Sex Steroid Actions in Male Bone

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Sex steroids are chief regulators of gender differences in the skeleton, and male gender is one of the strongest protective factors against osteoporotic fractures. This advantage in bone strength relies mainly on greater cortical bone expansion during pubertal peak bone mass acquisition and superior skeletal maintenance during aging. During both these phases, estrogens acting via estrogen receptor- α in osteoblast lineage cells are crucial for male cortical and trabecular bone, as evident from conditional genetic mouse models, epidemiological studies, rare genetic conditions, genome-wide meta-analyses, and recent interventional trials. Genetic mouse models have also demonstrated a direct role for androgens independent of aromatization on trabecular bone via the androgen receptor in osteoblasts and osteocytes, although the target cell for their key effects on periosteal bone formation remains elusive. Low serum estradiol predicts incident fractures, but the highest risk occurs in men with additionally low T and high SHBG. Still, the possible clinical utility of serum sex steroids for fracture prediction is unknown. It is likely that sex steroid actions on male bone metabolism rely also on extraskeletal mechanisms and cross talk with other signaling pathways. We propose that estrogens influence fracture risk in aging men via direct effects on bone, whereas androgens exert an additional antifracture effect mainly via extraskeletal parameters such as muscle mass and propensity to fall. Given the demographic trends of increased longevity and consequent rise of osteoporosis, an increased understanding of how sex steroids influence male bone health remains a high research priority. (Endocrine Reviews 35: 906-960, 2014)

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Abbreviations: AF-1, activation function 1; AF-2, activation function 2; AIS, androgen insensitivity syndrome; AP-1, activating protein-1; AR, androgen receptor; ARE, androgen response element; ARKO, AR knockout; BMD, bone mineral density; BMP, bone morphogenetic protein; ChIP, chromatin immunoprecipitation; COMT, catechol-O-methyltransferase; Ctsk, cathepsin K; DBD, DNA-binding domain; DHEA, dehydroepiandrosterone; DHT, dihydrotestosterone; Dmp1, dentin matrix protein 1; DXA, dual-energy x-ray absorptiometry; E2, 17 \$\beta\$-estractiol; EDC, E2 dendrimer conjugate; ER, estrogen receptor; ERE, estrogen response element; ERKO, estrogen-receptor knockout; Fast, Fas ligand; GPR30, G protein-coupled receptor 30; GWAS, genome-wide association studies; \$hpg\$, hypogonadal (mice); HRpQCT, high-resolution pQCT; IGFBP, IGF binding protein; LBD, ligand-binding domain; LRP, low-density lipoprotein receptor-related protein; LySM, lysozyme M; NERKI, nonclassical ER knock-in; NTD, N-terminal domain; OPG, osteoprotegerin; pQCT, peripheral QCT; QCT, quantitative computed tomography; RANK, receptor activator of nuclear factor \$\pi\$ RANKL, RANK ligand; ROS, reactive oxygen species; RUNX2, Runt-related transcription factor 2; SARM, selective AR modulator; SERM, selective ER modulator; tau, transcriptional activation unit; Tfm, testicular feminization; vBMD, volumetric BMD.

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I. Introduction

steoporosis in men, although less common than in women, still constitutes a major burden for public health (1, 2). Despite the higher competing risk of mortality from other causes in older men, the remaining lifetime risk of osteoporotic fractures after age 50 may be as high as 20–25% (vs 45–55% in women) in high-risk Caucasian populations (3–5). For hip fractures, a recent systematic review found that the male-female incidence ratio was also about 1:2 and was remarkably constant globally, despite greater than 10-fold variation between geographic regions (Figure 1) (6). Accordingly, men account for around one-third of fractures and associated expenditures (7, 8). Of the estimated 9 million incident osteoporotic fractures worldwide in the year 2000, men accounted for 30% of fractures at the hip, 39% at the spine, 25% at the humerus, 20% at the forearm, and 54% at other sites (7).

Fracture incidence increases exponentially around the age of menopause in women. In older men, the fracture rate rises in parallel, but with a delay of about 15 years (9, 10). Excess mortality endures for up to 10 years after hip fracture but more so in men, due to underlying frailty and comorbidities that appear characteristic of those men who do develop osteoporosis (11, 12). This is in line with the observation that the absolute risk of a second fracture is equal in both genders (ie, men have a lower risk of developing osteoporosis, but when they do so, they have a higher relative refracture risk) (13). Long-term care and institutionalization rates after fracture also seem higher in elderly men, and this greatly adds to the economic and societal burden of osteoporosis (14). Still, the general public, male patients, and many physicians perceive osteoporosis as a female disease. This contributes to underdiagnosis and undertreatment, which is considerable in both genders, but even more so in men (15–18). Recently, Boonen et al (19) provided formal evidence in a randomized controlled trial that this therapeutic inertia is unjustified because fracture risk in male osteoporosis can be reduced with similar effectiveness as that shown in women.

Thus, apart from younger age, male gender by itself is one of the strongest protective factors that approximately halves the risk of osteoporosis. As early as the 1940s, gender differences in fracture risk were attributed to the effects of estrogens in women and androgens in men. By the end of the 1990s, sex steroids, and especially estrogens, were deemed central mediators involved not only in postmenopausal osteoporosis but also in the slow, continued bone loss in older age (so-called involutional or type II osteoporosis), in male osteoporosis, and even in agerelated secondary hyperparathyroidism (20, 21). However, this unitary model centered on estrogen deficiency has been modified with the recognition of generic aging mechanisms and other master regulator pathways involved in bone turnover (eg, Wnt/sclerostin, receptor activator of nuclear factor kappa B [RANK]/RANK ligand [RANKL]/ osteoprotegerin [OPG], Runt-related transcription factor 2 [RUNX2], IGF-1, etc) (22–24). These pathways probably control bone metabolism in both genders (25, 26), and although many of them directly interact with sex steroid signaling, they have not been demonstrated as intrinsically sexually dimorphic (see also Section VII). Androgens and estrogens are therefore still considered the chief regulators of gender differences in bone metabolism.

The aim of this review is to critically summarize advances in our understanding of sex steroid actions on the male skeleton, and whether and how this contrasts to the situation in women. We will emphasize progress made since our previous review a decade ago (27). In *Sections II–VI*, we will discuss sex steroid physiology in relation to bone and how this regulates male bone strength and fracture risk at the organ, cellular, and genetic level during peak bone mass acquisition and subsequent maintenance. The clinical approach to the diagnosis and treatment of osteoporosis in men has been reviewed elsewhere (1, 28, 29) and will not be covered here.

In Section VIII, it will become clear that attention in this field has extended beyond the skeleton: sex steroids affect all aspects of body composition, which is overall more robust in men. Not only are men almost 10% taller than women, with 25% greater peak bone mineral content on average (1, 21), but also muscle mass and power are almost 50% higher and body fat percentage nearly 50% lower in young adult men compared to women (30, 31). In turn, interactions between bone, muscle, and fat are in-

Figure 1.

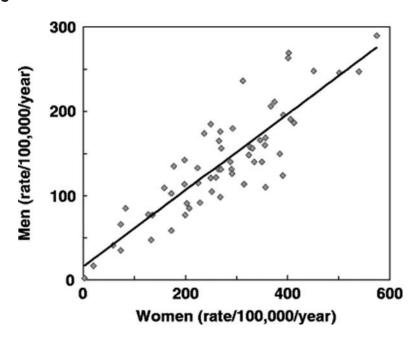


Figure 1. In a recent systematic review, age-standardized hip fracture rates (/100 000/y) in men were approximately half those in women (correlation: r=0.82; P<.001). Thus, in countries where higher rates were observed in women, higher rates were also found in men and vice versa. [Reproduced from J. A. Kanis et al: A systematic review of hip fracture incidence and probability of fracture worldwide. *Osteoporos Int.* 2012;23:2239–2256 (6), with permission. © International Osteoporosis Foundation and National Osteoporosis Foundation.]

creasingly studied. In old age, decreasing bioavailable androgen and estrogen levels contribute to detrimental changes in these body composition domains (32–34). In that sense, sex steroid signaling may be considered an attractive drug target in the quest for shifting mesenchymal precursor cells toward osteogenic and myogenic differentiation and away from adipogenesis (see also Section V and Figure 5) (35). Unraveling these pathways may inform the design of better, preferentially gender-neutral drugs that not only inhibit bone resorption or stimulate bone formation, but also reinforce the musculoskeletal system as a whole, prevent falls, and decrease adiposity and its consequences on glucose and lipid metabolism. Given the demographic trends of increased longevity and consequent rise of osteoporosis, sarcopenia, and obesity in many societies worldwide, this is a high research priority.

II. Sex Steroid Physiology in Relation to Bone

A. Circulating sex steroids and SHBG

Androgens and estrogens are respectively C19 and C18 metabolites of cholesterol. The predominant gonadal androgen in men is T, 95% of which is secreted by the testes. The remaining 5% is produced by the adrenals after conversion of the precursor dehydroepiandrosterone

(DHEA). In peripheral tissues, T can be converted by 5α -reductase enzymes into the more potent androgen dihydrotestosterone (DHT). T can also be converted into 17β estradiol (E2) by the aromatase (CYP19A1) enzyme. Approximately 20% of circulating E2 in men is made by the testes; the remaining 80% is derived from DHEA in peripheral tissues (36, 37). Therefore, the actions of T can be mediated via T directly or DHT via the androgen receptor (AR), or T can be converted into E2 and act via the estrogen receptor (ER) α or β .

SHBG binds 50–60% of circulating T and E2 with high affinity, whereas 40–50% is bound nonspecifically to albumin and other proteins. Only 1–3% of the active sex steroids are unbound, and this is termed the free fraction (36). The free and non-SHBG-bound fractions together are referred to as the bioavailable sex steroid fraction. These fractions are often calculated using

law-of-mass-action equations, based on the concentrations of total T or E2, SHBG, albumin, and fixed association constants. Several studies have demonstrated the limitations of these formulas, which tend to overestimate the free and/or bioavailable levels relative to the laboratory measurements, especially in the case of T (38–41). However, because direct measurements of free and bioavailable sex steroid concentrations are laborious and require considerable expertise, they are reserved for research purposes but not considered accurate enough in the clinical setting (42).

The circulating levels of total T and E2 decrease only marginally with age in men. However, the age-related increase in SHBG is more pronounced, resulting in a greater decrease of bioavailable or free sex steroid levels (43, 44). Importantly, older men generally have higher circulating E2 levels than postmenopausal women (45), making it plausible that E2 contributes to the conservation of the male skeleton and/or other tissues during aging.

Serum sex steroid levels are routinely measured by immunoassays, but due to limited specificity, especially at low E2 concentrations, mass spectrometry has been recommended (46–48). This is an important consideration when interpreting the results of both human and rodent studies using immunoassays for measurement of sex ste-

roid levels (see *Section II.E* for discussion of circulating sex steroids in rodents). Nevertheless, serum E2 levels measured either by a sensitive immunoassay or by mass spectrometry provide almost identical correlations with bone mineral density (BMD) in men (48, 49).

B. Local sex steroid metabolism in bone tissue

Not only sex steroid receptors, but also all the necessary enzymes to convert the adrenal sex steroid precursors DHEA and androstenedione into active androgens and estrogens, are expressed in bone tissue and osteoblast cell lines (27), supporting the notion of local skeletal paracrine and/or intracrine synthesis and action of sex steroids. In comparison, little is known about sex steroid-metabolizing enzymes in osteocytes and osteoclasts.

Transgenic male mice overexpressing human aromatase have an increased bone mass, but other, adverse systemic effects were also observed due to elevated circulating E2 levels (50). In contrast, aromatase overexpression specifically in osteoblasts increases bone mass without elevating serum E2 levels or systemic effects (51). Male mice genetically ablated of 5α -reductase type 1 have slightly reduced bone mass (52), although the relevance of this finding to human physiology remains to be established (see *Section IX.B*).

Androgens and estrogens are also inactivated locally, and the resulting metabolites can be measured in serum and urine. More specifically, androgens are glucuronidated by uridine glucuronosyl transferases, and this process can regulate the intracellular levels of active androgens and their local biological activity. Circulating levels

Figure 2.

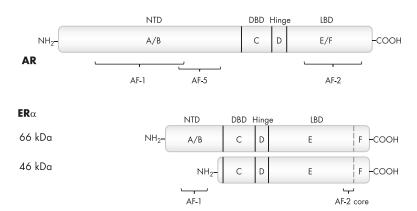


Figure 2. Diagram of the structure of AR and $ER\alpha$ with their different domains: the NTD (A/B domain), the C-domain containing the DBD, the D-domain containing the hinge region, and the C-terminal E- and F-domains containing the LBD. The AR A/B-domain contains two surfaces for interactions with coactivators AF-1 and AF-5 (also called transcriptional activation unit, tau-1 and tau-5) (60). The LBD contains AF-2, which is involved in N-terminal/C-terminal interactions and binding of coregulatory proteins, but it has weak activating capacity in AR. $ER\alpha$ also contains an AF-1 and AF-2 and has two isoforms: the full-length 66-kDa protein and a less abundant 46-kDa isoform lacking AF-1.

of these specific glucuronidated androgen metabolites showed closer associations with BMD at several sites in older men than the serum levels of the active androgens (53). Polymorphism in the gene for catechol-O-methyltransferase (COMT; an estrogen-degrading enzyme) have also been associated with peak bone mass development (54, 55) as well as prevalent fractures in elderly men (56, 57).

In conclusion, the concept of intraskeletal sex steroid metabolism limits the interpretation of circulating levels of androgens and estrogens because they might not always reflect androgen and estrogen bioactivity within bone. Future studies should assess sex steroid levels not only in serum but also in the tissue of interest, although better methods to measure intraskeletal sex steroids remain to be established (58).

C. Classical androgen and estrogen receptor signaling

Androgens and estrogens can be defined as high-affinity ligands for AR or ER α /ER β , which are ligand-inducible transcription factors of the nuclear receptor superfamily (NR3C4, NR3A1, and NR3A2, respectively). These receptors are comprised of a modular structure involving an intrinsically disordered N-terminal domain (NTD), a DNA-binding domain (DBD) consisting of two zinc fingers, the flexible hinge region, and the C-terminal ligand-binding domain (LBD) (Figure 2) (59, 60). Their classical action is initiated in the cytoplasm when ligands bind in a hydrophobic pocket of the LBD. This induces conformational changes revealing a nuclear translocation

signal in the hinge region. The LBD also harbors activation function 2 (AF-2), a ligand-dependent interaction surface for coregulatory proteins or intra- and intermolecular interactions with the NTD (N-terminal/ C-terminal interactions). Ligand-independent receptor activation can also occur through protein interactions and/or post-translational modifications of activation function 1 (AF-1), situated in the NTD. In both murine and human osteoblasts, a 46-kDa alternative splice variant of ER α lacking AF-1 has been demonstrated.

In the nucleus, these nuclear receptors bind as dimers to specific DNA sequences termed androgen or estrogen response elements (AREs, EREs), interact with other transcription factors, and recruit coregulatory proteins, chromatin

remodeling complexes, and components of the transcription machinery. Two types of AREs exist: 1) canonical AREs that form a clear palindromic repeat of the 5'-AGAACA-3' consensus sequence; and 2) selective AREs with a more relaxed resemblance to direct or palindromic repeats of the same hexamer (61). Selective AREs play a role in the male reproductive system but not in the musculoskeletal system, as shown in AR knock-in mice with abrogated binding to selective AREs (62).

Nuclear receptors can also bind chromatin indirectly via tethering to other proteins: ER, for example, may associate with the activating protein-1 (AP-1; a complex containing Fos/Jun) or specificity proteins (eg, SP-1), which can bind to specific DNA motifs different from EREs. Both AR and ER α are able to form heterodimers with other transcription factors to modulate their activity in osteoblasts as shown for, eg, RUNX2 (see Section VII.C). Finally, coactivators or corepressors can modulate tissue-specific activity of AR/ER on target genes. The in vivo importance of coactivators in bone was illustrated by a study in ovariectomized steroid receptor coactivator-1 knockout mice; physiological doses of E2 restored cortical bone, but supraphysiological doses were required to restore trabecular bone (63). Further research on AR/ER coactivators as tissue-specific modulators of sex steroid responses in bone is warranted.

D. Ligand-independent and nongenomic actions

Nonclassical AR/ER actions fall into two categories. In nongenomic (synonym, nongenotropic) signaling, the liganded receptor activates second messengers like kinases, phosphatases, cytoplasmatic calcium release through ion channels, or nitric oxide synthesis. The second category of ligand-independent activation occurs without occupancy of the ligand-binding pocket; this mode is triggered mostly via phosphorylation of AF-1.

A hallmark of the former nongenomic effects is that these can occur within seconds or minutes. AR has been found to exert nongenomic effects via components of the SRC, ERK, PI3K, and AKT pathways. A particular form of nongenomic actions may occur through transmembrane receptors. For example, G protein-coupled receptor 30 (GPR30; see Section IV.E) has been proposed as a high-affinity membrane G protein-coupled ER (renamed GPER1), although this hypothesis is not unequivocally supported (64). ER α isoforms may signal at the plasma membrane as well (65). A transmembrane AR has been proposed but remains controversial, and a role in skeletal physiology has not been demonstrated.

The latter, ligand-independent activation mode has clearly been demonstrated for $ER\alpha$ but is more controversial for AR. Kinases responsible for AR/ER phosphorylation (eg, MAPK, AKT, MEK1/2, and ERK1/2) may be

activated by several growth factors (eg, IGF-1, keratinocyte growth factor, epidermal growth factor) or by feedback from nongenomic sex steroid signaling. This may be sufficient to mediate ER α nuclear translocation, whereas for AR it may enhance androgen-induced translocation. In vitro ER α transfection in osteoblastic cells has been noted to activate transcription in the apparent absence of estrogens (66), whereas this is not the case for AR and androgens. In vivo, ERE reporter mice showed ER-dependent bone luciferase activity even when they were immature, gonadectomized, or treated with aromatase inhibitors (67), thus supporting ligand-independent ER α actions in bone.

To what extent these nonclassical mechanisms contribute to the skeletal effects of androgens, estrogens, or their respective receptors will be discussed in *Section IV.E.*

E. Rodent models in the study of sex steroids and bone: a critical appraisal

Animal models have proven invaluable to investigate the cellular and molecular mechanisms of sex steroid actions in bone. But before discussing the results of these studies, some caveats will be reviewed.

1. Differences in circulating sex steroid physiology

Circulating sex steroid physiology differs between humans, mice, and rats, mainly because the latter two lack circulating SHBG. This is thought to contribute to the observation that rodents have very low, highly fluctuating serum T levels. Indeed, SHBG transgenic mice have very high T levels in parallel with their SHBG overexpression (68).

Although it is clear that estrogens are important for the male rodent skeleton, circulating E2 levels measured with the "gold standard" of mass spectrometry are below the limit of detection of sensitive assays (69, 70), suggesting that local aromatization and intraskeletal effects are important. Even pharmacological doses of continuous T do not increase serum E2 in male mice, presumably because aromatase expression in rodents is mainly limited to the gonads and certain areas of the brain (71). Given the increased recognition in human studies that immunoassays may be unreliable and confounding to much of the literature, mass spectrometry will also be needed to reexamine serum T and E2 concentrations in different rodent models (72). Administration of E2 to (orchidectomized) male mice does produce measurable E2 serum levels (70), which are thus by definition unphysiological. This results in a marked increase in trabecular bone volume and formation (73, 74), which is unparalleled in humans (for example, unseen in male-to-female transsexuals given pharmacological E2 doses) (75). Together with the aforementioned findings on skeletal bioluminescence in ERE reporter mice

(67), these data argue against a purely endocrine role of circulating E2 in mediating genomic ER actions in rodents and point to the importance of intracrine effects of estrogens via local aromatization and/or ligand-independent ER activation in bone (see *Sections II.B*, *II.D*, and *IV.E*).

Gonadectomy is widely used as a preclinical model of osteoporosis, but because other gonadal endocrine bonestimulating factors are lost as well, eg, inhibin A (76), control experiments with hormone replacement are necessary (eg, with T or nonaromatizable DHT). The role of the adrenal glands in rodents as a source for androgens and estrogens has been suggested as negligible, although several studies have suggested that adrenal androstenedione may represent an additional source of (aromatizable) androgens in mice and rats (27, 77, 78). To overcome limitations of assessing androgenic/estrogenic status based on serum sex steroid concentrations, the weight of sex steroid-sensitive organs is commonly used, as an in vivo bioassay (seminal vesicles and levator ani muscle in males and uterus weights in females). Unfortunately, there is no good tissue indicator of estrogen bioactivity in males.

2. Genetic rodent models and their limitations

Genetic models preventing androgen or estrogen actions at the level of the receptor preclude confounding by metabolites, eg, from aromatization or adrenal origin. Ubiquitous androgen- or estrogen-receptor knockout (ARKO, ERKO) mice have greatly contributed to our understanding of sex steroid actions in bone. Unfortunately, ubiquitous models suffer from disrupted negative feedback on the hypothalamic-pituitary axis as well as altered serum IGF-1 levels in some models (Table 1). As mentioned above, the low serum sex steroid concentrations in these animals (especially E2) were probably not reliably measured with previous immunoassays; serum E2 is undetectable with mass spectrometry in both wild-type and ARKO males (<0.6-1.3 pg/mL; our unpublished obser-

vations). Furthermore, serum levels may not accurately reflect intraskeletal metabolism or receptor activation. For example, IGF-1 expression and IGF-1 receptor levels have been found down-regulated in other tissues in ARKO mice (79).

The first genetic models used to study androgen actions in bone were male testicular feminization (Tfm) rats that spontaneously developed a single base mutation in the AR LBD. Their bone phenotype is intermediate between females and males, which is, however, confounded by decreased IGF-1 and high estrogens (Table 1). Tfm mice, on the other hand, harbor a single base deletion and frameshift in the AR NTD but still express a truncated C-terminal AR peptide initiated upstream of the DBD with residual DNA- and hormone-binding activity (27).

Five different ubiquitous ARKO mouse models have been developed (97) that differ in their deletion of either exon 1, 2, or 3, resulting in abrogation of the NTD (98), the first zinc finger (80, 99), or the second zinc finger (100) of the DBD, respectively. The concern that AR is still expressed and may retain some of its actions in Tfm or resistant ARKO models is alleviated in the ARKO models that totally lack AR expression (80, 98, 99). Mice with abrogation of the AR second zinc finger (AR $^{\Delta ZF2}$ mice) purposely retain normal ligand binding and nongenomic effects on second messengers like ERK phosphorylation (100, 101) (see also Section IV.E). However, the possibility that the absence of an entire domain in this model results in protein misfolding and off-target effects cannot be completely excluded. Moreover, the floxed allele in $AR^{\Delta ZF2(flox/Y)}$ male mice results in increased baseline ARsensitivity (102).

The first reported ER α KO mouse model from the group of Korach and Smithies lacks the normal 66-kDa protein but still expresses the less abundant 46-kDa normal isoform as well as an unexpected 61-kDa splice vari-

Table 1. Summary of Endocrine Changes in Male Global Genetic Rodent Models Used in Studies on Sex Steroids and Male Bone Metabolism

	FSH/LH	Androgens	Estrogens ^a	IGF-1	Refs.
Tfm rats	<u> </u>	<u> </u>	<u> </u>	\downarrow	27
Tfm mice	<u></u>	,	<u> </u>	NR	71
ARKO	<u></u>	↓	<lod< td=""><td>= *</td><td>80-84</td></lod<>	= *	80-84
Aromatase KO	<u>`</u>	<u></u>	\downarrow	↓ (↑ F)	85-87
$ER\alphaKO$	<u></u>	<u> </u>	↑	.	84, 88, 89
$ER\beta KO$	=	=	=	= (↑ F)	89-93
NERKI	NR	NS (↑ F)	=	↓	94-96

Abbreviations: LOD, limit of detection; ↑, increased; ↓, decreased; =, no change; NR, not reported; ↑F, increase in female mice reported in some studies; = *, serum levels were reported as not decreased, but intraskeletal signaling may still be disturbed; NS, serum T was nonsignificantly increased due to high variability, and weights of androgen-responsive tissues were not reported.

^a Low circulating E2 was measured with immunoassays in most studies, and changes paralleled aromatase activity; with mass spectrometry, however, serum E2 is below the limit of detection (<0.6-1.3 pg/mL; our unpublished results) in male wild-type and ARKO mice.

ant, which both have possible activity of AF-2 but not AF-1 (27). Subsequent models represent truly null mutants based on deletion of the first zinc finger of the ER α DBD. Whether a putative splice variant lacking the DBD in the original ER β KO mice could still harbor residual activity has not been clearly demonstrated, but the possibility has been used to explain differences to other ER β KO models (93, 103, 104). Due to a lack of negative feedback on the hypothalamic-pituitary axis, ERαKO models have increased levels of gonadotropins, androgens, and estrogens (84, 88)—whereas circulating T was normal in the ER α KO man (105). Male ER β KO mice do not have disturbed hypothalamic-pituitary feedback (90-93), although IGF-1 was increased in young ER β KO females in one study (90). Male mice with a knock-in ER α (NERKI mice) with abrogated DNA binding but capability to mediate nonclassical signaling had nonsignificantly increased T and were suggested to have elevated E2 serum levels on a mixed genetic background (94, 95).

The main revolution in our understanding of the mechanisms of action of androgens and estrogens comes from the use of conditional genetic mouse models. Bone cellspecific ARKO/ERKO models have the potential to avoid disrupted feedback on other pathways (eg, systemic IGF-1) or indirect effects on bone via ER and AR in other tissues (eg, decreased voluntary physical activity in ARKO mice) (106, 107). However, Cre-LoxP technology has its own pitfalls, including independent effects of inserting a Cre somewhere in the genome or LoxP sites in a gene, incomplete deletion (which is typically still only around 80% in efficient conditional models), off-target Cre activity in cells and tissues other than those targeted, "leaky" Cre activity in inducible models or confounding effects of hormone therapy used to induce the recombinase (which probably explains why no studies on sex steroids and bone have yet used tamoxifen- or progesterone-inducible Cre systems). Moreover, the phenotype of noninducible models could result from embryonic or perinatal imprinting effects, even when this phenotype becomes evident at later ages.

In summary, sex steroid physiology differs between male mice, rats, and humans, mainly because of the absence of SHBG and differences in peripheral, extraskeletal aromatization. Genetic models exclude AR or ER activation altogether but harbor many caveats including the presence of residual receptor isoforms, nonclassical activation, baseline confounding phenotypes in Cre or LoxP controls, intrinsic perturbations of the hypothalamic-pituitary axis in global, or eg, neuron-specific models (108), and indirect effects on bone via other tissues. As such, use of selective ligands in gonadectomized, completely null genetic models with careful attention to con-

trols should probably be regarded as the highest level of evidence in the face of conflicting results. Future studies using inducible Cre systems are needed to avoid developmental effects and dissect the timeframe in which AR/ER actions on bone are determined.

III. Structural Basis of Male Bone Strength

Fracture risk is determined by bone strength on the one hand, and fall risk and defense mechanisms (eg, soft tissue absorption of kinetic energy) on the other hand. Bone strength in turn is determined not only by BMD but also by bone dimensions, microstructure, and material properties (2). BMD is usually measured by dual-energy x-ray absorptiometry (DXA), but because it is a projectional (two-dimensional) technique, it cannot truly differentiate between cortical and trabecular bone. Quantitative computed tomography (QCT), on the other hand, can separately measure the volumetric BMD (vBMD) of both bone compartments and provides bone geometrical parameters together with indices of bone strength. Still, this technology is limited by its resolution (about 400 μ m) to accurately measure bone microstructural properties. This has been overcome by the advent of high-resolution peripheral QCT (HRpQCT), which permits the noninvasive measurement of the microstructural properties of the trabecular and cortical bone compartments at the distal radius and distal tibia with a resolution of 82 µm or, more recently, even less. This is sufficient to quantify the larger pores in bone (cortical porosity). HRpQCT also provides the highest quality data to generate finite element analysis models for estimation of bone strength. Emerging new technologies like microindentation may be able to determine bone material properties and sexual dimorphism herein, but further validation is needed.

In the following paragraphs, we will discuss the structural basis of male bone strength, which determines the decreased risk of osteoporotic fractures in men. Understanding how gender dimorphism in bone geometry and microstructure is established during adolescence and young adulthood may offer insights in fracture susceptibility and its prevention later in life. Sex steroids probably determine sexual dimorphism in these (HR)pQCT parameters, but because current evidence is limited by its mostly cross-sectional and observational nature, sex steroid regulation of cortical and trabecular peak bone mass acquisition is better understood from murine and sporadic human genetic models discussed in *Section IV*.

A. Favorable cortical bone size and trabecular microstructure in young adult men

Men are on average 10% taller than women because of their greater bone length. Even after correcting for bone length, men have greater bone width than women, with cortical bone placed further away from the neutral axis of the bone. This results in an exponential increase in strength because the rigidity of a tubular bone scales as the fourth power of its diameter (2). Using QCT, Riggs et al (109) investigated sexual dimorphism in bone geometry in an age- and sex-stratified, population-based, cross-sectional sample. At both central (lumbar spine, femoral neck) and peripheral (distal radius, distal tibia) sites, young adult men had a 25-33% larger cross-sectional bone area than young adult women, and these relative differences were maintained throughout life. Trabecular vBMD was greater in men at all sites, whereas cortical vBMD was mostly similar between sexes (109). The larger bone width in men results from greater periosteal expansion during puberty and early adulthood in men, whereas girls predominantly increase their cortical thickness by limiting endocortical expansion.

Serum levels of calculated free T were positively associated with the cortical cross-sectional area and periosteal circumference at both the tibia and the radius of young adult Swedish men (110). Also in young men, higher serum

SHBG levels were associated with larger cortical bone area and periosteal and endocortical circumference, independently from free sex steroid levels, suggesting an independent role of SHBG in determining cortical bone size at the age of peak bone mass (110, 111). The reduced bone size in women may predispose elderly women to a greater fracture risk than elderly men after similar age-related bone losses have occurred (see *Section IX*).

Structural differences between male and female bone have more recently also been assessed by HRpQCT. Khosla et al (112) found that young adult men had a 26% greater trabecular bone volume with more plate-like, thicker trabeculae but similar values for trabecular number and separation compared to women (Figure 3, A–C). Bone area and medullary area were also larger in young men compared with young women. Cortical thickness at the age of peak bone mass was similar in both sexes (Figure 3D), and cortical vBMD was slightly lower (5%) in young men (112). These gender differences have been confirmed by several other cross-sectional HRpQCT studies (113–119).

Correlation analyses between serum levels of sex steroids and the differences in bone structure during pubertal growth showed that the increases in trabecular bone volume and trabecular thickness in boys were correlated to

Figure 3.

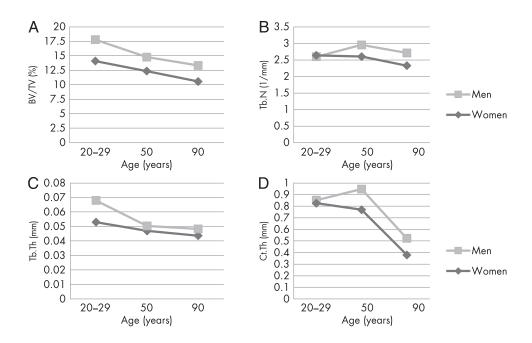


Figure 3. Age-related changes in structural determinants of bone strength in men and women based on a cross-sectional study using high-resolution peripheral CT at the ultradistal radius. A, BV/TV, trabecular bone volume/tissue volume; B, Tb.N, trabecular number; C, Tb.Th, trabecular thickness; D, Ct.Th, cortical thickness. [Adapted and reproduced from S. Khosla et al: Effects of sex and age on bone microstructure at the ultradistal radius: a population-based noninvasive in vivo assessment. *J Bone Miner Res.* 2006;21:124–131 (112), with permission. © American Society for Bone and Mineral Research.]

higher levels of both serum T and IGF-1 (116). The larger bone width in boys, evidenced by the larger periosteal circumference, was associated mainly with serum T. Although serum E2 was not an independent predictor of skeletal dimorphism in this multivariate model (116), part of the effects of T could still be mediated by aromatization into E2 locally in bone, which may not always be reflected by serum E2 levels (see *Section II.B*). Indeed, human cases of ER α and aromatase deficiency have clearly illustrated that estrogen actions are crucial in peak bone mass acquisition in both genders (see *Section IV*). Other studies, however, found no association between serum levels of sex steroids and trabecular or cortical microstructure in young adult men (112, 119).

Intracortical porosity (calculated as the pore volume normalized to the cortical bone volume) appears significantly greater in men in their 20s and 30s than in women (113, 115, 117-119). Although a more porous cortex would be a disadvantage in biomechanical terms, this was counterbalanced by a larger cross-sectional bone area. Size-dependent indices of bone strength such as stiffness and estimated failure load were higher in young adult men compared to young adult women, whereas apparent modulus, which is size-independent, was similar in both sexes (113, 115, 117–119). These findings indicate the importance of geometry with respect to bone strength. Cortical thickness, cross-sectional area, and cortical vBMD of the tibia and/or radius were all greater in young adult men than in late adolescent men, showing that early adulthood is a period of ongoing cortical modeling with further increases in cortical thickness and estimated bone strength (119, 120).

In summary, men have a structural advantage over women from early adult life characterized by wider bones due to greater cortical bone expansion and greater trabecular bone volume due to thicker trabeculae translating into superior indices of bone strength (Figure 3), which may protect against fractures later in life. The larger periosteal expansion and trabecular bone volume in men have been independently associated with serum T, SHBG, and IGF-1 in some (but not all) studies, although genetic disruption of ER α actions (discussed in *Section IV.A*) is certainly also detrimental for male peak bone mass acquisition.

B. Better maintenance of trabecular and cortical bone structure in men across the lifespan

Studies using central and peripheral QCT and HRpQCT have shown that, in general, age-related deterioration in bone geometry, vBMD, and bone microstructure affects men and women equally, but because older men maintain their sex steroid levels better, they do not

experience an equivalent to the accelerated phase of bone loss associated with menopause in women (ie, no "andropause" equivalent).

1. Cortical geometry

Riggs et al (109) nicely demonstrated, using QCT in both a cross-sectional and a 3-year longitudinal study (121) in an age- and sex-stratified population sample of men and women, that the total bone cross-sectional area continues to increase over a lifetime by about 15% in both sexes. This is, however, outpaced by endocortical resorption, estimated from the 25-40% increase in total medullary area at the femoral neck and distal radius, resulting in an age-related decrease in cortical thickness in both sexes (Figure 3D). Longitudinal pQCT data from the InCHIANTI study confirm that older women show a dramatically greater medullar expansion and resulting greater cortical thinning compared with older men, although periosteal expansion (estimated from the increase in total cross-sectional area) is slightly greater in older women than in older men (122). These changes in cortical geometry were also confirmed by cross-sectional agestratified HRpQCT studies (112–115, 117).

