Word count: Abstract: 249 Text: 3145 Refs: 40 Tables: 0 Figures: 2 Supp Text Supp Tables: 6 Supp Figs: 8

# Genome-Wide Association Study Reveals First Locus for Anorexia Nervosa and Metabolic Correlations

#### Authors

Laramie Duncan, PhD Zeynep Yilmaz, PhD Raymond Walters, PhD Jackie Goldstein, PhD Verneri Anttila, PhD Brendan Bulik-Sullivan, PhD Stephan Ripke, MD, PhD Eating Disorders Working Group of the Psychiatric Genomics Consortium Laura Thornton, PhD Anke Hinney, PhD Mark Daly, PhD Patrick Sullivan, MD, FRANZCP Eleftheria Zeggini, PhD Gerome Breen, PhD Cynthia Bulik, PhD\* \*Corresponding author

**Previous presentation:** This work was presented at the World Congress of Psychiatric Genetics, November 2016, Jerusalem, Israel.

Location of work: Dr. Bulik, Center of Excellence for Eating Disorders, Department of Psychiatry, CB #7160, 101 Manning Drive, Chapel Hill, NC 27599, USA and Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm Sweden. cbulik@med.unc.edu. Work also conducted at the Analytic and Translational Genetics Unit in the Department of Medicine at Massachusetts General Hospital and the Broad Institute of MIT and Harvard.

## Abstract

*Objective:* To conduct a genome-wide association study (GWAS) of anorexia nervosa and to calculate genetic correlations with a series of psychiatric, educational, and metabolic phenotypes. *Method:* Following uniform quality control and imputation using the 1000 Genomes Project (phase 3) in 12 case-control cohorts comprising 3,495 anorexia nervosa cases and 10,982 controls, we performed standard association analysis followed by a meta-analysis across cohorts. Linkage disequilibrium score regression (LDSC) was used to calculate genome-wide common variant heritability  $[h_{SNP}^2$ , partitioned heritability, and genetic correlations ( $r_g$ )] between anorexia nervosa and other phenotypes.

*Results:* Results were obtained for 10,641,224 single nucleotide polymorphisms (SNPs) and insertion-deletion variants with minor allele frequency > 1% and imputation quality scores > 0.6. The  $h_{SNP}^2$  of anorexia nervosa was 0.20 (SE=0.02), suggesting that a substantial fraction of the twin-based heritability arises from common genetic variation. We identified one genome-wide significant locus on chromosome 12 (rs4622308, p=4.3x10<sup>-9</sup>) in a region harboring a previously reported type 1 diabetes and autoimmune disorder locus. Significant positive genetic correlations were observed between anorexia nervosa and schizophrenia, neuroticism, educational attainment, and high density lipoprotein (HDL) cholesterol, and significant negative genetic correlations between anorexia nervosa is a complex heritable phenotype for which we have found the first genome-wide significant locus. Anorexia nervosa also has large and significant genetic correlations with both psychiatric phenotypes and metabolic traits. Our results encourage a reconceptualization of this frequently lethal disorder as one with both psychiatric and metabolic etiology.

## Introduction

Anorexia nervosa is a serious eating disorder characterized by restriction of energy intake relative to requirements, resulting in abnormally low body weight. It has a lifetime prevalence of approximately 1%, disproportionately affects females (1, 2), and has no well replicated evidence of effective pharmacologic or psychological treatments, despite high morbidity and mortality (3, 4). Twin studies consistently support a genetic basis for the observed familial aggregation in anorexia nervosa, with heritability estimates of 48%-74% (5). Although initial genome-wide association studies (GWASs) were underpowered (6, 7), the available evidence strongly suggested that signals for anorexia nervosa would be detected with increased sample size (6).

The aim of the current study was to combine existing samples to conduct a more powerful GWAS of anorexia nervosa. To further characterize the nature of the illness, we applied linkage disequilibrium score regression (LDSC) (8) to calculate genome-wide common variant heritability  $(h_{SNP}^2)$ , partitioned heritability, and genetic correlations  $(r_g)$  between anorexia nervosa and other phenotypes. These include the other major psychiatric disorders with large GWAS, namely schizophrenia, bipolar disorder, major depressive disorder, autism, and attention deficit hyperactivity disorder (ADHD). We then used  $r_g$  estimates between anorexia nervosa and 159 additional phenotypes (as described below) to characterize the phenome-wide genetic architecture of AN.

### Methods

<u>Cases and controls.</u> Our sample included 3,495 anorexia nervosa cases and 10,982 controls. Case definition required a diagnosis of lifetime anorexia nervosa (restricting or binge–purge subtype) or lifetime eating disorders 'not otherwise specified' anorexia nervosa-subtype (i.e., exhibiting the core features of anorexia nervosa). A lifetime history of bulimia nervosa was allowed given the frequency of diagnostic crossover (9). Amenorrhea was not required as it does not increase diagnostic specificity (10) [it has been removed as a diagnostic criterion in the DSM-5 (11)]. Extensive information on diagnostic and consensus procedures for the samples included in the Children's Hospital of Philadelphia/Price Foundation Collaborative Group (CHOP/PFCG) cohort

are available in (12). The cases included from the Genetic Consortium for Anorexia Nervosa/Wellcome Trust Case Control Consortium-3 (GCAN/WTCCC3) GWAS came from 12 previously collected clinical or population cohorts. Given that these were archived samples, the calculation of reliability statistics on diagnoses was not possible. Mitigating that concern, however, is that anorexia nervosa is a highly homogeneous phenotype with typical kappa values ranging from .81-.97 (13). Moreover, the approach taken here is consistent with successful GWAS meta-analysis efforts across psychiatric diagnoses, in which larger samples have overcome the challenges posed by imperfect diagnoses and small individual variant effect sizes.

