

Si-Tricalcium Phosphate Cement: Preparation, Characterization and Bioactivity in SBF

Mariana Motisuke^{a,b,*}, Raúl García Carrodegua^c, Cecília Amélia de Carvalho Zavaglia^{b,d}

^aInstituto de Ciência e Tecnologia – ICT, Universidade Federal de São Paulo – UNIFESP, CEP 12.231-280, São José dos Campos, SP, Brazil

^bInstituto Nacional de Ciência e Tecnologia em Biofabricação – INCT/Biofabris, Universidade Estadual de Campinas – UNICAMP, Campinas, SP, Brazil

^cInstituto de Cerámica y Vidrio – ICV,

Consejo Superior de Investigaciones Científicas – CSIC, Madrid, España

^dLaboratório de Biomateriais e Biomecânica – LABIOME, Faculdade de Engenharia Mecânica – FEM, Universidade Estadual de Campinas – UNICAMP, Campinas, SP, Brazil

Received: March 10, 2011; Revised: August 3, 2011

There are evidences considering the effectiveness of Si on enhancing biological properties of calcium phosphates; however, there are not many works relating to the Si- α -TCP bone cement. The influence of silicon doping on the properties of α -TCP cement was analyzed. Si-TCP was obtained by a solid state reaction employing CaCO_3 , CaHPO_4 and CaSiO_3 and powder was analyzed by XRD, FTIR, XRF and BET specific area. Cement samples were analyzed for their surface of fracture morphology, mechanical resistance and SBF bioactivity. Cement mechanical resistance was not satisfactory for biomedical application; nonetheless, sample's surface was coated by an apatite layer after immersion in SBF. Notwithstanding, to ensure that silicon is the element responsible for increasing the material's bioactivity it is necessary to evaluate the in vivo performance of the bone cement obtained in this work.

Keywords: calcium phosphate cements, tricalcium phosphate, wollastonite, silicon-doped tricalcium phosphate, biomedical application

1. Introduction

Ionic substitution into calcium phosphate's crystalline structure has been reported as an important path on the development of orthopedic implants and tissue engineering scaffolds once it is possible to control material's thermal stability, in vivo solubility and cell adhesion and proliferation¹⁻¹². For instance, silicon doped α -tricalcium phosphate (Si- α -TCP) is receiving great attention due to the increased bioactivity and the lower synthesis temperature promoted by the partial substitution of silicon into the tetrahedral phosphorus sites of tricalcium phosphate⁴⁻¹².

Usually, Si- α -TCP is synthesized either by wet precipitation from calcium nitrate and ammonium phosphate solutions in the proper ratio and in the presence of ammonia and colloidal silica or organic silicon compounds, followed by thermal decomposition of the resulting powder^{9,10}; by high temperature solid state reaction from mixtures of CaCO_3 , CaHPO_4 or $(\text{NH}_4)_2\text{PO}_4$, and Ca_2SiO_4 , (or CaSiO_3)^{7,13} or from mixtures of β - $\text{Ca}_3(\text{PO}_4)_2$ and CaSiO_3 ^{11,14}

Wet precipitation method often yields to Ca/(Si + P) ratio different from 1.50 unless strict control of pH, temperature, concentration, and aging of the precipitate are maintained. On the other hand, solid state reaction method allows an accurate control of the Ca/(Si + P) ratio. In spite of that, both methods possess a serious handicap; the "pure" commercial precursors commonly available (especially CaCO_3 and CaHPO_4) always contain a few tenths per cent of Mg which have been recognized as an effective inhibitor of $\beta \rightarrow \alpha$ transformation in TCP, and acts as a stabilizer of the low temperature phase β -TCP^{11,15,16}.

Extra "pure" commercial reagents are very expensive and not often available.

There are many evidences considering the effectiveness of Si- α -TCP and Si-HA on enhancing biological properties of these materials⁴⁻¹², however, there are not many works relating to the Si- α -TCP bone cement which should also have enhanced biological properties. Thus, the process by which the transformation of Si- α -TCP into apatite occurs must be investigated. There are evidences but not too many certainties that silicon is responsible for changes on the setting reaction process and in the final properties of the bone cement^{6,7,11}. Thus, the objective of this report is to characterize the calcium phosphate cement obtained from silicon doped tricalcium phosphate (Si-TCP).

