

NANO REVIEW

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Smart nanomaterial and nanocomposite with advanced agrochemical activities

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Abstract

Conventional agriculture solely depends upon highly chemical compounds that have negatively ill-affected the health of every living being and the entire ecosystem. Thus, the smart delivery of desired components in a sustainable manner to crop plants is the primary need to maintain soil health in the upcoming years. The premature loss of growth-promoting ingredients and their extended degradation in the soil increases the demand for reliable novel techniques. In this regard, nanotechnology has offered to revolutionize the agrotechnological area that has the imminent potential over conventional agriculture and helps to reform resilient cropping systems withholding prominent food security for the ever-growing world population. Further, in-depth investigation on plant-nanoparticles interactions creates new avenues toward crop improvement via enhanced crop yield, disease resistance, and efficient nutrient utilization. The incorporation of nanomaterial with smart agrochemical activities and establishing a new framework relevant to enhance efficacy ultimately help to address the social acceptance, potential hazards, and management issues in the future. Here, we highlight the role of nanomaterial or nanocomposite as a sustainable as well stable alternative in crop protection and production. Additionally, the information on the controlled released system, role in interaction with soil and microbiome, the promising role of nanocomposite as nanopesticide, nanoherbicide, nanofertilizer, and their limitations in agrochemical activities are discussed in the present review.

Keywords: Conventional agriculture, Agrochemicals, Nanomaterial, Crop improvement, Sustainable

Introduction

Globally, people are employed in agriculture for the cultivation of fundamental food crops and various essential forms of products such as fibers, fuels, fodders, and raw materials. Limited resources and an exponentially growing population, which is estimated to mark 9.6 billion by 2050, enforce the areas derived demanding the elaboration of very sustainable agriculture while permitting declination of global hunger and poverty [1, 2]. To fulfill this demand of relentlessly expanding population, there is an urgent prerequisite to enhance food production by more than 50% [2, 3]. Due to the limited number of natural resources (water, land, soil, forest, etc.) and ceiling in crop productivity, there is a huge demand for effective

agricultural approaches that are viable and liable economically and eco-friendly. To overcome these dilemmas, synthetic agrochemicals (herbicides, insecticides, fungicides, and fertilizers) have been developed and used to increase agricultural yields [4, 5]. However, the application of such agrochemicals had been instrumental for elevating food quality and quantity in past decades to evaluate the long-term ill effect of such agrochemicals on soil health and the ecosystem [6]. However, research on nanoparticle application as chemical alternatives for utility in the agriculture sector has become enhancing popularity over the past decade, later referred to as nanoagrochemicals [7]. The intentional and directional delivery in the environment, nanoagrochemicals may be considered specific in terms of expectable environmental issues, as they would represent the single diffuse cause of engineered nanoparticles (NPs) [8, 9]. Given this, one such initiative taken is the forefront of smart nanomaterials for revolutionizing

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current agriculture practices that contain good reactivity due to their substantial surface area to volume ratio and exceptional physicochemical characteristics that offer the novel advantage of modification according to increasing demand [2].

Modern agriculture is renovating into sustainable agriculture with the use of these modern age materials that are empowering to attain maximum output from limited resources [10]. Generally, agrochemical is essential to increase crop productivity but contrary, their application decline soil fertility by hindering soil mineral balance [11]. Moreover, the direct foliar or sprayed application can be cost-effective and very high, which run off and need to be controlled [12]. The nanomaterials-based chemicals developed in agriculture regulate nutrient depletion rate, yield reduction, input cost for crop raising, protection, production, and minimizing post-harvest loss [3]. Nanocomposites have become a key component of nanomaterials for scrutinizing and stimulating the plant life cycle because of their intrinsic unique thermal, electrical, chemical, and mechanical properties. The translocation in size-dependent lies in the range of 0.1–1000 nm within plant parts and altered according to surface compositions, a charge of NPs (highly negatively

charged shows more translocation), and plant size exclusion limit [10, 13]. These routes of penetration are confirmed via different in vitro (Filter paper, hydroponics, agar media, Hoagland solution, Mursashige and Skoog media, nutrient solution) and in vivo (foliar uptake, branch feeding, trunk injection, and root uptake) experiments using nanopesticide, nanoherbicide, nanoherbicides, and nanogrowth-promoting compounds [2, 9]. However, in certain cases the size exclusion is high so, it's difficult to limits the specific passage and concentration that affect the growth phase of plants both positively and negatively (Fig. 1).

Many successful examples of utilizing smart nanomaterial in agriculture have been reported in recent years including multi-walled carbon nanotubes [5, 14], metal-based nanocomposites [15], silver inhibits fungus germination [16], and many more. This new-age nanoformulation has the potential to fine-tune the physiology just entering the soil–plant complex that can be solely exploited to spotify the lateral effect [17].

The nanoparticle-based products (NMs) including smart agrochemical delivery systems having nanocomposites as chief ingredients are being constantly developed. Much intensive research is still required to achieve

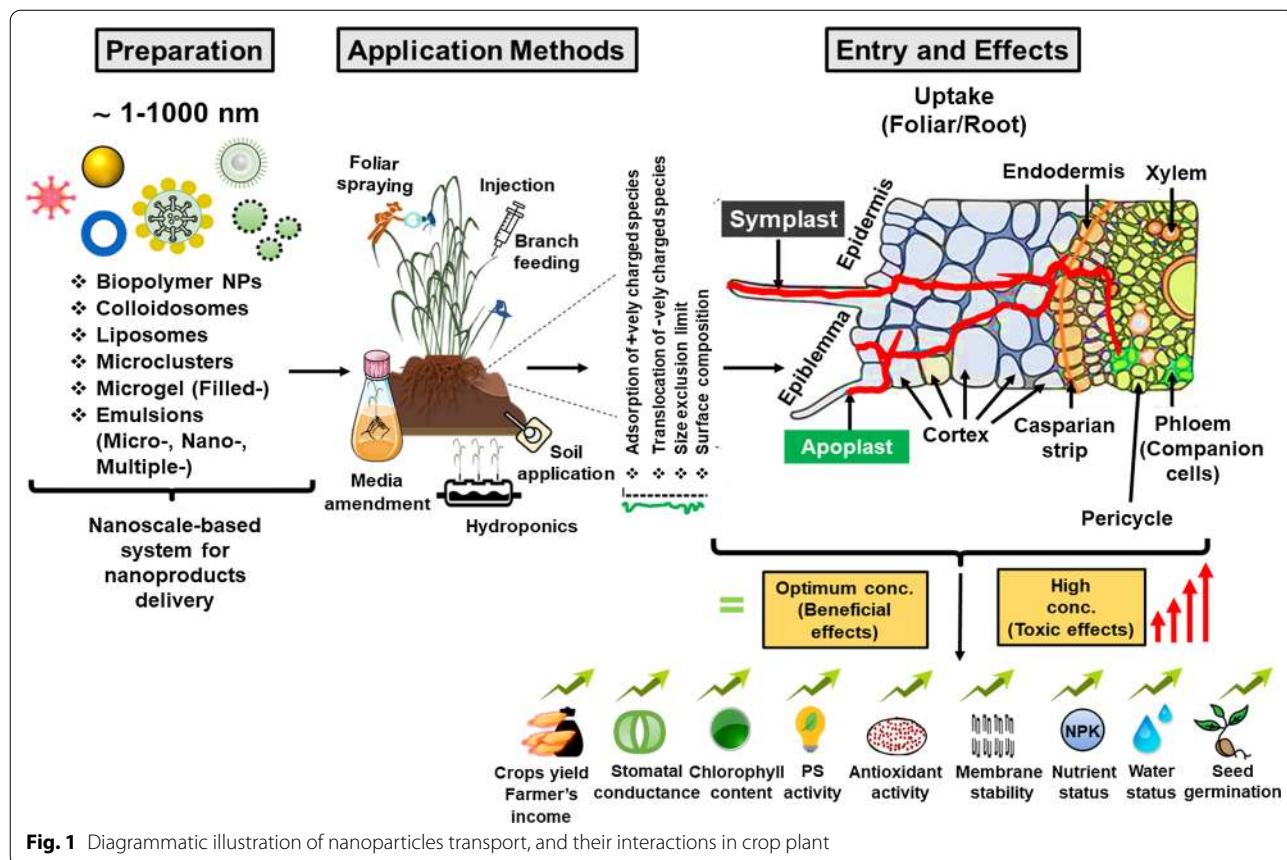


Fig. 1 Diagrammatic illustration of nanoparticles transport, and their interactions in crop plant

the practical advantages of nanoagrochemicals with improved working design, regulation of commercialization, and risk assessment of nanofertilizer, nanopesticide, and nanoherbicide [18, 19]. New crop cultivars, that can sustain heat, drought, salinity, and other unresolved challenges in farming systems disturb the whole spectrum of major cultivation practices worldwide. Moreover, it is expected that the implementation of NMs in the natural environment decline the chemicals-based hazardous level [12]. We surely believe, their application in agriculture will narrow down the gap between sustainable and chemical-based agriculture systems. Besides this, it boosts food production and quality globally in an eco-friendly manner by resolving water and soil contamination [20]. Thus, practically they could provide novel avenues regarding developing new NMs-based products [14]. Conventional agrochemical has offered numerous drawbacks regarding the non-selectively and adsorption rate of active ingredients (AIs).

It has been reported that more than 99.9% pesticides are failed to be delivered at target sites and cause a hazardous impact on the health of the soil, water, air with enhances pathogenic resistance and biodiversity loss [12, 21, 22]. Overall, we aimed to highlight the current information on facts that nanomaterial or nanocomposite deliver an efficient solution to upgrade and advanced the agriculture innovations, food systems, sustainable crop protection, and production. Moreover, information on the controlled released system, role in interaction with soil and microbiome, the promising role of nanocomposite as nanopesticide, nanoherbicide, nanofertilizer, and limitation in agrochemical activities are also discussed in the present review.

Nanostructure compounds with the controlled released system (CRS)

Due to several advantages over conventional chemical application approaches, many researchers have put forward the model of the controlled release system [15, 23–29] to offer substitutes to reduce environmental pollution. The controlled release (CR) allows efficient delivery of an AI more actively in soil and plant for the desired interval of time, resulting in the decreases of the amounts of agrochemicals used, energy, manpower, or other resources crucial to operate the application instruments as well as in enhancement in safety to humans who deal with their application [26, 29–32]. Additionally, CR shows many advantages over conventional methods including decrease phytotoxicity, reduce agrochemical loss due to volatilization, lixiviation, drift, improper handling, and degradation in soil and controlled delivery coincides with a suitable concentration in the plant to

prevent unpredictable losses in form of evaporation, leaching and weather (Fig. 2) [16, 33].

Comprehensive characterization is a significant prerequisite to predict or explain the efficiency and behaviour of smart nano-loaded agrochemicals. In particular, retention of AIs, behaviour, composition and phase, zeta potential, and internal structure of polymeric nanocarriers, and their release in particle environment conditions are summarized as important properties [30, 34–36]. The rate of loading and release for AIs from nanocarriers plays a central role in predicting or assessing their efficacy. These can be evaluated by ingredients concentration remaining within polymeric matrix and amount of released ingredients [37, 38]. The mechanism of release can be achieved via different modes such as:

Diffusion via relaxation/swelling of NPs

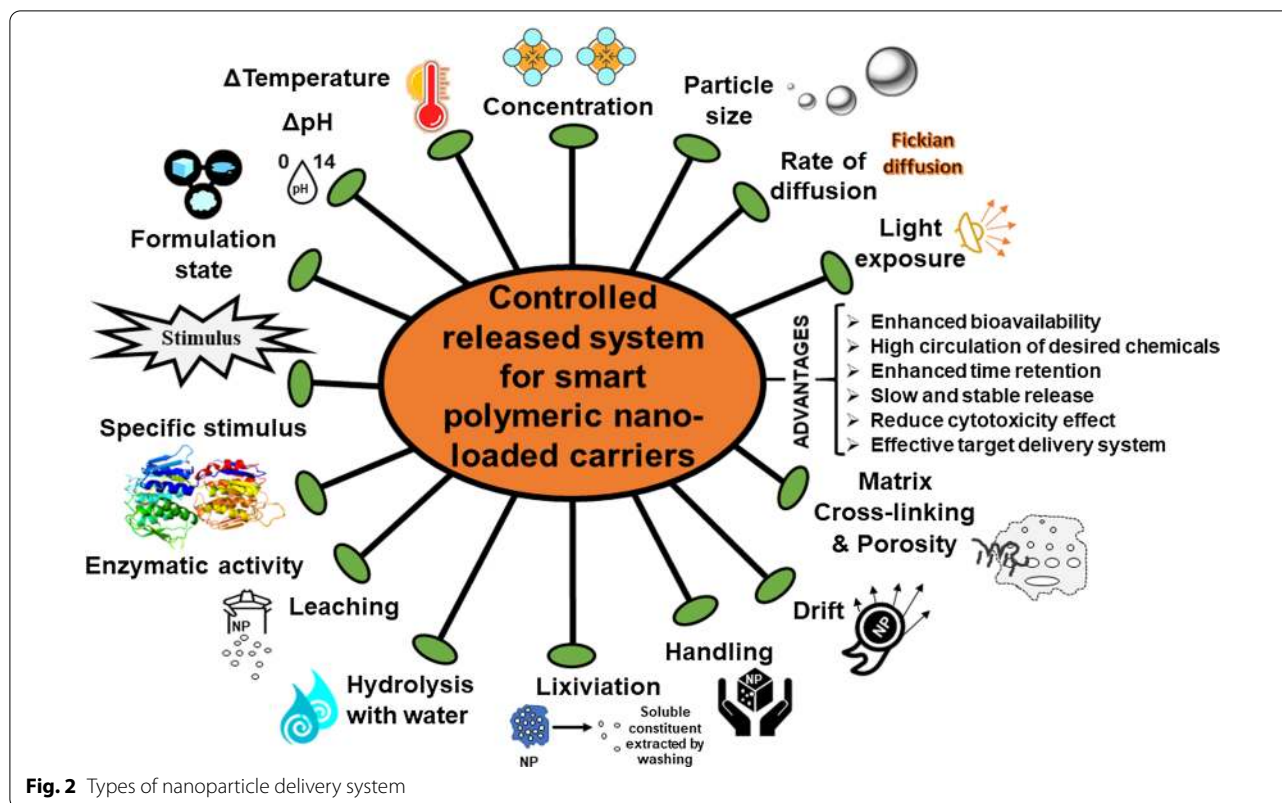
In the concentration gradient phenomena (or fickian diffusion), the release would occur at a high rate when nanocarriers are diluted using either concentrated or solid formulations even under irrigation or rainfall events. The diffusion can be slowdown by enhancing the nanoparticle size or enhancing the distance within media in which diffusion of AI occurs observed in poly lactic acid (PLA) loaded metazachlor [32, 39, 40]. Similarly, enhanced cross-linking has been suggested as an efficient method to delay diffusion by increasing the tortuosity or decreasing the porosity via the polymer matrix, as indicates by methomyl-loaded chitosan (azidobenzaldehyde-carboxymethyl) pesticide before and after polymer crosslinking [40–43].

Burst release

The most commonly rapid release method in which AI release undesirably, if an initial high amount of AI is not favorable for the application of target. The phenomena would show enhance the concentration of AIs present near or on the surface of the NPs indicates high significant burst release. For example, PLA-loaded metazachlor (herbicides) nanocapsule or surface coating has been recommended to inhibits the initial rapid burst that is frequently noted for nanospheres [35].

Degradation

Nanoparticle release can be triggered or accelerated by physical, chemical, and biological degradation that can be achieved by hydrolysis with water, light exposure, temperature, pH, specific stimulus, and enzymatic activities. For example, PLGA (Poly lactic co-glycolic acid) NPs show increased hydrolytic degradation with enhancing surface area- volume ratio for water, and their diffusion rate might be fine-tuned with appropriate nanocarriers [44]. Moreover, the mPEG (methoxy polyethylene



glycol) incorporated in PLGA-NPs increases the degradation rate of NPs via enhanced hydrophilicity and ultimately accessibility for hydrolysis in hydrolytic degradation type. In enzymatic degradation, the events lead by the activities of phosphatases, glycosidases, and protease viz: PCL (poly(ϵ -caprolactone) degradation enhance with the activity of lipase activity [44]. Similarly, γ -PGA (poly (γ -glutamic acid) degradation mediated by γ -GTP (γ -glutamyl transpeptidase) is considered as a most common enzyme that causes rapid degradation [38]. In another study, zein nanoparticle shows rapid and extensive degradation and release of encapsulated ciprofloxacin antibiotic, in presence of trypsin enzyme than collagenase [37].

In some cases, stimuli-response release can be observed using photosensitive polymers such as micellar or UV (Ultraviolet) labile core-shell NPs were produced to PEG and nitrobenzyl to carboxymethyl chitosan. Thus, stimuli-based nanocomposite can intelligently react to the stimulus produced by the target or the adjoining environment that eventually triggers the AIs release to regulate the pest effectively [45, 46]. However, physical stability in some NPs altered by pH, when the polymer is weak basic or acidic such that electrostatic and charge will reliable on pH [40, 41, 47]. For instance, carboxymethyl cellulose and feather keratin were loaded with

ivermectin. The diffusion rate was observed to be faster at low pH (Fickian transport) and higher pH (non-Fickian) [46].

Nanoformulations as a promising tool in an agricultural system

Agrochemicals includes pesticides, herbicides, fungicides, bactericides, nematocides, rodenticides that are used to target pest, weed, pathogenic fungus, bacteria, nematodes and rodents (Fig. 3) [48–50]. Globally, the herbicide market is expanding and is estimated to lies between \$27.21 and \$39.15 billion in a compound annual growth rate (CAGR) of 6.25% in the expected period 2016–2022. Besides this, the global pesticides market was accounted to reach \$70.57 billion by 2021 at a CAGR of 5.15% estimated between 2016 and 2021. Besides this, the global market of encapsulated pesticides grows exponentially at reach benchmark of US \$800 million by 2025 expectedly and gains 11.8% CAGR in the tenure of 2019–2025 [18, 19, 48, 49].

