Spatial and temporal country-wide survey of temephos resistance in Brazilian populations of *Aedes aegypti*

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The organophosphate temephos has been the main insecticide used against larvae of the dengue and yellow fever mosquito (Aedes aegypti) in Brazil since the mid-1980s. Reports of resistance date back to 1995; however, no systematic reports of widespread temephos resistance have occurred to date. As resistance investigation is paramount for strategic decision-making by health officials, our objective here was to investigate the spatial and temporal spread of temephos resistance in Ae. aegypti in Brazil for the last 12 years using discriminating temephos concentrations and the bioassay protocols of the World Health Organization. The mortality results obtained were subjected to spatial analysis for distance interpolation using semi-variance models to generate maps that depict the spread of temephos resistance in Brazil since 1999. The problem has been expanding. Since 2002-2003, approximately half the country has exhibited mosquito populations resistant to temephos. The frequency of temephos resistance and, likely, control failures, which start when the insecticide mortality level drops below 80%, has increased even further since 2004. Few parts of Brazil are able to achieve the target 80% efficacy threshold by 2010/2011, resulting in a significant risk of control failure by temephos in most of the country. The widespread resistance to temephos in Brazilian Ae. aegypti populations greatly compromise effective mosquito control efforts using this insecticide and indicates the urgent need to identify alternative insecticides aided by the preventive elimination of potential mosquito breeding sites.

Key words: insecticide resistance survey - dengue - distance interpolation - distribution maps - mosquito larvae

Vector-borne neglected (tropical) diseases such as dengue are an increasing worldwide issue of concern, particularly given current rates of urbanisation, international travel and trade, and climate change, all of which favor the spread of such diseases and their vectors (Hsieh & Chen 2009, Guzman et al. 2010, Gubler 2012). Mass gatherings and large sporting events are also associated with higher risks of health incidents. The 2014 FIFA World Cup held in Brazil is an example that drew attention and incited debate that focused particularly on dengue due to potential vector outbreaks (Hay 2013). The concern is understandable and justifiable, even if the

risks were generally small (Lowe et al. 2014, van Panhuis et al. 2014). As a result, no serious incident came to past. The 2016 Olympic Games to be held in Rio de Janeiro are bound to draw a similar level of international attention.

The lack of effective vaccines or pharmaceutical treatments for dengue, typical of the neglected diseases, places mosquito vector control in the forefront of prevention efforts for this disease (Gubler 2004, Halstead 2012). This scenario prevails throughout the affected tropical and subtropical regions of the world, which roughly encompasses about half of the global population (Guzman et al. 2010). Control of the dengue mosquito vector [Aedes aegypti (L.)], which also transmits chikungunya, zika and yellow fever (YF) (thus the common name "yellow fever mosquito"), relies heavily on insecticide use - but there are few compounds available and their use is usually guided by the countries' health officials (OPAS 1995, Funasa 2001, 2002, Braga & Valle 2007, Araújo et al. 2013, Macoris et al. 2014, Tomé et al. 2014).

The organophosphate temephos is globally the most commonly used insecticide against mosquito larvae due to its high efficacy, low cost and low vertebrate toxic-

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Received 23 October 2015 Accepted 17 March 2016 ity (WHO 2009). The result of this overreliance on temephos in controlling YF mosquito larvae is evolution and spread of temephos resistance among populations of this pest species. Such resistance has been detected in various countries since 1995 (Macoris et al. 1995, Mazzari & Georghiou 1995, Rawlins & Wan 1995, Bisset Lazcano et al. 2009, Melo-Santos et al. 2010, Bisset et al. 2013, Grisales et al. 2013). Furthermore, the use of temephos for the control of larvae of *Ae. aegypti* also apparently led to incidental selection for temephos resistance in co-occurring mosquito species populations (Campos & Andrade 2003, Alves et al. 2011, Phophiro et al. 2011, Amorim et al. 2013), as has also been reported among other co-occurring arthropod pest species (Guedes et al. 2016).

Routine applications of temephos against mosquito larvae in Brazil began in the 1980s (Funasa 1994, 2001, Sucen 1997). The initial suppression of Ae. aegypti in Brazil by 1955 was followed by its subsequent return in the 1970s (Schatzmayr 2000, Lourenço-de-Oliveira et al. 2004). Dengue became endemic in the country and has become an increasingly serious problem since 1986 despite established vector control programs in the country that still continue today (Lourenço-de-Oliveira et al. 2004, Maciel-de-Freitas et al. 2014). By the 1990s, concern emerged in Brazil regarding likely control failures and detection of temephos-resistant mosquito populations, which led to systematic surveys of insecticide resistance in the country and a series of reports on the phenomenon (Macoris et al. 1995, 2003, 2007, 2014, Campos & Andrade 2003, Lima et al. 2003, 2006, Melo-Santos et al. 2010, Gambarra et al. 2013, Diniz et al. 2014).

A few studies on the underlying mechanisms of temephos resistance followed the initial detection of this phenomenon in Brazil. Despite of an initial report of altered (acetylcholinesterase) target site sensitivity detected in a Brazilian population of Ae. aegypti resistant to temephos from Uberlândia (MG), current evidence suggests the prevalence of enhanced detoxification by metabolising enzymes in an apparently mixed pattern (Braga & Valle 2007, Melo-Santos et al. 2010, Lima et al. 2011, Gambarra et al. 2013). Congruent findings have been reported from other countries as well (Bisset Lazcano et al. 2009, Bisset et al. 2013, Grisales et al. 2013). Furthermore, recent transcriptome (i.e., the set of all mRNA molecules from a cell) evidence indicates upregulation of detoxification enzymes in insecticide-resistant mosquitoes (Reyes-Solis et al. 2014, Saavedra-Rodriguez et al. 2014). These findings reinforce the perception that multiple metabolic genes are involved in temephos resistance in Ae. aegypti, but with the prevalence of esterase rather than glutathione-S-transferase gene expression (Reyes-Solis et al. 2014, Saavedra-Rodriguez et al. 2014).