Individuals with conditions including schizophrenia, intellectual disability, and medical or neurological conditions causing weight loss were excluded, as per previous reports (6, 7). All sites had documented permission from local ethical committees and all participants provided informed consent.

Consistent with procedures established by the Psychiatric Genomics Consortium (PGC) (14, 15), we collected individual-level genotype (GWAS array) and phenotype (binary case-control status) data from contributing previous GWAS consortia and groups (for a description see **Supplementary Table S1**). In particular, the previous reports on anorexia nervosa GWAS from CHOP/PFCG data (7) and the GCAN/WTTTC3 (6) provide further details about cohort ascertainment and participant characteristics not described below or in the **Supplementary Text**.

Although most of the cases included in the published anorexia nervosa GWASs were included in this analysis, many of the controls used in previous GWAS could not be used for subsequent analyses. To summarize, our analysis includes the CHOP/PFCG data (7) plus *cases* from 12 of the 15 strata included in the GCAN/WTCCC3 analysis of anorexia nervosa (6). Three datasets (Italy-North, Sweden, and Poland) in Boraska et al. were dropped from our analysis because appropriately matched controls could not be found and/or where case plus control numbers were < 100. After removing these three datasets and combining the US and Canadian cases, we included 11 GCAN/WTCCC3-based datasets plus the CHOP/PFCG dataset in our analyses. For the nine datasets requiring new controls, we first evaluated diverse control datasets from Psychiatric Genomics Consortium (PGC) collaborators for potentially suitable controls based on

geographic location and Illumina genotyping. We then performed quality control (QC) steps (below, and with additional details in the **Supplementary Text**), using visual inspection of principal components plots (comparing cases to controls) as well as QQ and Manhattan plots (for evidence of systematic bias) to identify suitably matched controls. All samples in this report are of European ancestry. As shown in **Supplementary Figure S1**, all of the datasets (except for Finland) form a gradient of clusters when visualized in a scatter plot of the first two principal components, as expected based on known population genetic features (16).

Quality control and analysis. Following uniform quality control and imputation using the 1000 Genomes Project (phase 3) (17) in the anorexia nervosa case-control cohorts, we performed association analysis using an additive model using the dosage data for each cohort. Following adjustment for unbalanced case and control numbers across our 12 strata [see (18)], our summed effective balanced sample size was 5082 cases and 5082 controls. Accordingly, our power was 83.1% for a genotype relative risk of 1.25, at an allele frequency of 0.2 at p<5x10<sup>-8</sup> (http://zzz.bwh.harvard.edu/gpc). Analysis within datasets was performed in PLINK with the first ten principal components as covariates. METAL (18) was used to conduct fixed-effects meta-analysis across the twelve datasets using inverse-variance weighting. Results were obtained for 10,641,224 single nucleotide polymorphisms (SNPs) and insertion-deletion variants with minor allele frequency > 1% and imputation quality scores > 0.6 [see **Supplementary Figure S2** for quantile-quantile (QQ) plot]. The GWAS statistic inflation ( $\lambda$ ) was 1.080 with a sample size adjusted  $\lambda_{1000}$  of 1.008, consistent with minimal population stratification or other systematic biases. Plotting was performed in R (19) (see **Supplementary Text** for additional methods and quality control details and **Supplementary Table S1** for individual study details).

## Statistical significance

The primary analysis in this paper is the GWAS, which analyzes each SNP for association to phenotype. The international standard for statistical significance is  $p < 5x 10^{-8}$ , which corrects for the approximately one million independent statistical tests conducted. Focused secondary analyses are now the expectation for primary GWAS reports, and we describe statistical significance thresholds for them individually. We used the accepted and expected methods of multiple testing correction. Gene-based and pathway analyses were also conducted. For these

analyses, statistical significance was set using the Bonferroni correction, which is conservative given non-independence among the gene-based and pathway statistical tests. For the gene-based analyses, we defined statistical significance as gene p-value  $< 2.6 \times 10^{-6} (0.05/19,222 \text{ genes})$  tested). For pathway analyses: p-value  $< 1.8 \times 10^{-5} (0.05/2,714 \text{ pathways tested})$ .

Analytical methods for estimating heritability and genetic correlations, and for gene-based and pathway analyses, are presented in the **Supplementary Text**. Regarding the rationale for these particular secondary analyses, we note that these are often considered to be standard analyses for GWAS reports across medicine. In this particular application, we estimate SNP heritability for anorexia nervosa because it is important to quantify the combined effects of common variants on anorexia nervosa and compare it with other complex disorders and traits, within and outside psychiatry.