2. Materials and Methods

2.1. TCP powders synthesis and milling

Silicon doped tricalcium phosphate, Si-TCP, was synthesized by solid state reaction of an appropriate mixture of CaCO_3 (Synth, Brazil; 0.16 wt. (%) SiO_2 ; 0.57 wt. (%) MgO ; 0.17 wt. (%) Al_2O_3 ; 0.18 wt. (%) SrO), CaHPO_4 (Synth, Brazil; 0.27 wt. (%) Mg) and CaSiO_3 (Vetec, Brazil; 0.17 wt. (%) Al_2O_3 ; 0.16 wt. (%) MnO ; 0.66 wt. (%) Fe_2O_3) at 1400 °C during 6 hours. Heating rate was 10 °C/min and after the dwelling time the sample was left inside the furnace to cool down to room temperature. Afterwards, the powder was ball milled for one

*e-mail: motisuke@unifesp.br

week. TCP BET specific surface area and particle size distribution were determined using Micromeritics, ASAP 2010 and Malvern Mastersizer S, respectively.

2.2. Calcium phosphate cement

Cement samples were prepared using an aqueous solution containing 2.5 wt. (%) of Na_2HPO_4 and 1.5 wt. (%) of $\text{C}_6\text{H}_8\text{O}_7$ (citric acid) with a liquid-to-powder ratio equal to $0.60 \text{ mL}\cdot\text{g}^{-1}$. After molding in Teflon molds ($6 \times 12 \text{ mm}$ and $12 \times 6 \text{ mm}$) samples were left in a 100% relative moisture environment for 24 hours. Then, cement cylinders were polished, demolded and immersed in SBF for 24 and 168 hours at 37°C . After each immersion period, samples were gently rinsed with distilled water, immersed in acetone to stop the setting reaction and dried at 100°C for 6 hours.

To estimate pH evolution during cement setting reaction, Si-TCP pH evolution as a function of time was evaluated by preparing a Si-TCP suspension ($100 \text{ mg}\cdot\text{mL}^{-1}$) in a buffer solution ($\text{pH} = 7.4$). The suspension was maintained at 37°C and the pH was measured in different periods of times during 14 days.

Cements samples were mechanical tested (MTS, Test Star II) and the morphology of the surface of fracture was analyzed by scanning electron microscopy (JEOL-6400).

2.3. X-ray diffraction

Crystalline phase purity was analyzed by X-ray diffraction (Rigaku DMAX-2200, $\text{CuK}\alpha$, Ni filter, $20 \text{ a } 40^\circ (2\theta)$, $0.02^\circ/\text{s}$, 40 kV e 40 mA). JCPDS files used for phase identification were 09-0348 for α -TCP, 09-0169 for β -TCP and 46-0905 for CDHA (calcium deficient hydroxyapatite). β -TCP mass fraction present in the cement starting material was determined using the internal pattern method¹⁷.

2.4. Fourier transformed infrared

TCP powder and cement were diluted in KBr and FTIR analysis was carried out on a Perkin Elmer 1600 FT-IR spectrometer with a scanning range from 450 to 4000 cm^{-1} and resolution of 2 cm^{-1} .

2.5. X-ray fluorescence

TCP Ca/P ratio was determined using a MagiX Super Q Version 3.0 X-ray fluorescence spectrometer (Philips, The Netherlands) provided with Rh X-ray tube and power generator of 2.4 kW . 0.3000 g of each mixture were weighed, mixed with 5.5 g of spectral grade $\text{Li}_2\text{B}_4\text{O}_7$ and melted in a Pt/Au crucible and formed into disks in a special controlled furnace Per1'X3 (Philips, The Netherlands). Calibration curves were prepared using certified composition standards of natural and synthetic calcium phosphates and calcium silicates.

2.6. Bioactivity in SBF

After setting for 168 hours cements samples ($12 \times 6 \text{ mm}$ discs) were immersed once again in SBF at 37°C for analyzing the occurrence of apatite precipitation on the sample' surface after one week. After the immersion time, samples were gently rinsed with distilled water and dried overnight. To verify the presence of apatite precipitation the sample surface was gold coated and was analyzed by scanning electron microscopy (JEOL -6400).

3. Results and Discussions

Si-TCP was obtained from the solid state reaction employed; however, due to high magnesium levels of the precursors, Si-TCP was obtained with 19 wt. (%) of β -TCP. This element is a well established stabilizer of the β -TCP phase since it raises β -TCP thermal stability to almost 1400°C depending on Mg content^{15,16}. As it is observed

on the XRD pattern of Figure 1 there is a β -TCP characteristic XRD line at approximately $28^\circ 2$ -theta.

Indeed, the formation of tricalcium phosphate was also verified on FTIR analysis since the absorption bands present on sample's spectrum (Figure 2) are characteristic of α -TCP as displayed on Table 1^{18,19}.

