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Software tools of the Computis European project to process mass spectrometry images

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Keywords: imaging mass spectrometry, image processing, visualization, clustering, image registration, standard format

Abstract

Among the needs usually expressed by teams using mass spectrometry imaging, often arise user-friendly software able to quickly manage huge data volume and to provide efficient assistance for the interpretation of data. To answer this need, the Computis European project developed several complementary software tools to process mass spectrometry imaging data.

Data Cube Explorer provides a simple spatial and spectral exploration for MALDI-ToF and ToF-SIMS data. SpectViewer offers visualization functions, assistance to the interpretation of data, classification functionalities, peak list extraction to interrogate biological database, image overlay and can process data issued from MALDI-ToF, ToF-SIMS and DESI equipments. EasyReg2D is able to register two images, in ASCII format, issued from different technologies.

The collaboration between teams being hampered by the multiplicity of equipments and data formats, the project also developed a common data format (imzML) to facilitate the exchange of experimental data and their interpretation by the different software tools. The BioMap platform for visualization and exploration of MALDI-ToF and DESI images was adapted to parse imzML files, enabling its access to all project partners and more globally to a larger community of users.

Considering the huge advantages brought by the imzML standard format, a specific editor (vBrowser) for imzML files and converters from proprietary formats to imzML were developed to enable the use of imzML format by a broad scientific community. This initiative is paving the way towards the development of a large panel of software tools able to process mass spectrometry imaging datasets in the future.

Introduction

Since the emergence of mass spectrometry imaging almost twenty years ago in SIMS,¹⁻²⁻³⁻⁴ and MALDI technologies,⁵⁻⁶⁻⁷ this technique appeared as a major discovery tool in life sciences for detection, localization and identification of molecules in biological tissues,⁸⁻⁹ and for understanding the cellular processes involved in both health and disease,¹⁰.

Since then, the technology was perfected,¹¹ to improve the sensitivity, accuracy and spatial resolution of mass spectrometry imaging instruments,¹²⁻¹³⁻¹⁴⁻¹⁵ and to optimize ionization and reproducibility with appropriate sample preparation and treatment methods not introducing analyte delocalization or degradation,¹⁶⁻¹⁷⁻¹⁸⁻¹⁹⁻²⁰. The range of molecules was considerably extended so that it is now possible to measure molecular weights above 200 kDa,²¹⁻²² to reach mass measurement accuracy of sub-parts per million,²³⁻²⁴ and to detect a particular compound in the order of low fmol/ μ^2 ,²⁵.

Datasets acquired with state-of-the-art instrumentation often include thousands of mass spectra, each of which comprising thousands of mass channels, therefore mass spectrometry imaging outputs regularly amount gigabytes of data and it is essential to develop automated software to analyze the huge spectra rapidly and efficiently,²⁶⁻²⁷ and specific methods adapted to process such amount of data,²⁸⁻²⁹⁻³⁰⁻³¹.

For imaging mass spectrometry, available software is limited to proprietary software developed by the equipment manufacturers and linked to specific equipment (flexImaging from Bruker Daltonics, TissueView from AB Sciex, SurfaceLab from IonTof), and very few free of charge software such as BioMap or MITICS,³². If all these software tools provide image reconstruction and exploration functions, few of them include processing functionalities such as clustering or multivariate analysis,³³⁻³⁴⁻³⁵⁻³⁶ to help users in data analysis and interpretation.

To contribute to the improvement impetus of mass spectrometry imaging technology, the Computis European project (2006-2010) <http://www.computis.org/> was devoted to the development of high-resolution imaging instruments, software affording a large panel of analytical functions, and to use these new capacities in biological and medical applications.

The Computis project dedicated large efforts to the development of software tools for data processing and visualization. Many basic but largely used features such as zooming, region-of-interest analysis, image cropping, intensity-scale and color palette for image display, intensity profiles, peak and pixel picking, true-data or binned-data display, were developed. More specialized processing functions such as denoising and baseline subtraction, clustering, multivariate analysis and image registration, were also elaborated with the constraint to identify methods adapted to the management of enormous amounts of spectral data obtained from the imaging of tissue sections.

Three software tools were developed during the Computis project: Data Cube Explorer for a simple exploration of data and classification with the Kohonen network, SpectViewer for exploration, classification, data interpretation and connection with biological databases, and EasyReg2D for image registration.

One of the goals of the Computis project was to develop software compatible with data of all partners. Nevertheless, the multiplicity of data formats - issued from the use by the partners of various devices from several manufacturers in MALDI and SIMS imaging - quickly resulted difficult to manage. The need clearly appeared to have a common format allowing the comparison of images and datasets from the different partners.

To address this limitation, the partners examined the different standards available: netCDF/ANDI-MS,³⁷ from ASTM International, mzXML,³⁸⁻³⁹ from the Institute of Systems Biology, mzData,⁴⁰ from Human Proteome Organization (HUPO) and the joint format

mzML,⁴¹⁻⁴² (released in 2008 but under development at the time). However, it appeared that mzML could not completely describe a 2D MS imaging experiment as some 2D parameters (including x/y position, scan direction/pattern, pixel size) were not available and that data storage was not efficient enough. Therefore the Computis European project developed the imzML data format,⁴³⁻⁴⁴⁻⁴⁵ for imaging MS data and Justus Liebig University took the leadership of this development.

