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## Similarity in mycorrhizal communities associating with two widespread terrestrial orchids decays with distance — [Source link](#)

[Xiaoke Xing](#), [Gao Yue](#), [Zeyu Zhao](#), [Michael Waud](#) ...+7 more authors




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1 **Similarity in mycorrhizal communities associating with two**  
2 **widespread terrestrial orchids decays with distance**

3  
4 Xiaoke Xing<sup>1#\*</sup>, Yue Gao<sup>1#</sup>, Zeyu Zhao<sup>1</sup>, Michael Waud<sup>2</sup>, Karl J. Duffy<sup>3</sup>, Marc-André  
5 Selosse<sup>4,5</sup>, Marcin Jakalski<sup>5</sup>, Na Liu<sup>1</sup>, Hans Jacquemyn<sup>2</sup>, Shunxing Guo<sup>1</sup>

6 <sup>1</sup>*Key Laboratory of Bioactive Substances and Resources Utilization of Chinese Herbal*  
7 *Medicine, Ministry of Education, Institute of Medicinal Plant Development, Chinese*  
8 *Academy of Medical Sciences and Peking Union Medical College, Beijing 100193,*  
9 *China.*

10 <sup>2</sup>*KU Leuven, Department of Biology, Plant Conservation and Population Biology, B-*  
11 *3001 Leuven, Belgium*

12 <sup>3</sup>*Department of Biology, University of Naples Federico II, via Cinthia, Naples 80026,*  
13 *Italy.*

14 <sup>4</sup>*Institut de Systématique, Évolution, Biodiversité (ISYEB – UMR 7205 – CNRS,*  
15 *MNHN, UPMC, EPHE), Muséum national d’Histoire naturelle, Sorbonne Universités,*  
16 *57 rue Cuvier, 75005, Paris, France.*

17 <sup>5</sup>*University of Gdańsk, Faculty of Biology, ul. Wita Stwosza 59, 80-308 Gdańsk, Poland.*

18  
19 # Shared first authorship

20  
21 **Corresponding author: Xiaoke Xing**

22 E-mail: xkxing2009@hotmail.com Tel: 86-10-57833242

23 **Abstract**

24 **Aim:** Interactions with mycorrhizal fungi are increasingly recognized as an important  
25 factor underlying the distribution and abundance of orchid species. However, the  
26 geographic distribution of orchid mycorrhizal fungi (OMF) and how their communities  
27 vary over large geographical areas are less well understood. Because climatic and  
28 environmental similarity may decrease with geographical distance or because some  
29 OMF have limited dispersal capabilities, similarities in orchid mycorrhizal  
30 communities can be expected to decrease with increasing distances separating orchid  
31 populations. However, up till now empirical evidence is largely lacking.

32 **Location:** Eurasia

33 **Taxa:** *Gymnadenia conopsea* (L.) R. Brown and *Epipactis helleborine* (L.) Crantz

34 **Methods:** High-throughput sequencing was used to perform a cross-continental  
35 comparison of OMF that associate with two widespread Eurasian terrestrial orchids,  
36 *Epipactis helleborine* and *Gymnadenia conopsea*. Both phylogenetic and non-  
37 phylogenetic measures of community dissimilarity and their components were  
38 calculated and related to geographic distances using Mantel tests.

39 **Results:** Our results showed that in both orchid species similarity in mycorrhizal  
40 communities decreased significantly with geographical distance. Decomposing the  
41 contribution of spatial turnover and nestedness to overall dissimilarity showed that the  
42 observed dissimilarity was mainly the result of species replacement between regions,  
43 and not of species loss. Similarly, a strong relationship was observed between  
44 phylogenetic community dissimilarity and geographic distance. Decomposing PCD

45 values into a nonphylogenetic and phylogenetic component showed that orchid  
46 populations located closely next to each other were likely to contain the same OTUs,  
47 but that the non-shared taxa came from different phylogenetic clades. Species indicator  
48 analyses showed that the majority of OMF OTUs were restricted to particular  
49 geographic areas. However, some OTUs occurred in both continents, indicating that  
50 some fungi have very wide distributions.

51 **Main conclusions:** Overall, these results demonstrate that orchid mycorrhizal  
52 communities differ substantially across large geographic areas, but that the distribution  
53 of orchids is not necessarily restricted by the distribution of particular OMF. Hence,  
54 widespread orchid species can be considered mycorrhizal generalists that are flexible  
55 in the OMF with which they associate across large geographic areas.

56

57 **KEYWORDS** *Epipactis*, fungal community, *Gymnadenia*, mycorrhizal specificity,  
58 orchid mycorrhiza, spatial turnover

## 59 **1 INTRODUCTION**

60 Mycorrhizal symbioses have been considered as one of the most important symbiotic  
61 association in terrestrial ecosystems (van der Heijden et al., 2015). In this mutualism,  
62 the soil fungus contributes mineral nutrition and water to the plant that, in turn,  
63 contributes photosynthetically fixed carbon back to the fungus, by way of a dual organ  
64 made of roots colonized by fungal hyphae, the mycorrhiza (Smith & Read, 2008).  
65 Whether a given species is a specialist or generalist largely depends on its ability to  
66 associate with a large number of partners and whether its partners have a narrow or a  
67 broad geographical range. Many studies have shown that plants are often mycorrhizal  
68 generalists (Smith & Read, 2008), in that they can interact with many taxonomically  
69 disparate mycorrhizal taxa. Conversely, there are also cases of plants that are  
70 mycorrhizal specialists (van der Heijden et al., 2015), although the precise factors  
71 leading to specialist or generalist interactions are not well understood (Shefferson et al.,  
72 2019). Interacting with a broad range of partners may increase niche availability and  
73 allow survival in a large diversity of environments (Batstone et al., 2018).

74       Since the early discoveries of Noël Bernard (1899; see Selosse et al., 2017), it is  
75 widely accepted that orchid species are dependent on mycorrhizal fungi during the early  
76 stages of plant development (Rasmussen & Rasmussen 2009; Dearnaley et al., 2016).  
77 Most orchid species maintain associations with mycorrhizal fungi into adulthood as  
78 well (Cameron et al., 2006; Rasmussen & Rasmussen, 2009; Waterman et al., 2011).  
79 The fungi that form mycorrhizas with green orchids usually are members of the  
80 Tulasnellaceae, Ceratobasidiaceae and Serendipitaceae (Rasmussen, 1995; Smith &

81 Read, 2008; Dearnaley et al., 2012; Jacquemyn et al., 2017b). However, recent research  
82 has indicated that many orchid species, including photosynthetic orchids,  
83 simultaneously associate with a large diversity of ectomycorrhizal fungi (i.e. fungi  
84 usually found as mycorrhizal on tree species) from the Thelephoraceae, Sebacinaceae,  
85 Inocybaceae, and Tuberaceae (Waterman et al., 2011, Kottke et al., 2010; Zhang et al.,  
86 2012; Yagame et al., 2013; Jacquemyn et al., 2015; Waud et al., 2016). Although the  
87 ecological function and relevance of these fungi still has to be elucidated (Jacquemyn  
88 & Merckx, 2019), the available knowledge suggests that at least in some photosynthetic  
89 species their presence has an ecological function (Jacquemyn et al., 2017a), providing  
90 further support for recent claims that fungi may have more complex niches than  
91 previously assumed (Selosse et al., 2018). Therefore, orchids may harbor a large fungal  
92 diversity, with broadly distributed orchids having the possibility to sample diverse  
93 partners over large geographical ranges.

