Volume 17, Number 1, April 2016

Pages: 270-274

ISSN: 1412-033X E-ISSN: 2085-4722 DOI: 10.13057/biodiv/d170138

Short Communication: Genetic identification of local pigs, and imported pigs (Landrace and Duroc) based on cytochrome b sequence analysis

TETY HARTATIK^{1,Ā}, BAYU DEWANTORO PUTRA SOEWANDI², SLAMET DIAH VOLKANDARI³, ARNOLD CHRISTIAN TABUN⁴, SUMADI¹, WIDODO¹

¹Faculty of Animal Science, Gadjah Mada University, Yogyakarta 55281, Indonesia.. Tel.: +62-274-4333373; fax: +62-274-521578. ▼email: tetyharuta@yahoo.com

²Indonesian Research Institute for Animal Production. PO Box 221, Bogor 16002, West Java, Indonesia

³Research Center for Biotechnology, Indonesian Institute of Sciences (LIPI). Jl Raya Bogor Km 46, Cibinong, Bogor 16911, West Java, Indonesia

⁴Kupang State Agriculture Polytechnic. Jl. Adisucipto Penfui, Kupang 1152, East Nusa Tenggara, Indonesia

Manuscript received: 4 March 2016. Revision accepted: 4 April 2016.

Abstract. Hartatik T, Soewandi BDP, Volkandari SD, Tabun AC, Sumadi, Widodo. 2016. Genetic identification of local pigs, and imported pigs (Landrace and Duroc) based on cytochrome b sequence analysis. Biodiversitas 17: 270-274. The aim of this study was to identify the genetics of local pigs and imported pigs (Landrace and Duroc) based on qualitative analysis. Thirty-eight pigs were used in this study and consisted of 11 local pigs (from Bali), six Landrace pigs and four Duroc pigs (from Malang), nine Landrace pigs (from Bali) and eight local pigs (from Kupang). Qualitative traits in pigs such as coat color, body shape (back shape, belly shape, and ears) and hair cover were observed. The cytochrome b (Cyt b) gene of mitochondrial DNA was analyzed using PCR-restriction fragment length polymorphism. The PCR analysis resulted in a 464 base pairs (bp) amplified band, and this was digested using TaqI restriction enzyme. The PCR-RFLP analysis resulted in two bands, 246 and 218 bp (monomorphic). The alignment analysis showed four points of single nucleotide polymorphisms. Bali and Kupang pigs had a specific pattern on exterior characteristics such as curved back shape, belly hanging shape, small ears and thick hair, and had many variations on coat color such as black, cream, spotted and mottled. The differences in coat color and body shape, and the corresponding mtDNA Cyt b sequence (with four SNPs) is a marker for genetic variation in pigs.

Keywords: Coat color, mtDNA, cytochrome b, qualitative analysis, genetic variation

INTRODUCTION

Indonesia has a great variety of domestic animals such as native chickens, ducks, goats, sheep, cattle and pigs. There are five species of local pig in Indonesia, Sus barbatus (Babi Berjanggut), Sus celebenis (Babi Sulawesi Berkutil), Sus verrucous (Babi Jawa Berkutil), Sus scorfa (Babi Alang-Alang) and *Babyroussa babyrussa* (Babirusa) (Rothschild et al. 2011). Genetic variations of local pigs in Indonesia are important to investigate. Since these pigs have important roles for cultural activities in the society, it is necessary to maintain their sustainability. The local Kupang pigs were used for traditional ceremonies such as weddings and religious ceremonies (Johns et al. 2010). For Bali cultural activities, pigs were used as the oblation for the religious activities. The oblation pig was a young intact boar up to eight or nine months old which had been fattened for five to six months (Soewandi 2013). The study of genetic diversity based on mitochondrial DNA (mtDNA) as a maternal line has been a reportedly useful tool as a molecular marker. Mitochondrial DNA is maternally inherited and experiences nucleotide changes faster than DNA (Brown et al. 1979), which makes mtDNA an ideal tool for studying population genetics (Bailey et al. 2000). Mitochondrial DNA is a useful genetic marker for both intra- and interspecies studies (Brown et al. 1979; Kikkawa et al. 1995).