In the imzML format, the MS imaging data is divided in two separate files. The mass spectral data is stored in a binary file to ensure efficient storage. All metadata (instrumental parameters and sample details) are stored in a XML file with an extended controlled vocabulary to include specific parameters of mass spectrometry imaging. The two files (XML and binary) are connected by offset values in the XML file and are unambiguously linked by a universally unique identifier. The resulting datasets are comparable in size to the raw data and the separate metadata file allows flexible handling of large datasets. Specifications and example files for imzML can be downloaded at <http://www.imzML.org>.

Considering the advantages brought by the imzML standard format and its interest for the mass spectrometry community, the Computis project developed a specific editor for imzML files and converters from proprietary formats to imzML format to help the scientific community to use the imzML format. As all data formats of the Computis partners could not be read by the BioMap platform developed by Novartis, BioMap was also adapted to parse imzML files, enabling its access to all project partners and more globally to a larger community of users.

Data Cube Explorer

Data Cube Explorer is a user-friendly tool under Windows to provide an easy spectral and spatial exploration of MALDI-ToF and ToF-SIMS imaging mass spectrometry datasets. It enables zooming within spectra and scrolling through the dataset masses for images with a manual greyscale tuning to improve image contrast. Regions Of Interest can be selected with the display of the associated spectra. Developed by the Dutch Foundation for Fundamental Research of Matter (FOM-Amolf), Data Cube Explorer,⁴⁶⁻⁴⁷⁻⁴⁸ can be downloaded at <http://www.maldi-msi.org>.

The “self-organizing map” functionality classifies images according to the intensity of all pixel places and automatically selects a given number of images as different as possible. The classification method used is unsupervised competitive learning, also known as Kohonen neural network,⁴⁹. For performing this analysis, the image data set is converted into a set of images, defined by a start mass and an end mass, the mass bin per image and the step size to move to a next image. A threshold value is used to distinguish between images containing noise and images containing real image data.

One after the other, the images are fed into the network; a winning output image is calculated, where winning means that the “distance” of the fed image to the output images is minimal. The distance is defined as root mean square of the difference in the pixel intensities of all pixel places. The winning output image (and the images around it defined by the neighborhood value) are adapted to the input image with an learning rate factor. During the process both the neighborhood value and the learning rate are reduced (to respectively 1 and 0).

Figure 1 illustrates the use of Data Cube Explorer on a rodent urinary bladder dataset (image from Justus Liebig University). An image of the peak at m/z 171.10 Th is presented as well as

spectra associated to the entire image, and the two Regions of Interest selected in the image. The self-organizing map is able to bring out the most interesting images in the dataset.