Temephos resistance monitoring in populations of the YF mosquito were underway in Brazil by the late 1990s in response to the increasing incidence of dengue in the country (Braga & Valle 2007). Scientific reports of the incidence of temephos resistance have increased since then (Campos & Andrade 2003, Lima et al. 2003, 2006, Macoris et al. 2003, 2007, Melo-Santos et al. 2010, Gambarra et al. 2013, Diniz et al. 2014), but no comprehensive dataset is currently available and no area-wide

description of the phenomenon of temephos resistance and its spread has been attempted despite the strategic importance of such information in guiding control policies, protocols and decision-making by Brazilian health officials. The current effort took advantage of the dataset gathered by the National Network of Insecticide Resistance Monitoring (MoReNAa) in *Ae. aegypti* under the tutelage of the National Program of Dengue Control from the Office of Health Surveillance of the Brazilian Ministry of Health (Brasília, DF, Brazil). The objective of our study was to recognise the spatial and temporal spread of temephos resistance in Brazil for the past 12 years, which we hypothesized, has been acute and has likely encompassed the entire country since 2010.

Our spatial and temporal survey of temephos resistance was performed using standardised procedures for insect sampling and temephos bioassays from the WHO (1981) that were countersigned by the laboratories involved (from MoReNAa) with the support of the Centers for Disease Control and Prevention (CCD, USA), Pan-American Health Organization and the World Health Organization (Braga et al. 2004, Macoris et al. 2005, Braga & Valle 2007). The data obtained was subjected to kriging to select suitable semivariogram models for distance interpolation with the goal of generating geospatial maps of the frequency of temephos resistance in Brazilian populations of *Ae. aegypti*.

MATERIALS AND METHODS

Insects and insecticide - Mosquito populations were sampled through the MoReNAa (Table I, Fig. 1) as described by Macoris et al. (2003). Briefly, between 100-200 oviposition traps (i.e., ovitraps) were used for this purpose in each city. The ovitraps were placed outdoors in a grid pattern for four weeks, always in the second semester of each year (Fay & Eliason 1966, Jakob & Bevier 1969, Funasa 1999). Egg clutches thus collected were used to establish laboratory colonies of over 3,000 individuals from each city (i.e., sampling site). First-generation larvae raised in the laboratory were used in the bioassays (Lima et al. 2003, Macoris et al. 2003). Technical grade temephos (> 90% pure) was obtained from the Brazilian Ministry of Health and diluted with acetone at the desired concentration for subsequent use in the diagnostic bioassays.

Diagnostic bioassays of temephos resistance - The diagnostic bioassays were performed following the standardised procedures of the WHO (1981, 1992). The concentration of temephos required to identify resistant insects (i.e., the diagnostic concentration) was initially established as 14.0 µg a.i./L but was subjected to yearly calibration and validation with the standard susceptible Rockefeller strain, as described by Braga et al. (2004) and Macoris et al. (2005). The diagnostic concentration was applied as a 1 mL solution to each of the experimental containers, reaching a final 250 mL volume of contaminated water solution (except for the controls, for which only 1 mL acetone was used). Deionised and distilled water were used to prepare the bioassay solutions. Twenty-five individuals (3rd-4th instar mosquito larvae) were placed in 250 mL transparent glass contain-

TABLE I

Sample site identification and geographical coordinates of collection sites for populations of the yellow fever mosquito *Aedes aegypti* used in the spatio-temporal survey of temephos resistance in Brazil