2. Cortical and trabecular vBMD

Loss of trabecular and cortical vBMD with age is more pronounced in women than in men, although the rates of trabecular bone loss vary according to site. Remarkably, loss of trabecular vBMD starts already in young adult life in both women and men and continues throughout life with acceleration in women at perimenopause. In men, the rate of trabecular bone loss appears to attenuate in older men at the distal radius and tibia but not at the spine. In contrast, cortical vBMD remains relatively stable until midlife in both sexes with substantial loss of cortical vBMD after menopause in women, but only from about age 75 in men (109, 112, 117, 121, 122). In the Osteoporotic Fractures in Men (MrOS) study, men over 85 years of age had 22% lower trabecular vBMD but similar cortical vBMD of the femoral neck compared with men aged 65–69 years (123). At the femoral shaft, bone cross-sectional area increased by 9% in the oldest men, consistent with ongoing periosteal apposition with age, but because the endocortical area increased by 22%, the cortical area decreased significantly. It is plausible that the decreases in cortical thickness and cortical and trabecular vBMD after the age of 50 in women and in old age in men are driven by sex steroid deficiency both in women (by menopause) and at least partly in men (by the age-related decline in bioavailable sex steroids). However, the continuous decrease in trabecular bone even in young adult men and women, accounting for between one-third and one-half of

the total trabecular bone loss over life (121), occurs during sex steroid sufficiency, and the mechanism responsible for this remains unexplained.

3. Trabecular bone microstructure

Age-related microstructural changes at the trabecular compartment in the radius include similar decreases in trabecular bone volume in men (-26%) and women (-23to -27%) (Figure 3A), although the microstructural changes underpinning this trabecular bone loss differ between the sexes (112, 117). Women undergo loss of trabeculae, resulting in an increase in trabecular spacing, whereas in men between the ages of 20 and 49, the number of trabeculae increased while at the same time trabecular thickness decreased dramatically (Figure 3, B and C). These findings are consistent with the idea that in men, bone loss is mainly due to thinning of the trabeculae because of decreased rates of bone formation, whereas in women, the increased rates of bone resorption associated with menopause contribute to the loss of trabeculae (124). Importantly, this pattern of trabecular bone loss appears to vary with skeletal site. In contrast to the findings at the radius, Macdonald et al (117) and Hansen et al (115) recently demonstrated in population-based samples of men and women that in the distal tibia, both trabecular thinning and loss of trabeculae contributed to the loss of trabecular bone volume in both men and women. These studies also suggested site-specific differences in the onset of trabecular bone loss. There is clearly a need for prospective HRpQCT studies to confirm the age of onset, exact nature, and underlying mechanism of microstructural deterioration in men and women.

4. Cortical porosity

Age-related microstructural changes at the cortical compartment are dominated by significant increases in cortical porosity in both men and women (113, 115, 117, 125), coinciding with (and possibly explaining to a large extent) the decline in cortical vBMD. The increase in cortical porosity in men is less pronounced than in women and does not occur until old age in men, in accordance with the findings for loss of cortical vBMD. Recent data demonstrate that serum E2 levels associate inversely with cortical porosity in elderly Swedish men, indicating that E2 might regulate cortical bone microstructure (126).

In summary, men have favorable bone structure at peak bone mass, and age-related decreases in bone structure and strength at both the trabecular and cortical compartment are delayed and less pronounced in aging men compared to women. The patterns of age-related structural changes are gender-specific and also differ between weight-bearing (tibia) and non-weight-bearing (radius) sites. The larger number of thicker trabeculae and better maintenance of cortical thickness together with their larger bone width all contribute to the observed lower fracture incidence in men. This occurs most specifically at the radius, where the risk in childhood is greater in boys (at least in part due to a transient deficit during cortical bone expansion) (116), whereas there is a marked increase in Colles' fracture early after menopause in women compared with an almost complete absence in men (127). Longitudinal HRpQCT studies are needed to more accurately determine age-related changes in trabecular and cortical bone microstructure in men and women because the available cross-sectional studies cannot determine true age-related changes because of possible confounding by secular trends, as nicely illustrated by the InCHIANTI study (122).

The relative contributions of androgens, estrogens, and SHBG in determining bone loss and fracture risk in older men will be the focus of *Section IX*. More studies are needed to determine the role of sex steroids in age-related microstructural bone decay.

IV. Regulation of Skeletal Sexual Dimorphism

In this section, we will examine the relative contributions of androgens and estrogens for cortical and trabecular bone development. Puberty is a critical period for peak bone mass acquisition and establishment of sexual dimorphism in bone mass and geometry. This period may be even more important than age-related bone loss in determining the risk of osteoporotic fractures in later life (128). Cell-specific ER α KO and ARKO mice have recently advanced our understanding of sex steroid regulation in cortical and trabecular bone. These studies will be discussed in detail in *Section V*. The principal conclusions on the effects of sex steroids and their receptors in male and female peak bone mass acquisition are summarized schematically in Figure 4.

A. Estrogen receptor α : crucial for peak bone mass

1. Role in cortical and trabecular bone development in females

Estrogens have traditionally been viewed as promoting trabecular and cortical bone development in females. Compared to males, however, periosteal bone expansion, endocortical widening, and trabecular thickness are lower in females (see *Section III*). The importance of estrogens is confirmed in aromatase knockout mice: females have poor trabecular and cortical bone development despite increased T and IGF-1 (85, 87). Female ER α KO mice have been reported as having paradoxically increased trabecu-

Figure 4.

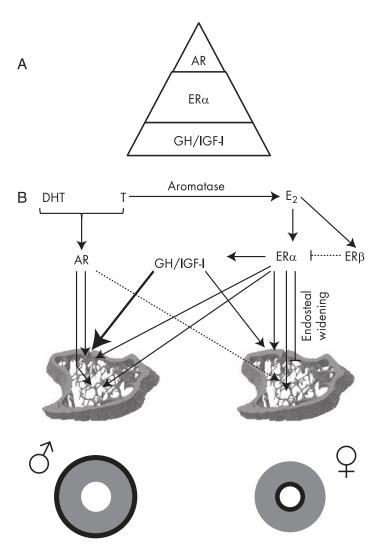


Figure 4. Schematic representation of the effects of sex steroids and their receptors in the development of male and female pubertal bone development. A, Pyramid hierarchy of sexual dimorphism in bone. Given the severely disordered bone development in case reports of $ER\alpha$ mutations or aromatase deficiency, compared to the normal female-like bone structure in XY females with AIS, the effects of $ER\alpha$ should be considered critical, whereas AR provides further benefits that, however, are unable to compensate for severely decreased $ER\alpha$ activation. Even more severe is the skeletal phenotype of developmental disruption of the GH/IGF-1 axis in both genders, and skeletal sexual dimorphism is completely lacking in GH-receptor knockout mice (with very low circulating IGF-1 levels) (78). B, Androgens like T can be converted via aromatization to estrogens and can thus activate both AR and ER α . In males, both AR and ER α stimulate cortical and trabecular bone development. During puberty, males have greater periosteal expansion than females (bottom left, black outer circle), which depends both on AR (in late puberty) and ER α (in early puberty), but especially on IGF-1 (probably via central aromatization of androgens) (78, 82). Trabecular bone formation is increased by $ER\alpha$ in males (220), whereas both $ER\alpha$ and AR can inhibit trabecular bone resorption. Additionally, AR inactivation has been shown to slow mineralization in early growth, when bone turnover is high (170). In females, $ER\alpha$ is responsible for both trabecular and cortical bone mass accretion, but their cortical thickness relies more on limiting endocortical expansion (bottom right, black inner circle). $ER\alpha$ inhibits endosteal bone resorption and stimulates periosteal bone formation (179), although the latter is lower compared to males (82). Females have higher endosteal bone formation (78) due to actions of $ER\alpha$ (95). Estrogens also stimulate trabecular bone formation in female mice (219, 221, 222). Both androgens and estrogens can decrease trabecular bone resorption in female mice (130, 212), although the physiological importance of androgens in female mice is probably limited. ER β is thought to repress $ER\alpha$'s stimulatory actions on cortical and trabecular bone in female mice and to play no role in male mice (91, 150, 151), but definitive evidence for a role in humans is lacking.

lar bone in the peripheral skeleton due to very high levels of T stimulating AR (90, 93, 129). This confirms other studies suggesting that compensatory actions via AR can maintain trabecular but not cortical bone mass in female rodents (130, 131), although this probably has minor physiological importance. Complete $ER\alpha\beta KO$ female mice again have a clear cortical and trabecular bone deficit, and $ER\alpha$ is required for the cortical and trabecular bone response to estrogens in ovariectomized mice (93, 129).

Osteoclast-, osteoblast-, and osteocyte-specific ERαKO models have now been used to avoid systemic endocrine alterations. These studies have shown that ER α actions are important to stimulate periosteal and trabecular bone formation and inhibit endocortical and trabecular bone resorption via different bone cells (see Section VI). Female mice also have increased endosteal bone formation compared to males (78), which may rely mainly on nonclassical actions (see Section IV.E) of ER α (95). In summary, ER α activation is crucial for cortical and trabecular bone development in female mice.

2. Role in cortical and trabecular bone development in males

The historic dichotomous view that estrogens were important for female bone health and androgens for male bone health were challenged by a unique report of a man with an ER α null mutation (105). He presented with tall stature, incomplete epiphyseal closure in adulthood, markedly decreased BMD, normal T, and increased E2, gonadotropin, and IGF-1 levels, demonstrating the importance of ER α for peak bone mass and growth plate closure in men. Further pQCT and histomorphometric evaluation in this man showed significant impairment of cortical thickness (due to increased endosteal expansion, whereas radial periosteal circumference was similar), as well as deficient cortical and trabecular vBMD with thinner trabeculae and decreased

bone formation (132). Similarly, men with aromatase deficiency have high T but suboptimal bone mass, which cannot improve without estrogen replacement (133). A pQCT study in an adolescent male with congenital aromatase deficiency revealed that increases in BMD during estrogen therapy were not the result of trabecular or cortical vBMD gains but rather increases in cortical thickness and cross-sectional bone area, suggesting that estrogens are required for optimal periosteal pubertal bone expansion typical of the male bone phenotype (134).

In mice, ER α and ER $\alpha\beta$ KO males also have cortical bone deficits despite increased serum T, in line with observations in male rats that pure androgens cannot compensate for a complete loss of estrogen signaling in cortical bone development and maintenance (89, 93, 135). Trabecular bone development, on the other hand, has been reported as normal (89, 136) or even increased in male $ER\alpha KO$ mice due to increased serum T acting via the AR (84, 93, 129), suggesting that AR is sufficient for trabecular bone development in male mice. On the other hand, E2 is a strong anabolic stimulus for trabecular bone in male mice, and this effect is $ER\alpha$ -dependent (129, 137), although it is currently impossible to define physiological E2 levels in male mice (see Section II.E). Male aromatase knockout mice have clearly reduced trabecular bone volume and increased endocortical bone resorption (77). In more recent conditional knockout models, some studies also suggest that ER α is involved in male bone development (see Section V). The main conclusion from both human and mouse genetic models is that male cortical and trabecular peak bone mass acquisition is dependent on both AR and ER α actions.

B. Estrogen receptor β

Estrogens act via at least two receptors (ER α and ER β), so the question is which of these is chiefly responsible for the effects of estrogens. ER β seems to play no role in bone development in male mice because male ER β KO mice have normal bones and ER $\alpha\beta$ KO mice show no difference compared to ER α KO males (89, 91, 129, 137, 138). Female ER β KO mice also show normal trabecular bone response to E2 after ovariectomy (129, 139), and selective ER β agonists are unable to protect rats from ovariectomyinduced bone loss (140, 141). Thus, ER α is definitely the dominant mediator of the skeletal actions of estrogens. Still, three independent groups have shown in female ER β KO mice that longitudinal and transversal bone growth are enhanced and age-related trabecular and endocortical bone resorption are decreased, the latter of which was associated with increased ER α mRNA levels (91, 92, 139, 142, 143). In a fourth ER β KO model, however, a transiently increased trabecular bone volume but no effect on longitudinal or radial bone growth was observed in female mice (93). Some studies also suggest that ER β is present in cancellous bone whereas cortical bone has little or no ER β (63, 144, 145). Less rapid trabecular bone loss and a greater increase in cortical area have in turn been observed in long-term follow-up of ovariectomized adult ER β KO mice (146). Female ER β KO also display enhanced fracture healing and increased mandibular condylar growth (147, 148). Overall, ER β appears to have a minor inhibiting effect on the positive actions of ER α in female mice.

Conflicting results have been presented on whether or not ER β can also partly compensate for bone loss due to the absence of ER α when ovariectomized complete ER α KO female mice were treated with E2 (129, 149). These data led Lindberg et al (150) to propose a "yinyang" relationship in female mice, with a moderate repressive function of ER β on normal ER α signaling, whereas ER β might partially compensate when ER α is lost (93, 150). Interestingly, ER β expression also appeared higher in stromal cells isolated from the ER α KO man (132).

Recently, to avoid putative activity from ER β transcript variants (104) and systemic influences, Khosla's group (151) selectively deleted ER β in osteoblasts (*Col1a1* 2.3 kb-Cre) and osteoprogenitor cells (*Prx1*-Cre). The latter increased trabecular bone volume by 93% in female mice (151).

We conclude that $ER\alpha$ is the dominant mediator of estrogen actions in bone. Studies in $ER\beta$ KO mice support a repressive function of $ER\beta$ in bone of females. The reason behind the gender difference in the role of $ER\beta$ is unknown. Evidence for a role of $ER\beta$ in male mice is lacking. In humans, several observational studies suggest that $ER\beta$ dinucleotide repeat polymorphisms are associated with BMD (although with opposite effects in different studies) (152–158) and a lower risk of femoral fracture (159), but this has not been confirmed in meta-analyses of genome-wide association studies (GWAS) (25, 26, 160–162). Thus, a definite role for $ER\beta$ in humans remains hitherto unconfirmed.

C. $ER\alpha$ AF-1 and AF-2

Recently, the effects of $ER\alpha$ in bone and other tissues have been further analyzed in terms of whether they require $ER\alpha$ AF-1 or AF-2 (see *Section II.C*).

Both gonadectomized male and female mice lacking AF-2 showed no response to E2, demonstrating the importance of the ligand-dependent AF-2 for the effects of E2 on cortical and trabecular bone. Importantly, female mice lacking AF-1 of ER α still displayed normal response in cortical bone, but not in trabecular bone, and showed only

weak effects on the uterus (149). Similarly, $ER\alpha AF-2$ was required for skeletal E2 responsiveness in male mice, whereas the role of $ER\alpha AF-1$ was again tissue-specific and was seen for cortical but not trabecular bone (136). Thus, selective $ER\alpha$ agonists with minimal effects on $ER\alpha AF-1$ could retain efficacy on cortical bone (which constitutes most of the skeleton) with minimal stimulation of reproductive tissues. Unfortunately, none of the existing selective ER modulators (SERMs) tested in a recent study were able to exert these actions in $ER\alpha AF-1$ null males (136). Surprisingly, the $ER\alpha$ antagonist ICI 182,780 (fulvestrant, Faslodex) behaves as a partial agonist on trabecular bone and uterine weights (but not cortical bone or other estrogen-sensitive organs) in ovariectomized $ER\alpha AF-2$ null mice (163).

The role of AF-1 and AF-2 in mediating ER α 's modulation of the response to mechanical loading is discussed in *Section VIII.D.* Regarding growth plate closure, ER α AF-1 seems not to be required—in fact, ER α AF-1 null females show accelerated growth plate closure, suggesting that ER α AF-1 opposes normal ER α -mediated growth plate closure (164). In ovariectomized ER α AF-2 null mice, on the other hand, ICI 182,780 behaved as an inverse agonist, increasing growth plate height in the absence of estrogens (163).

Taken together, these data suggest that estrogen actions do not require full-length $ER\alpha$, which opens perspectives for tissue-selective $ER\alpha AF-1$ or -AF-2 modulators.

D. Role of the androgen receptor

1. Providing an edge to cortical and cancellous bone

Following the realization that estrogens were crucial for male peak bone mass as evidenced by the severe consequences of $ER\alpha$ mutation in a man (105), confirmed by studies in male $ER\alpha KO$ and aromatase knockout mice discussed above, doubts arose as to whether aromatization alone would be sufficient to explain greater peak bone mass development in men.

It is important to emphasize here that estrogen deficiency has severe consequences in both genders, whereas androgens may merely serve to enhance bone (and muscle) mass in men compared to women. In other words, the worst consequence of impaired AR actions will probably still only be a lack of sexual dimorphism and consequent feminization of bone development, ie, lack of male-type musculoskeletal enhancement. This prediction was confirmed in a study in pubertal male ARKO and wild-type female mice, which had identical cortical bone parameters (165). Trabecular bone development, on the other hand, was worse in ARKO males compared to females. Male aromatase knockout mice also show additional trabecular

and endocortical bone resorption after orchidectomy (77). Further attesting to the pubertal skeletal actions of androgens independent of aromatization is a unique pQCT study in a man with concomitant aromatase deficiency and mild hypogonadism, in whom adding T to E2 replacement increased BMD, vBMD, cortical thickness, periosteal expansion, and normalized bone turnover (166). In the ER α KO man, all pQCT parameters were markedly lower compared to a control population except periosteal circumference, which was not significantly decreased (132).

Another question is whether androgens are directly responsible for the male skeletal phenotype or whether this is caused by other genetic determinants present on the Y chromosome. This can be examined, for example, in transsexual men and women, in whom gonadectomy and prolonged cross-gender hormone therapy reproduce the effects of karyotype on cortical bone size, trabecular vBMD, and muscle mass and strength (75, 167). Similarly, bone size has been reported intermediate between male and female as judged by pQCT in an XY woman with androgen insensitivity syndrome (AIS) (168). In contrast, BMD is much lower in both XX and XY women if gonadal dysgenesis is present, implying that gonadal factors (presumably sex steroids) are dominant over other chromosomal determinants (169).

2. Evidence from Tfm and ARKO rodent models

Following earlier studies in Tfm rats (reviewed in Ref. 27), it was shown in orchidectomized Tfm mice and also in orchidectomized growing ARKO mice that T could no longer prevent bone turnover and trabecular bone loss or stimulate periosteal bone formation, pointing to a direct role for AR in these processes (71, 82). A similar conclusion was reached in double AR-ER α KO and ER $\alpha\beta$ KO mice in which AR activation was sufficient for normal trabecular bone development and maintenance, whereas both AR and ER α activation was required for optimal cortical bone development (84). The observation that E2 is a more potent stimulus for trabecular bone than DHT (135) is in apparent contradiction with these findings, but the physiological relevance of pharmacological E2 doses in male rodents with undetectable circulating E2 levels remains questionable (see also Section II.E) (74, 135). Thus, AR actions independent of aromatization contribute to greater periosteal bone formation and enhanced trabecular bone development during puberty in male mice. This fits with the observed gender dimorphism in adults at the age of peak bone mass (see Section III).

The effects in knockout mice should be considered developmental; studies on the role of perinatal androgen secretion are discussed in the next paragraph, and studies on cortical and trabecular bone maintenance using selec-

tive ER or AR agonists in adult male ER α KO or ARKO mice are discussed in *Section IX*. Importantly, the effect of androgens on skeletal sexual dimorphism, in particular periosteal expansion, may partially depend on ER α (via aromatization) and stimulation of the GH/IGF-1 axis (see *Section VIII.A*).

Collectively, the murine and human data continue to support the dual mode of action of T on the male skeleton via both AR and ER α after aromatization (27). Nevertheless, key actions of AR (especially on periosteal bone formation) remain unexplained by cell-specific ARKO models (170–172) (see *Section V*), implying that AR may mediate some of its effects on bone via yet unidentified target cells (see *Section VIII*).

3. Importance of the perinatal T surge

Although skeletal sexual dimorphism is not so evident before puberty, sex steroids influence adult development even in the perinatal period. Especially in the first 2 weeks of life, there is a peak in serum T to low-adult male levels in boys due to high FSH and LH secretion (173). As a result, male mammals undergo neonatal imprinting, which determines male-pattern GH secretion, body weight, longitudinal bone growth, and hepatic steroid metabolism during puberty (174). An important question is whether the perinatal T surge and imprinting of the GH/ IGF-1 axis also regulates radial bone expansion during puberty. Sims et al (175) addressed this in an elegant study comparing hypogonadal mice (hpg, which have a large spontaneous deletion in the gene for GnRH) with intact, orchidectomized, and T-treated mice. At 3 weeks of age (before the start of puberty), hpg mice had lower osteoblast surface and higher osteoid volume and osteoclast surface. At 9 weeks, high-turnover osteopenia was similarly present in both hpg and orchidectomized mice, and T or DHT treatment from 3 weeks inhibited trabecular bone resorption and stimulated periosteal bone formation equally in both groups. However, a short bone phenotype remained as in previous studies (174). Thus, the perinatal T surge and consequent imprinting of the GH/IGF-1 axis has important effects at the growth plate but appears redundant for cortical or trabecular bone (175).

E. Nongenomic signaling

As mentioned in *Section II.D*, sex steroid receptor signaling can deviate from its classical mechanism in two ways: it can be activated in the absence of ligand, or it can have rapid nongenomic effects via cytoplasmatic second messengers.

As mentioned above, $ER\alpha$ transfection in osteoblasts can activate transcription in the apparent absence of estrogens (66), although this is not evident for AR and an-

drogens. Estrogen-independent ER activation in the skeleton was suggested to occur in immature, gonadectomized, or aromatase inhibitor-treated ERE-luciferase reporter mice (67), but intracrine signaling could be an alternative explanation. Several studies discussed in Section VIII show that ligand-independent actions of ER α mediate the osteogenic response to mechanical loading (176– 178). Also, in the study of Almeida et al (179), the unliganded $ER\alpha$ was found responsible for stimulation of osteoblastogenesis and potentiation of Wnt/β-catenin signaling. However, the skeletal phenotype of gonadectomized mice does not differ significantly from ER α KO, $ER\alpha\beta KO$, or ARKO animals, showing that in vivo, remaining ligand-independent ER α or AR actions generally do not compensate for sex steroid deficiency. In the following section, we will discuss studies on the latter, nongenomic mechanisms of sex steroid receptor actions in the skeleton.

1. Knock-in models with impaired DNA binding show that classical pathways have a dominant phenotypic role in bone

Mice expressing a mutant AR without the second zinc finger (AR $^{\Delta ZF2}$) have a similar overall phenotype as other ARKO models but, notwithstanding the limitations of this model (see *Section II.E*), allow the study of nongenomic AR effects. DHT treatment in male AR $^{\Delta ZF2}$ KO mice results in AR-dependent ERK-1/2 phosphorylation and a small further decrease in cortical bone growth (101), whereas DHT has no effect in complete ARKO models (82).

Knock-in mice with an ER α incapable of DNA binding (nonclassical ER knock-in [NERKI]) present an opportunity to study the contribution of ERE-dependent classical signaling. Female NERKI mice had deficits in cortical but not trabecular bone (although the latter may be confounded by elevated T) (94), whereas male NERKI mice had decreases in both trabecular and cortical bone development (95, 96). NERKI mice have decreased osteoblast activation markers and Wnt signaling (consistent with a direct role for estrogens in osteoblastogenesis) (66, 180). They also lost trabecular bone as expected after ovariectomy (94, 181), confirming that classical ERE-dependent mechanisms are responsible for the protean manifestations of estrogen deficiency. Remarkably, ovariectomized NERKI mice gained cortical vBMD compared to wildtype controls (who lost cortical bone as expected), and estrogens suppressed this (94). This again suggests a generally repressive function of non-ERE-mediated signaling on classical ER α signaling, with a paradoxical partial compensation in the complete absence of classical EREdependent signaling. The inhibitory effects of estrogens on

osteoblastogenesis, oxidative stress (see Section VIII.D), and osteoblast apoptosis were unaffected in NERKI mice and thus appear mediated by nongenomic ER α actions via ERKs (181). Interestingly, the majority (72%) of estrogen-regulated genes were unaltered in NERKI mice, suggesting that classical pathways control only a minority of the transcriptional response to estrogens, but that these are the genes critical for the ultimate phenotype (182).

In summary, both AR and ER α can induce non-ARE/ERE dependent signaling, but in both cases, the skeletal effects are small and in opposite direction to the dominant, classical effects.

2. Nongenomic sex steroid receptor agonists

In a seminal series of papers, Kousteni et al (183–185) showed that ER α , ER β , and AR through ligand-dependent, nongenomic signaling can induce bone-sparing and antiapoptotic effects in osteoblasts and osteocytes, regardless of whether the ligands were androgens or estrogens. These effects were accompanied by rapid and transient phosphorylation of common transcription factors (such as ERKs, c-Fos/c-Jun, CCAAT enhancer-binding protein β). Furthermore, a novel class of synthetic ligands called estrens could reproduce these nongenomic bone-sparing effects without stimulating reproductive organs. Substances acting via this new mechanism of action were termed ANGELS (activators of nongenotropic estrogen-like signaling). The same authors later showed that these ER-agonists, similarly to E2 treatment in NERKI mice, preferentially activate kinase signaling and stimulate osteoblasts via Wnt and bone morphogenetic protein (BMP) signaling (186). However, independent groups suggested that estrens do induce sex organ hypertrophy and activate classical signaling through both the AR and ER α (187-190).

E2 dendrimer conjugate (EDC) is a macromolecule in which E2 is attached to a large, positively charged, nondegradable poly(amido)amine dendrimer. This affords unhindered ligand access to ERs and initiation of nonnuclear signaling, without triggering classical nuclear transcription. Recently, the group of Manolagas (191) showed that EDC protects ovariectomized mice from cortical bone loss without affecting trabecular bone or the uterus. EDC achieved this by preventing ovariectomyinduced increased bone turnover and oxidative stress via the aforementioned non-nuclear signaling pathways. The authors suggested that the lack of effect on trabecular bone argues against a direct action of EDC on osteoclast apoptosis in vivo, contrary to their earlier in vitro findings (192). In summary, these promising results with EDC and the cortical bone phenotype of NERKI mice provide further impetus to exploit the benefits of sex steroid signaling as a gender-neutral, tissue-specific, pharmacotherapeutic strategy.

3. GPR30: a membrane ER present in bone cells

As mentioned above, it is still debated whether or not GPR30 is an ER (see Section II.D). GPR30 is expressed in osteoclasts, osteoblasts, and osteocytes but appears to be redundant for the effects of estrogens on most tissues, including fat, reproductive organs, and cortical and trabecular bone in female mice (193, 194), although it may have an effect in the growth plate. Female *Gpr30*-KO mice are shorter and have less body weight and decreased IGF-1 serum levels (195). Ovariectomized *Gpr30*-KO mice have diminished estrogen-mediated growth plate closure (194). In another *Gpr30*-KO mouse model, males also showed delayed growth plate closure and greater body length, but in contrast to previous studies, lean, fat, and bone mass were increased although serum IGF-1 was normal (196). Interestingly, GPR30 expression in the growth plate in humans seems to decline as puberty progresses in both genders (197). Thus, GRP30 may be important to mediate estrogenic inhibition of growth plate chondrocytes, but further studies are needed to delineate the contribution of GPR 30 to sex steroid regulation of male bone metabolism.

F. Independent effects of gonadotropins

Earlier observations that accelerated bone turnover in women precedes menopause by several years, when serum E2 is still normal, led to the hypothesis that other endocrine alterations during the early menopausal transition may be involved, such as FSH, progesterone, androgens, or inhibins (198). This topic received substantial attention when the group of Zaidi (199, 200) observed that neither $FSH\beta$ nor FSH-receptor null female mice exhibit bone loss despite severe hypogonadism, whereas osteoclastic bone resorption is increased in haploin sufficient FSH $\beta^{+/-}$ mice via osteoclastic and immune cell FSH receptors that enhance TNF-α production and osteoclast formation and function. More recently, they and others showed that an exogenous FSH β -subunit neutralizing antibody or an endogenous FSH β immunization strategy prevents ovariectomy-induced bone loss (201, 202). These findings have, however, been challenged by others. Previous reports had suggested that the effects of LH, FSH, and chorionic gonadotropin on bone in mice were indirect via their stimulatory actions on sex steroid synthesis (203). This led to concerns that increased serum LH and T concentrations in the aforementioned models may explain the observed bone preservation (204), which was confirmed by another group (205). Another way of examining this issue is to examine hpg mice, which have a bone deficit entirely correctable by sex steroid replacement (175). Another study

in ovariectomized rats showed that GnRH analog (decapeptyl) treatment did not rescue bone loss, except for a partial recovery of trabecular bone volume (206). Furthermore, female mice overexpressing human FSH on the *hpg* background showed a striking accrual in bone mass via a nonestrogen, ovary-dependent pathway, which probably includes T, inhibin A, and potentially other factors (207). Importantly, the presence of FSH receptors in bone or cultured osteoblasts or osteoclasts could not be confirmed (207). A direct interventional study using aromatase inhibition and the GnRH agonist leuprolide in postmenopausal women found that FSH suppression did not rescue the effects of estrogen deficiency (198).

In summary, the suggested independent effects of gonadotropins remain highly controversial. In any case, it should be noted that all previous reports have been limited to females. In the eagerly awaited bone substudy of the recent investigation by Finkelstein et al (208), an independent role of FSH suppression in men could not be confirmed (209). Thus, for male bone metabolism, any role of gonadotropins independent of sex steroids remains to be proven.

V. Target Cells of Sex Steroids in Bone

Although ER α and AR have been established to be important mediators of the estrogenic and androgenic effects in bone, the target cells for these effects are not completely known. The same goes for the target cell/s for the ER α modulating effect of ER β observed in female mice. ER α , ERB, and AR expression have been described in all resident bone cells including osteoblasts, osteocytes, osteoclasts, and bone marrow stromal cells (although AR expression in osteoclasts remains controversial; see Section V.A.2) (27). During recent years, the Cre-LoxP system has allowed conditional deletion of ER α , ER β , and AR in specific bone cells, which has made it possible to clarify the roles of different cells for the effects of sex steroids in bone. This approach is also favorable because the negative feedback regulation of serum sex steroid levels in these bone cell-specific mouse models is not disrupted, leading to normal sex steroid levels. These cell-specific ER α KO, ER β KO, and ARKO mouse models will be discussed in this Section. Although these mouse models have considerably advanced our understanding of direct sex steroid actions in bone, the Cre-LoxP technique comes with its own limitations (see Section II.E). The promoter driving the recombinase determines at what stage of cell differentiation Cre expression becomes activated (Figure 5). It is important to realize that the phenotype of the mouse models where ER or AR is deleted in early osteoprogenitor cells may also rely on downstream effects in osteoblasts or even osteocytes. However, not all phenotypes of osteoblast- and osteocyte-specific knockout models are reproduced by knockout in earlier stages. Thus, it remains unclear to what extent alternative cell maturation pathways exist, whether opposing effects at different maturation stages cancel each other out, or whether the recombined cells have an altered survival rate compared to the cells where no recombination has occurred.

A. Osteoclasts

1. Estrogen receptor- α

One of the pathways through which estrogens have been shown to reduce bone resorption is by promoting apoptosis and inhibiting differentiation of osteoclasts (192, 210, 211). ER α has been deleted in osteoclasts using the cathepsin K (Ctsk) promoter, which deletes ER α in mature osteoclasts (ER α ^{Oc;Ctsk}) (212), and the lysozyme M (LysM) promoter, which deletes ER α in early osteoclast precursors (ER α ^{Oc;LysM}_{Martin-Millan} and ER α ^{Oc;LysM}_{Seitz} (192, 213) (Figure 5).

The female $ER\alpha^{Oc;Ctsk}$ and $ER\alpha^{Oc;LysM}_{Martin-Millan}$ mice had decreased trabecular bone mass, whereas the cortical bone mass was normal (Table 2) (192, 212, 213). Male mice lacking $ER\alpha$ in osteoclasts have both normal cortical and trabecular bone (192, 212).

The osteoclast number was increased, whereas there was no change in osteoblast number in the female $ER\alpha^{Oc;Ctsk}$ and $ER\alpha^{Oc;LysM}_{Martin-Millan}$ mice (192, 212). The female $ER\alpha^{Oc;Ctsk}$ had increased bone formation and resorption rate, indicating a high bone turnover (212) (Table 2). The molecular mechanism explaining the increased osteoclast number in $ER\alpha^{Oc;Ctsk}$ females was suggested to be disrupted osteoclast apoptosis via Fas ligand (FasL) (212). However, this could not be shown for the mice lacking $ER\alpha$ in the monocyte/macrophage lineage (192). Furthermore, it should not be assumed that FasL is transcriptionally regulated by $ER\alpha$ in osteoclasts, because Krum et al (214) found evidence for a paracrine mechanism by which $ER\alpha$ regulates FasL in osteoblasts, not osteoclasts (see *Section VII*).

 $ER\alpha$ in osteoclasts is hence shown to be of importance for trabecular but not cortical bone in female mice, whereas it has no effect in male mice.

2. Androgen receptor

Thus far, AR expression has not been demonstrated in human osteoclasts, although it has been reported in avian and rodent osteoclasts (27). Studies on androgen modulation of osteoclast generation or activity remain conflicting (27, 215, 216). Because there is no published report of a mouse model lacking the AR in osteoclasts, this is a high research priority.

Figure 5.

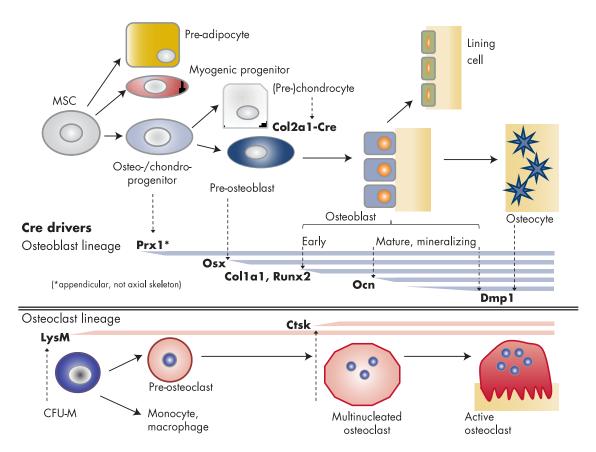


Figure 5. Overview of Cre promoter mouse strains (in bold) that have been crossed with floxed $ER\alpha$, $ER\beta$, or AR transgenic mice to generate conditional knockout models discussed in *Section V*. Osteoblast and osteoclast differentiation are schematically represented above and below, respectively, and the maturation stage at which the respective promoters become active is indicated by the blue and red horizontal bars, respectively. Note that Cre-mediated target gene excision occurring at a specified maturation stage also affects all downstream differentiation stages. *Col2a1*, collagen, type II, α 1; *Prx1*, paired related homeobox 1; *Osx*, osterix (transcription factor Sp7); *Col1a1*, collagen, type I, α 1; MSC, mesenchymal stem cell; CFU-M, colony-forming unit-monocyte.