#### Results

<u>GWAS.</u> One locus achieved genome-wide significance for a single variant, as shown in the Manhattan plot in **Figure 1**, in which the threshold for significance,  $p < 5x10^{-8}$ , is denoted with dotted line. The top locus (chromosome 12q13.2) overlaps six genes (*IKZF4*, *RPS26*, *ERBB3*, *PA2G4*, *RPL41*, and *ZC3H10*), and is located near six additional genes (*ESYT1*, *SUOX*, *RAB5B*, *CDK2*, *PMEL*, and *DGKA*). The top SNP was rs4622308 (p=4.3x10<sup>-9</sup>, odds ratio (OR)=1.2, standard error (SE)=0.03, minor allele frequency in cases (MAF<sub>cases</sub>)=0.48, minor allele frequency in controls (MAF<sub>controls</sub>)=0.44). We found no evidence for heterogeneity in effect sizes across cohorts (Q=12.58, p=.32) and estimated that 12.59 percent of variation was due to heterogeneity instead of chance (I<sup>2</sup>=12.59). The effects across studies are shown in the forest plot of rs4622308 in **Supplementary Figure S3**.

The results of conditional regression analyses are consistent with the existence of one signal at the top locus (see **Supplementary Figure S4**). The top SNP rs4622308 is in high linkage disequilibrium (LD) ( $r^2=0.86$ ; D'>0.99) with rs11171739, which has been found to be associated in GWASs of type 1 diabetes (20) and rheumatoid arthritis (21). The risk associated alleles of both SNPs (C-C) are typically found on the same haplotype, i.e., the direction of effect for the risk allele is consistent across anorexia nervosa and these other disorders. Several other immune-

related phenotypes: vitiligo, alopecia areata, and asthma (see **Supplementary Figure S5**) also have associations in the region, although these are (somewhat) LD independent of rs4622308.

Information for the top ten loci is given in **Supplementary Table S2.** The second (rs200312312 on chromosome 5, p= $6.7 \times 10^{-8}$ ), third (rs117957029 on chromosome 12, p= $1.6 \times 10^{-7}$ ), and fourth (rs11174202 on chromosome 12, p= $3.1 \times 10^{-7}$ ) most significant loci in our analyses also have consistent evidence for association across multiple cohorts (see **Supplementary Figure S6** for area plots of these loci). The fourth best locus is intronic in the *FAM19A2* gene.

<u>Gene-based and pathway analyses.</u> Multiple genes, all but one of which were in the region around the top SNP (rs4622308), reached gene-based significance (reflecting the high LD in the region). The remaining significant gene was *FAM19A2*, a putative chemokine/cytokine, and the 4<sup>th</sup> best locus in our SNP based analyses. No pathways were significant (see **Supplementary Table S3** for the complete gene-based and pathway analysis results). As has typically been reported for other psychiatric disorders, candidate genes from previous studies did not reach gene-based significance [or in our other analyses; for a detailed review of the candidate gene literature see (5)].

<u>Gene expression.</u> Interrogation of databases such as GTeX (22) did not indicate that any of the genes in the top region have distinct patterns of brain gene expression. Searches using both GTeX and the SNP tag lookup function in MRbase (www.mrbase.org/beta) indicated that that the top SNP (rs4622308) is not, directly or via LD tagging, an eQTL or mQTL. In addition, differential expression in an exploratory mouse model did not suggest a distinct pattern of gene expression (**Supplementary Figure S7**).

Linkage disequilibrium score regression (LDSC). LDSC (8, 23) was used to calculate  $h_{SNP}^2$ , partitioned heritability, and  $r_g$  between anorexia nervosa and other psychiatric, medical, and educational phenotypes. Heritability estimates reported here afford comparison of AN to other major psychiatric disorders. We made comparisons to the psychiatric disorders that have been examined with adequately sized GWAS to afford reliable estimates of heritability, including schizophrenia, bipolar disorder, major depressive disorder, autism, and ADHD. The genetic

correlation estimates between AN and 159 additional phenotypes (with publically available GWAS summary statistics) further characterize the genetic architecture of AN, by providing the magnitude and direction of shared genetic effects between AN and diverse psychological, medical, metabolic, and educational phenotypes.

In our cohort,  $h_{SNP}^2$  for anorexia nervosa was 0.20 (SE=0.021), comparable to  $h_{SNP}^2$  estimates for other psychiatric disorders (**see Supplementary Figure S8**). Partitioned heritability estimates for annotation categories and cell types were not significant after multiple testing correction (for complete results see **Supplementary Table S4**).

A wide range of genetic correlations between anorexia nervosa and other phenotypes were statistically significant. Of 159 phenotypes tested, 29 had false discovery rate (FDR)<0.05 (uncorrected p-values reported below). See **Figure 2** for depiction of these genetic correlations and text below for selected examples. All 159 genetic correlations and relevant references are available in **Supplementary Table S5**.

Notable significant genetic correlations between anorexia nervosa and psychiatric traits and disorders included neuroticism ( $r_g$ =0.39, SE=0.14, p=4.4x10<sup>-3</sup>), schizophrenia ( $r_g$ =0.29, SE=0.07, p=4.4x10<sup>-5</sup>), and results from a meta-analysis across psychiatric phenotypes ( $r_g$ =0.22, SE=0.07, p=3.4x10<sup>-3</sup>). Genetic correlations between anorexia nervosa and educational phenotypes such as years of education ( $r_g$ =0.34, SE=0.08, p=5.2x10<sup>-6</sup>) and attending college ( $r_g$ =0.30, SE=0.07, p=4.4x10<sup>-5</sup>) were also positive and significant. Obsessive compulsive disorder GWAS data were unavailable to us but a previous analysis reported a positive  $r_g$  with anorexia nervosa of 0.53 (SE=0.11, SE=0.13, p=5.5x10<sup>-6</sup>) (24).

