade inflammation as well [194]. On the contrary, systemic inflammation accompanied by increased circulation of proinflammatory cytokines is an important feature of cancer and contributes significantly to loss of muscle mass and the development of cancer cachexia [195].

Based on all of these findings, the changes in levels of pro-inflammatory cytokines seem to be abrupt and much more pronounced in cancer cachexia compared to other forms of muscle atrophy, such as those following unloading/disuse or associated with aging. Interestingly, men respond differently than women to these forms of muscle atrophy.

All of the information about the differences between the two sexes and the corresponding references cited in this review are summarized in Table 1.

Table 1. Sex-related differences in muscle phenotype under physiological or pathological conditions.

Muscle Conditions	Sex-Related Differences in Muscle Traits	Reference(s)
Physiological	Mass	[30]
	Energy metabolism	[32–34]
	Mitochondrial content	[42]
	Protein turnover	[47,50]
	Insulin sensitivity	[66,88]
	Muscle regeneration	[61,62,65]
Disuse	Muscle weight	[38,93]
	Muscle force	[92,98]
	Myofilament cross bridge kinetics	[99]
	Recovery	[96]
Aging	Muscle weight	[126,127]
	Muscle force	[128,129]
	Myofilament cross bridge kinetics	[130]
	Frailty	[136–138]
Cancer Cachexia	Muscle weight	[162,163]
	Muscle force	[37,162,163]
	Overall survival	[158,165]

7. Conclusions

7.1. Inflammation-Driven Muscle Atrophy: Are Cytokines the Culprit?

Here we have presented, in a comparative way, sex differences in three forms of sarcopenia (Figure 2). Aging seems to affect men more severely in terms of muscle mass loss, but it affects women more insofar as muscle function is preferably considered. Disuse affects muscle atrophy in women more than men [38], whereas cancer cachexia is the opposite [98,162]. One difference between disuse and cachexia is the absence or presence of a significant degree of inflammation due, in the latter, to tumor–host interactions [196,197]. Inflammatory cells deeply affect SC behavior and muscle homeostasis [198,199] and are promising new targets to treat muscle diseases [200]. Even though inflammation does not necessarily correspond to an increased presence of inflammatory cells in muscle infiltrates [201], pro-inflammatory cytokines directly target striated muscles, triggering muscle wasting [202] and inhibiting muscle regeneration [60,203]. In addition to the cytokines released by the immune system, the levels of circulating myokines are strongly dependent on the amount of muscle mass present, which is overtly different between sexes. Based on the above, we propose that, in addition to obvious differences in hormone and growth factors, differences in myokines and cytokines must be taken into account when considering the mechanisms of differential muscle atrophy observed in the two sexes in different forms of muscle atrophy.

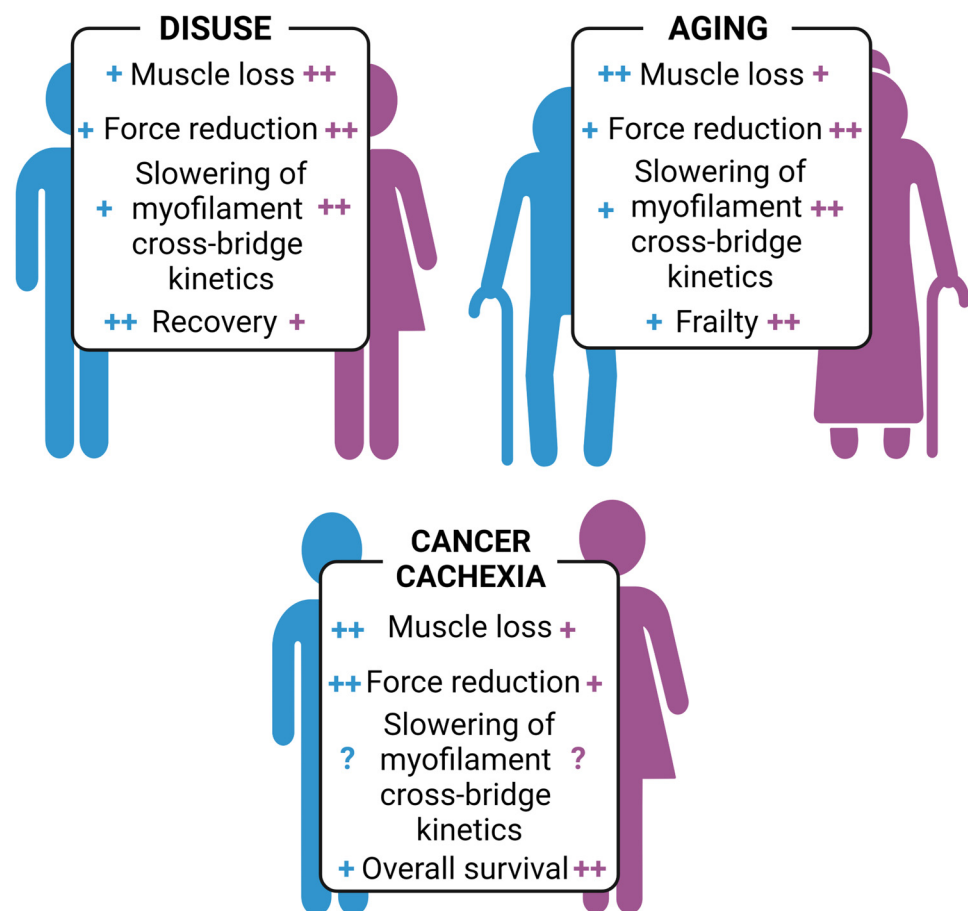


Figure 2. Sex differences in the response to disuse/unloading, age-related sarcopenia, and cancer cachexia. ++ as compared to + refers to a more pronounced feature in the corresponding sex (blue color for men, pink color for women); ? means no consistent data are available.

7.2. Which Direction Shall We Go?

The US NIH already requires taking into account sex as a biological variable in preclinical studies [204]. However, we propose a step forward in this direction: comparisons based on the sex of the organism should be systematically planned in both clinical and experimental studies dealing with muscle atrophy from now on. Remarkably, there is an issue even with studies addressing sex differences in a variety of biological disciplines; as beautifully demonstrated by Garcia-Sifuentes and Maney, often when a sex-specific effect is claimed based on experimental data, the authors do not actually statistically test the differences [204]. This often makes it difficult to actually state if and to what extent sex differences exist, and it calls for further investigation on this important aspect of biology.

In clinical trials, the study groups are not systemically stratified by the sex of the patients, which is often due to the small size of the cohort studied. Nonetheless, it was already reported that the results may change significantly depending on the sex of the patient. For instance, the treatment with the common immunosuppressant rapamycin has sex-specific effects [15], highlighting the importance of taking into account sex differences for precision medicine. The same is true for physical exercise [205] as an intervention against cancer. The conclusion of a ponderous survey on the effectiveness, acceptability, and safety of exercise for cancer cachexia in adults is that “further high-quality randomized controlled trials are still required to test exercise alone or as part of a multimodal intervention to improve people’s well-being throughout all phases of cancer care”, suggesting that additional clinical and basic studies are needed to implement exercise efficacy [206]. Because men and women respond differently to both endurance and resistance exercise training [207,208]—which seems obvious based on the profound differences in their musculature, which are summarized in the first section of this review—the sex of the patient represents a major variable to be taken into account for future studies. The challenges and opportunities for future research on sex differences have been discussed [209]. In addition, guidelines and methods to test sex differences were recently published [210].

Furthermore, an effort should be made to clarify the role of inflammation in different conditions as opposed to that of reduced mechanical, contraction-mediated stimuli. Indeed, depending on the specific condition, muscle wasting may be due to the inflammatory factors present at high levels in a given disease state plus the secondary sarcopenia due to other factors, likely leading to a positive feedback loop [211]. For instance, in intensive care units (ICU), patients experience high inflammation typical of critical illness combined with bed rest, both contributing to inflammatory disequilibrium; similarly, the elderly may show chronic inflammation combined with reduced activity due to poor muscle performance. To better address the relative contribution of each of these factors to muscle wasting, it will be interesting to compare similar conditions, ideally differing in one variable. For instance, is the amount of muscle wasting in space flights (i.e., a “purely” microgravity condition) the same as in bed rest in an ICU (which is characterized by inflammation induced by injury or a severe disease)?

7.3. Final Remarks

Only in recent years has the importance of personalized medicine, also known as precision medicine, gained momentum [212], and tailored treatments have emerged in clinical trials [14]. In pre-clinical research, exploring sex differences in various disease conditions may be the gateway to successful treatments [16]. For example, any protective factors discovered in one sex could be exploited to lower disease morbidity and severity or avoid mortality in the opposite sex [158]. In particular, the understanding of sex-dependent responses to different forms of muscle atrophy and inflammation is of pivotal importance for the design of innovative, tailored, and efficient interventions.

Author Contributions: Conceptualization, M.B. and D.C.; writing—original draft preparation, C.D.P., B.L.-O. and D.C.; writing—review and editing, B.L.-O., A.R., V.M., C.S.R., M.B. and D.C.; supervision, M.B. and D.C. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not Applicable.