B. Osteoblast lineage

1. Osteoblasts—ERα

To clarify the importance of ER α in osteoblast lineage cells, six different promoters have been used to delete ER α at different stages in osteoblast/osteocyte differentiation. These promoters have deleted ER α in osteoblast progenitors and all downstream cells (Prx1 and osterix promoter), from the mature osteoblast stage onward (Col1a1 [2.3-kb fragment], Runx2, and osteocalcin promoter) or only in late osteoblasts/osteocytes (dentin matrix protein 1 [Dmp1] promoter) (217) (Figure 5).

In males, cortical bone was transiently reduced when $ER\alpha$ was deleted in osteoblast precursors and downstream cells ($ER\alpha^{\mathrm{Ob;Prx1}}$ mice) (179, 218), whereas no cortical phenotype was evident when $ER\alpha$ was deleted in mature osteoblasts ($ER\alpha^{\mathrm{Ob;Osteocalcin}}_{\mathrm{Määttä}}$ and $ER\alpha^{\mathrm{Ob;Col1a1}}$) (Table 2) (179, 219). These studies demonstrate that $ER\alpha$ in osteoblast precursors regulates cortical bone in young male mice.

Extensive studies by Almeida et al (179) revealed that, in male mice, trabecular bone was not affected when ER α was deleted in osteoblast precursors (ER $\alpha^{\text{Ob;Prx1}}$ and $ER\alpha^{Ob;Osx1}$) or using *Col1a1*-Cre. In contrast, a study by Määttä et al (219) demonstrated that trabecular bone volume was reduced in male mice when $ER\alpha$ was deleted in mature osteoblasts (ER $\alpha_{M\ddot{a}\ddot{a}tt\ddot{a}}^{Ob;Osteocalcin}).$ Although the latter study stated that the osteocalcin-Cre mouse strain used did not have a bone phenotype in itself, this was, as emphasized in a recent review by Manolagas et al (218), not excluded quantitatively. However, a role of ER α in late osteoblasts/osteocytes for trabecular bone in males is supported by the finding that male $ER\alpha_{Windahl}^{Ocy;Dmp1}$ mice have reduced trabecular bone (see Section V.B.2) (220). Based on these studies, we conclude that there is moderate evidence for a role of ER α in mature osteoblasts/osteocytes in the regulation of trabecular bone in male mice.

Cortical bone in female mice is decreased when deleting $ER\alpha$ from the early osteoblast progenitor stage

Table 2. Skeletal Phenotypes of Osteoclast-, Osteoblast- or Osteocyte-Specific Conditional Sex Steroid Receptor Ablation Models

			Male		Female			
Cell Type	Mouse Strain	Ref.	Trabecular	Cortical	F/R	Trabecular	Cortical	F/R
ER-α								
Osteoclast	$ER lpha^{Oc;Ctsk}$	212	\rightarrow	\rightarrow	ND	\downarrow	\rightarrow	Tr: F + R ↑
								Ocl \uparrow , Obl \rightarrow
	$ER \alpha_{Martin-Millan}^{Oc;LysM}$	192	ND	ND	ND	\downarrow	\rightarrow	Tr: Ocl \uparrow , Obl \rightarrow
	$ER\alpha_{Soitz}^{Oc;LysM}$	213	ND	ND	ND	\rightarrow	\rightarrow	ND
Osteoblast	$ER \alpha^{Ob;Prx1}$	179	\rightarrow	((↓))	ND	\rightarrow	\downarrow	Co: Endosteal R ↑, periosteal F ↓
	$ER \alpha^{Ob;Osx1}$	179, 218	\rightarrow	\rightarrow	ND	\rightarrow	\downarrow	ND
	$ER \alpha^{Ob;Col1a1}$	179	\rightarrow	\rightarrow	ND	\rightarrow	\rightarrow	ND
	$ER \alpha^{Ob;Runx2}$	213	ND	ND	ND	\downarrow	ND	ND
	$ERlpha_{M\"a\"att\"a}^{Ob;Osteocalcin}$	219	\downarrow	\rightarrow	ND	\downarrow	\downarrow	Tr: Obl ↓, Ocl ↓
	$ER lpha_{Melville}^{Ob;Osteocalcin}$	221	ND	ND	ND	\downarrow	\downarrow	Tr: Obl \downarrow , Ocl \rightarrow
Osteocyte	$ER lpha_{Windahl}^{Ocy;Dmp1}$	220	\downarrow	\rightarrow	Tr: $F\downarrow$, $R{ ightarrow}$	(↓)	\rightarrow	ND
	$ER\alpha^{Ocy;Dmp1}_{Kondoh}$	222	ND*	ND*	ND	\downarrow	\rightarrow	Tr: Obl \downarrow , Ocl \rightarrow
ER-β								
Osteoblast	$ERoldsymbol{eta}^{Ob;Prx1}$	151	ND	ND	ND	↑	\rightarrow	Tr: Obl, Ocl \rightarrow
AR								
Osteoblast	AR ^{Ob;Prx1}	223	\downarrow	\rightarrow	ND	\rightarrow	\rightarrow	ND
	AR ^{Ob;Osteocalcin}	130	\downarrow	\rightarrow	Tr: $F \rightarrow$, R \uparrow	\downarrow	\rightarrow	Tr: Obl \rightarrow , Ocl \rightarrow
	AROb;Osteocalcin	170	\downarrow	((↓))	Tr: F→, R ↑	ND	ND	ND
	AR ^{Ob;Col1a1}	171	\downarrow	\rightarrow	Tr: R ↑	ND	ND	ND
Osteocyte	AR ^{Ocy;Dmp1}	172	\downarrow	\rightarrow	\rightarrow	ND	ND	ND

Abbreviations: Tr, trabecular; Co, cortical; F, bone formation; R, bone resorption; Obl, osteoblast; Ocl, osteoclast; \uparrow , increased; \downarrow , decreased; \rightarrow , unchanged; ND, not determined; ND*, only BMD determined by DXA; (\downarrow), reduced E2 response in ovariectomized mice but no phenotype in gonadal intact mice; ((\downarrow)), transient reduction.

 $(ER\alpha^{Ob;Prx1}$ and $ER\alpha^{Ob;Osx1})$ (179) as well as from the mature osteoblast stage when using the osteocalcin promoter $(ER\alpha_{Aaitta}^{Ob;Osteocalcin}, ER\alpha_{Melville}^{Ob;Osteocalcin})$ (219, 221) but not with the Col1a1 promoter $(ER\alpha^{Ob;Col1a1})$ (179) (Table 2). Collectively, these studies clearly demonstrate that $ER\alpha$ in the osteoblast lineage is crucial for cortical bone in female mice. Dynamic histomorphometry in female and male $ER\alpha^{Ob;Prx1}$ and female $ER\alpha^{Ob;Osx1}$ mice revealed that there was a reduction in cortical thickness as a result of a decreased mineral apposition rate at the periosteal but not the endosteal bone surface (179). The importance of $ER\alpha$ in osteoblast progenitors at the periosteal surface was concluded to be ligand-independent, and the effects of $ER\alpha$ were suggested to be induced by mechanical strain (179).

Studies by Almeida et al (179) revealed that trabecular bone was not affected in female mice when $ER\alpha$ was deleted in osteoblast precursors and downstream cells ($ER\alpha^{\text{Ob;Prx1}}$ and $ER\alpha^{\text{Ob;Osx1}}$) or in mature osteoblasts using Col1a1-Cre. In contrast, in three independent models ($ER\alpha^{\text{Ob;Osteocalcin}}_{\text{Määttä}}$, $ER\alpha^{\text{Ob;Osteocalcin}}_{\text{Melville}}$, and $ER\alpha^{\text{Ob;Runx2}}$) trabecular bone was reduced in female mice when $ER\alpha$ was deleted from the mature osteoblast stage (213, 219, 221). Although it was not excluded quantitatively in any of these three studies that the osteocalcin-Cre or Runx2-Cre mouse models used have a bone phenotype themselves, collectively we propose that there is at least mod-

erate evidence that $ER\alpha$ in mature osteoblasts contributes to the regulation of trabecular bone in female mice.

As described above, the Cre-LoxP technique has some limitations, and the discordant findings described above regarding the role of ER α in osteoblasts (especially for trabecular bone) could therefore possibly be due to residual ER α expression in the osteoblasts of some mouse models. Alternatively, the cells that are recombined may have a survival disadvantage compared to the fraction of cells that are not recombined. This could lead to a decreased amount of downstream cells with deletion of ER α . Another explanation is that the bone phenotype seen is due to unspecific deletion of ER α also in other cell types. Finally, it is possible that the Cre mouse strains used have a phenotype in themselves, confounding the results. In any case, genetic manipulation of mice, powerful as it is, requires controls that are not always obvious; without such controls, one can come to erroneous conclusions (218).

2. Osteocytes—ERa

Osteocytes are differentiated osteoblasts that are found within mature bone and represent a possible target cell for estrogens. There are two publications (220, 222) where ER α was deleted in osteocytes and late osteoblasts by using the Dmp1 promoter (ER $\alpha_{\rm Windahl}^{\rm Ocy;Dmp1}$ and ER $\alpha_{\rm Kondoh}^{\rm Ocy;Dmp1}$ mice). The young adult male ER $\alpha_{\rm Windahl}^{\rm Ocy;Dmp1}$ mice had a reduced trabecular bone volume but a normal cortical

bone, which is in line with the results from the male $ER\alpha_{M\ddot{a}\ddot{a}tt\ddot{a}}^{Ob;Osteocalcin}$ mice (219). The reduced trabecular bone volume in the male $ER\alpha_{Windahl}^{Ocy;Dmp1}$ mice was due to reduced bone formation (220). The bone phenotype in the male $ER\alpha_{Kondoh}^{Ccy;Dmp1}$ mice was, however, only analyzed with DXA, which showed normal BMD (222).

The female $ER\alpha_{Kondoh}^{Ocy;Dmp1}$ mice had reduced trabecular bone volume but normal cortical bone (222), whereas the female $ER\alpha_{Windahl}^{Ocy;Dmp1}$ mice had both normal trabecular and cortical bone (220). However, when female $ER\alpha_{Windahl}^{Ocy;Dmp1}$ mice were treated with slightly pharmacological doses of E2, they had a substantially attenuated estrogenic response in trabecular bone, suggesting that the deletion of $ER\alpha$ in the osteocytes in these female mice also affects trabecular bone (220). These results are in agreement with the decreased trabecular bone volume observed in the female mouse models lacking $ER\alpha$ in mature osteoblasts (179, 213, 219, 221) (Table 2).

In summary, $ER\alpha$ in osteocytes is not required for cortical bone but seems to regulate trabecular bone in both genders.

3. Osteoblasts—ERB

Thus far, Prx1 and Col1a1 (2.3-kb fragment) have been used to delete $ER\beta$ in specific bone cells ($ER\beta^{Ob;Prx1}$ and $ER\beta^{Ob;Col1a1}$) in only one study, and only the phenotype of female mice has been reported (151). The $ER\beta^{Ob;Prx1}$ mice showed an increase in trabecular bone volume, whereas no difference was observed in the cortical bone. This increase in trabecular bone was accompanied by an increase in osteoblast numbers without any difference in osteoclast numbers, suggesting an increased bone formation rate (Table 2). The female $ER\beta^{Ob;Col1a1}$ had increased BMD in the spine and femur (151). The increase in trabecular bone confirms the inhibitory actions of $ER\beta$ in bone of female mice (150) (see *Section IV.B*).

4. Osteoblasts—AR

The AR has been deleted both in osteoblast progenitors (223) and in mature osteoblasts (130, 170, 171). AR deletion in osteoblast progenitors using the *Prx1* promoter (AR^{Ob;Prx1}) in male mice reduces trabecular bone volume and number but not cortical thickness (223). Deletion of the AR in mature osteoblasts using the *Col1a1* promoter 2.3-kb fragment (AR^{Ob;Col1a1}) (171) or the osteocalcin promoter (AR^{Ob;Osteocalcin} and AR^{Ob;Osteocalcin}) (130, 170) also reduces trabecular bone volume and number in male mice, whereas cortical bone was again unaffected in the AR^{Ob;Col1a1} and the AR^{Ob;Osteocalcin} mice (130, 171) (Table 2). However, a slight and transient reduction in cortical bone was seen in the male AR^{Ob;Osteocalcin} mice when bone turnover peaked at the age of 6 weeks, but no dif-

ference was observed at later ages (170). The difference between AR^{Ob;Osteocalcin} mice could be explained by the fact that Chiang et al (170) deleted exon 3, whereas Määttä et al (130) deleted exon 2 of the AR gene. Although exon 2 deletion leads to a total ablation of AR activity (99, 130), the deletion of exon 3 results in an AR transcript that has preserved DNA-binding capacity and could possibly still mediate AR signaling (see *Section II.E*). The male AR^{Ob;Osteocalcin} mice had an increase in osteoclast but not osteoblast number in trabecular bone, suggesting an increased resorption (130) (Table 2).

In female $AR^{Ob;Prx1}$ mice, the bone phenotype was normal (223), whereas the female $AR^{Ob;Osteocalcin}_{M\ddot{a}\ddot{a}tt\ddot{a}}$ mice had a decreased trabecular bone volume due to a reduced trabecular number (130). Cortical bone was normal in female $AR^{Ob;Prx1}$ and $AR^{Ob;Osteocalcin}_{M\ddot{a}\ddot{a}tt\ddot{a}}$ mice (130, 223).

In conclusion, AR in mature osteoblasts appears to have an important role in the maintenance of trabecular bone in adult male mice, whereas AR in osteoblasts seems to have no significant effect on cortical bone. The impact on trabecular bone in male ARKO mice is probably mainly mediated via AR in osteoblasts. In contrast, the AR target cell responsible for the reduced cortical bone dimensions in male ARKO mice remains currently unknown (Figure 6).

5. Osteocytes—AR

AR expression increases toward osteocyte differentiation, and thus osteocytes are likely target cells for the skeletal effects of androgens (172). AR deletion in late osteoblasts and osteocytes using the Dmp1 promoter (AROcy;Dmp1) decreases trabecular bone volume in the appendicular skeleton in 32-week-old male AR Ocy; Dmp1 mice, due to a reduced trabecular number. Trabecular number was also reduced in the vertebra of male AR Ocy; Dmp1 mice, although trabecular bone volume was normal. Again, cortical bone was unaffected (172). No differences in bone formation, resorption, or the response to mechanical loading could be identified. The decreased trabecular bone volume and normal cortical bone observed in these male mice are in accordance with the results from the mice lacking AR in osteoprogenitor cells and osteoblasts (130, 170, 171, 223). In conclusion, the AR in osteocytes is important for trabecular bone in older mice but not for cortical bone development.

6. Osteoblast overexpression of AR

The bone phenotype resulting from overexpression of the AR in osteoblasts has been evaluated by using two different fragments of the *Col1a1* promoter: 3.6-kb and 2.3-kb fragments (AR-transgene^{Ob;3.6-Col1a1} and AR-transgene^{Ob;2.3-Col1a1}). The 3.6-kb promoter is expressed throughout the osteoblast lineage (and also in stromal precursors and periosteal fibroblasts), whereas the 2.3-kb promoter is expressed in

Figure 6.

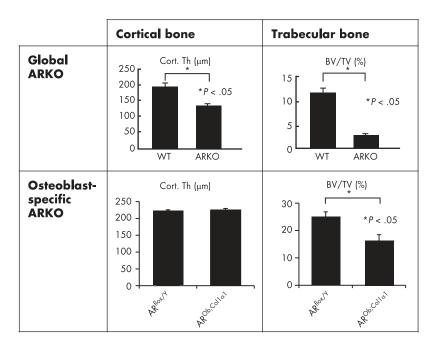


Figure 6. Cortical thickness (Cort.Th) and trabecular bone volume fraction (BV/TV) in global ARKO mice (81) compared to osteoblast-specific ARKO (AR^{Ob;Col1a1}) mice (171). Cortical bone deficits in global ARKO mice are not reproduced in osteoblast-specific conditional models, although clearly and selectively related to androgens (see *Section IV.D*). Thus, the target cell for androgen actions on cortical bone remains to be identified. Trabecular bone is decreased in both global and conditional ARKO models, although less pronounced in the conditional models, probably due to inherent incomplete deletion. [Adapted and reproduced from H. Kawano et al: Suppressive function of androgen receptor in bone resorption. *Proc Natl Acad Sci USA*. 2003;100:9416–9421 (81), with permission. © National Academy of Sciences. And from A. J. Notini et al: Osteoblast deletion of exon 3 of the androgen receptor gene results in trabecular bone loss in adult male mice. *J Bone Miner Res*. 2007;22:347–356 (171), with permission. © American Society for Bone and Mineral Research.]

mature osteoblasts (224, 225). The male AR-transgene Ob; Col1a1 mice in both models had an increased trabecular bone volume due to an increased trabecular number (224, 225). Bone formation rate and mineral apposition rate in the trabecular bone were reduced, indirectly indicating that the increased trabecular bone volume results from decreased bone resorption (224). In the calvaria, AR overexpression resulted in cortical thickening that was most pronounced at the periosteal surface (224). Transgenic males demonstrated a dramatic lack of labeling at the endosteal surface and an increase on the anterior periosteal surface of the femur (224). Because the effect of decreased endosteal bone formation was dominant, cortical bone area and overall bone strength were reduced in both of the AR-transgene Ob; Col1a1 mice (224, 225). The female AR-transgene Ob;2.3-Col1a1 mice did not show any bone phenotype, whereas the female AR-transgene Ob;3.6-Col1a1 only had a very mild phenotype (224, 225).

It is evident from these overexpression studies that the AR in osteoblasts has a direct role in modulating trabecular bone resorption and cortical bone formation in male mice. The decreased trabecular bone resorption is consistent with global and conditional ARKO models. The tendency to increase periosteal bone formation and limit endosteal contraction also confirms a direct role for AR in these processes, although this is not mirrored in conditional ARKO models. The rather complex bone phenotype in the ARtransgene Ob; Col1a1 mice suggests that excessive AR signaling in osteoblasts does not have a positive effect on cortical bone development. Taken together with the results from the global and osteoblast-specific ARKO mice, it is reasonable to believe that neither too high nor too low AR activity is favorable for bone development.

C. Chondrocytes

1. Estrogen receptor- α

Estrogens have long been known to affect both the longitudinal growth spurt in early puberty and the epiphyseal closure, and thereby cessation of growth, in late puberty. These seemingly opposite effects of estrogens on longitudinal growth depend on the maturational stage of the skeleton and the E2 serum levels; low levels of E2 induce the pubertal

growth spurt, whereas higher E2 levels later in puberty induce growth plate closure (226). $ER\alpha$ is chiefly responsible for the effects of estrogens on growth, as discussed in *Section IV.B.*

Growth plate physiology is somewhat different between mice and humans because fusion occurs shortly after puberty in humans but not in mice. Still, it is possible to fuse the growth plates in mice by long-term E2 treatment (227), and their growth plate height decreases with aging. It was previously believed that there was a discrepancy between the man lacking ER α (105, 132) and the mouse model lacking ER α (228) because the human had unfused growth plates and continued to grow, whereas the mouse had fused growth plates. This discrepancy was shown to be due to the first ER α -deleted mouse still expressing low levels of truncated forms of ER α , lacking the $ER\alpha AF-1$ (see Sections II. C and II. E). The E2 levels in this mouse model were elevated, and the truncated ER α was able to induce growth plate closure (164, 228). A study of mice completely devoid of ER α isoforms showed that these mice indeed resembled the ER α KO man in that they had wider growth plates, prolonged growth, and greater ultimate length (164).

Estrogen is a potent regulator of the GH/IGF-1 axis (see Section VII.A), and the effects of estrogens on longitudinal growth and epiphyseal fusion are generally believed to be mediated either indirectly via central regulation of the GH/IGF-1 axis or directly via ERα expressed in the growth plate. To investigate what mechanisms control longitudinal growth, a mouse model lacking $ER\alpha$ specifically in chondrocytes $(ER\alpha^{Chond;Col2a1})$ has been developed (229). Both female and male $ER\alpha^{Chond;Col2a1}$ mice showed a normal growth pattern during sexual maturation, when estrogen activity is low, suggesting that local ER α in the growth plate is not crucial for pubertal growth (229). To investigate whether local ER α is of importance for growth plate closure during late sexual maturation, when estrogen activity is high, the $ER\alpha^{Chond;Col2a1}$ mice were gonadectomized and treated with a high E2 dose. In control mice, this treatment reduced growth plate height, but this effect was not observed in either female or male $ER\alpha^{Chond;Col2a1}$ mice (229). This shows that local ER α in the growth plate is required for the ability of estrogens to reduce growth plate height in both female and male mice (Figure 7). Osteocalcin is also expressed in hypertrophic chondrocytes, which leads to a deletion of ER α in these cells in the ER $\alpha_{\rm M\ddot{a}\ddot{a}tt\ddot{a}}^{\rm Ob;Osteocalcin}$ mice. Analyses of the growth plates in the female ER $\alpha_{\rm M\ddot{a}\ddot{a}tt\ddot{a}}^{\rm Ob;Osteocalcin}$ mice showed that the growth plate height was increased compared to controls (219), which is in agreement with the lack of E2 effect in the ER $\alpha^{\rm Chond;Col2a1}$ mice. Moreover, female ER $\alpha^{\rm Chond;Col2a1}$ mice continued to grow after 4 months of age, whereas essentially no growth was observed in the control mice, resulting in a significant difference in bone lengths at 12 months of age (229).

Male complete ER α KO mice have reduced bone lengths at 17 weeks of age, which was not seen in the ER α ^{Chond;Col2a1} mice (229). This reduction in length was associated with a disturbed GH secretory pattern and low serum IGF-1 levels. Therefore, indirect effects of ER α , probably via the GH/IGF-1 axis, are crucial for a normal growth during sexual maturation. In contrast, local ER α in the growth plate cartilage is crucial for high-dose E2-mediated growth plate closure in adult mice and for reducing longitudinal bone growth in elderly female mice (Figure 7).

Figure 7.

Proposed Role of ERα For Longitudinal Bone Growth B Late during sexual maturation (humans) or Treatment with high E2 doses (humans + mice) (High E2) Hypothalamus Pituitary FRα GH FIGH FIGH FRA Reduced/Cessation of longitudinal bone growth

Figure 7. A, Longitudinal growth during early sexual maturation is mediated by nonbone/noncartilage $ER\alpha$, probably by indirect effects of $ER\alpha$ via the $ER\alpha$ via the $ER\alpha$ in the growth plate cartilage is crucial for high dose $ER\alpha$ in the growth plate closure in adult mice and for reducing longitudinal bone growth in elderly female mice. [Reproduced from A. E. Börjesson et al: The role of estrogen receptor $ER\alpha$ in growth plate cartilage for longitudinal bone growth. *J Bone Miner Res.* 2010;25:2690–2700 (229), with permission. © American Society for Bone and Mineral Research.]

2. Androgen receptor

The AR is expressed in growth plate chondrocytes, but there is no clear evidence that it should be of importance for growth and growth plate closure. So far, there are no reports of a mouse depleted of AR in chondrocytes. Most studies of ARKO mice show normal bone lengths, although one study reported male ARKO mice as having a slightly increased femur length (84), and osteoblast-specific AR-overexpressing mice showed reduced femur length (224). Therefore, although most studies in humans and mice indicate that the effect of T on the growth plate is due to its aromatization and stimulation of ER α , a chondrocytespecific ARKO model could still be of interest.

D. Conclusions from conditional mouse models

1. Target cells of sex steroid actions in males

The studies from total ER α KO, ARKO, and aromatase knockout

mice, together with the different conditional knockout and overexpression models, show that both $ER\alpha$ and AR are required for normal trabecular and cortical bone development (Figure 8A). However, the trabecular and cortical bone compartments are not always regulated by the same bone cells.

 $ER\alpha$ in early osteoblasts has been found to mediate a transient positive effect on cortical bone during growth, although there is no evidence that this effect requires estrogens (179). $ER\alpha$ in mature osteoblasts, osteocytes, and osteoclasts appears not to be important for cortical bone in male mice (Figure 8A). Importantly, although osteoblast-lineage cells may be involved (170, 224), the AR-expressing cell type mediating the regulation of male cortical bone remains largely unknown. It is possible that the AR in osteoclasts or in periosteal cell populations is important, but it is also possible that at least some of the AR bone effects are mediated via target cells outside bone.

For trabecular bone, we suggest that the effect in males comes from T acting via the AR in osteoblast precursors, mature osteoblasts, and (at older ages) osteocytes, which decreases trabecular bone resorption by indirectly affecting osteoclasts (Figure 8A). Estrogens may also have an effect on trabecular bone in male mice by acting via $ER\alpha$ in mature osteoblasts/osteocytes, by increasing bone formation. $ER\alpha$ exerts no actions in osteoclasts in male mice; AR actions in osteoclasts require further investigation.

The regulation of longitudinal growth in male mice is clearly regulated by $ER\alpha$, where the growth plate closure is regulated by $ER\alpha$ in chondrocytes, whereas the longitudinal growth during early sexual maturation is mediated by nonbone/noncartilage $ER\alpha$, probably involving the GH/IGF-1 axis via a central mechanism (Figure 7).

2. Target cells of sex steroid actions in females

 $ER\alpha$ is the most important sex steroid receptor in female bone (Figure 8B and Table 2). We propose that estrogen regulation of trabecular bone is mediated via the $ER\alpha$ in osteoclasts, which directly leads to a reduction of bone resorption. $ER\alpha$ in mature osteoblasts/osteocytes increases trabecular bone formation, which can be inhibited by $ER\beta$.

For cortical bone, $ER\alpha$ actions are evident in early and mature osteoblasts, whereas $ER\alpha$ in osteoclasts and osteocytes is not required. The estrogenic effect mediated by $ER\alpha$ in the early osteoblasts affects endocortical osteoclasts, thereby decreasing endocortical resorption. Ligand-independent activation of the $ER\alpha$ in early osteoblasts increases periosteal bone formation and is also required for the periosteal response to mechanical loading (see also *Section VII.E*).

ER α in chondrocytes in female mice is clearly required for growth plate closure and cessation of longitudinal growth, whereas the longitudinal growth during early sex-

Figure 8.

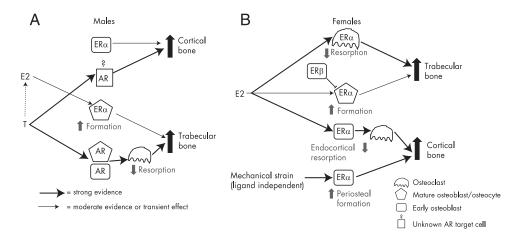


Figure 8. Proposed pathways for ER and AR actions on bone cells. A, In males, ER α in early osteoblasts has a transient effect on cortical bone mass. Some but not all studies indicate that estrogens, via the ER α in mature osteoblasts/osteocytes, lead to an increased bone formation rate that in turn leads to an increase in trabecular bone mass. Acting via the AR in osteoblast lineage cells, T affects the osteoclasts to decrease their resorption rate, which maintains trabecular bone mass. It is known that androgens via the AR stimulate cortical bone expansion, but the target cell for this is still unclear. B, In females, E2 acts via the ER α in osteoclasts to decrease bone resorption, which maintains trabecular bone mass. In addition, some but not all studies indicate that trabecular bone mass is increased by E2 acting via ER α in mature osteoblasts/osteocytes, which induces an increased bone formation rate. ER β in osteoblasts can inhibit this positive effect of ER α in osteoblasts. E2 acting via the ER α in early osteoblasts can affect endocortical osteoclasts to decrease their resorption rate, which leads to an increased cortical bone mass. Cortical bone mass can also be increased via mechanical strain activating ER α in early osteoblasts, in a ligand-independent manner, to increase periosteal bone formation. AR actions have also been reported to contribute to female bone mass, but the physiological significance of these findings is difficult to establish.

ual maturation seems to be mediated by $ER\alpha$ in cells of nonbone/noncartilage origin, probably involving the GH/IGF-1 axis via a central mechanism (Figure 7).

VI. Direct Skeletal Target Genes of the Androgen and Estrogen Receptor

From the previous section, it is clear that ER α exerts direct skeletal effects in osteoblasts, osteocytes, osteoclasts, and chondrocytes, whereas direct AR actions have been demonstrated in osteoblasts and osteocytes. The logical next question regards the downstream mechanism by which these receptors act. As discussed in Section IV.E, important aspects of their effects likely rely on non-ARE/EREdependent mechanisms at the cell membrane, in the cytoplasm, or in the nucleus (like ER α and AR modulation of RUNX2 activity) (see Section VII.C). This section will, however, focus on the traditional direct transcriptional targets of AR and ER in bone cells. Consistent with their role in regulating bone turnover, androgen or estrogen deficiency has been associated with up-regulation of osteoblast and osteoclast markers. For example, a recent microarray study using whole bones isolated from osteocalcin-driven ARKO mice confirmed androgen-responsive expression of Col1a1, Bglap, Ctsk, and Tnfsf11 (RANKL) (230). An elegant example also comes from Wiren et al (231), who examined RNA expression from cortical bone of osteoblast-specific, AR-overexpressing male mice and calvarial osteoblasts continuously treated with DHT. Up-regulation was found for genes like Tnfrs11b (osteoprotegerin), Runx2, Tgfb2 (TGFβ2), Fos, and Jun, and down-regulation of Il6, Il1a, Tnfrsf11a (encoding RANK), and several other nuclear factor-κB, TNF, Wnt, and BMP family members (231). More recently, Khosla's group (232) studied associations between gene expression in human bone samples and estrogen status. But to ascertain whether these transcriptional differences are a direct or indirect consequence of AR/ER signaling and to identify the associated target cells, other techniques are needed. In an excellent recent review on this topic, Krum (233) defined a direct transcriptional target as a gene of which the expression is regulated by AR or ER binding at a promoter or enhancer of that gene. However, evidence from techniques such as chromatin immunoprecipitation (ChIP) remains a major hiatus in the bone field, with a few notable exceptions for ER α target genes in osteoblasts (Table 3). These have advanced our knowledge from previous studies examining early changes in gene expression (after 24 h or less) in cell lines or ex vivo cell cultures (27, 233). To our knowledge, no studies have examined early changes in differential gene expression induced by sex steroids after in vivo stimulation (234).

A. Estrogen receptor target genes

1. ER target genes in osteoclasts

Several E2 targets in osteoclasts have been known from studies in the 1990s, yet surprisingly, systematic studies using microarray approaches or ChIP studies are lacking. In vitro studies using mammalian osteoclasts found E2induced up-regulation of c-Fos and c-Jun within minutes and down-regulation of Ctsk, lysosomal enzyme secretion (cathepsins and tartrate-resistant alkaline phosphatase), and IL-1R within a few hours (27, 233). C-Fos and c-Jun are essential transcription factors for osteoclast development and also belong to the AP-1 family through which $ER\alpha$ signals indirectly (see Section II.C); hence, this has been suggested to represent an amplification mechanism for estrogen signaling (233). Regulation of Ctsk in rabbit osteoclasts has been observed after only 3 hours and is affected by transcription blockers but not the translation inhibitor cycloheximide (233). However, another study using murine osteoclasts could not confirm altered Ctsk expression within 3 hours of E2 treatment (214). Estrogen regulation of FasL was suggested as a mechanism from osteoclast-specific ER α KO (212), but this study relied again on whole bones. However, Krum et al (214, 235) found evidence for a paracrine mechanism by which estrogens regulate osteoblast FasL to induce osteoclast apoptosis (see Section VI.A.2). Another mechanism of estrogen signaling in osteoclasts comes from the group of Manolagas (192), who showed that E2 can stimulate os-

Table 3. Direct Transcriptional Target Genes of $ER\alpha$ in Osteoblasts, Confirmed by ChIP of Target Gene Promoter DNA

Target Gene Regulation		Stimulation	Cell Origin	Refs.	
Fasl	<u> </u>	3 h	Mouse calvaria	214	
ALPL	<u>`</u>	3 h	U2OS-ER α (249), mouse calvaria	236	
Svep1	<u></u>	24 h	Mouse preosteoblastic MBA-15 cell line	245	
IGFBP4	·	24 h	Primary human Ob	246	
ESR1	<u> </u>	15-90 min to 24 h	SaOS2 cell line	247	

teoclast apoptosis via DNA-binding independent actions of $ER\alpha$.

2. $ER\alpha$ target genes in osteoblasts

Krum et al (214, 236) revealed, using ChIP-on-chip, that ER α controls expression of many key osteoblast genes (including alkaline phosphatase (*Alpl*), *Nfatc1* and *Fasl*), nuclear receptors (including Gper1/Gpr30 and Nr5a2), and other transcription factors (including Pax7, Sox5, Foxo1, and Foxo4). Interestingly, whereas FoxA1 serves as a critical pioneering factor for ER α in breast cancer cells, GATA4 may serve this role in osteoblasts (237, 238). More recently, the same authors demonstrated that ER α regulation of matrix metalloproteinase 3 was responsible for osteoblast membrane-bound FasL cleavage and release to mediate osteoclast apoptosis (235). Spelsberg's group (239-244) has identified retinoblastoma-binding protein 1 and TGF- β early inducible gene 1 as highly regulated by $ER\alpha$ and $ER\beta$, respectively, and these factors directly modulate RUNX2 and OPG in osteoblasts as well as important osteoclast pathways. However, confirmation by ChIP is lacking (233). Others found that Svep1 is directly $ER\alpha$ -regulated in a preosteoblast cell line, in collaboration with other transcription factors recruited to its promoter by E2 stimulation, including transcription factor IIB, nuclear factor-κB, and specificity protein 1 (245). The identification of the IGF binding protein 4 (IGFBP4) gene as a direct ER α target (246) underlines the importance of the interplay between sex steroid and IGF signaling. Finally, ER α and AP-1 regulation of ER α itself has been demonstrated in human osteoblasts (247), confirming earlier studies identifying a positive feedback regulation (248).

3. ERB target genes

In a classical study from the group of Spelsberg (249) using inducible ER α or ER β U2OS osteosarcoma cell lines, 24-hour treatment with E2 detected 63 and 59 genes regulated solely by ER α and ER β , respectively, and only 17 by both ERs. Some of the identified ER α target genes were similar to those identified by ChIP (like *Alpl*), but for many of the historically described target genes of estrogens (eg, c-Fos/Jun, cathepsins, IL-1 receptor), we do not know for certain whether they are selective for ER α or ER β .