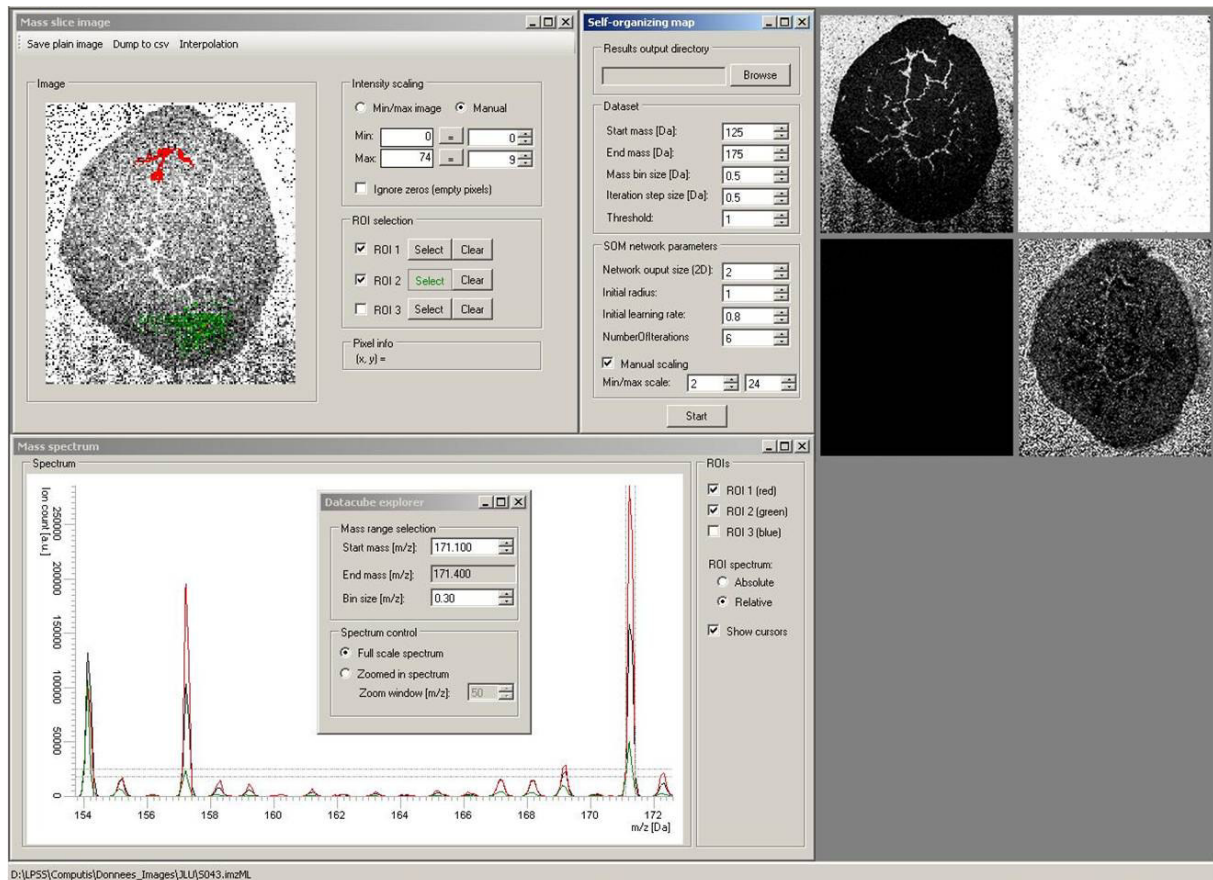


Figure 1: Application of Data Cube Explorer on a rodent urinary bladder dataset

Bottom: Selection of a mass interval $171.1 < m/z < 171.4$ in the spectrum; display of the sample average spectrum (black line) and the spectra of the selected regions of interest (red and green lines)

Top left: Image of the mass interval selected in the spectrum and selection of two Regions of Interest by coloring the area by moving the computer mouse

Top center: Parameters for the self-organizing map computation (zone of the spectrum, number of images, parameters of the self-organizing map algorithm)

Top right: Images selected by the self-organizing map method (143.1 Th, 153.1 Th, 156 Th, 171.2 Th)

SpectViewer

SpectViewer is a software module developed by French Atomic Energy Commission (CEA) under Windows and Linux for processing and visualization of MALDI-ToF, ToF-SIMS and DESI mass spectrometry imaging, with several tools providing user assistance for the interpretation of data. SpectViewer handles datasets in Analyze 7.5 (Applied Biosystems), GRD generated by SurfaceLab6 software (IonTof), BrukerFlex (Bruker), Orbitrap (ThermoFisher) and imzML formats. SpectViewer is available through partnership with French Atomic Energy Commission.

In addition to classical data display and exploration functionalities (spectrum and image display, peak and pixel picking, zooming on spectra and images, ROI selection), SpectViewer

extracts the peak list for faster molecule identification with public biological databanks, and provides some more specialized treatments such as denoising spectra or structure analysis by clustering methods. SpectViewer offers a fast processing and display of original data: no binning for MALDI and DESI data, no binning or user-defined binning for SIMS data. It also enables to overlay the images of two or three peaks of the dataset to compare their respective locations in the sample.

As assistance to the interpretation of data, SpectViewer computes several indicators:

- The relative variance spectrum is an indicator of how much the image corresponding to a peak deviates from a uniform Poisson noise image. This simple and fast tool greatly highlights peaks that have a highly contrasted spatial distribution.
- Moran index,⁵⁰ is an autocorrelation indicator measuring the spatial auto-covariance of a point with the neighbour points. This index evaluates if a local pattern is clustered, dispersed or random. It highlights peaks associated to clustered spatial areas, which is particularly adapted to detect thin local structures such as membranes, in images.
- The correlation matrix can be calculated between all peaks (computed from binned data) or between one given m/z and all other bins of m/z . The correlation spectrum associated to one given m/z brings out correlated m/z , which are often co-localised or complementary with the given m/z .

SpectViewer also includes several clustering tools to perform spatial (i.e. pixel-based) classification or spectral (i.e. m/z -based) classification. The K-means clustering is one of the simplest and fastest classification methods. The time for running a K-means clustering,⁵¹ usually lasts only some seconds. Stochastic K-means,⁵² optimizes both the cluster position and the cluster number. The random projection tree clustering,⁵³⁻⁵⁴ consists in classifying data by random tree coupled to random projection sorting rules and a dimensionality reduction method. The hierarchical clustering performs clustering inside a zone defined by a preceding clustering,⁵⁵.

The diffusion map method,⁵⁶⁻⁵⁷⁻⁵⁸ uses the eigenfunctions of a Markov transition matrix, defining a random walk on the data, to represent spectral data as a cloud of points in the Euclidean space. The eigenvectors of the Markov matrix are used as coordinates of the data set. The diffusion distance between a pair of points is calculated as a L2 Euclidean distance weighted by the eigenvalues of the Markov matrix. By keeping only the top eigenvectors, it is possible to reduce the dimensionality of the problem with a limited precision loss. Then a clustering analysis can be performed in the reduced data.

