

**Article title:****Spatial Cognition in Mice and Rats: Similarities and Differences in Brain and Behaviour****Article type:**

OPINIONS provide a forum for thought-leaders, hand-picked by the editors, to provide a more individual perspective on the field in question. (Average extent = 2,000-4,000 words, < 5 figures/tables, ~5 pages)

PRIMERS are meant to be understood by a very general audience, such as undergraduates just entering a given field. (Average extent = 3,000 words, 3 figures/tables, ~4 pages)

OVERVIEWS provide a broad and relatively non-technical treatment of important topics at a level suitable for advanced students and for researchers without a strong background in the field. (Average extent = 4,000-8,000 words, 10-16 figures/tables, 10-14 pages)

ADVANCED REVIEWS, aimed at researchers and advanced students with a strong background in the subject, review key areas of research in a citation-rich format similar to that of leading review journals. (Average extent = 4,000-6,000 words, < 10 figures/tables, ~10 pages)

FOCUS ARTICLES describe specific real-world issues, examples, implementations, etc. These articles are usually technical in nature. (Average extent = 3,000-4,000 words, < 7 figures/tables, ~5 pages)

SOFTWARE FOCUS articles describe specific software packages of high utility in a certain field, with an emphasis on their capabilities and implementations rather than methodology. (Average extent = 3,000 words, 3 figures/tables, ~4 pages)

Authors:

Full name and affiliation; email address if corresponding author; any conflicts of interest

First author

Vincent Hok*, (1) Laboratory of Cognitive Neuroscience, CNRS and Aix-Marseille University, Marseille, France, (2) Fédération 3C, CNRS and Aix-Marseille University, Marseille, France. vincent.hok@univ-amu.fr

Second author

Bruno Poucet*, (1) Laboratory of Cognitive Neuroscience, CNRS and Aix-Marseille University, Marseille, France, (2) Fédération 3C, CNRS and Aix-Marseille University, Marseille, France. bruno.poucet@univ-amu.fr

Third author

Éléonore Duvelle, (4) UCL Psychology and Language Sciences, Faculty of Brain Sciences, London, United Kingdom.

Fourth author

Étienne Save, (1) Laboratory of Cognitive Neuroscience, CNRS and Aix-Marseille University, Marseille, France, (2) Fédération 3C, CNRS and Aix-Marseille University, Marseille, France.

Last author

Francesca Sargolini, (1) Laboratory of Cognitive Neuroscience, CNRS and Aix-Marseille University, Marseille, France, (2) Fédération 3C, CNRS and Aix-Marseille University, Marseille, France, (3) Institut Universitaire de France, Paris, France.

Keywords

Spatial cognition, Hippocampus, Behaviour, Place cells, Mice, Rats.

Abstract

The increasing use of mice models in cognitive tasks that were originally designed for rats raises crucial questions about cross-species comparison in the study of spatial cognition. The present review focuses on the major neuroethological differences existing between mice and rats with a particular attention given to the neurophysiological basis of space coding. Whereas little difference is found in the basic properties of space representation in these two species, it appears that the stability of this representation changes more drastically over time in mice than in rats. We consider several hypotheses dealing with attentional, perceptual and genetic aspects and offer some directions for future research that might help in deciphering hippocampal function in learning and memory processes.

Introduction

As pointed out by Hans J. Hedrich¹, the Norway rat (*Rattus norvegicus*) was the first mammalian species to be domesticated for scientific purpose as early as the first half of the 19th century. With the advent of molecular techniques in the late '80s and the development of transgenic mouse models, mice account nowadays for three-quarters of the mammals used in biomedical research². For this reason, the mouse was the second mammal to have its genome fully sequenced, right after the human genome³.

Historically, the rat has been most commonly used by physiologists, with a special attention given to learning and memory^{4,5}, whereas the mouse became the model of choice for genetic studies. By allowing manipulation of specific genes thought to be involved in cognitive processes, the knockout approach increased drastically the use of mice in behavioural research⁶⁻⁸. This has led to the confounding situation in which mice have been used extensively in behavioural paradigms originally

1
2
3 designed for rats, with virtually no consideration of the differences between these two species, as if
4 they were fully interchangeable (see on this specific matter Refs ^{4,9–11}).

5
6 Rodents' abilities in spatial navigation tasks have been widely investigated for decades. This has led
7 to the production of a vast amount of experimental data on brain and behaviour available in both
8 species. In order to **evaluate putative cognitive differences between these two species, we will first**
9 **present the primary concepts in spatial navigation and discuss the nature of the spatial**
10 **representation in the rodent brain. We then examine throughout the last sections the interactions**
11 **between behaviour and brain activity that might explain interspecies differences when tested in**
12 **common spatial tasks.**

13 14 15 16 **SPACE PERCEPTION**

17
18 As early as the beginning of the 20th century, scientists began to investigate in details the
19 mechanisms supporting the ability of animals to find their way back to their nest (*e.g.* Ref ¹²). It
20 appeared quickly that the selection of appropriate navigational strategies was primarily determined
21 by the perception of space, that is, by the nature of the cues that could be used for navigation^a.

22 23 24 **Cues for navigation**

25
26 Cues useful for navigation are of two sorts: External cues provided by the environment (*allothetic*
27 *cues*) and self-motion related cues (*idiothetic cues*). Allothetic cues encompass visual, tactile,
28 auditory and olfactory signals whereas idiothetic cues are provided mainly by vestibular and
29 proprioceptive inputs^{14,15}. Note that a given sensory modality organ can provide both types of
30 information: for instance, vision can convey allothetic information about static environmental
31 landmark as well as idiothetic cues through the optic flow generated during self-motion.

32
33
34 In laboratory conditions, allothetic cues can be easily manipulated in order to trigger changes in
35 behaviour and brain activity in freely-moving animals (*e.g.* Refs ^{16,17}). Conversely, manipulation of
36 idiothetic cues cannot be achieved without partially restraining the animals (*e.g.* head fixed
37 preparation¹⁸). These cues are otherwise always available (even in complete darkness), and are
38 sometimes sufficient for an animal to estimate distance and orientation parameters. For instance,
39 Wallace and Whishaw¹⁹ recorded trajectories from rats moving around a circular table-top in
40 either light or complete darkness conditions. Although their speed was lower in the dark, rats
41 managed to head to their departure point with the same precision in both conditions. In addition,
42 rats demonstrated knowledge of the distance to the goal, as their speed significantly decreased at
43 the midpoint of the homeward trip, regardless of the length of the trip^{19,20}. In this case, both
44 direction and distance controlled the trajectory, independently from the availability of allothetic
45 information. The ability of animals to keep track of their position with respect to a departure point
46 is termed path integration and can prove, to some extent, to be sufficient for an animal to achieve
47 accurate navigation^{19,21}. Indeed, the principal limitation of this navigation strategy comes from the
48 iterative nature of the process leading to accumulation of errors with increasing distance
49 traveled^{22,23}.

50
51
52
53
54
55
56 ^a Following Gallistel¹³, navigation is defined as "the process of determining and maintaining a course or
57 trajectory from one place to another."
58
59
60

1
2
3 Although rodents are able to use idiothetic signals to navigate, they usually rely heavily on allothetic
4 cues when available. In general, the visual modality is the most used. Olfactory and tactile signals
5 can also help localisation, particularly when visual cues are less salient (*e.g.* olfactory-based
6 navigation in the dark²⁴; cooperation of olfactory and vision²⁵; auditory cues²⁶). However, landmarks
7 can be unstable and allothetic cues may sometimes not be sufficient to disambiguate two similar
8 environments (such a situation is probably more likely to happen in laboratory conditions). In natural
9 conditions, animals combine allothetic and idiothetic signals to navigate, depending on their
10 reliability. We generally refer to this process as multisensory integration^{15,18}.
11
12

13 **Multisensory integration**

14
15 To assess the relative contribution of each type of information to self-localisation, a common
16 paradigm consists in causing a conflict between different sensory sources. For instance, in the
17 experiment by Etienne and collaborators²⁷, hamsters first learned to go from their nest to a feeder
18 located in the middle of a 220 cm diameter circular arena by following a baited spoon directed by
19 the experimenter. Once there, the hamsters filled in their cheek pouches with food and came back
20 to the nest. During training, a light spot was presented at the opposite side of the nest. During the
21 test, this visual cue was rotated by either 90° or 180°, thus creating a conflict between visual and
22 idiothetic cues. If hamsters relied exclusively on idiothetic cues, they would directly return to their
23 nest. If they relied on the visual cue, they would aim at the opposite direction of the spotlight. The
24 authors found that animals did neither one nor the other, but chose a position that was
25 intermediary between the one indicated by self-motion cues and the one deduced from the visual
26 cue. Interestingly, the deviation from the actual nest position depended upon the degree of conflict
27 between self-motion cues and the visual cue. When the spotlight was rotated 90° (small conflict),
28 the final position was far away from the nest position, as if the animals used preferentially the visual
29 cue over idiothetic cues. On the contrary, when the spotlight was rotated 180° (large conflict), the
30 final position was closer to the nest, thus suggesting that the idiothetic cues were given a larger
31 weight than the visual cue. Overall, these results suggest that navigation relies on a weighted
32 multisensory integration process²⁸. In this context, the contribution of each sensory modality
33 depends on the degree of confidence that can be attributed to them. There are several other
34 examples suggesting that mammals navigate by combining allothetic and idiothetic cues^{15,22,26,29}. In
35 certain conditions, rats can show a hierarchy in the use made of different sensory modalities to
36 guide navigation, vision being predominant over olfactory or self-motion cues³⁰. However, the
37 relative importance given to certain sensory sources over others greatly depends on their reliability
38 within a reference frame^{31,32}.
39
40
41
42
43
44
45

46 To summarize, rodents can navigate using a combination of allothetic and idiothetic information, in
47 a flexible and opportunistic manner, allowing switches between various strategies continuously in
48 the course of navigation.
49
50
51
52
53
54
55
56
57
58
59
60

NAVIGATION STRATEGIES

"No wild animal roams at random over the country; each has a home-region, even if it has not an actual home."³³ *Home range*^b concept (or *home-region* as previously defined by Ernest T. Seton) has been widely used in order to define the "area traversed by the individual in its normal activities of food gathering, mating, and caring for young."³⁴ In the wild, mice and rats show territorial behaviour like many other mammals^{35,36}, and show *home base* behaviour in laboratory settings³⁷. In this context, home base refers to the location in which the animal spends a disproportionate period of time and from which it performs excursions³⁸. During this home base behaviour, it appears that exploratory behaviour is organized³⁹ and that specific locomotor patterns can be identified⁴⁰.

