

Research and Reviews: Research Journal of Biology

Short review on Production of Biodiesel from Animal Tallow via Enzymatic Transesterification

Uzwal Prakash*

Biju Patnayak University of technology, Odissa

Short Review

Received: 27/02/2015

Revised : 03/03/2015

Accepted:10/03/2015

*For Correspondence

Uzwal Prakash, Biju Patnayak
University of technology, Odissa,
Tel. +91-9912961554; E-mail:
uzwal4333@gmail.com

Keywords: Biodiesel, Enzyme,
Transesterification

ABSTRACT

In this article authors explained about effectiveness of enzymatic transesterification of animal fat using the experimental enzyme catalyst NS88001 [1,2]. He evaluated biodiesel yield with the effect of Oil: Alcohol, reaction temperature and reaction time. They worked on the different Oil: Alcohol ratio and observed their yield effect on biodiesel production [3,4]. Reaction performed in this procedure slowly at the starting stage and then increased rapidly due to the initial mixing and dispersion of alcohol in to oil substrate and effectively activation of catalytic enzyme NS88001 [5-25]. After alcohol dispersion, rapidly interaction of fatty acid with enzyme takes place resulted maximum yield conversion [1,26-30]. 114.95 – 65.59 % yield conversion were observed when they increased the reaction time from 4Hrs to 16Hrs at 45 °C [1].

Review

In the high demand of fossil fuel we cannot depend on the limited resource we need to think about some other alternatives of renewable energy and biodiesel is one of them which can be used as a current resource of energy and fulfill the demand up to some extent. We can use biodiesel for compression ignition engine instead of diesel. We know that the use of biodiesel is sulfur free, non-toxic and biodegradable in nature so; these characteristics make it eco-friendly. There are several plants, animal fats, yellow grease, waste cooking and algae biomass which can be used as raw material for biodiesel production. The main component of fats and oils are triacylglycerols which are made up of different fatty acids and glycerol being the backbone. There is variation in all type of fatty acid due to their source component. Chemical process use to convert oils and fats in to biodiesel known as transesterification. Main objective of his work is to investigate the effectiveness of the enzymatic transesterification process for the production of biodiesel from animal tallow using methanol in a solvent free system [31-50].

Raw material, enzyme and chemicals

Animal tallow (10 Kgs; Stored at -20 °C)

Enzyme catalyst: NS88001

Chemicals: Methanol, Tetrahydrofuran, N,O-Bis (Trimethylsilyl)-trifluoroacetamide and hilditch reagent.

Followed: Purification of crude animal tallow, (110 ° C, 50 rpm, 1 Hrs.)

They performed an enzymatic trans-esterification at different oil: alcohol molecular ratio, time and temperature and analyze product ratio with the help of gas chromatography. He explained different analysis result at different oil: alcohol ratio, reaction time and temperature and analyzed their result. According to his result if you would consider yield you found that in oil: alcohol ratio 1:4 molar ratio is more effective, it resulted (80.42%) mean yield; In reaction time we found 16 Hrs is more productive in their comparison it reflected 49% mean yield. But in reaction temperature 45 °C is more productive [1].

DISCUSSION

Fatty acid [51-64] used in this experiment contains oleic acid (44%), palmitic acid (28%) and stearic acid (26%) as well as lower percentages of myristic acid (1%) and linoleic acid (1%). High content of Oleic acid improves quality of biodiesel.

The benefit of animal tallow oil is to increase methyl ester oxidation which might increase the cetane number which tends to delay the ignition time in the engine.

They considered 1:4 oil : alcohol molar ratio is standard because they found above 1:4 ration yield was going to decrease from 38.74, 25.41 and 44.10% at the reaction temperatures of 40, 45 and 50 °C and the similar type of changes observed in reaction time.

