



Conference on ‘Roles of sleep and circadian rhythms in the origin and nutritional management of obesity and metabolic disease’

Symposium 1: Relevance of circadian rhythms and sleep to obesity and metabolic disease

Sleep, circadian rhythm and body weight: parallel developments

Margriet S. Westerterp-Plantenga*

Department of Human Biology, Nutrim, Maastricht University, MUMC+, P.O. Box 616, 6200 MD, Maastricht, The Netherlands

Circadian alignment is crucial for body-weight management, and for metabolic health. In this context, circadian alignment consists of alignment of sleep, meal patterns and physical activity. During puberty a significant reduction in sleep duration occurs, and pubertal status is inversely associated with sleep duration. A consistent inverse association between habitual sleep duration and body-weight development occurs, independent of possible confounders. Research on misalignment reveals that circadian misalignment affects sleep-architecture and subsequently disturbs glucose–insulin metabolism, substrate oxidation, leptin- and ghrelin concentrations, appetite, food reward, hypothalamic–pituitary–adrenal-axis activity and gut-peptide concentrations enhancing positive energy balance and metabolic disturbance. Not only aligning meals and sleep in a circadian way is crucial, also regular physical activity during the day strongly promotes the stability and amplitude of circadian rhythm, and thus may serve as an instrument to restore poor circadian rhythms. Endogenicity may play a role in interaction of these environmental variables with a genetic predisposition. In conclusion, notwithstanding the separate favourable effects of sufficient daily physical activity, regular meal patterns, sufficient sleep duration and quality sleep on energy balance, the overall effect of the amplitude and stability of the circadian rhythm, perhaps including genetic predisposition, may integrate the separate effects in an additive way.

Sleep disruption: Circadian misalignment: Overweight: Insulin sensitivity: Metabolic disorders

Energy- and food-reward homeostasis are the essential components for maintaining body weight and body composition. Disruption of such homeostasis may lead to metabolic disorders, including obesity, diabetes and cancer^(1–3). The organism’s ability to coordinate daily patterns of activity, feeding, energy utilisation and energy storage, supported by a synchronised pattern of release of the relevant endocrine components across the daily 24-h cycle, is crucial for energy homeostasis^(4–6). Circadian alignment implies synchronisation of behavioural and physiological rhythms by the master circadian

clock, i.e. the suprachiasmatic nucleus (SCN) of the hypothalamus. The master clock is connected to peripheral tissues of the body that contain the molecular clock machinery required for local circadian oscillation and rhythmic gene expression^(7,8). However, metabolic processes are decoupled from the primarily light-driven SCN when food intake is desynchronised from normal diurnal patterns of activity^(4,5,9), and a dissociation of feeding patterns from SCN-based timing occurs, resulting in changes in energy availability, substrate oxidation, storage and metabolic status^(1,2,7,10–13). If feeding becomes

Abbreviations: GLP-1, glucagon-like peptide-1; HOMA-IR, homeostasis model assessment of insulin resistance; HPA axis, hypothalamic–pituitary–adrenal axis; IR, insulin resistance; QS, quality sleep; REM, rapid eye movement; SCN, suprachiasmatic nucleus; SWS, slow wave sleep; WT, wrist temperature.

*Corresponding author: Professor M. S. Westerterp-Plantenga, email m.westerterp@maastrichtuniversity.nl

the dominant entraining stimulus, adaptation to the changed food-intake patterns occurs, facilitated by an autonomous food-entrainable oscillator that governs behavioural rhythms^(2,5,7,8,11,12). Moreover, core circadian clock genes involved in reciprocal transcriptional feedback with genetic regulators of metabolism, and directly responsive to cellular energy supply, are involved^(7,8). In addition, the reward and motivational value of food is a potent synchroniser for the SCN clock⁽¹⁴⁾. This suggests that energy metabolism and motivational properties of food can influence the clock mechanism of the SCN. Food-related cues may entrain clock genes of the SCN directly or indirectly, and play an integral role as a food-entrainable oscillator, responsible for anticipation of meal-time⁽⁸⁾. This close interaction is likely to be critical for normal circadian regulation of metabolism, and may underlie the disruption of proper metabolic rhythms observed in metabolic disorders, such as obesity and type-II diabetes^(4,10,15–19).

This review deals with effects of disruption of sleep and quality sleep (QS), and with effects of circadian misalignment on energy- and food-reward homeostasis, endocrinological factors, body weight and body composition. Before considering these topics in adults, the development of sleeping hours and body weight during puberty, and the consequences, will be addressed.

Development of sleep duration and body weight during puberty

During puberty, sex differences in anthropometric and endocrine variables are observed in the transition from Tanner stages 1–5^(20–23). Puberty is initiated through pulsatile gonadotropin-releasing hormone release from the hypothalamus and activation of the gonadal axis^(1,24,25). The subsequent development of secondary sex characteristics originates from shared neuronal systems, with the hypothalamus as integration point⁽¹⁾. The hypothalamus regulates the sleep–wake and feeding circuits^(1,26). Circuits are connected through the hypocretin-1 hormone that regulates feeding and locomotor activity via the nucleus accumbens, as well as signal transduction on the light–dark cycle to the SCN. Changes in hypothalamic functioning, such as disturbed hypocretin-1 signalling, are associated with disturbance of the circadian cycle and feeding behaviour, affecting energy balance and body composition^(1,26). Furthermore, during puberty a significant reduction in sleep duration occurs^(20–23), and pubertal status appears to be inversely associated with sleep duration^(20–23). Moreover, studies report a consistent inverse association between habitual sleep duration and body-weight development^(21–23), independent of BMI at the start of puberty, fat mass and obesity-associated allele genotype (rs9939609), parent BMI, as well as changes in physical activity and hours of television viewing. Therefore, the changes in hypothalamic functioning during puberty may explain the relationships between the changes in BMI and in sleep duration during puberty. Since these developments occur in parallel, cause and effect cannot be distinguished.