Continental of wild boars and domestic pigs were clearly divided into eastern and western clades (Larson et al. 2005; Wu et al. 2007; Leutkemeier et al. 2010) using dloop mitochondrial DNA. A previous study based on Single Nucleotide Polymorphism (SNP) revealed that population of wild boars from Near Eastern Asia (Turkey, Iran and Armenia) and Europe (Spain, Belgium and Russia) are genetically different (Manunza et al. 2013). Asian pig populations were comprised of three groups. One group is represented by Erhualian and Meishan breed, while the second represented by Lanyu pigs and the third represented by the Asian wild boars. The Asian domestic populations were derived from multiple Asian ancestral origins whereas the European domestic populations represent a single ancestral European lineage (Leutkemeier et al. 2010).

Most genetic studies on wild boars in East Asia were carried out using mtDNA sequence analysis, which revealed several subclades (Larson et al. 2005; Hongo et al. 2002; Cho et al. 2009; Ramayo et al. 2010; Ji et al. 2011; Larson et al. 2010). Previous studies based on both mtDNA and nuclear genes demonstrated no population substructures exists in neither wild boars nor domestic pigs

in East Asia and showed a very high level of admixture between them (Ji et al. 2011). Korean wild boars were clearly clustered within Asian wild boar groups, sharing the same cluster with populations from Myanmar and Thailand (Cho et al. 2009) and the Vietnamese wild pig haplotype (Hongo et al. 2002). On the other hand, Larson et al. (2010) ascertained that wild boars in South Korea belong to groups unique within East Asia, and remain differentiated from domestic pigs.

Based on the mitochondrial DNA, the Vietnamese wild boars were clustered into two groups, group I was genetically distinct to Asian wild boars and group II that was genetically close to Asian wild boars. There are three types of haplotype in Vietnamese domestic pigs and two types of haplotypes in Vietnamese wild boars in Central Highland (Long et al. 2014). Chinese pig breeds were originally from the wild boars in the South China and the Yangtze River Region (Yu et al. 2013). The others studies showed that Chinese native pig breeds had a single origin (Lan and Shin 1993; Huang et al. 1999).

Study on the cytochrome b gene of indigenous pigs with PCR-SSCP methods had previously identified four SNPs located at positions 47 (T/C),49 (G/A), 52 (C/T) and 56 (G/A) and revealed the Asian origin of the Indigenous pigs (Ghungroo, Meghalaya local and Nagaland local) with A1 haplotype which revealed absence of mixed haplotype (Saikia et al. 2015). Other study in India, based on the analysis of sequence generated from the partial fragment (421 bp) mtDNA cytochrome b (*Cyt* b) gene exhibited unambiguous (>3%) genetic variation between Indian wild and domestic pigs. They observed nine forensically informative nucleotide sequence (FINS) variations between Indian wild and domestic pigs (Gupta et al. 2013).

This study was conducted to determine the genetic variation of local pigs (Bali and Kupang) compared with imported pigs (Landrace and Duroc) based on coat color, body shape and mtDNA *Cyt* b by using PCR-RFLP and sequence analysis.

MATERIALS AND METHODS

Samples and DNA extraction

Thirty-eight pigs were studied, consisting of 11 local pigs (Bali), eight local pigs (Kupang, NTT), 15 Landrace pigs and four Duroc pigs. Exterior characteristics of the pigs such as coat color, body shape (back shape, belly shape, and ears) and hair cover were observed. Blood and ear tissue samples were collected for DNA analysis. Blood samples were taken through jugular venipuncture and preserved in K₃EDTA solution tubes. Samples were stored frozen (-20 °C) until needed. The DNA from blood or ear tissue samples was extracted by using the standard SDS/Proteinase K modified method according to Sambrook et al. (1989).

