

Spectral Properties of Leaves Deficient in Iron, Sulfur, Magnesium, and Manganese

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ABSTRACT

In crop plants, deficiency of an essential element may drastically reduce growth rate and yield. Research on the use of leaf spectral properties in the detection of crop mineral deficiency is needed. The objective of this study was to examine the effects of Fe, S, Mg, and Mn deficiency on reflectance (R), absorptance (A), and transmittance (T) spectra of barley (*Hordeum vulgare* L.), wheat (*Triticum aestivum* L.), corn (*Zea mays* L.), and sunflower (*Helianthus annuus* L.) leaves. Plants were grown in the greenhouse in nutrient solution. Chlorophyll (Chl), Fe, S, Mg, and Mn concentrations and spectral properties were determined on the youngest fully expanded leaf when deficiency symptoms were clearly manifested. In all species, mineral deficiency affected leaf concentration of the deficient element and also of other elements. Nutrient deficiency reduced Chl concentration and A, and increased R and T. Iron deficiency severely affected all species, and corn was the species most sensitive to all deficiencies. Reflectance, A, and T spectra of leaves were correlated with leaf Chl concentration. Our results suggested that all nutritional deficiencies reduce leaf Chl concentration, and subsequently this reduction increases leaf R and T, decreases leaf A, and shortens the red-edge position, defined as the inflection point that occurs in the rapid transition between red and near-infrared. Modifications in leaf spectral properties were not characteristic of nutrient deficiency, but were always observed in the same wavelengths.

DEFICIENCY OF AN ESSENTIAL ELEMENT may drastically reduce growth rate and yield in crop plants. Mineral deficiency causes visible abnormalities in pigmentation, size, and shape of leaves and leaf photosynthetic rate, also leading to the appearance of various other symptoms. Leaf photosynthetic rate is linked to the amount of absorbed radiation, which depends on incident radiation and leaf absorptance. Absorptance is affected by leaf external and internal reflectance and by leaf pigment content, essentially represented by chlorophyll (Maas and Dunlap, 1989).

Several authors have found empirical relationships between leaf spectral properties and leaf morphological and physiological conditions, including leaf thickness (Gausman and Allen, 1973), presence of pubescence (Nielsen et al., 1984), N content (Walburg et al., 1982), water content (Gausman et al., 1971), and chlorophyll concentration (Gausman, 1982; Ercoli et al., 1993). Other studies have documented spectral and morphological changes in plants grown with abnormal concentrations of mineral elements in the nutrient solution. Al-Abbas et al. (1974) showed that corn plants growing under N-

Mg-, S-, K-, P-, and Ca-deficient conditions had higher leaf R and T and lower A in the visible region of the spectrum (400–700 nm) than normal plants. Milton et al. (1989, 1991) studied spectral reflectance in *Hosta ventricosa* Co-, Ni-, and Zn-deficient plants, and in soybean [*Glycine max* (L.) Merr.] As-, P-, and Se-deficient plants. They showed that, with lower than normal Co, Ni, Zn, As, and P content, R increased in the 500- to 650-nm portion of the spectrum and the red edge (the inflection point that occurs in the rapid transition between red and near-infrared; Horler et al., 1983) shifted to shorter wavelengths. In contrast, Se-dosed soybean plants displayed virtually opposite results. Furthermore, in studies by Thomas and Oerther (1972) on sweet pepper (*Capsicum annuum* L.) it was found that leaf R increased with decreasing leaf N content. Adams et al. (1993), reporting on soybean, showed that a decrease in leaf Mn concentration led to an increase in R and shifted the red-edge position to shorter wavelengths. However, use of spectral properties as a diagnostic tool in nutrient deficiency diagnosis requires further understanding of the relationship between spectral properties and nutrient concentration in plant tissue. Aspects of this relationship have been described for corn with varying N fertilizer levels (Ercoli et al., 1993; Blackmer et al., 1994).

Our objective was to examine the effects of Fe, S, Mg, and Mn deficiency on mineral and Chl concentrations and on R, A, and T spectra of attached leaves of barley, wheat, corn, and sunflower. In addition, this study assessed whether nutrient deficiency produces changes in detailed curve shape that could be used diagnostically for detection of the deficiency itself.