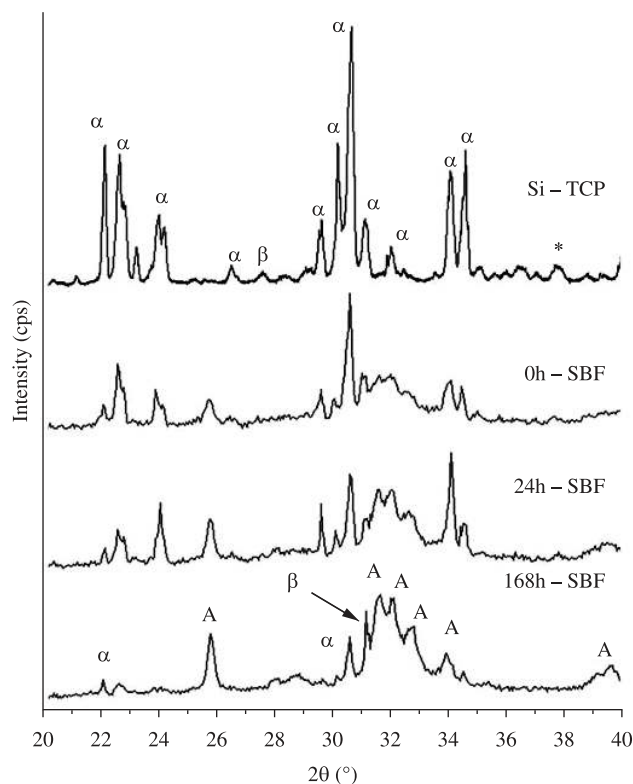


Figure 1. XRD patterns from Si-TCP and cement samples after 0, 24 and 168 hours of SBF immersion. Legend = α : α -TCP, β : β -TCP, A: apatite, *: CaO.

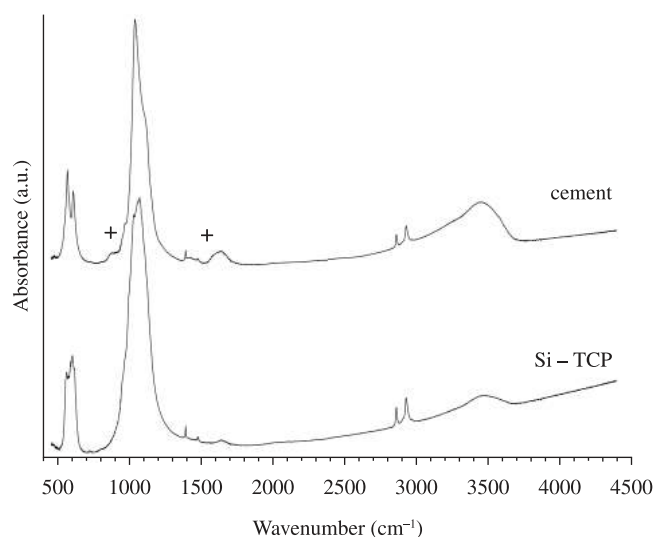


Figure 2. FTIR spectrum of α -TCP powders and cement sample after 168 hours of setting. "+" represents CO_3^{2-} -absorbance bands.

Table 1. FTIR absorption bands of α -TCP¹⁸.

Absorption	Bond	Wavenumber (cm ⁻¹)
ν_1	P-O	963
ν_2	OPO	462
ν_3	P-O	1120
		1100
		1084
		1025
		990
ν_4	OPO	597
		583
		572
		559

Table 2. Ca/(P + Si) and (Ca + Mg)/(P + Si) ratios for Si-TCP.

Ca (wt. (%))	P (wt. (%))	Si (wt. (%))	Mg (wt. (%))	Ca/ (P + Si)	(Ca + Mg)/ (P + Si)
38.70	18.68	1.27	0.18	1.49	1.50

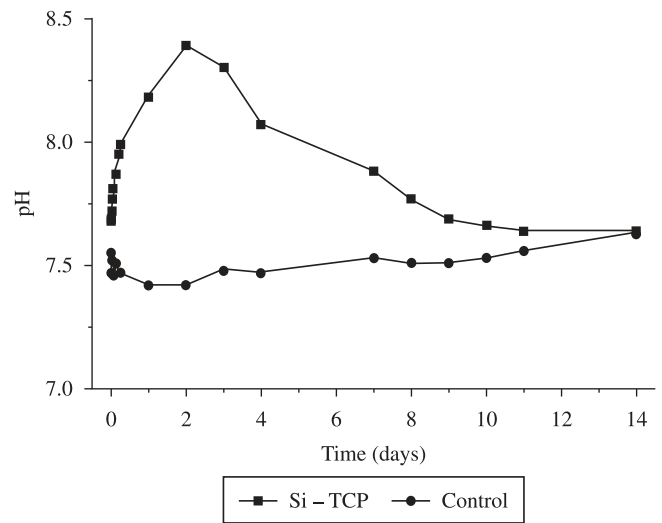


Figure 3. Si-TCP pH evolution in a buffer solution (pH = 7.4). CaO in aqueous solutions forms Ca(OH)₂ leading to an pH increase which could be responsible for Si-TCP lower reactivity during cement setting reaction.

