

Species Distribution and Antifungal Susceptibility Profile of *Candida* spp. Bloodstream Isolates from Latin American Hospitals

Patrício Godoy, Iris Nora Tiraboschi*, Luiz Carlos Severo**, Beatriz Bustamante***, Belinda Calvo****, Leila Paula de Almeida, Daniel Archimedes da Matta, Arnaldo Lopes Colombo/+

Hospital São Paulo, Escola Paulista de Medicina, Universidade Federal de São Paulo, Rua Napoleão de Barros 740 - 7º andar, 04023-062 São Paulo, SP, Brasil *Hospital de Clínicas "José de San Martín", Universidad de Buenos Aires, Buenos Aires, Argentina ** Instituto Especializado em Pesquisa e Diagnóstico, Santa Casa de Porto Alegre, Porto Alegre, RS, Brasil ***Hospital "Cayetano Heredia", Lima, Perú ****Hospital Universitario de Maracaibo, Universidad del Zulia (Luz), Maracaibo, Venezuela

From March 1999 to March 2000, we conducted a prospective multicenter study of candidemia involving five tertiary care hospitals from four countries in Latin America. Yeast isolates were identified by classical methods and the antifungal susceptibility profile was determined according to the National Committee for Clinical Laboratory Standards microbroth assay method.

*During a 12 month-period we were able to collect a total of 103 bloodstream isolates of *Candida* spp. *C. albicans* was the most frequently isolated species accounting for 42% of all isolates. Non-*albicans* *Candida* species strains accounted for 58% of all episodes of candidemia and were mostly represented by *C. tropicalis* (24.2%) and *C. parapsilosis* (21.3%). It is noteworthy that we were able to identify two cases of *C. lusitanae* from different institutions. In our casuistic, non-*albicans* *Candida* species isolates related to candidemic episodes were susceptible to fluconazole. Continuously surveillance programs are needed in order to identify possible changes in the species distribution and antifungal susceptibility patterns of yeasts that may occurs after increasing the use of azoles in Latin American hospitals.*

Key words: candidemia - antifungal susceptibility - *Candida* spp.

A progressive increase in the frequency of candidemia has been observed, particularly among patients receiving antibiotics, immunosuppressive therapy, or parenteral nutrition, as well as among patients exposed to invasive medical procedures such as intravascular catheter, hemodialysis, and abdominal surgery (Lunel et al. 1999).

Despite *Candida albicans* is still considered the most frequently isolated species of candidemic patients, the emergence of non-*albicans* *Candida* species is clearly a concern. The resistance of non-*C. albicans* isolates to currently available antifungal drugs represents a major challenge for future empirical therapeutic and prophylactic strategies (Krcmery & Barnes 2002).

Due to the lack of information related to the epidemiology of candidemia existing in our region, we conducted a multicenter study to evaluate the species distribution and antifungal susceptibility patterns of *Candida* spp. bloodstream isolates in Latin American hospitals.

PATIENTS AND METHODS

Data collection and clinical specimens - From March 1999 to March 2000, we conducted a prospective multicenter study of candidemia involving five tertiary

care hospitals from four countries in Latin America: Hospital Universitario de Maracaibo, Universidad del Zulia (Luz), Maracaibo, Venezuela, Hospital "Cayetano Heredia", Lima, Peru, Hospital de Clínicas "José de San Martín", Universidad de Buenos Aires, Argentina, Instituto Especializado em Pesquisa e Diagnóstico, Santa Casa de Porto Alegre, Brazil and Hospital São Paulo, Escola Paulista de Medicina, Universidade Federal de São Paulo, Brazil.

The centers were requested to send the isolated strains to the reference laboratory (Laboratório Especial de Micologia, Division of Infectious Diseases, Unifesp, São Paulo, Brazil) for further identification and antifungal susceptibility testing. In order to be included in this study the patient must have had at least one blood culture positive for *Candida* spp., drawn from a peripheral vein, and clinical evidence of sepsis.

Yeast identification procedures - The purity and viability of yeast original cultures were checked by plating yeast colonies on CHROMagar *Candida* (CHROMagar Microbiology Paris, France). *C. albicans* isolates were identified if they exhibited green colonies on CHROMagar *Candida* and produced chlamydoconidia on corn meal-Tween 80 agar. Non-*albicans* *Candida* species isolates were identified on the basis of their micromorphology on corn meal-Tween 80 agar and biochemical tests evaluated by using the commercial system ID 32C, bioMérieux Marcy l'Etoile, France (Warren & Hazen 1995, Baumgartner et al. 1996).