Exploring space

Most mammalian species show increased exploratory activity when confronted with novelty and rodents are no exception^{41,42}. This behaviour consists in moving towards unknown places or objects and gathering different types of information from several sensory modalities. Exploratory behaviour diminishes with habituation, albeit in different ways across species⁴³ and strains⁴⁴. Interestingly, this diminution seems to depend on the integrity of the hippocampus (*e.g.* Refs⁴⁵⁻⁴⁸, but see Ref⁴⁹ for contrasting results), a structure known for its major implication in spatial navigation⁵⁰. Exploration is a central concept in spatial cognition, as this specific behaviour allows the animal to gain spatial knowledge and build representations of its environment^{50,51}. It is a form of latent learning⁵², which refers to the acquisition of knowledge occurring in the absence of explicit reward⁵³.

Organisation of exploration

Exploration behaviour can be triggered by a wide set of stimulus (*e.g.* a new environment⁵⁴, a new object in a familiar environment⁴⁷, a new spatial arrangement of objects^{47,55}, or even a change in the environment topology⁵⁶). Novelty detection often interferes with the ongoing behavioural activities that animals have to perform, as if acting on the current goal of the animal, prioritising the gathering of new knowledge over feeding^{41,42,57} or other behaviours⁵⁸. However, despite its instinctive component and seemingly random structure, behavioural studies demonstrate that novelty exploration is actually quite organised^{59,60} while still enabling the expression of inter-individual differences⁶¹.

Basically, when exploring a new environment for the first time, a rodent will make excursions from its departure point to unexplored parts of its environment, most often following the borders, and regularly returning to a place termed 'home base'^{39,62,63}. Specific behaviours such as rearing or grooming are more likely to occur at the home base³⁹. The home base is usually the place where the animal was first released in the environment⁶⁴, but it has to provide sufficient shelter to be effective³⁸. Regarding this latter observation, Whishaw and colleagues suggested that exploration would mainly serve to optimise safety. Exploration has similar patterns in the absence of visual cues: in the dark, rats placed in a new environment will still organise their displacements around a chosen home base. Their displacements show invariant characteristics, *e.g.* a dissociation between the

^b *Home range* is not to be confused with the *territoriality* concept, the latter being the protected part of the home range. However, these two concepts largely overlap in some instance (*i.e.* territory can include the entire home range or only the nest; see Ref³⁴ for further discussion on this matter).

1
2
3 outward trajectory (*i.e.* away from the home base) and the return trajectory (towards the home
4 base)⁶⁵.

5
6 More recently, a thorough characterization of mice exploratory behaviour was performed by Fonio
7 and collaborators⁶⁰. The authors demonstrated that exploration of a new and large circular arena
8 could be decomposed in several behavioural patterns, the order of which was highly reproducible
9 among individuals. These behavioural patterns progressively take place within the three-dimensional
10 space: mice first make short back and forth trips from their home base following the wall (one-
11 dimensional motion). Once they complete a full turn, they begin making small incursions inside the
12 environment (two-dimensional) that progressively become independent from the home base. They
13 end up performing jumping movements (three-dimensional). The authors highlighted the fact that in
14 their experiment, exploration was free: the departure point of exploration trips was the mouse
15 home cage, where *ad libitum* water and food were provided, and the time left for exploration was
16 exceptionally long (45h in total). In common rodent experiments, exploration is forced and
17 constrained in time and space, which might explain why the full pattern of exploratory behaviour is
18 usually not observed. The importance of environmental limits (and probably geometrical
19 information) is evidenced by the necessity for the mice to first entirely explore the borders before
20 performing incursions towards the centre of the environment. In addition to providing shelter,
21 borders²¹ and geometrical layout⁶⁶ probably serve as anchor points necessary to build a spatial map
22 of an environment^{50,67}.

23 24 25 26 27 28 *Object exploration*

29
30 The spontaneous exploration of objects is usually seen as good indication that mammals memorise
31 and manipulate representations of space and objects in space (*e.g.* Ref ⁶⁸ in hamsters; Ref ⁶⁹ in rats).
32 Indeed, the selective exploration of new objects in a known environment can only be possible if one
33 has previously stored the arrangement of objects in this environment and is able to compare the
34 current layout with the memorised representation. Many studies rely on spontaneous exploration to
35 assess the memory for the nature or the position of objects, which relates to the 'what' or 'where'
36 aspects of episodic-like memory^{70,71}. In rodents, the hippocampus appears to be selectively involved
37 in processing memory for object locations⁴⁷.

38 39 40 41 *Exploration and task performance*

42
43 Interestingly, Olton and collaborators showed the importance of exploration (also termed 'shaping'
44 in that context) prior to testing. Rats that were not given the opportunity to explore a radial arm
45 maze before testing, did not perform better than chance in the task⁷². Therefore, exploration (or
46 simple pre-exposure to an environment), even in the absence of food, seems necessary for
47 subsequent performance in navigation tasks⁷²⁻⁷⁵. As an example, Chai and White tested rats in their
48 ability to discriminate neighbouring locations in a radial arm maze⁷⁵. In this task, rats were confined
49 to a specific arm of the maze, where they could either find food or not. When later tested with a free
50 choice between adjacent arms that include the food-paired arm, rats demonstrated preference for
51 this arm only if previously exposed to the entire maze^{75,76}. If not pre-exposed to the maze, the
52 knowledge acquired when restrained in an arm was not sufficient to build a representation of the
53 environment and of the spatial configurations of the maze arms. In that regard, it is interesting to
54 note that in complex environments rats spend more time exploring the topologically relevant parts
55
56
57
58
59
60

1
2
3 of a maze (*i.e.* the intersections), probably reflecting encoding of information on the connectivity
4 layout of the environment^{56,77}.

5
6 Evidence reviewed above shows the crucial role of exploration in building a representation of space,
7 and, by extension, in developing accurate navigation strategies. A navigation strategy can be defined
8 by a set of rules to follow in order to reach a spatial goal when one is placed in a particular situation.
9 Spatial information processing can endow the animal with navigation strategies allowing different
10 degrees of behavioural flexibility and complexity. For example, turning left at the green sign is a
11 response strategy whereas going to a specific place defined by its relationships with surrounding
12 cues is a place strategy. Although there are different ways to categorize strategies, they are sharing
13 common features^{13,50,78}.

14 15 16 17 **Guidance**

18
19 In certain navigation situations, the goal is either directly visible or cued. In that case, the best
20 strategy, or at least the less cognitive demanding, is to orient towards the goal and approach it. This
21 type of strategy is termed target approaching (when the goal itself is visible) or beacon approaching
22 (if a cue is located at the goal position), or more generally cue, guidance or taxon strategy. It only
23 requires learning of a single stimulus-response association.

24
25 Contrary to most functions described here, it is generally accepted that the hippocampus is not
26 involved in guidance strategy, or at least that hippocampal lesions do not impair performance in cue-
27 guided tasks⁷⁹⁻⁸². The ability for rats with hippocampal lesions to perform a guidance strategy is
28 often used as a control for non-spatial aspects of behaviour (*e.g.* sensory or motor abilities).

29 30 31 32 **Response strategy**

33
34 In some instances, the goal is neither visible nor directly cued but can be reached by means of
35 associations between elements of the environment and actions (each association being
36 independent from the others). This response (or stimulus-triggered response) strategy, also
37 termed egocentric strategy⁸³, has been first studied by Edward C. Tolman in his search to identify
38 the nature of the information used by animals to solve a spatial task^{84,85}. A commonly used
39 place/response task is the cross-maze task, in which rats are trained to retrieve food from one arm
40 using either a place or a response strategy. During training, access to the north arm is blocked.
41 Animals are then placed on the starting point of the south arm, and allowed to consume the food
42 pellet located at the end of the east arm. In this phase, turning right (action) when facing the
43 intersection (stimulus) will be sufficient to reach the goal. During the probe trial, access to the
44 south arm is blocked. Animals are released from the north arm, and allowed to choose either the
45 east arm (place learning) or the west arm (response learning). Similarly to the cue strategy,
46 hippocampal lesions do not impair performance when the response strategy can be used to
47 navigate towards a goal⁸⁶. Conversely, the striatum is likely to be one of the key structures
48 involved in this strategy⁸⁷. Overall, it seems that in the intact animal, these two structures acquire
49 different types of information simultaneously and in parallel⁸⁸, at least during the early phases of
50 acquisition of the spatial task⁸⁹.

51 52 53 54 55 56 **Routes**

1
2
3 When specific actions can be associated to specific states as in the response strategy, but the
4 knowledge of the state is not sufficient to select the action, one can use a route strategy. The route
5 strategy has also been termed sequential egocentric strategy or sequence-based navigation⁹⁰. It
6 relies on a sequence of stimulus-response actions and it can also be used in a modified version of the
7 Morris water maze in which neither proximal nor distal cues are present⁹⁰. In this task, the animal
8 has to cross three identical intersections before reaching its goal but the action to be performed at
9 each of these intersections is different. Therefore, a sequence of stimulus – response associations
10 must be learned and each choice must be selected according to its position in the sequence. We
11 note that a route strategy is more complex than a succession of cue and response strategies,
12 because the order of the stimuli in the sequence is important. Many structures are likely to be
13 involved in this strategy, which holds a sequential (and possibly a timing) component. The CA1 field
14 of the hippocampus would be one of the structures involved, along with other cortical and
15 subcortical structures⁹⁰.

20 Place navigation

21
22 The strategy which probably requires the highest level of spatial information processing is the place
23 (or map-based) strategy. It consists in localising the goal and oneself using the spatial relationships
24 between elements of the environment. Contrary to the response strategy, it enables flexible
25 behaviour, *i.e.* adaptability in the face of environmental changes. It was postulated to rely on a
26 ‘cognitive map’, as defined by Edward C. Tolman⁵².

27
28 Tolman⁵² suggested that animals can manipulate representations of their environment and that they
29 were not simply stimulus-response machines, in contradiction with the behaviourist approach,
30 largely dominant at that time. Namely, Tolman proposed that rats could rely on a cognitive map to
31 navigate, or, in other words, a neural representation of places and of the relationships between
32 these places, independent of the current position of the subject. Tolman advanced several
33 arguments to support this view. First, the rats are able to find shortcuts and to perform detours.
34 Second, the rats show vicarious-trial-and-error behaviour (*i.e.* rats would occasionally pause and
35 look back and forth at an intersection in a maze) when facing a choice. Third, the rats display several
36 forms of latent learning. For example, that exploration improves further performance in a task⁹¹ is
37 evidence that the animal acquires information in the absence of an explicit reward. Another instance
38 of such latent learning is provided by the observation that rats can incidentally learn what type of
39 reward is available even when not currently motivated for this reward^{92,93}. Fourth, the rats express
40 hypothesis-based (or strategy-based) behaviour. This behaviour corresponds to a form of learning
41 that shows a sudden shift from a near-random to near-perfect performance, contrary to what is
42 observed with trial-and-error learning. Such a change in behaviour would underlie a non-incremental
43 neural process, *i.e.* a sudden change of hypothesis about the world (see Ref⁵³ for a review of these
44 arguments).