REFERENCES

1. Kumar S, Ghaly AE, Brooks MS, Budge SM, Dave D (2013) Effectiveness of Enzymatic Transesterification of Beeftallow Using Experimental Enzyme Ns88001 with Methanol and Hexane. *Enz Eng* 2:116. doi: 10.4172/2329-6674.1000116
2. Vinod Kumar, Firdaus Jahan and R.K. Saxena (2012) Biocatalysts towards ever greener for production of biodiesel from non-edible oil and comparison of fuel characteristics. 3rd World Congress on Biotechnology, OMICS Group, Hyd.
3. Soumen Ghosal (2014) Effect of shoot age, liming and potassium application for summer season lac cultivation on Palas (*Butea monosperma*) trees. 2nd Int Conf Horticulture Sci, OMICS, Hyd.
4. Raymond Le Van Mao, Abdulhafed Muntasar and HaiTao Yan (2013) Conversion of ethers of light alcohols into hydrocarbons over ZSM-5 zeolite catalysts. World Cong Petrochem Chem Engg, USA.
5. R. K. Pathak, Nitin.e.pereira (2013) Various Sources for Production Of Biodisel. *Poll Cont.*
6. López T, Moreno A, Ortiz-Islas E, Pecchi G, Bersani D, et al. (2015) Inclusion of FeCl₃ in Sol-Gel TiO₂: Spectroscopic Studies. *J Nanomed Nanotechnol* 6:255. doi: 10.4172/2157-7439.1000255.
7. Shintani H (2014) Effect of Humidity, OH Radicals, UV Irradiation, Adhesive and Adsorption Effect on Photocatalyst Sterilization. *Chem Sci J* 5:e104. doi: 10.4172/2150-3494.1000e104.
8. Lim HK, Park NJ, Hwang YK, Lee KI, Hwang IT (2015) Improvement and Immobilization of a new Endo-β-1,4-xylanases KRICT PX1 from *Paenibacillus* sp. HPL-001. *J Bioprocess Biotech* 5:215. doi: 10.4172/2155-9821.1000215
9. Obregón WD, Cisneros JS, Ceccacci F Quiroga E (2015) A Highly Stable Biocatalyst Obtained from Covalent Immobilization of a Non-Commercial Cysteine Phytprotease. *J Bioprocess Biotech* 5:211. doi: 10.4172/2155-9821.1000211
10. Panda N, Jena AK (2015) Cu/Fe-Catalyzed Carbon-Carbon and Carbon-Heteroatom Cross-Coupling Reactions. *Organic Chem Curr Res* 4:130. doi: 10.4172/2161-0401.1000130
11. Ahmad K, Ali M, Ibrahim A, Baig WM (2014) Optimising the Yield of Silicon Carbide Synthesised from Indigenous Biomass Husk using Different Catalysts. *J Material Sci Eng* 3: 147. doi: 10.4172/2169-0022.1000147
12. Gawande MB (2014) Sustainable Nanocatalysts for Organic Synthetic Transformations. *Organic Chem Curr Res* 1:e137. doi: 10.4172/2161-0401.1000e137
13. Sandhya S, Rayalu S, Bruno B (2014) Solar Light Induced Photo Catalytic Disinfection of Gram Positive and Negative Microorganisms from Water with Highly Efficient AuTiO₂ Nanoparticle. *J Bioprocess Biotech* 4:176. doi: 10.4172/2155-9821.1000176
14. Pandey PC, Prakash A, Pandey AK, Pandey D (2014) 3-Aminopropyltrimethoxysilane and 3-Glycidoxypropyltrimethoxysilane Mediated Synthesis of Graphene and its Nanocomposite: Potential Bioanalytical Applications. *J Anal Bioanal Tech* S7:012. doi: 10.4172/2155-9872.S7-012
15. Cendrowski K, Peruzynska M, Markowska-Szczupak A, Chen X, Wajda A, et al. (2013) Mesoporous Silica Nanospheres Functionalized by TiO₂ as a Photoactive Antibacterial Agent. *J Nanomed Nanotechnol* 4:182. doi: 10.4172/2157-7439.1000182