B. Androgen receptor target genes

As discussed in *Section V*, there are doubts about the in vivo role of AR in osteoclasts. In an early in vitro study, 24-hour treatment of avian osteoclasts up-regulated genes for TGF- β and down-regulated genes for cathepsin B and tartrate-resistant acid phosphatase (250), but further evidence of AR regulation of osteoclast genes is lacking.

No direct AR target genes have been identified by ChIP, although ChIP has been used to demonstrate that AR binding to RUNX2 abrogates its recruitment to Runx response elements (251). In murine calvarial osteoblasts, in vitro stimulation with androgens for 10 hours also up-regulates TGF- β (27, 233). In a more recent microarray study, 12 genes were specifically reported as induced \geq 2-fold after 12-hour DHT stimulation; of five genes tested, three (MYBL2, HOXD11, and ADCYAP1R1) were confirmed by quantitative RT-PCR, and an AR antagonist only abrogated up-regulation of the latter two (252). There is evidence for androgen regulation of IL-6 and OPG (although both stimulation and repression of the latter have been reported), but there is no definitive evidence that these are direct transcriptional targets (233).

In summary, studies using ChIP have begun to elucidate direct transcriptional targets of ER α in osteoblasts. The direct transcriptional targets of AR in bone cells and of ER α in osteoclasts and osteocytes remain, however, poorly investigated. Although most evidence points to a convergence of sex steroid signaling on master regulators like RUNX2, identification of highly regulated downstream targets such as TGF- β early inducible gene 1 or retinoblastoma-binding protein 1 is useful as a proxy for AR/ER activity.

VII. Interaction of Sex Steroids with Other Signaling Pathways

Sex steroids probably control musculoskeletal sexual dimorphism largely by converging onto more fundamental signaling pathways, which by themselves would seem unlikely to have gender-specific set points. This view is supported by the fact that the only gender-specific locus (at Xp22.31) associated with BMD in GWAS meta-analyses is also associated with male serum T levels, whereas no autosomal gene-by-sex interactions have yet been confirmed (25, 26, 40). Furthermore, interfering with master regulatory pathways in bone can abolish skeletal sexual dimorphism, as shown for GH receptor knockout mice, for example (78). Numerous animal and human studies have suggested gender-specific effects in other pathways (eg, Wnt signaling, discussed in Section VII.D), but whether these observations represent downstream consequences of cross talk with sex steroids rather than independent causes of sexually dimorphic bone metabolism requires further investigation.

A. Growth hormone and IGF-1

1. Physiology

Bone matrix is a storehouse replete with polypeptide growth factors, among which IGFs and TGF- β feature

most abundantly. Under control of pituitary GH, the liver produces systemic IGFs that exert endocrine actions on bone and cartilage, but osteoblasts and chondrocytes also secrete these factors locally in an autocrine/paracrine pathway (253). Osteoclast digestion of bone's extracellular matrix releases growth factors like IGFs, which act as coupling factors by stimulating osteoblast refilling of the resorption cavity (254). Human bone contains about 10fold higher concentrations of IGF-2 than IGF-1, but the latter is much more potent. The activity and storage of IGFs is regulated by no less than six IGFBPs: IGFBP-3 and -5 can form a ternary complex with the acid labile subunit. Because IGFBPs are produced in molar excess, less than 1% of IGF-1 circulates as a free hormone (255). However, IGFBPs also prolong the half-life of IGF-1, thus enabling its activity, as evidenced by reduced bone length and width in humans and mice with defects in the acid labile subunit (256). The precise role of IGFBPs may be even more complex and remains incompletely understood; they may also enhance IGF activity by enhancing extracellular matrix storage or even have intracellular and/or IGF-independent effects. Studies in rodents dissecting the skeletal influence of different components of the GH/IGF-1 system have been reviewed elsewhere (253, 254). Briefly, genetic models of GH or IGF-1 deficiency display osteopenia with reduced periosteal bone expansion and longitudinal bone growth. Conditional overexpression of IGF-1 or deletion of the GH or IGF-1 receptor, either ubiquitously, in the liver or locally in osteoblasts and chondrocytes has shown that: 1) IGF-1 has both GH-dependent and -independent effects; 2) GH has direct effects on osteoblasts as well as on hepatic IGF-1 production; and 3) IGF-1 acts both in an endocrine and intracrine/paracrine manner. Furthermore, IGF-2 knockout mice have only modest reductions in cortical bone size compared to the previous models (257). Finally, paracrine overexpression of IGFBP-3, -4, and -5 has been shown to impair osteoblast function (255).

2. Sex steroid regulation of local IGF signaling in bone

IGF-1, IGF-2, their receptors, and several IGFBPs have been suggested to be regulated by androgens and estrogens in vitro in some studies, although the data remain as conflicting as a decade ago (27, 66, 258). For example, E2 treatment increased IGF-1 expression in osteoblastic U2OS cells stably transfected with ER α but not a NERKI vector, suggesting an ERE-dependent pathway (66). However, E2 had no direct effect on IGF-1 expression in rat osteoblasts in vitro; rather, ER α suppressed IGF-1 induction by other hormones indirectly via contact with other transcription factors (258). In a highly AR-expressing human fetal osteoblast cell line, DHT increased cell proliferation and expression of IGF-1 and IGFBP-2 and -3 lev-

els, whereas IGFBP-4 decreased (259). Another group, however, showed that DHT increased TGF- β but not IGF-1 or IGF-2 in conditioned media of murine calvarial osteoblast cultures, although IGF-2 responsiveness was increased due to increased expression of its receptor (260, 261). IGFBP-4 has been confirmed by ChIP as a direct ER α target gene in osteoblasts (see *Section V.B*). Gender differences in the skeletal response to acid labile subunit deficiency have been described in mice, but regulation by sex steroids has not been examined (256). Estrogens may also enhance GH signaling in human osteoblasts, at least in vitro (262). No firm conclusions can be drawn from the conflicting evidence available.

3. Indirect skeletal effects of sex steroids via the somatotropic axis

The GH/IGF-1 axis is considered critical in determining sexual dimorphism in both longitudinal growth and periosteal bone expansion. The perinatal T surge is believed to cause imprinting of the hypothalamic GnRH pulse generator and consequent male-type GH secretion patterns during adulthood (see also Section IV.D) (175). At the start of puberty, the GH pulse amplitude rises, IGF-1 increases, and both contribute to an acceleration in longitudinal and transversal bone growth. This occurs later in boys but lasts longer, resulting in enhanced pubertal crosssectional and longitudinal bone development. However, more recent studies show that gender differences in adult GH secretion are determined not only perinatally and during puberty but also during adulthood. Indeed, experimental alterations in the gonadal steroid environment in adulthood rapidly alter GH pulsatility patterns (263). Male-pattern GH secretion may also be more directly involved in periosteal bone expansion than previously realized, as evidenced by a striking feminization of bone geometry in male mice with conditional GH-receptor knockout in osteoblasts (264).

As mentioned in *Section II.E*, Tfm rats display decreased circulating IGF-1, but this is not the case in orchidectomized or ARKO mice (82, 84); thus, liver-derived IGF-1 cannot account for decreased periosteal bone formation in the latter two models (although disrupted local IGF-1 production has not been ruled out). Conversely, male ER α KO, aromatase knockout, ER α β KO and aromatase inhibitor-treated mice all show decreased IGF-1 and radial bone expansion despite increased androgens (see *Section II.E* and Table 1); thus, androgens seem to regulate systemic IGF-1 in males via aromatization and activation of ER α . Female ER β KO mice had increased IGF-1 levels in some studies; the inhibitory effect of ER β on female bone (see *Section IV.B*) could thus be related to restraints on IGF-1.

Callewaert et al (78) dissected the relative contributions of androgens, aromatization, and IGF-1 for skeletal sexual dimorphism in a careful longitudinal study. Skeletal sexual dimorphism was mainly established in early puberty (age 3–5 wk), when male mice have higher periosteal and lower endocortical bone formation, along with higher serum IGF-1 compared to females (78). From age 6 weeks, circulating IGF-1 was not significantly different between genders, and body weight and bone cross-sectional area increased in parallel. GH-receptor knockout mice, on the other hand, showed very low IGF-1 and no skeletal sexual dimorphism. Androgen deficiency did not decrease IGF-1 and only altered radial bone expansion during late puberty. Treatment with an aromatase inhibitor, however, significantly reduced IGF-1 and radial bone expansion from early puberty in male mice. Along the same lines, E2 was found to be a major positive regulator of hepatic IGF-1 synthesis in male GH-receptor knockout mice, which fully restored periosteal bone formation in these mice (265). Ovariectomy increased radial bone expansion during early puberty in females independent of IGF-1 levels, as confirmed by others (266). Thus, previous observations that (bioavailable) E2 levels correlate with the earlier onset of puberty in females by direct activation of their GH/IGF-1 axis and consequent bone development may be confounded by other factors, eg, leptin or signals from the central nervous system.

In conclusion, these mouse studies suggest that skeletal sexual dimorphism is mainly established in early puberty, when the stimulation of radial bone expansion by androgens in males appears to depend partially on both aromatization and IGF-1, whereas in females, estrogens stimulate endocortical bone formation and limit periosteal expansion independent of IGF-1 (Figure 3). In late puberty, androgens maintain periosteal bone formation in males independently of systemic IGF-1 (267). This fits with human observations that aromatizable androgens are more effective to stimulate bone growth in boys with delayed puberty, whereas only aromatase- or ER α -deficient men lack a pubertal growth spurt but not AIS individuals (254). Estrogens appear to exert a biphasic influence on periosteal bone formation and longitudinal bone growth (see Section V.C). In both genders, low levels during early puberty have stimulatory effects on the GH/ IGF-1 axis, but this effect lasts longer in boys. In late puberty, growth plate chondrocytes and periosteal expansion are directly inhibited by high estrogen levels, which are reached more quickly in girls (268).

4. Involvement of GH/IGF-1 in male osteoporosis

From the previous section, it is clear that sex steroids interact with the GH/IGF-1 axis to determine sexual di-

morphism during pubertal bone development. It is not clear, however, whether sex steroids also influence bone maintenance in older men via the somatotropic axis there is even uncertainty whether the somatotropic axis plays a role altogether (33). Observational studies have shown that IGF-1 decreases with age in both genders, but both positive association and no association with bone turnover or BMD in men have been reported, as well as associations only in women or only in men in different studies (269-271). Recently, Ohlsson et al (272) found that older men with IGF-1 below the median had increased risk of incident hip and vertebral fractures. However, most of these studies showed no association between IGF-1 and sex steroid levels (270, 272). Although treatment with an aromatase inhibitor decreases IGF-1 in younger men and aged male rodent models, this has either not been confirmed (273) or has been left unreported in elderly men (274–276). This issue should be further examined in human interventional studies.

B. Vitamin D and PTH

PTH regulates formation of the active vitamin D metabolite 1,25-(OH)₂-vitamin D, and both maintain serum calcium by stimulating bone resorption, intestinal calcium absorption, and renal calcium reabsorption (277). Secondary hyperparathyroidism, low serum 25-OH-D and increased levels of 1,25-(OH)₂-vitamin D have shown independent and additive associations with bone loss in men (278, 279). At least in postmenopausal women, estrogens stimulate PTH-induced calcium absorption and reabsorption, although their overriding effect is a PTH increase due to decreased calcium mobilization from the skeleton (23). Whether the same holds true in men remains unconfirmed.

In MrOS USA, 25-OH-vitamin D levels were associated with hip BMD loss only when <20 ng/mL in men >75 years of age (280). In a case-control analysis, 25-OH-vitamin D levels were associated with incident hip fracture only, and not with other types of fractures (281). Isolated vitamin D deficiency without sex steroid deficiency was not related to any skeletal measure, and incident nonspine fractures were not related to low bioavailable T/E2 or high SHBG alone; only concurrent vitamin D deficiency and low bioavailable E2 were associated with these fractures (282). The authors concluded that the adverse effects of sex steroid deficiency were more pronounced in older men with low 25-OH-vitamin D levels. However, interventional studies are needed to confirm causality because in some other studies, androgen and vitamin D deficiency were associated (283, 284). In a small randomized trial, vitamin D supplementation increased (total and bioactive) T levels in healthy overweight men in a weight reduction program (285). Interestingly, DHT has been shown to

increase 1α -hydroxylation of 25-OH-vitamin D in osteoblasts from men, whereas E2 had no effect (286). In mice, it was shown that males have higher calcium excretion along with increased expression of renal calcium transporters, which are both decreased by orchidectomy and restored by T replacement; PTH and 1,25-(OH)₂-vitamin D were not affected (287). A cross-sectional study in MrOS USA and Sweden found that both T and E2 were independently associated with lower serum phosphorus levels in men (288).

In summary, sex steroid deficiency might amplify the association between vitamin D deficiency and fracture risk in older men. There might exist more direct interactions between the gonadal and calciotropic axis in men, which could have important repercussions for skeletal metabolism, but the existing data remain hypothesis-generating.

C. Biphasic modulation of RUNX2 activity

The interplay between RUNX2 and sex steroids has been excellently reviewed by Frenkel et al (289). Briefly, both ER α and AR have been shown to directly bind and modulate the transcriptional activity of RUNX2 (formerly CBFA1, a master transcription factor in osteoblast and chondrocyte differentiation) (Figure 9). In late-stage MC3T3-E1 osteoblast cultures (when RUNX2 activity is maximal; Figure 9A, right side), liganded ER α can bind with its DBD to RUNX2, hijacking its recruitment and activity on Runx DNA response elements (290). In early MC3T3-E1 osteoblasts, however, ER α binding has been found stimulatory to RUNX2 activity and repressive to ERE activation (291, 292) (Figure 9A, left side). These findings may seem contradictory but have been suggested as indicative of a stage-specific, biphasic regulation by

which $ER\alpha$ keeps RUNX2 in check, preventing both insufficient and excessive RUNX2 activity. AR, also through its DBD, binds RUNX2 even more strongly than $ER\alpha$, and a mutual repression of their subnuclear localization and transactivational capacity has been reported (251, 292) (Figure 9B). RUNX2 in turn regulates expression of several major osteoblast proteins such as osteocalcin, Col1a1, bone sialoprotein, osteopontin, osterix, and matrix metallopeptidases. Furthermore, the aromatase gene has recently been reported as a downstream target of RUNX2 in bone (293). Because AR, ER α , and RUNX2 are beneficial for osteoblast development and bone formation, a repressive action of AR and ER α on RUNX2 target genes may seem counterintuitive at first. However, attenuation of RUNX2 activity may prevent the deleterious effects of excessively high bone turnover as seen in transgenic mice overexpressing RUNX2 in osteoblasts, whereas transgenic mice expressing a dominant-negative RUNX2 resist ovariectomy-induced bone loss (294).

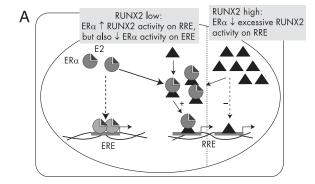
In summary, sex steroid nuclear receptors and RUNX2 can heterodimerize and are proposed to exert a mutual and biphasic modulation of each other's activity, which prevents both overstimulation and (as demonstrated for $ER\alpha$) underactivity (289).

D. Wnt signaling and sclerostin

1. Physiology

Genetic investigations into the cause of several high bone mass disorders revealed an important role for the Wingless-type mouse mammary tumor virus integration site (Wnt) pathway in bone formation control (295). Briefly, canonical Wnt signaling involves several ligands

Figure 9.



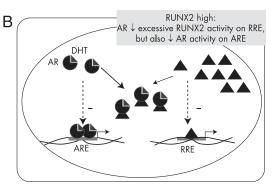


Figure 9. A, ER α and B, AR may interact with other nuclear transcription factors, thus binding and influencing chromatin indirectly. This is illustrated by the interactions between ER α and AR on the one hand and RUNX2 on the other hand. ERE, estrogen response element; ARE, androgen response element; RRE, Runx response element. [Adapted and reproduced from T. L. McCarthy et al: Runx2 integrates estrogen activity in osteoblasts. *J Biol Chem.* 2003;278:43121–43129 (291), with permission. © American Society for Biochemistry and Molecular Biology. And from B. Frenkel et al: Regulation of adult bone turnover by sex steroids. *J Cell Physiol.* 2010;224:305–310 (289), with permission. © Wiley Periodicals Inc.]

binding at the cell surface with Frizzled and low-density lipoprotein receptor-related proteins (LRP) 5 and LRP6 as coreceptors, activating downstream factors like GSK3 β and LEF1 and ultimately resulting in intracellular β -catenin activation. The result is a striking increase in bone formation, which requires regulation by endogenous inhibitors like sclerostin (encoded by the SOST gene and almost exclusively expressed in osteocytes), Dickkopf-1, soluble frizzled-related proteins, and Wnt-inhibitory factors.

2. Sexual dimorphism in mouse models of the Wnt/β-catenin pathway

Several studies have shown gender differences resulting from genetic manipulation of the Wnt/ β -catenin pathway, but few studies have yet examined whether this is directly related to sex steroid signaling or their interaction with bone mass, turnover, or response to mechanical loading (see Section VII.E). For example, trabecular bone volume showed greater increases in female compared to male Sost knockout mice (296). Conversely, progression of bone loss was generally faster and more severe in female compared to male Dmp1-Cre conditional β -catenin knockout mice (297), and in a haploinsufficient model, female mice showed markedly decreased trabecular bone volume, whereas males were unaffected (298). Along the same lines, Gsk3\beta and Lef1 haploinsufficient females displayed enhanced and reduced trabecular bone mass, respectively, whereas males were unaffected (299). Interestingly, Lef1 haploinsufficient males did show low bone mass on a *Tfm* background, although the AR may not be completely disrupted in this model (see Section II.E). On the other hand, secreted frizzled-related protein overexpression decreased trabecular bone to a greater extent in male mice (300). Osteocyte-specific deletion of the transcription factor myocyte enhancer factor 2c (involved in sclerostin expression) reduced sclerostin to a greater extent in male mice, resulting in more pronounced cortical and trabecular bone gains in males (301). Female mice with activating *Lrp5* mutations were slightly more load-responsive and less sensitive to disuse compared to males (302).

As mentioned above, several studies have suggested that sex steroid actions alter expression of Wnt family members, although direct regulation needs to be confirmed by ChIP. At least in vitro, Almeida et al (179) showed that ERα stimulation of periosteal cells occurs via potentiation of Wnt signaling. Androgens have also been shown to increase differentiation of MC3T3 preosteoblasts with an accompanying increase in Wnt signaling, and soluble frizzled-related protein can inhibit this (303). Several in vivo studies have suggested sex steroid regulation of sclerostin. Ovariectomy decreased skeletal scleros-

tin expression in one study (304), whereas another study came to the opposite conclusion: ovariectomy increased skeletal sclerostin expression, whereas E2 prevented this (305). The former study found some variation, depending on the type of bone and the time point examined.

In conclusion, male mice have displayed either enhanced or decreased sensitivity to manipulation of the Wnt/ β -catenin pathway, depending on which component is genetically modified. ER α and AR have been shown to potentiate Wnt signaling in osteoblasts in vitro. Future in vivo studies are needed to examine whether sex steroid actions involve a direct, short-term regulation of Wnt/ β -catenin signaling, and whether this contributes to skeletal gender differences. Clinical trials may in the future reveal whether or not sexual dimorphism in the skeletal response to Wnt/ β -catenin manipulation also exist in humans, although this pathway is most likely of great importance in both genders.

3. Role of circulating sclerostin

Sclerostin serum levels have been examined in several studies, but the results differ markedly depending on the immunoassay used and should thus be interpreted cautiously (306). In animal models (304) and more recently in humans (307), discordance has been noted between circulating sclerostin and expression locally in bone. Several studies found higher (instead of lower) sclerostin serum levels in men compared to women, and a positive (instead of a negative) association with age, wholebody bone mineral content, and HR-pQCT parameters (306, 308-314). The gender difference in circulating sclerostin seems to be established during puberty, and serum sclerostin tends to decline in late puberty in both genders (314). In the MINOS study, older men with higher serum sclerostin had lower fracture risk, even independent of BMD and age (310). Although not unequivocally (312), most observational (310) and also interventional studies showed that E2, and not T, regulates serum sclerostin in elderly men and both circulating sclerostin and intraskeletal mRNA in postmenopausal women (232, 315). This correlated with bone resorption but not formation markers. Taken together, these results suggest that circulating sclerostin is more closely related to the amount of bone (and probably osteocytes) present and may also reflect increased bone turnover per se, but sclerostin serum levels do not necessarily reflect its known role as a local inhibitor of bone formation. Circulating sclerostin was related to estrogen status in both genders in most studies, but whether sex steroids also regulate local sclerostin signaling in male bone and whether this represents a direct AR/ER effect requires further study.

The interaction between sex steroids and the Wnt/sclerostin pathway in the osteogenic response to mechanical loading is discussed in the next section.

E. Physical activity and response to mechanical loading

In a seminal study, the group of Lanyon (316) showed that female ER α KO mice displayed decreased periosteal and endosteal bone formation after mechanical loading and that strain induced less proliferation in osteoblast-like cells derived from these animals. Load-induced cancellous bone formation on the other hand is normal in these female ER α KO mice (317). Strain-induced osteoblast proliferation is also prevented by an ER α antagonist (177). The antiapoptotic effect of mechanical stimulation on osteoblasts and osteocytes, however, seems to require ligand-independent actions of both ER α and ER β (176). In vivo, the cortical osteogenic response in female mice was enhanced by tamoxifen (an ER α agonist in bone, but a pure ER β antagonist), and this effect could be abrogated by adding fulvestrant (which inhibits both ERs and had no effect on its own). ER β KO had the opposite effect, ie, increased cortical loading response, in line with the known inhibition of normal ER α functions by ER β (see Section IV.B) (317). Ovariectomy on its own does not reproduce this effect (318). Along the same lines, several studies in rats showed that ovariectomy increases and E2 decreases load- or exercise-induced bone formation (319–323), whereas another study showed no interaction (324). A recent study confirmed that these effects are indeed ligandindependent and further showed that ER α AF-1, but not AF-2, mediated the osteogenic response of cortical bone to mechanical loading (178). Furthermore, loading did not induce in vivo luciferase activity in ERE-Luc reporter mice (178). Thus, the osteoanabolic effects of mechanical loading are enhanced in female mice by ER α actions via its AF-1 and nonclassical signaling.

In male mice, however, the group of Lanyon (317) showed that ER α KO increased both the cortical and trabecular response to mechanical loading (contrary to the situation in female mice), whereas ER β KO increased the cortical response (similarly to what was observed in females). In an earlier study using the Korach's ER α KO model and ulna loading, however, there was no effect of ER α on the osteogenic response to loading in male mice (325). Apart from methodological issues, the latter difference could also imply that ER α -AF-1 might be involved in males, too, but this requires confirmation. In ARKO male mice, the cortical response to loading is increased (325). Whether this effect is ligand-dependent or not requires confirmation.

The exact mechanism upstream of ER α linking it to mechanical stimuli remains unidentified. The early Wnt/

β-catenin response to mechanical loading seems to require ER α , at least in vitro (326). Almeida et al (179) confirmed this in periosteal cells from $ER\alpha^{Ob;Prx1}$ and $ER\alpha^{Ob;Osx1}$ mice, which showed blunted proliferative and osteoblastogenic responses to Wnt3 stimulation. E2 on the other hand had no effect on proliferation and even attenuated Wnt-induced osteoblastogenesis, showing that the receptor has independent or even opposite effects to those of its ligand (179). Suggested downstream effects explaining the increased loading response include sensitization of the IGF-1 receptor via ERα and down-regulation of Sost/ sclerostin via ER β (whereas ER α tends to antagonize this) in female mice (327). In male ARKO mice, a stronger inhibitory effect of loading on sclerostin expression and a higher nitric oxide production by osteocytes in vitro were observed (325).

There are several methodological caveats in these studies, including the reliability of strain-gauge techniques in one cortical site (but not in other sites or in trabecular bone), reporting relative differences, which obscures baseline absolute differences, and confounding, for example, by background physical activity (328). For example, ARKO mice have decreased physical activity and lack of motivation, whereas forced activity improves their musculoskeletal deficit (106). Interestingly, spontaneous locomotor activity was increased in gonadally intact but not orchidectomized central nervous system (Nestin-Cre) ARKO mice, suggesting a role for increased estrogen levels in this model (108). This is in line with evidence that female mice and rats have higher voluntary running activity than their male counterparts (329, 330).

The osteocyte would be an obvious candidate target cell given its mechanosensitivity and mechanotransduction capabilities. Strain has been shown to induce nuclear and membrane localization of ER α in vitro in osteocytes (331), and E2 enhances connexin 43-based gap junction intercellular communication in the osteocyte-like MLO-Y4 cell line (332). However, both the osteocyte-specific ER α KO and ARKO mice showed no altered loading responses (172, 220). Instead, ER α stimulation of mechanoresponsiveness in periosteal cells has been proposed to be responsible for the cortical bone deficits observed in osteoprogenitor- and early osteoblast-specific conditional knockout models (179).

In summary, the cortical osteogenic response to mechanical loading in female mice requires $ER\alpha$ via its AF-1 and ligand-independent, non-ERE-mediated signaling. More studies are needed in male mice, but there seems to be an opposite, inhibitory effect of AR. Many questions remain regarding the target cells, the molecular mechanism, as well as the importance of these findings (not the least of which is their generalizability to humans). Further

studies on the interaction between hormones, mechanical signals, and physical activity in bone regulation are eagerly awaited.

VIII. Indirect Effects of Sex Steroids on Bone

Because the skeletal phenotype of bone cell-specific ARKO and ERKO models are generally mild compared to their ubiquitous counterparts, it follows that sex steroids might have important indirect effects on bone via other tissues. The primary suspects in this regard are muscle and the nervous system. In vivo evidence from muscle- and neuron-specific ARKO/ERKO models will be reviewed in the following paragraphs. Perhaps other tissues known to affect bone metabolism like fat, the gut, the kidney, etc, may also be involved in sex steroid control of bone metabolism, which needs to be investigated in future studies.

A. Muscle

Given the well-known trophic and anabolic effects of androgens on muscle in humans (208), as evidenced most strongly by androgen abuse in weight lifters (333), the possibility arose that some of the effects of androgens in rodent models might indirectly result from bone-muscle interactions (334). In ARKO mice, males (but not females) have impaired muscle development (83). Additional $ER\alpha KO$ further reduces muscle mass in ARKO males (84), in line with other evidence for an independent effect of estrogens on muscle in mice and rats (135, 335). In humans, however, T alone via the AR seems sufficient for maintaining muscle mass in older men because neither aromatase nor 5α -reductase inhibition diminishes the efficiency of T replacement (208, 336, 337). However, bone and muscle development are highly integrated and controlled by common signals including IGF-1 (338). Musclespecific ARKO mice do not show altered bone metabolism, and even appendicular muscle mass is only slightly reduced, possibly because only perineal muscles display high AR content and androgen regulation in mice (339-341). In human studies, the effects of androgens on bone could be examined by correcting for muscle parameters. In male-to-female transsexuals, for example, bone mass seems to be maintained during pharmacological estrogen therapy, despite substantial muscle loss related to androgen deficiency (75). Such associations are interesting, but they offer no definitive proof that estrogens alone are sufficient for skeletal maintenance in older men.

Although the effects of androgens on bone in mice are unlikely to be mediated via the AR in muscle, it remains unclear whether sarcopenia in older men with late-onset hypogonadism contributes directly or is merely associated with bone loss and fractures (32, 33, 342). Future studies are necessary to examine whether maintaining or increasing muscle mass is useful to prevent falls, bone loss, and ultimately fractures. In any case, exercise seems an effective countermeasure for hypogonadal bone loss in preclinical models (106). Conversely, hypogonadism is commonly seen in clinical situations of disuse, and nandrolone or T have been found beneficial for bone and muscle decay in the absence of mechanical stimuli in some preclinical studies (343–345), but not others (346). Whether these effects are mediated by muscle or skeletal AR requires further investigation.

B. Nervous system

It has long been known that bone cells are connected to sympathetic, parasympathetic, and sensory nerve fibers and contain various receptors for neurotransmitters, although the notion that these nervous systems control bone metabolism is still controversial. Sex steroid receptors in turn are expressed in neurons throughout the nervous system, opening the possibility that sex steroids may affect the skeleton via the nervous system (108, 347). Interestingly, ovariectomy has been demonstrated to produce a dramatic decrease of innervation in the rat tibia (348). In some mouse models, bone loss due to estrogen deficiency was diminished when nervous signaling was disrupted, although other studies could not confirm this (349).

Farr et al (350) recently demonstrated, for the first time in humans, an increase in sympathetic activity in postmenopausal women, which was negatively associated with trabecular microstructure and bone strength.

A role of estrogen signaling in nervous tissue for bone metabolism was recently suggested by an investigation in female mice lacking $ER\alpha$ in neuronal cells (351). Pan-neuronal $ER\alpha$ inactivation using Nestin-Cre resulted in increased bone mass, indicating that estrogen signaling in neuronal cells may have a negative impact on bone mass in contrast to the positive effects of peripheral estrogen signaling. The exact mechanism underlying the increased bone mass observed in this study was not completely understood, but a subsequent study suggested that pro-opiomelanocortin neurons were involved (mainly found in the arcuate nucleus in the ventral hypothalamus, a region previously implicated in regulation of bone mass) (352).

In conclusion, apart from their well-known actions on the gonadotropic and somatotropic neuroendocrine axis, estrogens may influence bone indirectly via $ER\alpha$ signaling in other neuronal cells. Future mechanistic studies in mice as well as more studies in humans are needed to confirm whether and how both estrogens and androgens affect neuronal regulation of bone metabolism.

C. Adipose tissue

According to the traditional view, obesity, both through increased mechanical forces imposed on the skeleton and estrogen production via adipocyte aromatization, has beneficial effects on BMD, and thus fracture risk. This paradigm has, however, been challenged recently by reports demonstrating unaltered fracture susceptibility in obese men despite higher BMD (353). Moreover, the relationship between adiposity and bone mass may depend on gender, age, menopausal status as well as type of adipose depot (sc vs visceral) (354). Premenopausal women predominantly accumulate sc fat that is positively correlated with bone mass; men, however, accumulate visceral fat, which associates negatively with BMD (35). Marrow fat can also be assessed with novel imaging techniques and has, in most reports, been found inversely correlated with bone parameters (35). However, thus far no prospective study has related marrow fat to fracture.

T promotes a reduction in total body fat and visceral adiposity in elderly men (355–357). Finkelstein et al (208) have suggested that increased body fat in male late-onset hypogonadism is dependent on reduced E2. However, anabolic doses of nonaromatizable androgens have also demonstrated independent lipolytic actions of AR (358, 359), which may depend on shifting mesenchymal precursor cells away from adipogenesis and toward osteogenic and myogenic differentiation (360). Global inactivation of ER α and AR in mice of both sexes results in obesity (84, 107, 361, 362). In contrast, ERβKO mice of both sexes have normal adipose tissue under normal diet conditions (363). Sex steroid receptors are expressed in adipocytes, and recently, adipocyte-specific ARKO mice were shown to have impaired insulin secretion and increased susceptibility to visceral obesity (364). Plasma levels and adipocyte expression of adipokines like leptin, adiponectin, and resistin have been associated with circulating sex steroid levels in humans, and receptors for these adipokines are expressed in osteoblasts and osteoclasts and regulate bone mass in transgenic mouse models (353).

In conclusion, adipose tissue influences the skeleton, and this effect seems depot-specific. Sex steroids also exert depot-specific effects on adipocytes and may regulate adipokine secretion. Whether effects of sex steroids on bone also involve indirect effects via AR or ER in adipocytes remains to be established.

D. Oxidative stress

Reactive oxygen species (ROS) are natural products from the aerobic metabolism of cells, but they can also be generated from oxidation of fatty acids or as a response to external stimuli, eg, inflammatory cytokines, growth factors, or UV light. Although high levels of ROS are harmful, low levels are important for triggering essential signaling cascades. Oxidative stress is caused by excess ROS, which cumulatively leads to age-related damage in many tissues including bone. Generic aging mechanisms including oxidative stress and their link with sex steroids were recently the subject of an excellent review by Manolagas (22), to which we refer for a more extensive summary on this topic.

ROS influence the generation and apoptosis of osteoclasts, osteoblasts, and osteocytes, and it has been shown that oxidative stress is temporally linked to reduced bone mass and strength in aging mice (which remain sex steroid sufficient) (210). Estrogens and androgens may protect against bone loss by increasing defenses against excessive oxidative stress. Gonadectomy of both female and male mice increases ROS in the bone marrow. These changes can be reversed by E2 (females), DHT (males), or an antioxidant (210). Antioxidant treatment also prevents the gonadectomy-induced spinal BMD loss in both male and female mice (210). In vitro studies using osteoblastic cells confirm that E2 and DHT attenuate osteoblast apoptosis induced by oxidative stress (210). There is also evidence that E2 and DHT restrain osteoclastogenesis and stimulate osteoclast apoptosis by reducing oxidative stress via promotion of antioxidative pathways (210). The effect of estrogens on oxidative stress does not appear to involve binding of the ER α to DNA response elements because E2-treated ovariectomized NERKI and control mice show a similar reduction of ROS in bone marrow (181). Instead, the normal increase in ROS in bone marrow in ovariectomized mice was reversed by EDC treatment (an estrogenic ligand incapable of mediating nuclear ER signaling; see Section IV.E). This suggests that oxidative stress is reduced by extranuclear ER α actions via ERKs (191).

In conclusion, age-related bone loss may partly result from oxidative stress, which may be exacerbated by combined androgen and estrogen steroid deficiency in lateonset male hypogonadism. This hypothesis should be examined further in human interventional studies.

E. Immune system

Since the 1990s, it has been known from seminal studies by Pacifici (365) and other groups that estrogen deficiency increases systemic and bone marrow levels of proinflammatory cytokines like IL-1, TNF- α , interferon- γ , IL-6, and IL-17, which may all contribute to osteoporosis. T-lymphocytes are considered critical in the field of osteoimmunology because T-cell-deficient mice have been shown to be resistant to bone loss induced by ovariectomy in several (but not all) studies (365). Androgen deficiency also produces a proinflammatory response (366), but it is un-

known whether immunodeficient mice resist orchidectomy-induced bone loss. Recently, Onal et al (367) showed that RANKL expression in B-lymphocytes but not T-lymphocytes contributes to ovariectomy-induced cancellous bone loss.