94       Although mycorrhizal dependency has been increasingly recognized as an  
95 important factor influencing both the distribution and abundance of orchid populations  
96 (McCormick & Jacquemyn, 2014; McCormick et al., 2018), at present little is known  
97 about the geographic distribution of orchid mycorrhizal fungi (OMF) (reviewed in  
98 Jacquemyn et al., 2017b). However, the widespread occurrence of orchids across the  
99 globe and in diverse ecosystems (Givnish et al., 2016) suggests that the OMF that are  
100 necessary for germination and seedling establishment are also widespread and not  
101 necessarily restricted to geographical regions. A major caveat in our current  
102 understanding of the biogeographical distribution of OMF is that most of the available

103 data are very fragmentary and that often only a few populations are sampled within a  
104 restricted geographic area, making it difficult to draw any general conclusions about  
105 the distribution of fungi associated with orchids across larger scales (Jacquemyn et al.,  
106 2017b).

107 The few available studies (Taylor et al., 2004; Selosse et al., 2002; Girlanda et al.,  
108 2006; Irwin et al., 2007; Otero et al., 2007; Roy et al., 2009; Davis et al., 2015;  
109 Těšitelová et al., 2015; Duffy et al., 2019) that have attempted to sample the large-scale  
110 distribution of mycorrhizal fungi associating with a particular orchid species have  
111 shown that the wide distribution of some orchid species may to some extent be  
112 explained by the widespread occurrence of its mycorrhizae. For example, Davis et al.  
113 (2015) showed that the Australian orchid *Pheladenia deformis* associates with one or  
114 two *Sebacina* sp., but that these fungi have a widespread distribution across the  
115 Australian continent. Because many orchid species show more generalist interactions  
116 and associate with several different fungi (Selosse et al., 2002; Girlanda et al., 2006;  
117 Roy et al., 2009; Jacquemyn et al., 2012) and because soil fungal communities can vary  
118 strongly in space (Talbot et al., 2014), this possibly leads to turnover in mycorrhizal  
119 partners across large geographic areas and a significant decrease in similarity in  
120 mycorrhizal communities with increasing distance, i.e. so-called distance decay of  
121 similarity (Nekola & White, 1999; Soininen et al., 2007; Talbot et al., 2014). Indeed,  
122 recent research has already indicated that OMF diversity decreases with increasing  
123 latitude (Duffy et al., 2019), and that the community composition of OMF varies  
124 according to habitat (Jacquemyn et al., 2016a).

125 To improve our knowledge about the geographic distribution of orchid mycorrhizal  
126 fungi, we performed a cross-continental, Eurasian comparison of the mycorrhizal  
127 communities associating with the roots of two widespread terrestrial orchids,  
128 *Gymnadenia conopsea* and *Epipactis helleborine*. Given that both species are  
129 mycorrhizal generalists that associate with a wide number of mycorrhizal fungi  
130 belonging to different fungal families (Jacquemyn et al., 2016b; Stark et al., 2009;  
131 Těšitelová et al., 2013; Waud et al., 2016; Schweiger et al., 2018), we hypothesized that  
132 the fungal communities associated with *G. conopsea* and *E. helleborine* show large  
133 geographic variation, leading to a decay of similarity in mycorrhizal communities with  
134 increasing distance. To better understand the causality of the processes underlying  
135 variation in OMF diversity, we decomposed the overall dissimilarity into two additive  
136 components that account for species replacement and species loss, respectively  
137 (Baselga, 2010).

138

## 139 **2 MATERIALS AND METHODS**

### 140 **2.1 Study species**

141 *Gymnadenia conopsea* (L.) R. Brown is a terrestrial, photosynthetic orchid that is  
142 widely distributed across Europe and Asia. Populations have been reported in Anatolia,  
143 the Caucasus, the Urals, Siberia and the Far East, including Japan, Korea and China  
144 (Meekers et al., 2012). It is one out of five species of *Gymnadenia* that occur in China,  
145 three of which are endemic. *Gymnadenia conopsea* can be found in a wide range of  
146 habitats, including forests, grasslands, and waterlogged meadows at altitudes varying



147 between 0 and 4700 m throughout Europe and temperate and subtropical zones of Asia  
148 (Meekers et al., 2012). In China, *G. conopsea* occurs mainly in the provinces Sichuan,  
149 Qinghai, Gansu, Tibet, Hebei, Shaanxi and Inner Mongolia. With the overexploitation  
150 of *G. conopsea* for traditional medicine as well as over-grazing and habitat destruction,  
151 natural populations of *G. conopsea* have declined rapidly in China. Currently, *G.*  
152 *conopsea* has been listed in the grade II section of endangered species in 2000 (Gesang  
153 & Gesang, 2010).

154 *Epipactis helleborine* (L.) Crantz occurs throughout large parts of Eurasia and North  
155 Africa (Delforge, 1995). *Epipactis helleborine* occurs in a broad range of habitat types,  
156 including dense forest floors, urban areas, open grasslands with scattered trees and  
157 calcareous soils from temperate to boreal zones (Salmia, 1986; Buttler, 1991; Delforge,  
158 1995; Hollingsworth & Dickson, 1997). In North America, *E. helleborine* has become  
159 a rapidly spreading species after it was introduced about 150 years ago (Owen, 1879,  
160 Soper & Garay, 1954) and it is currently considered as invasive.

161 Previous studies on the mycorrhizal fungi associating with *G. conopsea* (Stark et al.,  
162 2009; Těšitelová et al., 2013; Waud et al., 2016; Schweiger et al., 2018) and *E.*  
163 *helleborine* (Bidartondo et al., 2004; Ogura-Tsujita & Yukawa, 2008; Jacquemyn et al.,  
164 2016b) have shown that both species are mycorrhizal generalists that associate with a  
165 wide range of mycorrhizal fungi, including a dominance of ectomycorrhizal fungi in  
166 the second species. However, to the best of our knowledge, the mycorrhizal associates  
167 of these two orchid species in China remain unknown.

168

## 169 **2.2 Sampling**

170 *G. conopsea* and *E. helleborine* samples were collected from both Europe and China in  
171 July and August 2018 (Figure 1a,b). For *G. conopsea*, four populations (GITA, GITB,  
172 GBEA, GPLA) were collected from three countries of Europe (Italy, Belgium and  
173 Poland), while six populations (GB, GC, GG, GN, GS, GZ) were sampled in China  
174 (Figure 1a, Table 1). For *E. helleborine* samples, seven populations (EITA, EITB,  
175 EBEA, EBEB, EBEC, EBED, EPLA) growing in three countries of Europe (Italy,  
176 Belgium and Poland) and seven populations (EJL, ESX, ENJA, ENJB, ENJC, ENX,  
177 EBLGZ) growing in three provinces of China (Jilin, Shanxi and Yunnan) were collected  
178 respectively (Figure 1b, Table 1). For each population, 5 individual plants were  
179 randomly selected and 4 root fragments (3-5 cm) from each individual plant were  
180 collected. Slight yellowish or opaque roots, a typical feature of OMF infection, were  
181 selected, and surface cleaned several times with sterile water to minimize the detection  
182 of soil fungi and microscopically checked for mycorrhizal colonization. Roots were  
183 stored at -80°C prior to molecular analyses of mycorrhizal associates.