Informed Consent Statement: Not Applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Deng, X.; Berletch, J.B.; Nguyen, D.K.; Distèche, C.M. X chromosome regulation: Diverse patterns in development, tissues and disease. *Nat. Rev. Genet.* **2014**, *15*, 367–378. [[CrossRef](#)]
- Juchniewicz, P.; Piotrowska, E.; Kloska, A.; Podlacha, M.; Mantej, J.; Węgrzyn, G.; Tukaj, S.; Jakóbkiewicz-Banecka, J. Dosage Compensation in Females with X-Linked Metabolic Disorders. *Int. J. Mol. Sci.* **2021**, *22*, 4514. [[CrossRef](#)] [[PubMed](#)]
- Migeon, B.R. Why females are mosaics, x-chromosome inactivation, and sex differences in disease. *Genet. Med.* **2007**, *4*, 97–105. [[CrossRef](#)]
- Duman, R.S. Sex-specific disease-associated modules for depression. *Nat. Med.* **2017**, *23*, 1015–1017. [[CrossRef](#)]
- Shakeshaft, A.; Panjwani, N.; Collingwood, A.; Crudgington, H.; Hall, A.; Andrade, D.M.; Beier, C.P.; Fong, C.Y.; Gardella, E.; Gesche, J.; et al. Sex-specific disease modifiers in juvenile myoclonic epilepsy. *Sci. Rep.* **2022**, *12*, 2785. [[CrossRef](#)]
- Ortona, E.; Pierdominici, M.; Maselli, A.; Veroni, C.; Aloisi, F.; Shoenfeld, Y. Sex-based differences in autoimmune diseases. *Ann. Dell'istituto Super. Di Sanita* **2016**, *52*, 205–212. [[CrossRef](#)]
- Foltz, S.; Wu, F.; Ghazal, N.; Kwong, J.Q.; Hartzell, H.C.; Choo, H.J. Sex differences in the involvement of skeletal and cardiac muscles in myopathic *Ano5*^{-/-} mice. *Am. J. Physiol. Physiol.* **2022**, *322*, C283–C295. [[CrossRef](#)] [[PubMed](#)]
- Winham, S.J.; de Andrade, M.; Miller, V.M. Genetics of cardiovascular disease: Importance of sex and ethnicity. *Atherosclerosis* **2015**, *241*, 219–228. [[CrossRef](#)] [[PubMed](#)]
- Lopez, M.-C.; Efron, P.A.; Ozrazgat-Baslanti, T.; Zhang, J.; Cuschieri, J.; Maier, R.V.; Minei, J.P.; Baker, H.V.; Moore, F.A.; Moldawer, L.L.; et al. Sex-based differences in the genomic response, innate immunity, organ dysfunction, and clinical outcomes after severe blunt traumatic injury and hemorrhagic shock. *J. Trauma Acute Care Surg.* **2016**, *81*, 478–485. [[CrossRef](#)] [[PubMed](#)]
- Wyld, M.L.; De La Mata, N.L.; Viecelli, A.; Swaminathan, R.; O'Sullivan, K.M.; O'Lone, E.; Rowlandson, M.; Francis, A.; Wyburn, K.; Webster, A.C. Sex-Based Differences in Risk Factors and Complications of Chronic Kidney Disease. *Semin. Nephrol.* **2022**, *42*, 153–169. [[CrossRef](#)] [[PubMed](#)]
- Vinnik, T.; Kreinin, A.; Abildinova, G.; Batpenova, G.; Kirby, M.; Pinhasov, A. Biological Sex and IgE Sensitization Influence Severity of Depression and Cortisol Levels in Atopic Dermatitis. *Dermatology* **2020**, *236*, 336–344. [[CrossRef](#)] [[PubMed](#)]
- Lubin, J.H.; Muscat, J.; Gaudet, M.M.; Olshan, A.F.; Curado, M.P.; Maso, L.D.; Wunsch-Filho, V.; Sturgis, E.M.; Szeszenia-Dabrowska, N.; Castellsagué, X.; et al. An examination of male and female odds ratios by BMI, cigarette smoking, and alcohol consumption for cancers of the oral cavity, pharynx, and larynx in pooled data from 15 case-control studies. *Cancer Causes Control.* **2011**, *22*, 1217–1231. [[CrossRef](#)] [[PubMed](#)]
- Sylvester, M.A.; Brooks, H.L. Sex-Specific Mechanisms in Inflammation and Hypertension. *Curr. Hypertens. Rep.* **2019**, *21*, 53. [[CrossRef](#)] [[PubMed](#)]
- Giesen, N.; Chatterjee, M.; Scheid, C.; Poos, A.M.; Besemer, B.; Miah, K.; Benner, A.; Becker, N.; Moehler, T.; Metzler, I.; et al. A phase II clinical trial of combined BRAF/MEK inhibition for *BRAF*^{V600E}-mutated multiple myeloma. *Blood* **2023**. [[CrossRef](#)]
- Wolff, C.A.; Lawrence, M.M.; Porter, H.; Zhang, Q.; Reid, J.J.; Laurin, J.L.; Musci, R.V.; Linden, M.A.; Peelor, F.F.; Wren, J.D.; et al. Sex differences in changes of protein synthesis with rapamycin treatment are minimized when metformin is added to rapamycin. *Geroscience* **2020**, *43*, 809–828. [[CrossRef](#)] [[PubMed](#)]
- Man, J.J.; Beckman, J.A.; Jaffe, I.Z. Sex as a Biological Variable in Atherosclerosis. *Circ. Res.* **2020**, *126*, 1297–1319. [[CrossRef](#)] [[PubMed](#)]
- Lynch, M.A. Exploring Sex-Related Differences in Microglia May Be a Game-Changer in Precision Medicine. *Front. Aging Neurosci.* **2022**, *14*, 281. [[CrossRef](#)] [[PubMed](#)]
- Madaro, L.; Passafaro, M.; Sala, D.; Etzaniz, U.; Lugarini, F.; Proietti, D.; Alfonsi, M.V.; Nicoletti, C.; Gatto, S.; De Bardi, M.; et al. Denervation-activated STAT3-IL-6 signalling in fibro-adipogenic progenitors promotes myofibres atrophy and fibrosis. *Nature* **2018**, *20*, 917–927. [[CrossRef](#)]
- Riera, C.S.; Lozanoska-Ochser, B.; Testa, S.; Fornetti, E.; Bouché, M.; Madaro, L. Muscle Diversity, Heterogeneity, and Gradients: Learning from Sarcoglycanopathies. *Int. J. Mol. Sci.* **2021**, *22*, 2502. [[CrossRef](#)]
- Ji, H.; Kwan, A.C.; Chen, M.T.; Ouyang, D.; Ebinger, J.E.; Bell, S.P.; Niiranen, T.J.; Bello, N.A.; Cheng, S. Sex Differences in Myocardial and Vascular Aging. *Circ. Res.* **2022**, *130*, 566–577. [[CrossRef](#)]