The families represented by inorganic chemicals are triazines, phenoxy, and benzoic acid chloroacetanilides representing herbicides, phenylpyrrole, benzimidazoles, dithiocarbamates, and nitriales for fungicide, carbamate, organophosphates, organochlorines relating to insecticide. Smart nanoagrochemicals with nanoformulations

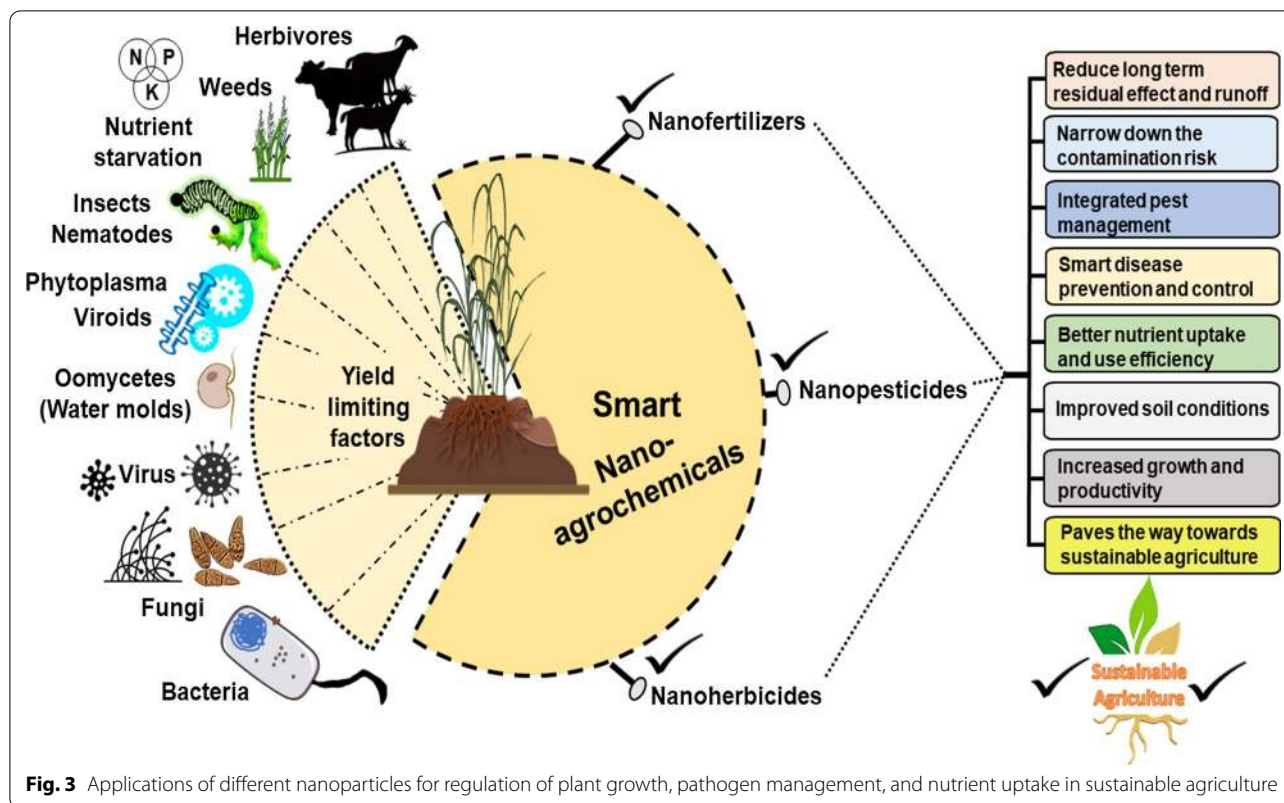


Fig. 3 Applications of different nanoparticles for regulation of plant growth, pathogen management, and nutrient uptake in sustainable agriculture

must offer a broad variety of benefits including enhanced durability, effectiveness, wettability, good dispersion, less toxicity, good biodegradable ability in soil and environment, and photogenerative nature with the least residues compared to conventional chemicals [51–53]. Over the past, extensive studies were carried out on nanoagrochemicals to access their significant role and contamination range in affecting soil–plant nutrient cycles [19].

Nanopesticide

The potential utility of nanochemicals in integrated pest management (IPM) depends upon targeted delivery of AIs with increased activity at least drug concentration and proficient monitoring of pesticides interactions with the surroundings. Under harsh conditions, the chemical stability can be achieved by efficient nanocarriers having enhanced dispersal range, wettability, and more protectivity to pesticides without risk of runoff [54–57]. Other noteworthy characteristics of pesticidal nanocompositions can be observed in thermal stability, large surface area, increased target affinity, and biodegradable nature after successful delivery. These delivery systems can be regulated for single goals or multiple combinations viz; spatially target release, time-controlled release, remotely or self-regulated release to overcome the biological barriers in the successful target

[21, 58–60]. However, the efficacy of nanoencapsulation or nanocarriers is (1) to prevent pre-degradation of AI in the carrier before their release in the target (2) to improve penetration and ease solubility of AIs within the target site (3) to monitor or regulate the degradation of AIs in the desired site [61, 62].

According to Kremer et al. [63] the adsorptive interaction between pesticides and NPs showing discrete molecular dynamics. Such interactions should have a positive impact on adsorption sites via physiological morphology, binding ability, antioxidant systems, and transportability of pesticides in plants [64]. In *Arabidopsis thaliana*, the antagonistic effect between silver NPs and Diclofop-methyl (post-emergence herbicide) in which herbicides presence decline or affected the Ag^+ from silver NPs. Moreover, a decrease in pesticide concentration is imperative to avoid their toxicity on non-selected organisms and narrow down contamination risk [65–67]. Several nanocompositions of pesticides have been developed such as nanoemulsions, nanosuspensions, and nanocapsulations. Such nanomaterials are prepared specifically to maintain the regulated release of AIs in several ways including magnetic release, ultrasound release, pH release, heat release, moisture release, DNA-based release, specific release, quick and slow-release [19].

In some cases, nanoparticle delivery in hollow silica NPs are used to prevent avermectin from UV radiation and provide photostability to nanopesticides causes long-term effects on the target organism. Several NPs used various forms of encapsulations including (1) Lipid nanomaterial-based encapsulation. (2) Metal–organic framework-based encapsulation. (3) Polymer-based encapsulation. (4) Clay nanomaterial-based encapsulation. (4) Greener encapsulation [9, 42, 43, 45, 47, 68–70].

Nanofertilizer

Besides plant protection, these smart NPs are extensively used to regulate the physiological process. For example, SiO₂ NPs (silicon dioxide NPs) elevates seed germination rate in *Lycopersicon esculentum* [71, 72], chitosan-polymethacrylic-NPK increase biomass, nutrient uptake and antioxidant enzymes in *Phaseolus vulgaris* [73, 74], Au-NPs (gold NPs) promotes seed germination, seedling growth, enzymatic activity and nutrient uptake in *Zea mays* [75, 76], SiO₂-NPs improve uptake of NPK, increase enzymatic activity and seed germination rate in *Hyssopus officinalis* and *Z. mays* [77–79], chitosan-CuNPs (copper NPs) enhance seed germination, activation of α -amylase, protease and activity of various antioxidant enzymes in *Z. mays* [2, 80, 81], chitosan-ZnNPs (zinc NPs) increase accumulation of zinc content and defense enzymes in *Triticum durum* [82, 83], chitosan- γ -polyglutamic acid-gibberellic acid NPs promotes seed germination, root development, leaf area, hormonal efficiency, extracellular enzymes and nutrient efficiency [83, 84], Chitosan-polymethacrylic acid-NPK NPs promotes protein content and nutrient uptake [74, 85], ZnO-NPs (zinc oxide NPs) increase activity of catalase (60.7%), superoxide dismutase (22.8%) and nutrient acquisition [86, 87], CeO₂-NPs (cerium oxide NPs) enhance seed germination and vigour, enzymatic activity and nutrient uptake in *Spinacia oleracea* and *Z. mays* [88–91], AuNPs increase chlorophyll content and antioxidant enzyme activities in *Brassica juncea* [92] and TiO₂ NPs (titanium oxide NPs) enhance chlorophyll content, nutrient uptake, activity of Rubisco and antioxidant enzymes in *S. oleracea* and *Cicer arietinum* [89, 93] (Table 1).

Nanoinsecticides

As the trends and demand of encapsulated NPs exponentially increased the regulatory pressure for their management also enhanced simultaneously. Encapsulated insecticides share more than 42% of total pesticide revenue up to 2017 [60, 94, 95]. Recently, in 2019 pesticide manual online classified encapsulated insecticides contain hazardous toxic AIs like pendimethalin, acetochlor, dichlobenil, tefluthrin, etofenprox, chlorpyrifos, carbosulfan, and furathiocarb at the commercial level [19]. The

toxicity level of AIs not only depends upon encapsulation material but it helps in adjusting the dynamics of the target species exposure to AIs in vivo conditions [21, 25, 96]. The use of styrene and methylmethacrylate as encapsulation wall material increased the nematocidal activity to suppress the growth of the wheat rust-causing pathogen, *Puccinia reconditea*. Similarly, the effect of urea–formaldehyde and polyuria resin wall on stomatal toxicity, contact toxicity, phoxim loaded microcapsule efficacy, and photolysis properties was reported by Zhang et al. [97]. In another study, improved pest efficiency and poor cytotoxicity of sodium alginate imidachloroprid encapsulation were observed that favored direct application of imidachloroprid [68].

Another study shows a decrease in picloram toxicity to soil microbiota with silica gel encapsulation in comparison to free-form picloform. The silica NPs bioavailability to the non-selected organism can be enhanced by tuning the wall properties of the silica shell [98]. In a study, Jacques et al. [99] reported the atrazine toxicity in encapsulated polymeric and lipid nanocompositions against nematodes, *Caenorhabditis elegans*, but comparably no toxicity was observed in tripolyphosphate/chitosan-based encapsulation that itself can be attributed to low toxicity. Moreover, the oil encapsulated PCL neem-derived nanoencapsulation did not exhibit any adverse effect of stomatal conductance, the photosynthetic ability of maize after exposure up to 300 days. These findings suggest the careful selection of wall material/encapsulation and physicochemical properties of AIs and their composition and application sites [19, 100].

The Si-NPs (silicon NPs) have been efficiently reported to protect infestation from stored beetle *Callosobruchus maculatus* in pulses like *Vigna unguiculata*, *V. mungo*, *V. radiate*, *Macrotyloma uniflorum*, *C. arietinum*, and *Cajanus cajan* [101]. Despite their excellent performance, nanopesticides show poor commercialization and stability. The pH, temperature, humidity, UV radiation influence AIs availability and influence physiochemical characteristics. Besides these quantity, quality, strict legislation, expensiveness and degradation period of AIs are emerging issues while using nanopesticides [19, 54, 79].

Nanofungicides

Beyond the nanocarriers application, nanomaterial as AIs for crop protection is a major aspect of research. The broad spectrum of antifungal properties of nanofungicides can improve their efficiency as a pesticide. For instance, copper, silver, and zinc NPs resolve the disadvantages of chemical AIs for pathogenic resistance with sharp antimicrobial activity and non-toxicity [19]. Moreover, chitosan-based NPs (Ch-NPs) showed effective antifungal activity and restrict growth reported by many

Table 1 Successful use of nanoformulation used in crop plant as plant growth promoters

Nanoformulation	Mode of applications	Targeted crop	Properties (size/shape/Molecular weight/ pH)	Effect on Plant physiological processes			Key references
				Growth phases	Enzyme activities	Nutrient uptake and release	
SiO ₂ NPs	Seed treatment	<i>L. esculentum</i> Mill	12 nm	Enhance seed germination	No visible effect on enzyme activity	–	[71, 72]
Nano-chitosan	Seed treatment	<i>C. arietinum</i> L	pH 4.8	Promote total biomass, germination, and vigor index up to (57%)	Increase activities of catalase and superoxide dismutase	–	[83, 170]
Chitosan-poly(2-methylacrylic acid)-NPK NP	Foliar spray	<i>P. vulgaris</i> L	20 nm	Enhance plant growth and total biomass	Enhance the antioxidant enzyme activities	Promote nutrient uptake and accumulation	[73, 74]
Au NPs	Seed imbibition	<i>Z. mays</i> L	10–30 nm, spherical	Promote germination and seedling growth	Promote activity of Superoxide dismutase, peroxidase, and catalase	Increased nutrient uptake of maize excluding iron	[75, 76]
SiO ₂ NPs	Seed imbibition	<i>H. officinalis</i> L	10–20 nm, spherical	Improve plant growth and seed germination	Enhance total soluble proteins	–	[77, 78]
SiO ₂ NPs	Seed Imbibition	<i>Z. mays</i> L	10–20 nm, spherical	Promotes seed germination	Enhanced the activities of antioxidant enzymes	Increase uptake of nitrogen, phosphorus, and potassium	[77, 79]
Chitosan-Cu NPs	Seed treatment	<i>Z. mays</i> L	Low molecular weight, 80%	Promote seedling growth	Enhance activity of α-amylase and protease	–	[2, 80]
Chitosan-Cu NPs	Foliar spray	<i>Z. mays</i> L	50–190 kDa, 80%	Promote seedling growth, overall plant height, and biomass	Enhance activities of defense enzymes	–	[2, 81]
Chitosan-Zn NPs	Foliar spray	<i>T. durum</i> L	60 kDa, 85%	Stomatal localization of nanoparticles	Promote defense enzyme activities	Enhance zinc content accumulation by 42%	[82, 83]
Nano-chitosan	Seed treatment	<i>Z. mays</i> L	pH 7.0–9.0	Enhance plant height, leaf area, and seed germination	Promote activities of glucose-6-phosphate dehydrogenase, succinate dehydrogenase, and superoxide dismutase	Enhance accumulation of potassium inside the plant	[102, 171]
Chitosan-γ-polyglutamic acid-gibberellic Acid-NPs	Seed treatment	<i>P. vulgaris</i> L	290 kDa, 75%–85%, pH 4.5	Promote seed germination, root development, and total leaf area	Enhance the hormonal efficiency, enhance extracellular enzymes, such as cutinase, lipase, and esterase	Increase efficiency of nutrients	[83, 84]
Chitosan-gibberellic acid NPs	Seed treatment	<i>P. vulgaris</i> L	27 kDa, 75%–85%, pH 4.5	Promote leaf area, carotenoid and chlorophyll content	Enhance the hormonal efficiency by 90%	Not significant effect on nutrient uptake	[172, 173]
Nano-chitosan	Seed treatment	<i>P. vulgaris</i> L	100–399 kDa	Increase seed germination and radical length	Enhance the activity of peroxidase and catalase	Increase Zinc uptake in plant	[174, 175]

Table 1 (continued)

Nanoformulation	Mode of applications	Targeted crop	Properties (size/shape/Molecular weight/ pH)	Effect on Plant physiological processes		Key references
				Growth phases	Enzyme activities	
Nano-chitosan	Seed treatment	<i>Capiscum annumum</i> L	110 kDa, 85–90%, pH 4.0	Enhance the root biomass (77%) and fresh leaf biomass (28%)	Increase the activity of catalase and peroxidase 33% and 23% respectively	[176, 177]
Chitosan-polymethacrylic acid-NPK NP	Seed treatment	<i>Pisum sativum</i> var. <i>Master B</i>	20 nm	Enhance mitotic cell division about 1.5 fold	Enhance total soluble proteins like legumin, convicilin and β , vicilin 1, 2 and 3	[74, 85]
ZnO NPs	Seed soaked	<i>Avena sativa</i> L	20–50 nm, spherical shape	Promote percent germination	–	[178, 179]
ZnO NPs	Seed imbibition	<i>T. aestivum</i> L	30–40 nm, spherical crystal	Improve shoot length and total plant biomass	Enhance the activity of superoxide dismutase (22.8%) and catalase (60.7%)	[86, 87]
Chitosan-thiamine NP	Seed treatment	<i>C. arietinum</i> L	27 kDa, 85%	Promote seed germination and plant growth	Enhance activities of peroxidase, polyphenol oxidase, chitinase, and protease enzyme	[83, 180]
CeO ₂ NPs	50 Mg-Ce per L hydroponic	<i>Z. mays</i> L	2–4 nm, crystal	Promote photosynthesis and gas exchange	Accumulation of hydrogen peroxidase enzyme	[91, 181]
CeO ₂ NPs	Foliar spray	<i>S. oleracea</i> L	4–7 nm	Enhance percent germination (4%) and vigor index	Catalase activity significantly increased	[89, 182]
Nano-chitosan	Foliar spray	<i>Coffea canephora</i> <i>Piere</i> var <i>Robusta</i>	600 kDa, 85%, pH 6.0	Increase (30–50%) chlorophyll content and photosynthetic rate (30%)	Enhance the enzymatic activities	[83]
Chitosan-polymethacrylic acid-NPK NP	Foliar spray	<i>T. aestivum</i> L	20 nm	Increase crop yield (50%) and harvest index (24%)	Increase polysaccharides and total saccharides (11%), nitrate reductase enzyme	[83]
AuNPs	Spray on leaves	<i>B. juncea</i> L. <i>Czern</i>	10–20 nm, spherical	Enhance chlorophyll content and plant growth	Enhance antioxidative enzymes, proline and hydrogen peroxide	[92]

Table 1 (continued)

Nanof ormulation	Mode of applications	Targeted crop	Properties (size/shape/Molecular weight/ pH)	Effect on Plant physiological processes			Key references
				Growth phases	Enzyme activities	Nutrient uptake and release	
Carbon (CNTs)	Seeds in culture media	<i>L. esculentum Mill</i>	Nanotubes	Enhance seed germination and vigor index	Increase activities of peroxidase, catalase, and superoxide dismutase	Percentage concentration of nutrient elements present in germinated tomato increased	[182]
TiO ₂ NPs	Seeds soaked	<i>S. oleracea L</i>	30–60 nm, crystal shape	Enhance seedling growth, biomass, and chlorophyll content	Promote activities of antioxidant enzymes (0.25%)	Enhance nutrient uptake	[88]
TiO ₂ NPs	Spray on leaves	<i>C. arifinum L</i>	5–20 nm	Reduce membrane damage during cold stress	Increase activity of Rubisco enzyme	Increase the mineral uptake in plant	[183]

research workers in the last decade. For example, Ch-NPs against *Alternaria alternata*, *Macrophomina phaseolina*, *Rhizoctonia solani* [102], *Pyricularia grisea*, *Alternaria solani*, *Fusarium oxysporum* [102, 103], *Pyricularia grisea*, Copper–chitosan NPs against *Fusarium solani* [104], Cu-chitosan NPs- against *R. solani* and *Sclerotium rolfsii* [105], chitosan-saponin NPs [102], oleoyl-chitosan NPs against *Verticillium dahliae* [106], salicylic acid-loaded chitosan NPs against *Fusarium verticillioides* [107], Ag-chitosan NPs against *R. solani*, *Aspergillus flavus* and *A. alternata* [108], silica-chitosan NPs against *Phomopsis asparagi* [109] chitosan-pepper tree (*Schinus molle*) essential oil (CS-EO) NPs against *Aspergillus parasiticus* [110], chitosan boehmite alumina nanocomposites films and thyme oil against *Monilinia laxa* [111] fungicide zineb (Zb) and chitosan-Ag NPs against *Neoscytalidium dimidiatum* [112], chitosan-Thyme-oregano, thyme-tea tree and thyme-peppermint EO mixtures against *Aspergillus niger*, *A. flavus*, *A. parasiticus*, and *Penicillium chrysogenum*, [113], chitosan-thymol NPs against *Botrytis cinerea* [39], chitosan-*Cymbopogon martinii* essential oil against *Fusarium graminearum* [114].

In comparison to conventional agrochemicals, the nanoparticle was confirmed to be highly effective in crop protection even at minute concentration viz: 0.43 and 0.75 mg/plate concentration of Ag-doped hollow titanium-oxide (TiO₂) nanoformulation against Potato pathogens such as *Venturia inaequalis* and *F. solani* [115] (Table 2). Moreover, several successful examples of NPs were studied extensively for abiotic stress tolerance in recent years [116–118]. To cope with drought tolerance, several reports published in past decades on the application of NPs such as TiO₂ application in *Linum usitatissimum* via elevating pigmentation and reducing the activity of Malondialdehyde (MDA) and Hydrogen peroxide (H₂O₂) [119], ZnO promotes effective seed germination in *Glycine max* [120], CuNPs improve pigmentation, biomass and grain yield in *Z. mays* [121]. In case of salinity stress, seed soaking, nutrient solutions, and seed priming methods are used for evaluation in *G. max*, *S. lycopersicum*, and *Gossypium hirsutum* respectively [122–124].