Region	State	City	Longitude	-11,438611	
North	Rondônia (RO)	Cacoal	-61,447222		
North	Rondônia (RO)	Guajará-Mirim	-65,339444	-10,782778	
North	Rondônia (RO)	Porto Velho	-63,903889	-8,761944	
North	Rondônia (RO)	Jaru	-62,466389	-10,438889	
North	Rondônia (RO)	Vilhena	-60,145833	-12,740556	
North	Acre (AC)	Rio Branco	-67,810000	-9,974722	
North	Amazonas (AM)	Manaus	-60,025000	-3,101944	
North	Roraima (RR)	Boa Vista	-60,673333	2,819722	
North	Pará (PA)	Ananindeua	-48,372222	-1,365556	
North	Pará (PA)	Belém	-48,504444	-1,455833	
North	Pará (PA)	Benevides	-48,244722	-1,361389	
North	Pará (PA)	Dom Elizeu	-47,505000	-4,285000	
North	Pará (PA)	Marabá	-49,117778	-5,368611	
North	Pará (PA)	Marituba	-48,341944	-1,355278	
North	Pará (PA)	Rondon do Pará	-48,067222	-4,776111	
North	Pará (PA)	Sta. Bárbara do Pará	-48,294444	-1,223611	
North	Pará (PA)	Santarém	-54,708333	-2,443056	
North	Pará (PA)	Tucuruí	-49,672500	-3,766111	
North	Amapá (AP)	Macapá	-51,066389	0,038889	
North	Tocantins (TO)	Araguaína	-48,207222	-7,191111	
North	Tocantins (TO)	Palmas	-48,360278	-10,212778	
Northeast	Maranhão (MA)	Bacabal	-44,791667	-4,291667	
Northeast	Maranhão (MA)	São Luís	-44,302778	-2,529722	
Northeast	Piauí (PI)	Parnaíba	-41,776667	-2,904722	
Northeast	Piauí (PI)	Teresina	-42,801944	-5,089167	
Northeast	Ceará (CE)	Caucaia	-38,653056	-3,736111	
Northeast	Ceará (CE)	Fortaleza	-38,543056	-3,717222	
Northeast	Ceará (CE)	Juazeiro do Norte	-39,315278	-7,213056	
Northeast	Rio Grande do Norte (RN)	Caicó	-37,097778	-6,458333	
Northeast	Rio Grande do Norte (RN)	Jardim do Seridó	-36,774444	-6,584444	
Northeast	Rio Grande do Norte (RN)	Parnamirim	-35,262778	-5,915556	
Northeast	Rio Grande do Norte (RN)	Mossoró	-37,344167	-5,187500	
Northeast	Rio Grande do Norte (RN)	Natal	-35,209444	-5,795000	
Northeast	Rio Grande do Norte (RN)	Pau dos Ferros	-38,204444	-6,109167	
Northeast	Paraíba (PB)	Alagoa Grande	-35,630000	-7,158333	
Northeast	Paraíba (PB)	Bayeux	-34,932222	-7,125000	
Northeast	Paraíba (PB)	João Pessoa	-34,863056	-7,115000	
Northeast	Paraíba (PB)	Santa Rita	-34,978056	-7,113889	
Northeast	Paraíba (PB)	Souza	-38,228056	-6,759167	
Northeast	Pernambuco (PE)	Araripina	-40,498333	-7,576111	
Northeast	Pernambuco (PE)	Cabo de Sto Agostinho	-35,035000	-8,286667	
Northeast	Pernambuco (PE)	Jaboatão dos Guararapes	-35,014722	-8,112778	
Northeast	Pernambuco (PE)	Moreno	-35,092222	-8,118611	
Northeast	Pernambuco (PE)	Olinda	-34,855278	-8,008889	
Northeast	Pernambuco (PE)	Petrolina	-40,500833	-9,398611	
Northeast	Pernambuco (PE)	Recife	-34,881111	-8,053889	
Northeast	Pernambuco (PE)	Tamandaré	-35,104722	-8,759722	
Northeast	Alagoas (AL)	Arapiraca	-36,661111	-8,739722 -9,752500	
Northeast	Alagoas (AL) Alagoas (AL)	Maceió	-35,735278	-9,732300 -9,665833	
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Region	State	City	Longitude	Latitude -10,908889	
Northeast	Sergipe (SE)	Barra dos Coqueiros	-37,038611		
Northeast	Sergipe (SE)	Itabaiana	-37,425278	-10,685000	
Northeast	Bahia (BA)	Barreiras	-44,990000	-12,152778	
Northeast	Bahia (BA)	Eunápolis	-39,580278	-16,377500	
Northeast	Bahia (BA)	Feira de Santana	-38,966667	-12,266667	
Northeast	Bahia (BA)	Ilhéus	-39,049444	-14,788889	
Northeast	Bahia (BA)	Itabuna	-39,280278	-14,785556	
Northeast	Bahia (BA)	Jacobina	-40,518333	-11,180556	
Northeast	Bahia (BA)	Jequié	-40,083611	-13,857500	
Northeast	Bahia (BA)	Potiguará	-39,876667	-15,594722	
Northeast	Bahia (BA)	Salvador	-38,510833	-12,971111	
Northeast	Bahia (BA)	Teixeira de Freitas	-39,741944	-17,535000	
Northeast	Bahia (BA)	Vitória da Conquista	-40,839444	-14,866111	
Midwest	Mato Grosso do Sul (MS)	Campo Grande	-54,646389	-20,442778	
Midwest	Mato Grosso do Sul (MS)	Corumbá	-57,653333	-19,009167	
Midwest	Mato Grosso do Sul (MS)	Coxim	-54,760000	-18,506667	
Midwest	Mato Grosso do Sul (MS)	Três Lagoas	-51,678333	-20,751111	
Midwest	Mato Grosso do Sul (MS)	Ponta Porã	-55,725556	-22,536111	
Midwest	Mato Grosso do Sul (MS)	Dourados	-54,805556	-22,221111	
Midwest	Mato Grosso (MT)	Cuiabá	-56,096667	-15,596111	
Midwest	Mato Grosso (MT)	Várzea Grande	-56,132500	-15,646667	
Midwest	Goiás (GO)	Aparecida de Goiânia	-49,243889	-16,823333	
Midwest	Goiás (GO)	Goiânia	-49,253889	-16,678611	
Midwest	Goiás (GO)	Itumbiara	-49,215278	-18,419167	
Midwest	Goiás (GO)	Luziânia	-47,950278	-16,252500	
Midwest	Goiás (GO)	Novo Gama	-48,039444	-16,059167	
Midwest	Goiás (GO)	Rio Verde	-50,928056	-17,798056	
Midwest	Goiás (GO)	Uruaçu	-49,140833	-14,524722	
Midwest	Distrito Federal (DF)	Brasília	-47,929722	-15,779722	
Southeast	Minas Gerais (MG)	Belo Horizonte	-43,937778	-19,920833	
Southeast	Minas Gerais (MG)	Formiga	-45,426389	-20,464444	
Southeast	Minas Gerais (MG)	Januária	-44,361667	-15,488056	
Southeast	Minas Gerais (MG)	Montes Claros	-43,861667	-16,735000	
Southeast	Minas Gerais (MG)	Teófilo Otoni	-41,505278	-17,857500	
Southeast	Minas Gerais (MG)	Ubá	-42,942778	-21,120000	
Southeast	Minas Gerais (MG)	Uberaba	-47,931944	-19,748333	
Southeast	Minas Gerais (MG)	Uberlândia	-48,277222	-18,918611	
Southeast	Espírito Santo (ES)	Cach. de Itapemirim	-41,112778	-20,848889	
Southeast	Espírito Santo (ES)	Cariacica	-40,420000	-20,263889	
Southeast	Espírito Santo (ES)	Colatina	-40,630556	-19,539444	
Southeast	Espírito Santo (ES)	Serra	-40,307778	-20,128611	
Southeast	Espírito Santo (ES)	Viana	-40,496111	-20,390278	
Southeast	Espírito Santo (ES)	Vila Velha	-40,292500	-20,329722	
Southeast	Espírito Santo (ES)	Vitória	-40,337778	-20,319444	
Southeast	Rio de Janeiro (RJ)	Cabo Frio	-42,018611	-22,879444	
Southeast	Rio de Janeiro (RJ)	C. dos Goytacazes	-41,324444	-21,754167	
Southeast	Rio de Janeiro (RJ)	Duque de Caxias	-43,311667	-22,785556	
Southeast	Rio de Janeiro (RJ)	Itaperuna	-41,887778	-21,205000	
Southeast	Rio de Janeiro (RJ)	Niterói	-43,103611	-22,883333	
Southeast	Rio de Janeiro (RJ)	Nova Iguaçu	-43,451111	-22,759167	
Southeast	Rio de Janeiro (RJ)	Rio de Janeiro	-43,207500	-22,902778	
Southeast	Rio de Janeiro (RJ)	São Gonçalo	-43,053889	-22,826944	
Southeast	Rio de Janeiro (RJ)	São João de Meriti	-43,372222	-22,803889	