Sleep and metabolic disorders in children and adolescents

Sleep, and sleep architecture are important factors for normal growth and development during childhood^(27–32). Especially total sleeping time and QS ((slow wave sleep (SWS) + rapid eye movement sleep (REM))/total sleeping time) are crucial sleep factors associated with outcomes on physical, cognitive, emotional and social development in children⁽³⁰⁾. Sleep deprivation and poor QS have been identified as an independent risk factor for the development of insulin resistance (IR)^(27–30). Studies on sleep in relation to IR in adolescents^(33–35) show for instance that acute sleep restriction reduced insulin sensitivity in adolescent boys⁽³⁴⁾. Observations on sleep duration in healthy black and white adolescents show an inverse association between sleep duration and homeostasis model assessment of IR (HOMA-IR). Interventions to extend sleep duration may reduce diabetes and obesity risk in youth⁽³⁵⁾. A cross-sectional analysis from two examinations conducted in the Cleveland Children's Sleep and Health Cohort (n 387; 43 % minorities) shows a quadratic U-shape association between sleep duration and HOMA-IR. When adjusted for age, sex, race, preterm status and activity, adolescents who slept 7.75 h had the lowest predicted HOMA-IR, while adolescents who slept 5.0 or 10.5 h had HOMA-IR indices that were approximately 20 % higher. It was concluded that shorter and longer sleep durations are associated with decreased insulin sensitivity in adolescents⁽³³⁾. Another lifestyle factor, namely physical activity, second for improving insulin sensitivity, may act directly as well as indirectly by its effects on sleep characteristics. With regard to food intake, studies have found associations between sleep deprivation and food choices^(34–39). Sleep-deprived individuals appear more prone to choose unhealthy foods high in energy and fat content^(34–39). Also, sleep-deprived individuals are reported to have more frequent meals or snacks between meals compared with individuals who had sufficient sleep⁽³⁶⁾. It has been assumed that sleep deprivation is associated with decreased leptin concentrations and increased ghrelin concentrations, thereby promoting the feeling of hunger and suppressing satiety^(40,41). A reduced sleep duration, reduced quality of sleep and REM sleep, or fragmented sleep enhance a positive energy balance through altered substrate oxidation, hormone concentrations, sleeping metabolic rate, appetitive behaviours and stress⁽⁴²⁾. Circadian misalignment affects sleep architecture and the glucose–insulin metabolism, substrate oxidation, the HOMA-IR index, leptin concentrations and hypothalamic–pituitary–adrenal (HPA)-axis activity^(42–45).

Sleep disruption, body weight and metabolic disorders in adults

Sleep and metabolism

Sleep and circadian rhythms have direct impacts on energy metabolism, and represent important mechanisms underlying the major health epidemics of obesity and

diabetes^(3,5). The majority of studies are observational in nature, and a few intervention studies have been reported. For instance, after partial sleep deprivation increased food intake and increased intake of energy from snacks with a higher carbohydrate or fat content has been shown^(46–49), and in short sleepers, increased respiratory quotient, implying increase in carbohydrate oxidation was observed⁽⁵⁰⁾. Sleep deprivation may affect energy balance through hormone changes^(40,51–53), although this was not observed in all studies^(46,54–56).

In addition to reduced sleep, reduced QS is associated with metabolic disorders. Reduced QS obtained by a single night of fragmented sleep without reducing total sleeping time, induced a shift in insulin concentrations, from being lower in the morning and higher in the afternoon, while glucagon-like peptide-1 (GLP-1) concentrations and fullness scores were decreased. The decreased GLP-1 concentrations and fullness scores in the afternoon were synchronously related with reduced Visual Analogue Scale fullness scores, and increased Visual Analogue Scale desire to eat scores after dinner. This may lead to increased food intake and snacking, thus contributing to a positive energy balance⁽⁴³⁾. Also an increased respiratory quotient was observed when sleep was fragmented⁽⁵⁷⁾. Reduced QS affects several neuroendocrine signals involved in the control of substrate utilisation, including cortisol concentrations. The sharp morning rise and the steep fall to lower evening levels are modulated by even a single night of reduced sleep^(40,53). After a single night of reduced QS, cortisol levels were significantly reduced after awakening and were elevated in the evening^(40,43,53). Taken together, a reduced sleep-duration, and reduced QS affect substrate oxidation, leptin- and ghrelin concentrations, sleeping metabolic rate, appetite, food reward, HPA-axis activity, gut-peptide concentrations as such, that a positive energy balance is enhanced, which increases the risk for overweight.

Sleep and body-weight management

Effects of changes in sleep duration during a dietary intervention for body-weight loss was assessed by Nedeltcheva *et al.*^(46,50). They showed that sleep restriction to 5.5 h sleep compared with 8.5 h sleep compromised the efficacy of a dietary intervention for weight loss. The combination of energy and sleep restriction in overweight adults resulted in decreased loss of fat and considerably increased loss of fat-free body mass. These results suggest that sleep plays a role in the preservation of human fat-free body mass during periods of reduced energy intake^(46,50). The effect on sparing fat-free mass was confirmed by Verhoef *et al.*⁽⁵⁸⁾ who assessed whether during a weight-loss weight-maintenance programme in overweight subjects, a possible increase in sleep duration would precede the diet-induced decreases in body weight. They observed a concomitant inverse correlation between changes in sleep duration and in body weight, and respectively fat mass.

In addition, Chaput *et al.*⁽⁵⁹⁾ observed that short-duration sleepers who maintained their short sleep duration habits experienced a greater increase in BMI and

fat mass over a 6-year follow-up period compared with short-duration sleepers who increased their sleep duration, suggesting that shifting sleep duration from a short length to a healthier length is associated with lower adiposity gain⁽⁵⁹⁾. Moreover, they showed that both sleep duration and sleep quality were significantly related to fat mass loss during dietary interventions in overweight and obese adults⁽⁶⁰⁾. Despite these significant correlations it is not possible to determine any direction of causation.

Circadian alignment and energy balance

The significance of circadian alignment and energy balance implies assessment of the significance of circadian alignment for sleep, and sleep architecture, food-intake regulation, and physical activity. This is mainly assessed by circadian misalignment experiments. In the following sections, effects of circadian misalignment on sleep architecture and food-intake regulation are considered. In addition the significance of physical activity for circadian alignment will be highlighted.

Circadian misalignment and sleep

Circadian misalignment may reduce total sleep time, but mainly affects sleep architecture. The circadian phase at which sleep occurs affects the distribution of sleep stages. The preferential distribution of REM sleep towards the latter part of the night is linked to a circadian oscillator, while the preferential distribution of SWS towards the beginning of a sleep episode is mediated by homeostatic processes, i.e. the length of prior wakefulness⁽⁶¹⁾. Circadian misalignment resulted in disruption of the normal phase relationship between SWS and REM sleep, so that REM sleep is relatively phase advanced to SWS⁽⁴⁴⁾. This abnormal circadian sequencing results in shortening of REM sleep latency and increasing REM sleep duration in a phase advanced stage. This short latency to REM sleep is typical of narcoleptic and depressive patients⁽⁶²⁾. Mood disorders, especially unipolar depression and seasonal affective disorder, have been linked to circadian rhythm abnormalities⁽⁶²⁾. Dysregulation in the HPA-axis, implying an overall increased cortisol secretion with a phase advance of the cortisol circadian rhythm is extremely frequent in individuals with depression^(63,64). Misalignment between timing of the clock and the timing of sleep, in either direction, has been associated with depression in vulnerable individuals⁽⁶⁵⁾.