21. Kozdag, G.; Ertas, G.; Emre, E.; Akay, Y.; Celikyurt, U.; Sahin, T.; Gorur, G.; Karauzum, K.; Yilmaz, I.; Ural, D.; et al. Low serum triglyceride levels as predictors of cardiac death in heart failure patients. *Tex. Hear. Inst. J.* **2013**, *40*, 521–528.
22. Liu, M.; Zhang, Z.; Zhou, C.; Ye, Z.; He, P.; Zhang, Y.; Li, H.; Liu, C.; Qin, X. Predicted fat mass and lean mass in relation to all-cause and cause-specific mortality. *J. Cachex-Sarcopenia Muscle* **2022**, *13*, 1064–1075. [[CrossRef](#)]
23. Nunes, E.A.; Stokes, T.; McKendry, J.; Currier, B.S.; Phillips, S.M. Disuse-induced skeletal muscle atrophy in disease and nondisease states in humans: Mechanisms, prevention, and recovery strategies. *Am. J. Physiol. Physiol.* **2022**, *322*, C1068–C1084. [[CrossRef](#)] [[PubMed](#)]
24. Priyadarsini, N.; Nanda, P.; Devi, S.; Mohapatra, S. Sarcopenia: An Age-Related Multifactorial Disorder. *Curr. Aging Sci.* **2022**. [[CrossRef](#)]
25. Garcia, M.; Seelaender, M.; Sotiropoulos, A.; Coletti, D.; Lancha, A.H. Vitamin D, muscle recovery, sarcopenia, cachexia, and muscle atrophy. *Nutrition* **2019**, *60*, 66–69. [[CrossRef](#)]
26. Berardi, E.; Madaro, L.; Lozanoska-Ochser, B.; Adamo, S.; Thorrez, L.; Bouche, M.; Coletti, D. A pound of flesh: What cachexia is and what it is not. *Diagnostics* **2021**, *11*, 116. [[CrossRef](#)] [[PubMed](#)]
27. Lecker, S.H.; Jagoe, R.T.; Gilbert, A.; Gomes, M.; Baracos, V.; Bailey, J.; Price, S.R.; Mitch, W.E.; Goldberg, A.L. Multiple types of skeletal muscle atrophy involve a common program of changes in gene expression. *FASEB J.* **2003**, *18*, 39–51. [[CrossRef](#)] [[PubMed](#)]
28. Bouchè, M.; Lozanoska-Ochser, B.; Proietti, D.; Madaro, L. Do neurogenic and cancer-induced muscle atrophy follow common or divergent paths? *Eur. J. Transl. Myol.* **2018**, *28*, 7931. [[CrossRef](#)] [[PubMed](#)]
29. Pigna, E.; Sanna, K.; Coletti, D.; Li, Z.; Parlakian, A.; Adamo, S.; Moresi, V. Increasing autophagy does not affect neurogenic muscle atrophy. *Eur. J. Transl. Myol.* **2018**, *28*, 7687. [[CrossRef](#)]
30. Rosa-Caldwell, M.E.; Greene, N.P. Muscle metabolism and atrophy: Let's talk about sex. *Biol. Sex Differ. BioMed Cent.* **2019**, *10*, 43. [[CrossRef](#)] [[PubMed](#)]
31. O'Reilly, J.; Ono-Moore, K.D.; Chintapalli, S.V.; Rutkowsky, J.M.; Tolentino, T.; Lloyd, K.C.K.; Olfert, I.M.; Adams, S.H. Sex differences in skeletal muscle revealed through fiber type, capillarity, and transcriptomics profiling in mice. *Physiol. Rep.* **2021**, *9*, e15031. [[CrossRef](#)] [[PubMed](#)]
32. Haizlip, K.M.; Harrison, B.C.; Leinwand, L.A. Sex-Based Differences in Skeletal Muscle Kinetics and Fiber-Type Composition. *Physiology* **2015**, *30*, 30–39. [[CrossRef](#)] [[PubMed](#)]
33. Mauvais-Jarvis, F. Sex differences in metabolic homeostasis, diabetes, and obesity. *Biol. Sex Differ.* **2015**, *6*, 1–9. [[CrossRef](#)] [[PubMed](#)]
34. Callahan, D.M.; Bedrin, N.G.; Subramanian, M.; Berking, J.; Ades, P.A.; Toth, M.J.; Miller, M.S. Age-related structural alterations in human skeletal muscle fibers and mitochondria are sex specific: Relationship to single-fiber function. *J. Appl. Physiol.* **2014**, *116*, 1582–1592. [[CrossRef](#)] [[PubMed](#)]
35. Brown, J.L.; Rosa-Caldwell, M.E.; Lee, D.E.; Blackwell, T.A.; Brown, L.A.; Perry, R.A.; Haynie, W.S.; Hardee, J.P.; Carson, J.A.; Wiggs, M.P.; et al. Mitochondrial degeneration precedes the development of muscle atrophy in progression of cancer cachexia in tumour-bearing mice. *J. Cachex-Sarcopenia Muscle* **2017**, *8*, 926–938. [[CrossRef](#)]
36. Wang, Y.; Pessin, J.E. Mechanisms for fiber-type specificity of skeletal muscle atrophy. *Curr. Opin. Clin. Nutr. Metab. Care* **2013**, *16*, 243–250. [[CrossRef](#)]
37. Norman, K.; Stobäus, N.; Reiß, J.; Schulzke, J.; Valentini, L.; Pirlich, M. Effect of sexual dimorphism on muscle strength in cachexia. *J. Cachex-Sarcopenia Muscle* **2012**, *3*, 111–116. [[CrossRef](#)]
38. De Jonghe, B. Paresis Acquired in the Intensive Care Unit A Prospective Multicenter Study. *JAMA* **2002**, *288*, 2859–2867. [[CrossRef](#)] [[PubMed](#)]
39. Maher, A.C.; Fu, M.H.; Isfort, R.J.; Varbanov, A.R.; Qu, X.A.; Tarnopolsky, M.A. Sex Differences in Global mRNA Content of Human Skeletal Muscle. *PLoS ONE* **2009**, *4*, e6335. [[CrossRef](#)] [[PubMed](#)]
40. Colom, B.; Alcolea, M.P.; Valle, A.; Oliver, J.O.; Roca, P.; García-Palmer, F.J. Skeletal Muscle of Female Rats Exhibit Higher Mitochondrial Mass and Oxidative-Phosphorylative Capacities Compared to Males. *Cell. Physiol. Biochem.* **2007**, *19*, 205–212. [[CrossRef](#)] [[PubMed](#)]
41. Montero, D.; Madsen, K.; Meinild-Lundby, A.-K.; Edin, F.; Lundby, C. Sexual dimorphism of substrate utilization: Differences in skeletal muscle mitochondrial volume density and function. *Exp. Physiol.* **2018**, *103*, 851–859. [[CrossRef](#)]
42. Miotto, P.M.; McGlory, C.; Holloway, T.M.; Phillips, S.M.; Holloway, G.P. Sex differences in mitochondrial respiratory function in human skeletal muscle. *Am. J. Physiol. Integr. Comp. Physiol.* **2018**, *314*, R909–R915. [[CrossRef](#)] [[PubMed](#)]
43. Rosa-Caldwell, M.E.; Lim, S.; Haynie, W.A.; Brown, J.L.; Deaver, J.W.; Da Silva, F.M.; Jansen, L.T.; Lee, D.E.; Wiggs, M.P.; Washington, T.A.; et al. Female mice may have exacerbated catabolic signalling response compared to male mice during development and progression of disuse atrophy. *J. Cachex-Sarcopenia Muscle* **2021**, *12*, 717–730. [[CrossRef](#)]
44. Thompson, J.R.; Swanson, S.A.; Casale, G.P.; Johanning, J.M.; Papoutsis, E.; Koutakis, P.; Miserlis, D.; Zhu, Z.; Pipinos, I.I. Gastrocnemius mitochondrial respiration: Are there any differences between men and women? *J. Surg. Res.* **2013**, *185*, 206–211. [[CrossRef](#)] [[PubMed](#)]
45. Murphy, R.M.; Tunstal, R.J.; Mehan, K.A.; Cameron-Smith, D.; McKenna, M.J.; Spriet, L.L.; Hargreaves, M.; Snow, R.J. Human skeletal muscle creatine transporter mRNA and protein expression in healthy, young males and females. *Mol. Cell. Biochem.* **2003**, *244*, 151–157. [[CrossRef](#)]

46. Beaudry, K.M.; Devries, M.C. Sex-based differences in hepatic and skeletal muscle triglyceride storage and metabolism. *Appl. Physiol. Nutr. Metab.* **2019**, *44*, 805–813. [[CrossRef](#)] [[PubMed](#)]
47. Smith, G.I.; Mittendorfer, B. Sexual dimorphism in skeletal muscle protein turnover. *J. Appl. Physiol.* **2016**, *120*, 674–682. [[CrossRef](#)] [[PubMed](#)]
48. Oh, T.S.; Choi, J.-W.; Choi, D.K.; Mukherjee, R.; Liu, H.; Yun, J.W. Gender Dimorphism in Skeletal Muscle Proteome Between Lean and Diet-induced Obese Rats. *Cell. Physiol. Biochem.* **2011**, *28*, 981–996. [[CrossRef](#)] [[PubMed](#)]
49. Hirsch, K.R.; Church, D.D.; Kim, I.; Park, S.; Wolfe, R.R.; Ferrando, A.A. Comparison of basal whole-body protein kinetics and muscle protein synthesis between young and older adults. *Physiol. Rep.* **2020**, *8*, e14633. [[CrossRef](#)]
50. Hansen, M.; Kjaer, M. Influence of Sex and Estrogen on Musculotendinous Protein Turnover at Rest and After Exercise. *Exerc. Sport Sci. Rev.* **2014**, *42*, 183–192. [[CrossRef](#)]
51. West, D.W.D.; Burd, N.; Churchward-Venne, T.A.; Camera, D.; Mitchell, C.; Baker, S.K.; Hawley, J.; Coffey, V.G.; Phillips, S. Sex-based comparisons of myofibrillar protein synthesis after resistance exercise in the fed state. *J. Appl. Physiol.* **2012**, *112*, 1805–1813. [[CrossRef](#)] [[PubMed](#)]
52. Jahn, L.A.; Barrett, E.J.; Genco, M.L.; Wei, L.; Spraggins, T.A.; Fryburg, D.A. Tissue Composition Affects Measures of Postabsorptive Human Skeletal Muscle Metabolism: Comparison across Genders 1. *J. Clin. Endocrinol. Metab.* **1999**, *84*, 1007–1010. [[CrossRef](#)]
53. Horstman, A.M.H.; Kouw, I.W.K.; van Dijk, J.-W.; Hamer, H.M.; Groen, B.B.L.; van Kranenburg, J.; Gorissen, S.H.M.; van Loon, L.J.C. The Muscle Protein Synthetic Response to Whey Protein Ingestion Is Greater in Middle-Aged Women Compared With Men. *J. Clin. Endocrinol. Metab.* **2018**, *104*, 994–1004. [[CrossRef](#)]
54. Coletti, D.; Teodori, L.; Lin, Z.; Beranudin, J.F.; Adamo, S. Restoration versus reconstruction: Cellular mechanisms of skin, nerve and muscle regeneration compared. *Regen. Med. Res.* **2013**, *1*, 4. [[CrossRef](#)] [[PubMed](#)]
55. Beltrà, M.; Pin, F.; Costamagna, D.; Duellen, R.; Renzini, A.; Ballarò, R.; Garcia-Castillo, L.; Iannuzzi, A.; Moresi, V.; Coletti, D.; et al. PGC-1 α in the myofibers regulates the balance between myogenic and adipogenic progenitors affecting muscle regeneration. *iScience* **2022**, *25*, 105480. [[CrossRef](#)]
56. Toschi, A.; Severi, A.; Coletti, D.; Catizone, A.; Musarò, A.; Molinaro, M.; Nervi, C.; Adamo, S.; Scicchitano, B.M. Skeletal Muscle Regeneration in Mice Is Stimulated by Local Overexpression of V1a-Vasopressin Receptor. *Mol. Endocrinol.* **2011**, *25*, 1661–1673. [[CrossRef](#)]
57. Benedetti, A.; Fiore, P.F.; Madaro, L.; Lozanoska-Ochser, B.; Bouché, M. Targeting PKC θ Promotes Satellite Cell Self-Renewal. *Int. J. Mol. Sci.* **2020**, *21*, 2419. [[CrossRef](#)]
58. Fiore, P.F.; Benedetti, A.; Sandonà, M.; Madaro, L.; De Bardi, M.; Saccone, V.; Puri, P.L.; Gargioli, C.; Lozanoska-Ochser, B.; Bouché, M. Lack of PKC θ Promotes Regenerative Ability of Muscle Stem Cells in Chronic Muscle Injury. *Int. J. Mol. Sci.* **2020**, *21*, 932. [[CrossRef](#)] [[PubMed](#)]
59. Zhenlin, L.; Parlakian, A.; Coletti, D.; Alonso-Martin, S.; Hourdé, C.; Joanne, P.; Gao-Li, J.; Blanc, J.; Ferry, A.; Paulin, D.; et al. Synemin acts as a regulator of signalling molecules in skeletal muscle hypertrophy. *J. Cell Sci.* **2014**, *127*, 4589–4601. [[CrossRef](#)]
60. Musarò, A.; Giacinti, C.; Pelosi, L.; Dobrowolny, G.; Barberi, L.; Nardis, C.; Coletti, D.; Scicchitano, B.M.; Adamo, S.; Molinaro, M. Stem cell-mediated muscle regeneration and repair in aging and neuromuscular diseases. *Eur. J. Histochem.* **2007**, *51*.
61. Neal, A.; Boldrin, L.; Morgan, J.E. The Satellite Cell in Male and Female, Developing and Adult Mouse Muscle: Distinct Stem Cells for Growth and Regeneration. *PLoS ONE* **2012**, *7*, e37950. [[CrossRef](#)] [[PubMed](#)]
62. Manzano, R.; Toivonen, J.M.; Calvo, A.C.; Miana-Mena, F.J.; Zaragoza, P.; Muñoz, M.J.; Montarras, D.; Osta, R. Sex, fiber-type, and age dependent in vitro proliferation of mouse muscle satellite cells. *J. Cell. Biochem.* **2011**, *112*, 2825–2836. [[CrossRef](#)]
63. Lee, D.-M.; Bajracharya, P.; Lee, E.J.; Kim, J.-E.; Lee, H.-J.; Chun, T.; Kim, J.; Cho, K.H.; Chang, J.; Hong, S.; et al. Effects of gender-specific adult bovine serum on myogenic satellite cell proliferation, differentiation and lipid accumulation. *Vitr. Cell. Dev. Biol.-Anim.* **2011**, *47*, 438–444. [[CrossRef](#)] [[PubMed](#)]
64. Horwath, O.; Moberg, M.; Larsen, F.J.; Philp, A.; Apró, W.; Ekblom, B. Influence of sex and fiber type on the satellite cell pool in human skeletal muscle. *Scand. J. Med. Sci. Sport.* **2020**, *31*, 303–312. [[CrossRef](#)] [[PubMed](#)]
65. Bahri, O.A.; Naldaiz-Gastesi, N.; Kennedy, D.C.; Wheatley, A.M.; Izeta, A.; McCullagh, K.J.A. The panniculus carnosus muscle: A novel model of striated muscle regeneration that exhibits sex differences in the mdx mouse. *Sci. Rep.* **2019**, *9*, 15964. [[CrossRef](#)] [[PubMed](#)]
66. Welle, S.; Tawil, R.; Thornton, C.A. Sex-Related Differences in Gene Expression in Human Skeletal Muscle. *PLoS ONE* **2008**, *3*, e1385. [[CrossRef](#)] [[PubMed](#)]
67. Ferguson, L.; Giza, C.C.; Serpa, R.O.; Greco, T.; Robert, H.; Folkerts, M.; Prins, M.L. Sex Differences in Neurophysiological Changes Following Voluntary Exercise in Adolescent Rats. *Front. Neurol.* **2021**, *12*, 685822. [[CrossRef](#)] [[PubMed](#)]
68. Yang, X.; Brobst, D.; Chan, W.S.; Tse, M.C.L.; Herlea-Pana, O.; Ahuja, P.; Bi, X.; Zaw, A.M.; Kwong, Z.S.W.; Jia, W.-H.; et al. Muscle-generated BDNF is a sexually dimorphic myokine that controls metabolic flexibility. *Sci. Signal.* **2019**, *12*, eaau1468. [[CrossRef](#)]
69. Jia, W.-H.; Wang, N.-Q.; Yin, L.; Chen, X.; Hou, B.-Y.; Qiang, G.-F.; Chan, C.B.; Yang, X.-Y.; Du, G.-H. Effect of skeletal muscle phenotype and gender on fasting-induced myokine expression in mice. *Biochem. Biophys. Res. Commun.* **2019**, *514*, 407–414. [[CrossRef](#)]