The application improves stress tolerance by enhancing chlorophyll content, biomass number, soluble sugar content, seed germination [125–127]. According to Shoemaker [128] application of AgNPs (silver NPs) in *Triticum aestivum* increases seedling growth and leaf area whereas foliar application of SeNPs (selenium NPs) improves antioxidant enzyme activity and thylakoid membrane stability in *Sorghum bicolor* under heat stress [129] (Table 3).

Nanoherbicide

These NPs inhibit the physiological processes and growth phases in several weed species. For example, Ch-NPs retard germination and growth phases in *Bidens pilosa* [130, 131] NPs atrazine disrupts PSII activity in *Amaranthus viridis* [132], Fe₃O₄ NPs (Iron oxide NPs) + purified diatomite + glyphosate decrease pH level in *Cynodon dactylon* [133], Zero valent Fe NPs (Iron NPs) retard germination in *Lolium perenne* [32]. The efficacy of metribuzan, (a commercial herbicide) was enhanced via using NPs to maintain the growth of the weed population including *Melilotus album*, *T. aestivum*, *Agrostis stolonifera*, and *Setaria macrocheata* [19].

The atrazine-loaded nanocarriers are used to penetrate the stomatal region, hydathodes and ensure their direct entry into vascular tissues. It ensures the targeting, cellular uptakes, and overcomes intracellular trafficking due to certain properties of NPs: (1) Interaction affinity. (2) Mechanical effect of form and size. (3) catalytic effect. (4) Surface charges/hydrophobicity. Fraceto et al. [19] describing decreased toxicity level of paraquat in non-targeted plants preferring Triphosphate/chitosan nanocarriers application over conventional spray system in *Brassica* sp. Similarly, in *B. pilosa* and *C. dactylon* mortality rate of seedlings was enhanced using encapsulated glyphosate magnetic nanocarriers [19, 131]. The nanoencapsulation uses low doses of herbicide and could effectively reduce the long-term residual effect of herbicides in target species as well as in agricultural land. Conclusively, nanoherbicide can enhance the delivery of AIs in plant tissues and comparatively declined the chance of environmental toxicity [60, 94, 95].

Impact on plants-soil microbiome

NPs face numerous experience transformation, dissolution aggregation in soil microbiota, adsorption with key regulators that mediate the fate of degradation for organic content, pH, divalent cations, and clay (most important for retention of NPs). According to Asadishad et al. [134], the toxicity of AgNPs depends upon microbial substrate-dependent respiration toward ammonia-oxidizing bacteria decreased with elevation pH content and clay content. Low pH causes the dissolution of AgNPs whereas high soil pH value enhances the negative charge site numbers and leads to increase Ag sorption [19]. In a study, similar results were reported about CuONPs (Copper oxide NPs) on low clay content and organic matter with coarse soil texture. Such acidic soil favors the dissolution of Ag and CuNPs with free ionic liberation, which can elevate the short-duration impact of NPs [9]. Zhai et al. [135] also concluded that nanoformulations of ionic pesticides can show the variable impact, more commonly associated with the fractional ion release. Other authors

Table 2 Successful application of nanocomposites for biotic stress tolerance

Pathogen type	Nanoparticles used	Plant disease	Mechanism of action	Key references
<i>Fungus</i>				
<i>Bipolaris sorokiniana</i>	AgNPs biosynthesized with <i>Serratia</i> sp.	Spot blotch pathogen of wheat	Enhance lignification of vascular bundles	[92]
<i>Gloeophyllum abietinum</i>	Green-synthesize AgNPs extracted with turnip leaf	Wood-rotting	Inhibit the conidia development	[184]
<i>Phytophthora capsici</i>	Ag core-DHPAC shell nanocluster	Blight diseases in Solanaceae	Reduce mycelial growth and sporangial production	[185]
<i>Escherichia coli</i> , <i>Bacillus Subtilis</i> and <i>F. oxysporum</i>	Cu(OH) ₂ NPs	Corn leaf blight	Decrease number of conidia	[125]
<i>F. oxysporum</i>	Cu ₃ (PO ₄) ₂ ·3H ₂ O nanosheets	Root fungal disease in watermelon	Inhibit the fungus growth	[125]
<i>F. graminearum</i>	Multiwalled carbon nanotubes, graphene oxide, reduced graphene oxide, and fullerene	Fusarium head blight in wheat	Inhibit spore germination of <i>Fusarium graminearum</i>	[96]
<i>F. oxysporum</i>	CeO ₂ NPs	Panama disease	Enhance antioxidant enzyme activity	[186]
<i>Aspergillus</i> spp.	SiNPs	Black mold	Inhibit fungus proliferation	[187]
<i>R. solani</i>	Calcium carbonate	Brown rot of stems	Reduce rot growth and recover sucrose level	[188]
<i>Phytophthora</i>	Green-synthesize AgNPs extracted with <i>Artemisia absinthium</i>	Seed rots	Effect zoospore development	[189]
<i>Bacteria</i>				
<i>X. perforans</i>	Ag nanoparticles along with graphene oxide	Bacterial spot of tomato	Significantly decrease the activity of <i>X. perforans</i>	[190]
<i>B. sorokiniana</i>	AgNPs biosynthesized with <i>Serratia</i> sp.	Spot blotch pathogen of wheat	Inhibit conidial germination	[191]
<i>Clavibacter michiganensis</i>	CuNPs and K ₂ SiO ₃ NPs	Bacterial ring rot in potato	Decrease bacterial cell viability	[192]
<i>X. perforans</i>	Photochemically active TiO ₂ NPs	Spot disease in tomato	Due to high photocatalytic activity, reduction in bacterial spot	[193]
<i>Ralstonia solanacearum</i>	MgONPs	Vascular wilt disease	Inhibit bacterial activity	[185]
<i>Xanthomonas campestris</i> pv. <i>campestris</i>	Silver (Ag) NPs	Bacterial blight	Enhance antioxidant enzyme activity	[194]
<i>Colletotrichum gloeosporioides</i>	Chitosan NP	Disease in Chile	Inhibition growth of mycelia	[195]
<i>A. alternata</i>	Chitosan NP	Leaf spot	Inhibit spore germination	[83]
<i>Xanthomonas alfalfae</i>	Synthesized Mg(OH) ₂ NPs	Bacterial leaf spot	Significantly decrease the activity of <i>X. alfalfae</i>	[11]
Target species	Nanoparticles used	Mechanism of action	References	
<i>Insects</i>				
<i>Aedes aegypti</i> and <i>Anopheles stephensi</i>	Microbial synthesized Ag, Au, and ZnO-NPs	Epithelial cell, midgut, cortex damage, and thorax shape change		[196]
<i>Aedes albopictus</i> and <i>Culex pipiens pallens</i>	Ag synthesized using <i>Cassia fistula</i> extract	Total protein level, acetylcholinesterase, and α- and decreased activity of β-carboxylesterase		[197]
<i>Chironomus riparius</i>	AgNPs	Modulates GST genes expression, upregulated mRNA expression in delta3, Sigma4 and Epsilon1 GST class		[198]
<i>C. riparius</i>	AgNPs	Downregulated activity of ribosomal gene protein, activation of gonadotrophin through upregulation of Balbiani ring protein gene (CrBR2.2) and gonadotrophin-releasing hormone gene (CrGnRH1)		[198]
<i>C. riparius</i>	AgNPs	Enhance expression of epsilon-1, sigma-4, and delta-3 and transcript levels of catalase, thioredoxin reductase 1, Mn superoxide dismutase		[198]

Table 2 (continued)

Target species	Nanoparticles used	Mechanism of action	References
<i>Drosophila melanogaster</i>	AgNPs	Reduce Cu-dependent enzyme activity, couple with membrane-bound Cu transport protein results in Cu sequestration	[199]
<i>D. melanogaster</i>	AgNPs	Cause pigmentation defects and flies locomotive ability	[200]
<i>D. melanogaster</i>	AgNPs	DNA-damage, autophagy, ROS-mediated apoptosis	[201]
<i>D. melanogaster</i>	Ag and TiO ₂ NPs	Effect developmental processes of flies	[202]
<i>Sitophilus oryzae</i>	Nanostructured Al ₂ O ₃	Absorbing wax layer that results in insect dehydration	[93]
<i>Aedes albopictus</i>	Ag NPs prepared using 3,5-dinitrosalicylic acid and salicylic acid	Total protein, esterase, phosphatase, and acetylcholine esterase enzyme activity decreased	[203]

noted the difference and similarities of ionic and nanoforms of AgNPs with variation in antibacterial activity or the effect on a soil-borne microbial community and their response in in-vitro conditions [19, 136, 137].

In long-term studies, Guilger et al. [66], ensuring routes predictably depend on biogenic NPs, that show the least effect on human cells and denitrification process but are likely to show more impact on plant fungus relationship. At the microscale level, denitrification is a prime microbial activity that gets affected by AgNPs by modulating hydric conditions, pH and creating a devoid zone for fundamental accessories (carbon, nitrate, and oxygen). However, by high soil redox potential value and sandy texture soil favored denitrification, whereas textured clay soils provided offers low redox potential and lies in range for biological transformation [19]. Such impact is correlated by the affinity of AgNPs to denitrification and physicochemical properties ex: surface charge, coating, size, sedimentation rate, dispersibility, and solubility [138]. The biogenic AgNPs are derived from the green process and have no effect on N-cycle reported by Kumar et al. [67]. While the effect of nanocapsules, nanogels, nanometal, and nonmetal particles on soil microbiota as non-selected microbes has been documented. Li et al. [139] evidenced the negative impact of nanopesticide CM- β -CD-MNPs-Diuron complex (carboxymethyl-hydroxypropyl- β -cyclodextrin magnetic NPs) on the activity of the urease enzyme.

The Diuron NPs complex causes declined in the population status of soil bacteria except for actinobacteria with an increase in reactive oxygen species. All these indicate toxicity of CM- β -CD-MNPs-Diuron exert stress on soil microbes and did not reduce even by using Diuron nanoencapsulation [12, 19]. The bionanopesticides treatment was confirmed to improve soil microbiome including weight gain and survival percentages in beneficial earthworm *Eudrilus eugeniae*. It also shows excellent

larvicidal, antifeedant, and pupicidal activities against *Helicoverpa armigera* and *Spodoptera* sp. at 100 ppm nanoformulation dose [19, 50, 55].

Drawbacks using nanoagrochemicals on plants

The nanopesticides are also showing some adverse effects on crop plants directly or indirectly. The most favorable and used AgNPs and their complex nanoparticle have been attributed to their diverse range in each class of pesticides due to low toxicity but still many reported published that explained the drawback of these smart nanoagrochemicals [61, 140, 141] (Table 4). For example, In *Vicia faba*, the AgNPs internalization in leaves can abrupt the stomatal conductance CO₂ assimilation rate and photosystem II [142]. Furthermore, the binding of AgNPs attaches with Chlorophyll forming a hybrid, that excites electrons 10 times due to fast electron-hole separation and plasmon resonance effect. In another study, AgNPs and AgNPs-graphene oxide GO (Ag@dsDNA GO) effect also observed in *L. esculentum* exhibit antibacterial activity toward *Xanthomonas perforans* [143]. Various reports were submitted in recent years such as ZnO NPs reduced root growth in *Allium cepa* [89], Ch-NPs + paraquat biomass reduction, lipid peroxidation, genotoxicity and leaf necrosis in *Brassica* sp. [144], SiO₂NPs affect biomass, germination, protein content, photosynthetic pigment in *Taraxacum officinale* and *Amaranthus retroflexus* [76], AgNPs cause lipid peroxidation, leaf damages and alters catalase activity in *G. max* [145], NPP ATZ + AMZ *Raphanus raphanistrum* suppresses plant growth [146].

Besides these, NPs show an adverse impact on plant physiology, soil microbiota, and declined enzymatic population. For instance; Al₂O₃ (Aluminium oxide) reduces bacterial growth and reduces seedling growth [147, 148], C60 fullerene restricts bacterial growth up to 20–30% [149], ZnNPs decrease enzymatic activities in soil and

Table 3 Successful application of nanocomposites for abiotic stress tolerance

Stress type	Nanoparticles	Plant species	Mode of application	Results	Key references	
Drought	TiO ₂	<i>L. usitatissimum</i> L.	Foliar spray	Enhance chlorophyll and carotenoid content as well as lowers the activity of MDA and H ₂ O ₂	[204]	
	TiO ₂	<i>T. aestivum</i> L. c.v Pishtaz	Foliar spray	Enhance starch content, growth, and yield	[76]	
	TiO ₂	<i>T. aestivum</i> L.	Amended soil	Increased seedling growth, antioxidant enzymes, total chlorophyll, and carotenoid content	[205]	
	ZnO	<i>G. max</i> L.	Seed soaking	Enhance germination rate and percent germination	[119]	
	Fe ₂ O ₃	<i>Mentha piperita</i> L.	Hoagland solution	Increase activity of antioxidant enzymes	[206]	
	Cu	<i>Z. mays</i> L.	Plant priming	Improve plant biomass, chlorophyll, anthocyanins, and grain yield	[207]	
	CNTs, graphene	<i>G. hirsutum</i> L.	Seed priming	Increase seedling growth and biomass	[123]	
	Chitosan NPs	<i>Hordeum vulgare</i> L.	Foliar spray	Increase proline content, CAT and SOD	[208]	
	Salinity	Ag	<i>Trigonella foenum-graecum</i>	Seed soaking	Improve percent germination, fresh and dry weight of seedlings	[209]
		ZnO	<i>Abelmoschus esculentus</i> L.	Foliar spray	Increase activity of superoxide dismutase, catalase, and photosynthetic pigments	[210]
ZnO and Si		<i>Mangifera indica</i> L.	Foliar spray	Enhance nutrient uptake, carbon assimilation in plants	[211]	
SiO ₂		<i>Solanum lycopersicum</i> L.	Seed soaking	Upregulation of stress tolerance genes	[212]	
SiO ₂		<i>Fragaria ananassa</i> L.	Soil application	Enhance growth, proline, chlorophyll, epicuticular wax layer and leaf relative water content	[213]	
SiO ₂		<i>Musa acuminata</i> L.	Seed priming	Enhance chlorophyll content, shoot growth	[214]	
Ag		<i>T. aestivum</i> L.	Seed priming	Enhance total soluble sugars, proline content, and peroxidase activity	[185]	
Fe ₂ O ₃		<i>Helianthus annuus</i> L.	Foliar spray	Enhance dry weight, leaf area, chlorophyll content	[215]	
Fe ₂ O ₃		<i>Dracocephalum moldavica</i> L.	Foliar spray	Increase the enzymatic activity of guaiacol peroxidase, catalase, ascorbate peroxidase, and glutathione reductase	[215]	
Mn		<i>C. annuum</i> L.	Nanoprimer	Improve plant growth	[216]	
CeO		<i>G. hirsutum</i> L.	Seed priming	Improve root growth and decrease ROS level	[121]	
CNTs, graphene		<i>Catharanthus roseus</i> L.	Murashige and Skoog medium	Enhance the number of leaves and flowers	[124]	
Chitosan-PVA and CuNPs		<i>S. lycopersicum</i> L.	Nutrient solution	Enhance chlorophyll, carotenoids, and lycopene content	[122]	
Heat		Ag	<i>T. aestivum</i> L.	Soil application	Promote the root number, seedling length, and leaf area	[127]
	Se	<i>S. bicolor</i> L. Moench	Foliar spray	Improve thylakoid membrane stability and activity of antioxidant enzymes	[217]	

Table 3 (continued)

Stress type	Nanoparticles	Plant species	Mode of application	Results	Key references
Heavy metal	Fe	<i>T. aestivum</i> L.	Soil application	Increase rate of photosynthesis, chlorophyll content, and plant growth	[87]
UV-B	Si	<i>T. aestivum</i> L.	Nutrient solution	Improve antioxidant defense system	[218]
Cold	TiO ₂	<i>C. arietinum</i> L.	Amended soil	Decrease MDA levels and electrolyte leakage index	[219]
Flooding	Al ₂ O ₃	<i>G. max</i> L.	Seed soaking	Increased hypocotyl length, mitochondrial membrane proteins	[220]

Table 4 Adverse effect of nanoparticles on targeted crop and soil health

NPs	Size (nm)	Targeted crop	Adverse effect on plant	Degradation time in soil (days)	Effect on soil	Key references
Al ₂ O ₃	50	<i>Nicotiana tabacum</i> L.	Reduce the germination percentage, biomass per seedling, and average root length	3	Reduce the activity of bacteria <i>Bacillus cereus</i> and <i>Pseudomonas stutzeri</i>	[147, 148]
C ₆₀ -fullerence	50	<i>G. max</i> (L.) Merr	Reduced biomass	60	Reduction of 20–30% in fast-growing protozoa and bacteria	[58, 149]
CuO, Ni, ZnO and Cr ₂ O ₃	100	<i>Oryza sativa</i> L.	Effect the activities of antioxidant enzymes in plant	24	Activity of enzyme dehydrogenase and urease reduced to 75% and 44% respectively	[221, 222]
ZnO and TiO ₂	10–20	<i>T. aestivum</i> L.	Reduced the root growth by 75%	60	Adversely affect the growth of earthworms, traces of ZnO and TiO ₂ were found inside the body	[61, 223]
Zn ²⁺ , Zn, and ZnO	50	<i>Z. mays</i> L.	50% reduction in photosynthesis, leaf stomatal conductance, transpiration rate, and intercellular CO ₂ concentration	56	Reduce enzymes like β-glucosidase, phosphatase, and dehydrogenase present in the soil	[51, 150]
nZVI (zero valent iron)	20–100	<i>Salix alba</i> L.	Effect seedling growth	7	At 750 mg/kg, mortality rate of <i>Lumbricus rubellus</i> and <i>Eisenia fetida</i> was 100%	[53, 224]
Au	25	<i>O. sativa</i> L.	Damage to the root cell wall due to accumulation of Au across xylem	30	Effect the soil microbes and edaphic factors of soil	[52, 225]
TiO ₂ , Ag, and CeO ₂	7–45	<i>A. cepa</i> L.	Increase in DNA damage as well as lipid peroxidation in roots	14	Reduced the survival, growth and fertility of nematodes	[226]
SnO ₂ , CeO ₂ and Fe ₃ O ₄	61 (SnO ₂), 50–100 (CeO ₂), 20–30 (Fe ₃ O ₄)	<i>Z. mays</i> L.	Fe ₃ O ₄ results in accumulation of Al in plant roots and negatively affects plant growth	63	Inhibits microbial growth	[141]
Ag	10–20	<i>P. vulgaris</i> L.	Disrupt chlorophyll synthesis, nutrient uptake, and hormone regulation	30	50% reduction in the activity of nitrifying bacteria	[158]

reduces transpiration rate and photosynthetic rate in *Z. mays* [150]. Conclusively, NPs are very reactive and variable in nature, so always a concerning risk for workers who may come across during their application.