Region	State	City	Longitude	-22,151389	
Southeast	Rio de Janeiro (RJ)	S. José do V. Rio Preto	-42,924444		
Southeast	Rio de Janeiro (RJ)	Três Rios	-43,209167	-22,116667	
Southeast	Rio de Janeiro (RJ)	Volta Redonda	-44,104167	-22,523056	
Southeast	São Paulo (SP)	Araçatuba	-50,432778	-21,208889	
Southeast	São Paulo (SP)	Barretos	-48,567778	-20,557222	
Southeast	São Paulo (SP)	Bauru	-49,060556	-22,314722	
Southeast	São Paulo (SP)	Botucatu	-48,445000	-22,885833	
Southeast	São Paulo (SP)	Campinas	-47,060833	-22,905556	
Southeast	São Paulo (SP)	Itapevi	-46,934167	-23,548889	
Southeast	São Paulo (SP)	Itu	-47,299167	-23,264167	
Southeast	São Paulo (SP)	Jandira	-46,902500	-23,527500	
Southeast	São Paulo (SP)	Marília	-49,945833	-22,213889	
Southeast	São Paulo (SP)	Presidente Prudente	-51,388889	-22,125556	
Southeast	São Paulo (SP)	Ribeirão Preto	-47,810278	-21,177500	
Southeast	São Paulo (SP)	Santana de Parnaíba	-46,917778	-23,444167	
Southeast	São Paulo (SP)	Santos	-46,333611	-23,960833	
Southeast	São Paulo (SP)	São Carlos	-47,890833	-22,017500	
Southeast	São Paulo (SP)	São José do Rio Preto	-49,379444	-20,819722	
Southeast	São Paulo (SP)	São Paulo (Pirituba)	-46,723611	-23,475000	
Southeast	São Paulo (SP)	São Paulo (Ipiranga)	-46,642222	-23,543889	
Southeast	São Paulo (SP)	São Sebastião	-45,409722	-23,760000	
Southeast	São Paulo (SP)	Sorocaba	-47,458056	-23,501667	
South	Paraná (PR)	Foz do Iguaçu	-54,588056	-25,547778	
South	Paraná (PR)	Londrina	-51,162778	-23,310278	
South	Paraná (PR)	Jacarezinho	-49,969444	-23,160556	
South	Paraná (PR)	Maringá	-51,938611	-23,425278	
South	Paraná (PR)	Palotina	-53,840000	-24,283889	
South	Rio Grande do Sul (RS)	Crissiumal	-54,101111	-27,499722	
South	Santa Catarina (SC)	Florianópolis	-48,549167	-27,596667	
South	Santa Catarina (SC)	Itapiranga	-53,712222	-27,169444	

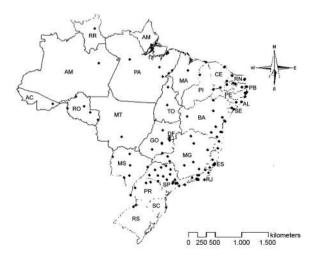


Fig. 1: distribution of the sampling sites of the populations of the yellow fever mosquito *Aedes aegypti* used in the spatio-temporal survey of temephos resistance in Brazil. Identification for each sampling site and its coordinates are listed in Table I.

ers containing temephos-contaminated water (except in the control treatments) and four replicates were used for each locally collected population. Mortality assessment of the mosquito larvae was performed after 24 h exposure. The larvae were considered dead if they were unable to rise to the surface when dorsally prodded.

Geostatistical analyses - These analyses were based on the geographical coordinates of each mosquito sampling site from which the mosquito populations were obtained and used to calculate the distance between sampling sites. The distances from the sampling sites and the mortality data obtained from the diagnostic bioassays were subjected to alternative kriging methods (stable, circular, spherical, exponential and Gaussian) to select suitable semivariogram functions for distance interpolation (Isaacs & Srivastava 1989). The semivariogram functions obtained using each group of models allowed the estimation of three parameters to determine their respective shapes: range (h_r) , partial sill (C), and nugget (C_p) . The range (h_r) and partial sill (C) refer to the point in the semivariogram function in which a plateau

is reached; the range (h) corresponds to the distance at which this phenomenon takes place, while the partial sill (C) refers to its respective semivariance value. The nugget (C₂) is the semivariogram value in which the model intercepts the y-axis (i.e., the mortality semivariance axis) corresponding to measurement errors or spatial sources of variation at distances smaller than the sampling interval (or both). Three additional parameters were calculated from these three basic parameters described above. These were: sill (C₀ + C), proportion [C/ (C_o + C)] and randomness (C_o/C) of the data. A crossvalidation procedure was subsequently used to select the best data adjustment to compare the observed and estimated data for each sampling point using the model of semivariogram function under test. This estimated error allows the best model selection as those leading to the error average closer to zero, aided by the randomness assessment (the higher, the better). The semivariance data obtained from the selected models were used to generate the spatial maps depicting the phenomenon of temephos resistance. All the spatial analyses were performed using ArcGIS 10 software (ESRI, Redlands, CA, USA).

RESULTS

General temephos mortality findings - The diagnostic bioassays assessing mosquito larvae mortality by temephos were performed to estimate the frequency of temephos-resistant individuals in the sampled insect populations. This frequency of resistant individuals is indicated as an average mortality score ranging from 80.31% between 1999-2000 and dropping to less than 50% between 2010-2011 (Table II). The number of insect samples tested per year ranged from 25 (from 2010-2011) to 74 (between 2000-2001) and had a broad range of mortality response within each year, resulting in a high standard deviation of larval mortality per year (Table III).