Increased REM sleep during both a phase advance and a phase delay is not favourable^(46,50,66–72), because this results in a relatively shorter REM sleep duration during the second part of the night, associated with higher cortisol concentrations, higher fasting insulin concentrations, and a higher HOMA-IR index^(44,64,73,74). Sleep during the circadian nadir (03.00–06.00 hours), is important in protecting normal physiological rhythms and function of the HPA-axis⁽⁷⁴⁾. Circadian misalignment, both a phase advance and a phase delay results in dysregulation of the HPA-axis. All in all, circadian misalignment appears to affect sleep-architecture, namely the distribution of sleep stages. REM sleep then becomes

phase advanced to SWS with reduced REM sleep latency. REM sleep duration increases during phase advance, and during phase delay, resulting in a shorter REM sleep duration during the second part of the night.

Circadian misalignment, endocrinology, energy homeostasis and meal patterns

The daily patterns of feeding, energy utilisation, and energy storage across the daily 24-h cycle, is based upon a neuro-endocrinological system^(75,76). Metabolically relevant hormones show circadian oscillation with different daily patterns. Cortisol secretion has a circadian rhythm with the nadir during the early biological night (i.e. a time according to the original circadian rhythm associated with the start of behavioural inactivity) and the peak in the biological morning (i.e. a time according to the circadian rhythm associated with the start of behavioural activity)⁽⁷⁷⁾. Glucose and insulin levels peak during the late biological night^(78,79). Leptin, which suppresses appetite, is secreted in a circadian manner^(80–83). In human subjects, night-time plasma leptin levels are high when appetite decreases, favouring fasting and nocturnal rest, and low during the day, when hunger increases. Gastric leptin levels oscillate in a circadian manner where leptin levels are high at night but low during the day^(84–86).

Ghrelin, which is produced in the stomach, pancreas and hypothalamus^(87,88), is involved in stimulating appetite via its action on neuropeptide Y in the lateral hypothalamus^(89,90) and can also alter clock function in the SCN *in vitro*^(91,92). Ghrelin oscillates with feeding⁽⁹³⁾, making this peptide a putative candidate for food-related entraining signals. In addition, elevated levels of ghrelin were found during the early part of the night in sleeping subjects, decreasing in the morning before awakening⁽⁹³⁾. Sleep deprivation can increase circulating ghrelin levels and this is accompanied by heightened hunger sensation⁽⁵⁴⁾. Thus, ghrelin may be a signal involved in the cross-talk between the peripheral and central circadian clock system.

In parallel to the circadian changes in neuropeptide levels and humoral signals from peripheral tissues, a circadian rhythm in macronutrient selection occurs. In human subjects, a carbohydrate-rich diet is favoured during breakfast and high-fat diets are preferred during evening meals⁽⁹⁴⁾. Carbohydrates are metabolised better during breakfast because, also in relation to the glucostatic theory, then the body metabolically responds more readily to a glucose stimulus, since the fasting glucose level then is relatively stable, and very clearly indicates the first transient glucose decline⁽⁹⁵⁾.

Disruption of the circadian system affects metabolic and cardiovascular changes^(2,45,96–98). In relation to appetite, release of some endocrine products shifts with meal patterns, such as glucose, insulin, GLP-1, ghrelin and leptin concentrations. Independently, disruption of the circadian system was associated with a significantly increased insulin response⁽⁴⁵⁾, possibly related to hyperglycaemia associated with a progressing IR associated with sleep restriction, and to increased sympathetic

nervous system activity⁽⁹⁶⁾. Changes in the magnitudes of glucose and insulin responses indicate a disturbed glucose and insulin metabolism^(86,97) and decreased GLP-1 concentrations indicate decreased satiety⁽⁴⁵⁾.

Cortisol levels do not show a meal-related pattern during misalignment. During circadian misalignment the cortisol curve is flattened compared to 24-h cycles^(11,45). In addition, with sleep loss, cortisol may exert its deleterious metabolic effects through remaining high night-time concentrations, which are associated with IR, suppressed immunity and increased inflammation⁽⁹⁸⁾.

Moreover, circadian disruption results in increased carbohydrate oxidation⁽⁴⁵⁾, probably due to both hyperinsulinaemia and hyperglycaemia⁽⁹⁹⁾. Often the increased carbohydrate oxidation occurs at the cost of protein oxidation, while fat oxidation remains constant⁽⁴⁵⁾.

The main effect of circadian misalignment, either phase advanced or phase delayed, is a concomitant disturbance of the glucose–insulin metabolism and substrate-oxidation. Chronically eating and sleeping at unusual circadian times may create a health risk through metabolic disturbance⁽⁴⁵⁾.

Consequently, meals need to be aligned in a circadian fashion. This requires timing and regularity of meals, with respect to food selection, meal frequency, meal intervals and meal size^(9,10,76). Beneficial metabolic effects of regular meal frequency on dietary thermogenesis, insulin sensitivity and fasting lipid profiles in healthy obese women align with effects of regular circadian patterns⁽⁴²⁾. After a regular v. an irregular meal pattern energy intake was lower, postprandial thermogenesis higher, and fasting total and LDL-cholesterol was lower⁽¹⁰⁰⁾. Peak insulin concentrations and area under the curve of insulin responses to the test meal were lower⁽¹⁰⁰⁾. Regular meal frequency creates more appropriate insulin sensitivity and lipid profiles compared with irregular meal frequency in healthy lean women⁽¹⁰¹⁾, and irregular meal frequency led to a lower postprandial energy expenditure compared with the regular meal frequency, while the mean energy intake was not significantly different between the two⁽¹⁰²⁾. The reduced diet-induced thermogenesis with the irregular meal frequency may lead to weight gain in the long term⁽¹⁰²⁾. With a regular meal frequency glucose excursions are blunted, net insulin production is reduced, and LDL-cholesterol concentrations tend to be lowered, mainly due to gastric emptying slowing down, and insulin production being reduced^(103–111). The net result is that lipid oxidation is favoured at the expense of glucose oxidation and lipid storage, and cholesterol synthesis is reduced. This may reduce adiposity and the level of circulating fatty acids, thereby leading to systematic, adaptive changes in both lipid and carbohydrate metabolism. Also examples of long-term responses to a sustained regular meal frequency such as improved glucose tolerance, and moderately reduced fasting plasma total and LDL-cholesterol, and a higher HDL:LDL cholesterol ratio are observed in normolipidaemic free-living subjects, as well as in type-II diabetes patients^(103–111). Also for cholesterol synthesis, meal frequency-dependent control of cholesterogenesis appeared to be mediated via hormonal mechanisms⁽⁹⁷⁾.