70. Molinero, A.; Fernandez-Perez, A.; Mogas, A.; Giral, M.; Comes, G.; Fernandez-Gayol, O.; Vallejo, M.; Hidalgo, J. Role of muscle IL-6 in gender-specific metabolism in mice. *PLoS ONE* **2017**, *12*, e0173675. [[CrossRef](#)] [[PubMed](#)]
71. Gentry, B.A.; Bs, J.A.F.; Phillips, C.; Brown, M. Hindlimb skeletal muscle function in myostatin-deficient mice. *Muscle Nerve* **2010**, *43*, 49–57. [[CrossRef](#)] [[PubMed](#)]
72. Oldham, J.M.; Osepchook, C.C.; Jeanplong, F.; Falconer, S.J.; Matthews, K.G.; Conaglen, J.V.; Gerrard, D.F.; Smith, H.K.; Wilkins, R.J.; Bass, J.J.; et al. The decrease in mature myostatin protein in male skeletal muscle is developmentally regulated by growth hormone. *J. Physiol.* **2009**, *587*, 669–677. [[CrossRef](#)]
73. Paul, R.G.; Hennebry, A.S.; Elston, M.S.; Conaglen, J.V.; McMahon, C.D. Regulation of murine skeletal muscle growth by STAT5B is age- and sex-specific. *Skelet. Muscle* **2019**, *9*, 1–13. [[CrossRef](#)]
74. Jia, W.-H.; Wang, N.-Q.; Yin, L.; Chen, X.; Hou, B.-Y.; Wang, J.-H.; Qiang, G.-F.; Chan, C.B.; Yang, X.-Y.; Du, G.-H. Effects of fasting on the expression pattern of FGFs in different skeletal muscle fibre types and sexes in mice. *Biol. Sex Differ.* **2020**, *11*, 1–13. [[CrossRef](#)] [[PubMed](#)]
75. Bredewold, R.; Veenema, A.H. Sex differences in the regulation of social and anxiety-related behaviors: Insights from vasopressin and oxytocin brain systems. *Curr. Opin. Neurobiol.* **2018**, *49*, 132–140. [[CrossRef](#)]
76. Adamo, S.; Pigna, E.; Lugarà, R.; Moresi, V.; Coletti, D.; Bouché, M. Skeletal Muscle: A Significant Novel Neurohypophyseal Hormone-Secreting Organ. *Front. Physiol.* **2019**, *9*, 1885. [[CrossRef](#)] [[PubMed](#)]
77. Costa, A.; Rossi, E.; Scicchitano, B.M.; Coletti, D.; Moresi, V.; Adamo, S. Neurohypophyseal hormones: Novel actors of striated muscle development and homeostasis. *Eur. J. Transl. Myol.* **2014**, *24*, 3790. [[CrossRef](#)]
78. Naro, F.; De Arcangelis, V.; Coletti, D.; Molinaro, M.; Zani, B.; Vassanelli, S.; Reggiani, C.; Teti, A.; Adamo, S. Increase in cytosolic Ca²⁺ induced by elevation of extracellular Ca²⁺ in skeletal myogenic cells. *Am. J. Physiol. Physiol.* **2003**, *284*, C969–C976. [[CrossRef](#)]
79. Alvisi, M.; De Arcangelis, V.; Ciccone, L.; Palombi, V.; Alessandrini, M.; Nemoz, G.; Molinaro, M.; Adamo, S.; Naro, F. V1a vasopressin receptor expression is modulated during myogenic differentiation. *Differentiation* **2008**, *76*, 371–380. [[CrossRef](#)] [[PubMed](#)]
80. Herbst, K.; Bhasin, S. Testosterone action on skeletal muscle. *Curr. Opin. Clin. Nutr. Metab. Care* **2004**, *7*, 271–277. [[CrossRef](#)] [[PubMed](#)]
81. Velders, M.; Diel, P. How Sex Hormones Promote Skeletal Muscle Regeneration. *Sport Med.* **2013**, *43*, 1089–1100. [[CrossRef](#)]
82. Alexander, S.E.; Pollock, A.C.; Lamon, S. The effect of sex hormones on skeletal muscle adaptation in females. *Eur. J. Sport Sci.* **2021**, *22*, 1035–1045. [[CrossRef](#)] [[PubMed](#)]
83. McMillin, S.L.; Minchew, E.C.; Lowe, D.A.; Spangenburg, E.E. Skeletal muscle wasting: The estrogen side of sexual dimorphism. *Am. J. Physiol. Physiol.* **2022**, *322*, C24–C37. [[CrossRef](#)]
84. Javed, A.A.; Mayhew, A.J.; Shea, A.K.; Raina, P. Association Between Hormone Therapy and Muscle Mass in Postmenopausal Women. *JAMA Netw. Open* **2019**, *2*, e1910154. [[CrossRef](#)]
85. Fuxjager, M.J.; Miles, M.C.; Schlinger, B.A. Evolution of the androgen-induced male phenotype. *J. Comp. Physiol. A* **2017**, *204*, 81–92. [[CrossRef](#)] [[PubMed](#)]
86. Handelsman, D.J.; Hirschberg, A.L.; Bermon, S. Circulating Testosterone as the Hormonal Basis of Sex Differences in Athletic Performance. *Endocr. Rev.* **2018**, *39*, 803–829. [[CrossRef](#)]
87. Tarnopolsky, M.A. Sex Differences in Exercise Metabolism and the Role of 17-Beta Estradiol. *Med. Sci. Sports Exerc.* **2008**, *40*, 648–654. [[CrossRef](#)]
88. Lindegaard, B.; Abildgaard, J.; Heywood, S.E.; Pedersen, B.K.; Febbraio, M.A. Female sex hormones are necessary for the metabolic effects mediated by loss of Interleukin 18 signaling. *Mol. Metab.* **2018**, *12*, 89–97. [[CrossRef](#)]
89. Brooks, N.E.; Myburgh, K.H. Skeletal muscle wasting with disuse atrophy is multi-dimensional: The response and interaction of myonuclei, satellite cells and signaling pathways. *Front. Physiol.* **2014**, *5*, 99. [[CrossRef](#)]
90. Lee, P.H.U.; Chung, M.; Ren, Z.; Mair, D.B.; Kim, D.-H. Factors mediating spaceflight-induced skeletal muscle atrophy. *Am. J. Physiol. Physiol.* **2022**, *322*, C567–C580. [[CrossRef](#)]
91. Lipes, J.; Mardini, L.; Jayaraman, D. Sex and Mortality of Hospitalized Adults After Admission to an Intensive Care Unit. *Am. J. Crit. Care* **2013**, *22*, 314–319. [[CrossRef](#)]
92. Yasuda, N.; Glover, E.I.; Phillips, S.; Isfort, R.J.; Tarnopolsky, M.A. Sex-based differences in skeletal muscle function and morphology with short-term limb immobilization. *J. Appl. Physiol.* **2005**, *99*, 1085–1092. [[CrossRef](#)]
93. Miles, M.P.; Heil, D.P.; Larson, K.R.; Conant, S.B.; Schneider, S.M. Prior Resistance Training and Sex Influence Muscle Responses to Arm Suspension. *Med. Sci. Sport Exerc.* **2005**, *37*, 1983–1989. [[CrossRef](#)]
94. Shaffer, N.C.; Huang, Y.; Abraham, D.S.; Cheng, Y.; Lu, W.; Gruber-Baldini, A.L.; Hochberg, M.C.; Guralnik, J.; Magaziner, J.; Orwig, D. Comparing Longitudinal Sarcopenia Trends by Definitions across Men and Women after Hip Fracture. *J. Am. Geriatr. Soc.* **2020**, *68*, 1537–1544. [[CrossRef](#)] [[PubMed](#)]
95. Shim, D.-G.; Kwon, T.-Y.; Lee, K.-B. Rectus femoris muscle atrophy and recovery caused by preoperative pretibial traction in femoral shaft fractures-comparison between traction period. *Orthop. Traumatol. Surg. Res.* **2017**, *103*, 691–695. [[CrossRef](#)]
96. Clark, B.C.; Manini, T.M.; Hoffman, R.L.; Russ, D.W. Restoration of Voluntary Muscle Strength after 3 Weeks of Cast Immobilization is Suppressed in Women Compared with Men. *Arch. Phys. Med. Rehabil.* **2009**, *90*, 178–180. [[CrossRef](#)]
97. Stroud, J.E.; Gale, M.S.; Zwart, S.R.; Heer, M.; Smith, S.M.; Montana, T.; Metz, G.A.S. Longitudinal metabolomic profiles reveal sex-specific adjustments to long-duration spaceflight and return to Earth. *Cell. Mol. Life Sci.* **2022**, *79*, 1–18. [[CrossRef](#)]