Semivariogram model selection - Suitable semivariogram models were obtained for each biannual dataset of temephos mortality using the diagnostic insecticide resistance bioassays. The selected semivariogram models are exhibited in Table III, along with their respective parameters for model selection. The plots from each model and the respective observed data are exhibited in Fig. 2.

TABLE II

Descriptive statistics of the diagnostic bioassays with temephos on larvae of the yellow fever mosquito *Aedes aegypti*

Year	G 1: '		Mor	C1	**		
	Sampling sites - (n)	Minimum	Maximum	Mean	SD	Skewness (g_i)	Kurtosis (g_2)
1999-2000	64	13.15	100.00	80.31	24.62	-1.22	3.40
2000-2001	74	10.80	100.00	71.53	26.34	-0.68	2.38
2002-2003	58	2.00	99.80	62.48	30.16	-0.51	2.08
2004-2005	59	1.50	98.45	53.41	33.69	-0.18	1.39
2006-2007	39	6.40	97.60	52.33	24.48	-0.16	1.97
2008-2009	46	6.00	96.70	50.60	24.99	0.05	1.82
2010-2011	25	7.50	88.20	49.99	28.16	-0.12	1.55

SD: standard deviation

TABLE III

Semivariogram models and parameters of larval mortality by temephos on populations of the yellow fever mosquito *Aedes aegypti*

Year	Kriging	Model	Nugget (C_0)	Partial sill (C)	$Sill (C_0 + C)$	Proportion (C/C+C ₀)	Range (h_r, m)	Randomness (C_0/C)	Mean errors
1999-2000	Ordinary	Gaussian	132.963	639.079	772.042	0.827778	593820.368	0.208054	-0.027
2000-2001	Simple	Gaussian	231.740	640.182	871.922	0.734219	632424.376	0.361991	-0.059
2002-2003	Simple	Exponential	391.601	972.709	1364.31	0.712968	3658678.194	0.402588	-0.203
2004-2005	Ordinary	Gaussian	224.524	176.033	400.557	0.439471	695175.201	1.275465	0.101
2006-2007	Ordinary	Exponential	162.384	669.389	831.773	0.804774	1175553.465	0.242585	-0.096
2008-2009	Ordinary	Circular	57.218	723.989	781.207	0.926757	947927.124	0.079032	0.266
2010-2011	Ordinary	Circular	367.832	262.731	630.563	0.416661	507101.080	0.714269	1.576

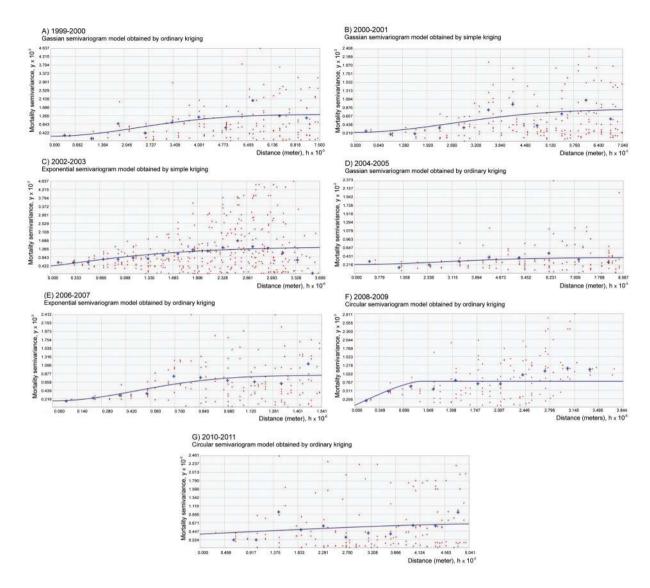


Fig. 2: semivariogram models [mortality semivariance (y) as a function of distance (x)] exhibited in Table II and obtained from the diagnostic bioassays of temephos resistance on larvae of the yellow fever mosquito *Aedes aegypti*. Observed points are represented as red symbols, and averages are represented as blue crosses.

Temporal spread of temephos resistance - Spatial interpolation using kriging allowed mapping the countrywide spread of temephos resistance in larvae of YF mosquitoes from 1999-2000 until 2010-2011, which is the last year the survey data were available. Initially the efficacy of temephos was high, causing larval mortality of > 80% throughout Brazil, except in the coastal area, which spans from Pará in the north to Piauí in the northeast and encompasses the state of Rio de Janeiro and neighboring parts of São Paulo and Minas Gerais (Fig. 3). However, the frequency of temephos resistant individuals in the insect populations increased steadily during each biannual survey, reflecting a significant reduction in temephos efficacy. This trend reached high levels (< 50% mortality) in about half the country as early as 2004-2005 (Fig. 3). Although the frequency

of temephos resistance seems to have been attenuated in the main problem areas observed between 2004-2005, temephos resistance continued to spread within Brazil. By 2010-2011 only Rondônia (in the North), São Paulo (Southeast), Paraná and Santa Catarina (South) exhibited satisfactory temephos efficacy against YF mosquito larvae. New focal areas of temephos resistance were detected in the 2010-2011 survey radiating from near Rio Branco (southern Acre in North Brazil, near Bolivia) and Brasilia (Central Brazil), leading to a country-wide resistance phenomenon.