Furthermore, circadian alignment including careful and fixed timing of food intake and meal frequency plays a role in substrate utilisation and in energy expenditure. Large metabolic fluctuations in carbohydrate and fat oxidation were shown in a gorging food-intake pattern, while in the nibbling pattern, carbohydrate and fat oxidation remained relatively constant during the active hours of the day⁽¹¹²⁾. Also, the diet-induced thermogenic response was related to meal frequency⁽¹¹³⁾. In a series of experimental studies, variation in energy intake was primarily explained by habitual meal frequency, macronutrient composition and number of blood glucose declines⁽¹¹⁴⁾. The variation in habitual meal frequency was explained by percentage energy from carbohydrate or from fat in the diet, while the protein in the diet attenuates the metabolic amplitudes^(115,116). Moreover the effect of protein intake on satiety is partly due to the optimal timing of protein intake^(115,116). In healthy young men, habitual meal frequency appeared to be of greater significance in energy-intake regulation than forced meal frequency^(117–120). Adiposity may increase when young lean male subjects switch from a four- to a three-meal pattern by removing their usual afternoon meal, partly mediated by a change in the macronutrient composition of the diet⁽¹²¹⁾. Assessment of the effect of omitting or adding the third meal, revealing that eating three meals compared with two meals had no effects on 24 h energy expenditure, diet-induced thermogenesis, activity-induced energy expenditure and sleeping metabolic rate. However, eating the same amount of energy divided over three meals compared with over two meals, did increase satiety, particularly during the day, and did increase fat oxidation, particularly during the night in healthy, normal-weight women⁽¹²²⁾.

Circadian alignment and physical activity

To include circadian rhythm in the assessment of circadian alignment and physical activity, wrist skin temperature has been used as a valid method of assessing circadian rhythms in human subjects⁽¹²³⁾. As a measure for circadianity, circadian temperature amplitude and stability have been used as variables. Tranel *et al.*⁽¹²³⁾ identified physiological and behavioural measures that were significantly associated with the circadianity of these temperature parameters. Moreover, they tested the hypothesis that circadian temperature amplitude and stability would significantly differ among groups of healthy young men of varying adiposities⁽¹²³⁾. Wrist skin temperatures taken at 10 min intervals for seven consecutive days were determined in eighteen optimal, twenty fair and twenty-one poor % body fat grouped young men and subsequently analysed using available validated software. Body composition, cardiorespiratory fitness, actigraphy, daily nutritional and sleep data, and fasting lipid, insulin and glucose concentration measures were also determined⁽¹²³⁾. Subjects with one or more health problems had significantly lower temperature amplitude and stability⁽¹²³⁾. This occurred in subjects with a single metabolic syndrome risk factor compared to those with no metabolic syndrome risk factors, and

in subjects with a poor % body fat⁽¹²³⁾. In addition, step-wise multivariate regression analyses showed that 50 % of the variance in temperature amplitude was explained by actigraphy (mean steps taken per day), cardiorespiratory fitness, and late night eating per week; and 57 % in temperature stability by mean steps taken per day, time spent in moderate-to-vigorous activity per day, fat mass, and late night eating per week⁽¹²³⁾. Physical activity was the most important measure associated with the differences in circadian rhythm parameters⁽¹²³⁾.

Genetic background for the amplitude and stability of circadian rhythm

In order to shed light on the genetic component of the circadian marker described above, relative genetic and environmental influences on wrist temperature (WT) was determined using classical twin models⁽¹²⁴⁾. A study was performed in fifty-three pairs of female twins (twenty-eight monozygotic and twenty-five dizygotic), with a BMI 25.9 (SD 3.78) and mean age 52 (SD 6) years. The sample was selected from the Murcia Twin Register. Circadian patterns were studied by analysing WT during one week every 10 min using Circadianware[®]⁽¹²⁴⁾. Genetic influences on WT variability were estimated by comparing correlations of monozygotic and dizygotic twin pairs and fitting genetic structural equation models to measured variables⁽¹²⁴⁾. Monozygotic twins showed higher intra-pair correlations than dizygotic twins for most of the parameters. Genetic factors were responsible for between 46 and 70 % of variance (broad sense heritability) in parameters such as mean temperature, mesor, acrophase, Rayleigh test, percentage of rhythmicity and 5 h of maximum temperature⁽¹²⁴⁾. The pattern of correlations and the genetic models point to moderate-to-high heritability for most of the WT parameters, suggesting a relevant genetic influence. The presence of these genetic factors points to endogenicity as the main cause of the coincidence of the WT rhythms⁽¹²⁴⁾. However, some WT parameters are still dependent on environment to a relevant extent and, hence, more amenable to change through external interventions⁽¹²⁴⁾.

Discussion

Circadian alignment is crucial for body-weight management, and for metabolic health. In this context, circadian alignment consists of alignment of sleep, meal patterns and physical activity. Research on these topics mainly consists of research on misalignment. For instance, circadian misalignment appears to affect sleep-architecture in that REM sleep becomes phase advanced to SWS with reduced REM sleep latency. REM sleep duration increases during phase delay, resulting in a shorter REM sleep duration during the second part of the night. When sleep architecture is affected by circadian misalignment, it affects substrate oxidation, leptin- and ghrelin concentrations, appetite, food reward, HPA-axis activity and gut-peptide concentrations as such, that a positive energy balance is enhanced. Phase-advanced misalignment leads to

increased night-time cortisol exposure, increased HOMA-IR index, increased carbohydrate- and decreased protein-oxidation, as well as to food-reward deficiency^(42,45). Phase-delayed misalignment increases REM sleep, glucose concentrations and carbohydrate oxidation, and decreased GLP-1 concentrations and protein-oxidation^(42,45). The main effect of circadian misalignment, either phase advanced or phase delayed, is a concomitant disturbance of the glucose–insulin metabolism and substrate-oxidation. Chronically eating and sleeping at unusual circadian times may create a health risk through metabolic disturbance⁽⁴⁵⁾.