Several significant negative genetic correlations emerged between anorexia nervosa and weightrelated phenotypes, suggesting shared genetic loci underlying these phenotypes and opposing effects for relevant alleles. Extreme high body mass index (BMI) was significantly negatively correlated with anorexia nervosa ( $r_g$ =-0.29, SE=0.08, p=2.0x10<sup>-4</sup>) as were obesity, BMI in the normal range, overweight, and hip circumference, with genetic correlations ranging from -0.2 to -0.3.

We also observed significant negative genetic correlations between anorexia nervosa and insulin and glucose related traits—e.g., exceeding those of BMI for both insulin resistance (HOMA-IR)  $(r_q=-0.50, SE=0.11, p=1.3 \times 10^{-5})$  and fasting insulin  $(r_q=-0.41, SE=0.09, p=5.2 \times 10^{-6})$ ; as well as a similar correlation with fasting glucose ( $r_q$ =-0.26, SE=0.07, p=3.0x10<sup>-4</sup>). Although BMI corrected HOMA-IR GWAS statistics were not available genome-wide, additional analyses with the available BMI corrected GWAS results for related phenotypes suggest that this metabolic signal is at least partly independent of BMI with leptin levels ( $r_g = -0.24$ , SE=0.11, p=0.03). Regarding cholesterol and lipid measures, a distinction between different lipid fractions emerges when comparing high density lipoprotein (HDL), low density lipoprotein (LDL), and very low density lipoprotein (VLDL) phenotypes. Genetic correlations between anorexia nervosa and HDL phenotypes were positive: e.g., total cholesterol in large HDL particles ( $r_a$ =0.39, SE=0.12, p=1.6x10<sup>-3</sup>); free cholesterol in large HDL particles ( $r_g$ =0.37, SE=0.12, p=2.2x10<sup>-3</sup>); and phospholipids in large HDL particles ( $r_g$ =0.30, SE=0.11, p=6.7x10<sup>-3</sup>). In contrast, VLDL cholesterol phenotypes were negatively correlated with AN, albeit with nominal significance (i.e., uncorrected p<.05): e.g., total lipids in VLDL ( $r_q$ =-0.30, SE=0.12, p=0.01); phospholipids in VLDL ( $r_g$ =-0.33, SE=0.13, p=4.4x10<sup>-3</sup>); and LDL cholesterol ( $r_g$ =-0.20, SE=0.08, p=0.011).

### Discussion

To our knowledge, this is the first report of a genome-wide significant association for anorexia nervosa. As is typical of many GWAS loci for complex disorders, the region implicated is broad, with a modest odds ratio of 1.2 but at a common allele (MAF<sub>controls</sub>=0.44) (25). Our genome-wide  $h_{SNP}^2$  estimate of 20% for anorexia nervosa supports a substantial role for common genetic variation. As we now expect (26), the  $h_{SNP}^2$  estimate reported here indicates that common variants account for a sizeable portion of twin-based heritability ( $h_{Twin}^2$  48-74%)<sup>6</sup>. Further, these results fit with the expectation that  $h_{Twin}^2$  should exceed  $h_{SNP}^2$ , because the former captures the effects of all types of genetic variation (common and rare, as well as variation not captured with current methods).

The observed pattern of genetic correlations with psychiatric, personality, educational, and metabolic phenotypes provides grounds for broadening our conceptualization of the disorder. First, the strong positive genetic correlations of anorexia nervosa with obsessive-compulsive disorder and neuroticism reinforce clinical and epidemiological observations. AN is commonly comorbid with OCD and twin studies have reported high twin-based genetic correlations (27). High neuroticism in adolescence predicts subsequent onset of AN (1). In addition, anorexia nervosa is commonly comorbid with multiple anxiety phenotypes, which often pre-date the onset of anorexia nervosa (28).

Second, the positive genetic correlations seen with schizophrenia and the cross psychiatric disorder phenotype firmly anchor anorexia nervosa with other psychiatric disorders and reflect the substantial evidence for partially shared genetic risk across many psychiatric disorders (29). Third, congruent with our results, positive associations been anorexia nervosa and educational attainment have been reported (30) and have been conjectured to reflect greater internal and external demands for academic success in highly educated families. Our results, in contrast, suggest that genetic factors may partially account for these reported associations.

Fourth, the identification of significant negative correlations between anorexia nervosa and BMIrelated and anthropometric measures could potentially serve as an important first step toward gaining a better understanding of the shared biology underlying extremes of weight dysregulation (i.e., obesity vs. anorexia nervosa). This is of critical importance as adequate explanations for how individuals with anorexia nervosa reach, sustain, and revert to exceedingly low BMIs have been elusive. Clinically, one of the most perplexing features of anorexia nervosa, is how patients' bodies seem to revert rapidly to a "low set point" after renourishment, which may represent the biological inverse of the reversion to high set points commonly seen in the unsuccessful treatment of obesity (31, 32). As noted by Bulik-Sullivan et al. (23) and Hinney et al. (33), these observations extend our understanding that the same genetic factors that influence normal variation in BMI, body shape, and body composition may also influence extreme dysregulation of these weight-related features in anorexia nervosa. This pattern of observations complements prior strong evidence for the involvement of neural mechanisms in obesity (34). Finally, positive correlations with "favorable" metabolic phenotypes (i.e., HDL and lipid

measures) and negative correlations with "unfavorable" metabolic phenotypes (i.e., fasting insulin, fasting glucose, HOMA-IR) encourage additional exploration of the role metabolic factors may play in extreme dysregulation of appetite and weight in anorexia nervosa.