Limitation and challenges at commercial scale implementation

As with documentation, the lack of finding on behavior and fate in the environment of nanoagrochemicals and their impact on faunal diversity may put challenges on their incorporation in agriculture. Instead of the benefits of using nanoencapsulation systems, their implementation requires caution, since it is mandatory to calculate their behavior in the environment and non-targeted communities to develop safer product development policies [54]. Although, it needs to develop smart nanoagrochemicals that are focused on biological nanoformulation and that offer a simple handling process, low cost, more AIs persistence with a sharp release system, and high degradation rate without leaving any residue [148]. Besides these, poor demonstrations at field conditions, cost-effectiveness, consumer acceptance, and feasibility of technology are major constraints on commercial implementation [152].

The limited management guidelines, inconsistency legislative framework, and regulatory models, and lack of public awareness campaign creates inconsistent marketing of such incipient nanoagricultural products. The national and international arrangement that fits at ground level is the only way that supports Nanotechnological development [49]. However, the community seeking approval for nanoagrochemicals must demonstrate the precautionary uses of these new products by proposing unjustifiable safety risks to the user and environment. Thus regulatory guidelines and frameworks are becoming primarily important to resolve the emerging issues of nanoagrochemicals [153]. Moreover, the need for collaboration, discussion, and information exchange forums among countries to ensure threat mitigating strategies should be considered as a milestone in nanoagrochemicals. So consolidates efforts of governmental organizations, scientists, and social communities are needed to preventing the adverse effect of nanoagrochemicals on humans and the environment [59].

In this scenario, the toxicity measuring instrumental setup is used in the characterization of toxicity type and their level to access the potential intrinsic hazards [59]. Currently, the main focus of experimental investigation on nanomaterial translocation in biotic/abiotic systems, monitoring and revealing interaction Among nanotoxicity and nanomaterial in the physical and chemical environment [48, 54, 151–153].

Transformation

Due to high reactivity, the interaction of nanocomponents with organic and inorganic components in the soil as well as for plants is undetermined and unregulated. The changing in physiochemical properties and transformation behavior after implementation creates chances of heavy metal toxicity. Biotransformation was demonstrated in *Cucumis sativa*, using CeO_2 bioavailability cause 20% to Ce(III) in the shoots and 15% of Ce(IV) being reduced to Ce(III) in the roots [154]. In another study, AgNPs were oxidized and forming the Ag-glutathione complex in the lettuce plant [154].

Accumulation of NPs

Because of variability in binding, the accumulation of NPs causes toxicity in plants, humans, and animals. In soybean, CeO_2 application shut down the Nitrogen fixation cycles and causes toxicity. However, ROS production, growth inhibition, cellular toxicity, and other phytotoxic effect were reported in *Amaranthus tricolor*. The application of C60 fullerene enhanced DDT accumulation in soybean, tomato, and zucchini plants [155].

Time to switch toward more sustainability

Most agrochemicals are not fully utilized by plants or seep off into the soil, air and water unintendedly causes toxic ill effects and accumulated through biomagnification. Moreover, global pesticide rise threatened biodiversity and led to the adverse effect on human intelligence quotient and fecundity in recent years. Still, it's also enhancement the resistance in weeds and plant pathogen against agrochemical turn them to super pathogen/weed. New doses after the changing in strategies of pathogens or new strain resurgence enhance cost-effectiveness and put the question on existing regulatory recommendations. [14, 106, 156–158].

The chemicals persist in soil particles, agricultural residues, irrigation water and migrates into the different layers of soils turns into a serious threat to the ecosystem. Leaching of synthetic pesticides, abructing soil-pest, soil-microbe activities, algal blooms formation, eutrophication, altering soil physiochemical properties [159], and salt toxicity via creating salt buildup in soil [160].

Low-cost oxides of Mg, Al, Fe, Ti, Ce, and Zn (Magnesium, Aluminium, Iron, Titanium, Cerium, Zinc) are ideal candidates and provides greater affinity, a large number of active sites, minimum intraparticle diffusion distance, and maximum specific surface area [160]. NP implementation help to successfully chase down the inorganic residues of various chemicals such as permethrin, 2–4 Dichlorophenoxy acetic acid (2–4-D), Dichlorodiphenyltrichloroethane (DCPT), Diuron (Adsorption),

Chlorpyrifos, Chloridazon, Methomyl (Photocatalysis) from the soil. Some nanocomposites are used for complete degradation of lethal agrochemicals for example silver-doped TiO_2 and gold doped TiO_2 , Zerovalent Fe (nZVI), endosulfan, TiO_2 , nZVI for atrazine, Ag for chlorpyrifos, Pd–Mg, Ni–Fe bimetallic system, nZVI for DDT, nZVI, nitrogen-doped TiO_2 , Fe–Pd (iron–palladium), Fe–S (Iron-sulfur) for Lindane [161] (Table 5).

Smart agrochemical: a step ahead toward more sustainability

Al-Barly et al. reported the slow release of nanocomposite fertilizers to depend upon phosphate and nitrogen content availability in soil [162]. TiO_2 NPs derived from *Moringa oleifera* leaf extract are used to control the red palm weevil (*Rhynchophorus ferrugineus*) and exhibits antioxidant and larvicidal activities. In the case of *Zanthoxylum rhoifolium*, nano-encapsulated essential oil was reported to maintain the population of *Bemisia tabaci* [19, 163]. Nanopesticides derived from pyrethrum insecticides cause an impact on the population status of honey bees. Except for these studies, agrochemical degradation can also be accomplished using adsorption, membrane filtration, catalytic degradation, oxidation, and biological treatment. Since, adsorption using smart Nanosorbents also relies on environmental factors including pH, temperature, and competitive adsorbing molecules [19]. At low pH, the protonated charged active site of NPs disturbs the binding ability of positively charge agrochemical whereas, high temperature creates hinders the electrochemical interactions between active sites and agrochemicals due to elevated vibrate energy of active site of adsorbent and kinetic energy of agrochemicals [79]. Moreover, chitosan-coated and cross-linked chitosan-Ag NPs used as composite microbeads that incorporated into reverse osmosis filters help in the effective removal of atrazine content from the water. According to Aseri et al. [164] integration of membrane filters and magnetic NPs-based beads enhances microbial elimination and resonance activation of water, respectively.

Secondly, targeting a not selected species with possible adverse effect is a key issue emerging that put a loophole of criticism for these smart nanoagrochemicals. For example; 1–10 mg L^{-1} of Polyhydroxybutyrate-co-hydroxyvalerate (PBHA) encapsulation for atrazine in lactuca sativa for 24 h reduced genotoxicity in plants [165], PCL atrazine nanocapsules ill effect on *Daphnia similis* and *Pseudokirchneriella subcapitata*, after exposure up to 24 h [166], Solid lipid NPs encapsulating simazine 0.025–0.25 mg mL^{-1} exhibits *Caenorhabditis elegans* Induction of mortality and decrease in the body length after exposure of 48 h [167]. The uncontrolled non-targeted release of AIs in plant cells causes

lysosomal damage with increasing pH. After the cellular compartment, nanoagrochemicals may bind or channelization into cell organelles and causes damage to protein, pigments, and DNA [98].

The binding ability of nanocompositions with selected and non-selected binding helps to recognize its distribution, bioavailability, toxicity level, and exclusion from the plant cell. Several proteins acquire a wide range of functional and structural properties including ligand bonding, metabolite production, catalysis, cellular and molecular reorganization [19]. The protein-nanopesticide complex can cause minor structural configuration and denaturation of proteins. Similarly, conformational changes and movement of the genomic DNA mediated through NPs also induced cytogenetic abnormalities. These nanopesticide toxicity are solely dependent upon the balance between key factors like biodegradability, concentration, and size of incorporated AIs. In *Prochilodus lineatus* 20 $\mu\text{g L}^{-1}$ concentration using PCL nanocapsules containing atrazine up to 24–48 h declined toxicity, as they did not induce carbonic anhydrase activity, alterations in glycemia and antioxidant response [168], in *Enchytraeus crypticus* causes a decrease in hatching due to the delayed number of adults and juveniles [19, 158, 169].

No doubt, intervention of nanoagrochemicals, resolve many threats mitigation put forward by the implementation of agrochemical but still more validation is required to lowering the agroecological risks. The persistent use of novel monitoring applications always knocks down the door of improvement of sustainable crop production and protection without creating the threats of NPs as a new contaminant.

Conclusion and future perspectives

During the entire course of million years of evolution, the green plants had evolved without any interference from other eukaryotes. However, for the last fifty years, continuous human activities have introduced many contaminants in the environment that altered the ecological balance and raised the eye-brows of researchers towards combating the new pathovars and pathotypes. These thrusting biological stresses have severely damaged global crop production. Concerning, the environmental penalty of conventional agrochemicals at present, nanoformulations seem to be a potential applicant for plant protection. The use of controlled biodegradable polymers especially polyhydroxyalkanoates shows significant and attractive properties of biocompatibility, biosorption rate, low-cost synthesis, thermoplastic nature, and ease in biodegradation rate that have popular advantages conventional chemical delivery systems. However, sustainable and efficient utilization with promising target delivery and low toxic effects are prerequisites of commercial

Table 5 Agrochemicals (insecticides, herbicides, and other fungicides) used to regulate the activity of crop pests under a sustainable agriculture approach

Chemical	Trade manufacture company	Nanocomposites used	Crop	Target	Key references
<i>Insecticides</i>					
<i>Inorganic</i>					
Chlorpyrifos	Dow Chemical Company	PVC	<i>G. hirsutum</i> L.	<i>Aphis gossypii</i> , <i>Spodoptera frugiperda</i> , and <i>Lygus lineolaris</i>	[227]
Chlorfenapyr	Dow AgroSciences	Chitosan/PLA	<i>Solanum melongena</i> L.	<i>Pseudococcidae</i>	[228]
	Super Bio Tech Marketing Company	Silica	<i>Brassica rapa</i>	<i>Helicoverpa armigera</i>	[94]
Avermectin	Super Bio Tech Marketing Company	Polystyrene nanoparticles (PHSN)	<i>G. hirsutum</i> L.	<i>Tetranychus urticae</i>	[229]
	Super Bio Tech Marketing Company	Polydopamine	<i>Brassica oleraceae</i>	<i>Thysanoptera</i>	[230]
Azadirachtin	Ecobiocides & Botanicals Pvt Ltd	Chitosan	<i>Ricinus communis</i> L.	<i>Spodoptera litura</i>	[231]
Deltamethrin	Crop Chemicals India Limited	Chitosan-coated beeswax SLN (Solid-liquid nanoparticles)	<i>G. hirsutum</i> L., <i>S. lycopersicum</i> L.	<i>Helicoverpa zea</i> , <i>Leucinodes orbonalis</i>	[185]
Imidacloprid	Chemets Wets & Flows Pvt. Ltd	Sodium alginate	<i>Nicotiana glauca</i> L.	<i>Cicadellidae</i>	[232]
Geraniol	Otto Chemie Pvt Ltd	Chitosan/Gum Arabic	<i>G. hirsutum</i> L.	<i>Bemisia tabaci</i>	[233]
Nicotine	Alchem International Pvt. Ltd	Chitosan/TPP	<i>S. lycopersicum</i> L.	<i>Musca domestica</i>	[234]
<i>Organic</i>					
Garlic essential oil	Arishtha Organics Pvt	PEG	<i>O. sativa</i> L.	<i>Tribolium castaneum</i>	[235]
<i>A. arborescens</i> L. essential oil	Priority Biocidal, LLC	SLN	<i>S. lycopersicum</i> L.	<i>Sitophilus zeamais</i>	[236]
Nanopermethrin	Jerobin J (Hamad medical corporation)	PEG	–	<i>Culex quinquefasciatus</i>	[237]
Geranium essential oils	India aroma oils and company	PEG	<i>T. aestivum</i> L.	<i>Rhyzopertha dominica</i>	[238]
Citrus peel essential oil	India aroma oils and company	PEG	<i>S. lycopersicum</i> L.	<i>Tuta absoluta</i>	[239]
<i>Rosmarinus officinalis</i> essential oil	Rosemary essential oil manufacturers & oem manufacturers India	PEG	<i>O. sativa</i> L.	<i>Tribolium castaneum</i>	[240]
<i>Herbicides</i>					
Paraquat	Syngenta	Montmorillonite	<i>G. max</i> L.	<i>Plantago lanceolata</i> L	[241]
	Syngenta	Chitosan/tripolyphosphate	<i>Brassica rapa</i> L.	Soil sorption microalga	[242]
Atrazine	Syngenta	Poly(ϵ -caprolactone)	<i>S. bicolor</i> L.	<i>Stellaria media</i> L., <i>Trifolium repens</i> L., <i>Lamium amplexicaule</i> L	[63]
		SLN	<i>Brassica napus</i>	<i>Raphanus raphanistrum</i> L	
		Poly (lactic-co-glycolic acid)	<i>Solanum tuberosum</i> L.	<i>Croton setigerus</i> L., <i>Oxalis corniculata</i> L	
Imazapic, Imazapyr	Avansagro chemicals shanghai limited	Alginate/chitosanChitosan/tripolyphosphate	<i>A. cepa</i> L.	<i>B. pilosa</i> L	[243]
Diuron	Adama Agan Ltd	Chitosan	<i>Z. mays</i> L.	<i>Echinochloa crus-galli</i> L. Beauv	[241]
2,4-D	FirmLimited Company	Nanosized rice husk		<i>T. repens</i> L., <i>Stellaria media</i> L	

Table 5 (continued)

Chemical	Trade manufacture company	Nanocomposites used	Crop	Target	Key references
<i>Fungicides</i>					
Tebuconazole	Super bio tech	PVP and PVP copolymer Bacterial ghosts	<i>Pinus taeda</i> L. <i>T. aestivum</i> L., <i>H. vulgare</i> L. and <i>Cucumis sativus</i> L.	<i>Gloeophyllum trabeum</i> <i>Erysiphe graminis</i> and <i>Sphaerotheca fuliginea</i>	[244]
Chlorothalonil	Super bio tech	Polymeric and SLN	<i>P. vulgaris</i> L.	<i>A. niger</i>	[11]
Kathon 930	Rallis india limited	PVP and PVP copolymer	<i>Pinus taeda</i> L.	<i>Gloeophyllum trabeum</i>	[244]
	Rohm and haas company	PVC	<i>Betula alleghaniensis</i> Britt	<i>Trametes versicolor</i>	[245]
Validamycin	Indochem agri science private limited	PHSN	<i>S. tuberosum</i> L.	<i>R. solani</i>	[246]
Pyraclostrobin	Shijiazhuangdai-lunchemical co. Ltd	Chitosan–PLA graft copolymer Chitosan/MSN	<i>Abelmoschus esculentus</i> L. <i>S. lycopersicum</i> L.	<i>Colletotrichum gossypii</i> <i>Puccinia asparagi</i>	[247] [248]
Ferbam	Loveland Products Canada Inc	Gold	<i>Camellia sinensis</i>	<i>Pythium aphanidermatum</i>	[249]
Carbendazim	Chemet Wets & Flows Pvt. Ltd	Chitosan/Pectin	<i>Z. mays</i> L.	<i>F. oxysporum</i>	[250]
Prochloraz	Lanfeng biochemical co., Ltd	PHSN	<i>C. sativus</i> L.	<i>B. cinerea</i>	[251]

implementation. Although, the studies on the soil–plant microbiome and nanoscale characterization highlight the impact of chemical agrochemical on the environment.

The use of nanocoated AIs biopesticides is expected to surpass the challenges of chemical residual management gap and premature degradation of AIs. Instead, these, applying new nanocomponents along with existing chemicals should follow regular checks on resistance strategies of targeted organisms, new resistance pathways, and revolutionized pest strains. Although, smart agrochemicals or nanoagrochemicals resolve so many issues and gives an instant solution.

To ensure these, it is essential to develop more international and national risk assessment, management, and mitigating strategies. Beyond these challenges, social acceptance with reduced environmental cost chiefly soil deterioration, microbiome disruption, depleted water resources need keen monitoring. Ecologically, the continuum uses of agrochemical put the question on survival challenges result in more resistance races creating a vicious loop in which pesticides concentration help to revolutionizing the organism more toward superiority.

For this, alternative strategies with strong monitoring are required, together recommendations of IPM practices help to eliminate shortcomings in individual practices. Despite the advancement in studies on nanoformulation and plant response more extensions in genomic, proteomics, physiological, and metabolic studies help to understand the interaction in the mechanism.

Abbreviations

NPs: Nanoparticles; NMs: Nanomaterials-based products; AIs: Active ingredients; CRS: Controlled release system; CR: Controlled release; PLA: Poly lactic acid; PLGA: Poly(lactic-co-glycolic acid); mPEG: Methoxy polyethylene glycol; PCL: Poly(ϵ -caprolactone); γ -PGA: (Poly (γ -glutamic acid)); γ -GTP: (γ -Glutamyl transpeptidase); UV: Ultraviolet; PEG: Polyethylene glycol; CAGR: Compound annual growth rate; IPM: Integrated pest management; Ag⁺: Silver; SiO₂-NPs: Silicon dioxide nanoparticles; Ch-polymethacrylic NPK: Chitosan polymethacrylic nitrogen phosphorus potassium; Au-NPs: Gold nanoparticles; ZnO NPs: Zinc oxide nanoparticles; CeO₂-NPs: Cerium dioxide nanoparticles; TiO₂-NPs: Titanium oxide nanoparticles; *S. oleracea*: *Spinacia oleracea*; Si NPs: Silicon nanoparticles; *V. mungo*: *Vigna mungo*; *V. radiate*: *Vigna radiate*; *C. arietinum*: *Cicer arietinum*; Ch-NPs: Chitosan nanoparticles; CS-EO: Chitosan essential oil; MDA: Malondialdehyde; H₂O₂: Hydrogen peroxide; PS II: Photosystem II; Fe₃O₄ NPs: Iron oxide nanoparticles; Fe NPs: Iron nanoparticles; *T. aestivum*: *Triticum aestivum*; *B. pilosa*: *Bidens pilosa*; *C. dactylon*: *Cynodon dactylon*; Ag-NPs: Silver nanoparticles; CM- β -CD-MNPs-Diuron complex: Carboxymethyl-hydroxypropyl- β -cyclodextrin magnetic nanoparticles diuron complex; Ag@dsDNA GO: Ag@dsDNA-graphene oxide; *L. esculentum*: *Lycopersicon esculentum*; *Z. mays*: *Zea mays*; CeO₂: Cerium dioxide; ROS: Reactive oxygen species; Mg: Magnesium; Al: Aluminium; Fe: Iron; Ti: Titanium; Ce: Cerium; Zn: Zinc; 2-4-D: 2-4 Dichlorophenoxy acetic acid; DCPT: DDT- Dichlorodiphenyltrichloroethane; nZVI: Zerovalent iron; Fe-Pd: Iron-palladium; Fe-S: Iron-Sulphur; PBHA: Polyhydroxybutyrate-co-hydroxyvalerate; *P. vulgaris*: *Phaseolus vulgaris*; *C. annuum*: *Capsicum annuum*; *S. oleracea*: *Spinacia oleracea*; *B. juncea*: *Brassica juncea*; CNTs: Carbon nanotubes; Cu₃(PO₄)₂: Copper(II) phosphate; *X. perforans*: *Xanthomonas perforans*; *B. sorokiniana*: *Bipolaris sorokiniana*; *X. alfalfa*: *Xanthomonas alfalfa*; *C. riparius*: *Chironomus riparius*; CrBR2.2: Ballbani ring protein gene; CrGrRH1: Gonadotrophin-releasing hormone gene; *D. melanogaster*: *Drosophila melanogaster*; *L. usitatissimum*: *Linum usitatissimum*; *G. max*: *Glycine max*; SLN: Solid lipid nanoparticles; *G. hirsutum*: *Gossypium hirsutum*; PVA: Poly vinyl alcohol; *S. lycopersicum*: *Solanum lycopersicum*; *S. bicolor*: *Sorghum bicolor*; PVC: Polyvinyl chloride; PHSN: Polystyrene nanoparticles; *O. sativa*: *Oryza sativa*; SnO₂: Stannic oxide; *H. vulgare*: *Hordeum vulgare*; *A. cepa*: *Allium cepa*; *T. repens*: *Trifolium repens*; *H. vulgare*: *Hordeum vulgare*; *S. tuberosum*: *Solanum tuberosum*; MSN: Mesoporous silica nanoparticles; *C. sativus*: *Cucumis sativus*; *B. cinerea*: *Botrytis cinerea*.