98. Callahan, D.M.; Tourville, T.W.; Miller, M.S.; Hackett, S.B.; Sharma, H.; Cruickshank, N.C.; Slauterbeck, J.R.; Savage, P.D.; Ades, P.A.; Maughan, D.W.; et al. Chronic disuse and skeletal muscle structure in older adults: Sex-specific differences and relationships to contractile function. *Am. J. Physiol. Physiol.* **2015**, *308*, C932–C943. [[CrossRef](#)]
99. Callahan, D.M.; Miller, M.S.; Sweeny, A.P.; Tourville, T.W.; Slauterbeck, J.R.; Savage, P.D.; Maughan, D.W.; Ades, P.A.; Beynonn, B.D.; Toth, M.J. Muscle disuse alters skeletal muscle contractile function at the molecular and cellular levels in older adult humans in a sex-specific manner. *J. Physiol.* **2014**, *592*, 4555–4573. [[CrossRef](#)] [[PubMed](#)]
100. Romanello, V.; Guadagnin, E.; Gomes, L.; Roder, I.; Sandri, C.; Petersen, Y.; Milan, G.; Masiero, E.; Del Piccolo, P.; Foretz, M.; et al. Mitochondrial fission and remodelling contributes to muscle atrophy. *EMBO J.* **2010**, *29*, 1774–1785. [[CrossRef](#)]
101. Carraro, U.; Coletti, D.; Kern, H. The EJtm Specials “The Long-Term Denervated Muscle”. *Eur. J. Transl. Myol.* **2014**, *24*. [[CrossRef](#)]
102. Oga, S.; Goto, K.; Sakamoto, J.; Honda, Y.; Sasaki, R.; Ms, K.I.; Kataoka, H.; Nakano, J.; Origuchi, T.; Okita, M. Mechanisms underlying immobilization-induced muscle pain in rats. *Muscle Nerve* **2020**, *61*, 662–670. [[CrossRef](#)] [[PubMed](#)]
103. Leermakers, P.A.; Kneppers, A.E.M.; Schols, A.M.W.J.; Kelders, M.C.J.M.; De Theije, C.C.; Verdijk, L.B.; van Loon, L.J.C.; Langen, R.C.J.; Gosker, H.R. Skeletal muscle unloading results in increased mitophagy and decreased mitochondrial biogenesis regulation. *Muscle Nerve* **2019**, *60*, 769–778. [[CrossRef](#)] [[PubMed](#)]
104. Meyer, G.A.; Thomopoulos, S.; Abu-Amer, Y.; Shen, K.C. Tenotomy-induced muscle atrophy is sex-specific and independent of NFκB. *eLife* **2022**, *11*, e82016. [[CrossRef](#)]
105. Deguise, M.-O.; De Repentigny, Y.; Tierney, A.; Beauvais, A.; Michaud, J.; Chehade, L.; Thabet, M.; Paul, B.; Reilly, A.; Gagnon, S.; et al. Motor transmission defects with sex differences in a new mouse model of mild spinal muscular atrophy. *Ebiomedicine* **2020**, *55*, 102750. [[CrossRef](#)]
106. Renzini, A.; Pigna, E.; Rocchi, M.; Cedola, A.; Gigli, G.; Moresi, V.; Coletti, D. Sex and HDAC4 Differently Affect the Pathophysiology of Amyotrophic Lateral Sclerosis in SOD1-G93A Mice. *Int. J. Mol. Sci.* **2022**, *24*, 98. [[CrossRef](#)]
107. Furukawa, T.; Nakao, K.; Sugita, H.; Tsukagoshi, H. Kugelberg-Welander Disease: With Particular Reference to Sex-Influenced Manifestations. *Arch. Neurol.* **1968**, *19*, 156–162. [[CrossRef](#)]
108. Tobias, M.; Marin, M.; Kelley, D. The roles of sex, innervation, and androgen in laryngeal muscle of *Xenopus laevis*. *J. Neurosci.* **1993**, *13*, 324–333. [[CrossRef](#)]
109. Kume-Kick, J.; Strand, F.L. Sex Hormones Affect Muscle Contractility and Motor Functional Recovery Following Peroneal Nerve Crush. *Exp. Neurol.* **1994**, *128*, 115–123. [[CrossRef](#)]
110. Musacchia, X.J.; Steffen, J.M.; Fell, R.D. Disuse atrophy of skeletal muscle: Animal models. *Exerc. Sport Sci. Rev.* **1988**, *16*, 61–88. [[CrossRef](#)]
111. Coletti, D.; Aulino, P.; Pigna, E.; Barteri, F.; Moresi, V.; Annibali, D.; Adamo, S.; Berardi, E. Spontaneous Physical Activity Downregulates Pax7 in Cancer Cachexia. *Stem Cells Int.* **2015**, *2016*, 1–9. [[CrossRef](#)]
112. He, W.A.; Berardi, E.; Cardillo, V.M.; Acharyya, S.; Aulino, P.; Thomas-Ahner, J.; Wang, J.; Bloomston, M.; Muscarella, P.; Nau, P.; et al. NF-κB mediated Pax7 dysregulation in the muscle microenvironment promotes cancer cachexia. *J. Clin. Investig.* **2013**, *123*, 4821–4835. [[CrossRef](#)] [[PubMed](#)]
113. Yoshihara, T.; Natsume, T.; Tsuzuki, T.; Chang, S.-W.; Kakigi, R.; Sugiura, T.; Naito, H. Sex differences in forkhead box O3a signaling response to hindlimb unloading in rat soleus muscle. *J. Physiol. Sci.* **2018**, *69*, 235–244. [[CrossRef](#)] [[PubMed](#)]
114. Trevino, M.B.; Zhang, X.; Standley, R.A.; Wang, M.; Han, X.; dos Reis, F.C.G.; Periasamy, M.; Yu, G.; Kelly, D.P.; Goodpaster, B.H.; et al. Loss of mitochondrial energetics is associated with poor recovery of muscle function but not mass following disuse atrophy. *Am. J. Physiol. Metab.* **2019**, *317*, E899–E910. [[CrossRef](#)] [[PubMed](#)]
115. Rosa-Caldwell, M.E.; Lim, S.; Haynie, W.S.; Jansen, L.T.; Westervelt, L.C.; Amos, M.G.; Washington, T.A.; Greene, N.P. Altering aspects of mitochondrial quality to improve musculoskeletal outcomes in disuse atrophy. *J. Appl. Physiol.* **2020**, *129*, 1290–1303. [[CrossRef](#)] [[PubMed](#)]
116. Rosa-Caldwell, M.E.; Lim, S.; Haynie, W.S.; Brown, J.L.; Lee, D.E.; Dunlap, K.R.; Jansen, L.T.; Washington, T.A.; Wiggs, M.P.; Greene, N.P. Mitochondrial aberrations during the progression of disuse atrophy differentially affect male and female mice. *J. Cachex-Sarcopenia Muscle* **2021**, *12*, 2056–2068. [[CrossRef](#)] [[PubMed](#)]
117. Powers, S.K.; Kavazis, A.N.; DeRuisseau, K.C. Mechanisms of disuse muscle atrophy: Role of oxidative stress. *Am. J. Physiol. Integr. Comp. Physiol.* **2005**, *288*, R337–R344. [[CrossRef](#)]
118. Bauer, J.; Morley, J.E.; Schols, A.M.W.J.; Ferrucci, L.; Cruz-Jentoft, A.J.; Dent, E.; Baracos, V.E.; Crawford, J.A.; Doehner, W.; Heymsfield, S.B.; et al. Sarcopenia: A Time for Action. An SCWD Position Paper. *J. Cachexia Sarcopenia Muscle* **2019**, *10*, 956–961. [[CrossRef](#)] [[PubMed](#)]
119. Rosenberg, I.H. Sarcopenia: Origins and clinical relevance. *J. Nutr.* **1997**, *127* (Suppl. S5), 990S–991S. [[CrossRef](#)] [[PubMed](#)]
120. Cruz-Jentoft, A.J.; Baeyens, J.P.; Bauer, J.M.; Boirie, Y.; Cederholm, T.; Landi, F.; Martin, F.C.; Michel, J.-P.; Rolland, Y.; Schneider, S.M.; et al. Sarcopenia: European consensus on definition and diagnosis: Report of the European Working Group on Sarcopenia in Older People. *Age Ageing* **2010**, *39*, 412–423. [[CrossRef](#)] [[PubMed](#)]
121. Cruz-Jentoft, A.J.; Bahat, G.; Bauer, J.; Boirie, Y.; Bruyère, O.; Cederholm, T.; Cooper, C.; Landi, F.; Rolland, Y.; Sayer, A.A.; et al. Sarcopenia: Revised European consensus on definition and diagnosis. *Age Ageing* **2019**, *48*, 16–31. [[CrossRef](#)]
122. Roubenoff, R.; Hughes, V.A. Sarcopenia: Current Concepts. *J. Gerontol. Ser. A* **2000**, *55*, M716–M724. [[CrossRef](#)] [[PubMed](#)]
123. Moresi, V.; Renzini, A.; Cavioli, G.; Seelaender, M.; Coletti, D.; Gigli, G.; Cedola, A. Functional Nutrients to Ameliorate Neurogenic Muscle Atrophy. *Metabolites* **2022**, *12*, 1149. [[CrossRef](#)]