The genome-wide significant locus we identify to be associated with anorexia nervosa is broad and multigenic (chr12:56,372,585-56,482,185). Mechanistic explanations about the role of the associated variant require additional functional data; nevertheless, we note the possible role for genes at this locus in the pathophysiology of anorexia nervosa. *PA2G4* is involved in growth regulation and acts as a corepressor of the androgen receptor (35). *ESYT1* [extended synaptotagmin-1 which binds and transports lipids (36)] is enriched in the postsynaptic density, which is implicated in the etiology of schizophrenia (37). Perhaps more convincing is that the sentinel marker for this locus, rs4622308, is in high LD with a known GWAS hit for type 1 diabetes (20), and rheumatoid arthritis (21), and the region around it harbors multiple other autoimmune associations. Multiple reports of shared effects between anorexia nervosa and immune phenotypes fit into a broader pattern of above-chance comorbidity across psychiatric and immune phenotypes (38, 39). Evidence suggests that this shared risk is at least partly genetic in origin (23, 39). A negative genetic correlation between anorexia nervosa and rheumatoid arthritis was previously reported (23), and our LDSC estimate—though only nominally significant—is in the negative direction as well (see **Supplementary Table S5**).

The primary strength of this investigation is to have extended prior work by increasing sample size via collaboration. Importantly, our combined sample size remains modest given contemporary understanding of complex trait genetics. Moreover, since our collection represents all of the currently GWAS genotyped anorexia nervosa samples in the world, no known genotyped replication samples exist. As such, we expect this to be the beginning of genomic discovery in eating disorders (25). Future work with additional and better-powered anorexia nervosa GWAS will clarify the magnitude of genetic relationships among metabolic and psychiatric phenotypes and methods such as that proposed by Pickrell et al. (40) will provide clues about the direction of causal relationships.

In summary, we identified the first robust genome-wide significant locus for anorexia nervosa, which is also a previously reported type 1 diabetes and general autoimmune disorder locus. Perhaps of greater importance, is that we find anorexia nervosa is a complex heritable phenotype with intriguingly large and significant genetic correlations not only with psychiatric disorders but also multiple metabolic traits. This encourages a reconceptualization of this frequently lethal disorder as both psychiatric and metabolic. Just as obesity is increasingly considered to be both a metabolic/endocrine and psychiatric disorder, approaching anorexia nervosa as both a psychiatric and metabolic condition could ignite interest in developing or repositioning pharmacologic agents for its treatment where currently none exist.

# Eating Disorders Working Group of the Psychiatric Genomics Consortium

Laramie Duncan, PhD Zeynep Yilmaz, PhD Raymond Walters, PhD Jackie Goldstein, PhD Verneri Anttila, PhD Brendan Bulik-Sullivan, PhD Stephan Ripke, MD, PhD Roger Adan, PhD Lars Alfredsson, PhD Tetsuya Ando, MD, PhD Ole Andreassen, MD, PhD Harald Aschauer, MD Jessica Baker, PhD Jeffrey Barrett, PhD Vladimir Bencko, MD, PhD Andrew Bergen, PhD Wade Berrettini, MD, PhD Andreas Birgegård, PhD Claudette Boni, PhD Vesna Boraska Perica, PhD Harry Brandt, MD Roland Burghardt, MD Laura Carlberg, MD Matteo Cassina, MD Carolyn Cesta Sven Cichon, PhD Maurizio Clementi, MD Sarah Cohen-Woods, PhD Joni Coleman, MSc Roger Cone, PhD

Philippe Courtet, MD Steven Crawford, MD Scott Crow, MD Jim Crowley, PhD Unna Danner, PhD Oliver Davis, MSc, PhD Martina de Zwaan, MD George Dedoussis, PhD Daniela Degortes, PhD Janiece DeSocio, PhD, RN, PMHNP-BC Danielle Dick, PhD Dimitris Dikeos, MD Christian Dina, PhD Bo Ding, PhD Monika Dmitrzak-Weglarz, PhD Elisa Docampo, MD, PhD Karin Egberts, MD Stefan Ehrlich, MD Geòrgia Escaramís, PhD Tõnu Esko, PhD Thomas Espeseth, PhD Xavier Estivill, MD, PhD Angela Favaro, MD, PhD Fernando Fernández-Aranda, PhD, FAED Manfred Fichter, MD, Dipl-Psych Chris Finan, PhD Krista Fischer, PhD James Floyd, PhD Manuel Föcker, MD Lenka Foretova, MD, PhD Monica Forzan, PhD Caroline Fox, MD Christopher Franklin, PhD Valerie Gaborieau Steven Gallinger, MD Giovanni Gambaro, MD, PhD Héléna Gaspar, PhD Ina Giegling, PhD Fragiskos Gonidakis, MD Philip Gorwood, MD, PhD Monica Gratacos, MD, PhD Sébastien Guillaume, MD, PhD Yiran Guo, PhD Hakon Hakonarson, MD, PhD Katherine Halmi, MD Rebecca Harrison