MATERIALS AND METHODS

Research was carried out in 1993 at the Department of Agronomy and Agroecosystem Management (Agronomia e Gestione dell'Agro-Ecosistema), University of Pisa, Italy. 'Aura' distichous barley (*Hordeum vulgare* L.), 'Pandas' winter wheat (*Triticum aestivum* L.), hybrid Laurus corn (*Zea mays* L.), and hybrid Oleica sunflower (*Helianthus annuus* L.) were grown on nutrient solution in the greenhouse. These four species, which are among the most widely cultivated crops in Italy, were chosen with the aim of ascertaining the effects of mineral deficiency in leaves of plants with similar or different botanical characteristics. For each species, treatments consisted of five different nutrient culture solutions, as follows: complete solution, minus Fe, minus S, minus Mg, and minus Mn.

Three seeds per species were sown in plastic pots (6 cm deep, 6 cm diam.) filled with agriperlite. The sowing date was 5 January for barley and wheat and 5 March for corn and sunflower. After germination, seedlings were thinned to

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Table 1. Phenological stage and days after emergence (DAE) of visible deficiency symptoms, degree of chlorosis (DOC), and nodal position of measured leaves (ML) from barley, wheat, corn, and sunflower plants grown under normal and Fe-, S-, Mg-, and Mn-deficient conditions.†

Crop	Deficiency	Stage of deficiency evidence		DAE	ML	DOC
		Reference scale‡	Description			
Barley	Normal-1§	32	2nd node detectable	75	5	None
	Fe	32	2nd node detectable	75	5	Moderate
	Normal-2	43	Boots just visibly swollen	85	6	None
	S	43	Boots just visibly swollen	85	6	Slight
	Mg	43	Boots just visibly swollen	85	6	Slight
	Mn	43	Boots just visibly swollen	85	6	Slight
Wheat	Normal-1	23	Main shoot and 3 tillers	63	4	None
	Fe	23	Main shoot and 3 tillers	63	4	Moderate
	Normal-2	43	Boots just visibly swollen	87	6	None
	S	43	Boots just visibly swollen	87	6	Slight
	Mg	43	Boots just visibly swollen	87	6	Slight
	Mn	43	Boots just visibly swollen	87	6	Slight
Corn	Normal-1	1	4 leaves	29	3	None
	Fe	1	4 leaves	29	3	Severe
	S	1	4 leaves	29	3	Moderate
	Normal-2	1	5 leaves	45	4	None
	Mg	1	5 leaves	45	4	Moderate
	Mn	1	5 leaves	45	4	Moderate
Sunflower	Normal-1	V4	4 leaves	31	3	None
	Fe	V4	4 leaves	31	3	Severe
	Normal-2	V10	10 leaves	47	8	None
	S	V10	10 leaves	47	8	Slight
	Normal-3	R1	Floral bud	55	11	None
	Mg	R1	Floral bud	55	11	Moderate
Mn	R1	Floral bud	55	11	Slight	

† For all species and deficiency treatments (with one exception), initial symptoms of mineral deficiency occurred on the youngest leaf; for Mg deficiency on sunflower, initial symptoms occurred on the oldest leaf.

‡ Phenological stages are according to Zadoks et al. (1974) for barley and wheat, Hanway (1963) for corn, and Schneiter and Miller (1981) for sunflower.

§ Normal-1, -2, and -3 differ only by phenological stage (matching that of the nutrient-deficient plants), not in nutrient solution.

one per pot and 160 pots were placed on 0.7 m² (1.2 by 0.6 m) plastic trays containing the nutrient solutions, with one species per tray. Basic composition of the nutrient solution followed Clark (1982) (in mg L⁻¹): NO₃-N, 321; Ca, 302; K, 283; Cl, 65; S, 58.5; NH₄-N, 39; Mg, 37.8; Na, 4.56; Fe, 4; P, 2; Mn, 0.974; B, 0.536; Zn, 0.3; Cu, 0.076; Mo, 0.155. Growth solution was changed every 3 d and pH was adjusted daily to 6.5 with HNO₃ or NaOH. Deionized water was used throughout the experiment and added as needed to maintain solution volume.