45
46 In the late ‘70s, the concept of cognitive map was amended following discoveries on its putative
47 neural bases (namely, the hippocampus⁵⁰). This updated theory, supported by neural data, led to a
48 large amount of research centred on the role of the hippocampus and related brain areas in spatial
49 cognition. To date, although few criticisms (*e.g.* Refs^{94–96}) and reformulations have been addressed
50 (*e.g.* Refs^{97–99}), the cognitive map concept offers one of the most fruitful experimental paradigm in
51 cognitive neuroscience.

BRAIN SUBSTRATES OF NAVIGATION IN MICE AND RATS

Given the extensive behavioural evidence of a flexible use of the different cues (see section Space Perception), it is fair to assume that space representation at the neural level shows a great dependence on multisensory integration. Indeed, such integration is present at the neural level in the hippocampus^{18,100}, where place cells have been first described in the rat¹⁰¹ and later on also in the mouse^{102,103}. We shall review in the following sections the principal differences between these two species in terms of hippocampal place cell activity with a careful look to their basic properties and experience dependent dynamics.

Sidebar title: Hippocampal place cells and the representation of space

Since its discovery in the early '70s by John O'Keefe, hippocampal place cells have been extensively studied in numerous spatial memory paradigms. These pyramidal cells are selectively active in restricted portions of space and change their firing activity (*i.e.* both firing rate and location) according to the nature of the environment being tested. Therefore, these place cells, along with other spatially tuned types of neurons (*e.g.* grid cells¹⁰⁴, head direction cells¹⁰⁵), are thought to provide the rat brain with a unique spatial signature characterizing a specific environment, and thereby a memory trace of the subject's place. Originally discovered in the rat¹⁰¹, place cells have been found since then in other mammalian species, including mouse^{102,103}, big brown bat¹⁰⁶, non-human primates¹⁰⁷ and human¹⁰⁸. Although there is little doubt on the role played by place cells across these various species in spatial processing, few constitutive differences remain, especially in the primate literature. For instance, it appears that hippocampal cells in the non-human primate brain are sensitive to whole-body motion¹⁰⁹ and spatial view¹¹⁰ during passive translocation, while such factors have somewhat limited impact on rodent place cell activity (*e.g.* see Ref¹¹¹ on the "local view" issue). However, it is possible that these discrepancies arise from the experimental design *per se* (passive translocation *versus* active exploration) rather than in any interspecies differences¹¹².

Basic properties of hippocampal pyramidal cells

As mentioned in the Introduction, the growing number of mice used in behavioural studies focusing on learning and memory raises the question of interoperability of the various behavioural tests used in this field of research. To this end, Routh and collaborators¹¹³ asked whether basic properties of hippocampal CA1 pyramidal cells share common features between rats (Sprague-Dawley) and mice (C57BL/6). In line with others^{114–116}, the authors found larger hippocampi in rats than in mice, this difference being partly due to a smaller width of the dentate gyrus in mice¹¹³. However, the total number of neurons might be similar in the two species, as the CA1 pyramidal neurons appeared to be more densely packed in mice^{113,117}.

Routh and colleagues found little difference between rats and C57BL/6 mice regarding the cellular morphologies and passive membrane properties of CA1 pyramidal neurons, except for a more hyperpolarized resting membrane potential, and a lower resonance frequency^c in mice neurons. Since resonance frequency is thought to be directly related to the magnitude of the

^c The membrane potential resonance property describes the ability of neurons to respond selectively to inputs at preferred frequencies¹¹⁸.

1
2
3 hyperpolarization-activated cation current (I_h)^{113,119}, mice would have less I_h active at rest compared
4 to rats. Furthermore, I_h has been shown to regulate dendritic integration of distal synaptic inputs to
5 CA1 pyramidal cells^{120–122}. Deletion of one of the two channels isoforms (HCN1) responsible of I_h
6 enhances behavioural performance in a hippocampal-dependent task, increases the power of theta
7 oscillations and synaptic plasticity at the entorhinal inputs to CA1 neurons¹²². This last result is of
8 particular importance given the central role of synaptic plasticity and long-term potentiation in
9 stabilizing the activity of hippocampal place cells¹²³. As discussed below, differences in molecular
10 composition of HCN channels might be a key component of place field instability generally observed
11 in mice.
12
13

14 15 **Basic spatial properties of place cells**

16
17 Cross-species comparison of the functional properties of place cells appears critical in understanding
18 the general principles underlying hippocampal function¹²⁴. However, there is relatively little
19 comparative information even for the basic spatial properties of place cells (*e.g.* firing rate, spatial
20 coherence, spatial information content and place field size). Several non-exclusive factors might
21 explain this lack of systematic comparison. First, over the 40 years of research on hippocampal place
22 cells, rat has been the dominant model. It is only from the mid '90s that mice models have been
23 used in learning and memory research with emphasis on the molecular and genetic aspects, but not
24 on the fundamental spatial properties of hippocampal place cells. Second, the wide variety of strains
25 in both species (*e.g.* inbred *versus* outbred) and genetic backgrounds used for transgenic research
26 reduces drastically our ability to draw systematic comparisons. Third, no single methodology has
27 been laid down to analyse the various parameters of the spatial discharge of hippocampal neurons.
28 For instance, there exists at least six different ways if only to mathematically define a place field (*i.e.*
29 the portion of space where the place cell is active) all species combined^{125–130}.
30
31
32
33

34
35 Despite all these limitations, it is possible to get a rough idea on the degree of similarity of basic
36 spatial properties of hippocampal place cells recorded in both mice (C57BL/6) and rats (Long-Evans)
37 using nearly identical criteria^{131,103,132–138}. It appears from this selected sample (see Table 1) that
38 average firing activity is similar in both species. The internal organization of the place field (*i.e.*
39 spatial coherence; a measure of the extent to which the firing rate in a pixel is predicted by the rates
40 of its neighbours¹³⁹) is nearly identical as well. It seems that the main difference concerns the spatial
41 information content, which is a measure of the extent to which a cell's firing can be used to predict
42 the position of the animal¹⁴⁰. This index is nearly twofold in rats. **However, inferring any particular**
43 **behavioural alteration from variations of this measure can prove cumbersome given its**
44 **dependency to other variables such as the place field size. Indeed, numerous experimental studies**
45 **reporting a loss of spatial information content report also an increased size of hippocampal place**
46 **fields (*e.g.* Ref ^{141–143}). However, it is unknown whether hippocampal place fields in mice are**
47 **broader than those observed in rats. In addition, works performed by Markus and colleagues¹⁴⁴**
48 **suggest that place field reliability is more important for spatial navigation than the size of the**
49 **place field *per se*. This issue will be developed in the following section.**
50
51
52
53
54
55
56
57
58
59
60

Table 1: Comparison of main properties of place cells in mice and rats

	Mice (range ^{Refs.})	Rats (range ^{Refs.})
Average firing (Hz)	1.1–2.27 ^{103,133,136–138}	0.79–1.73 ^{131,132,135}
Spatial coherence	0.51–0.71 ^{103,133,137,138}	0.64–0.67 ^{131,134,135}
Information (bits per spike)	0.7–0.85 ^{136–138}	1.43–2.11 ^{131,132,134,135}
Stability	0.3–0.45 ^{145,146}	0.5–0.7 ^{147,148}

Place cell activity over time

A great deal of studies that initially explored the relationship between place cell activity and behaviour involved lesioning or inactivating specific brain areas (see Ref¹⁴⁹ for a review on this specific matter). Most of these studies were performed in the rat and showed that performance deteriorated when place cell activity was altered^{150–152}. Studies conducted in transgenic mice reached a similar conclusion^{137,138,153–156}.

Aging studies provided further support for the idea that place cell activity was tightly linked to behavioural performance in rodents^{141,157–159}. More precisely, major differences are observed between young and aged animals when comparing place field stability across days^d. For instance, hippocampal place cells in young rats show strong place field stability over time¹⁶⁰ while aged animals show spontaneous rearrangements of place field locations (*i.e.* place cells remap) from time to time¹⁶¹. At this point, it is important to note that the same aging effect has been reported for mice place cells¹⁶². **However, a major interspecies difference is found when comparing place field stability (see Table 1); hippocampal place cell representation in mice does show a marked instability in normal conditions^{103,133,145,146}.**

Place field relative instability has been reported straight from the beginning of electrophysiological recordings in freely-moving mice^{103,133}, but has been specifically investigated by Kentros and collaborators a few years later¹⁴⁵. Since then, this particular aspect of place cells in mice has been reported in other electrophysiological¹⁴⁶ and calcium imaging^{163,164} studies. In the forthcoming sections we will review the different hypotheses that tried to explain interspecies differences regarding the place cells dynamics.

Attentional hypothesis

Kentros and collaborators¹⁴⁵ showed that mice place fields are unstable when the behavioural task did not require any particular attention (*i.e.* the animal was left free to explore an open environment). Conversely, when the animal had to perform a pellet chasing task or, to a greater extent, when it had to solve a spatial navigation task (*i.e.* the animal had to reach an unmarked zone in the environment to receive a reward), place cells showed highly reproducible patterns of activity between sessions. This work also showed that a positive correlation exists between the level of behavioural performance and the degree of place cells stability: the best performing animals had the

^d A place cell that fires at the same location in a familiar environment across multiple sessions is said to show a stable place field.