According to our experience, it is necessary to have several classification methods available as no method is able to classify perfectly all biological samples. The K-means algorithm is often giving good results at little CPU cost and provides structured classification images that can be physically interpreted. But in some cases, the K-means method is not able to classify data and more complex methods, such as the diffusion map method, are necessary.

The use of SpectViewer is presented on a coronal section of mouse brain tissue (part of the corpus callosum and caudate putamen of $256 \times 256 \mu\text{m}^2$) analyzed by French National Center for Scientific Research – Institute for Natural Substance Chemistry (CNRS-ICSN) in positive mode with Tof-SIMS equipment.

Figure 2a presents the total image with the associated total spectrum, the relative variance spectrum enabling to discriminate interesting peaks in the spectrum, and the correlation spectrum with m/z 385.34 Th. The relative variance highlights cholesterol (m/z 385.34 Th) in

the corpus callosum and m/z 158.92 Th in the caudate putamen. Peak at m/z 158.92 Th is also inversely correlated to m/z 385.34 Th. Both peaks provide complementary images.

Figure 2b displays a zoom of the total image with m/z included in 1 to 100 Th, as the red spots are particularly visible below 100 Th. In order to characterize the high-intensity spots in the caudate putamen (red spots in the total image), the spectra of pixel ($x=118, y=185$) in a red spot and pixel ($x=129, y=175$) in a green zone were displayed. By comparison of the spectra, it appears that $m/z = 66.01$ and 107.02 Th are localized in the spots.

Figure 4c shows a K-means classification on pixels with 4 clusters, associated to the cluster spectra. The cluster image shows cluster 0 in blue, cluster 1 in green, cluster 2 in orange and cluster 3 in red (color correspondence is indicated in the bar scale on the right side of the cluster image). The corpus callosum and the caudate putamen are well separated and subzones fitting with the spots are identified.