The key question is whether $ER\alpha$, $ER\beta$, and AR in T-lymphocytes and/or B-lymphocytes mediate some of the skeletal effects of sex steroid deficiency. Conditional AR inactivation has recently been shown to expand B-cell repertoires (368), and $ER\alpha$ actions in T-lymphocytes account for the protective effects of estrogens in experimental autoimmune encephalitis (369). Future studies examining sex steroid actions in osteoimmunology using similar conditional models are eagerly awaited.

IX. Bone Loss and Fractures in Men

In the previous sections, we have reviewed evidence, mostly from genetic mouse models and rare human mutations, on how androgens and estrogens, via the AR and ERs, promote male skeletal development. Although further validation of these insights in human studies is needed, the conclusions that emerge from the bulk of available evidence are that the effects of T on male cortical and

Figure 10.

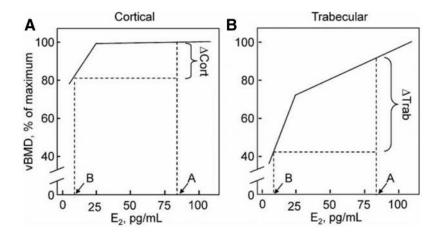


Figure 10. The possible threshold for skeletal E2 deficiency is more evident at cortical than trabecular sites in men. Schematic illustration is based on the data of Khosla et al (379) of the relationship between femoral neck cortical vBMD (A) and vertebral trabecular vBMD (B) and serum E2 levels. Note that whereas cortical vBMD is correlated with E2 levels at low E2 levels (below 25 pg/mL, corresponding to bioavailable E2 of 30 pmol/L), no relationship is evident at high E2 levels, consistent with a threshold below which cortical bone becomes estrogendeficient. By contrast, trabecular vBMD remains associated with E2 levels at both low and high levels, suggesting either the absence of a threshold or a threshold considerably higher than that present for cortical bone. Also shown are the predicted relative changes in cortical (Δ cort) and trabecular bone (Δ trab) as E2 levels fall from estrogen sufficiency (point A) to estrogen deficiency (point B). [Reproduced from S. Khosla et al: The unitary model for estrogen deficiency and the pathogenesis of osteoporosis: is a revision needed? *J Bone Miner Res.* 2011;26:441–451 (23), with permission. © American Society for Bone and Mineral Research.]

trabecular bone development rely on several mechanisms including: 1) direct actions via AR in osteoblasts and possibly other target cells; 2) ER α actions in osteoblast-lineage cells; 3) interaction with pubertal systemic GH/IGF-1 signaling; and 4) possibly via interaction with other signaling pathways and indirect effects (see Figures 4 and 8). These mechanisms can be linked to the greater cortical bone size and superior trabecular microstructure observed in young adult men compared to women. As mentioned in Section III, part of the advantage men have in terms of fracture risk later in life is probably determined by optimal pubertal peak bone mass acquisition.

In the last part of our review, we will now focus on age-related bone decay as well as fracture risk and the contribution of (acquired) sex steroid deficiency in late-onset male hypogonadism (see *Section II.A*).

A. Estrogen deficiency: the primary mediator of hypogonadal bone loss in older men

1. Observational studies

The importance of E2 for maintenance of bone mass in adult and elderly men has been established by numerous cross-sectional observational studies showing a strong association between serum levels of total and especially bio-

available E2, on the one hand, and BMD at various sites on the other (27, 370-374). Prospective studies also confirm that serum E2 levels associate with BMD loss in older men (373, 375-378). In a cohort of elderly men from Rochester, Minnesota, BMD at the radius and ulna declined by 0.49 to 0.66% per year, and these decreases were associated with both bioavailable E2 and bioavailable T levels, although the strongest correlations were found with bioavailable E2 (375). In this study, a threshold below which annual rates of bone loss started to decrease was found at a bioavailable E2 level of 40 pmol/L (11 pg/mL; corresponding to a total E2 level of 114 pmol/L [31 pg/mL]) (Figure 10). Other investigators independently reported similar threshold levels below which bone loss in older men accelerated at the lumbar spine or femoral neck (376, 378). Using QCT, further studies in men found that serum bioavailable E2 levels were consistently associated with vBMD at trabecular and cortical sites (379, 380). Interestingly, there appear to be differences in the dose relationships between serum E2 levels and cortical vs trabecular bone. In an elderly population of men, significant associations were observed between E2 levels and cortical vBMD when bioavailable E2 was below the median of 30 pmol/L (corresponding to a total E2 level of about 25 pg/mL), but not for bioavailable E2 levels above this value (379) (Figure 10A). In contrast, the correlations between trabecular vBMD and bioavailable E2 were rather similar for bioavailable E2 levels below or above 30 pmol/L (Figure 10B). Thus, the proposed threshold effect mentioned above appears to be more pronounced at cortical sites than trabecular sites in elderly men (379), although this was not confirmed in middleaged men (380).

Only one prospective study investigated the associations between sex steroids and vBMD loss at trabecular and cortical sites (121). In older men (>50 y), cortical bone loss correlated with bioavailable levels of both T and E2 at the distal radius, but not at the distal tibia. Serum bioavailable E2 levels associated with trabecular vBMD loss at the lumbar spine. Cross-sectional HRpQCT analyses in older men show moderate associations between serum bioavailable E2 and trabecular bone volume, trabecular number, and trabecular spacing at the distal radius (381), whereas serum total E2 levels associate inversely with cortical porosity at the distal tibia in the MrOS Sweden (126). In the STRAMBO cohort, a threshold was found of 25 pmol/L (quite similar to the one proposed above), below which cortical thickness and cortical vBMD were decreased when combined with a low apparent free T concentration (382). Lastly, data from the MINOS study (383) and the BACH study (384) showed that circulating E2 levels associated positively with estimated bone strength at the femur neck and distal radius in men. Longitudinal HRpQCT studies are required, however, to carefully elucidate the role of sex steroids in the age-related changes in trabecular and cortical microstructure and related bone strength indices in men.

Studies in older men with prostate cancer receiving androgen deprivation therapy (by chronic administration of GnRH agonists) further demonstrate the importance of E2 for skeletal maintenance in older men. These patients have reduced BMD and increased fracture risk (385) because of significantly suppressed levels of serum T and E2. In contrast, in patients treated with antiandrogen monotherapy, serum concentrations of T and E2 are increased, BMD is maintained, and bone turnover is unaffected (386, 387), indicating the importance of estrogen actions. Also in male-to-female transsexuals, androgen suppression and pharmacological estrogen therapy seems to be sufficient to maintain BMD (75, 388, 389), although this requires confirmation in prospective pQCT studies.

Several genetic studies also support the primary role of estrogens for bone health in men. Older men with more TTTA repeats in the aromatase (CYP19A1) gene have higher serum E2, less rapid BMD loss, and reduced fracture risk (377, 390, 391). In a sex steroid-related candidate gene study, a single nucleotide polymorphism in CYP19A1 conferred elevated E2 levels, higher lumber spine BMD, and reduced fracture incidence in older men (391). Bone quantitative ultrasound parameters in middle-aged and elderly men were also associated with several polymorphisms in CYP19A1 (392). The fact that peripheral aromatase activity may modulate circulating E2 levels is particularly interesting because estrogens in men are mostly derived from peripheral conversion of androgens (36, 37). A polymorphism in the gene for COMT (an estrogen-degrading enzyme) is also associated with prevalent fractures in elderly men (56, 57). Moreover, men with higher CAG repeat length in the AR gene have superior calcaneal ultrasound parameters because of estrogen activity after aromatization of their increased T levels (393). Finally, GWAS meta-analyses have identified the ESR1 locus (encoding $ER\alpha$) to be strongly associated with lumbar spine and femoral neck BMD (394, 395), as well as trabecular and cortical vBMD in middle-aged and older European men (396). Finally, Paternoster et al (162) recently provided the first genetic evidence using GWAS that a single nucleotide polymorphism in ESR1 was associated with cortical but not trabecular vBMD in men.

Taken together, these observational studies suggest that estrogens play a vital role in maintaining bone mass in adult and elderly men (Table 4).

2. Intervention studies

Although highly indicative, the above-mentioned association studies do not provide evidence for a causal role of E2 for skeletal health in aging men. The relative contributions of androgens and estrogens to male bone metabolism have been elegantly demonstrated in direct intervention studies. Administration of an aromatase inhibitor in older men with low T levels increases T but decreases E2 levels and reduces BMD, suggesting that the increase in T cannot overcome the detrimental effect of selective E2 deficiency on the skeleton (274). Conversely, SERMs have proven efficient in treating bone loss associated with androgen deprivation therapy in prostate cancer patients (397). Raloxifene treatment modestly increases BMD in men on androgen deprivation therapy (398), whereas toremifene not only increased BMD and reduced bone turnover markers, but also reduced the incidence of new vertebral fractures after 2 years in a randomized, placebo-

Table 4. Different Lines of Evidence Regarding the Role of Estrogen or Androgen Actions in Male Hypogonadal Bone Loss

	E2/ER $lpha$	T/AR	Refs.		
Population-based studies	Independent association with BMD, BMD losses, (micro)structure and fracture risk in multiple independent studies	Same associations less strong, not independent or weakened after correcting for E2 levels; or limited to cortical bone sites and BMD-independent fracture prevention	121, 126, 370–384, 417, 418 ^a 271, 373, 380, 382, 413, 416, 419, 420, 422, 427 ^b		
Genetic evidence	Polymorphisms in <i>ESR1</i> , <i>CYP19A1</i> , or <i>COMT</i> associated with E2, BMD, and/ or fracture risk	Long CAG repeats inhibit AR but increase T and E2; not negatively associated with BMD	56, 57, 162, 377, 390- 392, 394–396 ^a		
			393, 425 ^b		
Clinical studies	Androgen (and estrogen) deprivation therapy for prostate cancer increases	Antiandrogen monotherapy does not increase bone loss or fracture risk	385, 387, 398, 399 ^a		
	bone loss and fracture risk; SERMs prevent bone loss and fractures in this setting	SARMs do not reduce BMD loss or bone turnover in human trials	359, 446 ^b		
	Pharmacological E2, even with androgen suppression, prevents bone loss in male-to-female transsexuals	In female-to-male transsexuals, effects of T on BMD are equivocal; cortical bone size may increase and cortical vBMD decrease, but may depend on aromatization	75, 388ª 167, 389, 423, 424 ^b		
Intervention studies	Aromatase inhibition increases T but decreases E2 and BMD	DHT therapy decreases BMD, probably due to E2 suppression 5α-reductase inhibitors reduce muscle-	274, 275 ^a 438 ^b 336, 337, 447–450 ^b		
		anabolic effects of T but do not affect bone	., ,		
	GnRH suppression with T, E2, or T + aromatase inhibition shows dominant role for E2 in short-term BTM and HRpQCT changes	Androgens selectively linked to bone formation markers	276, 404, 405 ^{a,b}		

Abbreviation: BTM, bone turnover marker.

39controlled trial (399). Treatment with the sex steroid precursor DHEA in older men with low levels of DHEA sulfate modestly increased BMD at the femoral neck in some (400, 401) but not all (402, 403) studies. Jankowski et al (400) showed that the changes in regional hip BMD during DHEA treatment were associated with the serum levels of E2 at the end of the intervention. Short-term pharmacological suppression of endogenous E2 and T production in elderly men (through treatment with a GnRH agonist in combination with an aromatase inhibitor), followed by complete or selective add-back of E2 and T, shows that E2 significantly restores bone resorption and formation markers, although T also independently maintains the bone formation marker osteocalcin (276). Leder et al (404) randomized adult men to chemical castration and T replacement, with or without an aromatase inhibitor, to show that especially estrogens but possibly also androgens independently regulate bone resorption, whereas bone formation markers showed more complex patterns and only bordered significance. Most recently, a 16-week intervention study in 20- to 50-year-old men with gonadotropin inhibition, aromatase inhibition, and different doses of T treatment was performed by the same investigators in order to investigate the effects of T and E2 on vBMD and bone microstructure as assessed by HRpQCT (208, 405). Changes in vBMD and bone microstructural parameters were similar across the different T dose groups, indicating a lack of effect of physiological androgen levels in the near-absence of E2. In contrast, this suppression of E2 levels reduced trabecular and cortical vBMD at the tibia and radius and altered bone microstructure in a pattern consistent with trabecularization of the cortex.

These results strongly indicate that estrogen deficiency is the primary mediator of bone loss in hypogonadal men.

3. Observational studies with fracture endpoints

Several large-scale, prospective cohort studies using fractures as the hard endpoint further support a dominant role for E2 in male bone health (406). However, it should be mentioned that the early fracture studies, both cross-sectional and prospective, showed conflicting results regarding the relationship between serum E2 and fracture risk (407–414) (Table 5). This was most probably due to

^a These references pertain to estrogens/ER.

^b These references pertain to androgens/AR.

Table 5. Overview of Longitudinal Studies Investigating the Relationship Between Serum Levels of Sex Steroids and SHBG and Fractures

First Author, Year (Ref.), Study	No. of Subjects/ Age, y	Follow-Up Time, y	No. of Subjects With Fracture/ Fracture Type	Sex Steroid Assay	E2	т	SHBG
Nyquist, 1998 (453)	242/mean, 67	7	22/all	RIA	-	No	No
Barrett-Connor, 2000 (407), Rancho Bernardo study	352/median, 66	8	28/vertebral	RIA	Yes	No	_
Goderie-Plomp, 2004 (410), Rotterdam study	178/mean, 66	6.5	45/vertebral	RIA	No	No	No
Amin, 2006 (416), Framingham study	793/mean, 71	18	39/hip	RIA	Yes	No	_
Bjørnerem, 2007 (408), Tromsø study	1364/mean, 63	8.4	105/nonvertebral	IA	No	No	Yes
Meier, 2008 (427), Dubbo study	609/mean, 73	5.8	113/low-trauma	MS	No	Yes	Yes
Mellström, 2008 (417), MrOS Sweden	2639/mean, 75	3.3	209/all	MS	Yes	No	Yes
Roddam, 2009 (414), EPIC-Oxford study	464/mean, 51	5	155/all	RIA	Yes	No	No
LeBlanc, 2009 (418), MrOS USA	1978/mean, 73	4.7	342/nonvertebral	MS	Yes	Noa	Yes
Woo, 2012 (373), MrOS Hong Kong	1489/mean, 73	4	108/all	MS	Yes	No	No

Abbreviations: –, not reported; IA, immunoassay; MS, mass spectrometry.

the limited number of fractures in these studies, as well as reliance on immunoassays, which have inadequate specificity at lower sex steroid concentrations (46, 48, 415) (see *Section II.A*).

Amin et al (416) assessed the risk for hip fracture in aging men from the Framingham Study who were stratified in tertiles according to serum E2 levels. During 18 years of follow-up, 39 men sustained a hip fracture, and men in the low E2 group had three times greater risk of hip fracture compared to men in the highest E2 group. This was consistent with a threshold E2 level of around 66 pmol/L (18 pg/mL), below which fracture risk increased exponentially. The most conclusive data addressing this

issue have been reported by investigators from the MrOS study. In the MrOS Sweden cohort, Mellström et al (417) analyzed baseline serum sex steroid levels by mass spectrometry in 2639 elderly men and evaluated fractures over a relatively short follow-up of 3.3 years. Serum free E2 levels were independently associated with fracture risk. Fracture incidence correlated negatively with serum E2 levels, but only below a level of 59 pmol/L (16 pg/mL), whereas no relationship between fracture incidence and E2 levels was seen above this level (Figure 11) (417). These findings were subsequently extended in men \geq 65 years old in MrOS USA (n = 1978) by LeBlanc et al (418), who found that men in the lowest quartile of bioavailable E2

(<12 pg/mL) had the greatest risk of nonvertebral fractures. A third very similar study in the MrOS Hong Kong cohort observed comparable findings (373). Also in this study, nonlinear relationships were described between total E2 and bioavailable E2 and fracture risk, with thresholds at 16 and 12 pg/mL, respectively, similar to the ones described previously for fractures (416–418).

In all studies, the association between serum free/bioavailable E2 levels and fracture risk was attenuated after adjustment for BMD, suggesting that this is partly explained by effects on bone density and partly by additional effects. The threshold E2 level observed in the different fracture studies is also somewhat lower than the threshold described previously for BMD and BMD loss.

Figure 11.

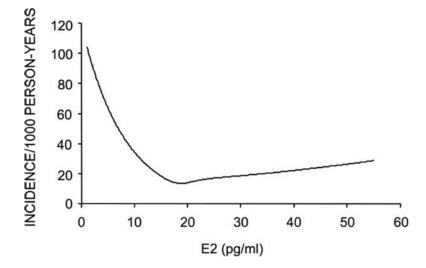


Figure 11. Annual incidence of fractures in relation to serum E2 levels in older men. Poisson regression models were used to determine the relationship between serum E2 and fracture risk. [Reproduced from D. Mellström et al: Older men with low serum estradiol and high serum SHBG have an increased risk of fractures. *J Bone Miner Res.* 2008;23:1552–1560 (417), with permission. © American Society for Bone and Mineral Research.]

^a Men with low bioavailable T and high SHBG have increased fracture risk.

This may indicate that the mechanism through which E2 affects fracture risk differs to some extent from that affecting bone maintenance in aging men. Alternatively, the different E2 assays used may also explain the difference in threshold values because in the association studies serum E2 was most often measured by immunoassay, whereas mass spectrometry was used in recent fracture studies.

In summary, these results provide compelling evidence for the essential effects of estrogens on skeletal health in aging men. Accumulating evidence shows a threshold below which E2 is associated with lower BMD, accelerated bone loss, and elevated fracture risk in older men.

B. High SHBG and low T: the second and third fiddle in male osteoporosis?

1. Independent actions of androgens in aging men

Although the above-mentioned data indicate that E2 is pivotal for maintaining bone health in aging men, this does not preclude independent contributions of androgens. Yet, observational studies have reported conflicting results on the relation between serum T levels and bone turnover markers, BMD, or BMD loss, with the associations being absent, less strong compared to E2, limited to cortical bone sites, or lost after adjusting for E2 or other confounders (271, 380, 382, 413, 419, 420). Most of these studies are limited by the small number of older men with frank hypogonadism, ie, ability only to show lack of association between BMD and T variations within the normal range for older men. A more strict definition of lateonset hypogonadism from the European Male Ageing Study (EMAS) study is based on the presence of at least three sexual symptoms, total T levels of less than 11 nmol/L, and free T level of less than 220 pmol/L (421). These criteria more accurately associated with ultrasound-estimated BMD than biochemically low T alone (422). Although T therapy in female-to-male transsexuals has no clear effect on BMD (167, 389, 423), it increased cortical bone size in a recent prospective QCT study (167). In another study, however, BMD loss was again seen when an aromatase inhibitor was added (424).

CAG repeats, which reduce AR sensitivity, have been shown to exert a positive effect on BMD in EMAS by increasing T and consequently E2 levels (393), whereas previous smaller studies were inconsistent (425). Genetic evidence recently implicated T in male bone health by identifying a locus on the X chromosome that was genome-wide significantly associated with both serum T levels (40) and lumbar spine BMD specifically in men (25), although this could be due to aromatization.

The influence of T on bone turnover markers is complex, with T treatment inducing a decrease in bone resorp-

tion, which is followed by a decrease in bone formation (426). Still, T was suggested to be independently related to indices of bone formation and resorption in the intervention studies described above (276, 404, 405).

The contribution of serum T to fracture risk in men is more complex than that of E2. In several studies, serum levels of total or bioavailable T were not independently related to fracture incidence in older men (373, 416–418) (Table 5). Nonetheless, Meier et al (427) found that T was independently and more strongly associated with low-trauma fractures than serum E2 in 609 older men during a 5.8-year follow-up, although spine and hip BMD in that study related to E2 but not T levels. Also in the Framingham Study, men with low T faced additional hip fracture risk in the presence of similarly low E2 levels (416), and in MrOS Hong Kong, older men within the lowest quartiles of both E2 and T faced approximately twice the risk of incident nonvertebral fractures (373).

Despite the general lack of association with BMD, androgens may still play a role in cortical density (382) and cortical appositional growth because older men with high T levels have been reported to have larger cortical bone areas (378, 380, 413). Whether or not this occurs through effects on peak bone mass acquisition (see Sections III and IV) or during aging is unknown. Furthermore, we agree with the view of Khosla (406), who suggested that androgen status probably affects fracture risk in elderly men to a large extent via nonskeletal effects, such as on muscle mass and propensity to fall. Indeed, in the MrOS USA study, it was found that serum T levels affect falling; men with T levels in the lowest quartile had a 40% higher fall risk than those in the highest quartile (428). Falls are independent predictors of fracture risk in older men (429). Serum T levels have been associated with lean mass, muscle strength, and frailty in cross-sectional (32) and prospective studies (34, 430). Also, intervention studies have shown that T increased lean body mass and muscle strength in men (33, 208). Moreover, (free) T levels have been associated with cognition in men (431-433), although randomized trials have not confirmed this. Clinical trials with T or selective AR modulators (SARMs) looking at BMD, sarcopenia, frailty, fractures, and falls as endpoints are needed to provide direct evidence for the clinical efficacy (and safety) of T for prevention and treatment of age-related musculoskeletal involution, falls, and fractures.

2. Randomized trials

The use of androgen replacement in elderly men to preserve musculoskeletal integrity, physical functioning, and quality of life remains controversial and poorly supported by evidence (406). Nevertheless, an increasing number of

older men are prescribed T for age-related symptoms or age-related decline in T levels (33). However, T replacement in aging men is only warranted in the case of lateonset hypogonadism, with consistent symptoms and signs and unequivocally low T levels (434). The effects of T therapy on bone health and physical function outcomes have been reviewed recently (33). Its clinical efficacy on BMD in older men with low or borderline serum T levels varies depending on the route of administration, with im but not transdermal T increasing lumbar spine BMD (435). Also, short follow-up and relatively normal baseline serum T levels of the participants may explain the lack of effect in some studies. Lumbar spine BMD increased moderately in most studies after at least 1 year of T therapy, whereas BMD increased at the hip in some studies but not others (355–357, 436, 437).

Whether T exerts its effects directly via the AR or indirectly on the ERs after aromatization cannot be determined from these studies. However, a 2-year study in older men showed that transdermal DHT (which cannot be aromatized) reduced spinal but not hip BMD, concomitant with decreases in serum E2 levels (438). These findings are at odds with studies in adult rodents and ARKO, ER α KO, and/or ER β KO mice treated with selective ligands, which showed that both ER α and AR can mediate trabecular bone maintenance during aging, whereas both together are required for cortical bone maintenance (73, 131, 135, 137). It is important to realize that all of the human intervention studies described above (274, 276, 404, 405, 438) have examined the role of androgens on the background of very low instead of normal E2 levels. Moreover, effects on periosteal apposition may occur too slowly to be detected in short-term experimental studies.

There is growing concern regarding the effects of T therapy on cardiovascular outcomes and mortality. A generally modest association has been observed between low endogenous T and incident cardiovascular endpoints in men (439). A randomized clinical trial of T therapy in men with impaired mobility and a high prevalence of cardiovascular diseases was, however, stopped prematurely due to adverse cardiovascular events (440), raising concerns about the safety of T therapy. In multivariable analysis, these cardiovascular events were related to the change in free T levels, and not E2 (441). The ongoing Testosterone Trial in Older Men (ClinicalTrials.gov identifier: NCT00799617), a large, randomized trial of T in older men with unequivocally low T levels and symptoms – sexual dysfunction, physical dysfunction, or low vitalitywill determine whether T improves these symptoms and also address safety issues.

Preclinical animal studies have demonstrated increased bone and muscle mass and decreased fat mass without

adverse effects on reproductive organs with SARMs (358, 442-445). Yet the few available human trials with SARMs haven't shown any beneficial effect on BMD or bone turnover markers, despite improvements in lean mass and physical performance (359, 446). Furthermore, the lack of aromatizability of most (nonsteroidal) SARMs remains a concern, given the possibility of (further) suppressing endogenous T and the importance of aromatization for the male skeleton.

3. Role of 5α -reductase

Part of the effects of T on the skeleton may be mediated by the more potent androgen DHT after conversion by 5α -reductase enzymes. 5α -Reductase type 2 is expressed predominantly in male reproductive tissues, whereas type 1 is moderately expressed in bone and also in other tissues (52). Finasteride, the most commonly used 5α -reductase inhibitor, is specific for type 2 and does not affect bone turnover, BMD, fracture risk (447–449), or the effects of androgens on muscle mass and the skeleton (337, 450). The skeletal effects of dutasteride (a dual type 1 and 2 inhibitor) are less well studied, but available evidence suggests that it also does not reduce BMD, increase bone turnover markers (448), or impair the muscle anabolic effects of T (336), suggesting that the conversion of T to DHT is not necessary for the effects of T on the musculoskeletal system in humans. 5α -Reductase type 1-inactivated male mice have slightly reduced bone mass (52) (see Section II.B), but the relevance of this finding to human bone physiology requires further investigation.

4. Role of SHBG

SHBG levels have also been associated with bone turnover markers, BMD, and BMD loss in older men (372, 378, 411, 412, 451). Polymorphisms in the promoter of the SHBG gene associated with serum levels of SHBG and hip BMD in elderly men (452). Several observational studies showed increased prevalent fractures with increased levels of SHBG (409, 411, 412). High SHBG levels were also independently associated with incident fracture risk in older men in most (408, 417, 418, 427), but not all studies (373, 410, 453) (Table 5). Interestingly, the MrOS USA study found evidence of an interaction between bioavailable T and SHBG: older men with low bioavailable T and high SHBG were at substantially increased risk of fracture, and the highest fracture risk occurred in men with low bioavailable E2, low bioavailable T, and high SHBG (418). Similar results were found when analyzing bone loss in the same cohort: men who had the lowest bioavailable E2, the lowest bioavailable T, and the highest SHBG experienced the fastest annualized rate of bone loss (378). A significant threshold was also found for SHBG at

Figure 12.

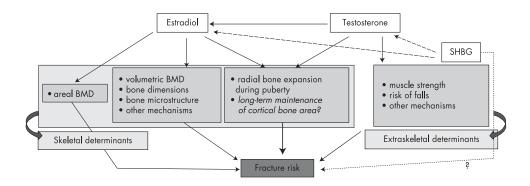


Figure 12. Proposed mechanism for the effects of E2, T, and SHBG on fracture risk in older men.

around 50 nmol/L; men with SHBG levels above this level experienced significantly faster rates of bone loss. This threshold is consistent with findings from the MrOS Sweden cohort in which men with SHBG levels in the highest quartile had an increased fracture risk (417). In the latter cohort, the greatest fracture risk was seen in men with low serum E2 and high SHBG levels. Serum SHBG levels are also related to nonskeletal fracture determinants like frailty (433, 454).

SHBG may affect fracture risk indirectly; by binding T and E2, SHBG reduces the free and bioavailable sex steroid fraction. However, there is discussion about whether SHBG may also enhance sex steroid sufficiency or deficiency, regulate the androgen/estrogen balance, or regulate mostly total sex steroid levels (because hypothalamic feedback mechanisms may correct free/bioavailable concentrations in vivo) (455, 456). Alternatively, it has been speculated that high levels of SHBG may directly potentiate sex steroid actions via putative membrane uptake of SHBG-sex steroid complexes or sequestration of sex steroids interstitially or intracellularly (41). Nonetheless, the results summarized here should be interpreted with caution because of strong correlations between each of the sex steroids and SHBG, particularly for the calculated free and bioavailable levels because these are derived from equations that already include SHBG (see also Section II.A). Furthermore, putative nonclassical actions of SHBG remain highly controversial, and it is not known whether these mechanisms bear any relevance to bone tissue. Finally, the observed associations could be due to residual confounding, eg, by IGF-1 levels, which may be associated with SHBG levels and predict fracture risk in older men (272).

In summary, strong evidence suggests that E2 plays a pivotal role in bone maintenance and fracture risk in aging men. These findings also suggest that each hormone—E2, T, and SHBG—or combinations thereof play a role in

male bone health. The proposed mechanisms for the effects of serum E2, T, and SHBG on bone loss and fracture risk in men are depicted in Figure 12. Circulating E2 is consistently associated with BMD and bone loss and inversely predicts fracture risk. Because the association between serum E2 and fractures is only partially attenuated after adjustment for BMD, other mechanisms besides a BMD-dependent effect can also contribute to E2's effect on fracture risk. These can include effects on bone dimensions, vBMD, and bone microstructure, which cannot be captured by DXA. T has a moderate effect on fracture risk that appears independent of BMD but may be related to maintenance of bone formation markers, cortical bone area, and especially to extraskeletal factors such as muscle mass, strength, and propensity to fall. Nevertheless, because T also functions as the substrate for aromatization into E2, it can affect BMD and thus fracture risk indirectly via E2. Finally, SHBG may indirectly influence fracture risk by regulating the bioavailable concentrations of E2 and T or, alternatively, affect fracture risk directly.

Regardless of their importance in pathophysiology, recent guidelines on male osteoporosis (28) do not advise routine measurement of E2, T, or SHBG in the absence of symptomatic hypogonadism because: 1) mass spectrometry is required to accurately determine low E2 levels, but this is not routinely available in most clinical settings; 2) there are no validated algorithms available to incorporate the sex steroid or SHBG in clinical decision making, eg, demonstrating that their measurement improves fracture prediction beyond DXA results and clinical risk factors (457); and 3) there are no randomized trials of T showing fracture prevention, whereas traditional antiresorptive drugs with established antifracture efficacy in women have demonstrated similar efficacy in men, regardless of normal or low T levels (19, 458). However, T replacement is an option in older men with morning serum T repeatedly < 6.9 nmol/L (200 ng/dL) who are not candidates for other osteoporosis treatments (28), in order to prevent the deleterious effects on bone that are mainly related to the concomitant estrogen deficiency.

X. Summary and General Conclusions

Although male osteoporosis constitutes a major burden for public health and can be effectively treated, underrecognition and undertreatment of this condition remains. Yet male gender is one of the strongest protective factors that approximately halves the risk of hip and other osteoporotic fractures (Figure 1). During the last decade, studies using (p)QCT and more recently HR-pQCT have refined our insights into the structural basis of sex differences in bone strength across the human lifespan (Figure 3).

The effects of androgens and estrogens on male bone health can be divided into two phases: 1) peak bone mass acquisition; and 2) subsequent maintenance. The male advantage in bone strength is mainly established during the first phase by placing cortical bone further from its central axis due to greater periosteal bone formation. Young adult women achieve similar cortical thickness by limiting endosteal expansion, but this doesn't provide the same biomechanical advantages. Men also have greater peak trabecular bone volume due to thicker, more plate-like trabeculae. However, these gender differences are probably site-specific and require further confirmation in more prospective studies.

How then do androgen and estrogen bioactivity regulate the establishment of this skeletal sexual dimorphism in cortical and trabecular bone? Human case reports of rare genetic conditions such as inactivating ER α mutations, aromatase deficiency, or AIS suggest that the lack of ER α activation is devastating for male cortical and trabecular bone acquisition, whereas androgen signaling via AR merely offers enhancement in these compartments in males. Genetic mouse models have allowed more systematic mechanistic studies, which confirm the dual mode of action of T on cortical and trabecular bone via both the AR and ER α . At the periosteal surface specifically, the stimulation by T in these models occurs mainly via the AR but also requires ER α as well as systemic GH/IGF-1 actions, which are regulated by ER α after aromatization (Figure 4).

Which target cells are then responsible for the skeletal effects of AR and ER α ? Throughout the osteoblast lineage, AR decreases trabecular bone resorption in male mice, whereas there is moderate evidence that ER α in mature osteoblasts and osteocytes regulates trabecular bone in male mice. ER α in early osteoblasts is crucial for the cortical bone in female mice, whereas it exerts a transient

effect on cortical bone mass in male mice (Table 2 and Figure 8). Direct effects of ER α in osteoclasts have been shown in female, but not male mice. In addition, ER β seems to exert some repressive actions in the female murine skeleton, but a role in male mice or in humans remains to be established. Longitudinal growth has been known to be regulated by ER α in males, but only recently it was confirmed in mice that ER α in growth plate chondrocytes mediates growth plate closure, whereas ER α in non-bone/ non-cartilage cells (probably via hypothalamic-pituitary regulation of the GH/IGF-1 axis) mediates longitudinal growth during early sexual maturation (Figure 7). Importantly, the target cell(s) for the AR effects on periosteal bone formation remains elusive, suggesting that sex steroid actions on the male skeleton may also rely on other organs. This is most evident for the effects of androgens on muscle in humans. Additional examples from mice include effects of ER α and AR on oxidative stress and possibly other organ systems, which requires further confirmation and investigation in humans.

How do AR and ERs regulate bone cells at the molecular level? With a few exceptions like FasL and alkaline phosphatase regulation by $ER\alpha$ in osteoblasts, the direct transcriptional targets of sex steroid actions in bone cells remain poorly understood. Nongenomic signaling pathways and specific actions of different $ER\alpha$ domains have shown promise as potential skeletal modulators in vivo. AR and $ER\alpha$ also cross talk with other signaling pathways and transcription factors in bone cells, as has been shown for GH/IGF and RUNX2. These molecular mechanisms require confirmation in humans.

Apart from their favorable influence on peak bone mass, how do sex steroid actions influence the second phase of male skeletal maintenance? Age-related bone loss is generally similar between men and women, only men lack an accelerated phase of bone loss compared to perimenopausal women, probably because men better maintain their bioactive sex steroids and especially estrogen levels. In rodent models, both ER α - and AR-mediated effects are required for male cortical and trabecular bone maintenance. In humans, however, recent epidemiological and interventional studies show that (bioactive) E2 levels in aging men are pivotal determinants of bone loss, microarchitectural decay, and increased fracture incidence. Yet the highest fracture risk is observed in men with a combination of low E2, low T, and high SHBG. Contrary to the situation for E2, the associations of T and SHBG with fractures appear largely independent of BMD, thereby suggesting that extraskeletal parameters like sarcopenia, frailty, and falls determine such associations (Figure 12). Interventional studies show that androgens alone cannot overcome bone loss due to (severe) estrogen defi-

ciency in aging men. On the other hand, none of the existing studies have examined the possibility of an independent skeletal effect of androgen deficiency in the face of (relatively) normal estrogen levels. Androgens could still have an independent effect on bone formation markers and at the periosteal surface, but the latter may not become evident in short-term interventional studies.

Regardless of their importance in pathophysiology, recent guidelines in male osteoporosis rightly point out that the role of T replacement in men with late-onset hypogonadism remains controversial. Evidence supporting the clinical utility of serum sex steroid measurements for fracture prediction beyond clinical risk factors and BMD also remains lacking. Further studies on sex steroid signaling in the musculoskeletal system are a high research priority because they may help in the design of additional, preferentially gender-neutral, therapeutic strategies to reach the ultimate goal of not merely preventing bone loss but also reinforcing the musculoskeletal system as a whole to prevent osteoporotic fractures in both genders.