In vitro susceptibility testing - Antifungal susceptibility tests were performed by using the broth microdilution assay according to the methodology recommended by

+Corresponding author. Fax: +55-11-5549.6585. E-mail: lemidipa@vento.com.br
Received 5 July 2002
Accepted 23 January 2003

the National Committee for Clinical Laboratory Standards (NCCLS), document M-27 A2. The following antifungal drugs, supplied by the manufacturers as pure standard compounds, were tested: amphotericin B, fluconazole, itraconazole and 5-flucytosine. Briefly, the medium used was RPMI-1640, with L-glutamine, without bicarbonate, and buffered at pH 7.0 with 0.165 M MOPS. The yeast inoculum suspension was prepared by using a spectrophotometer to obtain a final yeast concentration containing $0.5 - 2.5 \times 10^3$ cells/ml. The assays were incubated at 35°C for 48 h. The minimal inhibitory concentration (MIC) for amphotericin B was considered the lowest tested concentration able to prevent visible growth. MIC for azoles was considered as the lowest tested concentration with a significant reduction (approximately 50%) in growth compared to growth of a positive control (NCCLS 2002).

The breakpoints for azoles and 5-flucytosine MICs were those suggested by the NCCLS M27-A2. Due to a lack of consensus about the definition of MIC breakpoint for amphotericin B, arbitrary values were established according to values suggested by a previous study (Nguyen et al. 1998). Isolates with MICs ≤ 8 µg/ml for fluconazole, ≤ 4 µg/ml for 5-flucytosine, ≤ 0.125 µg/ml for itraconazole, and ≤ 1 µg/ml for amphotericin B were considered susceptible. Isolates with MICs between 16 and 32 µg/ml for fluconazole, and 0.25 to 0.5 µg/ml for itraconazole, were considered to have reduced (dose-dependent) susceptibility (DDS). MICs ≥ 64 µg/ml for fluconazole, ≥ 32 µg/ml for 5-flucytosine, ≥ 1 µg/ml for itraconazole, and ≥ 2 µg/ml for amphotericin B, were considered resistant (NCCLS M27-A2).

RESULTS

During a 12 month-period we were able to collect a total of 103 bloodstream isolates of *Candida* spp. *C. albicans* was the most frequently isolated species accounting for 42% of all isolates. As show in Table I, non-*albicans Candida* species strains accounted for 58% of

TABLE I
Etiologic agents of 103 candidemic episodes

Etiologic agent	N	%
<i>Candida albicans</i>	43	42
<i>Candida tropicalis</i>	25	24.2
<i>Candida parapsilosis</i>	22	21.3
<i>Candida glabrata</i>	8	7.7
<i>Candida guilliermondii</i>	3	2.9
<i>Candida lusitaniae</i>	2	1.9

all episodes of candidemia and were mostly represented by *C. tropicalis* (24.2%) and *C. parapsilosis* (21.3%).

Table II shows MIC₅₀ and MIC₉₀ values exhibited by the four antifungal drugs tested against the most frequently isolated *Candida* spp. strains. *C. albicans*, *C. tropicalis* and *C. parapsilosis* isolates were all susceptible to fluconazole and amphotericin B.

Only one *C. albicans* isolate was resistant to 5-flucytosine. Non-*albicans Candida* species isolates had higher azoles MIC values than *C. albicans* isolates but remained sensitive to low drug concentrations of amphotericin B. *C. glabrata* isolates had fluconazole MIC₅₀ and MIC₉₀ values of 2 and 4 µg/ml, respectively.

The two isolates of *C. lusitaniae* were susceptible to amphotericin B (MICs of 0.125 and 0.5 µg/ml), itraconazole (MICs of 0.03 and 0.015 µg/ml) and fluconazole (MICs of 0.25 and 0.5 µg/ml). Regarding to 5-flucytosine, one isolate was susceptible (MIC = 0.125 µg/ml) and the other was intermediate (MIC = 8 µg/ml).

Isolates of *C. guilliermondii* (3) were all susceptible to amphotericin B (MICs of 0.05 and 1 µg/ml), 5-flucytosine (MICs of 0.125 µg/ml) and fluconazole (MICs of 4 and 8 µg/ml). Otherwise, all three isolates had susceptibility dose dependent to itraconazole (MICs of 0.5 µg/ml).