All measurements were performed on the youngest fully expanded leaf when leaf mineral deficiency symptoms were clearly manifested. For each species, three leaves (one per plant) randomly selected from each nutrient solution deficiency were compared with three leaves from the control at the same phenological stage as the deficient plants, so that the comparison was made on leaves in the same nodal position. According to Berg et al. (1993), degree of chlorosis of the measured leaf was visually rated as follows: none, slight (10 to 30% interveinal chlorosis), moderate (30 to 100% interveinal chlorosis and veins green), severe (entire leaf chlorotic), and very severe (leaf nearly white with necrotic tip).

Reflected and transmitted radiation spectra of adaxial (upper) surfaces of attached leaves were measured using a LI-COR (Lincoln, NE) portable spectroradiometer, Model LI-1800, connected to an external integrating sphere by means of a quartz fiber-optic probe. Measurements were taken over the wavelength range from 400 to 1100 nm at a scanning interval of 1 nm. The integrating sphere included a 10-W glass halogen lamp as radiation source and a pressed barium sulfate (BaSO₄) reference standard. For all species, measurements were taken from an area measuring roughly 1.8 cm² (sample port) situated at the center of the right-hand leaf lamina. Reflectance and transmittance were calculated as the ratio of reflected radiation from and transmitted radiation through the sample surface

to reflected and transmitted radiation from the standard and expressed in percent. Absorbance was computed as: $A = 100 - (R + T)$. The inflection point that occurs in the rapid transition between red and near-infrared, termed the *red edge*, is thought useful in estimating leaf pigment content, discriminating crop type and maturity, and estimating the severity of stress and crop condition (Horler et al., 1983). First derivatives of the R, A, and T individual spectral curves between 678 and 740 nm were computed, and the inflection points (maxima on the first-derivative spectra) were used to define the position of the edge. The derivative spectrum was calculated by fitting a third-order polynomial into the data with the least squares method as suggested by Savitzky and Golay (1964).

Immediately following spectroradiometric measurements, leaf area, fresh and dry weight, and Chl *a* and *b* concentrations were determined. Leaf area was measured using an image analyzer (Leica Quantimet 500). Ten 50-mm² disk samples were collected from each leaf for Chl determination. Disks were placed in a test tube, stoppered and deep-frozen (using dry ice) for transport to the laboratory, and then stored at -18° C until Chl analysis. Absorbance of *N,N*-dimethylformamide leaf sample extract was measured at 664 and 647 nm on a spectrophotometer (Lambda 6 UV/VIS, Perkin-Elmer, Norwalk, CT) using cuvettes of 10-mm path length. Chlorophyll *a* and *b*, expressed in moles on leaf area basis, were determined according to the Moran (1982) formulae.

Leaves from 60 plants (one leaf per plant) were combined in three samples (replications) for Fe, S, Mg, and Mn analysis. Samples were ground to pass through a 40-mesh stainless steel screen. For Fe, Mg, and Mn analysis, samples (0.5 g) were wet-ashed by overnight predigestion in 14 mL of concentrated HNO₃-HClO₄ mixture (5:2 v/v basis) and digestion was completed in an aluminum block heater at 205° C (Ohki, 1984). Ashed extracts were brought to 25 mL volume and Fe, Mg, and Mn were determined by atomic absorption spectrometry

Table 2. Iron, S, Mg, and Mn concentration of the youngest fully expanded leaf of barley, wheat, corn, and sunflower plants grown under normal and Fe-, S-, Mg-, and Mn-deficient conditions. For each species and deficiency, comparison was made between leaves grown under normal and mineral-deficient conditions.