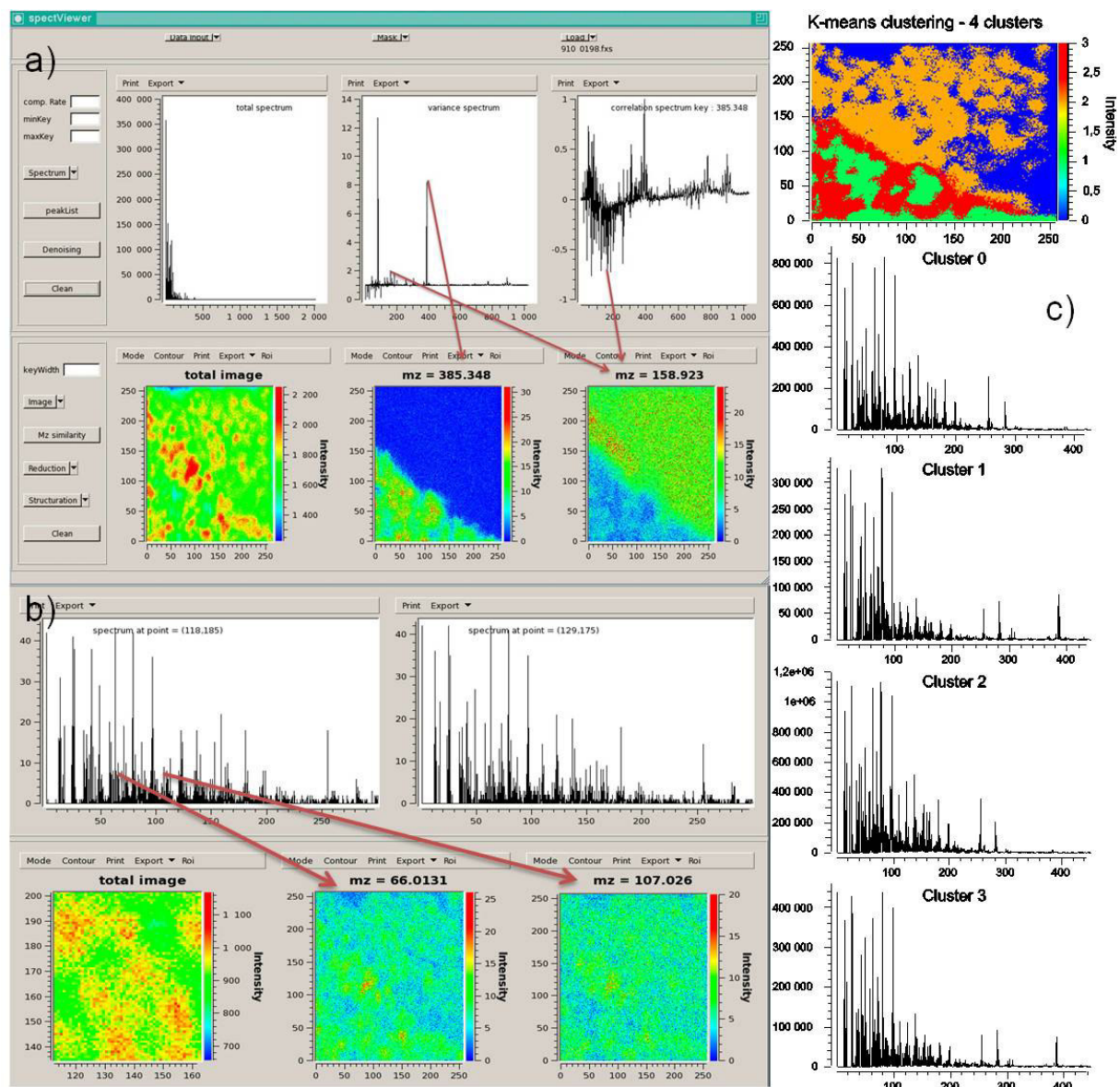


Figure 2: Application of SpectViewer on a mouse brain tissue dataset

EasyReg2D

EasyReg2D is a data fusion module developed by French Atomic Energy Commission (CEA) under Linux to register two images. The method supports multimodal registrations, for example microscopic image coming from a histopathology analysis with an image extracted from mass spectrometry data. Input image format is a plain ASCII file, containing a bidimensional array of gray intensity values (integers or floating numbers). Images in standard formats (jpeg, tiff, gif, bmp) can be converted in ASCII format by ImageJ free software <http://rsbweb.nih.gov/ij/>.

The geometric transformation registering the two images is an affine transform (6 degrees of freedom in 2D); the merit function is the mutual information,⁵⁹⁻⁶⁰. The merit function is minimized using a Quasi-Newton method. If needed, the graphical user interface allows the user to provide the optimizer with a good starting point (semi-manual registration).

Figure 3 shows the application of EasyReg2D on a microscopy image and a ToF-SIMS image of a rat brain tissue provided by French National Center for Scientific Research – Institute for Natural Substance Chemistry (CNRS-ICSN). Registration is initialized with two user-defined starting values (top). Initial images are superimposed without registration (middle) and after registration (bottom).