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Authors' contributions

The primary draft of the manuscript was prepared by AH and established by AC, AK, HK, and SM. SM reviewed the literature, AK and AC examined the manuscript and designed the table and figure section. AK, AC, and SM were involved in manuscript writing, and AH offered crucial advice and examined the entire manuscript on every step of writing. All authors read and approved the final version of the manuscript.

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References

- Zulfiqara F, Navarro M, Ashraf M, Akrame NA, Munné-Bosch B (2019) Nanofertilizer use for sustainable agriculture: advantages and limitations. *Plant Sci* 289:110270. <https://doi.org/10.1016/j.plantsci.2019.110270>
- Yu J, Wang D, Geetha N, Khawar KM, Jogaiah S, Mujtaba M (2021) Current trends and challenges in the synthesis and applications of chitosan-based nanocomposites for plants: a review. *Carbohydr Polym* 261:117904. <https://doi.org/10.1016/j.carbpol.2021.117904>
- Mittal D, Kaur G, Singh P, Yadav K, Ali SA (2020) Nanoparticle-based sustainable agriculture and food science: recent advances and future outlook. *Front Nanotechnol* 2:579954. <https://doi.org/10.3389/fnano.2020.579954>
- Husen H, Iqbal M (2019) *Nanomaterials and plant potential*. Springer, Cham. <https://doi.org/10.1007/978-3-030-05569-1>
- Husen H, Jawaid M (2020) *Nanomaterials for agriculture and forestry applications*. Elsevier, Cambridge. <https://doi.org/10.1016/C2018-0-02349-X>
- Husen H (2021) Harsh environment and plant resilience (*Molecular and Functional Aspects*). Springer, Cham. <https://doi.org/10.1007/978-3-030-65912-7>
- Husen H (2021) Plant performance under environmental stress (*Hormones, Biostimulants and Sustainable Plant Growth Management*). Springer, Cham. <https://doi.org/10.1007/978-3-030-78521-5>
- Bachheti RK, Fikadu A, Bachheti A, Husen A (2020) Biogenic fabrication of nanomaterials from flower-based chemical compounds, characterization and their various applications: a review. *Saudi J Biol Sci* 27:2551–2562. <https://doi.org/10.1016/j.sjbs.2020.05.012>
- Siddiqi MH, Al-Wahaibi MH (2014) Role of nano-SiO₂ in germination of tomato (*Lycopersicon esculentum* seeds Mill). *Saudi J Biol Sci* 21:13–17. <https://doi.org/10.1016/j.sjbs.2013.04.005>
- Sonika D, Saurav K, Aakash G, Uttam L, Ranjita T, Shankar J, Ganesh L, Deval PB, Niranjana P (2021) Current research on silver nanoparticles: synthesis, characterization, and applications. *J Nanomat* 2021:6687290. <https://doi.org/10.1155/2021/6687290>
- Salem SS, Fouda A (2021) Green synthesis of metallic nanoparticles and their prospective biotechnological applications: an overview. *Biol Trace Elem Res* 199:344–370. <https://doi.org/10.1007/s12011-020-02138-3>
- He X, Deng H, Hwang H (2019) The current application of nanotechnology in food and agriculture. *J Food Drug Anal* 27:1–21. <https://doi.org/10.1016/j.jfda.2018.12.002>
- Salem SS, Fouda MMG, Fouda A (2021) Antibacterial, cytotoxicity and larvicidal activity of green synthesized selenium nanoparticles using *Penicillium corylophilum*. *J Clust Sci* 32:351–361. <https://doi.org/10.3390/jof7050372>
- Husen A, Siddiqi KS (2014) Plants and microbes assisted selenium nanoparticles: characterization and application. *J Nanobiotechnol* 12:28. <https://doi.org/10.1186/s12951-014-0028-6>
- Khot LR, Sankaran S, Maja JM, Ehsani R, Schuster EW (2012) Applications of nanomaterials in agricultural production and crop protection: a review. *Crop Prot* 35:64–70. https://doi.org/10.1007/978-3-030-31938-0_7
- Nair R, Varghese SH, Nair BG, Maekawa T, Yoshida Y, Kumar DS (2010) Nanoparticulate material delivery to plants. *Plant Sci* 179:154–163. <https://doi.org/10.1016/j.plantsci.2010.04.012>
- Sharma A, Bachheti A, Sharma P, Bachheti RK, Husen A (2020) Phytochemistry, pharmacological activities, nanoparticle fabrication, commercial products and waste utilization of *Carica papaya* L.: a comprehensive review. *Curr Res Biotechnol* 2:145–160. <https://doi.org/10.1016/j.crbiot.2020.11.001>
- Pandey A, Srivastava S, Aggarwal N (2020) Assessment of the pesticidal behaviour of diacyl hydrazine-based ready-to-use nanofertilizers. *Chem Biol Technol Agric* 7:10. <https://doi.org/10.1186/s40538-020-0177-9>
- Fraceto LF, Pascoli M, de Albuquerque FP, Calzavara AK, Tinoco-Nunes B, Oliveira WHC, Gonçalves KC (2020) The potential of nanobiopesticide based on zein nanoparticles and neem oil for enhanced control of agricultural pests. *J Pest Sci* 93:793–806. https://doi.org/10.1007/978-3-030-61985-5_17
- Kamle M, Mahato DK, Devi S, Soni R, Tripathi V, Mishra AK, Kumar P (2020) Nanotechnological interventions for plant health improvement and sustainable agriculture. *Biotech* 10:1–1. <https://doi.org/10.1007/s13205-020-2152-3>
- Özkara A, Akyıl D, Konuk M (2016) Pesticides, environmental pollution, and health. In: *Environmental health risk-hazardous factors to living species 2016*, p 16. <https://doi.org/10.5772/63094>
- Titir G, Geetha G, Rita K, Amitava M (2020) Nanocomposites for delivering agrochemicals: a comprehensive review. *J Agric Food Chem* 68:3691–3702. <https://doi.org/10.1021/acs.jafc.9b06982>
- Aouada FA, de Moura MR, Orts WJ, Mattoso LHC (2010) Polyacrylamide and methylcellulose hydrogel as delivery vehicle for the controlled release of paraquat pesticide. *J Mater Sci* 45:4977–4985. <https://doi.org/10.1002/app.30339>
- Bortolin A, Aouada FA, de Moura MR, Ribeiro C, Longo E, Mattoso LHC (2012) Application of polysaccharide hydrogels in adsorption and controlled-extended release of fertilizers processes. *J Appl Polym Sci* 123:2291–2298. <https://doi.org/10.1002/app.34742>
- Ghazali SAISM, Hussein MZ, Sarijo SH (2013) 3,4-Dichlorophenoxyacetate intercalated into anionic clay for controlled release formulation of a new environmentally friendly agrochemical. *Nanoscale Res Lett* 8:362. <https://doi.org/10.1186/1556-276X-8-362>
- Chuxiang S, Ke S, Wei W, Zhao Y, Tian L, Yuxiang G, Hua Z, Yihua Y (2014) Encapsulation and controlled release of hydrophilic pesticide in shell cross-linked nanocapsules containing aqueous core. *Int J Pharm* 463:108–114. <https://doi.org/10.1016/j.ijpharm.2013.12.050>
- Wanyika H (2014) Controlled release of agrochemicals intercalated into montmorillonite interlayer space. *Sci World J* 2014(1–15):656287. <https://doi.org/10.1155/2014/656287>
- Cartmill AD, Cartmill DL, Alarcon A (2014) Controlled release fertilizer increased phytoremediation of petroleum-contaminated sandy soil. *Int J Phytorem* 16:285–301. <https://doi.org/10.1080/15226514.2013.773280>
- Carson LC, Ozores-Hampton M, Morgan KT, Sargent SA (2014) Effect of controlled-release and soluble fertilizer on tomato production and postharvest quality in seepage irrigation. *Hort Sci* 49:89–95. <https://doi.org/10.21273/HORTSCI.49.1.89>
- Sopena F, Maqueda C, Morillo E (2009) Controlled release formulations of herbicides based on micro-encapsulation. *Cienc Investig Agrar* 35:27–42. <https://doi.org/10.4067/S0718-16202009000100002>

31. Chevillard A, Angellier-Coussy H, Guillard V, Gontard N, Gastaldi E (2012) Controlling pesticide release via structuring agropolymer and nanoclays based materials. *J Hazard Mater* 205:32–39. <https://doi.org/10.1016/j.jhazmat.2011.11.093>
32. El-Temseh YS, Joner EJ (2013) Effects of nano-sized zero-valent iron (nZVI) on DDT degradation in soil and its toxicity to collembola and ostracods. *Chemosphere* 92:131–137
33. Aouada FA, de Moura MR (2015) Nanotechnology applied in agriculture: controlled release of agrochemicals. In: Rai M, et al (eds) *Nanotechnologies in food and agriculture*. Springer. <https://doi.org/10.1007/978-3-319-14024-7>
34. Fauzia S, Furqani F, Zein R, Munaf E (2015) Adsorption and reaction kinetics of tatzazine by using *Annona muricata* L. seeds. *J Chem Pharm Res* 7:573–582
35. Stloukal P, Kucharczyk P, Sedlarik V, Bazant P, Koutny M (2012) Low molecular weight poly(l-lactic acid) microparticles for controlled release of the herbicide metazachlor: preparation, morphology, and release kinetics. *J Agric Food Chem* 60:4111–4119. <https://doi.org/10.1021/jf300521j>
36. Zielińska A, Carreiró F, Oliveira AM, Neves A, Pires B, Venkatesh DN, Durazzo A, Lucarini M, Eder P, Silva AM, Santini A, Souto EB (2020) Polymeric nanoparticles: production, characterization, toxicology and ecotoxicology. *Molecules* 25:3731–3951. <https://doi.org/10.3390/molecules25163731>
37. Fu JX, Wang HJ, Zhou YQ, Wang JY (2009) Antibacterial activity of ciprofloxacin-loaded zein microsphere films. *Mater Sci Eng* 29:1161–1166. <https://doi.org/10.1016/j.msec.2008.09.031>
38. Hou Y, Hu J, Park H, Lee M (2012) Chitosan based nanoparticles as a sustained protein release carrier for tissue engineering applications. *J Biomed Mater Res Part A* 100:939–947. <https://doi.org/10.1002/jbm.a.34031>
39. Kalagatur NK, Nirmal Ghosh OS, Sundararaj N, Mudili V (2018) Antifungal activity of chitosan nanoparticles encapsulated with *Cymbopogon martinii* essential oil on plant pathogenic fungi *Fusarium graminearum*. *Front Pharmacol* 9:610. <https://doi.org/10.3389/fphar.2018.00610>
40. El-Hamshary H, Fouda MMG, Moydeen M, El-Newehy MH, Al-Deyab SS, Megheed M (2016) Synthesis and antibacterial of carboxymethyl starch-grafted poly (vinyl imidazole) against some plant pathogens. *Int J Biol Macromol* 72:1466–1472. <https://doi.org/10.1016/j.jbiomac.2014.10.051>
41. Ianchis R, Ninciuleanu CM, Gifu IC, Alexandrescu E, Somoghi R, Gabor AR, Preda S, Nistor CL, Nitu S, Petcu C, Icriverzi M, Florian PE, Roseanu AM (2017) Novel hydrogel-advanced modified clay nanocomposites as possible vehicles for drug delivery and controlled release. *NANO* 7:443. <https://doi.org/10.3390/nano7120443>
42. Zheng M, Falkeborg M, Zheng Y, Yang T, Xu X (2013) Formulation and characterization of nanostructured lipid carriers containing a mixed lipids core. *Colloids Surf Physicochem Eng Asp* 430:76–84. <https://doi.org/10.1016/j.jconrel.2013.01.018>
43. Pan Y, Tikekar RV, Nitin N (2016) Distribution of a model bioactive within solid lipid nanoparticles and nanostructured lipid carriers influences its loading efficiency and oxidative stability. *Int J Pharm* 511:322–330. <https://doi.org/10.1016/j.jpharm.2016.07.019>
44. Chawla JS, Amiji MM (2002) Biodegradable poly (ϵ - caprolactone) nanoparticles for tumor-targeted delivery of tamoxifen. *Int J Pharm* 249:127–138. [https://doi.org/10.1016/s0378-5173\(02\)00483-0](https://doi.org/10.1016/s0378-5173(02)00483-0)
45. Orellana-Tavra C, Baxter EF, Tian T, Bennett TD, Slater NKH, Cheatham AK, Fairen-Jimenez D (2015) Amorphous metal-organic frameworks for drug delivery. *Chem Commun* 51:13878–13881. <https://doi.org/10.1039/c5cc05237h>
46. Lin G, Chen X, Zhou H, Zhou X, Xu H, Chen H (2019) Elaboration of a feather keratin/carboxymethyl cellulose complex exhibiting pH sensitivity for sustained pesticide release. *J Appl Polym Sci* 136:47160. <https://doi.org/10.1002/app.47160>
47. Ramasamy T, Ruttala HB, Gupta B, Poudel BK, Choi HG, Yong CS, Kim JO (2017) Smart chemistry-based nanosized drug delivery systems for systemic applications: a comprehensive review. *J Control Release* 258:226–253. <https://doi.org/10.1016/j.jconrel.2017.04.043>
48. Henchion M, McCarthy M, Dillon EJ (2019) Big issues for a small technology: consumer trade-offs in acceptance of nanotechnology in food. *Innov Food Sci Emerg Technol* 58:102210. <https://doi.org/10.1016/j.ifset.2019.102210>
49. Lai RWS, Yeung KWY, Yung MMN (2018) Regulation of engineered nanomaterials: current challenges, insights and future directions. *Environ Sci Pollut Res Int* 25:3060–3077. <https://doi.org/10.1007/s11356-017-9489-0>
50. Kamaraj C, Gandhi PR, Elango G, Karthi S, Chung IM, Rajakumar G (2018) Novel and environmental friendly approach; Impact of Neem (*Azadirachta indica*) gum nanoformulation (NGNF) on *Helicoverpa armigera* (Hub.) and *Spodoptera litura* (Fab.). *Int J Biol Macromol* 107:59–69. <https://doi.org/10.1016/j.jbiomac.2017.08.145>
51. Vishnu D, Tatiana M, Arvind B, Svetlana NS, Saglara M, Ritu S, Andrey G, Viktoria ST, William OP, Karen AG, Hasmik SM (2018) Effects of zinc-oxide nanoparticles on soil, plants, animals and soil organisms: a review. *Environ Nanotech Monitor Manag* 9:76–84. <https://doi.org/10.1016/j.enmm.2017.12.006>
52. Zoya J, Kavaya D, Mansi M, Vinayak DF, Ayushi S (2019) Effect of accumulation of nanoparticles in soil health- a concern on future. *Front Nanosci Nanotech*. <https://doi.org/10.15761/FNN.1000182>
53. Zand AD, Tabrizi AM, Heir AV (2020) Incorporation of biochar and nanomaterials to assist remediation of heavy metals in soil using plant species. *Environ Technol Innov* 20:101134. <https://doi.org/10.1016/j.eti.2020.101134>
54. Pandey S, Giri K, Kumar R (2018) Nanopesticides: opportunities in crop protection and associated environmental risks. *Proc Natl Acad Sci India Sect B Biol Sci* 88:1287–1308. <https://doi.org/10.1007/s40011-016-0791-2>
55. Pavela R, Benelli G (2016) Essential oils as eco-friendly biopesticides? Challenges and constraints. *Trends Plant Sci* 21:1000–1007. <https://doi.org/10.1016/j.tplants.2016.10.005>
56. Kremer RJ (2020) Bioherbicides and nanotechnology: current status and future trends. In: *Nano-biopesticides today and future perspectives*, Academic Press, pp 353–366. <https://doi.org/10.1016/B978-0-12-815829-6.00015-2>
57. Grillo R, Fraceto LF, Amorim MJ, Scott-Fordsmand JJ, Schoonjans R, Chaudhry Q (2020) Ecotoxicological and regulatory aspects of environmental sustainability of nanopesticides. *J Hazard Mater* 404:124148. <https://doi.org/10.1016/j.jhazmat.2020.124148>
58. Torre-Roche RDL, Hawthorne J, Deng Y, Xing B, Cai W, Newman LA, Wang Q, Ma X, Hamdi H, White JC (2013) Multiwalled carbon nanotubes and C₆₀ fullerenes differentially impact the accumulation of weathered pesticides in four agricultural plants. *Environ Sci Technol* 12:12539–12547. <https://doi.org/10.1021/es4034809>
59. Kah M, Tufenkji N, White JC (2019) Nano-enabled strategies to enhance crop nutrition and protection. *Nat Nanotechnol* 14:532–540. <https://doi.org/10.1038/s41565-019-0439-5>
60. de Oliveira RL, de Mello PR, Felisberto G, Checchio MV, Gratão PL (2019) Silicon mitigates manganese deficiency stress by regulating the physiology and activity of antioxidant enzymes in sorghum plants. *J Soil Sci Plant Nutr* 19:524–534. <https://doi.org/10.1007/s11738-013-1367-x>
61. Ahmed B, Ameen F, Rizvi A, Ali K, Sonbol H, Zaidi A, Musarrat J (2020) Destruction of cell topography, morphology, membrane, inhibition of respiration, biofilm formation, and bioactive molecule production by nanoparticles of Ag, ZnO, CuO, TiO₂, and Al₂O₃ toward beneficial soil bacteria. *ACS Omega* 5:7861–7876. <https://doi.org/10.1021/acsomega.9b04084>
62. Chung I, Rekha K, Venkidasamy B, Thiruvengadam M (2019) Effect of copper oxide nanoparticles on the physiology, bioactive molecules, and transcriptional changes in *Brassica rapa* ssp. *rapa* seedlings. *Water Air Soil Pollut* 230:48. <https://doi.org/10.1007/s11270-019-4084-2>
63. Kremer RJ (2020) Bioherbicides and nanotechnology: current status and future trends. In: *Nano-biopesticides today and future perspectives*. Academic Press, pp 353–366. <https://doi.org/10.1016/B978-0-12-815829-6.00015-2>
64. Fadoju OM, Osinowo OA, Ogunsuyi OI (2020) Interaction of titanium dioxide and zinc oxide nanoparticles induced cytogenotoxicity in *Allium cepa*. *Nucleus* 63:159–166. <https://doi.org/10.1007/s13237-020-00308-1>
65. Yousefu MS, Elamawi RM (2018) Evaluation of phytotoxicity, cytotoxicity, and genotoxicity of ZnO nanoparticles in *V. faba*. *Environ Sci Pollut Res Int* 5:1–13. <https://doi.org/10.1007/s11356-018-3250-1>