Crop	Mineral concentration (by growth medium treatment)							
	Fe		S		Mg		Mn	
	Normal	Deficient	Normal	Deficient	Normal	Deficient	Normal	Deficient
	Iron, $\mu\text{mol m}^{-2}$							
Barley	72	41*	94	47*	94	63*	94	77
Wheat	60	39*	85	76	85	71*	85	60*
Corn	82	32*	82	28*	115	40*	115	44*
Sunflower	65	39*	142	90*	179	96*	179	117*
	Sulfur, mmol m^{-2}							
Barley	8.9	9.0	10.9	8.9*	10.9	10.9	10.9	8.7*
Wheat	6.0	6.1	10.4	8.5*	10.4	10.4	10.4	8.0*
Corn	6.2	3.9*	6.2	4.9*	6.2	3.7*	6.2	3.5*
Sunflower	6.4	5.5	11.3	6.3*	11.4	7.0*	11.4	6.6*
	Magnesium, mmol m^{-2}							
Barley	2.5	3.4*	2.8	1.9*	2.8	0.9*	2.8	2.0*
Wheat	2.5	3.4*	3.3	2.4*	3.3	0.7*	3.3	1.8*
Corn	1.8	1.6	1.8	1.6	2.4	0.9*	2.4	1.6*
Sunflower	2.4	2.5	4.9	2.9*	3.3	1.4*	3.3	4.4*
	Manganese, $\mu\text{mol m}^{-2}$							
Barley	11	20*	13	6*	13	12	13	5*
Wheat	15	26*	17	8*	17	5*	17	2*
Corn	38	27*	38	27*	50	39*	50	25*
Sunflower	17	50*	33	13*	42	30*	42	7*

* Significant at the 0.05 probability level (*F*-test).

(Zeiss FMD3 spectrophotometer). For total S assay, following Hafez et al. (1991), 0.2 g of plant material was digested with 2 mL of concentrated HNO_3 containing $1.25 \text{ g L}^{-1} \text{ MgO}$ and 1 mL of 70% (v/v) HClO_4 and subsequently evaporated to dryness. Ashed extract was supplemented with 5 mL of 250 mM NaOH and brought to 50 mL volume. The liquid was filtered through a $0.45\text{-}\mu\text{m}$ filter and the filtrate was analyzed for $\text{SO}_4\text{-S}$ using an ion-chromatograph.

Since in the various species deficiency symptoms were detected at different stages and on leaves of different nodal position, analysis of variance was performed separately for each species and growth-medium deficiency to test the significance of the difference between control and individual mineral deficiency. Statistical analysis on spectral data was undertaken as bands of 10 nm width in the spectral range from 400 to 1100 nm.

Analysis of correlation was performed to assess the relationships between leaf spectral properties at the 429-, 555-, 678-, and 700-nm wavelengths and in the visible region and leaf Chl *a* and *b* concentration. These wavelengths were chosen because at 429 and 678 nm maximum *in vivo* Chl absorption occurred and at 555 and 700 nm the highest variations produced by mineral deficiencies were recorded. The model used to represent the relationship between leaf spectral properties (*y*) and Chl was $y = a[1 + b \exp(-kx)]$, where *x* is leaf Chl concentration expressed in moles per unit area (Ercoli et al., 1993). In the model, the expression $\exp(-kx)$ represents radiation attenuation within the leaf that decreases exponentially with increasing leaf thickness (*x*) on the basis of a specific extinction coefficient *k*. In the leaf, chlorophyll is the substance having the greatest effect on *A* in the visible region; therefore, the assumption was made that its content, expressed in moles per unit leaf area (*x*), would adequately represent leaf thickness. In the model, the coefficient *b* represents the percentage increase (for R and T) or decrease (for A) of the value of *a* with $x = 0$ and (*ab*) represents the range of values assumed by *y* with progressive increase of *x* from 0 to infinity ($ab = y_{\text{max}} - y_{\text{min}}$).

RESULTS AND DISCUSSION

Leaf Symptoms of Mineral Deficiency

The degree of chlorosis and the stage at which leaf symptoms of mineral deficiency were detected varied among species and mineral deficiencies (Table 1). Corn was found to be the most sensitive to all deficiencies, as in this plant symptoms were evident as early as the 4- to 5-leaf stage. In all species, Fe- and S-deficiency symptoms occurred in early growth stages while Mg- and Mn-deficiency symptoms occurred at early stages in corn and later in barley, wheat, and sunflower. Iron deficiency produced moderate or severe chlorosis at early stages in all species. Symptoms of Mg deficiency were slight in barley and wheat leaves and moderate in corn and sunflower leaves; symptoms of Mn- and S-deficiencies were slight in barley, wheat, and sunflower leaves and moderate in corn leaves.