Since the relationship between circadian alignment and energy homeostasis appears in that daily patterns in activity, feeding, energy utilisation and energy storage are strongly synchronised by the SCN^(4,5), and since the SCN clock is entrained by light–dark cycles as well as by daily feeding cycles, this close interaction is critical for circadian regulation of metabolism, and partly underlies the disruption of proper metabolic rhythms observed in metabolic disorders, such as obesity and type-II diabetes^(4,10). Aligning meals in a circadian way requires timing of food intake, including regularity of meals, i.e. of meal frequency and meal intervals^(9,10,76). Over the longer term, in the perspective of dietary intervention for body-weight loss, the combination of energy and sleep restriction in overweight adults resulted in decreased loss of fat and considerably increased loss of fat-free body mass^(46,50). Moreover, a concomitant inverse correlation between changes in sleep duration and in body-weight, and respectively fat mass was shown, during weight maintenance, showing a concomitant improvement of body-composition and sleep duration^(58,117).

The surprisingly strong effect of regular physical activity during the day on the stability and amplitude of circadian rhythm⁽¹²³⁾, may serve as an instrument to restore poor circadian rhythms. We suggest that primarily regular physical activity throughout and during the day stimulates the amplitude and stability of the circadian rhythm, which will be enhanced by a regular sleep pattern with sufficient sleep duration, taking into account the sleep hygiene⁽⁴⁶⁾, and a fixed and regular meal pattern. These environmental variables that support a stable circadian rhythm, likely operate in interaction with a genetic predisposition, suggested by a moderate-to-high heritability for most of the WT parameters indicating circadian rhythm⁽¹²⁴⁾. Endogeneity may be the main cause of the coincidence of the WT rhythms⁽¹²⁴⁾. However, the dependence of these WT parameters on environment make them amenable to change through external interventions⁽¹²⁴⁾.

In conclusion, notwithstanding the separate favourable effects of sufficient daily physical activity, regular meal patterns, sufficient sleep duration and QS on energy-balance, the overall effect of the amplitude and stability of the circadian rhythm, perhaps including genetic predisposition, may integrate the separate effects in an additive way.

Acknowledgements

Many thanks to my colleagues Dr Hanne Gonnisson, Dr Eveline Martens, Dr Claire Mazuy, Dr Rick

Hursel, Dr Sanne Verhoef, Dr Femke Rutters, and Dr Tanja Adam, for collaboration on the topics in this field. The technicians Loek Wouters and Paul Schoffelen are thankfully acknowledged.

Financial Support

None.

Conflicts of Interest

None.

Authorship

The paper was written solely by M. S. W-P.

References

1. Adamantidis A & de Lecea L (2008) Sleep and metabolism: shared circuits, new connections. *Trends Endocrinol Metab* **19**, 362–370.
2. Froy O (2007) The relationship between nutrition and circadian rhythms in mammals. *Front Neuroendocrinol* **28**, 61–71.
3. Wolk R & Somers VK (2007) Sleep and the metabolic syndrome. *Exp Physiol* **92**, 67–78.
4. Bechtold DA (2008) Energy-responsive timekeeping. *J Genet* **87**, 447–458.
5. Laposky AD, Bass J, Kohsaka A *et al.* (2008) Sleep and circadian rhythms: key components in the regulation of energy metabolism. *FEBS Lett* **582**, 142–151.
6. Garaulet M, Ordovas JM & Madrid JA (2010) The chronobiology, etiology and pathophysiology of obesity. *Int J Obes* **34**, 1667–1683.
7. Kohsaka A, Laposky AD, Ramsey KM *et al.* (2007) High-fat diet disrupts behavioral and molecular circadian rhythms in mice. *Cell Metab* **6**, 414–421.
8. Cagampang FR & Bruce KD (2012) The role of the circadian clock system in nutrition and metabolism. *Br J Nutr* **108**, 381–392.
9. Mendoza J (2007) Circadian clocks: setting time by food. *J Neuroendocrinol* **19**, 127–137.
10. Esquirol Y, Bongard V, Mabile L *et al.* (2009) Shift work and metabolic syndrome: respective impacts of job strain, physical activity, and dietary rhythms. *Chronobiol Int* **26**, 544–559.
11. Scheer FA, Hilton MF, Mantzoros CS *et al.* (2009) Adverse metabolic and cardiovascular consequences of circadian misalignment. *Proc Natl Acad Sci U S A* **106**, 4453–4458.
12. Mendoza J, Pevet P & Challet E (2008) High-fat feeding alters the clock synchronization to light. *J Physiol* **586**, 5901–5910.
13. Arble DM, Bass J, Laposky AD *et al.* (2009) Circadian timing of food intake contributes to weight gain. *Obesity (Silver Spring)* **17**, 2100–2102.
14. Kok P, Roelfsema F, Frolich M *et al.* (2008) Short-term treatment with bromocriptine improves impaired circadian