184

## 185 **2.3 Molecular analyses**

186 For DNA extraction, three pieces of colonized roots (2 cm long) were used per plant  
187 individual. Genomic DNA was extracted using the E.Z.N.A.® plant DNA Kit (Omega  
188 Bio-tek, Norcross, GA, U.S.) according to the manufacturer's instructions. To amplify  
189 the fungal internal transcribed spacer 2 (ITS2) region of fungi associated with *E.*  
190 *helleborine*, the fungal specific primer pair combination ITS86F (Turenne et al., 1999)

191 and ITS4 (White et al., 1990) was used, which has been used effectively for the  
192 detection of diverse mycobionts in previous studies (Jacquemyn et al., 2016b; Waud et  
193 al., 2016). The primer combination of ITS3 (White et al., 1990) and ITS4-OF (Taylor  
194 & McCormick, 2008) was used to amplify the ITS2 region of fungi associated with *G.*  
195 *conopsea* according to Waud et al. (2016). PCR reactions were performed in triplicate  
196 50µL mixture containing 5µL of 10 × Pyrobest Buffer, 4µL of 2.5mM dNTPs, 2 µL of  
197 each primer (10 µM), 0.3µL of Pyrobest DNA Polymerase (TaKaRa), and 30 ng of  
198 template DNA. The PCR program was as follows 95 °C for 5 min, 30 cycles at 95 °C  
199 for 30s, 56 °C for 30s, and 72 °C for 40s with a final extension of 72 °C for 10min.  
200 Amplicons were extracted from 2% agarose gels and purified using the AxyPrep DNA  
201 Gel Extraction Kit (Axygen Biosciences, Union City, CA, U.S.) according to the  
202 manufacturer's instructions and quantified using QuantiFluor™ -ST (Promega,  
203 U.S.).The purified amplicon mixture was subjected to high-throughput sequencing by  
204 Beijing Allwegene Tech, Ltd (Beijing, China) using the Illumina Miseq PE300  
205 sequencing platform (Illumina, Inc., CA, USA) that generated 300 bp long paired-end  
206 reads.

207

#### 208 **2.4 Data processing and OTU delimitation**

209 The extraction of high-quality sequences was firstly performed with the QIIME  
210 package (Quantitative Insights Into Microbial Ecology; v1.2.1). Raw sequences were  
211 selected based on sequence length, quality, primer and tag, wherein sequence quality  
212 was evaluated and enforced according to the following criteria. The raw sequences were

213 selected and the low-quality sequences were removed: (i) raw reads shorter than 110  
214 nucleotides were removed, (ii) the 300 bp reads were truncated at any site receiving an  
215 average quality score <20 over a 50 bp sliding window, discarding the truncated reads  
216 that were shorter than 50bp, (iii) exact barcode matching, 2 nucleotide mismatch in  
217 primer matching, reads containing ambiguous characters were removed, (iv) only  
218 sequences that overlap longer than 10 bp were assembled according to their overlap  
219 sequence. Reads that could not be assembled were discarded.

220 The unique sequence set was classified into operational taxonomic units (OTUs)  
221 under the threshold of 97% identity using UCLUST (Edgar, 2010). Chimeric sequences  
222 were identified and removed using USEARCH (version 10.1). The taxonomy of each  
223 representative ITS sequence was analyzed by UCLUST against the UNITE database  
224 using confidence threshold of 90%. To minimize the risk of retaining sequences that  
225 resulted from sequencing errors, global singletons or global doubletons (OTUs  
226 represented by only one or two sequence in the entire dataset) were removed as it has  
227 been shown that this improves the accuracy of diversity estimates (Ihrmark et al., 2012;  
228 Waud et al., 2014). Remaining OTUs were assigned taxonomic identities based on the  
229 BLAST (Altschul et al. 1990) results of the OTU representative sequences (selected by  
230 UPARSE) using the GenBank nucleotide (nt) and UNITE database (Edgar, 2013).

231

## 232 **2.5 Data analysis**

233 Prior to removal of OTUs known as non-mycorrhizal fungi, MOTHUR (Schloss et al.,  
234 2009) was used to generate rarefaction curves for each sample to estimate the overall

235 coverage of the fungal communities studied (Figure S1). The overall diversity and  
236 phylogenetic diversity of OMF detected in roots of *E. helleborine* (Figure S2) and *G.*  
237 *conopsea* (Figure S3) were examined, respectively. OTUs were manually screened for  
238 possible orchid associating mycorrhizal families based on the information of previously  
239 detected mycorrhizal fungi from the roots, germinating seeds and protocorms of various  
240 *Epipactis* species (Bidartondo et al., 2004; Selosse et al., 2004; Těšitelová et al. 2013,  
241 Jacquemyn et al., 2016b) (Table S3). Mycorrhizal symbionts of *G. conopsea* mainly  
242 belong to the fungal families Ceratobasidiaceae, Tulasnellaceae, Thelophoraceae,  
243 Serendipitaceae and Sebacinaceae (Table S1) (Stark et al., 2009, Těšitelová et al.,  
244 2013; Waud et al., 2016). Here we restricted our analysis to those fungal families known  
245 to associate with the two orchid species. Representative sequences for each mycorrhizal  
246 OTU were submitted to GenBank under the accession numbers: MN006041-  
247 MN006135 (*G. conopsea*) and MK955493-MK955537, MK956832-MK956905,  
248 MK959119-MK959180, MK961130-MK961204, MK962538-MK962579 and  
249 MK965742-MK965875 (*E. helleborine*).

250 To compare the phylogenetic diversity (PD) (Faith, 1992) of OMF between Europe  
251 and China, we first constructed a ML tree for all the mycorrhizal OTUs identified. The  
252 OTU sequences were aligned using Clustal X version 2.0 (Larkin et al., 2007). The best  
253 model of evolution was identified using the Akaike Information Criterion implemented  
254 in jModelTest 2 (Darriba et al., 2012). The GTR+G+I and K2+G+I models of evolution  
255 were identified as the best-fit models for the *E. helleborine* and *G. conopsea* data sets,  
256 respectively. The ML phylogeny was constructed with RAxML 7.2.8 (Stamatakis et al.,

257 2008). The phylogenetic distances between the OTUs from these trees were used to  
258 calculate PD of the OTUs associated with each individual orchid plant. All calculations  
259 were done using the software package ‘picante’ (Kembel et al., 2010) in R (R  
260 Development Team, 2016). Univariate analysis of variance (ANOVA) was used to test  
261 whether the number and PD of OTUs per plant associating with differed significantly  
262 between European and Asian samples.