Leaf Mineral Concentration

In all species, Fe, S, Mg, and Mn deficiency in growth medium greatly decreased leaf concentration of the respective deficient element, although not all differences were significant (Table 2). Accumulation of other elements in leaves also varied with the different availability of any of the elements, indicating that deficient supply of any given mineral element may limit uptake of other elements, but correlations among leaf concentrations of elements were not observed.

Deficiency of a given mineral element may produce biochemical disorders that modify accumulation of other elements, thereby increasing or decreasing concentration as compared with normal nutritional conditions. Thus, deficiency symptoms and morphological and physiologi-

cal modifications may not be produced by or characteristic of one deficient element, but may instead be produced by combined deficiency of more than one element, although all reduced mineral concentrations are caused by deficiency of only one element in the growth medium. In corn, for example, S deficiency caused a greater percent reduction in leaf Fe concentration (by 66%) than in leaf S concentration (by 21%).

Leaf Chlorophyll Concentration

Growth-medium mineral deficiencies decreased leaf Chl *a* and *b* concentration of all species by a percentage that differed according to species and mineral, ranging from 10 to 80% of the corresponding control (Table 3). Corn was the crop most sensitive to nutrient deficiency, as Chl *a* concentration of leaves from plants grown under Fe-, Mg-, and Mn-deficient conditions was 22% of the control, and that under S-deficient conditions was approximately 50%. In barley and wheat, the highest reduction in Chl was induced by Fe, followed by S and Mg, while Mn produced the least effect. Sunflower leaf Chl *a* concentration decreased to approximately 60% of normal values in plants grown under Mg-, Mn-, and S-deficient conditions and to 23% in Fe-deficient plants. Mineral deficiencies decreased leaf Chl *b* concentration differently from Chl *a* concentration.

The leaf Chl *a/b* ratio increased significantly only in leaves of Mg-deficient wheat and decreased significantly in leaves of corn with all deficiencies, of sunflower with Fe deficiency, and of barley and wheat with S deficiency.

No correlation was observed between leaf Chl *a*, and *b* concentration and leaf Fe, Mg, Mn, and S concentration. This result indicates that, although in all species growth-medium mineral deficiencies reduced leaf Chl concentration, such a reduction is not equal among species, because of differences in microelement requirements among the various species.

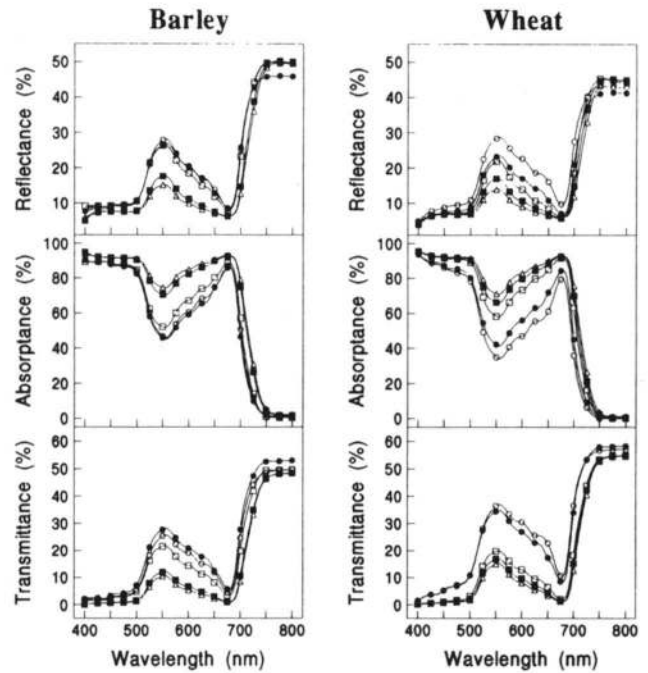


Fig. 1. Spectra of R, A, and T of the youngest fully expanded leaf of barley and wheat. Legend: open triangles, normal plants; open circles, Fe; solid circles, S; open squares, Mg; solid squares, Mn.