1
2
3 more stable place fields. The authors **assumed therefore** that attentional processes were
4 responsible for the increase of place field stability.
5

6 Such attentional effect is also observed in the rat but in very particular conditions. Zinyuk and
7 collaborators¹⁶⁵ trained rats to perform either a simple pellet chasing task or a navigation task on a
8 rotating arena. The continuous rotation of the arena in a cue-rich room allowed to dissociate the
9 stationary room-based from the rotating arena-based reference frame. The animals that were
10 trained in the simple pellet chasing task showed less stable place fields than the animals that were
11 trained in the navigation task when tested on the rotating arena (*i.e.* firing was more organized in
12 the task-relevant frame). In the same line, Fenton and Muller¹⁶⁶ showed that in a simple pellet-
13 chasing task, place cell firing was not nearly as reliable in the time domain as in the positional
14 domain (*i.e.* place cell discharge during different passes through the firing field is extremely variable,
15 a phenomenon called overdispersion). Fenton and collaborators¹⁴⁷ showed that attention could
16 constraint this temporal variability of place cell firing.
17
18
19

20 Overall, Kentros and colleagues¹⁴⁵ explain the natural instability of place fields in mice by arguing
21 that these animals pay less attention to distal environmental cues compared to rats. This idea is
22 supported by the work of Eichenbaum and colleagues¹⁶⁷ showing that place cells in the mice are
23 more easily controlled by local rather than distal cues. In this task, mice were allowed to explore a
24 plus-maze that contained a large set of controlled stimuli, including local cues consisting of a
25 distinctive surface on each maze arm. Additionally, distal cues, composed of distinct three-
26 dimensional objects, were fixed on a curtain surrounding the maze. On the test phase, local and
27 distal cues were rotated 90° in opposite directions. During this test phase, in control mice, place cells
28 appeared to follow local rather than distal cues. However, these results could be also interpreted
29 based on a hierarchical organization of sensory inputs, since the local cues were tactile and the distal
30 cues were relying on the visual modality.
31
32
33

34 *Hierarchical organization of sensory inputs*

35
36 Although attention positively modulates place field stability in mice, it should be noted that this
37 degree of stability remains relatively low when compared to recordings obtained in similar
38 conditions in rats (see Table 1). Las and Ulanovsky¹²⁴ speculate that these discrepancies can be
39 attributed to a differential use of sensory inputs in rats and mice. According to the authors, olfactory
40 cues might play a much more important role in place field formation in mice than in rats. Indeed,
41 theoretical¹⁶⁸ and experimental work¹⁶⁹ suggest that olfactory cues might control place field activity
42 to a greater extent than what has been previously thought¹⁷⁰. **In addition, experimental data from**
43 **several behavioural experiments** (reviewed in Ref ¹⁷¹) show that olfactory cues affect a wide set of
44 behaviours in mice, perhaps more strongly than in rats (but see subsection Genetic Differences
45 below). Added to the fact that visual acuity is poorer in mice¹⁷², Las and Ulanovsky¹²⁴ suggest that
46 rats would tend to develop more visually-based maps whereas mice would develop olfactory-based
47 map. The relative importance of the various sensory information in shaping the place cell activity in
48 mice remains however to be tested more thoroughly.
49
50
51
52
53

54 Another argument presented by Kentros and colleagues¹⁴⁵ in favour of genuine cognitive differences
55 between mice and rats relies on results showing poorer performance in the Morris water maze task
56 in mice⁹⁻¹¹. This task is thought to rely heavily on a distal cues triangulation process¹⁷³, **although rats**
57 **could use preferentially directional responding over true place navigation on occasions**^{174,175}.
58
59
60

1
2
3 Accordingly, rats use complex spatial strategies to find the hidden platform in the maze^{4,176}. On the
4 contrary, swimming patterns of the mice appeared more stereotyped, reflecting a preferential use of
5 sequence-based navigation¹⁷⁷. This last observation has to be considered along with further results
6 obtained in rats by Hamilton and colleagues¹⁷⁸ showing that lesions of the dorsal tegmental nuclei
7 (a brain structure known to contain head-direction cells) disrupt landmark-based navigation in this
8 task. Given the predominant influence of the head-direction system on place cell activity¹⁷⁹, one
9 can formulate the hypothesis that place field instability observed in mice might be closely linked
10 to an instability in the head-direction signals. Indeed, when comparing head-direction cells
11 characteristics between these two species, it appears that these cells are less reliably anchored to
12 salient environmental cues in mice¹⁸⁰. Nonetheless, particular caution should be taken in
13 interpreting mice behavioural data obtained in the water maze task, as the nature of the
14 behavioural strategy used during training could impact the way results are obtained in the probe
15 trials^{181–183}. For instance, adopting a spiralling search strategy during training can prove to be quite
16 effective to locate the platform but mice showing such behaviour will score poorly in the final
17 probe trial. Overall, it seems that mice use less robust and flexible strategies to solve spatial tasks
18 than rats do¹⁸⁴ but show nonetheless a certain capability to switch strategies when given the
19 opportunity⁹⁰.

24 *Behavioural factors underlying stable place field activity*

25
26 Exploratory behaviour (see section Exploring Space) is a complex response to novelty that results
27 from a compromise between the motivation to gather information about the surroundings and the
28 need to avoid predators^{185,186}. As tracking technology improves, it is now feasible to analyse carefully
29 the fine locomotor elements of exploratory behaviour in rodents⁴⁰. Several studies by Golani and
30 colleagues identified the moment-to-moment developmental sequence of forced^{187,37} and free
31 exploration⁶⁰ in rodents. Forced exploration refers to the procedure where the animal is placed
32 directly into the test box at the start of the session, whereas in the free exploration procedure the
33 animal has access to both the test box and its home cage¹⁸⁸.

34
35 In forced exploration, Long-Evans rats and BALB/cJtau mice show a gradual increase of excursion
36 length when placed in the arena. For both species, path length increases across individuals both
37 within and across multiple sessions, reflecting some habituation process. In contrast, in the same
38 conditions, C57BL/6Jtau mice show a complete reversed profile across the session (*i.e.* when
39 introduced in the arena, C57BL/6Jtau mice start with full circle excursions and only then proceed
40 with smaller radial movements). This behaviour is likely to reflect greater risk taking of C57BL/6 mice
41 compared to BALB/c¹⁸⁹. In free exploration, these strain differences are much less pronounced, as
42 C57BL/6 and BALB/c mice share common exploratory patterns⁶⁰.

43
44 To sum up, in classic studies of place cells in freely-moving rodents, the exploratory behaviour of
45 C57BL/6 mice appears rather different from that of rats and might contribute to some extent to the
46 differences observed in terms of place field stability. It is also important to note that a food pellet
47 chasing task will likely interfere with the proper completion of exploratory behaviour^{54,190}. Therefore,
48 systematic comparisons of place cells recordings in different mice strains showing behavioural
49 differences in forced but not in free exploration (*e.g.* C57BL/6 *versus* BALB/c) would shed light on the
50 contribution of specific locomotor patterns to stabilize place cell activity.

57 *Genetic differences*

1
2
3 Although belonging to the same subfamily Murinae, rats and mice share only 30% of their DNA
4 sequences⁵. Most of the genetic differences observed between these two species concern olfactory
5 receptors, which are nearly 40% more numerous in rat's genome. Other major differences involve
6 multiple biological processes such as pheromones detection, detoxification and proteolysis. Apart
7 from the qualitative differences in genome sequences, rats and mice might differ also in channel
8 subunit composition. For instance, as previously suggested by Routh and colleagues¹¹³, a particular
9 subunit composition of h channels (composed of HCN1 and HCN2 isoforms) in mouse would explain
10 the lower hyperpolarization-activated cation current (I_h) at the entorhinal—CA1 synapse (see
11 subsection Basic properties of hippocampal pyramidal cells above). Interestingly, Kandel and
12 colleagues¹⁹¹ performed hippocampal place cells recordings in HCN1 knockout mice in various
13 behavioural tasks. They found that CA1 hippocampal place fields in these mice were larger and more
14 stable than the controls. These electrophysiological data complement nicely the behavioural results
15 showing improved performance of the HCN1 knockout mice in a hippocampal-dependent task¹²².
16 Additionally, a recent study performed by Bittner and colleagues¹⁹² showed that active dendritic
17 integration in pyramidal neurons at the entorhinal—CA1 synapse is instrumental in forming new
18 place fields and that similar mechanisms might be involved in stabilizing place cell activity.
19
20
21
22

23
24 **To summarize, although sharing common basic neural features, rats and mice do show significant**
25 **differences when comparing brain representations of space. The relative instability of**
26 **representations in mice might lead to cognitive differences that are expressed not so much as**
27 **differences in behavioural performance as differences in navigation strategy selection. Molecular**
28 **variants of certain channels expressed in the mouse hippocampus might be directly related to this**
29 **phenomenon but constitute undoubtedly only a small fraction of the pertinent genetic factors that**
30 **are at play in space representation.**
31

32 33 Conclusion

34
35 Much of our review focused on the major neuroethological differences existing between mice and
36 rats in spatial cognition. Although sharing many behavioural characteristics in simple exploration
37 tasks^{10,37}, the neural representation of space differs largely between these two species in terms of
38 stability^{133,145,146,163,164}. This last observation correlates to some extent with interspecies differences
39 in navigational strategies used to solve spatial tasks^{4,177,184}. Additionally, it appears that a simple
40 modification in behavioural paradigms (*e.g.* free *versus* forced exploration) can induce important
41 behavioural changes within one single strain^{37,60}. On the other hand, growing evidence converge
42 towards molecular explanation for the origin of place field instability in mice^{113,122,191}. More
43 importantly, these constitutive differences appear unrelated to the positive attentional effect
44 observed on place field stability^{145,146} as forebrain deletion of HCN1 does not involve changes in
45 anxiety or attention¹²².
46
47
48

49
50 Additionally, we let deliberately aside the strain issues in our review. All the behavioural and
51 physiological data discussed in the previous section dealt with the C57BL/6 mouse genetic
52 background unless otherwise stated. However, numerous reports stressed out the importance of the
53 strain being used in spatial tasks, these between-strain differences leading sometimes to contrasting
54 results^{193–195,9,60,113,37}. Added to the fact that laboratory environment is likely influencing behavioural
55 results¹⁹⁶ and that interindividual variability in genetically identical mice emerges with time¹⁹⁷, all
56
57
58
59
60

1
2
3 these considerations strongly support the need of a greater behavioural and physiological
4 characterization of animal models used in learning and memory research⁴⁴.
5