16. Zacconi F, Arias HR (2013) Biocatalysts. *J Thermodyn Catal* 4:e121. doi: 2157-7544.1000e121
17. Bindig R, Butt S, Hartmann I (2013) Emission Abatement at Small-Scale Biomass Combustion Unit with High-Temperature Catalysts. *J Thermodyn Catal* 4:125. doi: 10.4172/2157-7544.1000125
18. Guo Y, Zu B, Dou X (2013) Zeolite-based Photocatalysts: A Promising Strategy for Efficient Photocatalysis. *J Thermodyn Catal* 4:e120. doi: 10.4172/2157-7544.1000e120
19. Babu ML, Sarma PN, Mohan SV (2013) Microbial Electrolysis of Synthetic Acids for Biohydrogen Production: Influence of Biocatalyst Pretreatment and Ph with the Function of Applied Potential. *J Microb Biochem Technol* S6:003. doi: 10.4172/1948-5948.S6-003
20. Zakaria S, Liew TK, Chia CH, Pua FL, Pin FS, et al. (2013) Characterization of Fe₂O₃/FeOOH Catalyzed Solvolytic Liquefaction of Oil Palm Empty Fruit Bunch (EFB) Products. *J Bioremed Biodeg* S4:001. doi: 10.4172/2155-6199.S4-001
21. Moinuddin Sarker, Mohammad Mamunor Rashid, and Mohammed Molla (2011) Abundant High-Density Polyethylene (HDPE-2) Turns into Fuel by Using of HZSM-5 Catalyst. *J fund renew energy appl* 1: 12.
22. Jinyi Deng, Subarna Banerjee, Susanta K. Mohapatra, York R. Smith¹ and Mano Misra (2011) Bismuth Iron Oxide Nanoparticles as Photocatalyst for Solar Hydrogen Generation from Water. *J fund renew energy appl* 1: 10.
23. Shohaimi NAM, Abu Bakar WAW, Jaafar J, Shukri NM (2013) Treatment of Acidic Petroleum Crude Oil Utilizing Catalytic Neutralization Technique of Magnesium Oxide Catalyst. *Mod Chem appl* 1:103. doi: 10.4172/2329-6798.1000103
24. Soloducho J, Cabaj J (2013) Electrochemical Nanosized Biosensors: Perspectives and Future of Biocatalysts. *J Anal Bioanal Techniques* S7:005. doi: 10.4172/2155-9872.S7-005
25. Anamika S, Pant KK (2013) Oxidative Steam Reforming of Bioethanol over Rh/CeO₂-Al₂O₃ Catalyst for Hydrogen Production. *J Thermodyn Catal* 4:119. doi: 10.4172/2157-7544.1000119
26. Bakou SN, Ella GSN, Aoussi S, Guiguand L, Cherel Y, et al. (2015) Fiber Composition of the Grasscutter (*Thryonomys swinderianus*, Temminck 1827) Thigh Muscle: An Enzyme-histochemical Study. *J Cytol Histol* 6:311. doi: 10.4172/2157-7099.1000311
27. Yilmaz İ, Doğan Z, Erdemli E, Gürsoy S, Bağ HG (2015) The Effects of Apricot on Large Intestine Oxidative Stress Enzymes in Rats. *J Cytol Histol* 6:306. doi: 10.4172/2157-7099.1000306
28. Ajayi AA, Peter-Albert CF, Adedeji OM (2015) Modification of Cell Wall Degrading Enzymes from Soursop (*Annona muricata*) Fruit Deterioration for Improved Commercial Development of Clarified Soursop Juice (A Review). *Med Aromat Plants* 4:178. doi: 10.4172/2167-0412.1000178
29. Obregón WD, Cisneros JS, Ceccacci F Quiroga E (2015) A Highly Stable Biocatalyst Obtained from Covalent Immobilization of a Non-Commercial Cysteine Phytolprotease. *J Bioprocess Biotech* 5:211. doi: 10.4172/2155-9821.1000211
30. Mamta Sahu, V.K.Gour And Swati Gupta (2008) JATROPHA OIL: AN ECO-FRIENDLY SUSTAINABLE BIO-FUEL SOURCE. *Jr. of Industrial Pollution Control* 24: 177-182.
31. Al hattab M, Ghaly A, Hammoud A (2015) Microalgae Harvesting Methods for Industrial Production of Biodiesel: Critical Review and Comparative Analysis. *J Fundam Renewable Energy Appl* 5:154. doi: 10.4172/2090-4541.1000154
32. Silva LS, Bezerra LR, Azevedo Silva AMD, Heloisa Carneiro, de Morais RKO, et al. (2015) Greenhouse Gases and Volatiles Fat Acids in vitro of Glycerin Generated in the Biodiesel Production Chain. *J Food Process Technol* 6:429. doi: 10.4172/2157-7110.1000429
33. Olalekan A (2015) Impact of Palm Kernel Oil (PKO) Biodiesel- Contaminated Catfish on Hepatic Function of Rat. *J Pollut Eff Cont* 3:127. doi: 10.4172/2375-4397.1000127
34. Kabbashi NA, Mohammed NI, Alam Md Z, Mirghani MES (2015) Effect of Process Parameters on Yield and Conversion of Jatropha Biodiesel in a Batch Reactor. *J Fundam Renewable Energy Appl* 5:155 doi: 10.4172/2090-4541.1000155