Polymerase Chain Reaction-Restriction Fragment Length Polymorphism (PCR-RFLP)

The 464 base pairs (bp) fragment of the mtDNA *Cyt* b gene was amplified by polymerase chain reaction (PCR)

using forward and reverse primers according to Wolf et al. (1999): L14735 (5'-AAA AAC CAC CGT TGT TAT TCA ACTA-3') and H15149 (5'-GCC CCT CAG AAT GAT ATT TGT CCT CA-3'). Polymerase chain reaction was performed with a final volume of 20 µL of reaction mixture containing 1 µL of DNA sample (10-100 ng), 1 µL of each primer (10 pmol/ µL), 10 µL PCR KIT (Kappa, Biosystem), and 7 µL of double distilled water. The amplification process was performed using Thermocycler (Infinigen, TC-25/H) with the following conditions: initial denaturation at 94°C for 2 min, followed by 35 thermal cycles of denaturation at 95°C for 36 sec, annealing at 51°C for 73 sec, extension at 72°C for 84 sec and the final extension at 72°C for 3 min (Prado et al. 2005). The PCR product was visualized on 1% agarose gels buffered with 1X Tris-Boric-EDTA buffer (1XTBE), stained with ethidium bromide and visualised under ultraviolet (UV) light. The PCR-amplified DNA fragment of the Cyt b gene was digested using the TaqI restriction enzyme to identify genetic patterns. The total volume of digestion was 12 µL containing 3 µL PCR product, 0.2 µL TaqI (Fermentas) enzyme (1U), 1.2 µL Tango buffer and 7.6 µL aquabidest sterile. The enzymatic digestion was conducted at 65 °C for two hours by the TaqI enzyme. The digestion products were separated on 10% polyacrylamid gels in 1XTBE buffer and run with 50 V for three hours for separation of the DNA fragments. The bands were stained with ethidium bromide before visualization under UV light. The size of the amplified bands was compared with DNA marker X174 DNA/BsuRI (HaeIII) (Fermentas).

Sequencing and analysis

A total volume of 30 μ L for each PCR product and 10 μ L Cyt b primer (10 pmol/ μ L) was prepared for sequencing. Sequencing the amplified bands of PCR products was performed by BioSM Macrogen (Korea). The DNA sequences were aligned by using BioEdit version 7.7 for identification of the single nucleotide polymorphism.

RESULTS AND DISCUSSION

The exterior characteristics of the pigs were determined based on expert judgment and included coat color and body shape. Landrace pigs had 100% white color while Duroc pigs had 100% reddish brown color. Bali and Kupang pigs had a huge diversity of color: 63.6% of Bali pigs had black coat color and 36.4% had mottled color, whereas 37.5% Kupang pigs had reddish brown color, 25% had cream color and 12.5% had spotted (white and black) color. Local pigs had specific characteristics of body shape at back and belly whereas imported pigs (Landrace and Duroc) had a straight body shape between back and belly. In addition, Bali and Kupang pigs had small, upright ears and thick hair. This is different from the Landrace and Duroc pigs that had sparse hair and big ears, with 50% having ears folded forward (Figure 1).

The specific DNA fragment of the mtDNA *Cyt* b gene in local pigs, Landrace pigs and Duroc pigs was amplified by using L14735 and H15149 primers. The PCR product of

the mtDNA *Cyt* b gene was 464 bp (Figure 2a). The product size of PCR-RFLP using the *Taq*I enzyme showed the same restriction pattern. There were two fragments of DNA, 246 bp and 218 bp (Figure 2b), which indicates that the sample population was monomorphic. In total of 38 samples, one haplotype of cytochrome b sequence was observed based on PCR-RFLP.

The sequence analysis of the mtDNA *Cyt* b gene of local pigs (Bali), Landrace pigs and Duroc pigs was compared to the complete mtDNA of *Cyt* b gene database available at GenBank (DQ.534707.2/Sus scrofa breed Taoyuan; NC. 014692.1/Sus scrofa taiwanensis; NC.012095.1/Sus scrofa domesticus; and GQ. 338965.1/Sus scrofa). Based on mtDNA *Cyt* b sequence alignment analysis (Figure 3), we found four points of single nucleotide polymorphism (SNP) which changed the nucleotides from C to T and G to A. However the local, Landrace and Duroc pigs had a similar sequence of mtDNA *Cyt* b in this study.