- growth hormone secretion in obese premenopausal women. *J Clin Endocrinol Metab* **93**, 3455–3461.
15. Schibler U, Ripperger J & Brown SA (2003) Peripheral circadian oscillators in mammals: time and food. *J Biol Rhythms* **18**, 250–260.
 16. Hirota T & Fukada Y (2004) Resetting mechanism of central and peripheral circadian clocks in mammals. *Zoolog Sci* **21**, 359–368.
 17. Knutsson A & Boggild H (2010) Gastrointestinal disorders among shift workers. *Scand J Work Environ Health* **36**, 85–95.
 18. Hoogerwerf WA (2009) Role of biological rhythms in gastrointestinal health and disease. *Rev Endocr Metab Disord* **10**, 293–300.
 19. Szosland D (2010) Shift work and metabolic syndrome, diabetes mellitus and ischaemic heart disease. *Int J Occup Med Environ Health* **23**, 287–291.
 20. Rutters F, Gerver WJ, Nieuwenhuizen AG *et al.* (2010) Sleep duration and body-weight development during puberty in a Dutch children cohort. *Int J Obes* **34**, 1508–1514.
 21. Knutson KL (2005) The association between pubertal status and sleep duration and quality among a nationally representative sample of U. S. adolescents. *Am J Hum Biol* **17**, 418–424.
 22. Thorleifsdottir B, Bjornsson JK, Benediktsson B *et al.* (2002) Sleep and sleep habits from childhood to young adulthood over a 10-year period. *J Psychosom Res* **53**, 529–537.
 23. Lumeng JC, Somashekar D, Appugliese D *et al.* (2007) Shorter sleep duration is associated with increased risk for being overweight at ages 9 to 12 years. *Pediatrics* **120**, 1020–1029.
 24. DiVall SA & Radovick S (2009) Endocrinology of female puberty. *Curr Opin Endocrinol Diab Obes* **16**, 1–4.
 25. Lewis K & Lee PA (2009) Endocrinology of male puberty. *Curr Opin Endocrinol Diab Obes* **16**, 5–9.
 26. Vanitallie TB (2006) Sleep and energy balance: interactive homeostatic systems. *Metabolism* **55**, S30–S35.
 27. Mesarwi O, Polak J, Jun J *et al.* (2013) Sleep disorders and the development of insulin resistance and obesity. *Endocrinol Metab Clin North Am* **42**, 617–634.
 28. Nixon GM, Thompson JM, Han DY *et al.* (2008) Short sleep duration in middle childhood: risk factors and consequences. *Sleep* **31**, 71–78.
 29. Spiegel K, Leproult R & Van Cauter E (1999) Impact of sleep debt on metabolic and endocrine function. *Lancet* **354**, 1435–1439.
 30. Van Cauter E (2011) Sleep disturbances and insulin resistance. *Diab Med* **28**, 1455–1462.
 31. de Jong E, Stocks T, Visscher TL *et al.* (2012) Association between sleep duration and overweight: the importance of parenting. *Int J Obes* **36**, 1278–1284.
 32. Matricciani L, Blunden S, Rigney G *et al.* (2013) Children's sleep needs: is there sufficient evidence to recommend optimal sleep for children? *Sleep* **36**, 527–534.
 33. Javaheri S, Storer-Isser A, Rosen CL *et al.* (2011) Association of short and long sleep durations with insulin sensitivity in adolescents. *J Pediatr* **158**, 617–623.
 34. Klingenberg L, Chaput JP, Holmback U *et al.* (2013) Acute sleep restriction reduces insulin sensitivity in adolescent boys. *Sleep* **36**, 1085–1090.
 35. Matthews KA, Dahl RE, Owens JF *et al.* (2012) Sleep duration and insulin resistance in healthy black and white adolescents. *Sleep* **35**, 1353–1358.
 36. Hicks RA, McTighe S & Juarez M (1986) Sleep duration and eating behaviors of college students. *Percept Mot Skills* **62**, 25–26.
 37. Hogenkamp PS, Nilsson E, Nilsson VC *et al.* (2013) Acute sleep deprivation increases portion size and affects food choice in young men. *Psychoneuroendocrinology* **38**, 1668–1674.
 38. St-Onge MP (2013) The role of sleep duration in the regulation of energy balance: effects on energy intakes and expenditure. *J Clin Sleep Med* **9**, 73–80.
 39. Wells TT & Cruess CD (2006) Effect of partial sleep deprivation on food consumption and food choice. *Psychol Health* **21**, 79–86.
 40. Leproult R, Copinschi G, Buxton O *et al.* (1997) Sleep loss results in an elevation of cortisol levels the next evening. *Sleep* **20**, 865–870.
 41. St-Onge MP, O'Keefe M, Roberts AL *et al.* (2012) Short sleep duration, glucose dysregulation and hormonal regulation of appetite in men and women. *Sleep* **35**, 1503–1510.
 42. Gonnissen HK, Hulshof T & Westerterp-Plantenga MS (2013) Chronobiology, endocrinology, and energy- and food-reward homeostasis. *Obes Rev* **14**, 405–416.
 43. Gonnissen HK, Hursel R, Rutters F *et al.* (2013) Effects of sleep fragmentation on appetite and related hormone concentrations over 24 h in healthy men. *Br J Nutr* **109**, 748–756.
 44. Gonnissen HK, Mazuy C, Rutters F *et al.* (2013) Sleep architecture when sleeping at an unusual circadian time and associations with insulin sensitivity. *PLoS ONE* **8**, e72877.
 45. Gonnissen HK, Rutters F, Mazuy C *et al.* (2012) Effect of a phase advance and phase delay of the 24-h cycle on energy metabolism, appetite, and related hormones. *Am J Clin Nutr* **96**, 689–697.
 46. Nedeltcheva AV, Kilkus JM, Imperial J *et al.* (2009) Sleep curtailment is accompanied by increased intake of calories from snacks. *Am J Clin Nutr* **89**, 126–133.
 47. Bosy-Westphal A, Hinrichs S, Jauch-Chara K *et al.* (2008) Influence of partial sleep deprivation on energy balance and insulin sensitivity in healthy women. *Obes Facts* **1**, 266–273.
 48. Weiss A, Xu F, Storer-Isser A, *et al.* (2010) The association of sleep duration with adolescents' fat and carbohydrate consumption. *Sleep* **33**, 1201–1209.
 49. St-Onge MP, Roberts AL, Chen J *et al.* (2011) Short sleep duration increases energy intakes but does not change energy expenditure in normal-weight individuals. *Am J Clin Nutr* **94**, 410–416.
 50. Nedeltcheva AV, Kilkus JM, Imperial J *et al.* (2010) Insufficient sleep undermines dietary efforts to reduce adiposity. *Ann Intern Med* **153**, 435–441.
 51. Taheri S, Lin L, Austin D *et al.* (2004) Short sleep duration is associated with reduced leptin, elevated ghrelin, and increased body mass index. *PLoS Med* **1**, e62.
 52. Chaput JP, Despres JP, Bouchard C *et al.* (2008) The association between sleep duration and weight gain in adults: a 6-year prospective study from the Quebec Family Study. *Sleep* **31**, 517–523.
 53. Omisade A, Buxton OM & Rusak B (2010) Impact of acute sleep restriction on cortisol and leptin levels in young women. *Physiol Behav* **99**, 651–656.
 54. Schmid SM, Hallschmid M, Jauch-Chara K *et al.* (2008) A single night of sleep deprivation increases ghrelin levels and feelings of hunger in normal-weight healthy men. *J Sleep Res* **17**, 331–334.
 55. Schmid SM, Hallschmid M, Jauch-Chara K *et al.* (2009) Short-term sleep loss decreases physical activity under free-living conditions but does not increase food intake under time-deprived laboratory conditions in healthy men. *Am J Clin Nutr* **90**, 1476–1482.