35. Olalekan A (2014) The Effect of Palm Kernel Oil (PKO) Biodiesel-Contaminated Soil on Morphological and Biochemical Properties of *Zea mays*. *J Plant Biochem Physiol* 2:138. doi: 10.4172/2329-9029.1000138
36. Liu M, Yusoff MM, Makky EA, Salihon J (2014) Bacterial Isolation from Palm Oil Plantation Soil for Biodiesel Production: Isolation and Molecular Identification as Inferred by 16s RNA. *J Biotechnol Biomater* 4:165. doi: 10.4172/2155-952X.1000165.
37. Gomez-Mares M, Martinez-Ortega ME, Arroyo-Ortega G, Reyes-Blas H, Hernandez-Paz J, et al. (2014) Comparative Study of the Effects of Diesel and Biodiesel Over POM, PPA and PPS Polymers Used in Automotive Industry. *J Material Sci Eng* 3: 142. doi: 10.4172/2169-0022.1000142
38. Alemán-Nava GS, Sandate-Flores L, Meneses-Jácome A, Dávila-Chavez R, Dallemand JF, et al. (2014) Bioenergy Sources and Representative Case Studies in Mexico. *J Pet Environ Biotechnol* 5:190. doi: 10.4172/2157-7463.1000190
39. Azad AK, Yousuf A, Ferdoush A, Mahub Hasan Md, Rezaul Karim Md, et al. (2014) Production of Microbial Lipids from Rice Straw Hydrolysates by *Lipomyces starkeyi* for Biodiesel Synthesis. *J Microb Biochem Technol* S8:008. doi: 10.4172/1948-5948.S8-008
40. Nguyen PLT, Go AW, Huynh LH, Ju YH (2014) In Situ Transesterification of Wet Activated Sludge under Subcritical Conditions. *J Pet Environ Biotechnol* 5:182. doi: 10.4172/2157-7463.1000182
41. Dave D, Ramakrishnan VV, Trenholm S, Manuel H, Pohling J, et al. (2014) Marine Oils as Potential Feedstock for Biodiesel Production: Physicochemical Characterization. *J Bioprocess Biotech* 4:168. doi: 10.4172/2155-9821.1000168
42. Dávila L, Brito A (2014) FFA Adsorption from Waste Oils or Non-Edible Oils onto an Anion-Exchange Resin as Alternative Method to Esterification Reaction Prior to Transesterification Reaction for Biodiesel Production. *J Adv Chem Eng* 4:105. doi: 10.4172/2090-4568.1000105
43. Abd El Baky HH, El-Baroty GS, Bouaid A (2014) Lipid Induction in *Dunaliella salina* Culture Aerated with Various Levels CO₂ and Its Biodiesel Production. *J Aquac Res Development* 5:223 doi: 10.4172/2155-9546.1000223
44. Kumar S, Ghaly AE, Brooks MS, Budge SM, Dave D (2013) Effectiveness of Enzymatic Transesterification of Beef tallow Using Experimental Enzyme Ns88001 with Methanol and Hexane. *Enz Eng* 2:116. doi: 10.4172/2329-6674.1000116
45. Martin MZ, Gunter LE, Jawdy SS, Wullschlegel SD, Wheeler CS, et al. (2013) Genetic Improvement, Sustainable Production and Scalable Small Microenterprise of *Jatropha* as a Biodiesel Feedstock. *J Bioremed Biodeg* S4:002. doi: 10.4172/2155-6199.S4-002
46. Ragauskas AME, Ragauskas AJ (2013) Re-defining the Future of FOG and Biodiesel. *J Phylogenetics Evol Biol* 4:e118. doi: 10.4172/2157-7463.1000e118
47. Mansourpoor M, Shariati A (2012) Optimization of Biodiesel Production from Sunflower Oil Using Response Surface Methodology. *J Chem Eng Process Technol* 3:141 doi: 10.4172/2157-7048.1000141
48. Li Q, Zheng L, Hou Y, Yang S, Yu Z (2011) Insect Fat, a Promising Resource for Biodiesel. *J Phylogenetics Evol Biol* S2:001. doi: 10.4172/2157-7463.S2-001
49. Owolabi RU, Osiyemi NA, Amosa MK, Ojewumi ME (2011) Biodiesel from Household/Restaurant Waste Cooking Oil (WCO). *J Chem Eng Process Technol* 2:112. doi: 10.4172/2157-7048.1000112
50. Narendrula R, Nkongolo KK (2015) Fatty Acids Profile of Microbial Populations in a Mining Reclaimed Region Contaminated with Metals: Relation with Ecological Characteristics and Soil Respiration. *J Bioremed Biodeg* 6: 274. doi: 10.4172/2155-6199.1000274
51. Venkateswar Reddy M, Venkata Mohan S (2015) Polyhydroxy alkanoyates Production by Newly Isolated Bacteria *Serratia ureilytica* Using Volatile Fatty Acids as Substrate: Bio-Electro Kinetic Analysis. *J Microb Biochem Technol* 7:026-032. doi: 10.4172/1948-5948.1000177