124. Anderson, L.J.; Liu, H.; Garcia, J.M. Sex Differences in Muscle Wasting. *Adv. Exp. Med. Biol.* **2017**, *1043*, 153–197. [[CrossRef](#)] [[PubMed](#)]
125. Hwang, J.; Park, S. Sex Differences of Sarcopenia in an Elderly Asian Population: The Prevalence and Risk Factors. *Int. J. Environ. Res. Public Health* **2022**, *19*, 11980. [[CrossRef](#)]
126. Gallagher, D.; Ruts, E.; Visser, M.; Heshka, S.; Baumgartner, R.N.; Wang, J.; Pierson, R.N.; Pi-Sunyer, F.X.; Heymsfield, S.B. Weight stability masks sarcopenia in elderly men and women. *Am. J. Physiol.-Endocrinol. Metab.* **2000**, *279*, E366–E375. [[CrossRef](#)]
127. Kasai, T.; Ishiguro, N.; Matsui, Y.; Harada, A.; Takemura, M.; Yuki, A.; Kato, Y.; Otsuka, R.; Ando, F.; Shimokata, H. Sex- and age-related differences in mid-thigh composition and muscle quality determined by computed tomography in middle-aged and elderly Japanese. *Geriatr. Gerontol. Int.* **2014**, *15*, 700–706. [[CrossRef](#)]
128. Haynes, E.M.K.; Neubauer, N.; Cornett, K.; O'Connor, B.P.; Jones, G.R.; Jakobi, J.M. Age and sex-related decline of muscle strength across the adult lifespan: A scoping review of aggregated data. *Appl. Physiol. Nutr. Metab.* **2020**, *45*, 1185–1196. [[CrossRef](#)] [[PubMed](#)]
129. Mizuno, T.; Matsui, Y.; Tomida, M.; Suzuki, Y.; Nishita, Y.; Tange, C.; Shimokata, H.; Imagama, S.; Otsuka, R.; Arai, H. Differences in the mass and quality of the quadriceps with age and sex and their relationships with knee extension strength. *J. Cachex-Sarcopenia Muscle* **2021**, *12*, 900–912. [[CrossRef](#)] [[PubMed](#)]
130. Miller, M.S.; Bedrin, N.G.; Callahan, D.M.; Previs, M.J.; Jennings, M.E., II; Ades, P.A.; Maughan, D.W.; Palmer, B.M.; Toth, M.J. Age-related slowing of myosin actin cross-bridge kinetics is sex specific and predicts decrements in whole skeletal muscle performance in humans. *J. Appl. Physiol.* **2013**, *115*, 1004–1014. [[CrossRef](#)]
131. Coletti, C.; Acosta, G.F.; Keslacy, S.; Coletti, D. Exercise-mediated reinnervation of skeletal muscle in elderly people: An update. *Eur. J. Transl. Myol.* **2022**, *32*, 10416. [[CrossRef](#)] [[PubMed](#)]
132. Larsson, L.; Yu, F.; Höök, P.; Ramamurthy, B.; Marx, J.O.; Pircher, P. Effects of aging on regulation of muscle contraction at the motor unit, muscle cell, and molecular levels. *Int. J. Sport Nutr. Exerc. Metab.* **2001**, *11*, S28–S43. [[CrossRef](#)] [[PubMed](#)]
133. Soh, Y.; Won, C.W. Sex differences in impact of sarcopenia on falls in community-dwelling Korean older adults. *BMC Geriatr.* **2021**, *21*, 716. [[CrossRef](#)] [[PubMed](#)]
134. Waters, D.L.; Qualls, C.R.; Cesari, M.; Rolland, Y.; Vlietstra, L.; Vellas, B. Relationship of Incident Falls with Balance Deficits and Body Composition in Male and Female Community-Dwelling Elders. *J. Nutr. Health Aging* **2018**, *23*, 9–13. [[CrossRef](#)] [[PubMed](#)]
135. Rodríguez-Mañas, L.; Féart, C.; Mann, G.; Viña, J.; Chatterji, S.; Chodzko-Zajko, W.; Harmand, M.G.-C.; Bergman, H.; Carcaillon, L.; Nicholson, C.; et al. Searching for an Operational Definition of Frailty: A Delphi Method Based Consensus Statement. The Frailty Operative Definition-Consensus Conference Project. *J. Gerontol. Ser. A* **2012**, *68*, 62–67. [[CrossRef](#)]
136. Gordon, E.; Peel, N.; Samanta, M.; Theou, O.; Howlett, S.; Hubbard, R. Sex differences in frailty: A systematic review and meta-analysis. *Exp. Gerontol.* **2016**, *89*, 30–40. [[CrossRef](#)]
137. Collard, R.M.; Boter, H.; Schoevers, R.A.; Voshaar, R.C.O. Prevalence of Frailty in Community-Dwelling Older Persons: A Systematic Review. *J. Am. Geriatr. Soc.* **2012**, *60*, 1487–1492. [[CrossRef](#)]
138. Mayerl, H.; Stolz, E.; Freidl, W. Frailty and depression: Reciprocal influences or common causes? *Soc. Sci. Med.* **2020**, *263*, 113273. [[CrossRef](#)] [[PubMed](#)]
139. Curtis, E.; Litwic, A.; Cooper, C.; Dennison, E. Determinants of Muscle and Bone Aging. *J. Cell. Physiol.* **2015**, *230*, 2618–2625. [[CrossRef](#)]
140. Tay, L.; Ding, Y.Y.; Leung, B.P.; Ismail, N.H.; Yeo, A.; Yew, S.; Tay, K.S.; Tan, C.H.; Chong, M.S. Sex-specific differences in risk factors for sarcopenia amongst community-dwelling older adults. *Age* **2015**, *37*, 1–12. [[CrossRef](#)] [[PubMed](#)]
141. Paturi, S.; Gutta, A.K.; Katta, A.; Kakarla, S.K.; Arvapalli, R.K.; Gadde, M.K.; Nalabotu, S.K.; Rice, K.M.; Wu, M.; Blough, E. Effects of aging and gender on muscle mass and regulation of Akt-mTOR-p70s6k related signaling in the F344BN rat model. *Mech. Ageing Dev.* **2010**, *131*, 202–209. [[CrossRef](#)]
142. Szulc, P.; Schoppet, M.; Goettsch, C.; Rauner, M.; Dschietzig, T.B.; Chapurlat, R.; Hofbauer, L.C. Endocrine and Clinical Correlates of Myostatin Serum Concentration in Men—the STRAMBO Study. *J. Clin. Endocrinol. Metab.* **2012**, *97*, 3700–3708. [[CrossRef](#)]
143. Peng, L.-N.; Lee, W.-J.; Liu, L.-K.; Lin, M.-H.; Chen, L.-K. Healthy community-living older men differ from women in associations between myostatin levels and skeletal muscle mass. *J. Cachex-Sarcopenia Muscle* **2018**, *9*, 635–642. [[CrossRef](#)] [[PubMed](#)]
144. Ratkevicius, A.; Joyson, A.; Selmer, I.; Dhanani, T.; Grierson, C.; Tommasi, A.M.; Devries, A.; Rauchhaus, P.; Crowther, D.; Alesci, S.; et al. Serum Concentrations of Myostatin and Myostatin-Interacting Proteins Do Not Differ Between Young and Sarcopenic Elderly Men. *J. Gerontol. Ser. A* **2011**, *66*, 620–626. [[CrossRef](#)]
145. Fife, E.; Kostka, J.; Kroc, Ł.; Guligowska, A.; Pigłowska, M.; Sołtysik, B.; Kaufman-Szymczyk, A.; Fabianowska-Majewska, K.; Kostka, T. Relationship of muscle function to circulating myostatin, follistatin and GDF11 in older women and men. *BMC Geriatr.* **2018**, *18*, 200. [[CrossRef](#)]
146. Park, Y.-M.; Jankowski, C.M.; Ozemek, C.; Hildreth, K.L.; Kohrt, W.M.; Moreau, K.L. Appendicular lean mass is lower in late compared with early perimenopausal women: Potential role of FSH. *J. Appl. Physiol.* **2020**, *128*, 1373–1380. [[CrossRef](#)]
147. La Colla, A.; Pronsato, L.; Milanesi, L.; Vasconsuelo, A. 17β-Estradiol and testosterone in sarcopenia: Role of satellite cells. *Ageing Res. Rev.* **2015**, *24*, 166–177. [[CrossRef](#)]
148. Norman, K.; Haß, U.; Pirlich, M. Malnutrition in Older Adults—Recent Advances and Remaining Challenges. *Nutrients* **2021**, *13*, 2764. [[CrossRef](#)]
149. Sieber, C.C. Malnutrition and sarcopenia. *Ageing Clin. Exp. Res.* **2019**, *31*, 793–798. [[CrossRef](#)] [[PubMed](#)]