6 **Notes**

7
8 [Please add any notes here]
9

10
11
12 **References**

- 13
14 1. Hedrich, H. J. Chapter 1 - History, Strains and Models. in Krinke, G. J. ed. The
15 laboratory rat. Academic Press, London; 2000 pp.3–16.
16
17 2. Pankevich, D. E., Wizemann, T. M., Mazza, A.-M. & Altevogt, B. M. *International*
18 *Animal Research Regulations: Impact on Neuroscience Research: Workshop Summary*.
19 (National Academies Press, 2012).
20
21 3. Chinwalla, A. T. *et al.* Initial sequencing and comparative analysis of the mouse
22 genome. *Nature* **420**, 520–562 (2002).
23
24 4. Frick, K. M., Stillner, E. T. & Berger-Sweeney, J. Mice are not little rats: species
25 differences in a one-day water maze task. *Neuroreport* **11**, 3461–3465 (2000).
26
27 5. Gibbs, R. A. *et al.* Genome sequence of the Brown Norway rat yields insights into
28 mammalian evolution. *Nature* **428**, 493–521 (2004).
29
30 6. Silva, A. J., Stevens, C. F., Tonegawa, S. & Wang, Y. Deficient hippocampal long-term
31 potentiation in alpha-calcium-calmodulin kinase II mutant mice. *Science* **257**, 201–206
32 (1992).
33
34 7. Silva, A. J., Paylor, R., Wehner, J. M. & Tonegawa, S. Impaired spatial learning in
35 alpha-calcium-calmodulin kinase II mutant mice. *Science* **257**, 206–211 (1992).
36
37 8. Bach, M. E., Hawkins, R. D., Osman, M., Kandel, E. R. & Mayford, M. Impairment of
38 spatial but not contextual memory in CaMKII mutant mice with a selective loss of
39 hippocampal LTP in the range of the θ frequency. *Cell* **81**, 905–915 (1995).
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 9. Whishaw, I. Q. A comparison of rats and mice in a swimming pool place task and
4
5 matching to place task: Some surprising differences. *Physiol. Behav.* **58**, 687–693
6
7 (1995).
8
- 9
10 10. Whishaw, I. Q. & Tomie, J.-A. Of Mice and Mazes: Similarities Between Mice and Rats
11
12 on Dry Land But Not Water Mazes. *Physiol. Behav.* **60**, 1191–1197 (1996).
13
- 14 11. Stranahan, A. M. Similarities and differences in spatial learning and object recognition
15
16 between young male C57Bl/6J mice and Sprague-Dawley rats. *Behav. Neurosci.* **125**,
17
18 791–795 (2011).
19
- 20 12. Watson, J. B. & Lashley, K. S. Homing and related activities of birds. Carnegie
21
22 Institution of Washington, Washington; 1915.
23
- 24 13. Gallistel, C. R. The organization of learning. MIT Press, Cambridge; 1990.
25
- 26 14. Save, E. & Poucet, B. Piloting. in Whishaw I. Q., Kolb B. eds. The behavior of the
27
28 laboratory rat: a handbook with tests. Oxford University Press, New York; 2004 pp.
29
30 392–400.
31
32
- 33 15. Arleo, A. & Rondi-Reig, L. Multimodal sensory integration and concurrent navigation
34
35 strategies for spatial cognition in real and artificial organisms. *J. Integr. Neurosci.* **6**,
36
37 327–366 (2007).
38
39
- 40 16. Save, E., Nerad, L. & Poucet, B. Contribution of multiple sensory information to place
41
42 field stability in hippocampal place cells. *Hippocampus* **10**, 64–76 (2000).
43
44
- 45 17. Jeffery, K. J. Integration of the sensory inputs to place cells: What, where, why, and
46
47 how? *Hippocampus* **17**, 775–785 (2007).
48
- 49 18. Ravassard, P. *et al.* Multisensory Control of Hippocampal Spatiotemporal Selectivity.
50
51 *Science* **340**, 1342–1346 (2013).
52
53
54
55
56
57
58
59
60

19. Wallace, D. G. & Whishaw, I. Q. Dead reckoning. Whishaw I. Q., Kolb B. eds. The behavior of the laboratory rat: a handbook with tests. Oxford University Press, New York; 2004 pp. 401–409.
20. Wallace, D. G., Hines, D. J., Pellis, S. M. & Whishaw, I. Q. Vestibular Information Is Required for Dead Reckoning in the Rat. *J. Neurosci.* **22**, 10009–10017 (2002).
21. Zhang, S., Schönfeld, F., Wiskott, L. & Manahan-Vaughan, D. Spatial representations of place cells in darkness are supported by path integration and border information. *Front. Behav. Neurosci.* **8**, (2014).
22. Etienne, A. S. & Jeffery, K. J. Path integration in mammals. *Hippocampus* **14**, 180–192 (2004).
23. Poucet, B. *et al.* Is there a pilot in the brain? Contribution of the self-positioning system to spatial navigation. *Front. Behav. Neurosci.* **9**, 292 (2015).
24. Lavenex, P. & Schenk, F. Olfactory traces and spatial learning in rats. *Anim. Behav.* **56**, 1129–1136 (1998).
25. Rossier, J. & Schenk, F. Olfactory and/or visual cues for spatial navigation through ontogeny: olfactory cues enable the use of visual cues. *Behav. Neurosci.* **117**, 412–425 (2003).
26. Rossier, J., Haerberli, C. & Schenk, F. Auditory cues support place navigation in rats when associated with a visual cue. *Behav Brain Res* **117**, 209–14 (2000).
27. Etienne, A. S., Teroni, E., Hurni, C. & Portenier, V. The effect of a single light cue on homing behaviour of the golden hamster. *Anim. Behav.* **39**, 17–41 (1990).
28. Rotenberg, A. & Muller, R. U. Variable place-cell coupling to a continuously viewed stimulus: evidence that the hippocampus acts as a perceptual system. *Philos Trans R Soc Lond B Biol Sci* **352**, 1505–13 (1997).

- 1
2
3 29. Berthoz, A. & Viaud-Delmon, I. Multisensory integration in spatial orientation. *Curr.*
4
5 *Opin. Neurobiol.* **9**, 708–712 (1999).
6
- 7
8 30. Maaswinkel, H. & Whishaw, I. Q. Homing with locale, taxon, and dead reckoning
9
10 strategies by foraging rats: sensory hierarchy in spatial navigation. *Behav. Brain Res.*
11
12 **99**, 143–152 (1999).
13
- 14 31. Biegler, R. & Morris, R. G. M. Landmark stability is a prerequisite for spatial but not
15
16 discrimination learning. *Nature* **361**, 631–633 (1993).
17
- 18 32. Biegler, R. & Morris, R. G. Landmark stability: further studies pointing to a role in
19
20 spatial learning. *Q. J. Exp. Psychol. B* **49**, 307–345 (1996).
21
22
- 23 33. Seton, E. T. Life-histories of northern animals : an account of the mammals of
24
25 Manitoba. Scribner, New York City; 1909.
26
- 27 34. Burt, W. H. Territoriality and Home Range Concepts as Applied to Mammals. *J.*
28
29 *Mammal.* **24**, 346–352 (1943).
30
31
- 32 35. Steiniger, F. Beiträge zur Soziologie und sonstigen Biologie der Wanderratte. *Z. Für*
33
34 *Tierpsychol.* **7**, 356–379 (1950).
35
- 36 36. Hediger, H. Wild animals in captivity. Butterworth Scientific, London; 1950.
37
- 38 37. Draï, D., Kafkafi, N., Benjamini, Y., Elmer, G. & Golani, I. Rats and mice share
39
40 common ethologically relevant parameters of exploratory behavior. *Behav. Brain Res.*
41
42 **125**, 133–140 (2001).
43
44
- 45 38. Whishaw, I. Q., Gharbawie, O. A., Clark, B. J. & Lehmann, H. The exploratory
46
47 behavior of rats in an open environment optimizes security. *Behav. Brain Res.* **171**,
48
49 230–239 (2006).
50
- 51 39. Eilam, D. & Golani, I. Home base behavior of rats (*Rattus norvegicus*) exploring a
52
53 novel environment. *Behav. Brain Res.* **34**, 199–211 (1989).
54
55
56
57
58
59
60

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
40. Golani, I., Benjamini, Y., Dvorkin, A., Lipkind, D. & Kafkafi, N. Locomotor and exploratory behavior. in Whishaw I. Q., Kolb B. eds. *The behavior of the laboratory rat: a handbook with tests*. Oxford University Press, New York; 2004 pp. 392–400.
 41. Chitty, D. & Shorten, M. Techniques for the Study of the Norway Rat (*Rattus norvegicus*). *J. Mammal.* **27**, 63–78 (1946).
 42. Chance, M. R. A. & Mead, A. P. Competition between Feeding and Investigation in the Rat. *Behaviour* **8**, 174–182 (1955).
 43. Poucet, B., Durup, M. & Thinus-Blanc, C. Short-term and long-term habituation of exploration in rats, hamsters and gerbils. *Behav. Processes* **16**, 203–211 (1988).
 44. Wahlsten, D., Rustay, N. R., Metten, P. & Crabbe, J. C. In search of a better mouse test. *Trends Neurosci.* **26**, 132–136 (2003).
 45. Roberts, W. W., Dember, W. N. & Brodwick, M. Alternation and exploration in rats with hippocampal lesions. *J. Comp. Physiol. Psychol.* **55**, 695–700 (1962).
 46. Leaton, R. N. Exploratory behavior in rats with hippocampal lesions. *J. Comp. Physiol. Psychol.* **59**, 325–330 (1965).
 47. Save, E., Poucet, B., Foreman, N. & Buhot, M.-C. Object exploration and reactions to spatial and nonspatial changes in hooded rats following damage to parietal cortex or hippocampal formation. *Behav Neurosci* **106**, 447–56 (1992).
 48. Harley, C. W. & Martin, G. M. Open Field Motor Patterns and Object Marking, but Not Object Sniffing, Are Altered by Ibotenate Lesions of the Hippocampus. *Neurobiol. Learn. Mem.* **72**, 202–214 (1999).
 49. Moses, S. N., Sutherland, R. J. & McDonald, R. J. Differential involvement of amygdala and hippocampus in responding to novel objects and contexts. *Brain Res. Bull.* **58**, 517–527 (2002).
 50. O'Keefe, J. & Nadel, L. *Hippocampus as a Cognitive Map*. Clarendon, Oxford; 1978.

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
51. Poucet, B., Chapuis, N., Durup, M. & Thinus-Blanc, C. A study of exploratory behavior as an index of spatial knowledge in hamsters. *Anim Learn Behav* **14**, 93–100 (1986).
 52. Tolman, E. C. Cognitive maps in rats and men. *Psychol. Rev.* **55**, 189–208 (1948).
 53. Johnson, A. & Crowe, D. A. Revisiting Tolman, his theories and cognitive maps. *Cogn. Crit.* **1**, 43–72 (2009).
 54. Lever, C., Burton, S. & O’Keefe, J. Rearing on hind legs, environmental novelty, and the hippocampal formation. *Rev Neurosci* **17**, 111–133 (2006).
 55. Wilz, K. J. & Bolton, R. L. Exploratory behavior in response to the spatial rearrangement of familiar stimuli. *Psychon. Sci.* **24**, 117–118 (1971).
 56. Alvernhe, A., Sargolini, F. & Poucet, B. Rats build and update topological representations through exploration. *Anim. Cogn.* **15**, 359–368 (2012).
 57. Cowan, P. E. Systematic patrolling and orderly behaviour of rats during recovery from deprivation. *Anim. Behav.* **25**, Part 1, 171–184 (1977).
 58. Bindra, D. Stimulus change, reactions to novelty, and response decrement. *Psychol. Rev.* **66**, 96–103 (1959).
 59. Avni, R., Zadicario, P. & Eilam, D. Exploration in a dark open field: A shift from directional to positional progression and a proposed model of acquiring spatial information. *Behav. Brain Res.* **171**, 313–323 (2006).
 60. Fonio, E., Benjamini, Y. & Golani, I. Freedom of movement and the stability of its unfolding in free exploration of mice. *Proc. Natl. Acad. Sci.* **106**, 21335–21340 (2009).
 61. Kabbaj, M., Devine, D. P., Savage, V. R. & Akil, H. Neurobiological Correlates of Individual Differences in Novelty-Seeking Behavior in the Rat: Differential Expression of Stress-Related Molecules. *J. Neurosci.* **20**, 6983–6988 (2000).
 62. Golani, I., Benjamini, Y. & Eilam, D. Stopping behavior: constraints on exploration in rats (*Rattus norvegicus*). *Behav. Brain Res.* **53**, 21–33 (1993).