52. Cantani A (2014) Immunological Properties of Breast Milk: A Prospective Study in 589 Children. *Interdiscip J Microinflammation* 1:127. doi: 10.4172/ijm.1000127
53. Kahouli I, Malhotra M, Tomaro-Duchesneau C, Saha S, Marinescu D, et al. (2015) Screening and In-Vitro Analysis of *Lactobacillus reuteri* Strains for Short Chain Fatty Acids Production, Stability and Therapeutic Potentials in Colorectal Cancer. *J Bioequiv Availab* 7:039-050. doi: 10.4172/jbb.1000212
54. Deepika D, Vegneshwaran VR, Julia P, Sukhinder KC, Sheila T, et al. (2014) Investigation on Oil Extraction Methods and its Influence on Omega-3 Content from Cultured Salmon. *J Food Process Technol* 5:401. doi: 10.4172/2157-7110.1000401
55. Johnson M, Bradford C (2014) Omega-3, Omega-6 and Omega-9 Fatty Acids: Implications for Cardiovascular and Other Diseases. *J Glycomics Lipidomics* 4:123. doi: 10.4172/2153-0637.1000123
56. Tripathi MK (2014) Effect of Nutrition on Production, Composition, Fatty acids and Nutraceutical Properties of Milk. *J Adv Dairy Res* 2:115. doi: 10.4172/2329-888X.1000115
57. Shukla G, Verma A, Singh J, Yadav H (2014) Prebiotic Inulin Alters the Colonic Mass, pH, Microflora and Short Chain Fatty Acids in 1,2-Dimethylhydrazine Dihydrochloride Induced Early Colon Carcinogenesis in Male Laca Mice. *J Prob Health* 2:121. doi: 10.4172/2329-8901.1000121
58. Peighambardoust SH, Aghamirzaei M (2014) Physicochemical, Nutritional, Shelf Life and Sensory Properties of Iranian Sangak Bread Fortified with Grape Seed Powder. *J Food Process Technol* 5:381. doi: 10.4172/2157-7110.1000381
59. Grela ER, Pietrzak K (2014) Production Technology, Chemical Composition and Use of Alfalfa Protein-Xanthophyll Concentrate as Dietary Supplement. *J Food Process Technol* 5:373. doi: 10.4172/2157-7110.1000373
60. Mendes J (2014) Polyunsaturated fatty acids play a role in depression: main action mechanisms. *J Biosafety Health Educ* 2:e115. doi:10.4172/2332-0893.1000e115
61. D'Arner J, Carbonell P, Pino S, Farías A (2014) Variation of Fatty Acids in *Isochrysis galbana* (T-Iso) and *Tetraselmis suecica*, Cultured Under Different Nitrate Availabilities. *Fish Aquac J* 5:106. doi: 10.4172/2150-3508.1000106
62. Brunst KJ, Enlow MB, Kannan S, Carroll KN, Coull BA, et al. (2014) Effects of Prenatal Social Stress and Maternal Dietary Fatty Acid Ratio on Infant Temperament: Does Race Matter?. *Epidemiology (Sunnyvale)* 4:167. doi: 10.4172/2161-1165.1000167
63. Cocchi M, Tonello L, Gabrielli F, Minuto C (2014) Human and Animal Brain Phospholipids Fatty Acids, Evolution and Mood Disorders. *J Phylogen Evolution Biol* 2:128. doi: 10.4172/2329-9002.1000128
64. Ding X, Yang Z, Han Y, Yu H (2014) Mitochondrial Long Chain Fatty Acid Oxidation Related Enzyme Changes in Different Preeclampsia- Like Mouse Models. *J Hypertens* 3:161. doi: 10.4172/2167-1095.1000161