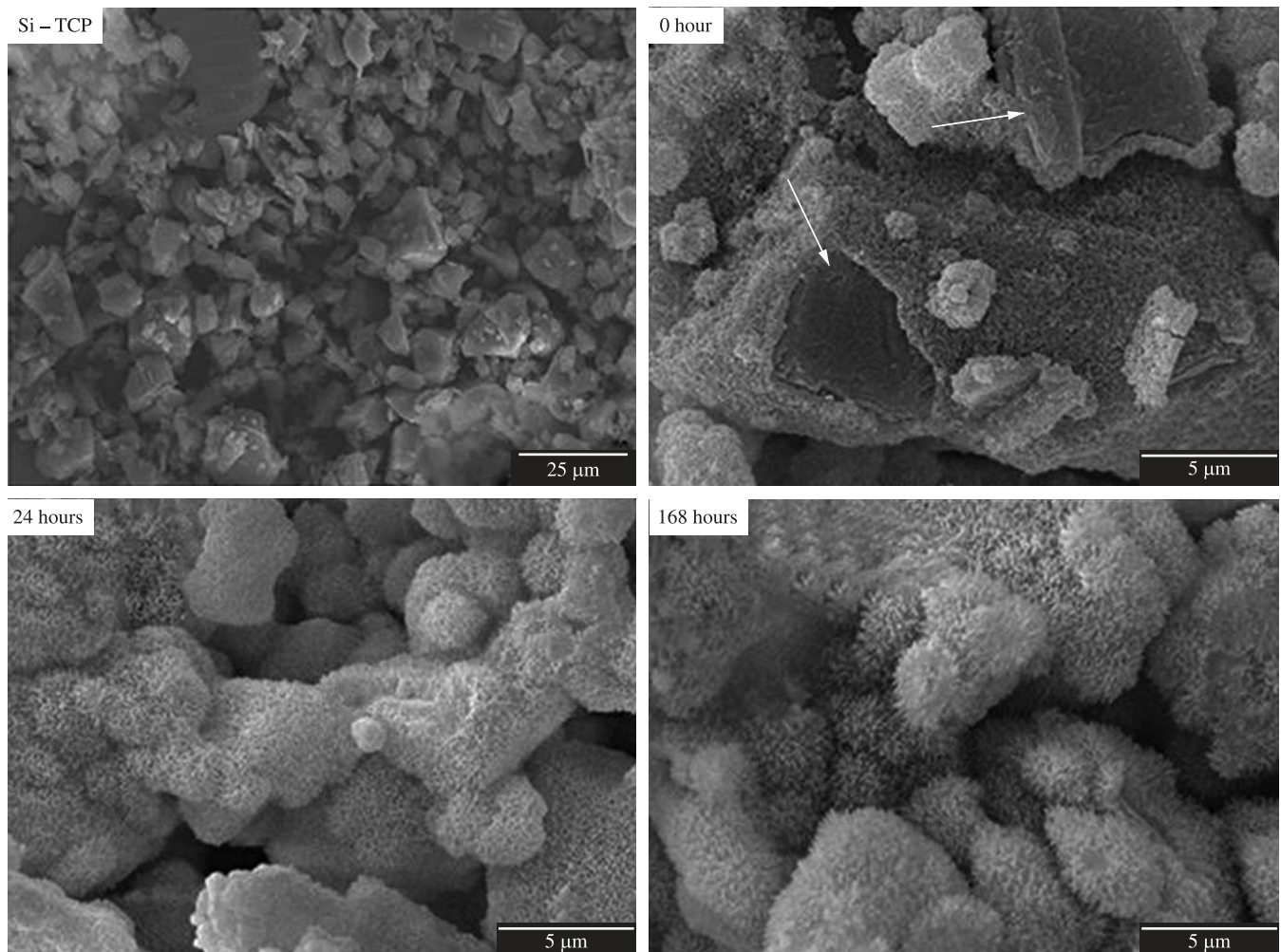


Figure 4. SEM micrographs of Si-TCP powder, cement sample after 24 hours 100% of relative moisture (0 hour) and cement samples after 24 hours and 168 hours of SBF immersion. The white arrows indicate α -TCP grains without reacting.