- 1
2
3 63. Draï, D., Benjamini, Y. & Golani, I. Statistical discrimination of natural modes of
4 motion in rat exploratory behavior. *J. Neurosci. Methods* **96**, 119–131 (2000).
5
6
7 64. Nemati, F. & Whishaw, I. Q. The point of entry contributes to the organization of
8 exploratory behavior of rats on an open field: An example of spontaneous episodic
9 memory. *Behav. Brain Res.* **182**, 119–128 (2007).
10
11
12
13 65. Wallace, D. G., Hamilton, D. A. & Whishaw, I. Q. Movement characteristics support a
14 role for dead reckoning in organizing exploratory behavior. *Anim. Cogn.* **9**, 219–228
15 (2006).
16
17
18
19
20 66. Cheng, K. A purely geometric module in the rat's spatial representation. *Cognition* **23**,
21 149–178 (1986).
22
23
24
25 67. Touretzky, D. S. & Redish, A. D. Theory of rodent navigation based on interacting
26 representations of space. *Hippocampus* **6**, 247–270 (1996).
27
28
29
30 68. Thinus-Blanc, C. *et al.* A study of spatial parameters encoded during exploration in
31 hamsters. *J. Exp. Psychol. Anim. Behav. Process.* **13**, 418–427 (1987).
32
33
34 69. Ennaceur, A. & Delacour, J. A new one-trial test for neurobiological studies of memory
35 in rats. 1: Behavioral data. *Behav Brain Res* **31**, 47–59 (1988).
36
37
38
39 70. Dere, E., Huston, J. P. & De Souza Silva, M. A. Integrated memory for objects, places,
40 and temporal order: Evidence for episodic-like memory in mice. *Neurobiol. Learn.*
41 *Mem.* **84**, 214–221 (2005).
42
43
44
45 71. Eacott, M. J., Easton, A. & Zinkivskay, A. Recollection in an episodic-like memory task
46 in the rat. *Learn Mem* **12**, 221–223 (2005).
47
48
49
50 72. Olton, D. S., Collison, C. & Werz, M. A. Spatial memory and radial arm maze
51 performance of rats. *Learn. Motiv.* **8**, 289–314 (1977).
52
53
54 73. Ellen, P., Parko, E. M., Wages, C., Doherty, D. & Herrmann, T. Spatial problem solving
55 by rats: Exploration and cognitive maps. *Learn. Motiv.* **13**, 81–94 (1982).
56
57
58
59
60

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
74. Sutherland, R. J., Chew, G. L., Baker, J. C. & Linggard, R. C. Some limitations on the use of distal cues in place navigation by rats. *Psychobiology* **15**, 48–57 (1987).
75. Chai, S.-C. & White, N. M. Effects of Fimbria-Fornix, Hippocampus, and Amygdala Lesions on Discrimination Between Proximal Locations. *Behav. Neurosci.* **118**, 770–784 (2004).
76. Gaskin, S., Chai, S.-C. & White, N. M. Inactivation of the dorsal hippocampus does not affect learning during exploration of a novel environment. *Hippocampus* **15**, 1085–1093 (2005).
77. Poucet, B. & Herrmann, T. Exploratory patterns of rats on a complex maze provide evidence for topological coding. *Behav Process.* **53**, 155–162 (2001).
78. Redish, A. D. *Beyond the Cognitive Map*. MIT Press, Cambridge; 1999.
79. Morris, R. G., Garrud, P., Rawlins, J. N. & O'Keefe, J. Place navigation impaired in rats with hippocampal lesions. *Nature* **297**, 681–683 (1982).
80. Jarrard, L. E., Okaichi, H., Steward, O. & Goldschmidt, R. B. On the role of hippocampal connections in the performance of place and cue tasks: Comparisons with damage to hippocampus. *Behav. Neurosci.* **98**, 946–954 (1984).
81. Rasmussen, M., Barnes, C. A. & McNaughton, B. L. A systematic test of cognitive mapping, working-memory, and temporal discontinuity theories of hippocampal function. *Psychobiology* **17**, 335–348 (2013).
82. Packard, M. G. & McGaugh, J. L. Double dissociation of fornix and caudate nucleus lesions on acquisition of two water maze tasks: Further evidence for multiple memory systems. *Behav. Neurosci.* **106**, 439–446 (1992).
83. Kesner, R. P., Farnsworth, G. & DiMattia, B. V. Double dissociation of egocentric and allocentric space following medial prefrontal and parietal cortex lesions in the rat. *Behav. Neurosci.* **103**, 956–961 (1989).

- 1
2
3 84. Tolman, E. C., Ritchie, B. F. & Kalish, D. Studies in spatial learning. II. Place learning
4 versus response learning. *J. Exp. Psychol.* **36**, 221–229 (1946).
5
6
7 85. Tolman, E. C. & Gleitman, H. Studies in spatial learning: VII. Place and response
8 learning under different degrees of motivation. *J. Exp. Psychol.* **39**, 653–659 (1949).
9
10 86. Packard, M. G. & McGaugh, J. L. Inactivation of Hippocampus or Caudate Nucleus
11 with Lidocaine Differentially Affects Expression of Place and Response Learning.
12 *Neurobiol. Learn. Mem.* **65**, 65–72 (1996).
13
14 87. McDonald, R. J. & White, N. M. A triple dissociation of memory systems:
15 Hippocampus, amygdala, and dorsal striatum. *Behav. Neurosci.* **107**, 3–22 (1993).
16
17 88. McDonald, R. J. & White, N. M. Parallel information processing in the water maze:
18 Evidence for independent memory systems involving dorsal striatum and hippocampus.
19 *Behav. Neural Biol.* **61**, 260–270 (1994).
20
21 89. Jacobson, T. K., Gruenbaum, B. F. & Markus, E. J. Extensive training and hippocampus
22 or striatum lesions: Effect on place and response strategies. *Physiol. Behav.* **105**, 645–
23 652 (2012).
24
25 90. Rondi-Reig, L. *et al.* Impaired Sequential Egocentric and Allocentric Memories in
26 Forebrain-Specific–NMDA Receptor Knock-Out Mice during a New Task Dissociating
27 Strategies of Navigation. *J. Neurosci.* **26**, 4071–4081 (2006).
28
29 91. Kimble, D. P. & Greene, E. G. Absence of latent learning in rats with hippocampal
30 lesions. *Psychon. Sci.* **11**, 99–100 (1968).
31
32 92. Spence, K. W. & Lippitt, R. An experimental test of the sign-gestalt theory of trial and
33 error learning. *J. Exp. Psychol.* **36**, 491–502 (1946).
34
35 93. Bendig, A. W. Latent learning in a water maze. *J. Exp. Psychol.* **43**, 134–137 (1952).
36
37 94. Bennett, A. T. Do animals have cognitive maps? *J. Exp. Biol.* **199**, 219–224 (1996).
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 95. Gibson, B. M. Cognitive maps not used by humans (*Homo sapiens*) during a dynamic
4
5 navigational task. *J. Comp. Psychol.* **115**, 397–402 (2001).
6
- 7 96. Bennett, M. R. & Hacker, P. M. S. History of cognitive neuroscience. Wiley-Blackwell,
8
9 Chichester; 2008.
10
- 11 97. Poucet, B. Spatial cognitive maps in animals: new hypotheses on their structure and
12
13 neural mechanisms. *Psychol Rev* **100**, 163–82 (1993).
14
- 15 98. Eichenbaum, H., Dudchenko, P., Wood, E., Shapiro, M. & Tanila, H. The
16
17 Hippocampus, Memory, and Place Cells: Is It Spatial Memory or a Memory Space?
18
19 *Neuron* **23**, 209–226 (1999).
20
- 21 99. Jacobs, L. F. & Schenk, F. Unpacking the cognitive map: The parallel map theory of
22
23 hippocampal function. *Psychol. Rev.* **110**, 285–315 (2003).
24
- 25 100. Chen, G., King, J. A., Burgess, N. & O’Keefe, J. How vision and movement combine in
26
27 the hippocampal place code. *Proc. Natl. Acad. Sci.* **110**, 378–383 (2013).
28
- 29 101. O’Keefe, J. & Dostrovsky, J. The hippocampus as a spatial map. Preliminary evidence
30
31 from unit activity in the freely-moving rat. *Brain Res.* **34**, 171–175 (1971).
32
- 33 102. McHugh, T. J., Blum, K. I., Tsien, J. Z., Tonegawa, S. & Wilson, M. A. Impaired
34
35 hippocampal representation of space in CA1-specific NMDAR1 knockout mice. *Cell*
36
37 **87**, 1339–1349 (1996).
38
- 39 103. Rotenberg, A., Mayford, M., Hawkins, R. D., Kandel, E. R. & Muller, R. U. Mice
40
41 Expressing Activated CaMKII Lack Low Frequency LTP and Do Not Form Stable
42
43 Place Cells in the CA1 Region of the Hippocampus. *Cell* **87**, 1351–1361 (1996).
44
- 45 104. Fyhn, M., Molden, S., Witter, M. P., Moser, E. I. & Moser, M.-B. Spatial representation
46
47 in the entorhinal cortex. *Science* **305**, 1258–64 (2004).
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 105. Taube, J. S., Muller, R. U. & Ranck, J. B. Head-direction cells recorded from the
4
5 postsubiculum in freely moving rats. I. Description and quantitative analysis. *J Neurosci*
6
7 **10**, 420–35 (1990).
8
9
10 106. Ulanovsky, N. & Moss, C. F. Hippocampal cellular and network activity in freely
11
12 moving echolocating bats. *Nat Neurosci* **10**, 224–233 (2007).
13
14 107. Matsumura, N. *et al.* Spatial- and task-dependent neuronal responses during real and
15
16 virtual translocation in the monkey hippocampal formation. *J. Neurosci.* **19**, 2381–2393
17
18 (1999).
19
20 108. Ekstrom, A. D. *et al.* Cellular networks underlying human spatial navigation. *Nature*
21
22 **425**, 184–8 (2003).
23
24 109. O’Mara, S. M., Rolls, E. T., Berthoz, A. & Kesner, R. P. Neurons responding to whole-
25
26 body motion in the primate hippocampus. *J. Neurosci.* **14**, 6511–6523 (1994).
27
28 110. Rolls, E. T. & O’Mara, S. M. View-responsive neurons in the primate hippocampal
29
30 complex. *Hippocampus* **5**, 409–424 (1995).
31
32 111. Muller, R. U., Bostock, E., Taube, J. S. & Kubie, J. L. On the directional firing
33
34 properties of hippocampal place cells. *J Neurosci* **14**, 7235–51 (1994).
35
36 112. Gavrilov, V. V., Wiener, S. I. & Berthoz, A. Discharge correlates of hippocampal
37
38 complex spike neurons in behaving rats passively displaced on a mobile robot.
39
40 *Hippocampus* **8**, 475–490 (1998).
41
42 113. Routh, B. N., Johnston, D., Harris, K. & Chitwood, R. A. Anatomical and
43
44 Electrophysiological Comparison of CA1 Pyramidal Neurons of the Rat and Mouse. *J.*
45
46 *Neurophysiol.* **102**, 2288–2302 (2009).
47
48 114. Kalisch, R. *et al.* Anxiety and Hippocampus Volume in the Rat.
49
50 *Neuropsychopharmacology* **31**, 925–932 (2005).
51
52
53
54
55
56
57
58
59
60