Konstantinos Hatzikotoulas, MD, PhD Joanna Hauser, MD, PhD Johannes Hebebrand, MD Sietske Helder, PhD Judith Hendriks, BSc Stefan Herms, PhD Beate Herpertz-Dahlmann, MD Wolfgang Herzog, MD Christopher Hilliard, BS Laura Huckins, PhD James Hudson, MD, ScD Julia Huemer, MD Hartmut Imgart, MD Hidetoshi Inoko, PhD Sigrid Jall Stephane Jamain, PhD Vladimir Janout, PhD Susana Jiménez-Murcia, PhD Craig Johnson, PhD Jenny Jordan, PhD Antonio Julià, PhD Anders Juréus, PhD Gursharan Kalsi, PhD Allan Kaplan, MSc, MD, FRCP(C) Jaakko Kaprio, MD, PhD Leila Karhunen, PhD Andreas Karwautz, MD, FAED Martien Kas, PhD Walter Kaye, MD Martin Kennedy, PhD James Kennedy, MD, FRCP(C) Anna Keski-Rahkonen, MD, PhD, MPH Kirsty Kiezebrink, BSc (Hons), PGDip, PhD, FHEA, RNutr Youl-Ri Kim, MD, PhD Lars Klareskog, MD Kelly Klump, PhD Gun Peggy Knudsen, PhD Bobby Koeleman, PhD Doris Koubek, MD Maria La Via, MD Mikael Landén, MD, PhD Stephanie Le Hellard, PhD Marion Leboyer. MD, PhD Robert Levitan, MD Dong Li, PhD Paul Lichtenstein, PhD

Lisa Lilenfeld, PhD Jolanta Lissowska, PhD Astri Lundervold, PhD Pierre Magistretti, PhD Mario Maj, MD, PhD Katrin Mannik, PhD Sara Marsal, MD, PhD Debora Kaminska, PhD Nicholas Martin, PhD Morten Mattingsdal, PhD Sara McDevitt, MB, MD, MRCPsych, MMedED Peter McGuffin, MD Elisabeth Merl, MD Andres Metspalu, PhD, MD Ingrid Meulenbelt, PhD Nadia Micali, MD, PhD James Mitchell, MD Karen Mitchell, PhD Palmiero Monteleone, MD Alessio Maria Monteleone, MD Grant Montgomery, PhD Preben Mortensen, MD, DrMedSc Melissa Munn-Chernoff, PhD Timo Müller, PhD Benedetta Nacmias, PhD Marie Navratilova, MUDr., PhD Ida Nilsson, PhD Claes Norring, PhD Ioanna Ntalla, PhD Roel Ophoff, PhD Julie O'Toole, MD Aarno Palotie, MD, PhD Jacques Pantel, PhD Hana Papezova, MD, PhD **Richard Parker** Dalila Pinto, PhD Raquel Rabionet, PhD Anu Raevuori, MD, PhD Andrzej Rajewski, MD, PhD Nicolas Ramoz, PhD N. William Rayner, PhD Ted Reichborn-Kjennerud, MD Valdo Ricca, MD Samuli Ripatti, PhD Franziska Ritschel, MSc Marion Roberts, PhD

Alessandro Rotondo, MD Dan Rujescu, MD Filip Rybakowski, MD, PhD Paolo Santonastaso, MD André Scherag, PhD Stephen Scherer, PhD, FRSC Ulrike Schmidt, MD, PhD Nicholas Schork, PhD Alexandra Schosser, PhD Laura Scott, PhD Jochen Seitz, MD Lenka Slachtova, PhD Robert Sladek, MD P. Eline Slagboom, PhD Margarita Slof-Op 't Landt, PhD Agnieszka Slopien, MD Tosha Smith, PhD Nicole Soranzo, PhD Sandro Sorbi, MD Lorraine Southam, BSc Vidar Steen, MD, PhD Eric Strengman, BS Michael Strober, PhD Jin Szatkiewicz, PhD Neonila Szeszenia-Dabrowska, MD, PhD Ioanna Tachmazidou, PhD Elena Tenconi, MD Alfonso Tortorella, MD Federica Tozzi, MD Janet Treasure, PhD, FRCP, FRCPsych Matthias Tschöp, MD Artemis Tsitsika, MD, PhD Konstantinos Tziouvas, MD, MSc Annemarie van Elburg, MD, PhD Eric van Furth, PhD Tracey Wade, PhD Gudrun Wagner, Dr, MSc, DPO Esther Walton, Dr. rer. nat., PhD Hunna Watson, PhD H-Erich Wichmann, PhD Elisabeth Widen, MD, PhD D. Blake Woodside, MD Jack Yanovski, MD, PhD Shuyang Yao, MSc, BSc Stephanie Zerwas, PhD Stephan Zipfel, MD

Laura Thornton, PhD Anke Hinney, PhD Gerome Breen, PhD Cynthia M. Bulik, PhD

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## URLs

SNP results, https://www.med.unc.edu/pgc.