Moreover, it is possible to verify that due to wollastonite (CaSiO_3) addition and to Mg substitution into some Ca^{2+} sites a tiny deviation on the system stoichiometry (calcium excess) has occurred leading to

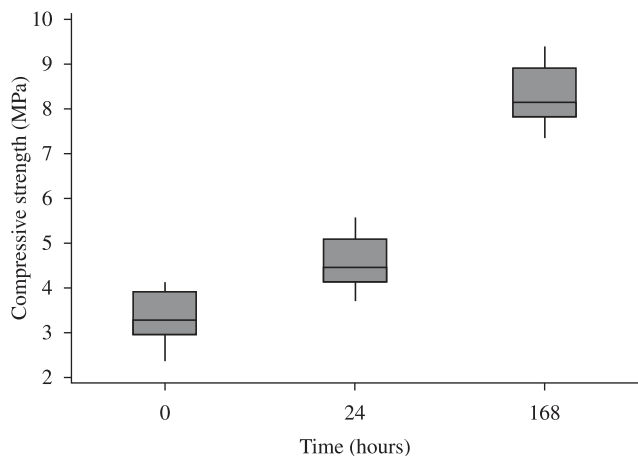


Figure 5. Cement samples compressive strength evolution with time. SBF immersion was responsible for the mechanical resistance enhancement. After ANOVA statistic analysis all samples can be considered to be statistically different ($p < 0.005$).

the formation of calcium oxide (CaO) after sintering at $1400\text{ }^\circ\text{C}$ (the * on Figure 1). Nonetheless, the values of $\text{Ca}/(\text{P} + \text{Si})$ and $(\text{Ca} + \text{Mg})/(\text{P} + \text{Si})$ ratios obtained by quantitative XRF (Table 2) was 1.49 and 1.50, respectively. The presence of some CaO in the final material was responsible for pH increase during the setting reaction (Figure 3).

The keys to the synthesis of pure Si- α -TCP are to ensure the right stoichiometry (i.e. atomic ratio $\text{Ca}/(\text{P} + \text{Si}) = 1.50$); to exclude the presence of Mg in the reagents employed, and to reach the proper temperature for maturing α -phase^{9,11,15}. The first requisite is easily accomplished by using the solid state synthetic method, which allows the accurate weighting of the required amounts of each reactant. The second one may be achieved by employing Mg-free reagents. A previous work¹³ proposed the synthesis of alpha-TCP¹⁶ precursors CaCO_3 and CaHPO_4 by wet precipitation in the presence of ethylenediamine tetraacetic acid (EDTA). In that work, it was verified that EDTA forms a stable complex with Mg, preventing its co-precipitation with the Ca salt. The effectiveness of the proposed procedure was demonstrated by the low Mg-contents of the obtained precursors and the phase purity of the Si- α -TCP prepared from them¹³.

After 168 hours of ball milling, Si-TCP powder achieved a main particle size of approximately $15.5\text{ }\mu\text{m}$, a $10\% < d < 90\%$ distribution between $0.84\text{--}39.66\text{ }\mu\text{m}$ and a specific BET surface area of $1.0293 \pm 0.0263\text{ m}^2\cdot\text{g}^{-1}$. Si-TCP powder morphology and the heterogeneous particle size distribution can be observed on Figure 4.