56. Morselli L, Leproult R, Balbo M *et al.* (2010) Role of sleep duration in the regulation of glucose metabolism and appetite. *Best Pract Res* **24**, 687–702.
57. Hursel R, Rutters F, Gonnissen HK *et al.* (2011) Effects of sleep fragmentation in healthy men on energy expenditure, substrate oxidation, physical activity, and exhaustion measured over 48 h in a respiratory chamber. *Am J Clin Nutr* **94**, 804–808.
58. Verhoef SP, Camps SG, Gonnissen HK *et al.* (2013) Concomitant changes in sleep duration and body weight and body composition during weight loss and 3-mo weight maintenance. *Am J Clin Nutr* **98**, 25–31.
59. Chaput JP, Despres JP, Bouchard C *et al.* (2011) Longer sleep duration associates with lower adiposity gain in adult short sleepers. *Int J Obes (Lond)* **36**, 752–756.
60. Chaput JP & Tremblay A (2012) Sleeping habits predict the magnitude of fat loss in adults exposed to moderate caloric restriction. *Obes Facts* **5**, 561–566.
61. Carskadon MA & Dement WC (2011) Monitoring and staging human sleep. In *Principles and Practice of Sleep Medicine*, pp. 16–26 [E Roth & WC Dement, editors]. St. Louis: Elsevier Saunders.
62. Lee ML, Swanson BE & de la Iglesia HO (2009) Circadian timing of REM sleep is coupled to an oscillator within the dorsomedial suprachiasmatic nucleus. *Curr Biol* **19**, 848–852.
63. Monteleone P & Maj M (2008) The circadian basis of mood disorders: recent developments and treatment implications. *Eur Neuropsychopharmacol* **18**, 701–711.
64. Van Cauter E, Leproult R & Plat L (2000) Age-related changes in slow wave sleep and REM sleep and relationship with growth hormone and cortisol levels in healthy men. *JAMA* **284**, 861–868.
65. Emens J, Lewy A, Kinzie JM *et al.* (2009) Circadian misalignment in major depressive disorder. *Psychiatry Res* **168**, 259–261.
66. Peterson MJ & Benca RM (2006) Sleep in mood disorders. *Psychiatr Clin North Am* **29**, 1009–1032.
67. Akerstedt T & Gillberg M (1984) A dose-response study of sleep loss and spontaneous sleep termination. *Psychophysiology* **23**, 293–297.
68. Brunner DP, Dijk DJ & Borbely AA (1993) Repeated partial sleep deprivation progressively changes in EEG during sleep and wakefulness. *Sleep* **16**, 100–113.
69. Tilley AJ & Wilkinson RT (1984) The effects of a restricted sleep regime on the composition of sleep and on performance. *Psychophysiology* **21**, 406–412.
70. Elmenhorst EM, Elmenhorst D & Luks N (2008) Partial sleep deprivation: impact on the architecture and quality of sleep. *Sleep Med* **9**, 840–850.
71. Wyatt JK, Ritz-De Cecco A & Czeisler CA (1999) Circadian temperature and melatonin rhythms, sleep, and neurobehavioral function in humans living on a 20-h day. *Am J Physiol* **277**, R1152–R1163.
72. Brass SD & Auerbach S (2009) A sleepy patient with REM rebound. *J Clin Sleep Med* **5**, 386–389.
73. Knutson KL, Van Cauter E, Zee P *et al.* (2011) Cross-sectional associations between measures of sleep and markers of glucose metabolism among subjects with and without diabetes: the Coronary Artery Risk Development in Young Adults (CARDIA) Sleep Study. *Diab Care* **34**, 1171–1176.
74. Wu H, Stone WS, Hsi X *et al.* (2011) Effects of different sleep restriction protocols on sleep architecture and daytime vigilance in healthy men. *Physiol Res* **59**, 821–829.
75. Koren D, Levitt Katz LE, Brar PC *et al.* (2011) Sleep architecture and glucose and insulin homeostasis in obese adolescents. *Diab Care* **34**, 2442–2447.
76. Huang W, Ramsey KM & Marcheva B (2011) Circadian rhythms, sleep, and metabolism. *J Clin Invest* **121**, 2133–2141.
77. Czeisler CA & Klerman EB (1999) Circadian and sleep-dependent regulation of hormone release in humans. *Recent Prog Horm Res* **54**, 97–132.
78. Kalsbeek A & Strubbe JH (1999) Circadian control of insulin secretion is independent of the temporal distribution of feeding. *Physiol Behav* **63**, 553–558.
79. Morgan L, Hampton S, Gibbs M *et al.* (2003) Circadian aspects of postprandial metabolism. *Chronobiol Int* **20**, 795–808.
80. van Aggel-Leijssen DP, van Baak MA, Tenenbaum R *et al.* (1999) Regulation of average 24 h human plasma leptin level; the influence of exercise and physiological changes in energy balance. *Int J Obes Relat Metab Disord* **23**, 151–158.
81. Schoeller DA, Cella LK, Sinha MK *et al.* (1997) Entrainment of the diurnal rhythm of plasma leptin to meal timing. *J Clin Invest* **100**, 1882–1887.
82. Lecoultré V, Ravussin E & Redman LM (2011) The fall in leptin concentration is a major determinant of the metabolic adaptation induced by caloric restriction independently of the changes in leptin circadian rhythms. *J Clin Endocrinol Metab* **96**, E1512–E1516.
83. Wong ML, Licinio J, Yildiz BO *et al.* (2004) Simultaneous and continuous 24-hour plasma and cerebrospinal fluid leptin measurements: dissociation of concentrations in central and peripheral compartments. *J Clin Endocrinol Metab* **89**, 258–265.
84. Cinti S, Matteis RD, Pico C *et al.* (2000) Secretory granules of endocrine and chief cells of human stomach mucosa contain leptin. *Int J Obes Relat Metab Disord* **24**, 789–793.
85. Bado A, Levasseur S, Attoub S *et al.* (1998) The stomach is a source of leptin. *Nature* **394**, 790–793.
86. Cinti S, de Matteis R, Ceresi E *et al.* (2001) Leptin in the human stomach. *Gut* **49**, 155.
87. Kojima M & Kangawa K (2002) Ghrelin, an orexigenic signaling molecule from the gastrointestinal tract. *Curr Opin Pharmacol* **2**, 665–668.
88. Cowley MA, Smith RG, Diano S *et al.* (2003) The distribution and mechanism of action of ghrelin in the CNS demonstrates a novel hypothalamic circuit regulating energy homeostasis. *Neuron* **37**, 649–661.
89. Chen HY, Trumbauer ME, Chen AS *et al.* (2004) Orexigenic action of peripheral ghrelin is mediated by neuropeptide Y and agouti-related protein. *Endocrinology* **145**, 2607–2612.
90. Hagemann D, Meier JJ, Gallwitz B *et al.* (2003) Appetite regulation by ghrelin – a novel neuro-endocrine gastric peptide hormone in the gut-brain-axis. *Z Gastroenterol* **41**, 929–936.
91. Yi CX, Challet E, Pevet P *et al.* (2008) A circulating ghrelin mimetic attenuates light-induced phase delay of mice and light-induced Fos expression in the suprachiasmatic nucleus of rats. *Eur J Neurosci* **27**, 1965–1972.
92. Yannielli PC, Molyneux PC, Harrington ME *et al.* (2007) Ghrelin effects on the circadian system of mice. *J Neurosci* **27**, 2890–2895.
93. Cummings DE, Purnell JQ, Frayo RS *et al.* (2001) A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. *Diabetes* **50**, 1714–1719.
94. Westerterp-Plantenga MS, Miedema MJ & Wijckmans-Duijsens NE (1996) The role of macronutrient selection in determining patterns of food intake in obese and non-obese women. *Eur J Clin Nutr* **50**, 580–591.
95. Dos Santos ML, Aragon FF, Padovani CR *et al.* (2006) Daytime variations in glucose tolerance in people with