TABLE II
Antifungal susceptibility profile of 98 ^a *Candida* bloodstream isolates

Species (no.)	Antifungal agent	Range	MIC (µg/ml)	
			MIC ₅₀	MIC ₉₀
<i>C. albicans</i> (43)	Amphotericin B	0.125-1	0.5	0.5
	Flucytosine	0.125-64	0.125	0.125
	Fluconazole	0.125-0.5	0.25	0.5
	Itraconazole	0.007-0.03	0.015	0.03
<i>C. tropicalis</i> (25)	Amphotericin B	0.25-1	0.5	1
	Flucytosine	0.125	0.125	0.125
	Fluconazole	0.125-0.5	0.25	0.5
	Itraconazole	0.007-0.06	0.015	0.03
<i>C. parapsilosis</i> (22)	Amphotericin B	0.25-1	0.5	1
	Flucytosine	0.125-1	0.125	0.125
	Fluconazole	0.125-2	0.5	2
	Itraconazole	0.007-0.06	0.03	0.03
<i>C. glabrata</i> (8)	Amphotericin B	0.5-1	0.5	0.5
	Flucytosine	0.125	0.125	0.125
	Fluconazole	2-8	2	4
	Itraconazole	0.015-0.5	0.125	0.5

a: isolates of *C. lusitaniae* (2) and *C. guilliermondii* (3) were not included on this Table.

DISCUSSION

Sandven (2000) reviewed 24 studies addressing episodes of candidemia in United States tertiary care hospitals and observed that the incidence of *C. albicans* isolates ranged from 38.8% to 79.4% of all episodes. The most prevalent non-*albicans Candida* species reported by the mentioned studies were *C. glabrata*, *C. tropicalis* and *C. parapsilosis*.

Indeed, in most recent studies of candidemia conducted in United States hospitals, *C. glabrata* accounted for 10% to 21% of all candidemic episodes (Pfaller et al. 1998, 1999, 2000, 2001).

Studies from Canada showed a different picture from the United States. In five out of six studies addressing the epidemiology of candidemia, the incidence of *C. albicans* isolates ranged from 53% to 74% and the non-*albicans Candida* species were correspondingly less prevalent. The high prevalence of *C. albicans* in Canada resembles the occurrence of this species in some European countries (Taylor et al. 1994, Karlowsky et al. 1997, Philips et al. 1997, Yamamura et al. 1999).

According to data from SENTRY and EORTC surveys, performed between 1997 and 1999, the incidence of *C. albicans* at European centers ranges from 49% (EORTC) to 59% (SENTRY) of all episodes of candidemia (Viscoli et al. 1999, Pfaller et al. 2001). Voss et al. (1996), after evaluating 626 episodes of candidemia in five Dutch's hospital observed that *C. albicans* isolates accounted for 60% of all episodes. However, in the same study, the incidence of non-*albicans Candida* species raised from 20 to 40% between 1987 and 1995.

In the present study we observed that *C. albicans* is the most frequent etiologic agent of candidemia in tertiary care hospitals from four Latin American countries, but non-*albicans Candida* species responded for almost 60% of all cases. In contrast with most series of candidemia from the United States and Europe, in which *C. glabrata* has been increasingly reported, this species represented only 7.5% of our candidemic episodes (Voss et al. 1996, Abi-Said et al. 1997). The low incidence of infections by primary azole resistant species of *Candida* may be related to the limited use of fluconazole in our hospitals due to cost considerations (Colombo et al. 1999). However, considering the lack of information regarding the consume of antifungal drugs in Latin American Hospitals, this hypothesis should be appropriately evaluated by further investigations.

In our casuistic, *C. tropicalis* was the second most frequently isolated species. The emergence of *C. tropicalis* as an important etiologic agent of candidemia in north hemisphere countries has been reported by several authors (Nguyen et al. 1996, Pfaller et al. 2000, 2001).