## **Acknowledgments**

We are grateful to the participants in these studies, without whom this work would not have been possible. We thank all study coordinators, volunteers, and research staff that enabled this work, the Price Family Collaborative Group (PFCG), and the computational infrastructure provided by the Psychiatric Genomics Consortium. Children's Hospital of Philadelphia/Price Foundation acknowledgements are as follows: We thank all patients and families enrolled in the study, as well as all healthy control who donated blood samples to Children's Hospital of Philadelphia (CHOP) for genetic research purposes. We thank the Price Foundation for their support of recruiting patients, collecting clinical information and providing DNA samples used in this study. We also thank the Klarman Family Foundation for supporting the study. We thank the technical staff at the Center for Applied Genomics (CAG) at CHOP for generating genotypes used for analyses and the nursing, medical assistant and medical staff for their invaluable

assistance with sample collection. Data on glycemic traits have been contributed by MAGIC investigators and have been downloaded from www.magicinvestigators.org

## Funding

Funding was provided to fifty-six investigators contributing to this report by the organizations noted below in the format of 'Investigator 1: funding source(s); Investigator 2: funding source(s); ...'. The views expressed are those of the authors and not necessarily those of the NHS, the NIHR, the Department of Health, NHSBT, or any of the other funders noted below. Laramie Duncan: NIMH 5U01MH094432-04, NIMH 3U01MH094432-03S1; Stephan Ripke: NIMH MH109528, NARSAD 23545; Mark Daly: NIMH MH109528; Youl-Ri Kim: Research of Korea Centers for Disease Control and Prevention Fund (code# HD16A1351); Susana Jiménez-Murcia: Instituto de Salud Carlos III (FIS PI14/290 and CIBERobn); Fernando Fernández-Aranda: Instituto de Salud Carlos III (FIS PI14/290 and CIBERobn); Philip Gorwood: EC Framework V 'Factors in Healthy Eating' from INRA/INSERM (4M406D), PHRC ENDANO (2008-A01636-49); Paolo Santonastaso: (for PAUDA Group) Veneto Region Grant BIOVEDA, Contract grant number: DGR 3984/08; Leila Karhunen: Academy of Finland (28327); Anu Raevuori: Academy of Finland grant number 259764; André Scherag: Federal Ministry of Education and Research (BMBF), Germany, FKZ 01EO1502; Andrew Bergen: Professional Services Agreement with the Regents of the University of California; Stephanie Le Hellard: Bergen Research Foundation, NFR (NORMENT-SFF), K.G. Jebsen Foundation, NCNG, the University of Bergen, Dr. Einar Martens Fund, the Research Council of Norway, to SLH, VMS and TE; Hakon Hakonarson: Institutional Development Fund to Center for Applied Genomics from CHOP, Electronic Medical Records and Genomics (eMERGE) Network (U01 HG006830) from National Human

Genome Research Institute of National Institutes of Health, Kurbert Family; Dong Li: Davis Foundation Postdoctoral Fellowship Program in Eating Disorders Research Award; Yiran Guo: Davis Foundation Postdoctoral Fellowship Program in Eating Disorders Research Award; Shuyang Yao: China Scholarship Council; Sietske Helder: European Commission (2008-2011) Early Stage Researcher from the Research Training Network INTACT (Individually Tailored Stepped Care for Women with Eating Disorders) in the Marie Curie Program (MRTN-CT-2006-035988); Stephen Scherer: Genome Canada, the government of Ontario, the Canadian Institutes of Health Research, University of Toronto McLaughlin Centre; Martina de Zwaan: German Federal Ministry for Education and Research (BMBF) 01GV0601 and 01GV0624; Anke Hinney: German Ministry for Education and Research (National Genome Research Net-Plus 01GS0820) and the German Research Foundation (DFG; HI865/2-1); Tracey Wade: Grants 324715 and 480420 from the National Health and Medical Research Council (NHMRC), Australian Twin Registry supported by Enabling Grant (ID 310667) from the NHMRC (University of Melbourne); Hana Papezova: Internal Grant Agency of the Ministry of Health of the Czech Republic IGA MZ ČR NT 14094-3/2013; Karen Mitchell: K01MH093750; Jessica Baker: K01MH106675; Zeynep Yilmaz: K01MH109782; Danielle Dick: K02AA018755-06, R01AA015416-08, National Institute on Alcohol Abuse and Alcoholism (NIAAA); Mikael Landén: Klarman Family Foundation; Sarah Cohen-Woods: Matthew Flinders Fellowship, Flinders University, Australia; Matthias Tschöp: Alexander von Humboldt Foundation, Helmholtz Alliance ICEMED-Imaging and Curing Environmental Metabolic Diseases through the Initiative and Networking Fund of the Helmholtz Association, the Helmholtz cross-program topic "Metabolic Dysfunction", Deutsche Forschungsgemeinschaft (DFG-TS226/1-1 and TS226/3-1, European Research Council Consolidator Grant (HepatpMetaboPath); Beate