- impaired glucose tolerance. *Diab Res Clin Pract* **74**, 257–262.
96. Fletcher EC (1997) Sympathetic activity and blood pressure in the sleep apnea syndrome. *Respiration* **64**, Suppl. 1, 22–28.
 97. Ribeiro DC, Hampton SM, Morgan L *et al.* (1998) Altered postprandial hormone and metabolic responses in a simulated shift work environment. *J Endocrinol* **158**, 305–310.
 98. DeSantis AS, DiezRoux AV, Hajat A *et al.* (2012) Associations of salivary cortisol levels with inflammatory markers: the Multi-Ethnic Study of Atherosclerosis. *Psychoneuroendocrinology* **37**, 1009–1018.
 99. Yki-Jarvinen H, Bogardus C & Howard BV (1987) Hyperglycemia stimulates carbohydrate oxidation in humans. *Am J Physiol* **253**, E376–E382.
 100. Farshchi HR, Taylor MA & Macdonald IA (2005) Beneficial metabolic effects of regular meal frequency on dietary thermogenesis, insulin sensitivity, and fasting lipid profiles in healthy obese women. *Am J Clin Nutr* **81**, 16–24.
 101. Farshchi HR, Taylor MA & Macdonald IA (2004) Regular meal frequency creates more appropriate insulin sensitivity and lipid profiles compared with irregular meal frequency in healthy lean women. *Eur J Clin Nutr* **58**, 1071–1077.
 102. Farshchi HR, Taylor MA & Macdonald IA (2004) Decreased thermic effect of food after an irregular compared with a regular meal pattern in healthy lean women. *Int J Obes Relat Metab Disord* **28**, 653–660.
 103. Arnold LM, Ball MJ & Duncan AW (1993) Effect of isoenergetic intake of three or nine meals on plasma lipoproteins and glucose metabolism. *Am J Clin Nutr* **57**, 446–451.
 104. Jenkins DJ & Jenkins AL (1995) Nutrition principles and diabetes. A role for “lente carbohydrate”? *Diab Care* **18**, 1491–1498.
 105. Jenkins DJ, Wolever TM, Vuksan V *et al.* (1989) Nibbling versus gorging: metabolic advantages of increased meal frequency. *N Engl J Med* **321**, 929–934.
 106. Jenkins DJ, Wolever TM, Ocana A, *et al.* (1990) Metabolic effects of reducing rate of glucose ingestion by single bolus versus continuous sipping. *Diabetes* **39**, 775–781.
 107. Wolever TM (1990) Metabolic effects of continuous feeding. *Metabolism* **39**, 947–951.
 108. Bertelsen J, Christiansen C, Thomsen C *et al.* (1993) Effect of meal frequency on blood glucose, insulin, and free fatty acids in NIDDM subjects. *Diab Care* **16**, 4–7.
 109. Thomsen C, Christiansen C, Rasmussen OW *et al.* (1997) Comparison of the effects of two weeks’ intervention with different meal frequencies on glucose metabolism, insulin sensitivity and lipid levels in non-insulin-dependent diabetic patients. *Ann Nutr Metab* **41**, 173–180.
 110. McGrath SA & Gibney MJ (1994) The effects of altered frequency of eating on plasma lipids in free-living healthy males on normal self-selected diets. *Eur J Clin Nutr* **48**, 402–407.
 111. Jones PJ, Leitch CA & Pederson RA (1993) Meal-frequency effects on plasma hormone concentrations and cholesterol synthesis in humans. *Am J Clin Nutr* **57**, 868–874.
 112. Verboeket-van de Venne WP & Westerterp KR (1991) Influence of the feeding frequency on nutrient utilization in man: consequences for energy metabolism. *Eur J Clin Nutr* **45**, 161–169.
 113. Verboeket-van de Venne WP, Westerterp KR & Kester AD (1993) Effect of the pattern of food intake on human energy metabolism. *Br J Nutr* **70**, 103–115.
 114. Westerterp-Plantenga MS, Kovacs EM & Melanson KJ (2002) Habitual meal frequency and energy intake regulation in partially temporally isolated men. *Int J Obes Relat Metab Disord* **26**, 102–110.
 115. Garrow JS, Durrant M, Blaza S *et al.* (1981) The effect of meal frequency and protein concentration on the composition of the weight lost by obese subjects. *Br J Nutr* **45**, 5–15.
 116. Westerterp-Plantenga MS, Nieuwenhuizen A, Tome D *et al.* (2009) Dietary protein, weight loss, and weight maintenance. *Annu Rev Nutr* **29**, 21–41.
 117. Westerterp-Plantenga MS, Goris AH, Meijer EP *et al.* (2003) Habitual meal frequency in relation to resting and activity-induced energy expenditure in human subjects: the role of fat-free mass. *Br J Nutr* **90**, 643–649.
 118. Melanson KJ, Westerterp-Plantenga MS, Saris WH *et al.* (1999) Blood glucose patterns and appetite in time-blinded humans: carbohydrate versus fat. *Am J Physiol* **277**, R337–R345.
 119. Melanson KJ, Westerterp-Plantenga MS, Campfield LA *et al.* (1999) Blood glucose and meal patterns in time-blinded males, after aspartame, carbohydrate, and fat consumption, in relation to sweetness perception. *Br J Nutr* **82**, 437–446.
 120. Melanson KJ, Westerterp-Plantenga MS, Campfield LA *et al.* (1999) Appetite and blood glucose profiles in humans after glycogen-depleting exercise. *J Appl Physiol* **87**, 947–954.
 121. Chapelot D, Marmonier C, Aubert R *et al.* (2006) Consequence of omitting or adding a meal in man on body composition, food intake, and metabolism. *Obesity* **14**, 215–227.
 122. Smeets AJ & Westerterp-Plantenga MS (2008) Acute effects on metabolism and appetite profile of one meal difference in the lower range of meal frequency. *Br J Nutr* **99**, 1316–1321.
 123. Tranel HR, Schroder EA, England J *et al.* (2015) Physical activity, and not fat mass is a primary predictor of circadian parameters in young men. *Chronobiol Int* **32**, 832–841.
 124. Lopez-Minguez J, Ordoñana JR, Sánchez-Romera JF *et al.* (2015) Circadian system heritability as assessed by wrist temperature: a twin study. *Chronobiol Int* **32**, 71–80.