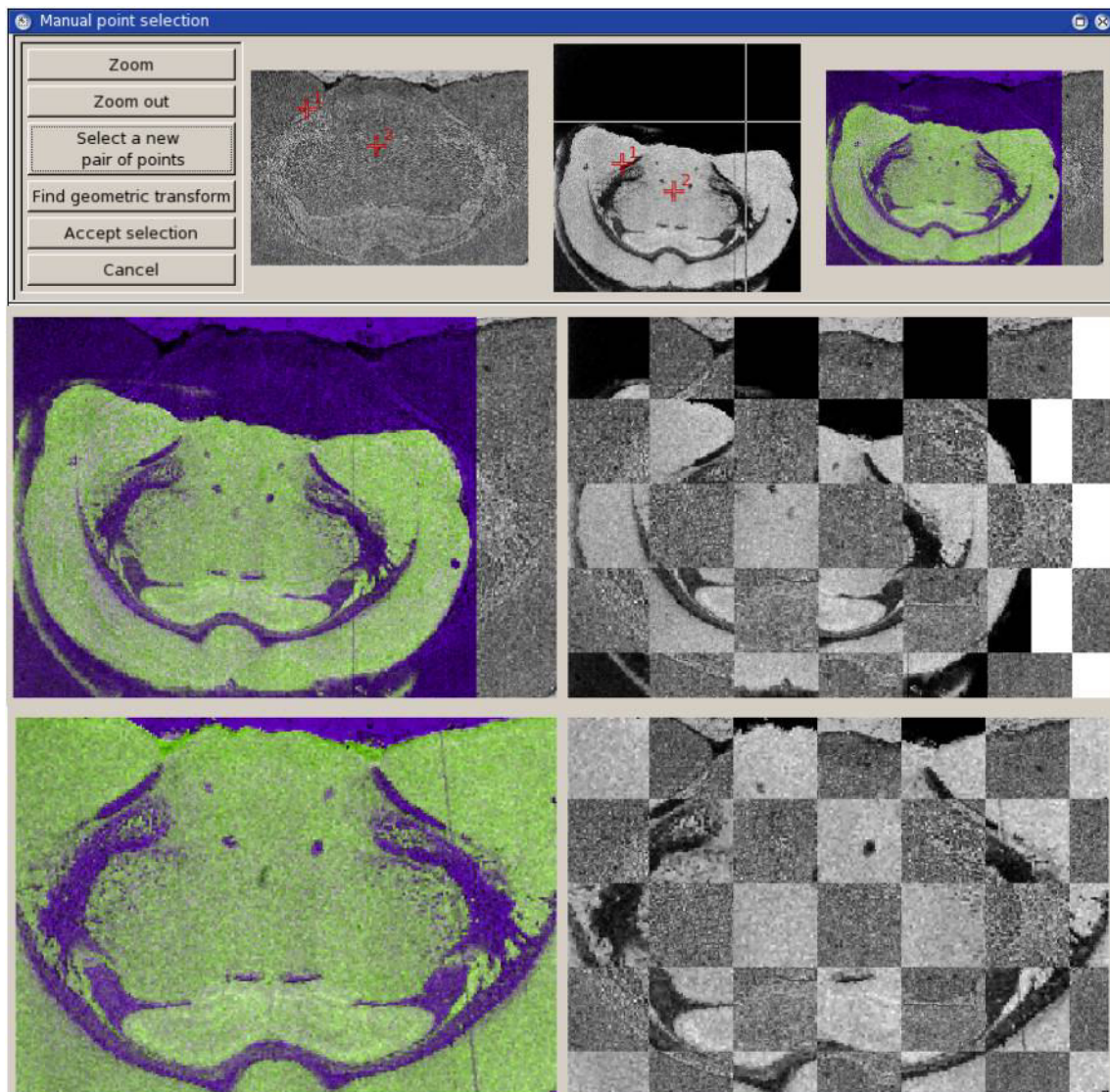


Figure 3: Application of EasyReg2D on a rat brain tissue dataset

BioMap

BioMap,⁶¹ is a free image analysis platform for Mass Spectrometry and Magnetic Resonance Imaging developed by Novartis under IDL. Visualization is based on multi-planar reconstruction allowing the extraction of arbitrary slices from a 3D-volume. Initially dedicated to data in Analyze format for MSI, Novartis adapted BioMap during the Computis project to also read imzML format. BioMap is available under Windows and Linux for MALDI-ToF and DESI data. It can be downloaded at <http://www.maldi-msi.org>

Well-known by proteomics teams for its visualization capacities and biology well-adapted functionalities, BioMap allows spectrum and image display with numerous colour tables, geometrical transformations (translation, rotation, flipping and resizing of images), adjustment of intensity, zooming on images and spectra, selection and treatment of multiple ROIs, statistical and histogram analysis, geometrical operations, and annotation of images. Simple calculations on images are available: spatial or temporal filtering, baseline correction, detrending (removal of a linear drift of the signal).

In order to manage large datasets with limited memory computers, BioMap offers the possibility to download and process only part of a dataset (a range of m/z). Display functions enabling to view simultaneously all images of a dataset or to create a movie are particularly useful to find the interesting m/z . Co-registration enables to superimpose and compare several images issued from different slices of a sample or different compatible samples, or to compare the position of patterns appearing at different m/z .

An example of BioMap is presented on a human cadaveric abdominal skin biopsy which was treated topically with an anti-acne development compound at m/z 466 (image from Novartis). The tissue was sectioned to provide a cross section of the full thickness of the skin (from epidermis to hypodermis/subcutaneous fat).