DISCUSSION

The temephos mortality dataset obtained from the diagnostic bioassays performed by the MoReNAa, although not carried out with the objective of spatial in-

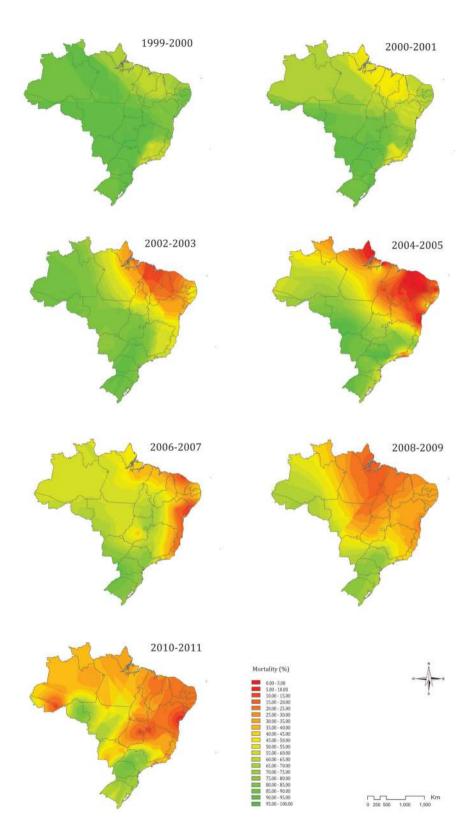


Fig. 3: contour maps of temephos resistance in Brazilian populations of the yellow fever mosquito (*Aedes aegypti*) generated using spatial interpolation. The colour legend indicates the represented range of mortality (%) of mosquito larvae obtained in the temephos resistance diagnostic bioassays. Colours tending toward red indicate lower larval mortality and, consequently, a higher frequency of temephos resistance.

terpolation to generate temephos resistance maps for Brazilian populations of *Ae. aegypti*, allowed such interpolations and the inferences necessary to generate the maps. The effort provided a means to clearly illustrate the temporal spread and spatial reach of temephos resistance in *Ae. aegypti* - which greatly increased during the 12-year period of assessment - within the Brazilian territory. Nonetheless, a more fine-tuned survey focusing on diagnostic bioassays of insecticide resistance using larger and better-distributed sampling sites would allow even more comprehensive assessments for eventual decision-making regarding policies and procedures to be adopted.

Temephos resistance among Brazilian populations of the YF mosquito is far from novel. All the Brazilian states have adopted the routine use of temephos (1% sand granule formulations) to manage Ae. aegypti by controlling its larvae since the early 1990's (Funasa 1994, 2001, Sucen 1997). The result of this continuous and consistent use of temephos throughout the country led to reports of temephos resistance as early as 1995 (Macoris et al. 1995). The increased incidence of dengue during the 1990s in Brazil attributed to the spread of Ae. aegypti enhanced concern regarding insecticide use against the mosquito and the susceptibility of mosquito populations (da Silva Jr et al. 2002, Braga & Valle 2007). The end result was the establishment of an insecticide-resistance monitoring program in the country that focused on populations of the YF mosquito (Braga & Valle 2007). Consistent detection of temephos resistance in different parts of the country soon followed (Campos & Andrade 2003, Lima et al. 2003, 2006, Macoris et al. 2003, 2007, Melo-Santos et al. 2010, Gambarra et al. 2013, Diniz et al. 2014).

Some of the studies of temephos resistance among Brazilian populations of the YF mosquito explored the mechanisms involved and the existence of fitness costs associated with this resistance. Fitness costs were indeed detected (Diniz et al. 2014). Unfortunately, the studies on the underlying mechanisms of temephos resistance were more confused and patchy, but they were suggestive of the prevailing involvement of enhanced insecticide detoxification as the main mechanism, with esterases likely playing a major role, although not an exclusive one (Braga & Valle 2007, Melo-Santos et al. 2010, Gambarra et al. 2013, Macoris et al. 2014). These findings seem consistent with mechanistic studies of temephos resistance performed with other Latin American populations of the same species (Bisset Lazcano et al. 2009, Bisset et al. 2013, Grisales et al. 2013, Reyes-Solis et al. 2014, Saavedra-Rodriguez et al. 2014).

The twelve-year effort of the MoReNAa achieved a great deal, but no summary of the country-wide survey effort had ever been performed; therefore, creating such a summary was the objective of the current work. The geostatistical tools used here allowed the recognition of both the temporal pattern of the spread of temephos resistance in the country and its gravity by 2011. Despite the early detection of temephos resistance in the mid-1990s, country-wide use of temephos continued; consequently, temephos resistance spread throughout the country during the following years, reaching serious levels by

2002-2003. At this point, nearly half of the country was already having problems because of temephos resistance in *Ae. aegypti*, particularly when considering "resistant" to mean mosquito populations that exhibit mortality levels below the 80% threshold-a threshold that incurs in a high likelihood of control failure (Davidson & Zahar 1973). The scenario has simply gotten worse in subsequent years. Now, nearly all the country (except a part in the South) exhibits temephos resistance.

The use of temephos as a mosquito larvicide in Brazil has been suppressed since late in 2010, which may reverse the spread of resistance and allow for future use of the compound. However, the high frequency of resistant individuals already established in the country potentially limits the extent of such (future) use, even if the fitness cost associated with temephos resistance prevails in the country. Effective, safe and cheap insecticides such as temephos, which are the underlying reasons for its global use as a mosquito larvicide, are hard to come by (Tomé et al. 2014). A few alternatives have emerged and are currently being explored, including a few pyrethroids, but these already exhibit insecticide resistance problems in wide areas in Brazil (e.g., Brito et al. 2013). More recently, insect growth regulators and the bioinsecticide Bacillus thuringiensis serovar israelensis (Bti) have been explored (Braga & Valle 2007, Fontoura et al. 2012, Araújo et al. 2013).

In conclusion, temephos resistance in Brazilian populations of the YF mosquito spread during the 12-year survey period, showing that resistance is now widespread and there is little hope of achieving effective mosquito control with this insecticide. Alternative insecticides aided by the preventive elimination of potential mosquito breeding sites are necessary. However, the use of these alternative insecticides will also lead to the eventual emergence of resistant mosquito populations - and may have already occurred in the country, considering their present rate of use. Therefore, continuing country-wide surveys are necessary to guide management decisions by the national health officers. In such a context, planned yearly systematic sampling and insecticide resistance diagnostic bioassays are necessary. Moreover, geostatistical analyses to map the levels and spread of the phenomenon are also necessary.

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REFERENCES

Alves SN, Tibúrcio JD, de Melo AL. Susceptibilidade de larvas de Culex quinquefasciatus a diferentes inseticidas. Rev Soc Bras Med Trop. 2011; 44(4): 486-489.