263 To test the hypothesis that similarities in mycorrhizal communities decrease with  
264 increasing distance, we first calculated the distances between all sampled populations  
265 using Vincenty’s inverse solution (Vincenty, 1975), while Sørensen’s dissimilarity  
266 index ( $\beta_{\text{sor}}$ ) and phylogenetic community dissimilarity (PCD) were used to assess  
267 dissimilarity in mycorrhizal communities between sampled populations. For each pair  
268 of populations, Sørensen’s pairwise dissimilarity was calculated as  $\beta_{\text{sor}} = \frac{b+c}{2a+b+c}$ , with  
269  $a$  the number of fungal OTUs common to both populations,  $b$  the number of OTUs  
270 occurring in the first, but not in the second population and  $c$  the number of species  
271 occurring in the second but not in the first population. Phylogenetic community  
272 dissimilarity was calculated using the formulas outlined in Ives & Helmus (2010). PCD  
273 values  $<1$  indicate that mycorrhizal communities are more similar than randomly  
274 selected communities, whereas PCD values  $> 1$  denote mycorrhizal communities that  
275 are more dissimilar than random communities sampled from the total species pool (Ives  
276 & Helmus, 2010). To test for a significant relationship between geographic distances  
277 and community dissimilarities, Mantel tests (Mantel, 1967) were performed using a  
278 total of 9,999 random permutations.

279 To get better insights into the precise factors determining dissimilarities, we  
280 further decomposed the Sørensen dissimilarity measures into components that assess  
281 the contribution of spatial turnover ( $\beta_{\text{sim}} = \frac{\min(b,c)}{a+\min(b,c)}$ ) and nestedness ( $\beta_{\text{nes}} =$   
282  $\frac{\max(b,c)-\min(b,c)}{2a+\min(b,c)+\max(b,c)} \times \frac{a}{a+\min(b,c)}$ ) (Baselga, 2010). Mantel tests were used again to  
283 assess the relationship between geographic distances and similarities derived from  
284 spatial turnover and nestedness. For both species, we also calculated the overall  
285 multiple-site dissimilarities ( $\beta_{\text{sor}}$ ) and its individual components ( $\beta_{\text{sim}}$  and  $\beta_{\text{nes}}$ ). Further,  
286 PCD values were partitioned into a nonphylogenetic component that reflects shared  
287 species between communities (PCDc) and a phylogenetic component that reflects the  
288 evolutionary relationships among non-shared species (PCDp) (Ives & Helmus, 2010).  
289 We then used Mantel tests to see whether PCDc and PCDp values were significantly  
290 related to the geographic distance separating communities. All calculations were  
291 performed using the R packages betapart (Baselga et al., 2018) and picante (Kembel et  
292 al., 2010).

293 Finally, Species Indicator Analyses were performed to examine whether certain  
294 OTUs were characteristic for a given geographic region. We used the multipatt function  
295 in the R package indicpecies (De Cáceres, Legendre & Moretti, 2010) to define  
296 indicator OTUs of both orchid species to a particular geographic region (Europe vs.  
297 Asia).

## 298 **3 RESULTS**

### 299 **3.1 Fungal OTUs**

300 In total, Illumina Miseq PE300 sequencing generated 1198133 (1943 OTUs) and  
301 848653 (1176 OTUs) fungal sequences for *E. helleborine* and *G. conopsea*,  
302 respectively. After analysis, 98.8% of the total number of sequences in *E. helleborine*  
303 (1183757 sequences, 1858 OTUs) and 95.5% of the total number of sequences in *G.*  
304 *conopsea* (810902 sequences, 1052 OTUs) could be assigned to Ascomycota and  
305 Basidiomycota. Rarefaction curves showed that the number of OTUs was relatively  
306 close to saturation for each individual plant (Figure S1).

307 ***G. conopsea*** - The most abundant fungi detected in *G. conopsea* belonged to the  
308 fungal families of Ceratobasidiaceae (42 OTUs), Tulasnellaceae (9 OTUs) and  
309 Serendipitaceae (9 OTUs), as well as to the ectomycorrhizal taxa Thelephoraceae (19  
310 OTUs), Inocybaceae (16 OTUs), Sebacinaceae (16 OTUs), Russulaceae (14 OTUs) and  
311 Tuberaceae (6 OTUs). In addition, a large number of other fungal taxa were also  
312 detected, including Cantharellaceae (6 OTUs), Tricholomataceae (17 OTUs),  
313 Tremellaceae (7 OTUs), Nectriaceae (24 OTUs) and Entolomataceae (17 OTUs)  
314 (Figure S4d). Members of Ceratobasidiaceae were most abundant (190338 sequences,  
315 25.95 %), followed by Tulasnellaceae (115167 sequences, 13.58%), Entolomataceae  
316 (84384 sequences, 9.95%), Inocybaceae (76731 sequences, 9.05%) and Nectriaceae  
317 (66383 sequences, 7.83%). Members of the ectomycorrhizal Sebacinaceae,  
318 Thelephoraceae, Inocybaceae, Russulaceae and Tuberaceae represented < 1% relative  
319 abundance (Figure S4a).



320 *E. helleborine* - The most abundant fungi detected in *E. helleborine* belonged to  
321 Helotiales (134 OTUs), Ceratobasidiaceae (22 OTUs) and Serendipitaceae (18 OTUs),  
322 as well as ectomycorrhizal taxa Thelephoraceae (62 OTUs), Inocybaceae (29 OTUs),  
323 Sebacinaceae (25 OTUs), Russulaceae (32 OTUs), Cortinariaceae (30 OTUs),  
324 Helvellaceae (14 OTUs), Tuberaceae (13 OTUs) and Hymenogastraceae (2 OTUs). In  
325 addition, a number of other fungal taxa previously shown to colonize *Epipactis* spp.  
326 were detected, including Tricholomataceae (16 OTUs), Gloniaceae (12 OTUs),  
327 Herpotrichiellaceae (9 OTUs), Pyronemataceae (5 OTUs) and Psathyrellaceae (3 OTUs)  
328 (Figure S5d). Members of Tulasnellaceae were represented by six OTUs, five of which  
329 were detected in European samples, and only one from an individual plant collected  
330 from China. In terms of relative abundances of sequences, members of Tuberaceae were  
331 most abundant (216586 sequences, 28.86 %), followed by Helotiales (176777  
332 sequences, 23.56 %), Russulaceae (101541 sequences, 13.53%), Pyronemataceae  
333 (44388 sequences, 5.92%), Helvellaceae (41300 sequences, 5.50%) and Sebacinaceae  
334 (38618 sequences, 5.15%) (Figure S5a).

335

### 336 **3.2 Mycorrhizal fungal communities**

337 *G. conopsea* - *G. conopsea* mainly associated with members of the  
338 Ceratobasidiaceae and Tulasnellaceae and ectomycorrhizal fungi from the  
339 Sebacinaceae and Thelephoraceae. These fungal associates represented in total 95  
340 OTUs (314182 sequences, 37.02% of the total sequences) (Table S1), of which 71  
341 (80672 sequences) and 50 (233510 sequences) were detected in Europe and China,

342 respectively (Table S2). Thirty-nine OTUs (47.6% of all OTUs) were shared between  
343 Europe and China. In terms of relative abundance, Ceratobasidiaceae and  
344 Tulasnellaceae were the most abundant in Europe and China. Members of the  
345 Tulasnellaceae were the most abundant fungal associates in Europe, while members of  
346 the Ceratobasidiaceae were the most abundant fungal associates of *G. conopsea* in  
347 China (Figure S4b, c, e). About half of the rhizoctonia fungal OTUs were detected in  
348 samples from both Europe and China (Figure 1a). In contrast, none of the fungal OTUs  
349 of Sebacinaceae was shared between Europe and China. The average number of  
350 mycorrhizal OTUs detected on the roots of individuals of *G. conopsea* in Europe (16.6,  
351 range: 9–27) was significantly smaller than that found on roots of plants in China (20.2,  
352 range: 7–29;  $F = 4.047$ ,  $p = 0.012$ ) (Table 1). However, PD values did not significantly  
353 ( $F = 2.706$ ,  $p > 0.05$ ) differ between plants from Europe ( $3.117 \pm 0.207$ ) and China  
354 ( $3.128 \pm 0.121$ ) (Table 1).