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References

- Khosla S, Amin S, Orwoll E. Osteoporosis in men. Endocr Rev. 2008;29:441–464.
- 2. Orwoll ES, Bilezikian JP, Vanderschueren D. Osteoporosis in Men: The Effects of Gender on Skeletal Health. 2nd ed. London, UK: Elsevier Science; 2010.
- 3. Nguyen ND, Ahlborg HG, Center JR, Eisman JA, Nguyen

TV. Residual lifetime risk of fractures in women and men. *J Bone Miner Res.* 2007;22:781–788.

- 4. Lippuner K, Johansson H, Kanis JA, Rizzoli R. Remaining lifetime and absolute 10-year probabilities of osteoporotic fracture in Swiss men and women. *Osteoporos Int.* 2009; 20:1131–1140.
- Ahmed LA, Schirmer H, Bjørnerem A, et al. The genderand age-specific 10-year and lifetime absolute fracture risk in Tromsø, Norway. Eur J Epidemiol. 2009;24:441–448.
- 6. Kanis JA, Odén A, McCloskey EV, et al. A systematic review of hip fracture incidence and probability of fracture worldwide. *Osteoporos Int.* 2012;23:2239–2256.
- 7. Johnell O, Kanis JA. An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. *Osteoporos Int.* 2006;17:1726–1733.
- 8. Burge R, Dawson-Hughes B, Solomon DH, Wong JB, King A, Tosteson A. Incidence and economic burden of osteoporosis-related fractures in the United States, 2005–2025. *J Bone Miner Res.* 2007;22:465–475.
- 9. Donaldson LJ, Reckless IP, Scholes S, Mindell JS, Shelton NJ. The epidemiology of fractures in England. *J Epidemiol Community Health*. 2008;62:174–180.
- 10. Bergström U, Björnstig U, Stenlund H, Jonsson H, Svensson O. Fracture mechanisms and fracture pattern in men and women aged 50 years and older: a study of a 12-year population-based injury register, Umeå, Sweden. *Osteoporos Int.* 2008;19:1267–1273.
- 11. Haentjens P, Magaziner J, Colón-Emeric CS, et al. Metaanalysis: excess mortality after hip fracture among older women and men. *Ann Intern Med*. 2010;152:380–390.
- 12. Kannegaard PN, van der Mark S, Eiken P, Abrahamsen B. Excess mortality in men compared with women following a hip fracture. National analysis of comedications, comorbidity and survival. *Age Ageing*. 2010;39:203–209.
- 13. Center JR, Bliuc D, Nguyen TV, Eisman JA. Risk of subsequent fracture after low-trauma fracture in men and women. *JAMA*. 2007;297:387–394.
- 14. Tarride JE, Guo N, Hopkins R, et al. The burden of illness of osteoporosis in Canadian men. *J Bone Miner Res.* 2012; 27:1830–1838.
- 15. Curtis JR, McClure LA, Delzell E, et al. Population-based fracture risk assessment and osteoporosis treatment disparities by race and gender. *J Gen Intern Med.* 2009;24: 956–962.
- 16. Nurmi-Lüthje I, Sund R, Juntunen M, Lüthje P. Post-hip fracture use of prescribed calcium plus vitamin D or vitamin D supplements and antiosteoporotic drugs is associated with lower mortality: a nationwide study in Finland. *J Bone Miner Res.* 2011;26:1845–1853.
- 17. Leslie WD, LaBine L, Klassen P, Dreilich D, Caetano PA. Closing the gap in postfracture care at the population level: a randomized controlled trial. *CMAJ*. 2012;184:290–296.
- 18. Wastesson JW, Ringbäck Weitoft G, Parker MG, Johnell K. Educational level and use of osteoporosis drugs in elderly men and women: a Swedish nationwide registerbased study. *Osteoporos Int.* 2013;24:433–442.
- 19. Boonen S, Reginster JY, Kaufman JM, et al. Fracture risk and zoledronic acid therapy in men with osteoporosis. *N Engl J Med*. 2012;367:1714–1723.
- 20. Riggs BL, Khosla S, Melton LJ 3rd. A unitary model for involutional osteoporosis: estrogen deficiency causes both

- type I and type II osteoporosis in postmenopausal women and contributes to bone loss in aging men. *J Bone Miner Res.* 1998;13:763–773.
- Riggs BL, Khosla S, Melton LJ 3rd. Sex steroids and the construction and conservation of the adult skeleton. *Endocr Rev.* 2002;23:279–302.
- 22. **Manolagas SC.** From estrogen-centric to aging and oxidative stress: a revised perspective of the pathogenesis of osteoporosis. *Endocr Rev.* 2010;31:266–300.
- 23. **Khosla S, Melton LJ 3rd, Riggs BL.** The unitary model for estrogen deficiency and the pathogenesis of osteoporosis: is a revision needed? *J Bone Miner Res.* 2011;26:441–451.
- 24. Khosla S, Bellido TM, Drezner MK, et al. Forum on aging and skeletal health: summary of the proceedings of an ASBMR workshop. *J Bone Miner Res.* 2011;26:2565–2578.
- 25. Estrada K, Styrkarsdottir U, Evangelou E, et al. Genomewide meta-analysis identifies 56 bone mineral density loci and reveals 14 loci associated with risk of fracture. *Nat Genet*. 2012;44:491–501.
- 26. Liu CT, Estrada K, Yerges-Armstrong LM, et al. Assessment of gene-by-sex interaction effect on bone mineral density. *J Bone Miner Res.* 2012;27:2051–2064.
- Vanderschueren D, Vandenput L, Boonen S, Lindberg MK, Bouillon R, Ohlsson C. Androgens and bone. *Endocr Rev.* 2004;25:389–425.
- Watts NB, Adler RA, Bilezikian JP, et al. Osteoporosis in men: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab*. 2012;97:1802–1822.
- 29. Laurent M, Gielen E, Claessens F, Boonen S, Vanderschueren D. Osteoporosis in older men: recent advances in pathophysiology and treatment. *Best Pract Res Clin Endocrinol Metab*. 2013;27:527–539.
- 30. Lindle RS, Metter EJ, Lynch NA, et al. Age and gender comparisons of muscle strength in 654 women and men aged 20–93 yr. *J Appl Physiol* (1985). 1997;83:1581–1587.
- 31. Visser M, Fuerst T, Lang T, Salamone L, Harris TB. Validity of fan-beam dual-energy x-ray absorptiometry for measuring fat-free mass and leg muscle mass. Health, Aging, and Body Composition Study–Dual-Energy X-ray Absorptiometry and Body Composition Working Group. *J Appl Physiol* (1985). 1999;87:1513–1520.
- 32. Gielen E, Verschueren S, O'Neill TW, et al. Musculoskeletal frailty: a geriatric syndrome at the core of fracture occurrence in older age. *Calcif Tissue Int.* 2012;91:161–177.
- 33. Giannoulis MG, Martin FC, Nair KS, Umpleby AM, Sonksen P. Hormone replacement therapy and physical function in healthy older men. Time to talk hormones? *Endocr Rev.* 2012;33:314–377.
- Renoud A, Ecochard R, Marchand F, Chapurlat R, Szulc P. Predictive parameters of accelerated muscle loss in men– MINOS study. Am J Med. 2014;127:554–561.
- 35. Fazeli PK, Horowitz MC, MacDougald OA, et al. Marrow fat and bone–new perspectives. *J Clin Endocrinol Metab*. 2013;98:935–945.
- 36. **Kaufman JM, Vermeulen A.** The decline of androgen levels in elderly men and its clinical and therapeutic implications. *Endocr Rev.* 2005;26:833–876.
- 37. Labrie F, Cusan L, Gomez JL, et al. Comparable

- amounts of sex steroids are made outside the gonads in men and women: strong lesson for hormone therapy of prostate and breast cancer. *J Steroid Biochem Mol Biol*. 2009;113:52–56.
- 38. de Ronde W, van der Schouw YT, Pols HA, et al. Calculation of bioavailable and free testosterone in men: a comparison of 5 published algorithms. *Clin Chem.* 2006;52: 1777–1784.
- 39. Ly LP, Sartorius G, Hull L, et al. Accuracy of calculated free testosterone formulae in men. *Clin Endocrinol (Oxf)*. 2010;73:382–388.
- 40. Ohlsson C, Wallaschofski H, Lunetta KL, et al. Genetic determinants of serum testosterone concentrations in men. *PLoS Genet*. 2011;7:e1002313.
- 41. Laurent MR, Vanderschueren D. Reproductive endocrinology: functional effects of sex hormone-binding globulin variants. *Nat Rev Endocrinol*. 2014;10:516–517.
- 42. Rosner W, Auchus RJ, Azziz R, Sluss PM, Raff H. Position statement: utility, limitations, and pitfalls in measuring testosterone: an Endocrine Society position statement. *J Clin Endocrinol Metab*. 2007;92:405–413.
- 43. Orwoll E, Lambert LC, Marshall LM, et al. Testosterone and estradiol among older men. *J Clin Endocrinol Metab*. 2006;91:1336–1344.
- 44. Bhasin S, Pencina M, Jasuja GK, et al. Reference ranges for testosterone in men generated using liquid chromatography tandem mass spectrometry in a community-based sample of healthy nonobese young men in the Framingham Heart Study and applied to three geographically distinct cohorts. *J Clin Endocrinol Metab*. 2011;96:2430–2439.
- 45. Naessen T, Sjogren U, Bergquist J, Larsson M, Lind L, Kushnir MM. Endogenous steroids measured by high-specificity liquid chromatography-tandem mass spectrometry and prevalent cardiovascular disease in 70-year-old men and women. *J Clin Endocrinol Metab*. 2010;95: 1889–1897.
- 46. Huhtaniemi IT, Tajar A, Lee DM, et al. Comparison of serum testosterone and estradiol measurements in 3174 European men using platform immunoassay and mass spectrometry; relevance for the diagnostics in aging men. *Eur J Endocrinol*. 2012;166:983–991.
- 47. Handelsman DJ, Wartofsky L. Requirement for mass spectrometry sex steroid assays in the Journal of Clinical Endocrinology and Metabolism. *J Clin Endocrinol Metab*. 2013;98:3971–3973.
- 48. Ohlsson C, Nilsson ME, Tivesten A, et al. Comparisons of immunoassay and mass spectrometry measurements of serum estradiol levels and their influence on clinical association studies in men. *J Clin Endocrinol Metab*. 2013;98: E1097–E1102.
- 49. Khosla S, Amin S, Singh RJ, Atkinson EJ, Melton LJ 3rd, Riggs BL. Comparison of sex steroid measurements in men by immunoassay versus mass spectroscopy and relationships with cortical and trabecular volumetric bone mineral density. *Osteoporos Int.* 2008;19:1465–1471.
- 50. Peng Z, Li X, Mäkelä S, Väänänen HK, Poutanen M. Skeletal changes in transgenic male mice expressing human cytochrome P450 aromatase. *J Bone Miner Res.* 2004;19: 1320–1328.
- Sjögren K, Lagerquist M, Moverare-Skrtic S, et al. Elevated aromatase expression in osteoblasts leads to increased

bone mass without systemic adverse effects. *J Bone Miner Res*. 2009;24:1263–1270.

- 52. Windahl SH, Andersson N, Börjesson AE, et al. Reduced bone mass and muscle strength in male 5α -reductase type 1 inactivated mice. *PLoS One*, 2011;6:e21402.
- 53. Vandenput L, Labrie F, Mellström D, et al. Serum levels of specific glucuronidated androgen metabolites predict BMD and prostate volume in elderly men. *J Bone Miner* Res. 2007;22:220–227.
- 54. Lorentzon M, Eriksson AL, Mellström D, Ohlsson C. The COMT val158met polymorphism is associated with peak BMD in men. *J Bone Miner Res.* 2004;19:2005–2011.
- 55. Lorentzon M, Eriksson AL, Nilsson S, Mellström D, Ohlsson C. Association between physical activity and BMD in young men is modulated by catechol-O-methyltransferase (COMT) genotype: the GOOD study. *J Bone Miner Res.* 2007;22:1165–1172.
- 56. Stolk L, van Meurs JB, Jhamai M, et al. The catechol-O-methyltransferase Met158 low-activity allele and association with nonvertebral fracture risk in elderly men. J Clin Endocrinol Metab. 2007;92:3206–3212.
- 57. Eriksson AL, Mellström D, Lorentzon M, et al. The COMT val158met polymorphism is associated with prevalent fractures in Swedish men. *Bone*. 2008;42:107–112.
- 58. Yarrow JF, Conover CF, Lipinska JA, Santillana CA, Wronski TJ, Borst SE. Methods to quantify sex steroid hormones in bone: applications to the study of androgen ablation and administration. *Am J Physiol Endocrinol Metab*. 2010;299:E841–E847.
- 59. Helsen C, Kerkhofs S, Clinckemalie L, et al. Structural basis for nuclear hormone receptor DNA binding. *Mol Cell Endocrinol*. 2012;348:411–417.
- Claessens F, Denayer S, Van Tilborgh N, Kerkhofs S, Helsen C, Haelens A. Diverse roles of androgen receptor (AR) domains in AR-mediated signaling. *Nucl Recept Signal*. 2008;6:e008.
- Sahu B, Pihlajamaa P, Dubois V, Kerkhofs S, Claessens F, Jänne OA. Androgen receptor uses relaxed response element stringency for selective chromatin binding and transcriptional regulation in vivo. *Nucleic Acids Res.* 2014;42: 4230–4240.
- 62. Schauwaers K, De Gendt K, Saunders PT, et al. Loss of androgen receptor binding to selective androgen response elements causes a reproductive phenotype in a knockin mouse model. *Proc Natl Acad Sci USA*. 2007;104:4961–4966.
- 63. Mödder UI, Sanyal A, Kearns AE, et al. Effects of loss of steroid receptor coactivator-1 on the skeletal response to estrogen in mice. *Endocrinology*. 2004;145:913–921.
- 64. Langer G, Bader B, Meoli L, et al. A critical review of fundamental controversies in the field of GPR30 research. *Steroids*. 2010;75:603–610.
- 65. Kim KH, Toomre D, Bender JR. Splice isoform estrogen receptors as integral transmembrane proteins. *Mol Biol Cell*. 2011;22:4415–4423.
- 66. Rudnik V, Sanyal A, Syed FA, et al. Loss of ERE binding activity by estrogen receptor-α alters basal and estrogenstimulated bone-related gene expression by osteoblastic cells. *J Cell Biochem*. 2008;103:896–907.
- 67. Ciana P, Raviscioni M, Mussi P, et al. In vivo imaging of

- transcriptionally active estrogen receptors. *Nat Med*. 2003;9:82-86.
- 68. Jänne M, Deol HK, Power SG, Yee SP, Hammond GL. Human sex hormone-binding globulin gene expression in transgenic mice. *Mol Endocrinol*. 1998;12:123–136.
- Toran-Allerand CD, Tinnikov AA, Singh RJ, Nethrapalli IS. 17α-estradiol: a brain-active estrogen? *Endocrinology*. 2005;146:3843–3850.
- 70. McNamara KM, Harwood DT, Simanainen U, Walters KA, Jimenez M, Handelsman DJ. Measurement of sex steroids in murine blood and reproductive tissues by liquid chromatography-tandem mass spectrometry. *J Steroid Biochem Mol Biol.* 2010;121:611–618.
- 71. Vandenput L, Swinnen JV, Boonen S, et al. Role of the androgen receptor in skeletal homeostasis: the androgen-resistant testicular feminized male mouse model. *J Bone Miner Res.* 2004;19:1462–1470.
- 72. Haisenleder DJ, Schoenfelder AH, Marcinko ES, Geddis LM, Marshall JC. Estimation of estradiol in mouse serum samples: evaluation of commercial estradiol immunoassays. *Endocrinology*. 2011;152:4443–4447.
- Vandenput L, Ederveen AG, Erben RG, et al. Testosterone prevents orchidectomy-induced bone loss in estrogen receptor-α knockout mice. *Biochem Biophys Res Commun*. 2001;285:70–76.
- 74. McDougall KE, Perry MJ, Gibson RL, Colley SM, Korach KS, Tobias JH. Estrogen receptor-α dependency of estrogen's stimulatory action on cancellous bone formation in male mice. *Endocrinology*. 2003;144:1994–1999.
- 75. Lapauw B, Taes Y, Simoens S, et al. Body composition, volumetric and areal bone parameters in male-to-female transsexual persons. *Bone*. 2008;43:1016–1021.
- 76. Perrien DS, Akel NS, Edwards PK, et al. Inhibin A is an endocrine stimulator of bone mass and strength. *Endocrinology*. 2007;148:1654–1665.
- 77. Matsumoto C, Inada M, Toda K, Miyaura C. Estrogen and androgen play distinct roles in bone turnover in male mice before and after reaching sexual maturity. *Bone*. 2006;38: 220–226.
- 78. Callewaert F, Venken K, Kopchick JJ, et al. Sexual dimorphism in cortical bone size and strength but not density is determined by independent and time-specific actions of sex steroids and IGF-1: evidence from pubertal mouse models. *J Bone Miner Res.* 2010;25:617–626.
- 79. Yu S, Zhang C, Lin CC, et al. Altered prostate epithelial development and IGF-1 signal in mice lacking the androgen receptor in stromal smooth muscle cells. *Prostate*. 2011; 71:517–524.
- Yeh S, Tsai MY, Xu Q, et al. Generation and characterization of androgen receptor knockout (ARKO) mice: an in vivo model for the study of androgen functions in selective tissues. *Proc Natl Acad Sci USA*. 2002;99:13498–13503.
- 81. Kawano H, Sato T, Yamada T, et al. Suppressive function of androgen receptor in bone resorption. *Proc Natl Acad Sci USA*. 2003;100:9416–9421.
- 82. Venken K, De Gendt K, Boonen S, et al. Relative impact of androgen and estrogen receptor activation in the effects of androgens on trabecular and cortical bone in growing male mice: a study in the androgen receptor knockout mouse model. *J Bone Miner Res.* 2006;21:576–585.
- 83. MacLean HE, Chiu WS, Notini AJ, et al. Impaired skeletal

- muscle development and function in male, but not female, genomic androgen receptor knockout mice. *FASEB J.* 2008;22:2676–2689.
- 84. Callewaert F, Venken K, Ophoff J, et al. Differential regulation of bone and body composition in male mice with combined inactivation of androgen and estrogen receptor-α. *FASEB J.* 2009;23:232–240.
- 85. Oz OK, Zerwekh JE, Fisher C, et al. Bone has a sexually dimorphic response to aromatase deficiency. *J Bone Miner Res.* 2000;15:507–514.
- 86. Oz OK, Hirasawa G, Lawson J, et al. Bone phenotype of the aromatase deficient mouse. *J Steroid Biochem Mol Biol.* 2001;79:49–59.
- 87. Miyaura C, Toda K, Inada M, et al. Sex- and age-related response to aromatase deficiency in bone. *Biochem Biophys Res Commun*. 2001;280:1062–1068.
- 88. Hewitt SC, Kissling GE, Fieselman KE, Jayes FL, Gerrish KE, Korach KS. Biological and biochemical consequences of global deletion of exon 3 from the ER α gene. *FASEB J*. 2010;24:4660–4667.
- Vidal O, Lindberg MK, Hollberg K, et al. Estrogen receptor specificity in the regulation of skeletal growth and maturation in male mice. *Proc Natl Acad Sci USA*. 2000;97: 5474–5479.
- 90. Lindberg MK, Alatalo SL, Halleen JM, Mohan S, Gustafsson JA, Ohlsson C. Estrogen receptor specificity in the regulation of the skeleton in female mice. *J Endocrinol*. 2001; 171:229–236.
- 91. Windahl SH, Vidal O, Andersson G, Gustafsson JA, Ohlsson C. Increased cortical bone mineral content but unchanged trabecular bone mineral density in female $ER\beta(-/-)$ mice. *J Clin Invest*. 1999;104:895–901.
- 92. Windahl SH, Hollberg K, Vidal O, Gustafsson JA, Ohlsson C, Andersson G. Female estrogen receptor β-/- mice are partially protected against age-related trabecular bone loss. *J Bone Miner Res.* 2001;16:1388–1398.
- 93. Sims NA, Dupont S, Krust A, et al. Deletion of estrogen receptors reveals a regulatory role for estrogen receptors-β in bone remodeling in females but not in males. *Bone*. 2002;30:18–25.
- 94. Syed FA, Mödder UI, Fraser DG, et al. Skeletal effects of estrogen are mediated by opposing actions of classical and nonclassical estrogen receptor pathways. *J Bone Miner Res.* 2005;20:1992–2001.
- 95. Syed FA, Fraser DG, Monroe DG, Khosla S. Distinct effects of loss of classical estrogen receptor signaling versus complete deletion of estrogen receptor α on bone. *Bone*. 2011; 49:208–216.
- 96. Syed FA, Fraser DG, Spelsberg TC, et al. Effects of loss of classical estrogen response element signaling on bone in male mice. *Endocrinology*, 2007;148:1902–1910.
- 97. Kerkhofs S, Denayer S, Haelens A, Claessens F. Androgen receptor knockout and knock-in mouse models. *J Mol Endocrinol*. 2009;42:11–17.
- 98. Matsumoto T, Takeyama K, Sato T, Kato S. Androgen receptor functions from reverse genetic models. *J Steroid Biochem Mol Biol.* 2003;85:95–99.
- 99. De Gendt K, Swinnen JV, Saunders PT, et al. A Sertoli cell-selective knockout of the androgen receptor causes spermatogenic arrest in meiosis. *Proc Natl Acad Sci USA*. 2004;101:1327–1332.

- 100. Notini AJ, Davey RA, McManus JF, Bate KL, Zajac JD. Genomic actions of the androgen receptor are required for normal male sexual differentiation in a mouse model. *J Mol Endocrinol*. 2005;35:547–555.
- 101. Pang TP, Clarke MV, Ghasem-Zadeh A, Lee NK, Davey RA, MacLean HE. A physiological role for androgen actions in the absence of androgen receptor DNA binding activity. *Mol Cell Endocrinol*. 2012;348:189–197.
- 102. MacLean HE, Chiu WS, Ma C, et al. A floxed allele of the androgen receptor gene causes hyperandrogenization in male mice. *Physiol Genomics*. 2008;33:133–137.
- 103. Dupont S, Krust A, Gansmuller A, Dierich A, Chambon P, Mark M. Effect of single and compound knockouts of estrogen receptors α (ER α) and β (ER β) on mouse reproductive phenotypes. *Development*. 2000;127:4277–4291.
- 104. Antal MC, Krust A, Chambon P, Mark M. Sterility and absence of histopathological defects in nonreproductive organs of a mouse ERβ-null mutant. *Proc Natl Acad Sci USA*. 2008;105:2433–2438.
- 105. Smith EP, Boyd J, Frank GR, et al. Estrogen resistance caused by a mutation in the estrogen-receptor gene in a man. *N Engl J Med.* 1994;331:1056–1061.
- 106. Ophoff J, Callewaert F, Venken K, et al. Physical activity in the androgen receptor knockout mouse: evidence for reversal of androgen deficiency on cancellous bone. *Biochem Biophys Res Commun*. 2009;378:139–144.
- 107. Rana K, Fam BC, Clarke MV, Pang TP, Zajac JD, MacLean HE. Increased adiposity in DNA binding-dependent androgen receptor knockout male mice associated with decreased voluntary activity and not insulin resistance. Am J Physiol Endocrinol Metab. 2011;301:E767– E778.
- 108. Raskin K, de Gendt K, Duittoz A, et al. Conditional inactivation of androgen receptor gene in the nervous system: effects on male behavioral and neuroendocrine responses. *J Neurosci.* 2009;29:4461–4470.
- 109. Riggs BL, Melton Iii LJ 3rd, Robb RA, et al. Population-based study of age and sex differences in bone volumetric density, size, geometry, and structure at different skeletal sites. *J Bone Miner Res.* 2004;19:1945–1954.
- 110. Lorentzon M, Swanson C, Andersson N, Mellström D, Ohlsson C. Free testosterone is a positive, whereas free estradiol is a negative, predictor of cortical bone size in young Swedish men: the GOOD study. *J Bone Miner Res.* 2005;20:1334–1341.
- 111. Vanbillemont G, Lapauw B, Bogaert V, et al. Sex hormone-binding globulin as an independent determinant of cortical bone status in men at the age of peak bone mass. *J Clin Endocrinol Metab.* 2010;95:1579–1586.
- 112. **Khosla S, Riggs BL, Atkinson EJ, et al.** Effects of sex and age on bone microstructure at the ultradistal radius: a population-based noninvasive in vivo assessment. *J Bone Miner Res.* 2006;21:124–131.
- 113. Burghardt AJ, Kazakia GJ, Ramachandran S, Link TM, Majumdar S. Age- and gender-related differences in the geometric properties and biomechanical significance of intracortical porosity in the distal radius and tibia. *J Bone Miner Res.* 2010;25:983–993.
- 114. Dalzell N, Kaptoge S, Morris N, et al. Bone micro-architecture and determinants of strength in the radius and tibia: age-related changes in a population-based study of normal

adults measured with high-resolution pQCT. Osteoporos Int. 2009;20:1683–1694.

- 115. Hansen S, Shanbhogue V, Folkestad L, Nielsen MM, Brixen K. Bone microarchitecture and estimated strength in 499 adult Danish women and men: a cross-sectional, population-based high-resolution peripheral quantitative computed tomographic study on peak bone structure. *Calcif Tissue Int.* 2014;94:269–281.
- 116. Kirmani S, Christen D, van Lenthe GH, et al. Bone structure at the distal radius during adolescent growth. *J Bone Miner Res.* 2009;24:1033–1042.
- 117. Macdonald HM, Nishiyama KK, Kang J, Hanley DA, Boyd SK. Age-related patterns of trabecular and cortical bone loss differ between sexes and skeletal sites: a population-based HR-pQCT study. *J Bone Miner Res.* 2011; 26:50–62.
- 118. Nishiyama KK, Macdonald HM, Moore SA, Fung T, Boyd SK, McKay HA. Cortical porosity is higher in boys compared with girls at the distal radius and distal tibia during pubertal growth: an HR-pQCT study. *J Bone Miner Res.* 2012;27:273–282.
- 119. Walsh JS, Paggiosi MA, Eastell R. Cortical consolidation of the radius and tibia in young men and women. *J Clin Endocrinol Metab*. 2012;97:3342–3348.
- 120. Ohlsson C, Darelid A, Nilsson M, Melin J, Mellström D, Lorentzon M. Cortical consolidation due to increased mineralization and endosteal contraction in young adult men: a five-year longitudinal study. *J Clin Endocrinol Metab*. 2011;96:2262–2269.
- 121. **Riggs BL, Melton LJ, Robb RA, et al.** A population-based assessment of rates of bone loss at multiple skeletal sites: evidence for substantial trabecular bone loss in young adult women and men. *J Bone Miner Res.* 2008;23:205–214.
- 122. Lauretani F, Bandinelli S, Griswold ME, et al. Longitudinal changes in BMD and bone geometry in a population-based study. *J Bone Miner Res.* 2008;23:400–408.
- 123. Marshall LM, Lang TF, Lambert LC, et al. Dimensions and volumetric BMD of the proximal femur and their relation to age among older U.S. men. *J Bone Miner Res.* 2006;21: 1197–1206.
- 124. Parfitt AM, Mathews CH, Villanueva AR, Kleerekoper M, Frame B, Rao DS. Relationships between surface, volume, and thickness of iliac trabecular bone in aging and in osteoporosis. Implications for the microanatomic and cellular mechanisms of bone loss. *J Clin Invest*. 1983;72:1396–1409.
- 125. **Zebaze RM**, **Ghasem-Zadeh A**, **Bohte A**, **et al.** Intracortical remodelling and porosity in the distal radius and post-mortem femurs of women: a cross-sectional study. *Lancet*. 2010;375:1729–1736.
- 126. Vandenput L, Lorentzon M, Sundh D, et al. Serum estradiol levels are inversely associated with cortical porosity in older men. J Clin Endocrinol Metab. 2014;99:E1322– E1326.
- 127. Haentjens P, Johnell O, Kanis JA, et al. Evidence from data searches and life-table analyses for gender-related differences in absolute risk of hip fracture after Colles' or spine fracture: Colles' fracture as an early and sensitive marker of skeletal fragility in white men. *J Bone Miner Res.* 2004; 19:1933–1944.
- 128. Hernandez CJ, Beaupré GS, Carter DR. A theoretical anal-

- ysis of the relative influences of peak BMD, age-related bone loss and menopause on the development of osteoporosis. *Osteoporos Int.* 2003;14:843–847.
- 129. Sims NA, Clément-Lacroix P, Minet D, et al. A functional androgen receptor is not sufficient to allow estradiol to protect bone after gonadectomy in estradiol receptor-deficient mice. *J Clin Invest*. 2003;111:1319–1327.
- 130. Määttä JA, Büki KG, Ivaska KK, et al. Inactivation of the androgen receptor in bone-forming cells leads to trabecular bone loss in adult female mice. *Bonekey Rep.* 2013;2: 440
- 131. Tivesten A, Movérare-Skrtic S, Chagin A, et al. Additive protective effects of estrogen and androgen treatment on trabecular bone in ovariectomized rats. *J Bone Miner Res.* 2004;19:1833–1839.
- 132. Smith EP, Specker B, Bachrach BE, et al. Impact on bone of an estrogen receptor-α gene loss of function mutation. *J Clin Endocrinol Metab.* 2008;93:3088–3096.
- 133. Vandenput L, Ohlsson C. Estrogens as regulators of bone health in men. *Nat Rev Endocrinol*. 2009;5:437–443.
- 134. Bouillon R, Bex M, Vanderschueren D, Boonen S. Estrogens are essential for male pubertal periosteal bone expansion. *J Clin Endocrinol Metab*. 2004;89:6025–6029.
- 135. Vandenput L, Boonen S, Van Herck E, Swinnen JV, Bouillon R, Vanderschueren D. Evidence from the aged orchidectomized male rat model that 17β-estradiol is a more effective bone-sparing and anabolic agent than 5α-dihydrotestosterone. *J Bone Miner Res.* 2002;17:2080–2086.
- 136. Börjesson AE, Farman HH, Engdahl C, et al. The role of activation functions 1 and 2 of estrogen receptor-α for the effects of estradiol and selective estrogen receptor modulators in male mice. *J Bone Miner Res.* 2013;28:1117–1126.
- 137. Movérare S, Venken K, Eriksson AL, et al. Differential effects on bone of estrogen receptor α and androgen receptor activation in orchidectomized adult male mice. *Proc Natl Acad Sci USA*. 2003;100:13573–13578.
- 138. Lindberg MK, Movérare S, Skrtic S, et al. Two different pathways for the maintenance of trabecular bone in adult male mice. *J Bone Miner Res.* 2002;17:555–562.
- 139. Lindberg MK, Weihua Z, Andersson N, et al. Estrogen receptor specificity for the effects of estrogen in ovariectomized mice. *J Endocrinol*. 2002;174:167–178.
- 140. Harris HA, Albert LM, Leathurby Y, et al. Evaluation of an estrogen receptor-β agonist in animal models of human disease. *Endocrinology* 2003;144:4241–4249.
- 141. Hillisch A, Peters O, Kosemund D, et al. Dissecting physiological roles of estrogen receptor α and β with potent selective ligands from structure-based design. *Mol Endocrinol*. 2004;18:1599–1609.
- 142. **Ke HZ, Brown TA, Qi H, et al.** The role of estrogen receptor-β, in the early age-related bone gain and later age-related bone loss in female mice. *J Musculoskelet Neuronal Interact.* 2002;2:479–488.
- 143. McDougall KE, Perry MJ, Gibson RL, et al. Estrogen-induced osteogenesis in intact female mice lacking ERβ. Am J Physiol Endocrinol Metab. 2002;283:E817–E823.
- 144. Bord S, Horner A, Beavan S, Compston J. Estrogen receptors α and β are differentially expressed in developing human bone. *J Clin Endocrinol Metab*. 2001;86:2309–2314.
- 145. Onoe Y, Miyaura C, Ohta H, Nozawa S, Suda T. Expres-

- sion of estrogen receptor β in rat bone. *Endocrinology*. 1997;138:4509–4512.
- 146. Seidlova-Wuttke D, Nguyen BT, Wuttke W. Long-term effects of ovariectomy on osteoporosis and obesity in estrogen-receptor-β-deleted mice. Comp Med. 2012;62: 8–13.
- 147. He YX, Liu Z, Pan XH, et al. Deletion of estrogen receptor β accelerates early stage of bone healing in a mouse osteotomy model. *Osteoporos Int.* 2012;23:377–389.
- 148. **Kamiya Y, Chen J, Xu M, et al.** Increased mandibular condylar growth in mice with estrogen receptor *β* deficiency. *I Bone Miner Res.* 2013;28:1127–1134.
- 149. Börjesson AE, Windahl SH, Lagerquist MK, et al. Roles of transactivating functions 1 and 2 of estrogen receptor-α in bone. *Proc Natl Acad Sci USA*. 2011;108:6288–6293.
- 150. Lindberg MK, Movérare S, Skrtic S, et al. Estrogen receptor (ER)- β reduces ER α -regulated gene transcription, supporting a "ying yang" relationship between ER α and ER β in mice. *Mol Endocrinol*. 2003;17:203–208.
- 151. Nicks KM, Fujita K, Fraser D, et al. Deletion of ERβ in early osteoprogenitor cells leads to a doubling of trabecular bone. In: Proceedings from the American Society for Bone and Mineral Research; October 4–7, 2013; Baltimore, MD. Oral Presentation 1090.
- 152. Ogawa S, Hosoi T, Shiraki M, et al. Association of estrogen receptor β gene polymorphism with bone mineral density. *Biochem Biophys Res Commun*. 2000;269:537–541.
- 153. Lau HH, Ho AY, Luk KD, Kung AW. Estrogen receptor β gene polymorphisms are associated with higher bone mineral density in premenopausal, but not postmenopausal southern Chinese women. *Bone*. 2002;31:276–281.
- 154. Shearman AM, Karasik D, Gruenthal KM, et al. Estrogen receptor β polymorphisms are associated with bone mass in women and men: the Framingham Study. *J Bone Miner Res.* 2004;19:773–781.
- 155. Scariano JK, Simplicio SG, Montoya GD, Garry PJ, Baumgartner RN. Estrogen receptor β dinucleotide (CA) repeat polymorphism is significantly associated with bone mineral density in postmenopausal women. *Calcif Tissue Int.* 2004;74:501–508.
- 156. Geng L, Yao Z, Yang H, Luo J, Han L, Lu Q. Association of CA repeat polymorphism in estrogen receptor β gene with postmenopausal osteoporosis in Chinese. *J Genet Genomics*. 2007;34:868–876.
- 157. Mendoza N, Quereda F, Presa J, et al. Estrogen-related genes and postmenopausal osteoporosis risk. *Climacteric*. 2012;15:587–593.
- 158. Currò M, Marini H, Alibrandi A, et al. The ESR2 AluI gene polymorphism is associated with bone mineral density in postmenopausal women. *J Steroid Biochem Mol Biol*. 2011;127:413–417.
- 159. Honma N, Mori S, Zhou H, et al. Association between estrogen receptor-β dinucleotide repeat polymorphism and incidence of femoral fracture. *J Bone Miner Metab*. 2013;31:96–101.
- 160. Zhang L, Choi HJ, Estrada K, et al. Multistage genome-wide association meta-analyses identified two new loci for bone mineral density. *Hum Mol Genet*. 2014;23:1923–1933.
- 161. Koller DL, Zheng HF, Karasik D, et al. Meta-analysis of genome-wide studies identifies WNT16 and ESR1 SNPs

- associated with bone mineral density in premenopausal women. *J Bone Miner Res.* 2013;28:547–558.
- 162. Paternoster L, Lorentzon M, Lehtimaki T, et al. Genetic determinants of trabecular and cortical volumetric bone mineral densities and bone microstructure. *PLoS Genet*. 2013;9:e1003247.
- 163. Movérare-Skrtic S, Börjesson AE, Farman HH, et al. The estrogen receptor antagonist ICI 182,780 can act both as an agonist and an inverse agonist when estrogen receptor α AF-2 is modified. *Proc Natl Acad Sci USA*. 2014;111: 1180–1185.
- 164. Börjesson AE, Windahl SH, Karimian E, et al. The role of estrogen receptor-α and its activation function-1 for growth plate closure in female mice. *Am J Physiol Endocrinol Metab*. 2012;302:E1381–E1389.
- 165. MacLean HE, Moore AJ, Sastra SA, et al. DNA-binding-dependent androgen receptor signaling contributes to gender differences and has physiological actions in males and females. *J Endocrinol*. 2010;206:93–103.
- 166. Rochira V, Zirilli L, Madeo B, et al. Skeletal effects of long-term estrogen and testosterone replacement treatment in a man with congenital aromatase deficiency: evidences of a priming effect of estrogen for sex steroids action on bone. *Bone*. 2007;40:1662–1668.
- 167. Van Caenegem E, Wierckx K, Taes Y, et al. Bone mass, bone geometry, and body composition in female-to-male transsexual persons after long-term cross-sex hormonal therapy. *J Clin Endocrinol Metab*. 2012;97:2503–2511.
- 168. Taes Y, Lapauw B, Vandewalle S, et al. Estrogen-specific action on bone geometry and volumetric bone density: longitudinal observations in an adult with complete androgen insensitivity. *Bone*. 2009;45:392–397.
- 169. Han TS, Goswami D, Trikudanathan S, Creighton SM, Conway GS. Comparison of bone mineral density and body proportions between women with complete androgen insensitivity syndrome and women with gonadal dysgenesis. *Eur J Endocrinol*. 2008;159:179–185.
- 170. Chiang C, Chiu M, Moore AJ, et al. Mineralization and bone resorption are regulated by the androgen receptor in male mice. *J Bone Miner Res.* 2009;24:621–631.
- 171. **Notini AJ, McManus JF, Moore A, et al.** Osteoblast deletion of exon 3 of the androgen receptor gene results in trabecular bone loss in adult male mice. *J Bone Miner Res.* 2007;22:347–356.
- 172. Sinnesael M, Claessens F, Laurent M, et al. Androgen receptor (AR) in osteocytes is important for the maintenance of male skeletal integrity: evidence from targeted AR disruption in mouse osteocytes. *J Bone Miner Res.* 2012;27: 2535–2543.
- 173. Konforte D, Shea JL, Kyriakopoulou L, et al. Complex biological pattern of fertility hormones in children and adolescents: a study of healthy children from the CALIPER cohort and establishment of pediatric reference intervals. *Clin Chem.* 2013;59:1215–1227.
- 174. Jansson JO, Ekberg S, Isaksson O, Mode A, Gustafsson JA. Imprinting of growth hormone secretion, body growth, and hepatic steroid metabolism by neonatal testosterone. *Endocrinology* 1985;117:1881–1889.
- 175. Sims NA, Brennan K, Spaliviero J, Handelsman DJ, Seibel MJ. Perinatal testosterone surge is required for normal

adult bone size but not for normal bone remodeling. *Am J Physiol Endocrinol Metab*. 2006;290:E456–E462.