Amorim LB, Helvecio E, de Oliveira CMF, Ayres CFJ. Susceptibility status of *Culex quinquefasciatus* (Diptera: Culicidae) populations to the chemical insecticide temephos in Pernambuco, Brazil. Pest Manag Sci. 2013; 69(12): 1307-1314.

Araújo AP, Araújo DFD, Helvecio E, Barros RA, Oliveira CMF, Ayres CFJ, et al. The susceptibility of *Aedes aegypti* populations displaing temephos resistance to *Bacillus thuringiensis israelensis*: a basis for management. Parasit Vectors. 2013; 6: 297.

- Bisset JA, Marín R, Rodríguez MM, Severson DW, Ricardo Y, French L, et al. Insecticide resistance in two *Aedes aegypti* (Diptera: Culicidae) strains from Costa Rica. J Med Entomol. 2013; 50(2): 352-361.
- Bisset Lazcano JA, Rodríguez MM, San Martín JL, Romero JE, Montoya R. Evaluación de la resistencia a insecticidas de una cepa de Aedes aegypti de El Salvador. Rev Panam Salud Publica. 2009; 26(3): 229-234.
- Braga IA, Lima JBP, Soares SS, Valle D. Aedes aegypti resistance to temephos during 2001 in several municipalities in the states of Rio de Janeiro, Sergipe, and Alagoas, Brazil. Mem Inst Oswaldo Cruz. 2004; 99(2): 199-203.
- Braga IA, Valle D. Aedes aegypti: vigilância, monitoramento da resistência e alternativas de controle no Brasil. Epidemiol Serv Saude. 2007; 16(4): 295-302.
- Brito LP, Linss JGB, Lima-Camara TN, Belinato TA, Peixoto AA, Lima JBP, et al. Assessing the effects of *Aedes aegypti* kdr mutations on pyrethroid resistance and its fitness cost. PLoS ONE. 2013; 8(4): e60878.
- Campos J, Andrade CFS. Susceptibilidade larval de populações de Aedes aegypti e Culex quinquefasciatus a inseticidas químicos. Rev Saude Publica. 2003; 37(4): 523-527.
- da Silva Jr J, Siqueira Jr JB, Coelho GE, Vilarinhos PT, Pimenta Jr FG. Dengue in Brazil: current situation and control activities. Epidemiol Bull. 2002; 23(1): 3-6.
- Davidson G, Zahar AR. The practical implications of resistance in malaria vectors to insecticides. Bull World Health Organization. 1973; 49(5): 475-483.
- Diniz MMCSL, Henriques ADS, Leandro RS, Aguiar DL, Beserra EB. Resistance of *Aedes aegypti* to temephos and adaptive disadvantages. Rev Saude Publica. 2014; 48(5): 775-782.
- Fay RW, Eliason DA. A preferred oviposition site as a surveillance method for *Aedes aegypti*. Mosq News. 1966; 26(4): 531-534.
- Fontoura NG, Bellinato DF, Valle D, Lima JBP. The efficacy of a chitin synthesis inhibitor against field populations of organophosphate-resistant *Aedes aegypti* in Brazil. Mem Inst Oswaldo Cruz. 2012; 107(3): 387-395.
- Funasa Fundação Nacional de Saúde. Dengue Instruções para pessoal de combate ao vetor: Manual de Normas Técnicas, 3rd ed. Brasília: Ministério da Saúde; 2001.
- Funasa Fundação Nacional de Saúde. Manual de normas técnicas - Instruções para pessoal de operações. Brasília: Ministério da Saúde; 1994.
- Funasa Fundação Nacional de Saúde. Programa nacional de controle da dengue. Brasília: Ministério da Saúde; 2002.
- Funasa Fundação Nacional de Saúde. Reunião técnica para discutir status de resistência de *Aedes aegypti* e definir estratégias a serem implantadas para monitoramento da resistência no Brasil. Brasília: Ministério da Saúde; 1999.
- Gambarra WPT, Martins WFS, de Lucena Filho ML, de Albuquerque IMC, Apolinário OKS, Beserra EB. Spatial distribution and esterase activity in populations of *Aedes (Stegomyia) aegypti* (Linnaeus) (Diptera: Culicidae) resistant to temephos. Rev Soc Bras Med Trop. 2013; 46(2): 178-184.
- Grisales N, Poupardin R, Gomez S, Fonseca-González I, Ranson H, Lenhart A. Temephos resistance in *Aedes aegypti* in Colombia compromises dengue vector control. PLoS Negl Trop Dis. 2013; 7(9): e2438.
- Gubler DJ. The changing epidemiology of yellow fever and dengue, 1900 to 2003: full circle? Comp Immunol Microbiol Infect Dis. 2004; 27(5): 319-330.
- Gubler DJ. The economic burden of dengue. Am J Trop Med Hyg. 2012; 86(5): 743-744.