150. Liu, G.; Lu, L.; Sun, Q.; Ye, X.; Sun, L.; Liu, X.; Zong, G.; Jin, Q.; Li, H.; Lin, X. Poor Vitamin D Status Is Prospectively Associated with Greater Muscle Mass Loss in Middle-Aged and Elderly Chinese Individuals. *J. Acad. Nutr. Diet.* **2014**, *114*, 1544–1551.e2. [[CrossRef](#)] [[PubMed](#)]
151. Ko, M.J.; Yun, S.; Oh, K.; Kim, K. Relation of serum 25-hydroxyvitamin D status with skeletal muscle mass by sex and age group among Korean adults. *Br. J. Nutr.* **2015**, *114*, 1838–1844. [[CrossRef](#)] [[PubMed](#)]
152. Baracos, V.E.; Martin, L.; Korc, M.; Guttridge, D.C.; Fearon, K.C.H. Cancer-associated cachexia. *Nat. Rev. Dis. Prim.* **2018**, *4*, 17105. [[CrossRef](#)] [[PubMed](#)]
153. Fearon, K.; Strasser, F.; Anker, S.D.; Bosaeus, I.; Bruera, E.; Fainsinger, R.L.; Jatoi, A.; Loprinzi, C.; MacDonald, N.; Mantovani, G.; et al. Definition and classification of cancer cachexia: An international consensus. *Lancet Oncol.* **2011**, *12*, 489–495. [[CrossRef](#)] [[PubMed](#)]
154. Aktas, A.; Lorton, C.M.; Griffin, O.; Higgins, K.; Roulston, F.; Stewart, G.; Corkery, N.; Barnes, E.; Walsh, D. Application of the 2011 international consensus cancer cachexia classification in routine oncology dietetic practice: An observational study. *Nutr. Clin. Pract.* **2022**. [[CrossRef](#)]
155. von Haehling, S.; Anker, M.S.; Prevalence and clinical impact of cachexia in chronic illness in Europe, USA, and Japan: Facts and numbers update 2016. *J. Cachex-Sarcopenia Muscle* **2016**, *7*, 507–509. [[CrossRef](#)]
156. Takenaka, Y.; Oya, R.; Takemoto, N.; Inohara, H. Predictive impact of sarcopenia in solid cancers treated with immune checkpoint inhibitors: A meta-analysis. *J. Cachex-Sarcopenia Muscle* **2021**, *12*, 1122–1135. [[CrossRef](#)]
157. Argilés, J.M.; Busquets, S.; Stemmler, B.; López-Soriano, F.J. Cancer cachexia: Understanding the molecular basis. *Nat. Rev. Cancer* **2014**, *14*, 754–762. [[CrossRef](#)]
158. Zhong, X.; Zimmers, T.A. Sex Differences in Cancer Cachexia. *Curr. Osteoporos. Rep.* **2020**, *18*, 646–654. [[CrossRef](#)]
159. Martin, L.; Senesse, P.; Gioulbasanis, I.; Antoun, S.; Bozzetti, F.; Deans, C.; Strasser, F.; Thoresen, L.; Jagoe, R.T.; Chasen, M.; et al. Diagnostic Criteria for the Classification of Cancer-Associated Weight Loss. *J. Clin. Oncol.* **2015**, *33*, 90–99. [[CrossRef](#)]
160. Ventura-Clapier, R.; Piquereau, J.; Veksler, V.; Garnier, A. Estrogens, Estrogen Receptors Effects on Cardiac and Skeletal Muscle Mitochondria. *Front. Endocrinol.* **2019**, *10*, 557. [[CrossRef](#)]
161. Cardinale, D.A.; Larsen, F.J.; Schiffer, T.A.; Morales-Alamo, D.; Ekblom, B.; Calbet, J.A.L.; Holmberg, H.-C.; Boushel, R. Superior Intrinsic Mitochondrial Respiration in Women Than in Men. *Front. Physiol.* **2018**, *9*, 1133. [[CrossRef](#)]
162. Baracos, V.E.; Reiman, T.; Mourtzakis, M.; Gioulbasanis, I.; Antoun, S. Body composition in patients with non-small cell lung cancer: A contemporary view of cancer cachexia with the use of computed tomography image analysis. *Am. J. Clin. Nutr.* **2010**, *91*, S1133–S1137. [[CrossRef](#)]
163. Wallengren, O.; Iresjö, B.-M.; Lundholm, K.; Bosaeus, I. Loss of muscle mass in the end of life in patients with advanced cancer. *Support. Care Cancer* **2014**, *23*, 79–86. [[CrossRef](#)] [[PubMed](#)]
164. Anoveros-Barrera, A.; Bhullar, A.S.; Stretch, C.; Esfandiari, N.; Dunichand-Hoedl, A.R.; Martins, K.J.; Bigam, D.; Khadaroo, R.G.; McMullen, T.; Bathe, O.F.; et al. Clinical and biological characterization of skeletal muscle tissue biopsies of surgical cancer patients. *J. Cachex-Sarcopenia Muscle* **2019**, *10*, 1356–1377. [[CrossRef](#)]
165. Burkart, M.; Schieber, M.; Basu, S.; Shah, P.; Venugopal, P.; Borgia, J.A.; Gordon, L.; Karmali, R. Evaluation of the impact of cachexia on clinical outcomes in aggressive lymphoma. *Br. J. Haematol.* **2019**, *186*, 45–53. [[CrossRef](#)] [[PubMed](#)]
166. Greenman, A.C.; Albrecht, D.M.; Halberg, R.B.; Diffie, G.M. Sex differences in skeletal muscle alterations in a model of colorectal cancer. *Physiol. Rep.* **2020**, *8*, e14391. [[CrossRef](#)]
167. de Castro, G.S.; Simoes, E.; Lima, J.D.; Ortiz-Silva, M.; Festuccia, W.T.; Tokeshi, F.; Alcântara, P.S.; Otoch, J.P.; Coletti, D.; Seelaender, M. Human Cachexia Induces Changes in Mitochondria, Autophagy and Apoptosis in the Skeletal Muscle. *Cancers* **2019**, *11*, 1264. [[CrossRef](#)]
168. Aversa, Z.; Pin, F.; Lucia, S.; Penna, F.; Verzaro, R.; Fazi, M.; Colasante, G.; Tirone, A.; Fanelli, F.R.; Ramaccini, C.; et al. Autophagy is induced in the skeletal muscle of cachectic cancer patients. *Sci. Rep.* **2016**, *6*, 30340. [[CrossRef](#)]
169. Camargo, R.G.; Ribeiro, H.Q.T.; Geraldo, M.V.; Matos-Neto, E.; Neves, R.X.; Carnevali, L.C., Jr.; Donatto, F.F.; Alcântara, P.S.M.; Otoch, J.P.; Seelaender, M. Cancer Cachexia and MicroRNAs. *Mediat. Inflamm.* **2015**, *2015*, 1–5. [[CrossRef](#)]
170. Lautaoja, J.H.; Lalowski, M.; Nissinen, T.A.; Hentilä, J.J.; Shi, Y.; Ritvos, O.; Cheng, S.; Hulmi, J.J. Muscle and serum metabolomes are dysregulated in colon-26 tumor-bearing mice despite amelioration of cachexia with activin receptor type 2B ligand blockade. *Am. J. Physiol. Metab.* **2019**, *316*, E852–E865. [[CrossRef](#)]
171. Zhong, X.; Narasimhan, A.; Silverman, L.M.; Young, A.R.; Shahda, S.; Liu, S.; Wan, J.; Liu, Y.; Koniaris, L.G.; Zimmers, T.A. Sex specificity of pancreatic cancer cachexia phenotypes, mechanisms, and treatment in mice and humans: Role of Activin. *J. Cachex-Sarcopenia Muscle* **2022**, *13*, 2146–2161. [[CrossRef](#)] [[PubMed](#)]
172. Herpich, M.C.; Franz, M.K.; Ost, M.; Otten, L.; Coleman, V.; Klaus, S.; Müller-Werdan, U.; Norman, K. Associations Between Serum GDF15 Concentrations, Muscle Mass, and Strength Show Sex-Specific Differences in Older Hospital Patients. *Rejuvenation Res.* **2021**, *24*, 14–19. [[CrossRef](#)]
173. Molfino, A.; Amabile, M.I.; Imbimbo, G.; Rizzo, V.; Pediconi, F.; Catalano, C.; Emiliani, A.; Belli, R.; Ramaccini, C.; Parisi, C.; et al. Association between Growth Differentiation Factor-15 (GDF-15) Serum Levels, Anorexia and Low Muscle Mass among Cancer Patients. *Cancers* **2020**, *13*, 99. [[CrossRef](#)] [[PubMed](#)]