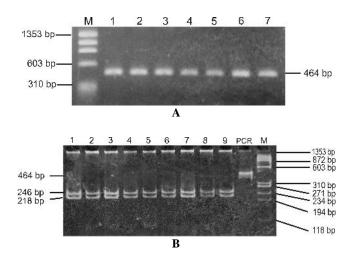


Figure 2. PCR product and RFLP with *Taq*I enzyme. A. PCR product of cyt b gene (464 bp); B. PCR-RFLP product using the *Taq*I enzyme: lanes 1-2: Duroc, lanes 3-5: Landrace, lanes 6-9: Bali pig, PCR: product, M: Marker X174 DNA/*Bsu*RI (*Hae*III) (Fermentas).



Figure 1. Coat color and body shape of pigs. A. Bali pig, B. Kupang pig, C. Duroc pig and D. Landrace pig

DQ534707.2 NC_014692.1 LANDRACE (BALI) DUROC (MALANG) BALI (LOCAL) GQ338965.1 NC_012095.1	TACACATCAGACACAACAACAGCTTTCTCATCAGTTACACACATCTGTCG TACACATCAGACACAACAACAGCTTTCTCATCAGTTACACACATCTGTCG TACACATCAGACACAACAACAGCTTTCTCATCAGTTACACACATCTGTCG TACACATCAGACACAACAACAGCTTTCTCATCAGTTACACACAC	150 150 150 150 150 150
DQ534707.2 NC_014692.1 LANDRACE (BALI) DUROC (MALANG) BALI (LOCAL) GQ338965.1 NC_012095.1	AGACGTAAATTACGGATGAGTTATTCGCTACCTACATGCAAACGGAGCAT AGACGTAAATTACGGATGAGTTATTCGCTACCTACATGCAAACGGAGCAT AGACGTAAATTACGGATGAGTTATTCGCTACCTACATGCAAACGGAGCAT AGACGTAAATTACGGATGAGTTATTCGCTACCTACATGCAAACGGAGCAT AGACGTAAATTACGGATGAGTTATTCGCTACCTACATGCAAACGGAGCAT AGACGTAAATTACGGATGAGTTATTCGCTATCTACATGCAAACGGAGCAT AGACGTAAATTACGGATGAGTTATTCGCTATCTACATGCAAACGGAGCAT ************************************	200 200 200 200 200 200 200
DQ534707.2 NC_014692.1 LANDRACE (BALI) DUROC (MALANG) BALI (LOCAL) GQ338965.1 NC_012095.1	CCATGTTCTTTATTTGCCTATTCATCCACGTAGGCCGAGGCCTATACTAC CCATGTTCTTTATTTGCCTATTCATCCACGTAGGCCGAGGCCTATACTAC CCATGTTCTTTATTTGCCTATTCATCCACGTAGGCCGAGGCCTATACTAC CCATGTTCTTTATTTGCCTATTCATCCACGTAGGCCGAGGCCTATACTAC CCATGTTCTTTATTTGCCTATTCATCCACGTAGGCCGAGGCCTATACTAC CCATGTTCTTTATTTGCCTATTCATCCACGTAGGCCGAGGTCTATACTAC CCATGTTCTTTATTTGCCTATTCATCCACGTAGGCCGAGGTCTATACTAC CCATGTTCTTTATTTGCCTATTCATCCACGTAGGCCGAGGTCTATACTAC **** ********************************	250 250 250 250 250 250 250

Figure 3. Multiple sequence alignment of partial mtDNA cyt b in pigs

The sequenced DNA data showed that Bali pig ancestors were from China, which possibly were similar to the *Sus scrofa taiwanensis* ancestor. In four positions of SNPs, the same nucleotide between Bali pig and DQ.534707.2 (Breed Taoyuan) and NC. 014692.1 (Breed Taiwanesis) was observed (Figure 3). A previous study by Sihombing (1997) revealed that the Bali pig ancestor came from a cross between a second type of *Sus scorfa vittatus* and the South China pig. This finding supports our data that Bali pigs have a genetic similarity to *Sus scrofa taiwanensis* and the Taoyuan pig. The mitochondria of the pigs was divided into two types (Asian type and Europe type), Taoyuan pig included to Asian type (Chang et al. 2008).

A previous study by Clop et al. (2004) proved that Landrace and Duroc breeds had an Asian allele. It was likely due to an introduction process of Landrace and Duroc pigs with the Asian allele. Evidence showed that there was an introduction of Asian pig breeds to commercial pig breeds (Clop et al. 2004; Giuffra et al. 2000). The introduction processes did not result in any differences in their performances, rather their mtDNA showed high genetic similarity. Mitochondrial DNA has been widely used to unravel evolutionary studies, due to its greater diversity compared to nuclear DNA.