355 ***E. helleborine*** - 432 OTUs (765472 sequences, 63.89% of the total sequences)  
356 previously described as fungal associates from the genus *Epipactis* were detected in this  
357 research (Table S3). 326 OTUs (359589 sequences) and 169 OTUs (405883 sequences)  
358 were detected in Europe and China, respectively, and 64 OTUs (196428 sequences;  
359 14.8% of all OTUs) were shared (Table S4). When excluding the Helotiales (134 OTUs,  
360 176777 sequences), of the remaining 298 OTUs (588695 sequences), 234 OTUs  
361 (299690 sequences) and 83 OTUs (289005 sequences) were detected in Europe and  
362 China, respectively, and 20 OTUs (36968 sequences; 6.7% of all OTUs) occurred in  
363 both regions. They belonged to nine different fungal families (Herpotrichiellaceae, 5

364 OTUs; Gloniaceae, 5 OTUs; Ceratobasidiaceae, 3 OTUs; Serendipitaceae, 2 OTUs;  
365 Thelephoraceae 2 OTUs; Psathyrellaceae, Tricholomataceae, Tulasnellaceae, and  
366 Pyronemataceae, 1 OTU respectively).

367 The relative abundance of the fungal families differed between Europe and China  
368 (Figure 1b). Members of Russulaceae were most abundant in Europe, while members  
369 of the Tuberaceae were most abundant in China. Members of Tulasnellaceae,  
370 Ceratobasidiaceae and Serendipitaceae were present in relatively low abundance in  
371 both Europe and China (Figure S5b, c, e). The average number of OTUs detected on  
372 the roots of individuals of *E. helleborine* in Europe (89.3, range: 43–112) was  
373 significantly higher than those on the roots of plants from China (22.1, range: 8–39;  $F$   
374 = 3.984,  $p < 0.001$ ) (Table 1). The difference is still significant when Helotiales is  
375 excluded ( $F = 3.926$ ,  $p < 0.001$ ; Europe 61.9, range: 36–78; China 9.0, range 3–15).  
376 Similarly, PD values of the mycorrhizal communities associating with *E. helleborine*  
377 in Europe ( $8.903 \pm 0.337$ ) were significantly ( $F = 3.980$ ,  $p < 0.001$ ) higher than those  
378 of the communities associating with *Epipactis* in China ( $3.189 \pm 0.122$ , Table 1).

379

### 380 **3.3 Comparison of mycorrhizal communities**

381 The estimated overall multiple-site dissimilarity was higher for *E. helleborine* ( $\beta_{\text{sor}} =$   
382 0.86) than for *G. conopsea* ( $\beta_{\text{sor}} = 0.76$ ). In both species most of the multiple-site  
383 dissimilarity was the result of spatial turnover ( $\beta_{\text{sim}} = 0.76$  and 0.74, respectively) and  
384 to a much smaller extent of nestedness ( $\beta_{\text{nes}} = 0.09$  and 0.02, respectively), indicating  
385 that the observed dissimilarity patterns are the result of taxon replacement and not by

386 taxon loss. In both orchid species, dissimilarity in mycorrhizal communities increased  
387 significantly with increasing geographic distance ( $r_M = 0.84$ ,  $P < 0.0001$  and  $r_M = 0.65$ ,  
388  $P = 0.02$  for *E. helleborine* and *G. conopsea*, respectively) (Fig. 2,3). When the  
389 contributions of spatial turnover and nestedness were quantified separately, spatial  
390 turnover contributed most to the observed patterns of mycorrhizal dissimilarity with  
391 nestedness showing little variation with increasing distance.

392 PCD values were generally low ( $<1$ ) for populations located close to each other,  
393 but increased ( $>1$ ) when populations were further apart (Fig. 2,3). For *E. helleborine*,  
394 the relationship between PCD values and geographic distances was highly significant  
395 ( $r_M = 0.82$ ,  $P < 0.0001$ ). Furthermore, the compositional component of PCD (PCDc)  
396 was also strongly and significantly correlated with geographic distance ( $r_M = 0.79$ ,  $P <$   
397  $0.0001$ ), while PCDp was less tightly correlated with geographic distance ( $r_M = 0.55$ ,  $P <$   
398  $= 0.0008$ ) (Fig. 2). In *G. conopsea*, both PCD and PCDc were significantly correlated  
399 to geographic distance (PCD:  $r_M = 0.62$ ,  $P = 0.0135$ ; PCDc:  $r_M = 0.59$ ,  $P = 0.0195$ ) (Fig.  
400 3), but there was no significant relationship between PCDp and geographic distance ( $r_M$   
401  $= -0.25$ ,  $P > 0.05$ ) (Fig. 3).

402 Species indicator analyses showed that a large number of fungal OTUs was  
403 significantly associated with one of the two geographic regions. In total, 208 and 23  
404 mycorrhizal OTUs were significantly associated with *E. helleborine* growing in Europe  
405 and China, respectively (Table S5). In *G. conopsea*, 28 OTUs were significantly  
406 associated with Europe, while 21 OTUs almost exclusively occurred in China (Table  
407 S6).

## 408 **4 DISCUSSION**

409 The results of this study show that there is a general increase in dissimilarity with  
410 increasing geographic distance in mycorrhizal communities associating with two  
411 widespread orchids, *Epipactis helleborine* and *Gymnadenia conopsea*. This pattern is  
412 consistent with previous work that has documented a decay in similarity for a wide  
413 range of organisms (Nekola & White, 1999; Soininen et al., 2007) and confirms earlier  
414 findings that patterns repeatedly observed for macro-organisms may also occur in  
415 microorganisms such as fungi (Talbot et al., 2014). In addition, our results showed that  
416 most of the observed dissimilarity was the result of spatial turnover and not of  
417 nestedness, indicating that both investigated orchid species are mycorrhizal generalists  
418 that show large geographic variation in their mycorrhizal communities.

419

### 420 **4.1 Mycorrhizal fungi associating with two widespread orchid species**

421 A recent review on the biogeography of orchid mycorrhizas has suggested that the  
422 fungal families that associate with orchids occur in a wide variety of habitats and that  
423 some of these fungal species have a very wide distribution (Jacquemyn et al., 2017b).