Rodero et al. (1999) identified *C. tropicalis* as the second most common isolate related to candidemic episodes during a multicenter study performed with the participation of 12 medical centers located in Argentina, where 89 cases were evaluated. In accordance with our findings, Colombo et al. (1999) reported that *C. parapsilosis* and *C. tropicalis* were the two most common non-*albicans Candida* species among 145 candidemic episodes documented

during a 18 month-period study including 6 different hospitals in Brazil. Costa et al. (2000), after evaluating 86 consecutive episodes of candidemia documented in a single tertiary care hospital from São Paulo, observed that *C. parapsilosis* and *C. tropicalis* responded for most of all non-*albicans Candida* isolates. Finally, Pfaller et al. (2001) reported that in Latin American hospitals participating of SENTRY, *C. parapsilosis* was the second most common *Candida* species causing blood stream infections.

Most of our *Candida* bloodstream isolates were susceptible to all antifungal drug tested. However, we had one strain of *C. albicans* and other of *C. lusitaniae* that were resistant and intermediate, respectively, to 5-FC. Isolates of *C. albicans* primarily resistant to 5-FC have been identified in different clinical materials by several authors, varying in frequency between 0% and 38% (Iwata 1992, Barchiesi et al. 2000). The relationship between resistance to 5-FC in *C. albicans* and serological type was investigated by Drouhet et al. (1975), who reported that the percentage of 5FC-resistant strains isolated from patients in Africa was higher than that recovered from patients in Europe. They suggested that the high incidence of resistance among the former strains was related to the predominance of serotype B isolates (Drouhet et al. 1975).

We were surprised by the low MIC values exhibited by our *C. glabrata* isolates. Several publications from north hemisphere countries have shown that *C. glabrata* isolates usually exhibit high fluconazole MIC values (Voss et al. 1996, Rex et al. 1997, Pfaller et al. 2001). According to SENTRY data, *C. glabrata* blood stream isolates have MIC₅₀ and MIC₉₀ values ranging from 4 to 16 µg/ml and 16 to 32 µg/ml, respectively. In our series, all *C. glabrata* isolates had MICs values ≤ 8. This aspect may be also related to the higher use of fluconazole in Europe and United States compared to Latin America.

It is noteworthy that we were able to identify two cases of candidemia due to *C. lusitaniae* (one reported in Brazil and the other in Argentina) and three cases of *C. guilliermondii*. Both pathogens have been rarely reported in different series of candidemia. Data from the Sentry Program (Pfaller et al. 2001) did not mention any specific data related to the identification of *C. lusitaniae* and *C. guilliermondii* candidemic episodes.

Luzzati et al. (2000) were able to identify three cases of invasive infection due to *C. lusitaniae* among 189 candidemic episodes reported in the University Hospital of Verona, during the period of 1992 and 1997. In Latin America, Colombo et al. (1999) reported only three cases of candidemia due *C. guilliermondii* among 145 episodes reported on six different tertiary care hospitals in Brazil.

Some authors advocate that *C. lusitaniae* isolates may present innate resistance to amphotericin B (Pfaller et al. 1994, Hadfield et al. 1997). However, the amphotericin B MIC results exhibited by both isolates of *C. lusitaniae* from our series were 0.125 and 0.5 µg/ml respectively. This aspect may be related to the limitations of the methodology we used to recognize isolates truly resistant to amphotericin B. The NCCLS methodology yields a range of MICs that span only three to four twofold serial dilutions, what make difficult a reliable discrimination between sus-

ceptible and resistant isolates (Rex et al. 1995). Considering the mentioned limitation of the NCCLS methodology, some authors have suggested to use the Etest with standardized RPMI medium supplemented with 2% glucose as the most sensitive and reliable means for detecting amphotericin B resistance (Wanger et al. 1995, Peyron et al. 2001).

Although *C. albicans* continues to account for approximately one-half of all episodes of candidemia reported worldwide, its frequency may vary widely from institution to institution. We were able to demonstrate that in South America most non-*albicans* *Candida* species are represented by *C. tropicalis* and *C. parapsilosis* isolates still susceptible to amphotericin B and fluconazole. However, continuously surveillance programs are needed in order to identify possible changes in the species distribution and antifungal susceptibility patterns of yeasts, particularly after increasing the use of azoles in Latin American hospitals.