- 1
2
3 115. Kovačević, N. *et al.* A Three-dimensional MRI Atlas of the Mouse Brain with Estimates
4 of the Average and Variability. *Cereb. Cortex* **15**, 639–645 (2005).
5
6
7 116. Ma, Y. *et al.* A three-dimensional digital atlas database of the adult C57BL/6J mouse
8 brain by magnetic resonance microscopy. *Neuroscience* **135**, 1203–1215 (2005).
9
10 117. Buzsáki, G. *et al.* Hippocampal network patterns of activity in the mouse. *Neuroscience*
11 **116**, 201–11 (2003).
12
13 118. Hutcheon, B. & Yarom, Y. Resonance, oscillation and the intrinsic frequency
14 preferences of neurons. *Trends Neurosci.* **23**, 216–222 (2000).
15
16 119. Narayanan, R. & Johnston, D. Long-Term Potentiation in Rat Hippocampal Neurons Is
17 Accompanied by Spatially Widespread Changes in Intrinsic Oscillatory Dynamics and
18 Excitability. *Neuron* **56**, 1061–1075 (2007).
19
20 120. Magee, J. C. Dendritic Ih normalizes temporal summation in hippocampal CA1
21 neurons. *Nat. Neurosci.* **2**, 508–514 (1999).
22
23 121. Robinson, R. B. & Siegelbaum, S. A. Hyperpolarization-Activated Cation Currents:
24 From Molecules to Physiological Function. *Annu. Rev. Physiol.* **65**, 453–480 (2003).
25
26 122. Nolan, M. F. *et al.* A Behavioral Role for Dendritic Integration: HCN1 Channels
27 Constrain Spatial Memory and Plasticity at Inputs to Distal Dendrites of CA1 Pyramidal
28 Neurons. *Cell* **119**, 719–732 (2004).
29
30 123. Dragoi, G., Harris, K. D. & Buzsáki, G. Place representation within hippocampal
31 networks is modified by long-term potentiation. *Neuron* **39**, 843–53 (2003).
32
33 124. Las, L. & Ulanovsky, N. Hippocampal neurophysiology across species. in Derdikman,
34 D. & Knierim, J. J. eds. *Space, Time and Memory in the Hippocampal Formation*.
35 Springer, Vienna; 2014. pp. 431–461.
36
37 125. Muller, R. U., Kubie, J. L. & Ranck, J. B. Spatial firing patterns of hippocampal
38 complex-spike cells in a fixed environment. *J Neurosci* **7**, 1935–50 (1987).
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 126. Mehta, M. R., Barnes, C. A. & McNaughton, B. L. Experience-dependent, asymmetric
4 expansion of hippocampal place fields. *Proc. Natl. Acad. Sci.* **94**, 8918–8921 (1997).
5
6
7 127. Hollup, S. A., Molden, S., Donnett, J. G., Moser, M.-B. & Moser, E. I. Place fields of
8 rat hippocampal pyramidal cells and spatial learning in the watermaze. *Eur J Neurosci*
9 **13**, 1197–208 (2001).
10
11
12 128. Mankin, E. A. *et al.* Neuronal code for extended time in the hippocampus. *Proc. Natl.*
13 *Acad. Sci.* **109**, 19462–19467 (2012).
14
15
16 129. Dragoi, G. & Tonegawa, S. Preplay of future place cell sequences by hippocampal
17 cellular assemblies. *Nature* **469**, 397–401 (2011).
18
19
20 130. Tanila, H., Sipilä, P., Shapiro, M. & Eichenbaum, H. Brain aging: impaired coding of
21 novel environmental cues. *J Neurosci* **17**, 5167–5174 (1997).
22
23
24 131. Poucet, B., Thinus-Blanc, C. & Muller, R. U. Place cells in the ventral hippocampus of
25 rats. *Neuroreport* **5**, 2045–2048 (1994).
26
27
28 132. Save, E., Cressant, A., Thinus-Blanc, C. & Poucet, B. Spatial firing of hippocampal
29 place cells in blind rats. *J Neurosci* **18**, 1818–26 (1998).
30
31
32 133. Rotenberg, A., Abel, T., Hawkins, R. D., Kandel, E. R. & Muller, R. U. Parallel
33 instabilities of long-term potentiation, place cells, and learning caused by decreased
34 protein kinase A activity. *J Neurosci* **20**, 8096–8102 (2000).
35
36
37 134. Save, E., Paz-Villagrán, V., Alexinsky, T. & Poucet, B. Functional interaction between
38 the associative parietal cortex and hippocampal place cell firing in the rat. *Eur J*
39 *Neurosci* **21**, 522–30 (2005).
40
41
42 135. Renaudineau, S., Poucet, B. & Save, E. Flexible use of proximal objects and distal cues
43 by hippocampal place cells. *Hippocampus* **17**, 381–395 (2007).
44
45
46 136. Cacucci, F., Wills, T. J., Lever, C., Giese, K. P. & O’Keefe, J. Experience-dependent
47 increase in CA1 place cell spatial information, but not spatial reproducibility, is
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 dependent on the autophosphorylation of alphaCaMKII. *J. Neurosci.* **27**, 7854–7859
4
5 (2007).
6
7
8 137. Renaudineau, S., Poucet, B., Laroche, S., Davis, S. & Save, E. Impaired long-term
9 stability of CA1 place cell representation in mice lacking the transcription factor
10 *zif268/egr1*. *Proc. Natl. Acad. Sci.* **106**, 11771–11775 (2009).
11
12
13 138. Rochefort, C. *et al.* Cerebellum Shapes Hippocampal Spatial Code. *Science* **334**, 385–
14 389 (2011).
15
16
17 139. Muller, R. U. & Kubie, J. L. The firing of hippocampal place cells predicts the future
18 position of freely moving rats. *J Neurosci* **9**, 4101–10 (1989).
19
20
21 140. Skaggs, W. E., McNaughton, B. L. & Gothard, K. M. An Information-Theoretic
22 Approach to Deciphering the Hippocampal Code. in Hanson, S. J., Cowan, J. D. &
23 Giles, C. L. eds. Morgan Kaufmann, San Mateo, CA; 1993. pp. 1030–1037.
24
25
26 141. Cacucci, F., Yi, M., Wills, T. J., Chapman, P. & O’Keefe, J. Place cell firing correlates
27 with memory deficits and amyloid plaque burden in Tg2576 Alzheimer mouse model.
28 *Proc Natl Acad Sci U A* **105**, 7863–7868 (2008).
29
30
31 142. Nakashiba, T., Young, J. Z., McHugh, T. J., Buhl, D. L. & Tonegawa, S. Transgenic
32 inhibition of synaptic transmission reveals role of CA3 output in hippocampal learning.
33 *Science* **319**, 1260–1264 (2008).
34
35
36 143. Russell, N. A., Horii, A., Smith, P. F., Darlington, C. L. & Bilkey, D. K. Long-Term
37 Effects of Permanent Vestibular Lesions on Hippocampal Spatial Firing. *J. Neurosci.*
38 **23**, 6490–6498 (2003).
39
40
41 144. Markus, E. J., Barnes, C. A., McNaughton, B. L., Gladden, V. L. & Skaggs, W. E.
42 Spatial information content and reliability of hippocampal CA1 neurons: effects of
43 visual input. *Hippocampus* **4**, 410–421 (1994).
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 145. Kentros, C. G., Agnihotri, N. T., Streater, S., Hawkins, R. D. & Kandel, E. R. Increased
4 attention to spatial context increases both place field stability and spatial memory.
5
6 *Neuron* **42**, 283–95 (2004).
7
8
9 146. Muzzio, I. A. *et al.* Attention Enhances the Retrieval and Stability of Visuospatial and
10 Olfactory Representations in the Dorsal Hippocampus. *PLoS Biol* **7**, e1000140 (2009).
11
12 147. Fenton, A. A. *et al.* Attention-like modulation of hippocampus place cell discharge. *J*
13 *Neurosci* **30**, 4613–4625 (2010).
14
15 148. Hok, V., Chah, E., Save, E. & Poucet, B. Prefrontal cortex focally modulates
16 hippocampal place cell firing patterns. *J Neurosci* **33**, 3443–3451 (2013).
17
18 149. Muir, G. M. & Taube, J. S. The Neural Correlates of Navigation: Do Head Direction
19 and Place Cells Guide Spatial Behavior? *Behav. Cogn. Neurosci. Rev.* **1**, 297–317
20 (2002).
21
22 150. Mizumori, S. J. Y., Miya, D. Y. & Ward, K. E. Reversible inactivation of the lateral
23 dorsal thalamus disrupts hippocampal place representation and impairs spatial learning.
24 *Brain Res.* **644**, 168–174 (1994).
25
26 151. Leutgeb, S. & Mizumori, S. J. Y. Excitotoxic Septal Lesions Result in Spatial Memory
27 Deficits and Altered Flexibility of Hippocampal Single-Unit Representations. *J.*
28 *Neurosci.* **19**, 6661–6672 (1999).
29
30 152. Cooper, B. G. & Mizumori, S. J. Y. Temporary Inactivation of the Retrosplenial Cortex
31 Causes a Transient Reorganization of Spatial Coding in the Hippocampus. *J. Neurosci.*
32 **21**, 3986–4001 (2001).
33
34 153. Nakazawa, K. *et al.* Requirement for hippocampal CA3 NMDA receptors in associative
35 memory recall. *Science* **297**, 211–218 (2002).
36
37 154. Nakazawa, K. *et al.* Hippocampal CA3 NMDA Receptors Are Crucial for Memory
38 Acquisition of One-Time Experience. *Neuron* **38**, 305–315 (2003).
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 155. Davis, S. *et al.* The Formation and Stability of Recognition Memory: What Happens
4 Upon Recall? *Front. Behav. Neurosci.* **4**, (2010).
5
6
7
8 156. Zhao, R., Fowler, S. W., Chiang, A. C. A., Ji, D. & Jankowsky, J. L. Impairments in
9 experience-dependent scaling and stability of hippocampal place fields limit spatial
10 learning in a mouse model of Alzheimer's disease. *Hippocampus* **24**, 963–978 (2014).
11
12
13
14 157. Mizumori, S. J., Lavoie, A. M. & Kalyani, A. Redistribution of spatial representation in
15 the hippocampus of aged rats performing a spatial memory task. *Behav Neurosci* **110**,
16 1006–1016 (1996).
17
18
19
20 158. Wilson, I. A. *et al.* Place cell rigidity correlates with impaired spatial learning in aged
21 rats. *Neurobiol Aging* **24**, 297–305 (2003).
22
23
24
25 159. Hok, V., Chah, E., Reilly, R. B. & O'Mara, S. M. Hippocampal dynamics predict
26 interindividual cognitive differences in rats. *J Neurosci* **32**, 3540–3551 (2012).
27
28
29
30 160. Thompson, L. T. & Best, P. J. Long-term stability of the place-field activity of single
31 units recorded from the dorsal hippocampus of freely behaving rats. *Brain Res.* **509**,
32 299–308 (1990).
33
34
35
36 161. Barnes, C. A., Suster, M. S., Shen, J. & McNaughton, B. L. Multistability of cognitive
37 maps in the hippocampus of old rats. *Nature* **388**, 272–275 (1997).
38
39
40
41 162. Yan, J., Zhang, Y., Roder, J. & McDonald, R. J. Aging effects on spatial tuning of
42 hippocampal place cells in mice. *Exp Brain Res* **150**, 184–193 (2003).
43
44
45
46 163. Ziv, Y. *et al.* Long-term dynamics of CA1 hippocampal place codes. *Nat. Neurosci.* **16**,
47 264–266 (2013).
48
49
50 164. Rubin, A., Geva, N., Sheintuch, L. & Ziv, Y. Hippocampal ensemble dynamics
51 timestamp events in long-term memory. *eLife* **4**, (2015).
52
53
54 165. Zinyuk, L., v S Kubik, Kaminsky, Y., Fenton, A. A. & s, J. B. Understanding
55 hippocampal activity by using purposeful behavior: place navigation induces place cell
56
57
58
59
60