Reflectance, Absorbance, and Transmittance

Figures 1, 2, and 3 illustrate R, A, and T spectra between 400 and 800 nm recorded for all species and growth media. In all cases, A decreased markedly from 400 to a minimum at 555 nm, then increased from 555 to 678 nm. Through the near-infrared (NIR) region, A decreased to a minimum at 750 nm. Beyond 750 nm, A remained practically constant at about 2%. Reflectance and T spectra were opposite to that of A, with a local maximum at 555 and 750 nm and a local minimum at 500 and 678 nm. Values of R, A, and T at 555 and 700 nm were similar and, as shown also by Chappelle et al.

Table 3. Chlorophyll *a* and *b* concentration and Chl *a/b* ratio of the youngest fully expanded leaf of barley, wheat, corn, and sunflower plants grown under normal and Fe-, S-, Mg-, and Mn-deficient conditions. For each species and deficiency, comparison was made between leaves grown under normal and mineral deficient conditions.

Crop	Chlorophyll <i>a</i> and <i>b</i> concentration and ratio (by growth medium treatment)							
	Fe		S		Mg		Mn	
	Normal	Deficient	Normal	Deficient	Normal	Deficient	Normal	Deficient
	Chl <i>a</i> , $\mu\text{mol m}^{-2}$							
Barley	457	165*	448	127*	448	196*	448	357*
Wheat	401	99*	433	148*	433	274*	433	354*
Corn	259	28*	259	128*	437	56*	437	97*
Sunflower	256	58*	408	242*	443	210*	443	286*
	Chl <i>b</i> , $\mu\text{mol m}^{-2}$							
Barley	149	50*	139	73*	139	62*	139	115*
Wheat	113	31*	137	72*	137	77*	137	112*
Corn	92	38*	92	66*	131	27*	131	39*
Sunflower	81	27*	149	90*	160	72*	160	94*
	Chl <i>a/b</i>							
Barley	3.1	3.3	3.2	1.7*	3.2	3.1	3.2	3.1
Wheat	3.6	3.2*	3.2	2.1*	3.2	3.6*	3.2	3.2
Corn	2.8	0.7*	2.8	1.9*	3.3	2.1*	3.3	2.5*
Sunflower	3.2	2.1*	2.7	2.7	2.8	2.9	2.8	3.0

* Significant at the 0.05 probability level (*F*-test).

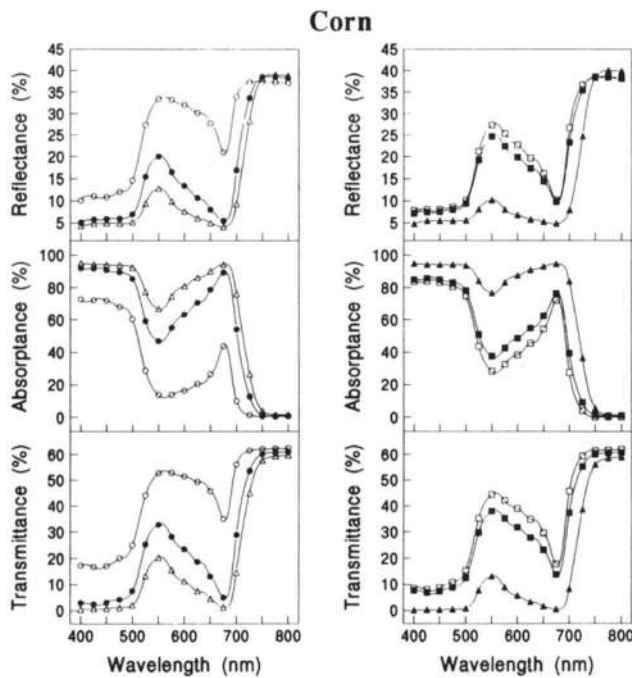


Fig. 2. Spectra of R, A, and T of the youngest fully expanded leaf of corn. Legend: open triangles, normal-1 plants (4-leaf stage); solid triangles, normal-2 plants (5-leaf stage); open circles, Fe; solid circles, S; open squares, Mg; solid squares, Mn.