REFERENCES

- Abi-Said D, Anaisse E, Uzun O, Raad I, Pinzowski H, Vartivarian S 1997. The epidemiology of hematogenous candidiasis caused by different *Candida* species. *Clin Infect Dis* 24: 1122-1128.
- Barchiesi F, Arzeni D, Caselli F, Scalesi G 2000. Primary resistance to flucytosine among clinical isolates of *Candida* spp. *J Antimicrob Chemother* 25: 408-409.
- Baumgartner C, Freydiere AM, Gille Y 1996. Direct identification and recognition of yeast species from clinical material by using Albicans ID and CHROMagar *Candida* plates. *J Clin Microbiol* 34: 454-456.
- Colombo AL, Nucci M, Salomão R, Branchini ML, Richtmann R, Derossi A, Wey BS 1999. High rate of non-*albicans* candidemia in Brazilian tertiary care hospitals. *Diagn Microbiol Infect Dis* 34: 281-286.
- Costa SF, Marinho I, Araujo EA, Manrique AE, Medeiros EA, Levin AS 2000. Nosocomial fungaemia: a 2-year prospective study. *J Hosp Infect* 45: 69-72.
- Drouhet E, Mercier-Soucy L, Montplaisir S 1975. Sensitivity and resistance of pathogenic yeasts to 5-fluoropyrimidines. Relation between the phenotypes of resistance to 5-fluorocytosine, the serotype of *Candida albicans* and the ecology of various species of *Candida* of human origin. *Ann Microbiol* 126: 25-39.
- Ghannoum M, Rex JH, Galgiani JN 1996. Susceptibility testing of fungi: current status of correlation in vitro data with clinical outcome. *J Clin Microbiol* 34: 489-495.
- Hadfield TL, Smith MB, Winn RE, Rinaldi MG, Guerra C 1997. Mycoses caused by *Candida lusitanae*. *Rev Infect Dis* 9: 1006-12.
- Iwata K 1992. Drug resistance in human pathogenic fungi. *Eur J Epidemiol* 8: 407-421.
- Karlowsky JA, Zhanel GG, Klym KA, Hoban DJ, Kabani AM 1997. Candidemia in a Canadian tertiary care hospital from 1976 to 1996. *Diagn Microbiol Infect Dis* 29: 5-9.
- Krcmery V, Barnes AJ 2002. Non-*albicans* *Candida* spp. causing fungaemia: pathogenicity and antifungal resistance. *J Hosp Infect* 50: 243-260.
- Lunel F M V, Meis JFGM, Voss A 1999. Nosocomial fungal infections: candidemia. *Diagn Microbiol Infect Dis* 34: 213-220.
- Luzzati R, Amalfitano G, Lazzarini L, Soldani F, Bellino S, Solbiati M, Danzi MC, Vento S, Todeschini G, Vivenza C, Concia E 2000. Nosocomial candidemia in non-neutropenic patients at an Italian tertiary care hospitals. *Eur J Clin Microbiol Infect Dis* 19: 602-607.
- NCCLS-National Committee for Clinical Laboratory Standards 2002. Publication M27-A2: Reference method for broth dilution antifungal susceptibility testing of yeasts; approved standard, Wayne, PA: NCCLS 22: 1-29.
- Nguyen MH, Clancy CJ, Yu VL, Yu YC, Morris AJ, Snyder DR, Sutton DA, Rinaldi MG 1998. Do in vitro susceptibility data predict microbiologic response to amphotericin B? Results of a prospective study of patients with *Candida* fungemia. *J Infect Dis* 177: 425-430.
- Nguyen MH, Peacock JE, Morris AJ, Tanner DC, Nguyen ML, Snyderman DR, Wagener MN, Rinaldi MG, Yu VL 1996. The changing face of candidemia: emergence of non-*Candida albicans* species and antifungal resistance. *Am J Med* 100: 617-623.
- Peyron F, Favel A, Nguyen AM, Gilly M, Regli P, Bolmström A 2001. Improved detection of amphotericin B-resistant isolates of *Candida lusitanae* by Etest. *J Clin Microbiol* 39: 339-342.
- Pfaller MA, Diekema DJ, Jones RN, Sader HS, Fluit AC, Hollis RJ, Messer AS 2001. International surveillance of bloodstream infections due to *Candida* species: frequency of occurrence and in vitro susceptibilities to fluconazole, ravuconazole, and voriconazole of isolates collected from 1997 through 1999 in the SENTRY Antimicrobial Surveillance Program. *J Clin Microbiol* 39: 3254-3259.
- Pfaller MA, Jones RN, Doern GV, Fluit AC, Verhoef J, Sader HS, Messer SA, Houston A, Coffman S, Hollis RJ 1999. International surveillance of blood stream infections due to *Candida* species in the European SENTRY program: species distribution and antifungal susceptibility including the investigational triazole and echinocandin agents. *Diagn Microbiol Infect Dis* 35: 19-25.
- Pfaller MA, Jones RN, Doern GV, Sader HS, Messer SA, Houston A, Coffman S, Hollis RJ 2000. Bloodstream infections due to *Candida* species: SENTRY Antimicrobial Surveillance Program in North America and Latin America, 1997-1998. *Antimicrob Agents Chemother* 44: 747-751.
- Pfaller MA, Jones RN, Messer SA, Edmond MB, Wenzel RP 1998. National surveillance of nosocomial blood stream infections due to species of *Candida* other than *Candida albicans*: frequency of occurrence and antifungal susceptibility in the SCOPE program. *Diagn Microbiol Infect Dis* 30: 121-129.
- Pfaller MA, Messer SA, Hollis RJ 1994. Strain delineation and antifungal susceptibilities of epidemiologically related and unrelated isolates of *Candida lusitanae*. *Diagn Microbiol Infect Dis* 20: 127-133.
- Phillips P, Shafran S, Garber G, Rotstein C, Smaill F, Fong I, Salit I, Miller M, Williams K, Conly JM, Singer J, Ioannou S 1997. Multicenter randomized trial of fluconazole versus amphotericin B for treatment of candidemia in non-neutropenic patients. *Eur J Clin Microbiol Infect Dis* 16: 337-345.
- Rex JH, Cooper CR, Merz WG, Galgiani JN, Anaissie EJ 1995. Detection of amphotericin B-resistant *Candida* isolates in a broth-based system. *Antimicrob Agents Chemother* 39: 906-909.
- Rex JH, Pfaller MA, Galgiani JN, Bortlett MS, Espinel-Ingroff A, Ghannoum MA, Lancaster M, Odds FC, Rinaldi MG, Walsh TJ, Barry AL 1997. Development of interpretive breakpoints for antifungal susceptibility testing: conceptual framework and analysis of in vitro correlation data for fluconazole, itraconazole and *Candida* infections. *Clin Infect Dis* 24: 235-247.
- Rodero L, Davel, G, Cordoba S, Soria M, Canteros C,