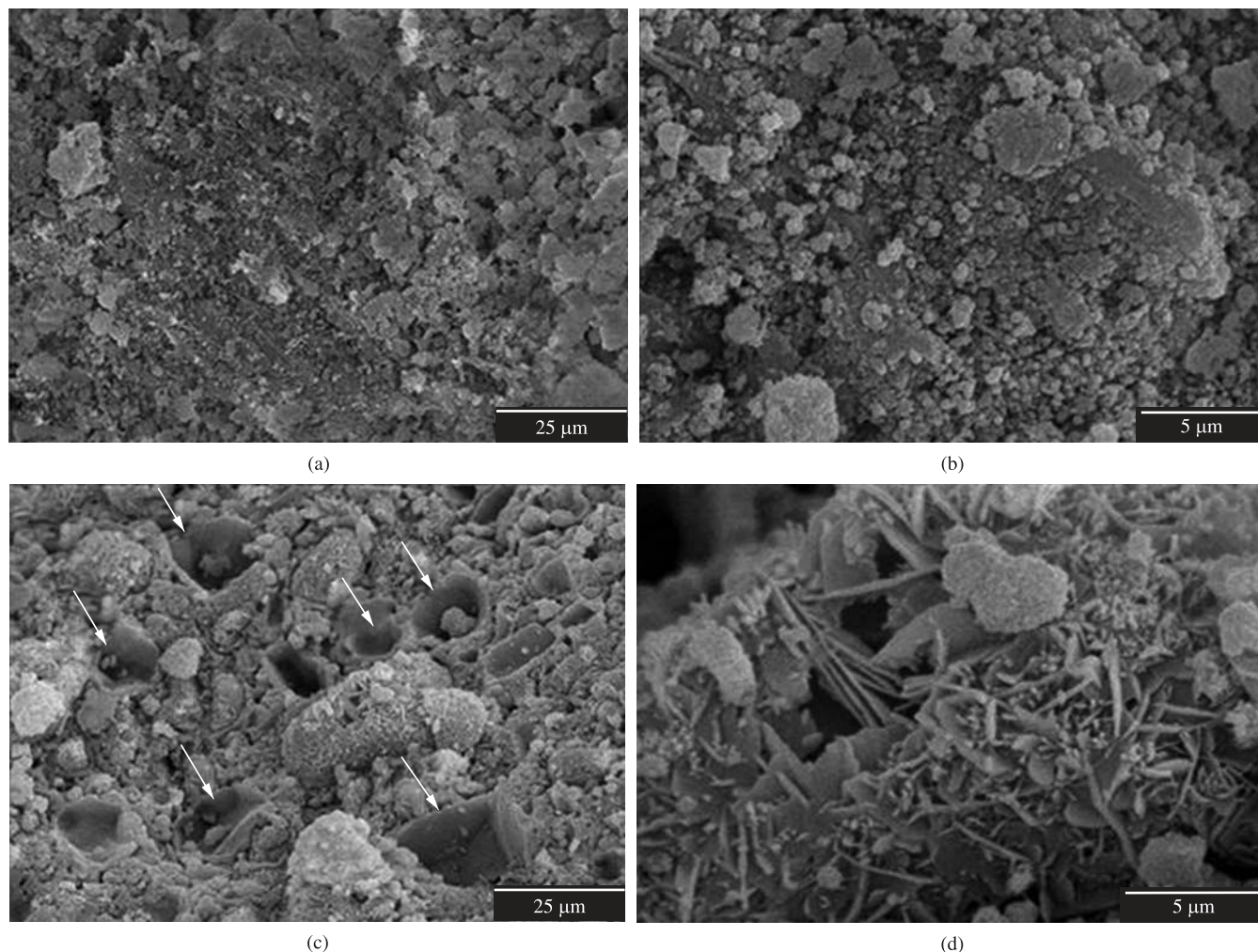


Figure 6. SEM micrographs of cement surface after 168 hours of setting (a and b) and an additional 168 hours of SBF immersion. The white arrows represent holes on the surface which are appeared as a result of the setting reaction continuation. The surface of these holes is also coated with an apatite layer.

Si-TCP cement setting reaction occurs as expected for α -TCP cement¹⁸. First, the superficial dissolution of the TCP grains occurs and after reaching the ionic saturation for apatite, tiny apatite crystals precipitate on this same surface. The white arrows of Figure 4 indicate TCP grains surrounded by an apatite layer which was fractured during the compressive mechanical testing. As the setting reaction evolves more TCP particles are dissolved and more apatite crystals are precipitated as can be observed on XRD patterns and SEM micrographs of the surface of fracture of cement samples after several setting times (Figure 1 and Figure 4, respectively). Moreover, comparing SEM micrographs for samples at 24 and 168 hours of reaction it was verified that the apatite crystals has grown as a function of immersion time.

Although the process involved on the cement setting reaction occurred as reported on the literature¹⁸, the rate by which Si-TCP evolves into apatite is very low compared to the values normally encountered^{20,21}. This fact could be attributed to many reasons: i) Si-TCP small surface area²⁰; ii) high β -TCP weight contents^{18,21} and; iii) the presence of CaO which raises environment pH making difficult TCP dissolution and apatite precipitation. Indeed, in Figure 3 it is possible to verify that after 48 hours pH value is 8.39 and it only reach physiological values after 14 days. For instance, on XRD diffractograms (Figure 1) α -TCP \rightarrow apatite reaction is not complete even after 7 days. Furthermore, it is important to highlight that the setting reaction resulted on a carbonated apatite phase due to the CO_3^{2-} characteristics bands at 850 to 900 and at 1350 to 1600 cm^{-1} highlighted with a "+" on Figure 2. Unfortunately, it was not possible to verify any Si-O absorbance bands on cement spectrums; thus, to verify the precipitation of silicon doped apatite during Si-TCP cement setting reaction it is necessary to investigate this fact more deeply.