To conclude, local pigs, Landrace and Duroc pigs have the same sequence of cytochrome b gene. The sequence analysis of seven breed of pigs shows four position of single nucleotide polymorphism.

ACKNOWLEDGEMENTS

This research was supported partly by a Graduate School Grant from the Faculty of Animal Science, Gadjah Mada University, Yogyakarta, Indonesia. The author expresses sincere gratitude to all of the farmers in the Fatumonas Village, Central Amfoang Sub-district, Kupang District, East Nusa Tenggara, Indonesia who provided their livestock for samples for this study. This research was also supported by The Livestock Department of Tabanan District and The Veterinary Bureau of Denpasar, Bali, Indonesia. We thank Drh. Ni Luh Putu Agustini, I Ketut Mayun, Dewa, Drh. I Ketut Mertanadi and Drh. Cok Ngurah Dharyatno for their contributions in collecting the sample resources.

REFERENCES

- Bailey JF, Healy B, Jianlin H, Sherchand L, Pradhan SL, Tsendsuren T, Foggin, JM, Gaillard C, Steane D, Zakharov I, Bradley DG. 2002. Genetic variation of mitochondrial DNA within domestic yak populations. In: Jianlin H, Richards C, Hanotte O, McVeigh C, Rege JEO (eds.). Yak production in Central Asian highlands; Proceedings of the Third International Congress on Yak. Lhasa, PR China, 4-9 September 2000.
- Brown WM, George MJr, Wilson AC. 1979. Rapid evolution of animal mitochondrial DNA. Proc Natl Acad Sci USA 76 (4): 1967-1971.
- Chang WH, Chu HP, Jiang YN, Li SH, Wang Y, Chen CH, Chen KJ, Lin CY, Ju YT. 2008. Genetic variation and phylogenetics of Lanyu and