424 Our results are in line with these observations. In our study, *Gymnadenia conopsea*  
425 mainly associated with fungi of the Ceratobasidiaceae and Tulasnellaceae, while  
426 members of Sebaciniales were present at low abundance. In addition, ectomycorrhizal  
427 taxa of the Thelephoraceae, Russulaceae, Inocybaceae and Cortinariaceae were also  
428 detected. When comparing the OMF community composition of *G. conopsea* between  
429 European and Asian populations, around a half of the dominant fungi found in Europe

430 were also found in China, indicating that these fungi have a very broad geographic  
431 distribution. Six out of nine Tulasnellaceae OTUs, 21 out of 42 Ceratobasidiaceae, and  
432 six out of nine Serendipitaceae OTUs were shared between Europe and China. By  
433 allowing seed germination the widespread distribution of these fungal taxa may  
434 therefore contribute to the widespread distribution of *G. conopsea*. In contrast, for  
435 members of the ectomycorrhizal Thelephoraceae, less than a quarter of the OTUs was  
436 found in both Europe and Asia, none of the Sebacinaceae members was found in both  
437 areas, indicating that the presence of these fungal members tends to vary across sites  
438 and therefore may be of lesser importance than rhizoctonia fungi. This pattern of  
439 interaction specificity strongly resembles a pattern that was recently coined ‘apparent  
440 generalism’, in which an orchid species specializes on one or few host species that  
441 contribute unique resources, but also associates with other host species that contribute  
442 functionally redundant resources (Shefferson et al., 2019).

443 In contrast, the mycorrhizal communities found in the roots of *E. helleborine*  
444 showed a high regional specificity and most of the fungal associates differed between  
445 European and Asian populations. Plants of *E. helleborine* mainly associated with  
446 members of Helotiales, fungi of Ceratobasidiaceae and Serendipitaceae, and  
447 ectomycorrhizal fungi of the Thelephoraceae, Inocybaceae, Sebacinaceae and  
448 Russulaceae. In addition, a large number of other ectomycorrhizal taxa known as fungal  
449 associates of *E. helleborine* and other *Epipactis* species were detected, mainly including  
450 Tuberaceae, Cortinariaceae, Tricholomataceae, and Helvellaceae. Only around 15%  
451 (64 out of 432 putative mycorrhizal OTUs) of the mycorrhizal OTUs were shared

452 between both regions. The dominant fungal associates also differed between European  
453 and Asian populations. For example, members of Russulaceae were the most dominant  
454 fungal associates in Europe, whereas members of Tuberaceae were most abundant in  
455 Asian populations, and not one OTU belonging to these two families was shared  
456 between the two regions. Similar patterns were observed for OTUs belonging to  
457 Cortinariaceae, Hymenogastraceae, Inocybaceae, Tulasnellaceae, Sebacinaceae and  
458 Helvellaceae. This pattern of interaction specificity was recently coined ‘true  
459 generalism’, in which an orchid species associates with multiple hosts that overlap  
460 functionally, and that are geographically interchangeable based on opportunity for  
461 encounter, leading to frequent host switching (Shefferson et al., 2019). The observed  
462 low selectivity towards mycorrhizal fungi and strong spatial turnover in fungal  
463 communities may also explain why this species occurs in a wide variety of habitats and  
464 seemingly easily colonizes new habitat and even can become invasive (Owen, 1879;  
465 Soper & Garay, 1954).

466 The two investigated species display different nutritional modes: *E. helleborine* is  
467 a partially heterotrophic (mixotrophic) orchid species (Bidartondo et al., 2004;  
468 Schiebold et al., 2017) that obtains a part of its carbon from its own photosynthesis and  
469 the other part from its mycorrhizal fungi. In this species, the fungus may provide 20 to  
470 100% of its carbon to the plant, depending on the time in the growth season (Gonneau  
471 et al., 2014). In contrast, *G. conopsea* is often considered autotrophic, although a limited  
472 amount of carbon may be derived from the fungus (Schweiger et al., 2018). To what  
473 extent this biological difference in resource acquisition explains why *E. helleborine* is

474 more opportunistic in fungal associations than *G. conopsea* remains unclear at this point  
475 and warrants further investigation.

476

#### 477 **4.2 Patterns of dissimilarity**

478 Variation in the species composition of ecological communities can be the result of  
479 spatial species turnover and nestedness (Harrison et al., 1992; Baselga, 2010).

480 Nestedness refers to the non-random loss of species and leads to progressive  
481 dissimilarity between the most species-rich communities and communities that contain  
482 only a few species anymore. Spatial turnover, on the other hand, refers to the repeated  
483 replacement of one species by another. Both processes can contribute to changes in  
484 community composition across large geographic areas and therefore it is important to  
485 assess the contribution of both processes to identify the potential causes determining  
486 variation in biotic communities (Baselga, 2010).

487 Our results showed that for both orchid species spatial turnover was the most  
488 important factor contributing to the observed variation in mycorrhizal communities. For  
489 both species, the relationships between PCD values and geographic distances were  
490 also significant, indicating that populations located close to each other have  
491 mycorrhizal communities that are more similar than randomly selected communities,  
492 whereas populations separated by large distances have mycorrhizal communities that  
493 are more dissimilar than random communities. Furthermore, the compositional  
494 component of PCD (PCDc) was strongly and significantly correlated with geographic  
495 distance, while PCDp was not significantly or only weakly correlated with geographic



496 distance. These results indicate that orchid populations located closely next to each  
497 other are likely to contain the same species (PCDc), but that the non-shared taxa come  
498 from different phylogenetic clades (PCDp).

499       The occurrence and geographic variation in local abundance of fungal strains can  
500 be influenced by local habitat conditions (Pandey et al., 2013) and OMF community  
501 composition may therefore be the result of complex interactions between different  
502 factors, including extrinsic factors (habitat type, geographic site, soil characteristics,  
503 etc.) and intrinsic factors (genetic differentiation, phylogeny of host plants; Swarts &  
504 Dixon, 2009; McCormick & Jacquemyn, 2014; Xing et al., 2017, Chen et al., 2019).  
505 Previous research has, for example, shown that variation in local environmental factors  
506 such as soil moisture content, pH, nutrient conditions (especially soil carbon, nitrogen  
507 and phosphorus) can generate pronounced differences in mycorrhizal communities  
508 (Bunch et al., 2013; Jacquemyn et al., 2015). For example, populations of the terrestrial  
509 orchid *Neottia ovata* occurring in forest and meadow habitats showed significantly  
510 different OM fungal communities (Oja et al., 2015). Mycorrhizal communities even  
511 vary between populations mainly due to differences in soil moisture content and pH  
512 (Jacquemyn et al., 2015). In the case of *E. helleborine*, differences in tree species  
513 composition and associated ectomycorrhizal communities may explain the observed  
514 differences between regions and the role of spatial turnover. More research is needed  
515 why some fungal families display very large geographic distributions, whereas others  
516 seem to be more restricted.

517

518 **5 CONCLUSION**

519 This cross-continental comparison of the mycorrhizal communities associated with two  
520 widespread terrestrial orchid *E. helleborine* and *G. conopsea* shows how similarities in  
521 fungal communities change with distance. The fungal community composition of the  
522 two orchid species differed significantly between Europe and China, leading to  
523 significant turnover in mycorrhizal communities and significant decay of similarity  
524 across large geographic distances. Nonetheless, some OTUs were found in both  
525 continents, suggesting that these fungi have very wide distributions that are not  
526 restricted by soil or local climate conditions. Strong turnover in fungal communities  
527 and significant decay of similarity with distance indicate that these orchids are  
528 generalists in their OMF communities across large geographic areas. More research is  
529 needed to understand the relative contribution of mycorrhizal taxa on the fitness of  
530 orchid that associate with multiple taxa, and whether orchid populations associated with  
531 particular OMF communities in one region are able to readily utilize OMF in another  
532 region.