- 176. Aguirre JI, Plotkin LI, Gortazar AR, et al. A novel ligand-independent function of the estrogen receptor is essential for osteocyte and osteoblast mechanotransduction. *J Biol Chem.* 2007;282:25501–25508.
- 177. Galea GL, Meakin LB, Sugiyama T, et al. Estrogen receptor α mediates proliferation of osteoblastic cells stimulated by estrogen and mechanical strain, but their acute down-regulation of the Wnt antagonist Sost is mediated by estrogen receptor β. *I Biol Chem.* 2013;288:9035–9048.
- 178. Windahl SH, Saxon L, Börjesson AE, et al. Estrogen receptor-α is required for the osteogenic response to mechanical loading in a ligand-independent manner involving its activation function 1 but not 2. *J Bone Miner Res.* 2013; 28:291–301.
- 179. Almeida M, Iyer S, Martin-Millan M, et al. Estrogen receptor-α signaling in osteoblast progenitors stimulates cortical bone accrual. *J Clin Invest*. 2013;123:394–404.
- 180. Mödder UI, Rudnik V, Liu G, Khosla S, Monroe DG. A DNA binding mutation in estrogen receptor-α leads to suppression of Wnt signaling via β-catenin destabilization in osteoblasts. I Cell Biochem. 2012;113:2248–2255.
- 181. Almeida M, Martin-Millan M, Ambrogini E, et al. Estrogens attenuate oxidative stress and the differentiation and apoptosis of osteoblasts by DNA-binding-independent actions of the ERα. *J Bone Miner Res.* 2010;25:769–781.
- 182. Chokalingam K, Roforth MM, Nicks KM, et al. Examination of ERα signaling pathways in bone of mutant mouse models reveals the importance of ERE-dependent signaling. *Endocrinology*. 2012;153:5325–5333.
- 183. Kousteni S, Bellido T, Plotkin LI, et al. Nongenotropic, sex-nonspecific signaling through the estrogen or androgen receptors: dissociation from transcriptional activity. *Cell.* 2001;104:719–730.
- 184. **Kousteni S, Chen JR, Bellido T, et al.** Reversal of bone loss in mice by nongenotropic signaling of sex steroids. *Science*. 2002;298:843–846.
- 185. **Kousteni S, Han L, Chen JR, et al.** Kinase-mediated regulation of common transcription factors accounts for the bone-protective effects of sex steroids. *J Clin Invest.* 2003; 111:1651–1664.
- 186. Kousteni S, Almeida M, Han L, Bellido T, Jilka RL, Manolagas SC. Induction of osteoblast differentiation by selective activation of kinase-mediated actions of the estrogen receptor. *Mol Cell Biol.* 2007;27:1516–1530.
- 187. Movérare S, Dahllund J, Andersson N, et al. Estren is a selective estrogen receptor modulator with transcriptional activity. Mol Pharmacol. 2003;64:1428–1433.
- 188. Windahl SH, Galien R, Chiusaroli R, et al. Bone protection by estrens occurs through non-tissue-selective activation of the androgen receptor. *J Clin Invest.* 2006;116:2500–2509
- 189. Manolagas SC, Jilka RL, Kousteni S, et al. Response to Windahl et al. *J Clin Invest*. 2006;116:2834.
- 190. Hewitt SC, Collins J, Grissom S, Hamilton K, Korach KS. Estren behaves as a weak estrogen rather than a non-genomic selective activator in the mouse uterus. *Endocrinology*. 2006;147:2203–2214.
- 191. Bartell SM, Han L, Kim HN, et al. Non-nuclear-initiated

- actions of the estrogen receptor protect cortical bone mass. *Mol Endocrinol*. 2013;27:649–656.
- 192. Martin-Millan M, Almeida M, Ambrogini E, et al. The estrogen receptor-α in osteoclasts mediates the protective effects of estrogens on cancellous but not cortical bone. *Mol Endocrinol*. 2010;24:323–334.
- 193. Heino TJ, Chagin AS, Sävendahl L. The novel estrogen receptor G-protein-coupled receptor 30 is expressed in human bone. *J Endocrinol*. 2008;197:R1–R6.
- 194. Windahl SH, Andersson N, Chagin AS, et al. The role of the G protein-coupled receptor GPR30 in the effects of estrogen in ovariectomized mice. *Am J Physiol Endocrinol Metab.* 2009;296:E490–E496.
- 195. Mårtensson UE, Salehi SA, Windahl S, et al. Deletion of the G protein-coupled receptor 30 impairs glucose tolerance, reduces bone growth, increases blood pressure, and eliminates estradiol-stimulated insulin release in female mice. *Endocrinology*. 2009;150:687–698.
- 196. Ford J, Hajibeigi A, Long M, et al. GPR30 deficiency causes increased bone mass, mineralization, and growth plate proliferative activity in male mice. *J Bone Miner Res.* 2011; 26:298–307.
- 197. Chagin AS, Sävendahl L. GPR30 estrogen receptor expression in the growth plate declines as puberty progresses. *J Clin Endocrinol Metab.* 2007;92:4873–4877.
- 198. Drake MT, McCready LK, Hoey KA, Atkinson EJ, Khosla S. Effects of suppression of follicle-stimulating hormone secretion on bone resorption markers in postmenopausal women. *J Clin Endocrinol Metab.* 2010;95:5063–5068.
- 199. Sun L, Peng Y, Sharrow AC, et al. FSH directly regulates bone mass. *Cell*. 2006;125:247–260.
- 200. **Iqbal J, Sun L, Kumar TR, Blair HC, Zaidi M.** Follicle-stimulating hormone stimulates TNF production from immune cells to enhance osteoblast and osteoclast formation. *Proc Natl Acad Sci USA*. 2006;103:14925–14930.
- 201. **Zhu LL, Blair H, Cao J, et al.** Blocking antibody to the β-subunit of FSH prevents bone loss by inhibiting bone resorption and stimulating bone synthesis. *Proc Natl Acad Sci USA*. 2012;109:14574–14579.
- 202. Geng W, Yan X, Du H, Cui J, Li L, Chen F. Immunization with FSHβ fusion protein antigen prevents bone loss in a rat ovariectomy-induced osteoporosis model. *Biochem Biophys Res Commun.* 2013;434:280–286.
- 203. Yarram SJ, Perry MJ, Christopher TJ, et al. Luteinizing hormone receptor knockout (LuRKO) mice and transgenic human chorionic gonadotropin (hCG)-overexpressing mice (hCG $\alpha\beta$ +) have bone phenotypes. *Endocrinology*. 2003;144:3555–3564.
- 204. Seibel MJ, Dunstan CR, Zhou H, Allan CM, Handelsman DJ. Sex steroids, not FSH, influence bone mass. *Cell.* 2006; 127:1079; author reply 1080–1081.
- 205. Gao J, Tiwari-Pandey R, Samadfam R, et al. Altered ovarian function affects skeletal homeostasis independent of the action of follicle-stimulating hormone. *Endocrinology*. 2007;148:2613–2621.
- 206. Rouach V, Katzburg S, Koch Y, Stern N, Somjen D. Bone loss in ovariectomized rats: dominant role for estrogen but apparently not for FSH. *J Cell Biochem*. 2011;112:128–137.
- 207. Allan CM, Kalak R, Dunstan CR, et al. Follicle-stimulating

- hormone increases bone mass in female mice. *Proc Natl Acad Sci USA*. 2010;107:22629-22634.
- 208. Finkelstein JS, Lee H, Burnett-Bowie SA, et al. Gonadal steroids and body composition, strength, and sexual function in men. *N Engl J Med*. 2013;369:1011–1022.
- 209. Uihlein AV, Finkelstein JS, Lee H, Leder BZ. FSH suppression does not affect bone turnover in eugonadal men. *J Clin Endocrinol Metab*. 2014;99:2510–2515.
- 210. Almeida M, Han L, Martin-Millan M, et al. Skeletal involution by age-associated oxidative stress and its acceleration by loss of sex steroids. *J Biol Chem.* 2007;282: 27285–27297.
- 211. **Manolagas SC.** Birth and death of bone cells: basic regulatory mechanisms and implications for the pathogenesis and treatment of osteoporosis. *Endocr Rev.* 2000;21:115–137.
- Nakamura T, Imai Y, Matsumoto T, et al. Estrogen prevents bone loss via estrogen receptor α and induction of Fas ligand in osteoclasts. *Cell*. 2007;130:811–823.
- 213. Seitz S, Keller J, Schilling AF, et al. Pharmacological estrogen administration causes a FSH-independent osteo-anabolic effect requiring ER α in osteoblasts. PLoS One. 2012; 7:e50301.
- 214. Krum SA, Miranda-Carboni GA, Hauschka PV, et al. Estrogen protects bone by inducing Fas ligand in osteoblasts to regulate osteoclast survival. *EMBO J.* 2008;27:535–545.
- 215. Michael H, Härkönen PL, Väänänen HK, Hentunen TA. Estrogen and testosterone use different cellular pathways to inhibit osteoclastogenesis and bone resorption. *J Bone Miner Res.* 2005;20:2224–2232.
- Turner AG, Notini AJ, Chiu WS, et al. Androgen receptor expression and function in osteoclasts. *Open Physiol J.* 2008;1:28–33.
- 217. Xiong J, Onal M, Jilka RL, Weinstein RS, Manolagas SC, O'Brien CA. Matrix-embedded cells control osteoclast formation. *Nat Med.* 2011;17:1235–1241.
- 218. Manolagas SC, O'Brien CA, Almeida M. The role of estrogen and androgen receptors in bone health and disease. *Nat Rev Endocrinol.* 2013;9:699–712.
- Määttä JA, Büki KG, Gu G, et al. Inactivation of estrogen receptor α in bone-forming cells induces bone loss in female mice. FASEB J. 2013;27:478–488.
- 220. Windahl SH, Börjesson AE, Farman HH, et al. Estrogen receptor-α in osteocytes is important for trabecular bone formation in male mice. *Proc Natl Acad Sci USA*. 2013; 110:2294–2299.
- 221. Melville KM, Kelly NH, Khan SA, et al. Female mice lacking estrogen receptor-α in osteoblasts have compromised bone mass and strength. J Bone Miner Res. 2014;29:370–379
- 222. Kondoh S, Inoue K, Igarashi K, et al. Estrogen receptor α in osteocytes regulates trabecular bone formation in female mice. *Bone*. 2014;60:68–77.
- 223. Ucer S, Warren A, Bartell S, et al. Unlike the estrogen receptor α, the androgen receptor in osteoblast progenitors is dispensable for optimal cortical bone accrual. In: Proceedings from the American Society for Bone and Mineral Research; October 4–7, 2013; Baltimore, MD. Presentation FR0446.
- 224. Wiren KM, Zhang XW, Toombs AR, et al. Targeted over-

- expression of androgen receptor in osteoblasts: unexpected complex bone phenotype in growing animals. *Endocrinology*, 2004;145:3507–3522.
- 225. Wiren KM, Semirale AA, Zhang XW, et al. Targeting of androgen receptor in bone reveals a lack of androgen anabolic action and inhibition of osteogenesis: a model for compartment-specific androgen action in the skeleton. *Bone*. 2008;43:440–451.
- 226. Börjesson AE, Lagerquist MK, Windahl SH, Ohlsson C. The role of estrogen receptor α in the regulation of bone and growth plate cartilage. *Cell Mol Life Sci.* 2013;70: 4023–4037.
- 227. Vanderschueren D, Boonen S, Ederveen AG, et al. Skeletal effects of estrogen deficiency as induced by an aromatase inhibitor in an aged male rat model. *Bone*. 2000;27:611–617.
- 228. Chagin AS, Lindberg MK, Andersson N, et al. Estrogen receptor-β inhibits skeletal growth and has the capacity to mediate growth plate fusion in female mice. *J Bone Miner Res.* 2004;19:72–77.
- 229. Börjesson AE, Lagerquist MK, Liu C, et al. The role of estrogen receptor α in growth plate cartilage for longitudinal bone growth. J Bone Miner Res. 2010;25:2690– 2700.
- 230. Russell PK, Clarke MV, Skinner JP, Pang TP, Zajac JD, Davey RA. Identification of gene pathways altered by deletion of the androgen receptor specifically in mineralizing osteoblasts and osteocytes in mice. *J Mol Endocrinol*. 2012;49:1–10.
- 231. Wiren KM, Semirale AA, Hashimoto JG, Zhang XW. Signaling pathways implicated in androgen regulation of endocortical bone. *Bone*. 2010;46:710–723.
- 232. Fujita K, Roforth MM, Demaray S, et al. Effects of estrogen on bone mRNA levels of sclerostin and other genes relevant to bone metabolism in postmenopausal women. *J Clin Endocrinol Metab*. 2014;99:E81–E88.
- 233. **Krum SA.** Direct transcriptional targets of sex steroid hormones in bone. *J Cell Biochem*. 2011;112:401–408.
- 234. Zaman G, Saxon LK, Sunters A, et al. Loading-related regulation of gene expression in bone in the contexts of estrogen deficiency, lack of estrogen receptor α and disuse. *Bone*. 2010;46:628–642.
- 235. Garcia AJ, Tom C, Guemes M, et al. $ER\alpha$ signaling regulates MMP3 expression to induce FasL cleavage and osteoclast apoptosis. *J Bone Miner Res.* 2013;28:283–290.
- 236. Krum SA, Miranda-Carboni GA, Lupien M, Eeckhoute J, Carroll JS, Brown M. Unique ERα cistromes control cell type-specific gene regulation. *Mol Endocrinol*. 2008;22: 2393–2406.
- Miranda-Carboni GA, Guemes M, Bailey S, et al. GATA4 regulates estrogen receptor-α-mediated osteoblast transcription. Mol Endocrinol. 2011;25:1126–1136.
- 238. Güemes M, Garcia AJ, Rigueur D, et al. GATA4 is essential for bone mineralization via ER α and TGF β /BMP pathways [published online June 16, 2014]. *J Bone Miner Res.* doi:10.1002/jbmr.2296.
- 239. Monroe DG, Secreto FJ, Hawse JR, Subramaniam M, Khosla S, Spelsberg TC. Estrogen receptor isoform-specific regulation of the retinoblastoma-binding protein 1 (RBBP1) gene: roles of AF1 and enhancer elements. *J Biol Chem*. 2006;281:28596–28604.

240. Hawse JR, Subramaniam M, Monroe DG, et al. Estrogen receptor β isoform-specific induction of transforming growth factor β -inducible early gene-1 in human osteoblast cells: an essential role for the activation function 1 domain. *Mol Endocrinol*. 2008;22:1579–1595.

- 241. Subramaniam M, Hawse JR, Bruinsma ES, et al. TGFβ inducible early gene-1 directly binds to, and represses, the OPG promoter in osteoblasts. *Biochem Biophys Res Commun.* 2010;392:72–76.
- 242. Monroe DG, Hawse JR, Subramaniam M, Spelsberg TC. Retinoblastoma binding protein-1 (RBP1) is a Runx2 coactivator and promotes osteoblastic differentiation. BMC Musculoskelet Disord. 2010;11:104.
- 243. Cicek M, Vrabel A, Sturchio C, et al. TGF-β inducible early gene 1 regulates osteoclast differentiation and survival by mediating the NFATc1, AKT, and MEK/ERK signaling pathways. *PLoS One*. 2011;6:e17522.
- 244. Hawse JR, Cicek M, Grygo SB, et al. TIEG1/KLF10 modulates Runx2 expression and activity in osteoblasts. *PLoS One*. 2011;6:e19429.
- 245. Glait-Santar C, Benayahu D. Regulation of SVEP1 gene expression by 17β-estradiol and TNFα in pre-osteoblastic and mammary adenocarcinoma cells. J Steroid Biochem Mol Biol. 2012;130:36–44.
- 246. Denger S, Bähr-Ivacevic T, Brand H, et al. Transcriptome profiling of estrogen-regulated genes in human primary osteoblasts reveals an osteoblast-specific regulation of the insulin-like growth factor binding protein 4 gene. *Mol Endocrinol*. 2008;22:361–379.
- 247. Lambertini E, Tavanti E, Torreggiani E, Penolazzi L, Gambari R, Piva R. ER α and AP-1 interact in vivo with a specific sequence of the F promoter of the human ER α gene in osteoblasts. *J Cell Physiol*. 2008;216:101–110.
- 248. Bord S, Ireland DC, Beavan SR, Compston JE. The effects of estrogen on osteoprotegerin, RANKL, and estrogen receptor expression in human osteoblasts. *Bone*. 2003;32: 136–141.
- 249. Monroe DG, Getz BJ, Johnsen SA, Riggs BL, Khosla S, Spelsberg TC. Estrogen receptor isoform-specific regulation of endogenous gene expression in human osteoblastic cell lines expressing either ERα or ERβ. *J Cell Biochem*. 2003;90:315–326.
- 250. Pederson L, Kremer M, Judd J, et al. Androgens regulate bone resorption activity of isolated osteoclasts in vitro. *Proc Natl Acad Sci USA*. 1999;96:505–510.
- 251. Baniwal SK, Khalid O, Sir D, Buchanan G, Coetzee GA, Frenkel B. Repression of Runx2 by androgen receptor (AR) in osteoblasts and prostate cancer cells: AR binds Runx2 and abrogates its recruitment to DNA. Mol Endocrinol. 2009;23:1203–1214.
- 252. Miki Y, Suzuki T, Hatori M, et al. Effects of aromatase inhibitors on human osteoblast and osteoblast-like cells: a possible androgenic bone protective effects induced by exemestane. *Bone*. 2007;40:876–887.
- 253. Ohlsson C, Mohan S, Sjögren K, et al. The role of liver-derived insulin-like growth factor-I. *Endocr Rev*. 2009;30: 494–535.
- 254. Olson LE, Ohlsson C, Mohan S. The role of GH/IGF-I-mediated mechanisms in sex differences in cortical bone size in mice. *Calcif Tissue Int*. 2011;88:1–8.
- 255. Giustina A, Mazziotti G, Canalis E. Growth hormone, in-

sulin-like growth factors, and the skeleton. *Endocr Rev*. 2008:29:535–559.

- 256. Courtland HW, DeMambro V, Maynard J, et al. Sex-specific regulation of body size and bone slenderness by the acid labile subunit. *J Bone Miner Res.* 2010;25:2059–2068.
- 257. Mohan S, Richman C, Guo R, et al. Insulin-like growth factor regulates peak bone mineral density in mice by both growth hormone-dependent and -independent mechanisms. *Endocrinology*. 2003;144:929–936.
- 258. Chang W, Parra M, Centrella M, McCarthy TL. Interactions between CCAAT enhancer binding protein δ and estrogen receptor α control insulin-like growth factor I (igf1) and estrogen receptor-dependent gene expression in osteoblasts. *Gene*. 2005;345:225–235.
- 259. Gori F, Hofbauer LC, Conover CA, Khosla S. Effects of androgens on the insulin-like growth factor system in an androgen-responsive human osteoblastic cell line. *Endocrinology*. 1999;140:5579–5586.
- 260. **Kasperk C, Fitzsimmons R, Strong D, et al.** Studies of the mechanism by which androgens enhance mitogenesis and differentiation in bone cells. *J Clin Endocrinol Metab.* 1990;71:1322–1329.
- 261. Kasperk CH, Faehling K, Börcsök I, Ziegler R. Effects of androgens on subpopulations of the human osteosarcoma cell line SaOS2. *Calcif Tissue Int*. 1996;58:376–382.
- 262. Bolamperti S, Mrak E, Moro G, et al. 17β-Estradiol positively modulates growth hormone signaling through the reduction of SOCS2 negative feedback in human osteoblasts. *Bone*. 2013;55:84–92.
- 263. Sanchez-Cardenas C, Fontanaud P, He Z, et al. Pituitary growth hormone network responses are sexually dimorphic and regulated by gonadal steroids in adulthood. *Proc Natl Acad Sci USA*. 2010;107:21878–21883.
- 264. Singhal V, Goh BC, Bouxsein ML, Faugere MC, DiGirolamo DJ. Osteoblast-restricted disruption of the growth hormone receptor in mice results in sexually dimorphic skeletal phenotypes. *Bone Res.* 2013;1:85–97.
- 265. Venken K, Schuit F, Van Lommel L, et al. Growth without growth hormone receptor: estradiol is a major growth hormone-independent regulator of hepatic IGF-I synthesis. *J Bone Miner Res.* 2005;20:2138–2149.
- 266. Govoni KE, Wergedal JE, Chadwick RB, Srivastava AK, Mohan S. Prepubertal OVX increases IGF-I expression and bone accretion in C57BL/6J mice. *Am J Physiol Endocrinol Metab.* 2008;295:E1172–E1180.
- 267. Callewaert F, Sinnesael M, Gielen E, Boonen S, Vanderschueren D. Skeletal sexual dimorphism: relative contribution of sex steroids, GH-IGF1, and mechanical loading. *J Endocrinol.* 2010;207:127–134.
- 268. Vanderschueren D, Venken K, Ophoff J, Bouillon R, Boonen S. Clinical review: sex steroids and the periosteum–reconsidering the roles of androgens and estrogens in periosteal expansion. *J Clin Endocrinol Metab.* 2006;91:378–382.
- 269. Vandenput L, Sjögren K, Svensson J, Ohlsson C. The role of IGF-1 for fracture risk in men. *Front Endocrinol (Lausanne)*. 2012;3:51.
- 270. Szulc P, Joly-Pharaboz MO, Marchand F, Delmas PD. Insulin-like growth factor I is a determinant of hip bone mineral density in men less than 60 years of age: MINOS study. *Calcif Tissue Int.* 2004;74:322–329.

- 271. Boonen S, Pye SR, O'Neill TW, et al. Influence of bone remodelling rate on quantitative ultrasound parameters at the calcaneus and DXA BMDa of the hip and spine in middle-aged and elderly European men: the European Male Ageing Study (EMAS). *Eur J Endocrinol*. 2011;165: 977–986.
- 272. Ohlsson C, Mellström D, Carlzon D, et al. Older men with low serum IGF-1 have an increased risk of incident fractures: the MrOS Sweden study. *J Bone Miner Res.* 2011; 26:865–872.
- 273. Lapauw B, T'Sjoen G, Mahmoud A, Kaufman JM, Ruige JB. Short-term aromatase inhibition: effects on glucose metabolism and serum leptin levels in young and elderly men. *Eur J Endocrinol.* 2009;160:397–402.
- 274. Burnett-Bowie SA, McKay EA, Lee H, Leder BZ. Effects of aromatase inhibition on bone mineral density and bone turnover in older men with low testosterone levels. *J Clin Endocrinol Metab*. 2009;94:4785–4792.
- Leder BZ, Finkelstein JS. Effect of aromatase inhibition on bone metabolism in elderly hypogonadal men. Osteoporos Int. 2005;16:1487–1494.
- 276. Falahati-Nini A, Riggs BL, Atkinson EJ, O'Fallon WM, Eastell R, Khosla S. Relative contributions of testosterone and estrogen in regulating bone resorption and formation in normal elderly men. *J Clin Invest*. 2000;106:1553– 1560.
- 277. **Bouillon R, Van Schoor NM, Gielen E, et al.** Optimal vitamin D status: a critical analysis on the basis of evidence-based medicine. *J Clin Endocrinol Metab.* 2013;98: E1283–E1304.
- 278. Curtis JR, Ewing SK, Bauer DC, et al. Association of intact parathyroid hormone levels with subsequent hip BMD loss: the Osteoporotic Fractures in Men (MrOS) Study. *J Clin Endocrinol Metab.* 2012;97:1937–1944.
- 279. Vanderschueren D, Pye SR, O'Neill TW, et al. Active vitamin D (1,25-dihydroxyvitamin D) and bone health in middle-aged and elderly men: the European Male Aging Study (EMAS). J Clin Endocrinol Metab. 2013;98:995–1005.
- 280. Ensrud KE, Taylor BC, Paudel ML, et al. Serum 25-hydroxyvitamin D levels and rate of hip bone loss in older men. *J Clin Endocrinol Metab*. 2009;94:2773–2780.
- 281. Cauley JA, Parimi N, Ensrud KE, et al. Serum 25-hydroxyvitamin D and the risk of hip and nonspine fractures in older men. *J Bone Miner Res.* 2010;25:545–553.
- 282. Barrett-Connor E, Laughlin GA, Li H, et al. The association of concurrent vitamin D and sex hormone deficiency with bone loss and fracture risk in older men: the osteoporotic fractures in men (MrOS) study. *J Bone Miner Res.* 2012;27:2306–2313.
- 283. Wehr E, Pilz S, Boehm BO, März W, Obermayer-Pietsch B. Association of vitamin D status with serum androgen levels in men. *Clin Endocrinol (Oxf)*. 2010;73:243–248.
- 284. Lee DM, Tajar A, Pye SR, et al. Association of hypogonadism with vitamin D status: the European Male Ageing Study. *Eur J Endocrinol*. 2012;166:77–85.
- 285. Pilz S, Frisch S, Koertke H, et al. Effect of vitamin D supplementation on testosterone levels in men. *Horm Metab Res.* 2011;43:223–225.
- 286. Somjen D, Katzburg S, Stern N, et al. 25 Hydroxy-vitamin D(3)-1 α hydroxylase expression and activity in cultured

- human osteoblasts and their modulation by parathyroid hormone, estrogenic compounds and dihydrotestosterone. *J Steroid Biochem Mol Biol.* 2007;107:238–244.
- 287. Hsu YJ, Dimke H, Schoeber JP, et al. Testosterone increases urinary calcium excretion and inhibits expression of renal calcium transport proteins. *Kidney Int.* 2010;77: 601–608.
- 288. Meng J, Ohlsson C, Laughlin GA, et al. Associations of estradiol and testosterone with serum phosphorus in older men: the Osteoporotic Fractures in Men study. *Kidney Int.* 2010;78:415–422.
- 289. Frenkel B, Hong A, Baniwal SK, et al. Regulation of adult bone turnover by sex steroids. *J Cell Physiol*. 2010;224: 305–310.
- 290. **Khalid O, Baniwal SK, Purcell DJ, et al.** Modulation of Runx2 activity by estrogen receptor-α: implications for osteoporosis and breast cancer. *Endocrinology*. 2008;149: 5984–5995.
- 291. McCarthy TL, Chang WZ, Liu Y, Centrella M. Runx2 integrates estrogen activity in osteoblasts. *J Biol Chem*. 2003;278:43121–43129.
- 292. Kawate H, Wu Y, Ohnaka K, Takayanagi R. Mutual transactivational repression of Runx2 and the androgen receptor by an impairment of their normal compartmentalization. *J Steroid Biochem Mol Biol.* 2007;105:46–56.
- 293. **Jeong JH, Jung YK, Kim HJ, et al.** The gene for aromatase, a rate-limiting enzyme for local estrogen biosynthesis, is a downstream target gene of Runx2 in skeletal tissues. *Mol Cell Biol.* 2010;30:2365–2375.
- 294. Maruyama Z, Yoshida CA, Furuichi T, et al. Runx2 determines bone maturity and turnover rate in postnatal bone development and is involved in bone loss in estrogen deficiency. *Dev Dyn.* 2007;236:1876–1890.
- 295. Ke HZ, Richards WG, Li X, Ominsky MS. Sclerostin and Dickkopf-1 as therapeutic targets in bone diseases. *Endocr Rev.* 2012;33:747–783.
- 296. Li X, Ominsky MS, Niu QT, et al. Targeted deletion of the sclerostin gene in mice results in increased bone formation and bone strength. J Bone Miner Res. 2008;23:860–869.
- 297. Kramer I, Halleux C, Keller H, et al. Osteocyte Wnt/β-catenin signaling is required for normal bone homeostasis. *Mol Cell Biol.* 2010;30:3071–3085.
- 298. Javaheri B, Stern AR, Lara N, et al. Deletion of a single β -catenin allele in osteocytes abolishes the bone anabolic response to loading. *J Bone Miner Res.* 2014;29:705–715.
- 299. Noh T, Gabet Y, Cogan J, et al. Lef1 haploinsufficient mice display a low turnover and low bone mass phenotype in a gender- and age-specific manner. *PLoS One*. 2009;4: e5438.
- 300. Yao W, Cheng Z, Shahnazari M, Dai W, Johnson ML, Lane NE. Overexpression of secreted frizzled-related protein 1 inhibits bone formation and attenuates parathyroid hormone bone anabolic effects. *J Bone Miner Res.* 2010; 25:190–199.
- Kramer I, Baertschi S, Halleux C, Keller H, Kneissel M. Mef2c deletion in osteocytes results in increased bone mass. J Bone Miner Res. 2012;27:360-373.
- 302. Saxon LK, Jackson BF, Sugiyama T, Lanyon LE, Price JS. Analysis of multiple bone responses to graded strains above functional levels, and to disuse, in mice in vivo show that the human Lrp5 G171V high bone mass mutation

increases the osteogenic response to loading but that lack of Lrp5 activity reduces it. *Bone*. 2011;49:184–193.