- exotic pig breeds in Taiwan analyzed by nineteen microsatellite markers. J Anim Sci 1-8.
- Cho IC, Han SH, Fang M, Lee SS, Ko MS, Lee H, Lim HT, Yoo CK, Lee JH, Jeon JT. 2009. The robust phylogeny of Korean wild boar (*Sus scrofa coreanus*) using partial D-loop sequence of mtDNA. Mol Cells 28: 423-430.
- Clop A, Amills M, Noguera JL, Fernandez A, Capote J, Ramon MM, Kelly L, Kijas JMH, Andersson L, Sanchez A. 2004. Estimating the frequency of Asian cytochrome B haplotypes in standard European and local Spanish pig breeds. Genet Selec Evol 36: 97-104.
- Giuffra E, Kijas JMH, Amargoer V, Carlborg O, Jeon JT, Andersson L. 2000. The origin of the domestic pig: independent domestication and subsequent introgression. Genetics 154 (4): 1785–1791.
- Gupta, S. K., A.Kumar, S.A. Hussain, Vipin, L. Singh. 2013. Cytochrome b based genetic differentiation of Indian wild pig (*Sus scrofa cristatus*) and domestic pig (*Sus scrofa domestica*) and its use in wildlife forensics. Sci Justice 53: 220-222
- Hongo H, Ishiguro N, Watanobe T, Shigehara N, Anezaki T, Long VT, Binh DV, Tien NT, Nam NH. 2002. Variation in mitochondrial DNA of Vietnamese pigs: relationships with Asian domestic pigs and Ryukyu wild boars. Zoo Sci 19: 1329-1335.
- Huang YF, Shi XW, Zhang YP. 1999. Mitochondrial genetic variation in Chinese pigs and wild boars. Biochem Genet 37: 335-343.
- Ji Y-Q, Wu D-D, Wu G-S, Wang G-D, Zhang Y-P. 2011. Multi-locus analysis reveals a different pattern of genetic diversity for mitochondrial and nuclear DNA between wild and domestic pigs in East Asia. PLoS ONE 6: e26416.
- Johns C, Cargill C, Patrick I, Geong M, Ly J. 2010. Commercial pig farming by small farmers in Nusa Tenggara Timur Indonesia. [Unpublished Report]. SADI-ACIAR, Canberra. [Indonesian]
- Lan H, Shi L. 1993. The origin and genetic differentiation of native breeds of pigs in Southwest China: an approach from mitochondrial DNA polymorphism. Biochem Genet 31: 51-60.
- Larson G, Dobney K, Albarella U, Fang M, Matisoo-Smith E, Robins J, Lowden S, Finlayson H, Brand T, Willerslev E, Rowley-Conwy P, Andersson L, Cooper A. 2005. Worldwide phylogeography of wild boar reveals multiple centers of pig domestication. Science 307: 1618-1621.
- Larson G, Liu R, Zhao X, Yuan J, Fuller D, Barton L, Dobney K, Fan Q, Gu Z,Liu X-H, Luo Y, Lv P, Andersson L, Li N. 2010. Patterns of East Asian pig domestication, migration, and turnover revealed by modern and ancient DNA. Proc Natl Acad Sci USA 107: 7686-7691.
- Leutkemeier ER, Sodhi M, Schook LB, Malhi RS. 2010. Multiple Asian pig origins revealed through genomic analyses. Mol Phylogen Evol 54: 680-686.
- Long LT, Mai NTP, Chung DC, Son HN, Si DM. 2014. The genetic relationship of Vietnamese pigs in central highlands assessed by cytochrome b. Open J Genet 4: 362-369.
- Kikkawa Y, Amano T, Suzuki H. 1995. Analysis of genetic diversity of domestic cattle in east and Southeast Asia in terms of variations in restriction sites and sequences of mitochondrial DNA. Biochem Genet 33: 51-60.
- Manunza A, Zidi A, Yeghoyan S, Balteanu VA, Carsai TC, Scherbakov O, Ramirez O, Castello A, Amills M, Mercade A. 2013. A high throughput genotyping approach reveals distinctive autosomal genetic signatures for European and Near eastern wild boar. Plos One 8 (2): e55891 doi: 10.1371/journal.pone.0055891.
- Prado M, Calo P, Cepeda A, Barros-Velázquez J. 2005. Genetic evidence of an Asian background in heteroplasmic Iberian cattle (*Bos taurus*): effect on food authentication studies based on polymerase chain reaction-restriction fragment length polymorphism analysis. Electrophoresis 26: 2918-2926.
- Ramayo Y, Shemeret'eva IN, Pérez-Enciso M. 2010. Mitochondrial DNA diversity in wild boar from the Primorsky Krai Region (East Russia). Anim Genet 42: 96-99.
- Rothschild MF, Ruvinsky A. 2001. The Genetics of the Pig. CAB International, London.
- Saikia J, Roy TC, Naskarl S, Das B, Ferdoci AM. 2015. Molecular characterization of Cytochrome B gene in indigenous pig. Indian J Anim Res 49: 196-198.
- Sambrook J, Fritsch EF, Maniatis T. 1989. Molecular Cloning, A Laboratory Manual. Cold Spring Harbour Laboratory Press, New York
- Sihombing DTH. 1997. Pig Animal Science. UGM Press, Yogyakarta [Indonesian].

- Soewandi BDP. 2013. Output estimation and identification growth hormone gene in pigs on Tabanan Regency Bali Province. [Thesis]. Gadjah Mada University, Yogyakarta. [Indonesian]
 Wolf C, Rentsch J, Hubner P. 1999. PCR-RFLP analysis of mitochondrial
- Wolf C, Rentsch J, Hubner P. 1999. PCR-RFLP analysis of mitochondrial DNA: a reliable method for species identification. J Agric Food Chem 47: 1350-1355.
- Wu GS, Yao YG, Qu KX, Ding ZL, Palanichamy MG, Duan ZY, Li N, Chen YS, Zhang YP, Li H. 2007. Population phylogenomic analysis of mitochondrial DNA in wild boars and domestic pigs revealed multiple domestication events in East Asia. Gen Biol 8: R245.
- Yu G, Xiang H, Wang J, Zhao X. 2013. The pylogenetic status of typical Chinese native pigs: analyzed by Asian and European pig mitochondrial genome sequences. J Anim Sci Biotechnol 4: 9.