66. Guilger M, Pasquoto-Stigliani T, Bilesky-Jose N, Grillo R, Abhilash PC, Fraceto LF, Lima R (2017) Biogenic silver nanoparticles based on *trichoderma harzianum*: synthesis, characterization, toxicity evaluation and biological activity. *Sci Rep* 7:44421. <https://doi.org/10.1038/srep44421>
67. Kumar S, Bhanjana G, Sharma A, Dilbaghi N, Sidhu MC, Kim KH (2017) Development of nanoformulation approaches for the control of weeds. *Sci Total Environ* 586:1272–1278. <https://doi.org/10.1016/j.scitotenv.2017.02.138>
68. Chevillard H, Angellier-Coussy V, Guillard N, Gontard EG (2012) Controlling pesticide release via structuring agropolymer and nanoclays based materials. *J Hazard Mater* 205:32–39. <https://doi.org/10.1016/j.jhazmat.2011.11.093>
69. Wilpiszewska K, Spychaj T, Paździoch W (2016) Carboxymethyl starch/montmorillonite composite microparticles: properties and controlled release of isoprotruron. *Carbohydr Polym* 136:101–106. <https://doi.org/10.1016/j.carbpol.2015.09.021>
70. Francis S, Joseph S, Koshy EP, Mathew B (2017) Green synthesis and characterization of gold and silver nanoparticles using *Mussaenda glabrata* leaf extract and their environmental applications to dye degradation. *Environ Sci Pollut Res Int* 24:17347–17357. <https://doi.org/10.1038/srep44421>
71. Nafees M, Ali S, Rizwan M, Aziz A, Adrees M, Hussain S, Junaid M (2020) Effect of nanoparticles on plant growth and physiology and on soil microbes. In: *Nanomaterials and environmental biotechnology*. Springer, Cham, pp. 65–85. https://doi.org/10.1007/978-3-030-34544-0_5
72. Hasaneen M, Abdel-aziz HMM, Omer AM (2016) Effect of foliar application of engineered nanomaterials: carbon nanotubes NPK and chitosan nanoparticles NPK fertilizer on the growth of French bean plant. *Biochem Biotechnol Res* 4:68–76
73. Mujtaba M, Khawar KM, Camara MC, Carvalho LB, Fraceto LF, Morsi RE, Wang D (2020) Chitosan-based delivery systems for plants: a brief overview of recent advances and future directions. *Int J Biol Macromol* 154:683–697. <https://doi.org/10.1016/j.ijbiomac.2020.03.128>
74. Mahakham W, Theerakulpisut P, Maensiri S, Phumying S, Samah AK (2016) Environmentally benign synthesis of phytochemicals-capped gold nanoparticles as nanopriming agent for promoting maize seed germination. *Sci Total Environ* 573:1089–1102. <https://doi.org/10.1016/j.scitotenv.2016.08.120>
75. Ullah H, Li X, Peng L, Cai Y, Mielke HW (2020) In vivo phytotoxicity, uptake, and translocation of PbS nanoparticles in maize (*Z. mays* L.) plants. *Sci Total Environ* 737:139558. <https://doi.org/10.1016/j.scitotenv.2020.139558>
76. Sharif-Rad J, Sharif-Rad M, Teixeira da Silva JA (2016) Morphological, physiological and biochemical responses of crops (*Z. mays* L., *Phaseolus vulgaris* L.), medicinal plants (*Hyssopus officinalis* L., *Nigella sativa* L.), and weeds (*Amaranthus retroflexus* L., *Taraxacum officinale* F. H. Wigg) exposed to SiO₂ nanoparticles. *J Agric Sci Technol* 18:1027–1040
77. Rezaei S, Ariaei P, Charmchian LM (2020) The effect of encapsulated plant extract of hyssop (*Hyssopus officinalis* L.) in biopolymer nanoemulsions of *Lepidium perfoliatum* and *Orchis mascula* on controlling oxidative stability of soybean oil. *Food Sci Nutr* 8:1264–1271. <https://doi.org/10.1002/fsn3.1415>
78. Sabir S, Zahoor MA, Waseem M, Siddique MH, Shafique M, Imran M, Muzammil S (2020) Biosynthesis of ZnO nanoparticles using *Bacillus subtilis*: characterization and nutritive significance for promoting plant growth in *Z. mays* L. Dose-Response 18:1559325820958911. <https://doi.org/10.1007/s00253-012-3934-2>
79. Saharan V, Mehrotra A, Khatik R, Rawal P, Sharma SS, Pal A (2013) Synthesis of chitosan based nanoparticles and their in vitro evaluation against phytopathogenic fungi. *Int J Biol Macromol* 62:677–683. <https://doi.org/10.1016/j.ijbiomac.2013.10.012>
80. Choudhary RC, Kumaraswamy R, Kumari S, Sharma S, Pal A, Raliya R, Biswas P, Saharan V (2017) Cu-chitosan nanoparticle boost defense responses and plant growth in maize (*Z. mays* L.). *Sci Rep* 7:9754. <https://doi.org/10.1038/s41598-017-08571-0>
81. Deshpande P, Dapkekar A, Oak MD, Paknikar KM, Rajwade JM (2017) Zinc complexed chitosan/TPP nanoparticles: a promising micronutrient nanocarrier suited for foliar application. *Carbohydr Polym* 165:394–401. <https://doi.org/10.1016/j.carbpol.2017.02.061>
82. Maluin FN, Hussein MZ (2020) Chitosan-based agronanochemicals as a sustainable alternative in crop protection. *Molecules* 25:1611. <https://doi.org/10.3390/molecules25071611>
83. Pereira A, Sandoval-Herrera I, Zavala-Betancourt S, Oliveira H, Ledezma-Pérez A, Romero J, Fraceto L (2017) γ -Polyglutamic acid/chitosan nanoparticles for the plant growth regulator gibberellic acid: characterization and evaluation of biological activity. *Carbohydr Polym* 157:1862–1873. <https://doi.org/10.1016/j.carbpol.2016.11.073>
84. Khalifa NS, Hasaneen MN (2018) The effect of chitosan-PMAA-NPK nanofertilizer on *Pisum sativum* plants. *Biotech* 8:193. <https://doi.org/10.1007/s13205-018-1221-3>
85. Mansoor N, Younus A, Jamil Y, Shahid M (2019) Impact of nanosized and bulk ZnO on germination and early growth response of *T. aestivum*. *Pak J Agric Sci* 56:879–884
86. Adrees M, Khan ZS, Hafeez M, Rizwan M, Hussain K, Asrar M, Ali S (2020) Foliar exposure of zinc oxide nanoparticles improved the growth of wheat (*T. aestivum* L.) and decreased cadmium concentration in grains under simultaneous Cd and water deficient stress. *Ecotoxic Environ Saf* 208:111627. <https://doi.org/10.1016/j.ecoenv.2020.111627>
87. Wu F, Fang Q, Yan S, Pan L, Tang X, Ye W (2020) Effects of zinc oxide nanoparticles on arsenic stress in rice (*Oryza sativa* L.): germination, early growth, and arsenic uptake. *Environ Sci Pollut Res Int* 27:26974–26981. <https://doi.org/10.1007/s11356-020-08965-0>
88. Singh D, Kumar A (2020) Quantification of metal uptake in *Spinacia oleracea* irrigated with water containing a mixture of CuO and ZnO nanoparticles. *Chemosphere* 243:125239. <https://doi.org/10.1016/j.chemosphere.2019.125239>
89. Spielman-sun E, Avellan A, Bland GD, Tappero RV, Acerbo AS, Unrine JM, Giraldo JP, Lowry GV (2019) Nanoparticle surface charge influences translocation and leaf distribution in vascular plants with contrasting anatomy. *Environ Sci Nano* 6:2508–2519. <https://doi.org/10.1039/C9EN00626E>
90. Fox JP, Capen JD, Zhang W, Ma X, Rossi L (2020) Effects of cerium oxide nanoparticles and cadmium on corn (*Z. mays* L.) seedlings physiology and root anatomy. *NanoImpact* 20:100264. <https://doi.org/10.1016/j.impact.2020.100264>
91. Prakash S, Deswal R (2020) Analysis of temporally evolved nanoparticle-protein corona highlighted the potential ability of gold nanoparticles to stably interact with proteins and influence the major biochemical pathways in Brassica juncea. *Plant Physiol Biochem* 146:143–156. <https://doi.org/10.1016/j.plaphy.2019.10.036>
92. Ghorbani R, Movafeghi A, Gangeali A, Nabati J (2021) Effects of TiO₂ nanoparticles on morphological characteristics of chickpea (*Cicer arietinum* L.) under drought stress. *Environ Stresses Crop Sci* 14:85–98. <https://doi.org/10.22077/ESCS.2020.2485.1654>
93. Buteler M, Gitto JG, Stadler T (2020) Enhancing the potential use of microparticulate insecticides through removal of particles from raw grain. *J Stored Prod Res* 89:101707. <https://doi.org/10.1016/j.jspr.2020.101707>
94. Kandil MAH, Sammour EA, Abdel-Aziz NF (2020) Comparative toxicity of new insecticides generations against tomato leafminer *Tuta absoluta* and their biochemical effects on tomato plants. *Bull Natl Res Cent* 44:126. <https://doi.org/10.1186/s42269-020-00382-0>
95. Li Z, Sellaoui L, Franco D, Netto MS, Georjina J, Dotto GL, Li Q (2020) Adsorption of hazardous dyes on functionalized multiwalled carbon nanotubes in single and binary systems: experimental study and physicochemical interpretation of the adsorption mechanism. *Chem Eng J* 389:124467. <https://doi.org/10.1016/j.molliq.2020.114348>
96. Zhang DX, Li BX, Zhang XP, Zhang ZQ, Wang WC, Liu F (2016) Phoxim microcapsules prepared with polyurea and urea-formaldehyde resins differ in photostability and insecticidal activity. *J Agric Food Chem* 64:2841–2846. <https://doi.org/10.1021/acs.jafc.6b00231>
97. Wibowo D, Zhao CX, Peters BC, Middelberg AP (2014) Sustained release of fipronil insecticide in vitro and in vivo from biocompatible silica nanocapsules. *J Agric Food Chem* 12:504–511. <https://doi.org/10.1021/jf504455x>
98. Jacques MT, Oliveira JL, Campos EVR, Fraceto LF, Ávila DS (2017) Safety assessment of nanopesticides using the roundworm *Caenorhabditis elegans*. *Ecotoxicol Environ Saf* 139:245–253. <https://doi.org/10.1016/j.ecoenv.2017.01.045>

99. Pasquoto-Stigliani T, Campos EVR, Oliveira JL, Silva CMG, Bilesky-José N, Guilger M, Troost J, Oliveira HC, Stolf-Moreira R, Fraceto LF, de Lima R (2017) Nanocapsules containing neem (*Azadirachta indica*) oil: development, characterization, and toxicity evaluation. *Sci Rep* 7:5929. <https://doi.org/10.1038/s41598-017-06092-4>
100. Mittal AK, Chisti Y, Banerjee UC (2013) Synthesis of metallic nanoparticles using plant extracts. *Biotechnol Adv* 31:346–356. <https://doi.org/10.1016/j.biotechadv.2013.01.003>
101. Sebastian A, Nangia A, Prasad MNV (2019) Cadmium and sodium adsorption properties of magnetite nanoparticles synthesized from *Hevea brasiliensis* Muell. Arg. Bark: relevance in amelioration of metal stress in rice. *J Hazard Mater* 371:261–272. <https://doi.org/10.1016/j.jhazmat.2019.03.021>
102. Sathiyabama M, Parthasarathy R (2016) Biological preparation of chitosan nanoparticles and its in vitro antifungal efficacy against some phytopathogenic fungi. *Carbohydr Polym* 151:321–325. <https://doi.org/10.1016/j.carbpol.2016.05.033>
103. Vokhidova NR, Sattarov ME, Kareva ND, Rashidova SS (2014) Fungicide features of the nanosystems of silk worm (*Bombyx mori*) chitosan with copper ions. *Microbiol* 83:751–753
104. Rubina MS, Vasil'kov AY, Naumkin AV, Shtykova EV, Abramchuk SS, Alghuthaymi MA, Abd-El Salam KA (2017) Synthesis and characterization of chitosan-copper nanocomposites and their fungicidal activity against two sclerotia-forming plant pathogenic fungi. *J Nanostruct Chem* 7:249–258. <https://doi.org/10.1007/s40097-017-0235-4>
105. Xing K, Liu Y, Shen X, Zhu X, Li X, Miao X, Qin S (2017) Effect of O-chitosan nanoparticles on the development and membrane permeability of *Verticillium dahliae*. *Carbohydr Polym* 165:334–343. <https://doi.org/10.1016/j.carbpol.2017.02.063>
106. Kumaraswamy RV, Kumari S, Choudhary RC, Sharma SS, Pal A, Raliya R, Saharan V (2019) Salicylic acid functionalized chitosan nanoparticle: a sustainable biostimulant for plant. *Int J Biol Macromol* 123:59–69. <https://doi.org/10.1016/j.jbiomac.2018.10.202>
107. Kaur P, Thakur R, Choudhary A (2012) An in vitro study of the antifungal activity of silver/chitosan nanoformulations against important seed borne pathogens. *Int J Sci Technol Res* 1:83–86
108. Cao L, Zhang H, Cao C, Zhang J, Li F, Huang Q (2016) Quaternized chitosan-capped mesoporous silica nanoparticles as nanocarriers for controlled pesticide release. *Nanomater* 6:126. <https://doi.org/10.3390/nano6070126>
109. Luque-Alcaraz AG, Cortez-Rocha MO, Velázquez-Contreras CA, Acosta-Silva AL, Santacruz-Ortega HDC, Burgos-Hernández A, Argüelles-Monal WM, Plascencia-Jatomea M (2016) Enhanced antifungal effect of chitosan/pepper tree (*Schinus molle*) essential oil bionanocomposites on the viability of *Aspergillus parasiticus* Spores. *J Nanomat* 1:1–10. <https://doi.org/10.1007/s11356-020-10716-0>
110. Cindi MD, Shittu T, Sivakumar D, Bautista-Banos S (2015) Chitosan boehmite-alumina nanocomposite films and thyme oil vapour control brown rot in peaches (*Prunus persica* L.) during postharvest storage. *Crop Prot* 72:127–131. <https://doi.org/10.1016/j.cropro.2015.03.011>
111. Ngoc UTP, Nguyen DH (2018) Synergistic antifungal effect of fungicide and chitosan-silver nanoparticles on *Neoscytalidium dimidiatum*. *Green Proc Synth* 7:132–138. <https://doi.org/10.1515/gps-2016-0206>
112. Hossaina F, Follettb P, Salmieria S, Vua KD, Frascinic C, Lacroixa M (2019) Antifungal activities of combined treatments of irradiation and essential oils (EOs) encapsulated chitosan nanocomposite films in *in vitro* and *in situ* conditions. *Int J Food Microbiol* 295:33–40. <https://doi.org/10.3389/fmicb.2021.613155>
113. Medina E, Caro N, Abugoch L, Gamboa A, Diaz-Dosque M, Tapia C (2019) Chitosan thymol nanoparticles improve the antimicrobial effect and the water vapour barrier of chitosan-quinoa protein films. *J Food Eng* 240:191–198. <https://doi.org/10.1016/j.jfoodeng.2018.07.023>
114. Arumugam G, Velayutham V, Shanmugavel S, Sundaram J (2016) Efficacy of nanostructured silica as a stored pulse protector against the infestation of bruchid beetle, *Callosobruchus maculatus* (Coleoptera: Bruchidae). *Appl Nanosci* 6:445–450. <https://doi.org/10.1007/s13204-015-0446-2>
115. Ahmad J, Qamar S, Kausar N, Qureshi MI (2020). Nanoparticles: the magic bullets in mitigating drought stress in plants. In: *Nanobiotechnology in agriculture*. Springer, Cham, pp 145–161. https://doi.org/10.1007/978-3-030-39978-8_8
116. Singh S, Husen A (2020) Behavior of agricultural crops in relation to nanomaterials under adverse environmental conditions. In: Husen A, Jawaid M (eds) *Nanomaterials for agriculture and forestry applications*. Elsevier, Cambridge, pp 219–256. <https://doi.org/10.1016/B978-0-12-817852-2.00009-3>
117. Singh S, Husen A (2019) Role of nanomaterials in the mitigation of abiotic stress in plants. In: Husen A, Iqbal M (eds) *Nanomaterials and plant potential*. Springer, Cham, pp 441–471
118. Husen A (2021) The Harsh environment and resilient plants: an overview. In: Husen A (Eds.) *Harsh environment and plant resilience*, pp 1–23. https://doi.org/10.1007/978-3-030-65912-7_1
119. Yusefi-Tanha E, Fallah S, Rostamnejadi A, Pokhrel LR (2020) Zinc oxide nanoparticles (ZnONPs) as a novel nanofertilizer: Influence on seed yield and antioxidant defense system in soil grown soybean (*Glycine max* cv. Kowsar). *Sci Total Environ* 738:140240. <https://doi.org/10.1016/j.scitotenv.2020.140240>
120. Van HC, Van ND, Nguyen HM, Le NT, Nguyen KH, Le HM (2020) Copper nanoparticle application enhances plant growth and grain yield in maize under drought stress conditions. *bioRxiv*. <https://doi.org/10.1101/2020.02.24.963132>
121. An J, Hu P, Li F, Wu H, Shen Y, White JC, Giraldo JP (2020) Emerging investigator series: molecular mechanisms of plant salinity stress tolerance improvement by seed priming with cerium oxide nanoparticles. *Environ Sci Nano* 7:2214–2228. <https://doi.org/10.1039/DOEN00387E>
122. Attia MS, Osman MS, Mohamed AS, Mahgoub HA, Garada MO, Abdelmouty ES (2021) Impact of foliar application of chitosan dissolved in different organic acids on isozymes, protein patterns and physio-biochemical characteristics of tomato grown under salinity stress. *Plants* 10:388. <https://doi.org/10.3390/plants10020388>
123. Sikder RK, Wang X, Zhang H, Gui H, Dong Q, Jin D, Song M (2020) Nitrogen enhances salt tolerance by modulating the antioxidant defense system and osmoregulation substance content in *Gossypium hirsutum*. *Plants* 9:450. <https://doi.org/10.3390/plants9040450>
124. Ioannou A, Gohari G, Papaphilippou P, Panahirad S, Akbari A, Dadpour MR, Fotopoulos V (2020) Advanced nanomaterials in agriculture under a changing climate: the way to the future? *Environ Exp Bot* 176:104048. <https://doi.org/10.1016/j.envexpbot.2020.104048>
125. Khan N, Bano AMD, Babar A (2020) Impacts of plant growth promoters and plant growth regulators on rainfed agriculture. *PLoS ONE* 15:e0231426. <https://doi.org/10.1371/journal.pone.0231426>
126. Soliman M, Qari SH, Abu-Elsaoud A, El-Esawi M, Alhathloul H, Elkesh A (2020) Rapid green synthesis of silver nanoparticles from blue gum augment growth and performance of maize, fenugreek, and onion by modulating plants cellular antioxidant machinery and genes expression. *Acta Physiol Plant* 42:1–16. <https://doi.org/10.3390/plant510040790>
127. Iftikhar A, Rizwan M, Adrees M, Ali S, Rehman MZ, Qayyum MF, Hussain A (2020) Effect of gibberellic acid on growth, biomass, and antioxidant defense system of wheat (*T. aestivum* L.) under cerium oxide nanoparticle stress. *Environ Sci Pollution Res* 27:33809–33820. <https://doi.org/10.1007/s11356-020-09661-9>
128. Shoemaker AG (2020) The effects of titanium dioxide nanoparticles on the growth and development of *Sorghum Bicolor* (L.) Moench. *Adv Agric Horticult Entomol* 132:1–15
129. Maruyama CR, Guilger M, Pascoli M, Bilesky-José M (2016) Nanoparticles based on chitosan as carriers for the combined herbicides imazapic and imazapyr. *Sci Rep* 6:19768. <https://doi.org/10.1038/srep19768>
130. Preisler AC, Pereira AES, Campos EVR, Dalazen G, Fraceto LF, Oliveira HC (2020) Atrazine nanoencapsulation improves pre-emergence herbicidal activity against *Bidens pilosa* without enhancing long-term residual effect on *Glycine max*. *Pest Manag Sci* 76:141–149. <https://doi.org/10.1002/ps.5482>
131. Sousa GFM, Gomes DG, Campos EVR, Oliveira JL, Fraceto LF, Stolf-Moreira R, Oliveira HC (2018) Post-emergence herbicidal activity of nanoatrazine against susceptible weeds. *Front Environ Sci* 6:1–6. <https://doi.org/10.1007/s10311-019-00912-x>
132. Xiang Y, Zhang G, Chi Y, Cai D, Wu Z (2017) Fabrication of a controllable nanopesticide system with magnetic collectability. *Chem Eng J* 328:320–330. <https://doi.org/10.1021/acscuschemeng.7b00348>