- 303. Liu XH, Kirschenbaum A, Yao S, Levine AC. Androgens promote preosteoblast differentiation via activation of the canonical Wnt signaling pathway. *Ann NY Acad Sci.* 2007; 1116:423–431.
- 304. Jastrzebski S, Kalinowski J, Stolina M, et al. Changes in bone sclerostin levels in mice after ovariectomy vary independently of changes in serum sclerostin levels. *J Bone Miner Res.* 2013;28:618–626.
- 305. Kim BJ, Bae SJ, Lee SY, et al. TNF-α mediates the stimulation of sclerostin expression in an estrogen-deficient condition. *Biochem Biophys Res Commun*. 2012;424:170–175.
- 306. Durosier C, van Lierop A, Ferrari S, Chevalley T, Papapoulos S, Rizzoli R. Association of circulating sclerostin with bone mineral mass, microstructure, and turnover biochemical markers in healthy elderly men and women. *J Clin Endocrinol Metab*. 2013;98:3873–3883.
- Roforth MM, Fujita K, McGregor UI, et al. Effects of age on bone mRNA levels of sclerostin and other genes relevant to bone metabolism in humans. *Bone*. 2014;59:1–6.
- 308. Mödder UI, Hoey KA, Amin S, et al. Relation of age, gender, and bone mass to circulating sclerostin levels in women and men. *J Bone Miner Res.* 2011;26:373–379.
- 309. Amrein K, Amrein S, Drexler C, et al. Sclerostin and its association with physical activity, age, gender, body composition, and bone mineral content in healthy adults. *J Clin Endocrinol Metab*. 2012;97:148–154.
- 310. Szulc P, Bertholon C, Borel O, Marchand F, Chapurlat R. Lower fracture risk in older men with higher sclerostin concentration: a prospective analysis from the MINOS study. *J Bone Miner Res.* 2013;28:855–864.
- 311. Szulc P, Boutroy S, Vilayphiou N, et al. Correlates of bone microarchitectural parameters and serum sclerostin levels in men: the STRAMBO study. *J Bone Miner Res.* 2013; 28:1760–1770.
- 312. Lapauw B, Vandewalle S, Taes Y, et al. Serum sclerostin levels in men with idiopathic osteoporosis. *Eur J Endocrinol*. 2013;168:615–620.
- 313. Kuipers AL, Zhang Y, Yu S, et al. Relative influence of heritability, environment and genetics on serum sclerostin. *Osteoporos Int.* 2014;25:905–912.
- Kirmani S, Amin S, McCready LK, et al. Sclerostin levels during growth in children. Osteoporos Int. 2012;23: 1123–1130.
- 315. Mödder UI, Clowes JA, Hoey K, et al. Regulation of circulating sclerostin levels by sex steroids in women and in men. *J Bone Miner Res.* 2011;26:27–34.
- Lee K, Jessop H, Suswillo R, Zaman G, Lanyon L. Endocrinology: bone adaptation requires oestrogen receptor-α.
 Nature. 2003;424:389.
- 317. Saxon LK, Galea G, Meakin L, Price J, Lanyon LE. Estrogen receptors α and β have different gender-dependent effects on the adaptive responses to load bearing in cancellous and cortical bone. *Endocrinology*. 2012;153:2254–2266.
- 318. Sugiyama T, Galea GL, Lanyon LE, Price JS. Mechanical loading-related bone gain is enhanced by tamoxifen but unaffected by fulvestrant in female mice. *Endocrinology*. 2010;151:5582–5590.

319. Sample SJ, Racette MA, Hao Z, Thomas CF, Behan M, Muir P. Functional adaptation in female rats: the role of estrogen signaling. *PLoS One*. 2012;7:e43215.

- 320. Saxon LK, Turner CH. Low-dose estrogen treatment suppresses periosteal bone formation in response to mechanical loading. *Bone*. 2006;39:1261–1267.
- 321. Järvinen TL, Kannus P, Pajamäki I, et al. Estrogen deposits extra mineral into bones of female rats in puberty, but simultaneously seems to suppress the responsiveness of female skeleton to mechanical loading. *Bone*. 2003;32:642–651
- 322. **Jagger CJ, Chow JW, Chambers TJ.** Estrogen suppresses activation but enhances formation phase of osteogenic response to mechanical stimulation in rat bone. *J Clin Invest*. 1996;98:2351–2357.
- 323. Tromp AM, Bravenboer N, Tanck E, et al. Additional weight bearing during exercise and estrogen in the rat: the effect on bone mass, turnover, and structure. *Calcif Tissue Int.* 2006;79:404–415.
- 324. Pajamäki I, Sievänen H, Kannus P, Jokihaara J, Vuohelainen T, Järvinen TL. Skeletal effects of estrogen and mechanical loading are structurally distinct. *Bone*. 2008;43: 748–757.
- 325. Callewaert F, Bakker A, Schrooten J, et al. Androgen receptor disruption increases the osteogenic response to mechanical loading in male mice. *J Bone Miner Res.* 2010; 25:124–131.
- 326. Armstrong VJ, Muzylak M, Sunters A, et al. Wnt/β-catenin signaling is a component of osteoblastic bone cell early responses to load-bearing and requires estrogen receptor α. *J Biol Chem.* 2007;282:20715–20727.
- 327. Sunters A, Armstrong VJ, Zaman G, et al. Mechano-transduction in osteoblastic cells involves strain-regulated estrogen receptor α-mediated control of insulin-like growth factor (IGF) I receptor sensitivity to ambient IGF, leading to phosphatidylinositol 3-kinase/AKT-dependent Wnt/LRP5 receptor-independent activation of β-catenin signaling. *J Biol Chem.* 2010;285:8743–8758.
- 328. Meakin LB, Sugiyama T, Galea GL, Browne WJ, Lanyon LE, Price JS. Male mice housed in groups engage in frequent fighting and show a lower response to additional bone loading than females or individually housed males that do not fight. *Bone*. 2013;54:113–117.
- 329. Eisenmann JC, Wickel EE, Kelly SA, Middleton KM, Garland T Jr. Day-to-day variability in voluntary wheel running among genetically differentiated lines of mice that vary in activity level. *Eur J Appl Physiol*. 2009;106:613–619.
- 330. Carrera O, Cerrato M, Vazquez R, Sineiro C, Gutierrez E. Gender dimorphic effects of voluntary running in laboratory rats depends on maturational status. *Q J Exp Psychol (Hove)*. 2011;64:823–832.
- 331. Zaman G, Jessop HL, Muzylak M, et al. Osteocytes use estrogen receptor α to respond to strain but their ER α content is regulated by estrogen. *J Bone Miner Res.* 2006; 21:1297–1306.
- 332. Ren J, Wang XH, Wang GC, Wu JH. 17β Estradiol regulation of connexin 43-based gap junction and mechanosensitivity through classical estrogen receptor pathway in osteocyte-like MLO-Y4 cells. *Bone*. 2013;53:587–596.
- 333. Pope HG Jr, Wood RI, Rogol A, Nyberg F, Bowers L,

Vanderschueren et al

- Bhasin S. Adverse health consequences of performanceenhancing drugs: an Endocrine Society scientific statement. Endocr Rev. 2014;35:341-375.
- 334. Dubois V, Laurent M, Boonen S, Vanderschueren D, Claessens F. Androgens and skeletal muscle: cellular and molecular action mechanisms underlying the anabolic actions. Cell Mol Life Sci. 2012;69:1651-1667.
- 335. Velders M, Diel P. How sex hormones promote skeletal muscle regeneration. Sports Med. 2013;43:1089–1100.
- 336. Bhasin S, Travison TG, Storer TW, et al. Effect of testosterone supplementation with and without a dual 5α -reductase inhibitor on fat-free mass in men with suppressed testosterone production: a randomized controlled trial. *JAMA*. 2012;307:931–939.
- 337. Borst SE, Yarrow JF, Conover CF, et al. Musculoskeletal and prostate effects of combined testosterone and finasteride administration in older hypogonadal men: a randomized, controlled trial. Am J Physiol Endocrinol Metab. 2014;306:E433-E442.
- 338. Lebrasseur NK, Achenbach SJ, Melton LJ 3rd, Amin S, Khosla S. Skeletal muscle mass is associated with bone geometry and microstructure and serum insulin-like growth factor binding protein-2 levels in adult women and men. J Bone Miner Res. 2012;27:2159-2169.
- 339. Ophoff J, Van Proeyen K, Callewaert F, et al. Androgen signaling in myocytes contributes to the maintenance of muscle mass and fiber type regulation but not to muscle strength or fatigue. Endocrinology. 2009;150:3558-3566.
- 340. Chambon C, Duteil D, Vignaud A, et al. Myocytic androgen receptor controls the strength but not the mass of limb muscles. Proc Natl Acad Sci USA. 2010;107:14327-14332.
- 341. Dubois V, Laurent MR, Sinnesael M, et al. A satellite cellspecific knockout of the androgen receptor reveals myostatin as a direct androgen target in skeletal muscle. FASEB *I*. 2014;28:2979–2994.
- 342. Verschueren S, Gielen E, O'Neill TW, et al. Sarcopenia and its relationship with bone mineral density in middleaged and elderly European men. Osteoporos Int. 2013; 24:87-98.
- 343. Wimalawansa SM, Chapa MT, Wei JN, Westlund KN, Quast MJ, Wimalawansa SJ. Reversal of weightlessnessinduced musculoskeletal losses with androgens: quantification by MRI. J Appl Physiol (1985). 1999;86:1841-1846.
- 344. Cardozo CP, Qin W, Peng Y, et al. Nandrolone slows hindlimb bone loss in a rat model of bone loss due to denervation. Ann NY Acad Sci. 2010;1192:303-306.
- 345. Yarrow JF, Conover CF, Beggs LA, et al. Testosterone dose dependently prevents bone and muscle loss in rodents after spinal cord injury. J Neurotrauma. 2014;31:834-845.
- 346. Libouban H, Blouin S, Moreau MF, Baslé MF, Audran M, Chappard D. Effects of risedronate in a rat model of osteopenia due to orchidectomy and disuse: densitometric, histomorphometric and microtomographic studies. Micron. 2008;39:998-1007.
- 347. Maggi A, Ciana P, Belcredito S, Vegeto E. Estrogens in the nervous system: mechanisms and nonreproductive functions. Annu Rev Physiol. 2004;66:291-313.
- 348. Burt-Pichat B, Lafage-Proust MH, Duboeuf F, et al. Dra-

- matic decrease of innervation density in bone after ovariectomy. Endocrinology. 2005;146:503-510.
- 349. Bouxsein ML, Devlin MJ, Glatt V, Dhillon H, Pierroz DD, Ferrari SL. Mice lacking β -adrenergic receptors have increased bone mass but are not protected from deleterious skeletal effects of ovariectomy. Endocrinology. 2009;150:
- 350. Farr JN, Charkoudian N, Barnes JN, et al. Relationship of sympathetic activity to bone microstructure, turnover, and plasma osteopontin levels in women. I Clin Endocrinol Metab. 2012:97:4219-4227.
- 351. Ohlsson C, Engdahl C, Börjesson AE, et al. Estrogen receptor- α expression in neuronal cells affects bone mass. Proc Natl Acad Sci USA. 2012;109:983-988.
- 352. Farman HH, Windahl SH, Clegg DJ, et al. Mice lacking estrogen receptor α in hypothalamic pro-opiomelanocortin (POMC) neurons display enhanced estrogenic response on bone mass. In: Proceedings from the American Society for Bone and Mineral Research; October 4-7, 2013; Baltimore, MD. Presentation FR0445.
- 353. Reid IR. Fat and bone. Arch Biochem Biophys. 2010;503: 20-27.
- 354. Ng AC, Melton LJ 3rd, Atkinson EJ, et al. Relationship of adiposity to bone volumetric density and microstructure in men and women across the adult lifespan. Bone. 2013;55:
- 355. Emmelot-Vonk MH, Verhaar HJ, Nakhai Pour HR, et al. Effect of testosterone supplementation on functional mobility, cognition, and other parameters in older men: a randomized controlled trial. JAMA. 2008;299:39-52.
- 356. Kenny AM, Kleppinger A, Annis K, et al. Effects of transdermal testosterone on bone and muscle in older men with low bioavailable testosterone levels, low bone mass, and physical frailty. J Am Geriatr Soc. 2010;58:1134-1143.
- 357. Bouloux PM, Legros JJ, Elbers JM, et al. Effects of oral testosterone undecanoate therapy on bone mineral density and body composition in 322 aging men with symptomatic testosterone deficiency: a 1-year, randomized, placebocontrolled, dose-ranging study. Aging Male. 2013;16: 38 - 47.
- 358. Sinnesael M, Callewaert F, Morreels M, et al. 7α -Methyl-19-nortestosterone vs. testosterone implants for hypogonadal osteoporosis: a preclinical study in the aged male orchidectomized rat model. Int J Androl. 2011;34:e601e611.
- 359. Dalton JT, Barnette KG, Bohl CE, et al. The selective androgen receptor modulator GTx-024 (enobosarm) improves lean body mass and physical function in healthy elderly men and postmenopausal women: results of a double-blind, placebo-controlled phase II trial. J Cachexia Sarcopenia Muscle. 2011;2:153-161.
- 360. Huang CK, Lai KP, Luo J, et al. Loss of androgen receptor promotes adipogenesis but suppresses osteogenesis in bone marrow stromal cells. Stem Cell Res. 2013;11:938-950.
- 361. Heine PA, Taylor JA, Iwamoto GA, Lubahn DB, Cooke PS. Increased adipose tissue in male and female estrogen receptor-α knockout mice. *Proc Natl Acad Sci USA*. 2000; 97:12729-12734.
- 362. Sato T, Matsumoto T, Yamada T, Watanabe T, Kawano H, Kato S. Late onset of obesity in male androgen receptor-

deficient (AR KO) mice. Biochem Biophys Res Commun. 2003;300:167-171.

- 363. Ohlsson C, Hellberg N, Parini P, et al. Obesity and disturbed lipoprotein profile in estrogen receptor-α-deficient male mice. *Biochem Biophys Res Commun.* 2000;278: 640–645.
- 364. McInnes KJ, Smith LB, Hunger NI, Saunders PT, Andrew R, Walker BR. Deletion of the androgen receptor in adipose tissue in male mice elevates retinol binding protein 4 and reveals independent effects on visceral fat mass and on glucose homeostasis. *Diabetes*. 2012;61:1072–1081.
- 365. Pacifici R. Role of T cells in ovariectomy induced bone loss-revisited. *J Bone Miner Res.* 2012;27:231–239.
- 366. Bellido T, Jilka RL, Boyce BF, et al. Regulation of interleukin-6, osteoclastogenesis, and bone mass by androgens. The role of the androgen receptor. *J Clin Invest*. 1995;95: 2886–2895.
- 367. Onal M, Xiong J, Chen X, et al. Receptor activator of nuclear factor κB ligand (RANKL) protein expression by B lymphocytes contributes to ovariectomy-induced bone loss. *J Biol Chem.* 2012;287:29851–29860.
- 368. Altuwaijri S, Chuang KH, Lai KP, et al. Susceptibility to autoimmunity and B cell resistance to apoptosis in mice lacking androgen receptor in B cells. *Mol Endocrinol*. 2009;23:444–453.
- 369. **Lélu K**, **Laffont S**, **Delpy L**, **et al.** Estrogen receptor α signaling in T lymphocytes is required for estradiol-mediated inhibition of Th1 and Th17 cell differentiation and protection against experimental autoimmune encephalomyelitis. *J Immunol*. 2011;187:2386–2393.
- 370. Kuchuk NO, van Schoor NM, Pluijm SM, Smit JH, de Ronde W, Lips P. The association of sex hormone levels with quantitative ultrasound, bone mineral density, bone turnover and osteoporotic fractures in older men and women. *Clin Endocrinol (Oxf)*. 2007;67:295–303.
- 371. Araujo AB, Travison TG, Leder BZ, McKinlay JB. Correlations between serum testosterone, estradiol, and sex hormone-binding globulin and bone mineral density in a diverse sample of men. *J Clin Endocrinol Metab*. 2008;93: 2135–2141.
- 372. Paller CJ, Shiels MS, Rohrmann S, et al. Relationship of sex steroid hormones with bone mineral density (BMD) in a nationally representative sample of men. *Clin Endocrinol* (Oxf). 2009;70:26–34.
- 373. Woo J, Kwok T, Leung JC, Ohlsson C, Vandenput L, Leung PC. Sex steroids and bone health in older Chinese men. *Osteoporos Int.* 2012;23:1553–1562.
- 374. Laurent M, Antonio L, Sinnesael M, et al. Androgens and estrogens in skeletal sexual dimorphism. *Asian J Androl*. 2014;16:213–222.
- 375. Khosla S, Melton LJ 3rd, Atkinson EJ, O'Fallon WM. Relationship of serum sex steroid levels to longitudinal changes in bone density in young versus elderly men. *J Clin Endocrinol Metab*. 2001;86:3555–3561.
- 376. Gennari L, Merlotti D, Martini G, et al. Longitudinal association between sex hormone levels, bone loss, and bone turnover in elderly men. *J Clin Endocrinol Metab*. 2003; 88:5327–5333.
- 377. Van Pottelbergh I, Goemaere S, Kaufman JM. Bioavailable estradiol and an aromatase gene polymorphism are determinants of bone mineral density changes in men over 70

- years of age. J Clin Endocrinol Metab. 2003;88:3075-3081.
- 378. Cauley JA, Ewing SK, Taylor BC, et al. Sex steroid hormones in older men: longitudinal associations with 4.5-year change in hip bone mineral density—the Osteoporotic Fractures in Men Study. *J Clin Endocrinol Metab*. 2010; 95:4314–4323.
- 379. Khosla S, Melton LJ 3rd, Robb RA, et al. Relationship of volumetric BMD and structural parameters at different skeletal sites to sex steroid levels in men. *J Bone Miner Res*. 2005;20:730–740.
- 380. Ward KA, Pye SR, Adams JE, et al. Influence of age and sex steroids on bone density and geometry in middle-aged and elderly European men. *Osteoporos Int.* 2011;22:1513–1523.
- 381. Khosla S, Melton LJ 3rd, Achenbach SJ, Oberg AL, Riggs BL. Hormonal and biochemical determinants of trabecular microstructure at the ultradistal radius in women and men. *J Clin Endocrinol Metab.* 2006;91:885–891.
- 382. Argoud T, Boutroy S, Claustrat B, Chapurlat R, Szulc P. Association between sex steroid levels and bone microarchitecture in men: the STRAMBO study. *J Clin Endocrinol Metab*. 2014;99:1400–1410.
- 383. Szulc P, Uusi-Rasi K, Claustrat B, Marchand F, Beck TJ, Delmas PD. Role of sex steroids in the regulation of bone morphology in men. The MINOS study. *Osteoporos Int.* 2004;15:909–917.
- 384. Travison TG, Araujo AB, Beck TJ, et al. Relation between serum testosterone, serum estradiol, sex hormone-binding globulin, and geometrical measures of adult male proximal femur strength. *J Clin Endocrinol Metab*. 2009;94:853–860.
- 385. Shahinian VB, Kuo YF, Freeman JL, Goodwin JS. Risk of fracture after androgen deprivation for prostate cancer. *N Engl J Med.* 2005;352:154–164.
- 386. Smith MR, Goode M, Zietman AL, McGovern FJ, Lee H, Finkelstein JS. Bicalutamide monotherapy versus leuprolide monotherapy for prostate cancer: effects on bone mineral density and body composition. *J Clin Oncol*. 2004; 22:2546–2553.
- 387. Wadhwa VK, Weston R, Mistry R, Parr NJ. Long-term changes in bone mineral density and predicted fracture risk in patients receiving androgen-deprivation therapy for prostate cancer, with stratification of treatment based on presenting values. *BJU Int.* 2009;104:800–805.
- 388. Mueller A, Zollver H, Kronawitter D, et al. Body composition and bone mineral density in male-to-female transsexuals during cross-sex hormone therapy using gonadotrophin-releasing hormone agonist. *Exp Clin Endocrinol Diabetes*. 2011;119:95–100.
- 389. Ruetsche AG, Kneubuehl R, Birkhaeuser MH, Lippuner K. Cortical and trabecular bone mineral density in transsexuals after long-term cross-sex hormonal treatment: a cross-sectional study. *Osteoporos Int.* 2005;16:791–798.
- 390. Gennari L, Masi L, Merlotti D, et al. A polymorphic CYP19 TTTA repeat influences aromatase activity and estrogen levels in elderly men: effects on bone metabolism. *J Clin Endocrinol Metab.* 2004;89:2803–2810.
- 391. Eriksson AL, Lorentzon M, Vandenput L, et al. Genetic variations in sex steroid-related genes as predictors of se-

Vanderschueren et al.

- rum estrogen levels in men. J Clin Endocrinol Metab. 2009; 94:1033-1041.
- 392. Limer KL, Pye SR, Thomson W, et al. Genetic variation in sex hormone genes influences heel ultrasound parameters in middle-aged and elderly men: results from the European Male Aging Study (EMAS). J Bone Miner Res. 2009;24: 314-323.
- 393. Huhtaniemi IT, Pye SR, Limer KL, et al. Increased estrogen rather than decreased androgen action is associated with longer androgen receptor CAG repeats. J Clin Endocrinol Metab. 2009;94:277-284.
- 394. Rivadeneira F, Styrkársdottir U, Estrada K, et al. Twenty bone-mineral-density loci identified by large-scale metaanalysis of genome-wide association studies. Nat Genet. 2009;41:1199-1206.
- 395. Styrkarsdottir U, Halldorsson BV, Gretarsdottir S, et al. Multiple genetic loci for bone mineral density and fractures. N Engl J Med. 2008;358:2355-2365.
- 396. Holliday KL, Pye SR, Thomson W, et al. The ESR1 (6q25) locus is associated with calcaneal ultrasound parameters and radial volumetric bone mineral density in European men. PLoS One. 2011;6:e22037.
- 397. Adler RA. Management of osteoporosis in men on androgen deprivation therapy. Maturitas. 2011;68:143-147.
- 398. Smith MR, Fallon MA, Lee H, Finkelstein JS. Raloxifene to prevent gonadotropin-releasing hormone agonist-induced bone loss in men with prostate cancer: a randomized controlled trial. J Clin Endocrinol Metab. 2004;89:3841-3846.
- 399. Smith MR, Morton RA, Barnette KG, et al. Toremifene to reduce fracture risk in men receiving androgen deprivation therapy for prostate cancer. *J Urol.* 2010;184:1316–1321.
- 400. Jankowski CM, Gozansky WS, Kittelson JM, Van Pelt RE, Schwartz RS, Kohrt WM. Increases in bone mineral density in response to oral dehydroepiandrosterone replacement in older adults appear to be mediated by serum estrogens. I Clin Endocrinol Metab. 2008;93:4767-4773.
- 401. Nair KS, Rizza RA, O'Brien P, et al. DHEA in elderly women and DHEA or testosterone in elderly men. N Engl *J Med.* 2006;355:1647–1659.
- 402. Weiss EP, Shah K, Fontana L, Lambert CP, Holloszy JO, Villareal DT. Dehydroepiandrosterone replacement therapy in older adults: 1- and 2-y effects on bone. Am I Clin Nutr. 2009;89:1459-1467.
- 403. von Mühlen D, Laughlin GA, Kritz-Silverstein D, Bergstrom J, Bettencourt R. Effect of dehydroepiandrosterone supplementation on bone mineral density, bone markers, and body composition in older adults: the DAWN trial. Osteoporos Int. 2008;19:699-707.
- 404. Leder BZ, LeBlanc KM, Schoenfeld DA, Eastell R, Finkelstein IS. Differential effects of androgens and estrogens on bone turnover in normal men. J Clin Endocrinol Metab. 2003;88:204-210.
- 405. Yu E, Taylor A, Wulcyzn K, et al. Hypogonadism with estrogen removal (HER): differential effects of androgens and estrogens on bone microarchitecture in adult men. I Bone Miner Res. 2012;27:S66.
- 406. Khosla S. Update in male osteoporosis. J Clin Endocrinol Metab. 2010;95:3-10.
- 407. Barrett-Connor E, Mueller JE, von Mühlen DG, Laughlin GA, Schneider DL, Sartoris DJ. Low levels of estradiol are

- associated with vertebral fractures in older men, but not women: the Rancho Bernardo Study. J Clin Endocrinol Metab. 2000;85:219-223.
- 408. Bjørnerem A, Ahmed LA, Joakimsen RM, et al. A prospective study of sex steroids, sex hormone-binding globulin, and non-vertebral fractures in women and men: the Tromso Study. Eur J Endocrinol. 2007;157:119-125.
- 409. Center JR, Nguyen TV, Sambrook PN, Eisman JA. Hormonal and biochemical parameters and osteoporotic fractures in elderly men. J Bone Miner Res. 2000;15:1405-
- 410. Goderie-Plomp HW, van der Klift M, de Ronde W, Hofman A, de Jong FH, Pols HA. Endogenous sex hormones, sex hormone-binding globulin, and the risk of incident vertebral fractures in elderly men and women: the Rotterdam Study. J Clin Endocrinol Metab. 2004;89:3261-3269.
- 411. Legrand E, Hedde C, Gallois Y, et al. Osteoporosis in men: a potential role for the sex hormone binding globulin. Bone. 2001;29:90-95.
- 412. Lormeau C, Soudan B, d'Herbomez M, Pigny P, Duquesnoy B, Cortet B. Sex hormone-binding globulin, estradiol, and bone turnover markers in male osteoporosis. Bone. 2004;34:933-939.
- 413. Mellström D, Johnell O, Ljunggren O, et al. Free testosterone is an independent predictor of BMD and prevalent fractures in elderly men: MrOS Sweden. I Bone Miner Res. 2006;21:529-535.
- 414. Roddam AW, Appleby P, Neale R, et al. Association between endogenous plasma hormone concentrations and fracture risk in men and women: the EPIC-Oxford prospective cohort study. J Bone Miner Metab. 2009;27:485-493.
- 415. Rosner W, Hankinson SE, Sluss PM, Vesper HW, Wierman ME. Challenges to the measurement of estradiol: an Endocrine Society position statement. J Clin Endocrinol Metab. 2013;98:1376-1387.
- 416. Amin S, Zhang Y, Felson DT, et al. Estradiol, testosterone, and the risk for hip fractures in elderly men from the Framingham Study. Am J Med. 2006;119:426-433.
- 417. Mellström D, Vandenput L, Mallmin H, et al. Older men with low serum estradiol and high serum SHBG have an increased risk of fractures. I Bone Miner Res. 2008;23: 1552-1560.
- 418. LeBlanc ES, Nielson CM, Marshall LM, et al. The effects of serum testosterone, estradiol, and sex hormone binding globulin levels on fracture risk in older men. J Clin Endocrinol Metab. 2009;94:3337-3346.
- 419. Fink HA, Ewing SK, Ensrud KE, et al. Association of testosterone and estradiol deficiency with osteoporosis and rapid bone loss in older men. J Clin Endocrinol Metab. 2006;91:3908-3915.
- 420. van den Beld AW, de Jong FH, Grobbee DE, Pols HA, Lamberts SW. Measures of bioavailable serum testosterone and estradiol and their relationships with muscle strength, bone density, and body composition in elderly men. J Clin Endocrinol Metab. 2000;85:3276-3282.
- 421. Wu FC, Tajar A, Beynon JM, et al. Identification of lateonset hypogonadism in middle-aged and elderly men. N Engl J Med. 2010;363:123-135.
- 422. Tajar A, Huhtaniemi IT, O'Neill TW, et al. Characteristics of androgen deficiency in late-onset hypogonadism: results

from the European Male Aging Study (EMAS). *J Clin Endocrinol Metab*. 2012;97:1508–1516.

- 423. Mueller A, Kiesewetter F, Binder H, Beckmann MW, Dittrich R. Long-term administration of testosterone undecanoate every 3 months for testosterone supplementation in female-to-male transsexuals. *J Clin Endocrinol Metab*. 2007;92:3470–3475.
- 424. Meriggiola MC, Armillotta F, Costantino A, et al. Effects of testosterone undecanoate administered alone or in combination with letrozole or dutasteride in female to male transsexuals. *J Sex Med.* 2008;5:2442–2453.
- 425. Van Pottelbergh I, Lumbroso S, Goemaere S, Sultan C, Kaufman JM. Lack of influence of the androgen receptor gene CAG-repeat polymorphism on sex steroid status and bone metabolism in elderly men. *Clin Endocrinol (Oxf)*. 2001;55:659–666.
- 426. Szulc P, Kaufman JM, Delmas PD. Biochemical assessment of bone turnover and bone fragility in men. Osteoporos Int. 2007;18:1451–1461.
- 427. Meier C, Nguyen TV, Handelsman DJ, et al. Endogenous sex hormones and incident fracture risk in older men: the Dubbo Osteoporosis Epidemiology Study. *Arch Intern* Med. 2008;168:47–54.
- 428. Orwoll E, Lambert LC, Marshall LM, et al. Endogenous testosterone levels, physical performance, and fall risk in older men. *Arch Intern Med.* 2006;166:2124–2131.
- 429. **Drake MT, Murad MH, Mauck KF, et al.** Clinical review. Risk factors for low bone mass-related fractures in men: a systematic review and meta-analysis. *J Clin Endocrinol Metab.* 2012;97:1861–1870.
- 430. Hyde Z, Flicker L, Almeida OP, et al. Low free testosterone predicts frailty in older men: the Health in Men Study. *J Clin Endocrinol Metab*. 2010;95:3165–3172.
- 431. Barrett-Connor E, Goodman-Gruen D, Patay B. Endogenous sex hormones and cognitive function in older men. *J Clin Endocrinol Metab*. 1999;84:3681–3685.
- 432. Yaffe K, Lui LY, Zmuda J, Cauley J. Sex hormones and cognitive function in older men. J Am Geriatr Soc. 2002; 50:707–712.
- 433. **Yeap BB, Almeida OP, Hyde Z, et al.** Higher serum free testosterone is associated with better cognitive function in older men, while total testosterone is not. The Health In Men Study. *Clin Endocrinol (Oxf)*. 2008;68:404–412.
- 434. Bhasin S, Cunningham GR, Hayes FJ, et al. Testosterone therapy in men with androgen deficiency syndromes: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab*. 2010;95:2536–2559.
- 435. Tracz MJ, Sideras K, Boloña ER, et al. Testosterone use in men and its effects on bone health. A systematic review and meta-analysis of randomized placebo-controlled trials. *J Clin Endocrinol Metab*. 2006;91:2011–2016.
- 436. Basurto L, Zarate A, Gomez R, Vargas C, Saucedo R, Galván R. Effect of testosterone therapy on lumbar spine and hip mineral density in elderly men. *Aging Male*. 2008; 11:140–145.
- 437. Wang YJ, Zhan JK, Huang W, et al. Effects of low-dose testosterone undecanoate treatment on bone mineral density and bone turnover markers in elderly male osteoporosis with low serum testosterone. *Int J Endocrinol*. 2013; 2013:570413.
- 438. Idan A, Griffiths KA, Harwood DT, et al. Long-term ef-

- fects of dihydrotestosterone treatment on prostate growth in healthy, middle-aged men without prostate disease: a randomized, placebo-controlled trial. *Ann Intern Med.* 2010;153:621–632.
- 439. **Ruige JB, Ouwens DM, Kaufman JM.** Beneficial and adverse effects of testosterone on the cardiovascular system in men. *J Clin Endocrinol Metab.* 2013;98:4300–4310.
- 440. Basaria S, Coviello AD, Travison TG, et al. Adverse events associated with testosterone administration. *N Engl J Med*. 2010;363:109–122.
- 441. Basaria S, Davda MN, Travison TG, Ulloor J, Singh R, Bhasin S. Risk factors associated with cardiovascular events during testosterone administration in older men with mobility limitation. *J Gerontol A Biol Sci Med Sci.* 2013;68:153–160.
- 442. Venken K, Boonen S, Van Herck E, et al. Bone and muscle protective potential of the prostate-sparing synthetic androgen 7α-methyl-19-nortestosterone: evidence from the aged orchidectomized male rat model. *Bone*. 2005;36: 663–670.
- 443. Miner JN, Chang W, Chapman MS, et al. An orally active selective androgen receptor modulator is efficacious on bone, muscle, and sex function with reduced impact on prostate. *Endocrinology*. 2007;148:363–373.
- 444. McCoy SC, Yarrow JF, Conover CF, et al. 17β -Hydroxyestra-4,9,11-trien-3-one (Trenbolone) preserves bone mineral density in skeletally mature orchiectomized rats without prostate enlargement. *Bone*. 2012;51:667–673.
- 445. Beck DT, Yarrow JF, Beggs LA, et al. Influence of aromatase inhibition on the bone-protective effects of testosterone. *J Bone Miner Res.* 2014;29:2405-2413.
- 446. Dobs AS, Boccia RV, Croot CC, et al. Effects of enobosarm on muscle wasting and physical function in patients with cancer: a double-blind, randomised controlled phase 2 trial. *Lancet Oncol*. 2013;14:335–345.
- 447. Matsumoto AM, Tenover L, McClung M, et al. The long-term effect of specific type II 5α -reductase inhibition with finasteride on bone mineral density in men: results of a 4-year placebo controlled trial. *J Urol.* 2002;167:2105–2108.
- 448. Amory JK, Anawalt BD, Matsumoto AM, et al. The effect of 5α-reductase inhibition with dutasteride and finasteride on bone mineral density, serum lipoproteins, hemoglobin, prostate specific antigen and sexual function in healthy young men. *J Urol.* 2008;179:2333–2338.
- 449. Jacobsen SJ, Cheetham TC, Haque R, Shi JM, Loo RK. Association between 5-α reductase inhibition and risk of hip fracture. *JAMA*. 2008;300:1660–1664.
- 450. Amory JK, Watts NB, Easley KA, et al. Exogenous testosterone or testosterone with finasteride increases bone mineral density in older men with low serum testosterone. *J Clin Endocrinol Metab*. 2004;89:503–510.
- 451. Bjørnerem A, Emaus N, Berntsen GK, et al. Circulating sex steroids, sex hormone-binding globulin, and longitudinal changes in forearm bone mineral density in postmenopausal women and men: the Tromsø study. *Calcif Tissue Int.* 2007;81:65–72.
- 452. Eriksson AL, Lorentzon M, Mellström D, et al. SHBG gene promoter polymorphisms in men are associated with serum sex hormone-binding globulin, androgen and andro-

- gen metabolite levels, and hip bone mineral density. *J Clin Endocrinol Metab*. 2006;91:5029–5037.
- 453. Nyquist F, Gärdsell P, Sernbo I, Jeppsson JO, Johnell O. Assessment of sex hormones and bone mineral density in relation to occurrence of fracture in men: a prospective population-based study. *Bone*. 1998;22:147–151.
- 454. Mohr BA, Bhasin S, Kupelian V, Araujo AB, O'Donnell AB, McKinlay JB. Testosterone, sex hormone-binding globulin, and frailty in older men. *J Am Geriatr Soc.* 2007; 55:548–555.
- 455. de Ronde W, van der Schouw YT, Muller M, et al. Associations of sex-hormone-binding globulin (SHBG) with non-SHBG-bound levels of testosterone and estradiol in

- independently living men. J Clin Endocrinol Metab. 2005; 90:157–162.
- 456. Khosla S. Editorial: sex hormone binding globulin: inhibitor or facilitator (or both) of sex steroid action? *J Clin Endocrinol Metab*. 2006;91:4764–4766.
- 457. Orwoll E, Lapidus JA, Wang Y, et al. Lack of value of serum sex steroid measures in the prediction of osteoporosis and fracture risk in community-dwelling, ambulatory older men. *J Bone Miner Res.* 2012;27:S66.
- 458. Orwoll E, Teglbjaerg CS, Langdahl BL, et al. A randomized, placebo-controlled study of the effects of denosumab for the treatment of men with low bone mineral density. *J Clin Endocrinol Metab.* 2012;97:3161–3169.