533

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**Table 1** Sampling sites, average mycorrhizal fungal OTU numbers and phylogenetic diversity (PD) in each population of *Epipactis helleborine* and *Gymnadenia conopsea* growing in Europe and China.

Population	Region, country	Latitude	Longitude	Number OTUs	PD
<i>Gymnadenia conopsea</i>					
GITA	Passo San Lanciano, Italy	42°10'47"	14°6'39"	15.6	3.03 ± 0.14
GITB	Passo San Leonardo, Italy	42°5'22"	14°1' 55"	14.8	3.15 ± 0.08
GBEA	Bonnerieu, Belgium	50°06'23"	4°43'5"	13.6	2.65 ± 0.46
GPLA	Kalina Lisiniec, Poland	50°21'44"	20°9'37"	22.5	3.63 ± 0.28
GB	Baihua Mountain, Beijing, China	39°49'19"	115°35'35"	17.2	3.20 ± 0.09
GC	Changbai Mountain, Jilin province, China	41°54'31"	128°0'18"	19.6	3.10 ± 0.41
GN	Lanping county, Yunnan province, China	29°24'36"	99°0'15"	23	3.47 ± 0.24
GS	Gongga Mountain, Sichuan Province, China	29°36'4"	102°0'42"	22.2	3.42 ± 0.25
GG	Min county, Gansu province, China	34°24'12"	104°18'12"	19.6	2.82 ± 0.43
GZ	Milin county, Tibet, China	29°7'8"	93°47'14"	19.4	2.76 ± 0.15
			Average:	18.8	3.11 ± 0.09
<i>Epipactis helleborine</i>					
EITA	Acquarotta, Italy	40°52'50"	14°35'52"	62.4	7.34 ± 0.31
EITB	Casafredda, Italy	40°57'20"	14°40'54"	88.2	9.70 ± 0.37
EBEA	Bierbeek, Belgium	50°48'50"	4°44'29"	101.6	9.94 ± 0.31
EBEB	Bierbeek, Belgium	50°48'37"	4°44'36"	102.0	9.26 ± 0.08
EBEC	Ave-et-Auffe, Belgium	50°06'04"	5°09'29"	97.6	9.19 ± 0.23
EBED	Belvaux, Belgium	50°06'04"	5°10'30"	86.6	8.40 ± 0.35
EPLA	Tunel, Poland	50°27'29"	19°58'41"	86.5	8.48 ± 0.21
EJL	Jiaohe, Jilin province, China	43°48'12"	127°2'42"	25.0	3.41 ± 0.17
ESX	Lingchuan county, Shanxi province, China	35°48'16"	113°24'42"	22.0	3.65 ± 0.25

ENJA	Lanping county, Yunnan province, China	29°24'54"	99°0'15"	17.0	2.74±0.42
ENJB	Lanping county, Yunnan province, China	29°24'35"	99°0'18"	25.4	3.44±0.23
ENJC	Lanping county, Yunnan province, China	29°24'26"	99°0'11"	26.0	2.95±0.27
ENX	Nixi Township, Shangri-la, Yunnan Province, China	28°18'18"	99°24'42"	21.4	3.16±0.28
EBLGZ	Balog Zon, Shangri-la, Yunnan Province, China	28°18'06"	99°24'36"	17.6	2.98±0.44
			Average:	55.2	6.01±0.36

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## Figure captions

**FIGURE 1** Sampling location and distribution of putative mycorrhizal fungi associated with populations of (a) *Gymnadenia conopsea* (closed circles) and (b) *Epipactis helleborine* (open circles), across their Eurasian distributions. Pie charts represent the relative number of reads belonging to each fungal family in each region sampled.

**FIGURE 2** Relationships between geographic distances and mycorrhizal dissimilarity ( $\beta_{\text{sor}}$ ,  $\beta_{\text{sim}}$  and  $\beta_{\text{nes}}$ ) and phylogenetic dissimilarity (PCD, PCDC, PCDp) of populations of *Epipactis helleborine* sampled across Eurasia.

**FIGURE 3** Relationships between geographic distances and mycorrhizal dissimilarity ( $\beta_{\text{sor}}$ ,  $\beta_{\text{sim}}$  and  $\beta_{\text{nes}}$ ) and phylogenetic dissimilarity (PCD, PCDC, PCDp) of populations of *Gymnadenia conopsea* sampled across Eurasia.

## Supporting Information

**FIGURE S1** Rarefaction curves of Fungal OTU (operational taxonomic unit) richness in individual plant of *Epipactis helleborine* (a) and *Gymnadenia conopsea* (b).

**FIGURE S2** Fungal diversity and phylogenetic diversity of the whole fungal communities detected in each populations of *Epipactis helleborine*.

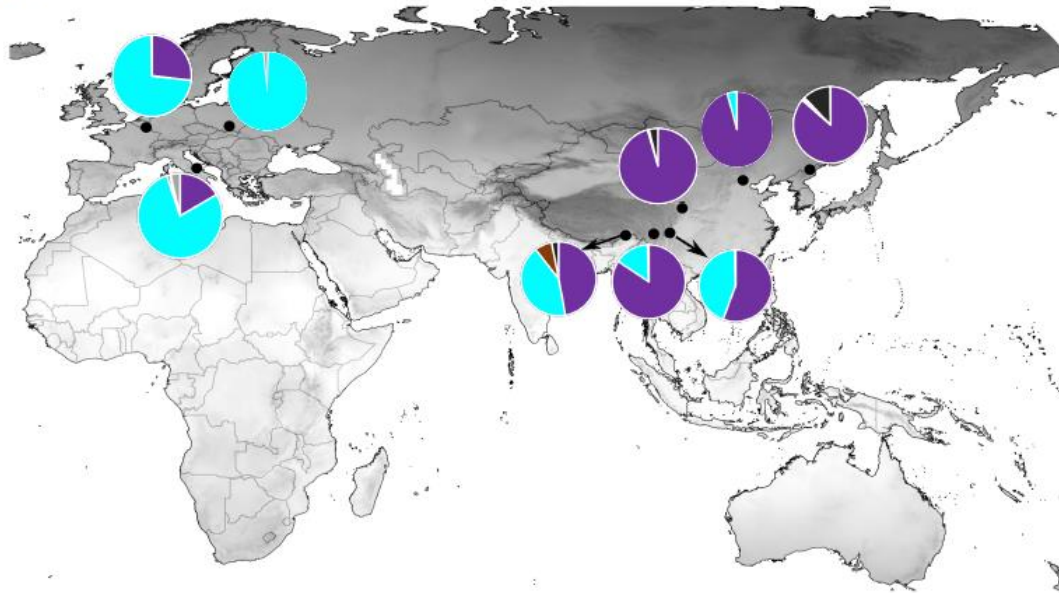
**FIGURE S3** Fungal diversity and phylogenetic diversity of the whole fungal communities detected in each populations of *Gymnadenia conopsea*.