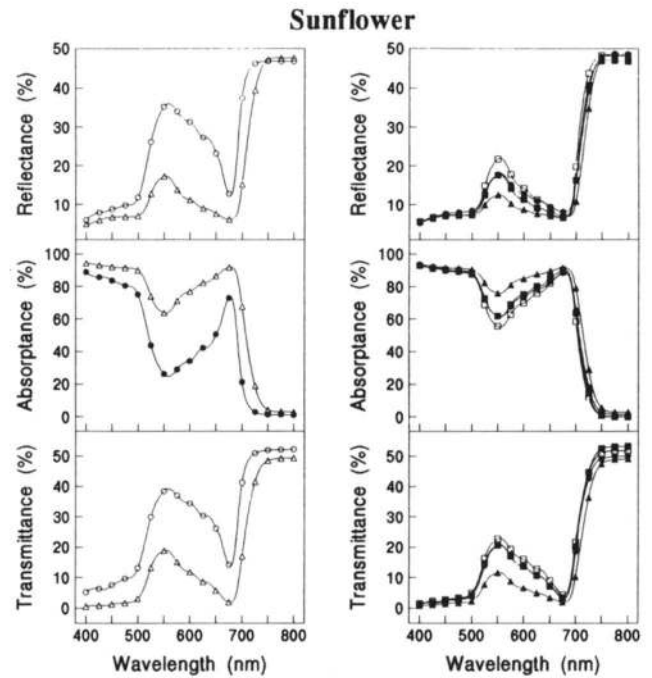


Fig. 3. Spectra of R, A, and T of the youngest fully expanded leaf of sunflower. Legend: open triangles, normal-1 plants (Stage V4); solid triangles, normal-23 plants (Stages V10 and R1 pooled); open circles, Fe; solid circles, S; open squares, Mg; solid squares, Mn. Spectra of normal plants in Stages V10 and R1 were averaged because only very slight differences occurred.

(1992), they were closely correlated with one another and with the integrated value over the entire 400- to 700-nm range (results not shown). Thus, values at these wavelengths are representative of the entire visible wavelength region.

All growth-medium mineral deficiencies caused pronounced modifications in leaf A, R, and T spectra in the visible region. The lowest variations produced by mineral deficiencies were recorded at wavelengths shorter than 500 nm and at 678 nm; the greatest variations recorded were near 555 and 700 nm. Figure 4 shows (for all species) the absolute difference between R, A, and T of leaves grown under normal and mineral-deficient growth medium conditions at 555 nm and 678 nm (555 nm being the wavelength of maximum variation among treatments and 678 nm being the wavelength of maximum *in vivo* Chl absorption). At 555 nm, all mineral deficiencies reduced A and increased R and T. Iron deficiency in the growth medium caused a marked decrease in leaf A in all species; in corn, the decrease was 54%, and in wheat and sunflower it was nearly 40%. With the other mineral deficiencies, leaf A decreased differently from species to species. The greatest reduction in leaf A was induced in barley and wheat by S deficiency and in corn and sunflower by Mg deficiency, while the smallest reduction was induced in barley and wheat by Mn deficiency and in corn and sunflower by S deficiency. Reflectance increased by a smaller percentage than T in all species and nutrient deficiencies, with the exception of the Mn-deficient barley leaf and Mg- and Mn-deficient wheat leaves. At 678 nm, where maximum Chl radiation absorption occurred, deficiency-induced modifications of R, A, and T were smaller than at 555 nm, with the

exception of corn grown in Fe-deficient growth medium, which showed similar modifications at 555 and 678 nm.

No differences in leaf A, R, and T between plants grown under normal and mineral-deficient conditions were observed in the NIR region (data not shown).