- Guedes RNC, Smagghe G, Stark JD, Desneux N. Pesticide-induced stress in arthropod pests for optimized integrated pest management programs. Annu Rev Entomol. 2016; 61: 43-62.
- Guzman MG, Halstead SB, Artsob H, Buchy P, Farrar J, Gubler DJ, et al. Dengue: a continuing global threat. Nat Rev Microbiol. 2010; 8(Suppl. 12): S7-S18.
- Halstead SB. Dengue vaccine development: a 75% solution? Lancet. 2012; 380(9853): 1535-1536.
- Hay S. Football fever could be a dose of dengue. Nature. 2013; 503(7477): 439.
- Hsieh YH, Chen CWS. Turning points, reproduction number, and impact of climatological events for multi-waver dengue outbreaks. Trop Med Int Health. 2009: 14(6): 628-638.
- Isaacs EH, Srivastava RM. An introduction to applied geostatistics. New York: Oxford University Press; 1989.
- Jakob WL, Bevier GA. Application of ovitraps in the US Aedes aegypti eradication program. Mosq News. 1969; 29(1): 55-61.
- Lima EP, de Oliveira Filho AM, Lima JWO, Ramos Jr NA, Cavalcanti LPG, Pontes RJS. Resistência do Aedes aegypti ao temefós em municípios do estado do Ceará. Rev Soc Bras Med Trop. 2006; 39: 259-263.
- Lima EP, Paiva MH, de Araújo AP, da Silva EV, da Silva UM, de Oliveira IN, et al. Insecticide resistance in *Aedes aegypti* populations from Ceará, Brazil. Parasit Vectors. 2011; 4: 5.
- Lima JBP, da Cunha MP, da Silva RC, Galardo AKR, Soares SS, Braga IA, et al. Resistance of *Aedes aegypti* to organophosphates in several municipalities in the states of Rio de Janeiro and Espírito Santo, Brazil. Am J Trop Med Hyg. 2003; 68(3): 329-333.
- Lourenço-de-Oliveira R, Vazeille M, de Filippis AMB, Failloux AB. *Aedes aegypti* in Brazil: genetically-differentiated populations with high susceptibility to dengue and yellow fever viruses. Trans R Soc Trop Med Hyg. 2004; 98(1): 43-54.
- Lowe R, Barcellos C, Coelho CAS, Bailey T, Coelho GE, Graham R, et al. Dengue outlook for the World Cup in Brazil: an early warning model framework driven by real-time seasonal climate forecasts. Lancet Infect Dis. 2014; 14(7): 619-626.
- Maciel-de-Freitas R, Avendanho FC, Santos R, Sylvestre G, Araújo SC, Lima JBP, et al. Undesirable consequences of insecticide resistance following *Aedes aegypti* control activities due to a dengue outbreak. PLoS ONE. 2014; 9(3): e92424.
- Macoris MLG, Andrighetti MTM, Nalon KCR, Garbeloto VC, Caldas Jr AL. Standardization of bioassays for monitoring resistance to insecticides in *Aedes aegypti*. Dengue Bull. 2005; 29: 176-182.
- Macoris MLG, Andrighetti MTM, Otrera VCG, de Carvalho LR, Caldas Jr AL, Brogdon WG. Association of insecticide use and alteration on *Aedes aegypti* susceptibility status. Mem Inst Oswaldo Cruz. 2007; 102(8): 895-900.
- Macoris MLG, Andrighetti MTM, Takaku L, Glasser CM, Garbeloto VC, Bracco JE. Resistance of *Aedes aegypti* from the state of *São Paulo*, Brazil, to organophosphate insecticides. Mem Inst Oswaldo Cruz. 2003; 98(5): 703-708.
- Macoris MLG, Andrighetti MTM, Wanderley DMV, Ribolla PEM. Impact of insecticide resistance on the field control of *Aedes aegypti* in the state of São Paulo. Rev Soc Bras Med Trop. 2014; 47(5): 573-578.
- Macoris MLG, Camargo MF, Silva IG, Takaku L, Andrighetti MTM. Modificação da susceptibilidade de *Aedes (Stegomyia) aegypti* ao temefós. Rev Pat Trop. 1995; 24(1): 31-40.
- Mazzari MB, Georghiou GP. Characterization of resistance to organophosphate, carbamate, and pyrethroid insecticides in field

- populations of *Aedes aegypti* from Venezuela. J Am Mosq Control Assoc. 1995; 11: 315-322.
- Melo-Santos MAV, Varjal-Melo JJM, Araújo AP, Gomes TCS, Paiva MHS, Regis LN, et al. Resistance to the organophosphate temephos: mechanisms, evolution, and reversion in na *Aedes aegypti* laboratory strain from Brazil. Acta Trop. 2010; 113: 180-189.
- OPAS Organización Panamericana de la Salud. Dengue y dengue hemorrágico en las Américas: guías para su prevención y control. Washington DC: OPS; 1995.
- Phophiro JS, Silva OS, Luna JED, Piccoli CF, Kanis LA, da Silva MAN. *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae): coexistence and susceptibility to temephos, in municipalities with occurrence of dengue and differentiated characteristics of urbanization. Rev Soc Bras Med Trop. 2011; 44(3): 300-305.
- Rawlins SC, Wan JO. Resistance in some Caribbean populations of *Aedes aegypti* to several insecticides. J Am Mosq Control Assoc. 1995; 11(1): 59-65.
- Reyes-Solis GC, Saavedra-Rodriguez K, Suarez AF, Black 4th WC. QTL mapping of genome regions controlling temephos resistance in larvae of the mosquito *Aedes aegypti*. PLoS Negl Trop Dis. 2014; 8(10): e3177.
- Saavedra-Rodriguez K, Strode C, Flores AE, Garcia-Luna S, Reyes-Solis G, Ranson H, et al. Differential transcription profiles in Ae-

- des aegypti detoxification genes after temephos selection. Insect Mol Biol. 2014; 23: 199-215.
- Schatzmayr HG. Dengue situation in Brazil by year 2000. Mem Inst Oswaldo Cruz. 2000; 95(Suppl. 1): S179-S181.
- Sucen Superintendência de Controle de Endemias. Plano de erradicação de *Aedes aegypti*: Guia de instruções. São Paulo: Secretaria de Estado da Saúde de São Paulo; 1997.
- Tomé HVV, Pascini TV, Dângelo RAC, Guedes RNC, Martins GF. Survival and swimming behavior of insecticide-exposed larvae and pupae of the yellow fever mosquito *Aedes aegypti*. Parasit Vectors. 2014; 7: 195.
- van Panhuis WG, Hyun S, Blaney K, Marques Jr ETA, Coelho GE, Siqueira Jr JB, et al. Risk of dengue for tourists and teams during the World Cup 2014 in Brazil. PLoS Negl Trop Dis. 2014; 8(7): e3065.
- WHO World Health Organization. Expert committee on vector biology and control. Vector resistance to pesticides: WHO Technical Report Series 818. Geneva: WHO; 1992.
- WHO World Health Organization. Global insecticide use for vector-borne disease control. 4th ed. Geneva: WHO/HTM/NTD/WHOPES/GCDPP; 2009.
- WHO World Health Organization. Instruction for determining the susceptibility or resistance of mosquito larvae to insecticides. Geneva: WHO/VBC; 1981.