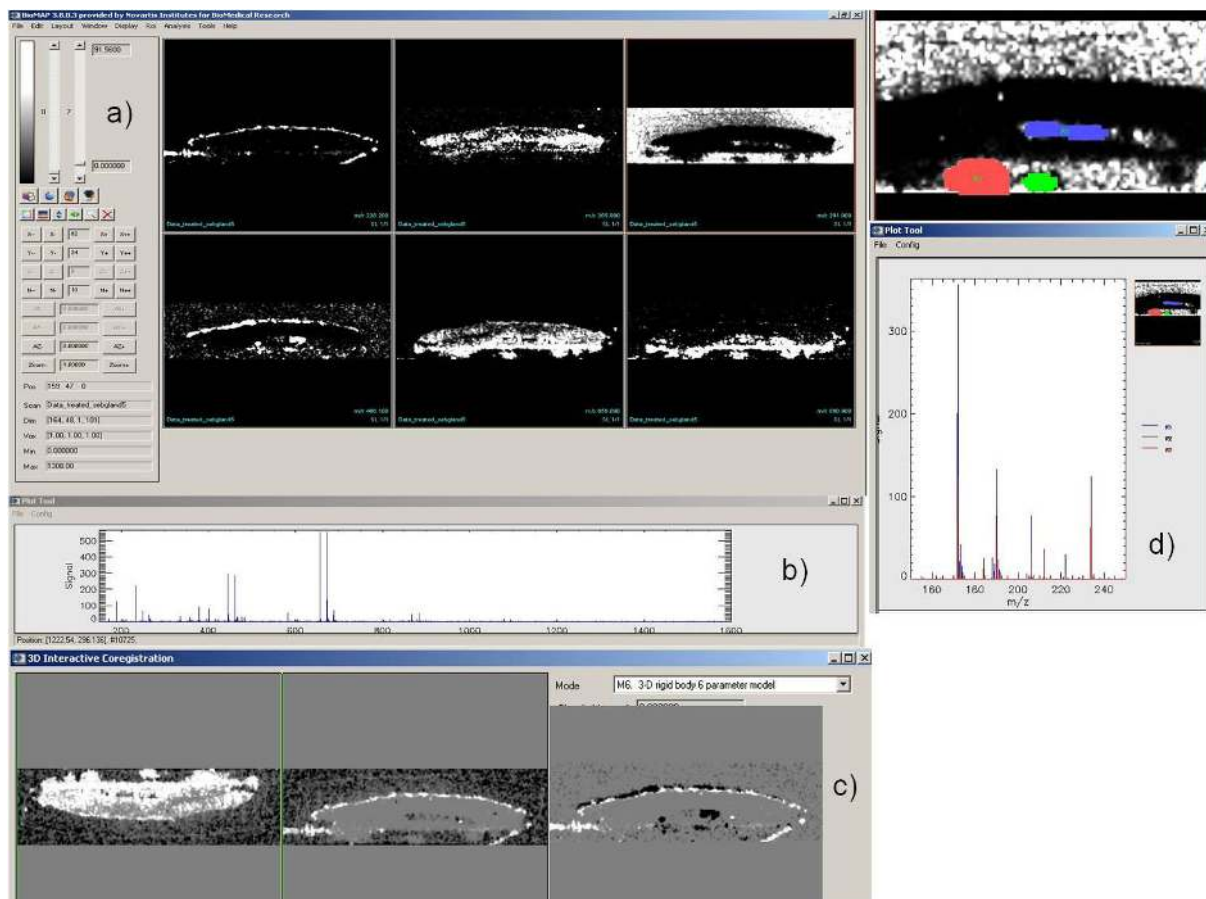


Figure 4: Application of BioMap on a sebaceous gland dataset

Figure 4a displays some characteristic m/z in positive mode, highlighting specific parts of the image: 227.22 Th,⁶² (halaminol A $C_{14}H_{29}NO$, a sphingoid base with an antimicrobial activity present at the skin surface), 265.9 Th, 290.44 Th (dihydrotestosterone), 466 Th (anti-acne compound), 655.63 Th,⁶³ (18:1 cholesteryl ester $C_{45}H_{73}D_5O_2$) and 860.9 Th. Figure 4b shows the total spectrum of the sample.

Figure 4c illustrates the co-registration function of Biomap. Complementary images at m/z 290.44 and 655.63 Th were co-registered after a rotation of 180° of the first image. The image at m/z 228.2 Th was superimposed with the one at m/z 291 Th and the one at m/z 466.1 Th to show the position of this thin zone compared to the others.

In figure 4d, three Regions Of Interest (ROI) were selected and the spectra of each ROI are displayed.

vBrowser editor and imzML plug-in

vBrowser provides a single frontend application to the networked resources. It renders a file-based GUI for the access of the resources, similar to an explorer-like browser. It supports Grid-based file systems like Grid-FTP, SSH-FTP, SRM, LFC and SRB out of the box while presenting them to the user in a homogeneous and familiar tree-like structure. vBrowser is developed by Power Computing & Communication and is part of the Dutch Virtual Laboratory for e-sciences,⁶⁴⁻⁶⁵.

The imzML plugin (Figure 5) from vBrowser provides advanced features for browsing imzML data files. It provides both tree-like and text based navigation of imzML metadata.

The imzML metadata can be represented as an organized tree, an XML text, or can be edited with any text editor.