174. Alcazar, J.; Frandsen, U.; Prokhorova, T.; Kamper, R.S.; Haddock, B.; Aagaard, P.; Suetta, C. Changes in systemic GDF15 across the adult lifespan and their impact on maximal muscle power: The Copenhagen Sarcopenia Study. *J. Cachex-Sarcopenia Muscle* **2021**, *12*, 1418–1427. [[CrossRef](#)]
175. Crunkhorn, S. Blocking GDF15 signalling reverses cachexia. *Nat. Rev. Drug Discov.* **2020**, *19*, 588. [[CrossRef](#)]
176. Garcia, J.M.; Garcia-Touza, M.; Hijazi, R.A.; Taffet, G.; Epner, D.; Mann, D.; Smith, R.G.; Cunningham, G.R.; Marcelli, M. Active Ghrelin Levels and Active to Total Ghrelin Ratio in Cancer-Induced Cachexia. *J. Clin. Endocrinol. Metab.* **2005**, *90*, 2920–2926. [[CrossRef](#)] [[PubMed](#)]
177. Hetzler, K.L.; Hardee, J.P.; Puppa, M.J.; Narsale, A.A.; Sato, S.; Davis, J.M.; Carson, J.A. Sex differences in the relationship of IL-6 signaling to cancer cachexia progression. *Biochim. et Biophys. Acta (BBA)-Mol. Basis Dis.* **2015**, *1852*, 816–825. [[CrossRef](#)]
178. Bonetto, A.; Aydogdu, T.; Jin, X.; Zhang, Z.; Zhan, R.; Puzis, L.; Koniaris, L.G.; Zimmers, T.A. JAK/STAT3 pathway inhibition blocks skeletal muscle wasting downstream of IL-6 and in experimental cancer cachexia. *Am. J. Physiol. Endocrinol. Metab.* **2012**, *303*, E410–E421. [[CrossRef](#)]
179. Bonafè, M.; Olivieri, F.; Cavallone, L.; Giovagnetti, S.; Mayegiani, F.; Cardelli, M.; Pieri, C.; Marra, M.; Antonicelli, R.; Lisa, R.; et al. A gender-dependent genetic predisposition to produce high levels of IL-6 is detrimental for longevity. *Eur. J. Immunol.* **2001**, *31*, 2357–2361. [[CrossRef](#)]
180. Hetzler, K.L.; Hardee, J.P.; LaVoie, H.A.; Murphy, E.A.; Carson, J.A. Ovarian function's role during cancer cachexia progression in the female mouse. *Am. J. Physiol.-Endocrinol. Metab.* **2017**, *312*, E447–E459. [[CrossRef](#)] [[PubMed](#)]
181. Cook, M.B.; Dawsey, S.M.; Freedman, N.D.; Inskip, P.D.; Wichner, S.M.; Quraishi, S.M.; Devesa, S.S.; McGlynn, K.A. Sex Disparities in Cancer Incidence by Period and Age. *Cancer Epidemiol. Biomark. Prev.* **2009**, *18*, 1174–1182. [[CrossRef](#)]
182. Cook, M.B.; McGlynn, K.A.; Devesa, S.S.; Freedman, N.D.; Anderson, W.F. Sex Disparities in Cancer Mortality and Survival. *Cancer Epidemiol. Biomark. Prev.* **2011**, *20*, 1629–1637. [[CrossRef](#)]
183. Wilkinson, N.M.; Chen, H.-C.; Lechner, M.G.; Su, M.A. Sex Differences in Immunity. *Annu. Rev. Immunol.* **2022**, *40*, 75–94. [[CrossRef](#)]
184. Straub, R.H. The Complex Role of Estrogens in Inflammation. *Endocr. Rev.* **2007**, *28*, 521–574. [[CrossRef](#)] [[PubMed](#)]
185. Polanczyk, M.J.; Carson, B.D.; Subramanian, S.; Afentoulis, M.; Vandenbark, A.A.; Ziegler, S.F.; Offner, H. Cutting Edge: Estrogen Drives Expansion of the CD4+CD25+ Regulatory T Cell Compartment. *J. Immunol.* **2004**, *173*, 2227–2230. [[CrossRef](#)] [[PubMed](#)]
186. Teilmann, S.C.; Clement, C.A.; Thorup, J.; Byskov, A.G.; Christensen, S.T. Expression and localization of the progesterone receptor in mouse and human reproductive organs. *J. Endocrinol.* **2006**, *191*, 525–535. [[CrossRef](#)]
187. Libert, C.; Dejager, L.; Pinheiro, I. The X chromosome in immune functions: When a chromosome makes the difference. *Nat. Rev. Immunol.* **2010**, *10*, 594–604. [[CrossRef](#)]
188. Moulton, V.R. Sex Hormones in Acquired Immunity and Autoimmune Disease. *Front. Immunol.* **2018**, *9*, 2279. [[CrossRef](#)] [[PubMed](#)]
189. Bupp, M.R.G.; Jorgensen, T.N. Androgen-Induced Immunosuppression. *Front. Immunol.* **2018**, *9*, 794. [[CrossRef](#)]
190. Jenny, N.S. Inflammation in aging: Cause, effect, or both? *Discov. Med.* **2012**, *13*, 451–460. [[PubMed](#)]
191. Shen, C.-Y.; Lu, C.-H.; Wu, C.-H.; Li, K.-J.; Kuo, Y.-M.; Hsieh, S.-C.; Yu, C.-L. Molecular Basis of Accelerated Aging with Immune Dysfunction-Mediated Inflammation (Inflamm-Aging) in Patients with Systemic Sclerosis. *Cells* **2021**, *10*, 3402. [[CrossRef](#)]
192. Mutin-Carnino, M.; Carnino, A.; Roffino, S.; Chopard, A. Effect of Muscle Unloading, Reloading and Exercise on Inflammation during a Head-down Bed Rest. *Int. J. Sport. Med.* **2013**, *35*, 28–34. [[CrossRef](#)] [[PubMed](#)]
193. Jurdana, M.; Jenko-Pražnikar, Z.; Mohorko, N.; Petelin, A.; Jakus, T.; Šimunič, B.; Pišot, R. Impact of 14-day bed rest on serum adipokines and low-grade inflammation in younger and older adults. *Age* **2015**, *37*, 1–11. [[CrossRef](#)] [[PubMed](#)]
194. Strollo, F.; Vernikos, J. Aging-like metabolic and adrenal changes in microgravity: State of the art in preparation for Mars. *Neurosci. Biobehav. Rev.* **2021**, *126*, 236–242. [[CrossRef](#)]
195. Webster, J.M.; Kempen, L.J.A.P.; Hardy, R.S.; Langen, R.C.J. Inflammation and Skeletal Muscle Wasting During Cachexia. *Front. Physiol.* **2020**, *11*. [[CrossRef](#)] [[PubMed](#)]
196. Gao, Y.; Arfat, Y.; Wang, H.; Goswami, N. Muscle Atrophy Induced by Mechanical Unloading: Mechanisms and Potential Countermeasures. *Front. Physiol.* **2018**, *9*, 235. [[CrossRef](#)]
197. de Castro, G.S.; Correia-Lima, J.; Simoes, E.; Orsso, C.E.; Xiao, J.; Gama, L.R.; Gomes, S.P.; Gonçalves, D.C.; Costa, R.G.F.; Radloff, K.; et al. Myokines in treatment-naïve patients with cancer-associated cachexia. *Clin. Nutr.* **2020**, *40*, 2443–2455. [[CrossRef](#)]
198. Shang, M.; Cappellesso, F.; Amorim, R.; Serneels, J.; Virga, F.; Eelen, G.; Carobbio, S.; Rincon, M.Y.; Maechler, P.; De Bock, K.; et al. Macrophage-derived glutamine boosts satellite cells and muscle regeneration. *Nature* **2020**, *587*, 626–631. [[CrossRef](#)]
199. Rizzo, G.; Di Maggio, R.; Benedetti, A.; Morroni, J.; Bouche, M.; Lozanoska-Ochser, B. Splenic Ly6Chi monocytes are critical players in dystrophic muscle injury and repair. *J. Clin. Investig.* **2020**, *5*, e130807. [[CrossRef](#)]
200. Lozanoska-Ochser, B.; Benedetti, A.; Rizzo, G.; Marrocco, V.; Di Maggio, R.; Fiore, P.; Bouche, M. Targeting early PKC θ -dependent T-cell infiltration of dystrophic muscle reduces disease severity in a mouse model of muscular dystrophy. *J. Pathol.* **2017**, *244*, 323–333. [[CrossRef](#)]
201. Berardi, E.; Aulino, P.; Murfuni, I.; Toschi, A.; Padula, F.; Scicchitano, B.M.; Coletti, D.; Adamo, S. Skeletal muscle is enriched in hematopoietic stem cells and not inflammatory cells in cachectic mice. *Neurol. Res.* **2008**, *30*, 160–169. [[CrossRef](#)] [[PubMed](#)]
202. Coletti, D.; Moresi, V.; Adamo, S.; Molinaro, M.; Sassoon, D. Tumor necrosis factor- α gene transfer induces cachexia and inhibits muscle regeneration. *Genes* **2005**, *43*, 120–128. [[CrossRef](#)] [[PubMed](#)]

203. Moresi, V.; Garcia-Alvarez, G.; Pristerà, A.; Rizzuto, E.; Albertini, M.C.; Rocchi, M.; Marazzi, G.; Sassoon, D.; Adamo, S.; Coletti, D. Modulation of Caspase Activity Regulates Skeletal Muscle Regeneration and Function in Response to Vasopressin and Tumor Necrosis Factor. *PLoS ONE* **2009**, *4*, e5570. [[CrossRef](#)] [[PubMed](#)]
204. Garcia-Sifuentes, Y.; Maney, D.L. Reporting and misreporting of sex differences in the biological sciences. *eLife* **2021**, *10*, e70817. [[CrossRef](#)]
205. Torregrosa, C.; Chorin, F.; Beltran, E.E.M.; Neuzillet, C.; Cardot-Ruffino, V. Physical Activity as the Best Supportive Care in Cancer: The Clinician's and the Researcher's Perspectives. *Cancers* **2022**, *14*, 5402. [[CrossRef](#)]
206. Grande, A.J.; Silva, V.; Neto, L.S.; Basmage, J.P.T.; Peccin, M.S.; Maddocks, M. Exercise for cancer cachexia in adults. *Cochrane Database Syst. Rev.* **2021**, 2021, CD010804. [[CrossRef](#)]
207. Besson, T.; Macchi, R.; Rossi, J.; Morio, C.Y.M.; Kunimasa, Y.; Nicol, C.; Vercruyssen, F.; Millet, G.Y. Sex Differences in Endurance Running. *Sport. Med.* **2022**, *52*, 1235–1257. [[CrossRef](#)]
208. Roberts, B.M.; Nuckols, G.; Krieger, J.W. Sex Differences in Resistance Training: A Systematic Review and Meta-Analysis. *J. Strength Cond. Res.* **2020**, *34*, 1448–1460. [[CrossRef](#)]
209. Kornstein, S. Exploring the Biological Contributions to Human Health: Does Sex Matter? *J. Women's Health Gender-Based Med.* **2001**, *10*, 433–439. [[CrossRef](#)]
210. Rich-Edwards, J.W.; Kaiser, U.B.; Chen, G.L.; Manson, J.E.; Goldstein, J.M. Sex and Gender Differences Research Design for Basic, Clinical, and Population Studies: Essentials for Investigators. *Endocr. Rev.* **2018**, *39*, 424–439. [[CrossRef](#)]
211. Winkelman, C. Inactivity and Inflammation in the Critically Ill Patient. *Crit. Care Clin.* **2007**, *23*, 21–34. [[CrossRef](#)] [[PubMed](#)]
212. Lu, C.Y.; Terry, V.; Thomas, D.M. Precision medicine: Affording the successes of science. *Npj Precis. Oncol.* **2023**, *7*, 3. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.