**FIGURE S4** Relative abundance of the most abundant fungal families detected in *Gymnadenia conopsea* in total (a), in Europe (b) and in China (c). (d) The number of operational taxonomic units (OTUs) for different fungal families. (e) Relative abundance of fungal families in each *G. conopsea* population.

**FIGURE S5** Putative mycorrhizal fungal families detected in *Epipactis helleborine* Relative abundance of each fungal family in the whole mycorrhizal communities in total (a), in Europe (b) and in China (c). (d) The number of operational taxonomic units (OTUs) for different mycorrhizal fungal families. (e) Relative abundance of mycorrhizal families in each *E. helleborine* population.

Fig. 1

(a)



(b)

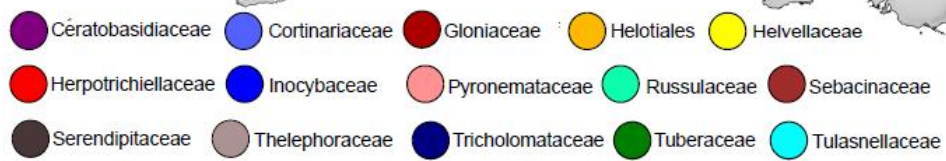
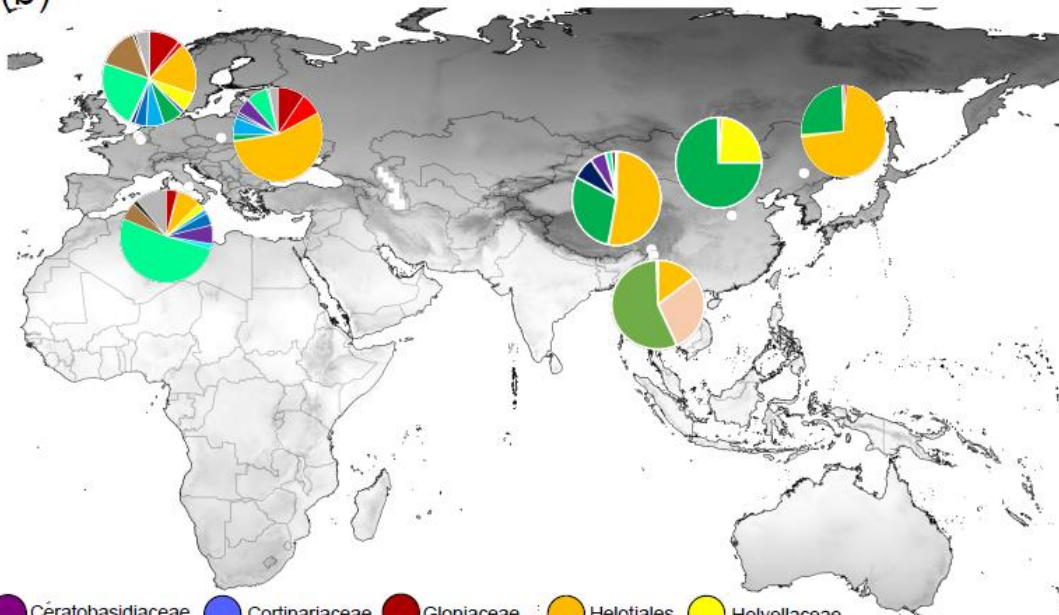


Fig. 2

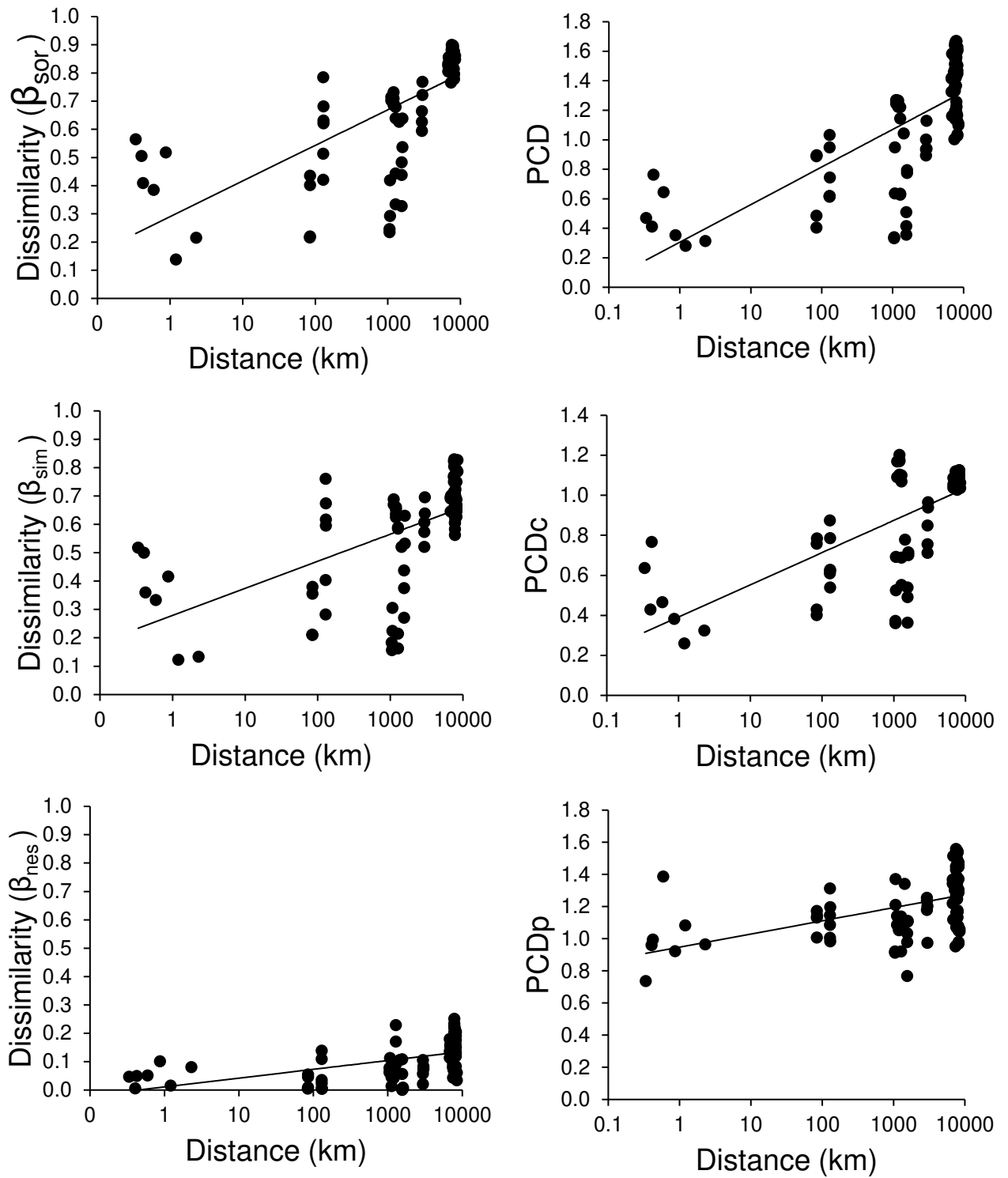


Fig. 3