133. Schlich K, Hund-Rinke K (2015) Influence of soil properties on the effect of silver nanomaterials on microbial activity in five soils. *Environ Pollut* 196:321–330. <https://doi.org/10.1016/j.envpol.2014.10.021>
134. Asadishad B, Chahal S, Akbari A, Cianciarelli V, Azodi M, Ghoshal S, Tufenkji N (2018) Amendment of agricultural soil with metal nanoparticles: effects on soil enzyme activity and microbial community composition. *Environ Sci Technol* 52:1908–1918. <https://doi.org/10.1021/acs.est.7b05389>
135. Zhai Y, Hunting ER, Wouters M, Peijnenburg WJGM, Vijver MG (2016) Silver nanoparticles, ions, and shape governing soil microbial functional diversity: nano shapes micro. *Front Microbiol* 7:1123. <https://doi.org/10.3389/fmicb.2016.01123>
136. Kędziora A, Speruda M, Krzyżewska E, Rybka J, Łukowiak A, Bugla-Płoskońska G (2018) Similarities and differences between silver ions and silver in nanoforms as antibacterial agents. *Int J Mol Sci* 19:444. <https://doi.org/10.3390/ijms19020444>
137. VandeVoort AR, Skipper H, Arai Y (2014) Macroscopic assessment of nanosilver toxicity to soil denitrification kinetics. *J Environ Qual* 43:1424–1430. <https://doi.org/10.2134/jeq2013.12.0524>
138. El-Temsah YS, Joner EJ (2012) Impact of Fe and Ag nanoparticles on seed germination and differences in bioavailability during exposure in aqueous suspension and soil. *Environ Toxicol* 27:42–49. <https://doi.org/10.1002/tox.20610>
139. Li F, Chen Y, Tang DM, Jian Z, Liu C, Golberg D, Yamada A, Zhou H (2014) Performance-improved Li–O₂ battery with Ru nanoparticles supported on binder-free multi-walled carbon nanotube paper as cathode. *Energy Environ Sci* 7:1648–1652. <https://doi.org/10.1039/C3EE44043E>
140. Dogaroglu ZG, Koleli N (2014) Effect of different zinc oxide nanoparticles on germination, plant growth and chlorophyll content of wheat. In: International congress on green infrastructure and sustainable societies/cities greinsus, p 78. <https://doi.org/10.13254/jjare.2020.0099>
141. Lei Y, Peiye L, Xiaopeng Z, Rong J, Lijuan Z (2020) Physiological and metabolic responses of maize (*Z. mays*) plants to Fe₃O₄ nanoparticles. *Sci Total Environ* 718:1–36. <https://doi.org/10.1016/j.scitotenv.2020.137400>
142. Falco WF, Scherer MD, Oliveira SL, Wender H, Colbeck I, Lawson T, Caires ARL (2019) Phytotoxicity of silver nanoparticles on *V. faba*: evaluation of particle size effects on photosynthetic performance and leaf gas exchange. *Sci Total Environ* 701:134816. <https://doi.org/10.1016/j.scitotenv.2019.134816>
143. Oliveira E, Núñez C, Santos HM, Fernández-Lodeiro J, Fernández-Lodeiro A, Capelo JL, Lodeiro C (2015) Revisiting the use of gold and silver functionalised nanoparticles as colorimetric and fluorometric chemosensors for metal ions. *Sens Actuators B Chem* 212:297–328. <https://doi.org/10.1016/j.snb.2015.02.026>
144. Scherer MD, Sposito JC, Falco WF, Grisolia AB, Andrade LH, Lima SM, Machado G, Nascimento VA, Gonçalves DA, Wender H, Oliveira SL (2019) Cytotoxic and genotoxic effects of silver nanoparticles on meristematic cells of Allium cepa roots: a close analysis of particle size dependence. *Sci Total Environ* 660:459–467. <https://doi.org/10.1016/j.scitotenv.2018.12.444>
145. Galazzi RM, Arruda MAZ (2018) Evaluation of changes in the macro and micronutrients homeostasis of transgenic and non-transgenic soybean plants after cultivation with silver nanoparticles through ionic approaches. *J Trace Elem Med Biol* 48:181–187. <https://doi.org/10.1016/j.jtemb.2018.04.004>
146. Pereira AE, Grillo R, Mello NF, Rosa AH, Fraceto LF (2014) Application of poly(epsilon-caprolactone) nanoparticles containing atrazine herbicide as an alternative technique to control weeds and reduce damage to the environment. *J Hazard Mater* 268:207–215. <https://doi.org/10.1016/j.jhazmat.2014.01.025>
147. Latef AAHA, Zaid A, Alhmad MFA, Abdelfattah KE (2020) The impact of priming with Al₂O₃ nanoparticles on growth, pigments, osmolytes, and antioxidant enzymes of Egyptian Roselle (*Hibiscus sabdariffa* L.) cultivar. *Agronomy* 10:681. <https://doi.org/10.3390/agronomy10050681>
148. Burklew CE, Ashlock J, Winfrey WB, Zhang B (2012) Effects of aluminum oxide nanoparticles on the growth, development, and microRNA expression of tobacco (*Nicotiana tabacum*). *PLoS ONE* 7:e34783. <https://doi.org/10.1371/journal.pone.0034783>
149. Ahmadi SZ, Ghorbanpour M, Aghae A, Hadian J (2020) Deciphering morpho-physiological and phytochemical attributes of *Tanacetum parthenium* L. plants exposed to C60 fullerene and salicylic acid. *Chemosphere* 259:127406. <https://doi.org/10.1016/j.chemosphere.2020.127406>
150. Caldelas MC, Poitrasson F, Viers J, Araus OJL (2020) Stable Zn isotopes reveal the uptake and toxicity of zinc oxide engineered nanomaterials in *Phragmites australis*. *Environ Sci Nano* 7:1927–1941. <https://doi.org/10.1039/D0EN00110D>
151. Lombi E, Donner E, Dusinska M (2019) One health approach to managing the applications and implications of nanotechnologies in agriculture. *Nat Nanotechnol* 14:523–531. <https://doi.org/10.1038/s41565-019-0460-8>
152. Mitter N, Hussey K (2019) Moving policy and regulation forward for nanotechnology applications in agriculture. *Nat Nanotech* 14:508–510. <https://doi.org/10.1038/s41565-019-0464-4>
153. Kumar JN, Bora A, Kumar RN, Amb MK, Khan S (2013) Toxicity analysis of pesticides on cyanobacterial species by 16S rDNA molecular characterization. *Proc Int Acad Ecol Environ Sci* 3:101
154. Hong JR (2014) Evidence of translocation and physiological impacts of foliar applied CeO₂ nanoparticles on cucumber (*Cucumis sativus*) plants. *Environ Sci Technol* 48:4376–4385. <https://doi.org/10.1021/es404931g>
155. Torre-Roche RDL (2012) Fullerene-enhanced accumulation of p, p'-DDE in agricultural crop species. *Environ Sci Technol* 46:9315–9323. <https://doi.org/10.1021/es301982w>
156. Kumar SS, Venkateswarlu P, Rao VR, Rao GN (2013) Synthesis, characterization and optical properties of zinc oxide nanoparticles. *Int Nano Lett* 3:1–6. <https://doi.org/10.1186/2228-5326-3-30>
157. Pejam F, Ardebili ZO, Ladan-Moghadam A, Danaee E (2021) Zinc oxide nanoparticles mediated substantial physiological and molecular changes in tomato. *PLoS ONE* 16(3):e0248778. <https://doi.org/10.1371/journal.pone.0248778>
158. Prazak R, Świącico A, Krzepilko A, Michalek S, Arczewska M (2020) Impact of Ag nanoparticles on seed germination and seedling growth of green beans in normal and chill temperatures. *Agriculture* 10:312. <https://doi.org/10.3390/agriculture10080312>
159. Sebastian A, Prasad MNV (2015) Trace element management in rice. *Agron* 5:374–404. <https://doi.org/10.3390/agronomy5030374>
160. Khan NM, Mobin M, Zahid A, Alamri S (2018) Fertilizers and their contaminants in soils, surface, and groundwater. *Environ Anthropol* 5:225–240. <https://doi.org/10.1016/B978-0-12-409548-9.09888-2>
161. Sebastian A, Nangia A, Majeti N, Vara P (2020) Advances in agrochemical remediation using nanoparticles Agrochemicals Detection. *Treat Remediat*. <https://doi.org/10.1016/B978-0-08-103017-2.00018-0465-484>
162. Al-Barly AMF, Hamza RZ (2015) Larvicidal, anti-oxidant activities and perturbation of transaminases activities of titanium dioxide nanoparticles synthesized using *Moringa oleifera* leaves extract against the red palm weevil (*Rhynchophorus ferrugineus*). *Eur J Pharm Med Res* 2:49–54. <https://doi.org/10.1080/21691401.2017.1408121>
163. Christofoli M, Costa ECC, Bicalho KU, de Cassia DV, Peixoto MF, Alves CCF, de Melo CC (2015) Insecticidal effect of nanoencapsulated essential oils from *Zanthoxylum rhoifolium* (Rutaceae) in *Bemisia tabaci* populations. *Ind Crops Prod* 70:301–308. <https://doi.org/10.1016/j.indcrop.2015.03.025>
164. Aseri A, Garg SK, Nayak A, Trivedi SK, Mohamed A (2015) Magnetic nanoparticles: magnetic nano-technology using biomedical applications and prospects. *Int J Pharm Sci Rev Res* 31:119–131. <https://doi.org/10.1002/adhm.201700845>
165. You G, Hou J, Xu Y, Miao L, Ao Y, Xing B (2021) Surface properties and environmental transformations controlling the bioaccumulation and toxicity of cerium oxide nanoparticles: a critical review. *Rev Environ Contam Toxicol* 253:155–206. https://doi.org/10.1007/978_2020_42
166. Grillo R, de Melo NF, de Araújo DR, de Paula E, Rosa AH, Fraceto LF (2010) Polymeric alginate nanoparticles containing the local anesthetic bupivacaine. *J Drug Target* 18:688–699. <https://doi.org/10.3109/10611861003649738>
167. Clemente Z, Castro VL, Moura MA, Jonsson CM, Fraceto LF (2014) Toxicity assessment of TiO₂ nanoparticles in zebrafish embryos under different exposure conditions. *Aquat Toxicol* 147:129–139. <https://doi.org/10.1016/j.aquatox.2013.12.024>
168. Ahsan SM, Rao CM, Ahmad MF (2018) Nanoparticle-protein interaction: the significance and role of protein corona. *Adv Exp Med Biol* 1048:175–198. https://doi.org/10.1007/978-3-319-72041-8_11

169. Severino P, da Silva CF, Andrade LN, de Lima OD, Campos J, Souto EB (2019) Alginate nanoparticles for drug delivery and targeting. *Curr Pharm Des* 25:1312–1334. <https://doi.org/10.2174/1381612825666190425163424>
170. Silva AM, Alvarado HL, Abrego G, Martins-Gomes C, Garduño-Ramirez ML, García ML, Calpena AC, Souto EB (2019) In vitro cytotoxicity of oleanolic/ursolic acids-loaded in PLGA nanoparticles in different cell lines. *Pharmaceutics* 11:362. <https://doi.org/10.3390/pharmaceutics11080362>
171. Khatri P, Chaudhary P, Gangola S, Bhatt P, Sharma A (2017) Nanochitosan supports growth of *Z. mays* and also maintains soil health following growth. *Biotech* 7:81. doi: <https://doi.org/10.1007/s13205-017-0668-y>.
172. Kubavat D, Trivedi K, Vaghela P, Prasad K, Vijay Anand GK, Trivedi H, Ghosh A (2020) Characterization of a chitosan-based sustained release nanofertilizer formulation used as a soil conditioner while simultaneously improving biomass production of *Z. mays* L. *Land Degrad Dev* 31:2734–2746. <https://doi.org/10.1002/ldr.3629>
173. Santo Pereira AE, Silva PM, Oliveira JL, Oliveira HC, Fraceto LF (2017) Chitosan nanoparticles as carrier systems for the plant growth hormone gibberellic acid. *Colloids Surf B Biointerfaces* 150:141–152. <https://doi.org/10.1016/j.colsurfb.2016.11.027>
174. El-Gazzar N, Almaary K, Ismail A, Polizzi G (2020) Influence of Funneliformis mosseae enhanced with titanium dioxide nanoparticles (TiO₂NPs) on Phaseolus vulgaris L under salinity stress. *PLoS ONE* 15:e0235355. <https://doi.org/10.1371/journal.pone.0235355>
175. Zayed MF, Eisa WH, Hezma AM (2017) Spectroscopic and antibacterial studies of anisotropic gold nanoparticles synthesized using *Malva parviflora*. *J Appl Spect* 83:1046–1050. <https://doi.org/10.1007/s10812-017-0406-6>
176. Mirbolook A, Rasouli-Sadaghiani M, Sepehr E, Lakzian A, Hakimi M (2020) Synthesized Zn (II)-amino acid and-chitosan chelates to increase Zn uptake by bean (*Phaseolus vulgaris*) plants. *J Plant Growth Regul.* <https://doi.org/10.1007/s00344-020-10151-y>
177. Asgari-Targhi G, Iranbakhsh A, Ardebili ZO (2018) Potential benefits and phytotoxicity of bulk and nano-chitosan on the growth, morphogenesis, physiology, and micropropagation of *Capsicum annuum*. *Plant Physiol Biochem* 127:393–402. <https://doi.org/10.1016/j.plaphy.2018.04.013>
178. Esyanti RR, Farah N, Bajra BD, Nofitasari D, Martien R, Sunardi S, Safitri R (2020) Comparative study of nano-chitosan and synthetic bactericide application on chili pepper (*Capsicum annuum* L.) infected by *Xanthomonas campestris*. *Agrivita* 42:13. <https://doi.org/10.17503/agrivita.v42i1.1283>
179. Maity A, Natarajan N, Vijay D, Srinivasan R, Pastor M, Malaviya DR (2018) Influence of metal nanoparticles (NPs) on germination and yield of Oat (*Avena sativa*) and Berseem (*Trifolium alexandrinum*). *Proc Natl Acad Sci India Sect B Biol Sci* 88:595–607. <https://doi.org/10.1007/s40011-016-0796-x>
180. Shahhoseini R, Azizi M, Asili J, Moshtaghi N, Samiei L (2020) Effects of zinc oxide nanoelicitors on yield, secondary metabolites, zinc and iron absorption of Feverfew (*Tanacetum parthenium* (L.) Schultz Bip.). *Acta Physiol Plant* 42:1–18. <https://doi.org/10.1007/s11738-020-03043-x>
181. Muthukrishnan S, Murugan I, Selvaraj M (2019) Chitosan nanoparticles loaded with thiamine stimulate growth and enhances protection against wilt disease in Chickpea. *Carbohydr Polym* 212:169–177. <https://doi.org/10.1016/j.carbpol.2019.02.037>
182. Zhang H, Lu L, Zhao X, Zhao S, Gu X, Du W, Wei H, Ji R, Zhao L (2019) Metabolomics reveals the “invisible” responses of spinach plants exposed to CeO₂ nanoparticles. *Environ Sci Technol* 53:6007–6017. <https://doi.org/10.1021/acs.est.9b00593>
183. Samadi S, Saharkhiz MJ, Azizi M, Samiei L, Ghorbanpour M (2020) Multi-walled carbon nanotubes stimulate growth, redox reactions and biosynthesis of antioxidant metabolites in *Thymus daenensis* celak in vitro. *Chemosphere* 249:126069. <https://doi.org/10.1016/j.chemosphere.2020.126069>
184. Mondal AH, Yadav D, Ali A, Khan N, Jin JO, Haq QMR (2020) Anti-bacterial and anti-candidal activity of silver nanoparticles biosynthesized using *Citrobacter* spp MS5 culture supernatant. *Biomolecules* 10:944. <https://doi.org/10.3390/biom10060944>
185. Tailor G, Yadav BL, Chaudhary J, Joshi M, Suvalka C (2020) Green synthesis of silver nanoparticles using *Ocimum canum* and their anti-bacterial activity. *Biochem Biophys Rep* 24:100848. <https://doi.org/10.1016/j.bbrep.2020.100848>
186. Francisco J, Frank LWT (2020) Biocontrol by *Fusarium oxysporum* using endophyte-mediated resistance. *Front Plant Sci.* <https://doi.org/10.3389/fpls.2020.00037>
187. Madany MM, Saleh AM, Habeeb TH, Hozzein WN, AbdElgawad H (2020) Silicon dioxide nanoparticles alleviate the threats of broomrape infection in tomato by inducing cell wall fortification and modulating ROS homeostasis. *Environ Sci Nano* 7:1415–1430. <https://doi.org/10.1039/C9EN01255A>
188. Enyedi NT, Makk J, Kótai L (2020) Cave bacteria-induced amorphous calcium carbonate formation. *Sci Rep* 10:8696. <https://doi.org/10.1038/s41598-020-65667-w>
189. Baran M, Keskin C, Atalar M, Baran A (2020) Environmentally friendly rapid synthesis of gold nanoparticles from *Artemisia absinthium* plant extract and application of antimicrobial activities. *J Inst Sci Technol* 11:365–375. <https://doi.org/10.21597/jist.779169>
190. Tan S, Wu X, Xing Y, Lilak S, Wu M, Zhao JX (2020) Enhanced synergetic antibacterial activity by a reduce graphene oxide/Ag nanocomposite through the photothermal effect. *Colloids Surf B: Biointerfaces* 185:110616. <https://doi.org/10.1016/j.colsurfb.2019.110616>
191. Ali M, Ahmed T, Wu W, Hossain A, Hafeez R, Islam M, Li B (2020) Advancements in plant and microbe-based synthesis of metallic nanoparticles and their antimicrobial activity against plant pathogens. *Nanomaterials* 10:1146. <https://doi.org/10.3390/nano10061146>
192. Orzali L, Valente MT, Scala V, Loreti S, Pucci N (2020) Antibacterial activity of essential oils and *Trametes versicolor* extract against *Clavibacter michiganensis* subsp. *michiganensis* and *Ralstonia solanacearum* for seed treatment and development of a rapid in vivo assay. *Antibiotics* 9:628. <https://doi.org/10.3390/antibiotics9090628>
193. Zhao L, Lu L, Wang A, Zhang H, Huang M, Wu H, Ji R (2020) Nanobiotechnology in agriculture: use of nanomaterials to promote plant growth and stress tolerance. *J Agric Food Chem* 68:1935–1947. <https://doi.org/10.1021/acs.jafc.9b06615>
194. Asma N, Mudassar I, Crispin H, Hassan W (2020) Biogenic AgNPs—a nano weapon against bacterial canker of tomato (bct). *Adv Agric* 2020(1–11):9630785. <https://doi.org/10.1155/2020/9630785>
195. Sonika D, Saurav K, Aakash G, Uttam L, Ranjita T, Shankar J, Ganesh L, Deval PB, Niranjana P (2021) Current research on silver nanoparticles: synthesis, characterization, and applications. *J Nanomat.* <https://doi.org/10.1155/2021/6687290>
196. Verma R, Chauhan A, Shandilya M, Li X, Kumar R, Kulshrestha S (2020) Antimicrobial potential of Ag-doped ZnO nanostructure synthesized by the green method using *Moringa oleifera* extract. *J Environ Chem Eng* 8:103730. <https://doi.org/10.1016/j.jece.2020.103730>
197. Jyoti K, Arora D, Fekete G, Lendvai L, Dogossy G, Singh T (2020) Antibacterial and anti-inflammatory activities of *Cassia fistula* fungal broth-capped silver nanoparticles. *Mater Technol* 1:11. <https://doi.org/10.1080/10667857.2020.1802841>
198. Khosrovyan A, Gabrielyan B, Kahru A (2020) Ingestion and effects of virgin polyamide microplastics on *Chironomus riparius* adult larvae and adult zebrafish *Danio rerio*. *Chemosphere* 259:127456. <https://doi.org/10.1016/j.chemosphere.2020.127456>
199. Alaraby M, Demir E, Domenech J, Velázquez A, Hernández A, Marcos R (2020) In vivo evaluation of the toxic and genotoxic effects of exposure to cobalt nanoparticles using *Drosophila melanogaster*. *Environ Sci Nano* 7:610–622. <https://doi.org/10.1039/C9EN00690G>
200. Raj A, Shah P, Agrawal N (2020) Impact of nanoparticles on behavior and physiology of *Drosophila melanogaster*. In: *Toxicology of nanoparticles: insights from Drosophila*. Springer, Singapore, pp. 59–67
201. Sahu S, Mishra M (2020) Hydroxyapatite nanoparticle causes sensory organ defects by targeting the retromer complex in *Drosophila melanogaster*. *NanolImpact* 19:100237. <https://doi.org/10.1016/j.nimpact.2020.100237>
202. Demir E (2020) An in vivo study of nanorod, nanosphere, and nanowire forms of titanium dioxide using *Drosophila melanogaster*: toxicity, cellular uptake, oxidative stress, and DNA damage. *J Toxicol Environ Health, Part A* 83:456–469. <https://doi.org/10.1080/15287394.2020.1777236>
203. Kumar D, Kumar P, Singh H (2020) Biocontrol of mosquito vectors through herbal-derived silver nanoparticles: prospects and challenges.