Apatite crystal size evolution was responsible for compressive strength enhancement with time of the cement samples. As displayed on Figure 5, at initial times the samples did not present a considerable mechanical resistance (3.35 ± 0.59 and 4.54 ± 0.62 MPa, at zero and 24 hours of SBF immersion, respectively). As the setting reaction evolves and the apatite crystals entanglement became more cohesive, compressive strength raises, however, as observed on the box plot for cement at 168 hours the maximum mechanical resistance achieved (8.32 ± 0.67 MPa) is very low when compared to the values reported on the literature, 30-60 MPa^{7,18}. This fact can be attributed to the high levels of β -TCP (19 wt. (%)) and the low reactivity of the Si-TCP powder. In fact, on Figure 1 it is possible to verify that even after 168 hours of reaction there still some visible XRD peaks of α -TCP. The presence of no reacted particles (β and α -TCP) acts as stress concentration points and the material fail faster. Moreover, the low mechanical resistance can be attributed to the high liquid-to-powder ratio employed (0.60 mL.g^{-1}) which increases porosity leading to more fragile materials.

Cement bioactivity in SBF can be verified in Figure 6 as a visible apatite layer was precipitated on sample's surface after 168 hours of immersion. The superficial "holes" (indicated with white arrows on Figure 6) were a result of the evolution of setting reaction. Probably, α -TCP particles which did not react after 168 hours were dissolved during the bioactivity assay. Moreover, the surface of the superficial "holes" is coated with apatite particles. Notwithstanding, to ensure that silicon is the element responsible for increasing the materials bioactivity^{7,12,14} it is necessary to evaluate the in vivo performance of the bone cement obtained in this work.

4. Conclusion

It was possible to synthesize at 1400 °C silicon doped tricalcium phosphate by mixing CaCO_3 , CaHPO_4 and CaSiO_3 . Nevertheless,

due to high Mg contamination, α -TCP was formed with 19 wt. (%) of β -TCP as impurity.

Cement setting reaction take place by the same steps of conventional α -TCP cement; however, the rate of reaction was very low and TCP \rightarrow apatite conversion was not complete even after 168 hours as a result of β -TCP and CaO contamination which dissolution processes compete with α -TCP dissolution and apatite precipitation.

Even though the cement developed in this work did not achieve a satisfactory mechanical performance, the bioactivity in SBF may be taken into account for encouraging further investigation and optimization of its properties. Notwithstanding, to ensure that silicon is the element responsible for increasing the materials bioactivity it is necessary to evaluate the in vivo performance of the bone cement obtained in this work.

Acknowledgement

The State of São Paulo Foundation (FAPESP) and the INCT-Biofabris for the financial support

References

- Wei X, Ozan U, Ankit A, Acar HY and Akinc M. Dissolution behavior of Si, Zn-codoped tricalcium phosphate. *Materials Science and Engineering C*. 2009; 29(1):126-135. <http://dx.doi.org/10.1016/j.msec.2008.05.020>
- Matsumoto N, Yoshida K, Hashimoto K and Toda Y. Thermal Stability of β -tricalcium phosphate doped with monovalent metal ions. *Materials Research Bulletin*. 2009; 44(9):1889-1894. <http://dx.doi.org/10.1016/j.materresbull.2009.05.012>
- Li X, Ito A, Sogo Y, Wang X and LeGeros RZ. Solubility of Mg-containing β -tricalcium phosphate at 25°C. *Acta Biomaterialia*. 2009; 5(1):508-517. <http://dx.doi.org/10.1016/j.actbio.2008.06.010>
- Pietak AP, Reid JW, Stott MJ and Sayer M. Silicon Substitution in the Calcium Phosphate Bioceramics. *Biomaterials*. 2007; 28(28):4023-4032. <http://dx.doi.org/10.1016/j.biomaterials.2007.05.003>
- Arcos D, Sanchez-Salcedo S, Izquierdo-Barba I, Ruiz L, Gonzalez-Calbet J and Vallet-Regi M. Crystallochemistry, textural properties, and in vitro biocompatibility of different silicon-doped calcium phosphates. *Journal of Biomedical Material Research Part A*. 2006; 78(4):762-771. <http://dx.doi.org/10.1002/jbm.a.30790>
- Camiré CL, Jegou Saint-Jean S, McCarthy I, Mochales-Palau C, Lidgren L, Planell JA et al. Production Methodology and Reactivity of Silica Substituted α phase Tricalcium Phosphate. In: *Proceedings of 7th World Biomaterials Congress*; 2004; Sydney, Australia. Sydney; 2004.
- Camiré CL, Saint-Jean SJ, Mochales C, Nevsten P, Wang JS, Lidgren L et al. Material Characterization and In Vivo Behavior of Silicon Substituted α -Tricalcium Phosphate Cement. *Journal of Biomedical Material Research Part B: Applied Biomaterials*. 2006; 76(2):424-431.
- Reid JW, Pietak A, Sayer M, Dunfield D and Smith TJ Phase formation and evolution in the silicon substituted tricalcium phosphate/apatite system. *Biomaterials*. 2005; 26(16):2887-2897. <http://dx.doi.org/10.1016/j.biomaterials.2004.09.005>
- Reid JW, Tuck L, Sayer M, Fargo K and Hendry JA. Synthesis and Characterization of Single-Phase Silicon-Substituted α -Tricalcium Phosphate. *Biomaterials*. 2006; 27(15):2916-2925. <http://dx.doi.org/10.1016/j.biomaterials.2006.01.007>
- Langstaff S, Sayer M, Smith TJ, Pugh SM, Hesp SA and Thompson WT. Resorbable bioceramics based on stabilized calcium phosphates. Part I: rational design, sample preparation and material characterization. *Biomaterials*. 1999; 20(18):1727-1741. [http://dx.doi.org/10.1016/S0142-9612\(99\)00086-1](http://dx.doi.org/10.1016/S0142-9612(99)00086-1)
- Carrodegua RG and De Aza S. Alpha-Tricalcium phosphate: synthesis, properties and biomedical applications. *Acta Biomaterialia*. 2011; in press.