- Hochenfellner F 1999. Estudio multicéntrico sobre candidiasis nosocomial en la República Argentina. *Rev Argent Microbiol* 31: 114-119.
- Sandven P 2000. Epidemiology of candidemia. *Rev Iberoam Micol* 17: 73-81.
- Taylor GD, Buchanan-Chell M, Kirkland T, McKenzie M, Wiens R 1994. Trends and sources of nosocomial fungemia. *Mycoses* 37: 187-190.
- Viscoli C, Girmenia C, Marinus A, Collette L, Martino P, Vandercam B, Doyen C, Lebeau B, Spence D, Kremery V, De Pauw B, Meunier F 1999. Candidemia in cancer patients: a prospective, multicenter surveillance study by the Invasive Fungal Infection Group (IFIG) of the European Organization for Research and Treatment of Cancer (EORTC). *Clin Infect Dis* 28: 1071-1079.
- Voss A, Kluytamans JA, Koeleman JG, Spanjaard L, Vandenbroucke-Grauls CMJE, Verbrugh HA, Voss MC, Weersink AYL, Hoogkamp-Korstanje JAA, Meis JFGM 1996. Occurrence of yeast bloodstream infections between 1987 and 1995 in five Dutch university hospitals. *Eur J Clin Microbiol Infect Dis* 15: 909-912.
- Wanger A, Mills K, Nelson PW, Rex JH 1995. Comparison of Etest and national committee for clinical laboratory standards broth microdilution method for antifungal susceptibility testing: enhanced ability to detect amphotericin B-resistant *Candida* isolates. *Antimicrob Agents Chemother* 39: 2520-22.
- Warren NG, Hazen KC 1995. *Candida*, *Cryptococcus* and other yeasts of medical importance. In PR Murray, *Manual of Clinical Microbiology*, ASM, Washington, p. 723-737.
- Yamamura DL, Rotstein C, Nicolle LE, Ionnou S 1999. Candidemia at selected Canadian sites: results from the Fungal Diseases Registry of the Canadian Infectious Disease Society. *CMAJ* 160: 493-499.