- 1
2
3 discharge in both task-relevant and task-irrelevant spatial reference frames. *Proc Natl*
4
5 *Acad Sci U A* **97**, 3771–6 (2000).
6
7
8 166. Fenton, A. A. & Muller, R. U. Place cell discharge is extremely variable during
9
10 individual passes of the rat through the firing field. *Proc Natl Acad Sci U A* **95**, 3182–7
11
12 (1998).
13
14 167. Cho, Y. H., Giese, K. P., Tanila, H., Silva, A. J. & Eichenbaum, H. Abnormal
15
16 hippocampal spatial representations in alphaCaMKII^{T286A} and CREB^{alphaDelta}-
17
18 mice. *Science* **279**, 867–869 (1998).
19
20 168. Kulvicius, T., Tamosiunaite, M., Ainge, J., Dudchenko, P. & Wörgötter, F. Odor
21
22 supported place cell model and goal navigation in rodents. *J. Comput. Neurosci.* **25**,
23
24 481–500 (2008).
25
26
27 169. MacDonald, C. J., Carrow, S., Place, R. & Eichenbaum, H. Distinct Hippocampal Time
28
29 Cell Sequences Represent Odor Memories in Immobilized Rats. *J. Neurosci.* **33**, 14607–
30
31 14616 (2013).
32
33
34 170. Quirk, G. J., Muller, R. U. & Kubie, J. L. The firing of hippocampal place cells in the
35
36 dark depends on the rat's recent experience. *J. Neurosci.* **10**, 2008–2017 (1990).
37
38
39 171. Schultz, E. F. & Tapp, J. T. Olfactory control of behavior in rodents. *Psychol. Bull.* **79**,
40
41 21–44 (1973).
42
43 172. Prusky, G. T., West, P. W. R. & Douglas, R. M. Behavioral assessment of visual acuity
44
45 in mice and rats. *Vision Res.* **40**, 2201–2209 (2000).
46
47
48 173. Tanila, H. Wading pools, fading memories—place navigation in transgenic mouse
49
50 models of Alzheimer's disease. *Front. Aging Neurosci.* **4**, 11 (2012).
51
52 174. Hamilton, D. A., Akers, K. G., Weisend, M. P. & Sutherland, R. J. How do room and
53
54 apparatus cues control navigation in the Morris water task? Evidence for distinct
55
56
57
58
59
60

- 1
2
3 contributions to a movement vector. *J. Exp. Psychol. Anim. Behav. Process.* **33**, 100–
4
5 114 (2007).
6
7
8 175. Hamilton, D. A. *et al.* The relative influence of place and direction in the Morris water
9
10 task. *J. Exp. Psychol. Anim. Behav. Process.* **34**, 31–53 (2008).
11
12 176. Gallagher, M. & Pelleymounter, M. A. Spatial learning deficits in old rats: A model for
13
14 memory decline in the aged. *Neurobiol. Aging* **9**, 549–556 (1988).
15
16 177. Wishaw, I. Q., Metz, G. A. S., Kolb, B. & Pellis, S. M. Accelerated nervous system
17
18 development contributes to behavioral efficiency in the laboratory mouse: A behavioral
19
20 review and theoretical proposal. *Dev. Psychobiol.* **39**, 151–170 (2001).
21
22
23 178. Clark, B. *et al.* Lesions of the dorsal tegmental nuclei disrupt control of navigation by
24
25 distal landmarks in cued, directional, and place variants of the Morris water task. *Behav.*
26
27 *Neurosci.* **127**, 566–581 (2013).
28
29
30 179. Calton, J. L. *et al.* Hippocampal place cell instability after lesions of the head direction
31
32 cell network. *J. Neurosci. Off. J. Soc. Neurosci.* **23**, 9719–9731 (2003).
33
34 180. Yoder, R. M. & Taube, J. S. Head direction cell activity in mice: robust directional
35
36 signal depends on intact otolith organs. *J. Neurosci. Off. J. Soc. Neurosci.* **29**, 1061–
37
38 1076 (2009).
39
40
41 181. Schenk, F. Comparison of spatial learning in woodmice (*Apodemus sylvaticus*) and
42
43 hooded rats (*Rattus norvegicus*). *J. Comp. Psychol.* **101**, 150–158 (1987).
44
45 182. Lipp, H.-P. & Wolfer, D. P. Genetically modified mice and cognition. *Curr. Opin.*
46
47 *Neurobiol.* **8**, 272–280 (1998).
48
49
50 183. Wolfer, D. P. & Lipp, H.-P. Dissecting the Behaviour of Transgenic Mice: Is it the
51
52 Mutation, the Genetic Background, or the Environment? *Exp. Physiol.* **85**, 627–634
53
54 (2000).
55
56
57
58
59
60

- 1
2
3 184. Cressant, A., Besson, M., Suarez, S., Cormier, A. & Granon, S. Spatial learning in
4
5 Long-Evans Hooded rats and C57BL/6J mice: Different strategies for different
6
7 performance. *Behav. Brain Res.* **177**, 22–29 (2007).
8
9
10 185. Suarez, S. D. & Gallup Jr., G. G. An ethological analysis of open-field behavior in rats
11
12 and mice. *Learn. Motiv.* **12**, 342–363 (1981).
13
14 186. Crusio, W. E. & van Abeelen, J. H. The genetic architecture of behavioural responses to
15
16 novelty in mice. *Heredity* **56 (Pt 1)**, 55–63 (1986).
17
18 187. Tchernichovski, O., Benjamini, Y. & Golani, I. The dynamics of long-term exploration
19
20 in the rat. Part I. A phase-plane analysis of the relationship between location and
21
22 velocity. *Biol. Cybern.* **78**, 423–432 (1998).
23
24 188. Welker, W. I. ‘free’ versus ‘forced’ exploration of a novel situation by rats. *Psychol.*
25
26 *Rep.* **3**, 95–108 (1957).
27
28
29 189. Augustsson, H. & Meyerson, B. J. Exploration and risk assessment: a comparative study
30
31 of male house mice (*Mus musculus musculus*) and two laboratory strains. *Physiol.*
32
33 *Behav.* **81**, 685–698 (2004).
34
35 190. Wells, C. E., Krikke, B., Saunders, J., Whittington, A. & Lever, C. Changes to open
36
37 field surfaces typically used to elicit hippocampal remapping elicit graded exploratory
38
39 responses. *Behav Brain Res* **197**, 234–238 (2009).
40
41
42 191. Hussaini, S. A., Kempadoo, K. A., Thuault, S. J., Siegelbaum, S. A. & Kandel, E. R.
43
44 Increased Size and Stability of CA1 and CA3 Place Fields in HCN1 Knockout Mice.
45
46 *Neuron* **72**, 643–653 (2011).
47
48
49 192. Bittner, K. C. *et al.* Conjunctive input processing drives feature selectivity in
50
51 hippocampal CA1 neurons. *Nat. Neurosci.* **18**, 1133–1142 (2015).
52
53
54 193. Upchurch, M. & Wehner, J. M. Differences between inbred strains of mice in Morris
55
56 water maze performance. *Behav. Genet.* **18**, 55–68 (1988).
57
58
59
60

- 1
2
3 194. Roullet, P. & Lassalle, J. M. Genetic variation, hippocampal mossy fibres distribution,
4
5 novelty reactions and spatial representation in mice. *Behav. Brain Res.* **41**, 61–70
6
7 (1990).
8
9
10 195. Bertholet, J.-Y. & Crusio, W. E. Spatial and non-spatial spontaneous alternation and
11
12 hippocampal mossy fibre distribution in nine inbred mouse strains. *Behav. Brain Res.*
13
14 **43**, 197–202 (1991).
15
16 196. Crabbe, J. C., Wahlsten, D. & Dudek, B. C. Genetics of Mouse Behavior: Interactions
17
18 with Laboratory Environment. *Science* **284**, 1670–1672 (1999).
19
20 197. Freund, J. *et al.* Emergence of Individuality in Genetically Identical Mice. *Science* **340**,
21
22 756–759 (2013).
23
24
25
26

Figure captions

27
28
29 [Please insert any figure captions here]
30
31
32

Tables

33
34
35 [Please insert any tables here]
36
37
38

Further Reading/Resources

39
40
41 [Please insert any further reading/resources here]
42
43
44
45

Related WIREs Articles

DOI	Article title
10.1002/wcs.1164	Hippocampus
10.1002/wcs.1171	Development of spatial cognition
10.1002/wcs.1198	Psychology of spatial cognition
10.1002/wcs.1272	Parallel and convergent processing in grid cell, head-direction cell, boundary cell, and place cell networks

46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



Hippocampal place fields. Spatial navigation in rats and mice relies on the activity of hippocampal place cells. Place fields in the two species, however, differ in subtle ways that may reflect different navigation strategies. †

216x382mm (96 x 96 DPI)