- Environ Sci Pollut Res 27:25987–26024. <https://doi.org/10.1007/s11356-020-08444-6>
204. Ahmad J, Qamar S, Kausar N, Qureshi MI (2020) Nanoparticles: the magic bullets in mitigating drought stress in plants. In: Nanobiotechnology in agriculture. Springer, Cham, pp 145–161
 205. Faraji J, Sepehri A (2020) Exogenous nitric oxide improves the protective effects of tio 2 nanoparticles on growth, antioxidant system, and photosynthetic performance of wheat seedlings under drought stress. *J Soil Sci Plant Nutr* 20:703–714. <https://doi.org/10.1007/s42729-019-00158-0>
 206. Tombuloglu H, Anil I, Akhtar S, Turumtay H, Sabit H, Slimani Y, Baykal A (2020) Iron oxide nanoparticles translocate in pumpkin and alter the phloem sap metabolites related to oil metabolism. *Sci Hortic* 265:109223. <https://doi.org/10.3390/nano10091654>
 207. Taran N, Storozhenko V, Sviatlova N, Batsmanova L, Shvartau V, Kovalevko M (2017) Effect of zinc and copper nanoparticles under drought resistance of wheat seedlings. *Nanoscale Res Lett* 12:60. <https://doi.org/10.1186/s11671-017-1839-9>
 208. Kolbert Z, Szöllösi R, Feigl G, Kónya Z, Rónavári A (2021) Nitric oxide signalling in plant nanobiology: current status and perspectives. *J Exp Bot* 72:928–940. <https://doi.org/10.1093/jxb/eraa470>
 209. Soliman M, Qari SH, Abu-Elsaud A, El-Esawi M, Alhathloul H, Elkesh A (2021) Rapid green synthesis of silver nanoparticles from blue gum augment growth and performance of maize, fenugreek, and onion by modulating plants cellular antioxidant machinery and genes expression. *Acta Physiol Plant* 42:1–16. <https://doi.org/10.3390/plants9040431>
 210. Alabdallah NM, Alzahrani HS (2020) The potential mitigation effect of ZnO nanoparticles on [*Abelmoschus esculentus* L. Moench] metabolism under salt stress conditions. *Saudi J Biol Sci* 27:3132–3137. <https://doi.org/10.1016/j.sjbs.2020.08.005>
 211. Elsheery NI, Helaly MN, El-Hoseiny HM, Alam-Eldein SM (2020) Zinc oxide and silicone nanoparticles to improve the resistance mechanism and annual productivity of salt-stressed mango trees. *Agronomy* 10:558. <https://doi.org/10.3390/agronomy10040558>
 212. Hoffmann J, Berni R, Hausman JF, Guerriero G (2020) A review on the beneficial role of silicon against salinity in non-accumulator crops: tomato as a model. *Biomolecules* 10:1284. <https://doi.org/10.3390/biom10091284>
 213. Zahedi SM, Karimi M, da Teixeira SJA (2020) The use of nanotechnology to increase quality and yield of fruit crops. *J Sci Food Agric* 100:25–31. <https://doi.org/10.1002/jsfa.10004>
 214. Mahmoud LM, Dutt M, Shalan AM, El-Kady ME, El-Boray MS, Shabana YM (2020) Silicon nanoparticles mitigate oxidative stress of in vitro-derived banana (*Musa acuminata* 'Grand Nain') under simulated water deficit or salinity stress. *South Afr J Bot* 132:155–163. <https://doi.org/10.1016/j.sajb.2020.04.027>
 215. Moradbeygi H, Jamei R, Heidari R, Darvishzadeh R (2020) Investigating the enzymatic and non-enzymatic antioxidant defense by applying iron oxide nanoparticles in *Dracocephalum moldavica* L. plant under salinity stress. *Sci Hortic* 272:109537. <https://doi.org/10.1016/j.scienta.2020.109537>
 216. Ye Y, Cota-Ruiz K, Hernández-Viezas JA, Valdés C, Medina-Velo IA, Turley RS (2020) Manganese nanoparticles control salinity modulated molecular responses in *Capsicum annum* L. through priming: a sustainable approach for agriculture. *ACS Sustain Chem Eng* 8:1427–1436. <https://doi.org/10.1021/acsschemeng.9b05615>
 217. Shoemaker AG (2020) The effects of titanium dioxide nanoparticles on the growth and development of *Sorghum Bicolor* (L.) Moench. *Adv Agric Hortic Entomol*. <https://doi.org/10.37722/AAHAE.202052>
 218. Thomas TD, Dinakar C, Puthur JT (2020) Effect of UV-B priming on the abiotic stress tolerance of stress-sensitive rice seedlings: priming imprints and cross-tolerance. *Plant Physiol Biochem* 147:21–30. <https://doi.org/10.1016/j.plaphy.2019.12.002>
 219. Kardavan GV, Karamian R (2020) Effects of TiO₂ nanoparticles and spermine on antioxidant responses of *Glycyrrhiza glabra* L. to cold stress. *Acta Bot Croat*. <https://doi.org/10.37427/botcro-2020-025>
 220. Iqbal MS, Singh AK, Singh SP, Ansari MI (2020) Nanoparticles and plant interaction with respect to stress response. *Nano Environ Biotechnol*. https://doi.org/10.1007/978-3-030-34544-0_1
 221. Joško I, Oleszczuk P, Futa B (2014) The effect of inorganic nanoparticles (ZnO, Cr₂O₃, CuO and Ni) and their bulk counterparts on enzyme activities in different soils. *Geoderma* 232:528–537. <https://doi.org/10.1016/j.geoderma.2014.06.012>
 222. Joško I, Dobrzyńska J, Dobrowolski R, Kusiak M, Terpiłowski K (2020) The effect of pH and ageing on the fate of CuO and ZnO nanoparticles in soils. *Sci Total Environ* 721:137771. <https://doi.org/10.1016/j.scitotenv.2020.137771>
 223. Dogaroglu ZG, Koleli N (2014) Effect of different zinc oxide nanoparticles on germination, plant growth and chlorophyll content of wheat. In: International congress on green infrastructure and sustainable societies/cities greinsus, vol 14, pp 78–84. <https://doi.org/10.1080/02772248.2013.803796>
 224. Mokarram-Kashtiban S, Hosseini SM, Tabari Kouchaksaraei M, Younesi H (2019) The impact of nanoparticles zero-valent iron (nZVI) and rhizosphere microorganisms on the phytoremediation ability of white willow and its response. *Environ Sci Pollut Res Int* 26:10776–10789. <https://doi.org/10.1016/j.chemosphere.2020.126909>
 225. Szymanski M, Dobrucka R (2020) Evaluation of phytotoxicity of bimetallic Ag/Au nanoparticles synthesized using *Geum urbanum* L. *J Inorg Organomet Polym Mater* 1:12. <https://doi.org/10.1016/j.chemosphere.2009.01.078>
 226. Heikal YM, Şuţan NA, Rizwan M, Elsayed A (2020) Green synthesized silver nanoparticles induced cytogenotoxic and genotoxic changes in *Allium cepa* L. varies with nanoparticles doses and duration of exposure. *Chemosphere* 243:125430. <https://doi.org/10.1016/j.chemosphere.2019.125430>
 227. Taheri SM, Aramideh S, Akbarian J, Pirsa S (2020) Effects of ZnO nanoparticles and kaolin in combination with Neem Azal-T/S against *Bemisia tabaci* and its parasitoid *Eretmocerus mundus* on cotton. *Chem Rev Lett* 3:131–139. <https://doi.org/10.22034/CRL.2020.235381.1066>
 228. Hafiz UR, Waqas A, Wahab N, Mansur AS, Anwaar A, Nauman K (2021) A comprehensive review on chlorpyrifos toxicity with special reference to endocrine disruption: evidence of mechanisms, exposures and mitigation strategies. *Sci Total Environ* 755:142649. <https://doi.org/10.1016/j.scitotenv.2020.142649>
 229. Lozano-Pérez AA, Pagán A, Zhurov V (2020) The silk of gorse spider mite *Tetranychus lintearius* represents a novel natural source of nanoparticles and biomaterials. *Sci Rep* 10:18471. <https://doi.org/10.1038/s41598-020-74766-7>
 230. Sun C, Yu M, Zeng Z, Francis F, Cui H, Verheggen F (2020) Biocidal activity of polylactic acid-based nano-formulated abamectin on *Acyrtosiphon pisum* (Hemiptera: Aphididae) and the aphid predator *Adalia bipunctata* (Coleoptera: Coccinellidae). *PLoS ONE* 15:e0228817. <https://doi.org/10.1371/journal.pone.0228817>
 231. Noudagar ME, Mujtaba MA, Safaei MR, Afzal A, Ahmed W, Banapurmath NR, Hossain N, Bashir S, Badruddin IA, Goodarzi M, Shahapurkar K (2021) Effect of Sr@ ZnO nanoparticles and Ricinus communis biodiesel-diesel fuel blends on modified CRDI diesel engine characteristics. *Energy* 215:119094. <https://doi.org/10.1016/j.energy.2020.119094>
 232. Singh BK, Pandey R, Singh AK, Mishra MK (2020) Effectiveness of flonicamid 50 wg and flupyradifurone 200 SL against leafhopper and whitefly in okra. *J Entomol Zool Stud* 8:181–185
 233. Da Silva CL, Henriques RO, Rios JV, Lerin LA, de Oliveira D, Furigo A (2020) Lipase-catalyzed esterification of geraniol and citronellol for the synthesis of terpenic esters. *App Biochem Biotechnol* 190:574–583. <https://doi.org/10.1007/s12010-019-03102-1>
 234. Attaullah MKZ, Muhammad AZ, Muhammad SM, Hina R, Humara NM, Muhammad Z, Kanwal R, Kishwar S, Muhammad I, Samina Q (2020) Insecticidal, biological and biochemical response of *Musca domestica* (Diptera: Muscidae) to some indigenous weed plant extracts. *Saudi J Biol Sci* 27:106–116. <https://doi.org/10.1016/j.sjbs.2019.05.009>
 235. Sabbour MMA (2020) Efficacy of nano-formulated certain essential oils on the red flour beetle *Tribolium castaneum* and confused flour beetle, *Tribolium confusum* (Coleoptera: Tenebrionidae) under laboratory and storage conditions. *Bull Natl Res Cent* 44:111. <https://doi.org/10.1186/s42269-020-00336-6>
 236. Raveau R, Fontaine J, Lounès H, Sahraoui A (2020) Essential oils as potential alternative biocontrol products against plant pathogens and weeds: a review. *Foods* 9:365. <https://doi.org/10.1186/s42269-020-00336-6>
 237. Nazima S, Prasanta KR, Diganta G, Dipankar D, Saidul I, Varun T, Bodhadiya D, Hemanta KG, Pronobesh C, Pakalapati SR (2020) Bio-nanoparticle

- assembly: a potent on-site biolarvicidal agent against mosquito vectors. *RSC Adv* 10:9356–9368. <https://doi.org/10.1039/C9RA09972G>
238. Abouelatta AM, Keratum AY, Ahmed SI, El-Zun HM (2020) Repellent, contact and fumigant activities of geranium (*Pelargonium graveolens* L'Hér) essential oils against *Tribolium castaneum* (Herbst) and *Rhyzopertha dominica* (F.). *Int J Tropical Insect Sci* 40:1021–1030. <https://doi.org/10.1007/s42690-020-00161-4>
239. Campolo O, Puglisi I, Barbagallo RN, Cherif A, Ricupero M, Biondi A, Zappala L (2020) Side effects of two citrus essential oil formulations on a generalist insect predator, plant and soil enzymatic activities. *Chemosphere* 257:127252
240. Ikawati S, Himawan T, Abadi AL, Tarno H (2020) Toxicity nano-insecticide based on clove essential oil against *Tribolium castaneum* (Herbst). *J Pest Sci* 46:222–228. <https://doi.org/10.1584/jpestics.D20-059>
241. Leslie B, Mark M, Leonard L, Matteo S (2020) Efficacy of various herbicides for the control of perennial *Plantago* spp. and effects on alfalfa damage and yield. *Agronomy* 10:1710. <https://doi.org/10.3390/agronomy10111710>
242. Thongpitak J, Pumas P, Pumas C (2020) Paraquat degradation by biological manganese oxide (BioMnOx) catalyst generated from living microalga *Pediastrum duplex* AARL G060. *Front Microbiol* 11:575361. <https://doi.org/10.3389/fmicb.2020.575361>
243. Francisco CF, María EAG, Claudia LA, Balam RR, Roberto LVG, Patricia RC, Rocio ACS, Alexey P, Yanis TM, Juan CGR, Nina B (2020) Argovit™ silver nanoparticles effects on *Allium cepa*: plant growth promotion without cyto genotoxic damage. *Nanomaterials* 10:1386. <https://doi.org/10.3390/nano10071386>
244. Broda M (2020) Natural compounds for wood protection against fungi—a review. *Molecules* 25:3538. <https://doi.org/10.3390/molecules25153538>
245. Kivrak I, Kivrak S, Karababa E (2020) Assessment of bioactive compounds and antioxidant activity of turkey tail medicinal mushroom *Trametes versicolor* (Agaricomycetes). *Int J Med Mushrooms* 22:559–571. <https://doi.org/10.1615/IntJMedMushrooms.2020035027>
246. Cui J, Sun C, Wang A, Wang Y, Zhu H, Shen Y, Li N, Zhao X, Cui B, Wang C, Gao F, Zeng Z, Cui H (2020) Dual-functionalized pesticide nanocapsule delivery system with improved spreading behavior and enhanced bioactivity. *Nanomaterials* 10:220. <https://doi.org/10.3390/nano10020220>
247. Luis AP, Ana AFP, Ramón G, Sandra M, Karen E (2020) Nanoparticles in agroindustry: applications, toxicity, challenges, and trends. *Nanomaterials* 10:1654. <https://doi.org/10.3390/nano10091654>
248. Marcela VH, Israel MB, Ramon GG, Enrique RG, Rosalia VOV, Luciano AJ, Irineo TP (2020) Nanoparticles as potential antivirals in agriculture. *Agriculture* 10:444. <https://doi.org/10.3390/agriculture10100444>
249. Arshad A, Temoor A, Wenge W, Afsana H, Rahila H, Md. Mahidul IM, Yanli W, Qianli A, Guochang S, Bin L (2020) Advancements in plant and microbe-based synthesis of metallic nanoparticles and their antimicrobial activity against plant pathogens. *Nanomaterials* 10:1146. <https://doi.org/10.3390/nano10061146>
250. Piela A, Żymańczyk DE, Brzezińska RM, Duda M, Grzesiak J, Saeid A, Klimek OM (2020) Biogenic synthesis of silica nanoparticles from corn cobs husks. Dependence of the productivity on the method of raw material processing. *Bioorg Chem* 99:103773. <https://doi.org/10.1016/j.bioorg.2020.103773>
251. Hafez YM, Attia KA, Kamel S, Alameri SF, El-Gendy S, Al-Doss AA, Abdelaal KA (2020) *Bacillus subtilis* as a bio-agent combined with nano molecules can control powdery mildew disease through histochemical and physiobiochemical changes in cucumber plants. *Physiol Mol Plant Pathol* 111:101489. <https://doi.org/10.1016/j.pmp.2020.101489>

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