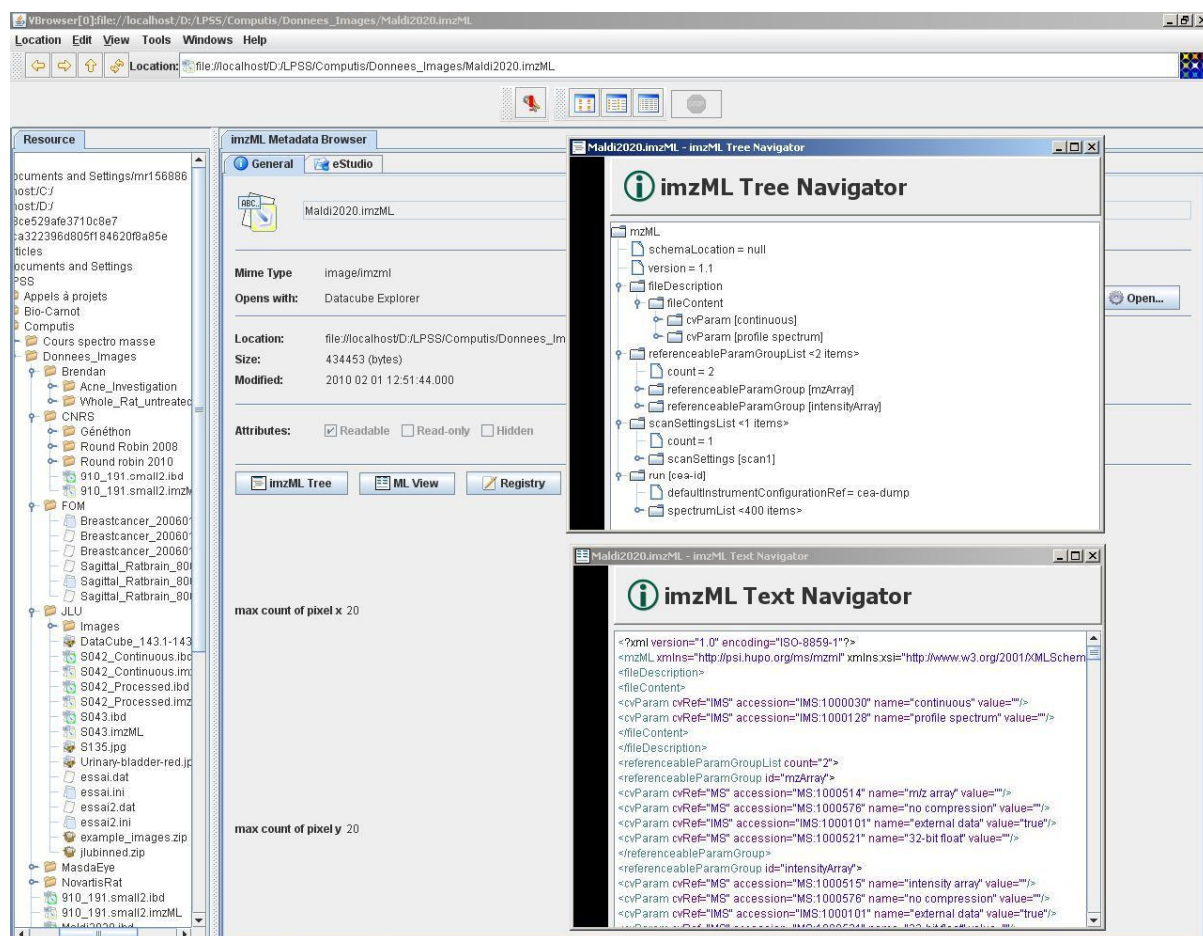


Figure 5: imzML metadata browser for file exploration and navigation in imzML data

imzML converters

Two converters were developed under Windows to generate imzML files out of proprietary formats. The “raw to imML converter” was developed by Justus Liebig University for raw files from Thermo Fisher Scientific. This tool can also combine multiple raw files into one imzML file, which can be used to process DESI imaging data. It can be freely downloaded on <http://www.imzml.org>.

The converter “ToimzML” was developed by French Atomic Energy Commission for Analyze 7.5 files from Applied Biosystems, GRD files generated by SurfaceLab6 (versions 6.0, 6.1 and 6.2) from Ion-ToF, BrukerFlex files and Orbitrap files from ThermoFisher. It is available on demand at marie-france.robbe@cea.fr, with a manual and example files.

Conclusions

To extend the panel of software tools able to process mass spectrometry imaging datasets, two user-friendly tools for visualization and data analysis with multiple processing functionalities were specially developed by the Computis European project. Data Cube Explorer is an easy visualization freeware for MALDI-ToF and ToF-SIMS images, with an image-classification tool. SpectViewer offers, in addition to visualization functions, an assistance to the interpretation of data, classification functionalities, image overlay, and peak list extraction to interrogate biological databases. SpectViewer is able to deal with original datasets (no binning) issued from MALDI-ToF, ToF-SIMS and DESI equipments. The capacities of

SpectViewer are complemented by EasyReg2D data fusion module for multimodal registration of images.

The well-known BioMap free platform was adapted to parse imzML files, thus enlarging the access to its multiple visualization and processing capacities, particularly dedicated to biologists, to a larger community of users.

Table 1 summarizes the main functionalities offered by the three visualization and processing software tools.

	BioMap	Data Cube Explorer	SpectViewer
Data	MALDI-ToF, DESI Binning	MALDI-ToF, ToF-SIMS Binning	MALDI-ToF, DESI, ToF-SIMS No binning or user- binning
Spectrum processing	Temporal filtering Baseline correction Detrending		Denoising Baseline correction
Spectrum display	1 spectrum 1D-zooming	1 spectrum 1D-zooming	Up to 6 spectra 2D-zooming
Image display	Up to 30 images Numerous color tables Intensity adjustment Zooming Geometrical transformations	1 image 1 grey table Intensity adjustment	Up to 9 images 3 color tables Intensity adjustment Zooming
Region of Interest	Several named ROIs Annotation of images Statistical analysis	3 ROIs	1 ROI
Assistance to find interesting peaks	Multi-image display Video		Relative variance Moran index Correlation matrix and spectrum
Clustering		Self-organizing map	K-means Stochastic K-means Hierarchical Random Projection Trees Diffusion map
Image registration	Registration with		Image overlay

	different models of 2 images (identical number of pixels)		Registration via EasyReg2D
Peak list			Parametrical peak list extraction

Table 1: Main functionalities of BioMap, Data Cube Explorer and SpectViewer

The joint efforts of the Computis project partners led to a major success: the development of software tools able to process imaging datasets issued from the majority of the mass spectrometry imaging instruments thanks to the definition of the common imzML format and the development of converters from equipment proprietary formats towards this standard format. In order to make easier the use of imzML format, the Computis project developed the vBrowser editor for imzML files to offer a comfortable lecture of imzML datasets with a tree-like display of imzML keywords.

Proteomics and mass spectrometry teams have now at their disposal a complete range of software tools to visualize and process mass spectrometry imaging data. Teams are no longer limited to the use of proprietary software to process their data; they can now choose the best suited software for their application and can process and compare images issued from different instruments with the same software.

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