12. Gaspar AMM, Saska S, Carrodegua RG, De Aza AH, Pena P, De Aza PN et al. Biological response to wollastonite doped α -tricalcium phosphate implants in hard and soft tissues in rats. *Key Engineering Materials*. 2009; 396-398:7-10.
13. Motisuke M, Carrodegua RG and Zavaglia CAC. Mg-free precursors for the synthesis of pure phase Si-doped α -Ca₃(PO₄)₂. *Key Engineering Materials*. 2008; 361-363: 199-202. <http://dx.doi.org/10.4028/www.scientific.net/KEM.361-363.199>
14. Carrodegua RG, De Aza AH, Jimenez J, De Aza PN, Pena P, López-Bravo A et al. Preparation and in vitro characterization of wollastonite doped tricalcium phosphate bioceramics. *Key Engineering Materials*. 2008; 361-363:237-240. <http://dx.doi.org/10.4028/www.scientific.net/KEM.361-363.237>
15. Reid JW, Fargo K, Hendry JA and Sayer M. The Influence of Trace Magnesium Content on the Phase Composition of Silicon-Stabilized Calcium Phosphate Powders. *Materials Letters*. 2007; 61(18):3851-3854. <http://dx.doi.org/10.1016/j.matlet.2006.12.046>
16. Carrodegua RG, De Aza A H, Turrillas X, Pena P and De Aza S. New approach to the $\beta \rightarrow \alpha$ polymorphic transformation in magnesium-substituted tricalcium phosphates and its practical implications. *Journal of the American Ceramic Society*. 2008; 91(4):1281-1286. <http://dx.doi.org/10.1111/j.1551-2916.2008.02294.x>
17. Cullity BD. *Elements of X-Ray Diffraction*. Indiana: Addison-Wesley Publishing Company, Inc.; 1977.
18. Ginebra MP, Fernández E, De Maeyer EA, Verbeeck RM, Boltong MG, Ginebra J et al. Setting Reaction and Hardening of an Apatitic Calcium Phosphate Cement. *Journal of Dental Research*. 1997; 76(4):905-912. <http://dx.doi.org/10.1177/00220345970760041201>
19. Dunfield D, Sayer M and Shurvell HF. Total Attenuated Reflection Infrared Analysis of Silicon-Stabilized Tri-Calcium Phosphate. *The Journal of Physical Chemistry B*. 2005; 109(42):19579-19583. <http://dx.doi.org/10.1021/jp0519823>
20. Ginebra MP, Driessens FCM and Planell JA. Effect of the Particle Size on the Micro and Nanostructural Features of a Calcium Phosphate Cement: a Kinetics Analysis. *Biomaterials*. 2004; 25(17):3453-3462. <http://dx.doi.org/10.1016/j.biomaterials.2003.10.049>
21. Ginebra MP, Fernandez E, Driessens FCM and Planell JA. Modeling of the Hydrolysis of α -Tricalcium Phosphate. *Journal of the American Ceramic Society*. 1999; 82(10):2808-2812. <http://dx.doi.org/10.1111/j.1151-2916.1999.tb02160.x>