Herpertz-Dahlmann: Ministry for Research and Education, Germany; Nicole Soranzo: Wellcome Trust (Grant Codes WT098051 and WT091310), EU FP7 (EPIGENESYS Grant Code 257082 and BLUEPRINT Grant Code HEALTH-F5-2011-282510), National Institute for Health Research Blood and Transplant Research Unit (NIHR BTRU) in Donor Health and Genomics at the University of Cambridge in partnership with NHS Blood and Transplant (NHSBT); Allan Kaplan: Ontario Mental Health Foundation, Ministry of Health of Ontario AFP Innovation Fund; Wade Berrettini: Price Foundation; Marion Roberts: Psychiatry Research Trust (registered charity no. 284286); Patrick Sullivan: R01 MH109528, D0886501, Swedish Research Council; Thomas Espeseth: Research Council of Norway (RCN), and South-East Norway Regional Health Authority (SEN); Michael Strober: Resnick Family Chair in Eating Disorders; Xavier Estivill: Spanish Ministry of Economy and Competitiveness (MINECO) no. SAF2013-49108-R, the Generalitat de Catalunya AGAUR 2014 SGR-1138, the European Commission 7th Framework Program (FP7/2007-2013) 262055 (ESGI); Lenka Foretova: MH CZ - DRO (MMCI, 00209805); Ole Andreassen: Research Council of Norway (#248778, # 223273), KG Jebsen Foundation; Timo Müller : Helmholtz Alliance ICEMED-Imaging and Curing Environmental Metabolic Diseases through the Initiative and Networking Fund of the Helmholtz Association, Helmholtz cross-program topic "Metabolic Dysfunction", Deutsche Forschungsgemeinschaft (DFG-TS226/1-1 and TS226/3-1); Cynthia Bulik: The Klarman Family Foundation, Wellcome Trust WT088827/Z/09, Swedish Research Council (VR Dnr: 538-2013-8864); Preben Mortensen: The Klarman Family Foundation, unrestricted grant from the Lundbeck Foundation, iPSYCH (Initiative for Integrative Psychiatric Research), Aarhus University for CIRRAU (Centre of Integrated Register-Based Research); Walter Kaye: The Price Foundation, NIMH R01 MH092793; Ted Reichborn-Kjennerud: Norwegian Research Council, Norwegian Foundation

for Health and Rehabilitation; Eleftheria Zeggini: The Wellcome Trust (WT098051); Martien Kas: ZonMW VIDI Grant (91786327) from The Netherlands Organization for Scientific Research (NWO); Tonu Esko: EU H2020 grants 692145, 676550, 654248, Estonian Research Council Grant IUT20-60, NIASC, EIT – Health and NIH-BMI Grant No: 2R01DK075787-06A1 and EU through the European Regional Development Fund (Project No. 2014-2020.4.01.15-0012 GENTRANSMED; Stefan Ehrlich: Deutsche Forschungsgemeinschaft (EH 367/5-1 and SFB 940), Swiss Anorexia Nervosa Foundation; Esther Walton: Deutsche Forschungsgemeinschaft (EH 367/5-1 and SFB 940), Swiss Anorexia Nervosa Foundation; Ulrike Schmidt: National Institute of Health Research Mental Health Biomedical Research Centre at South London and Maudsley National Health Service Foundation Trust and King's College London ; Martin Kennedy: University of Otago Research Grant;

Johannes Hebebrand: German Ministry for Education and Research (National Genome Research Net-Plus 01GS0820 and 01KU0903), German Research Foundation (DFG; HI865/2-1), European Community's Seventh Framework Programme (FP7/2007-2013) under grant agreement no. 245009 and no.262055; Stephan Zipfel: German Ministry for Education and Research (ANTOP-study project, number 01GV0624); Gerome Breen: National Institute for Health Research (NIHR) Biomedical Research Centre at South London, Maudsley NHS Foundation Trust and King's College London

## **Competing Financial Interests**

C.B. is grant recipient from and consultant to Shire. G.B. has received grant funding and consultancy fees from Eli Lilly. D.D. is speaker, consultant, or on advisory boards of various Pharmaceutical Companies including: AstraZeneca, Boehringer, Bristol Myers Squibb, Eli Lilly,

Genesis Pharma, GlaxoSmithKline, Janssen, Lundbeck, Organon, Sanofi, UniPharma, and Wyeth, and he has unrestricted grants from Lilly and AstraZeneca as director of the Sleep Research Unit of Eginition Hospital (National and Kapodistrian University of Athens, Greece). A.K. is on the Shire Canada BED Advisory Board. J.K. is a member of SAB of AssurexHealth Inc (unpaid). M.L. has received lecture honoraria from Lundbeck, AstraZeneca, and Biophausia Sweden, and served as scientific consultant for EPID Research Oy. No other equity ownership, profit-sharing agreements, royalties, or patents. P.S. is scientific advisor to Pfizer, Inc. J.T. received an honorarium for speaking at a diabetic conference for Lilly and royalties from a published book.

# Author contributions - See Supplementary Table S6

# Figure 1. Manhattan and regional plot of the genome-wide significant locus for anorexia

**nervosa.** A. Manhattan plot depicts a genome-wide significant locus on chromosome 12. The threshold for significance (see y-axis) is 7.3, which is  $-log_{10}(5x10^{-8})$ . B. Regional LOCUSZOOM plot of the top locus reveals numerous genes in the region. Results depicted here reflect the full meta-analysis. Per text, see Supplementary Figure 1 for area plot with phenotypic associations. The right axis gives recombination rate, depicted with a light blue line.







Figure 2. Genetic correlations between anorexia nervosa and diverse phenotypes reveal overlap across psychiatric, educational, weight, insulin, lipoprotein, and cholesterol phenotypes. The 24 correlations depicted here (of 159 phenotypes tested) have FDR<0.05. Bars are  $\pm$  standard error.