Leaf spectral properties at 429, 555, 678, and 700 nm were not correlated with mineral concentrations in leaves. This is consistent with the findings of Labovitz et al. (1983) and Milton et al. (1991). Milton et al. (1991) suggested an indirect effect of deficiency on plant R, hypothesizing an inhibition of nutrient or water uptake. Our data confirmed depleted uptake of other minerals in addition to the deficient mineral and indicated that reduction in leaf nutrient concentration reduced Chl formation. In all species, the first effect of each mineral deficiency was a reduction in leaf Chl concentration and consequently a decrease in A and an increase in R and T. A curvilinear relationship between leaf Chl *a* and *b* concentration and A, R, and T at 429, 555, 678, and 700 nm and in the entire visible region was observed, irrespective of deficiency and species. Relations between spectral properties at 429, 555, 678, and 700 nm and in the visible region and leaf Chl *a* and *b* concentration were always significant at 0.01 probability level (Table 4).

In agreement with Buschmann and Nagel (1993), in the visible region the best correlation between leaf Chl concentration and R, T, and A was recorded at 555 and 700 nm. Variations at these wavelengths were of the same amount and were the greatest in the spectra. At 555 nm, when Chl *a* concentration increased from 23 to 479 $\mu\text{mol m}^{-2}$, R decreased from 34 to 13%, T decreased from 51 to 12%, and A increased from 15

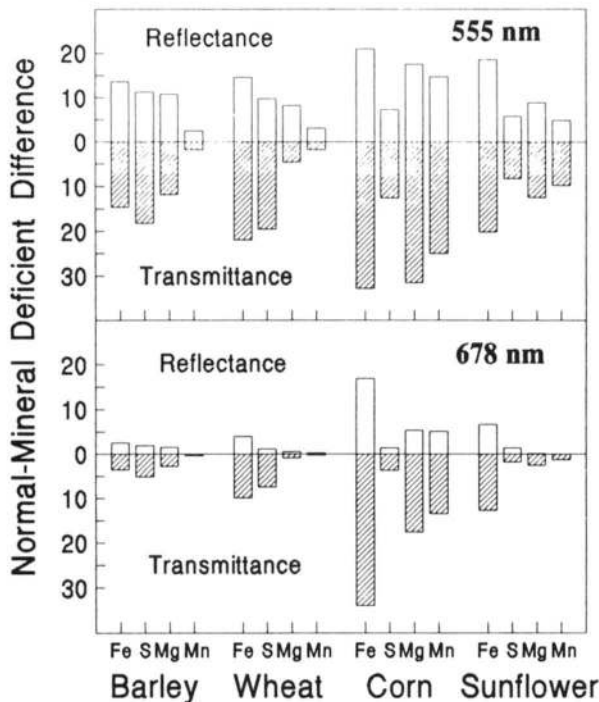


Fig. 4. Absolute difference (percent) of mean R and T between nutrient deficient and normal leaves of barley, wheat, corn, and sunflower at 555 and 678 nm. Values are means of a 10-nm waveband centered on 555 and 678 nm. The area between percent R and T is percent A. Mean R, A, and T of the mineral-deficient leaf differed from its control except for the Mn-deficient leaf in barley and the Mg- and Mn-deficient leaves in wheat and in sunflower.

to 76% (Fig. 5). Although R is the spectral property commonly utilized in remote sensing, A may be the most suitable spectral property for detecting small variations produced by changes in leaf physiologic characteristics, such as those due to nutrient deficiency, because the range of variability of A is higher than that of R or T.

Red-Edge Position

Red-edge position will be discussed only in R spectra, since it varied by only 2 to 3 nm among R, A, and T spectra (Table 5). All nutrient deficiencies produced a shift in red-edge position to shorter wavelengths, but this shift was found to differ among species. Red-edge shifts of more than 17 nm were observed for corn grown

Table 4. Correlation coefficients between leaf spectral properties at 429, 555, 678, and 700 nm and in the visible region (PAR) (400–700 nm) and leaf chlorophyll (Chl) *a* and *b* concentration.

Chl	PAR	Leaf spectral properties (by wavelength, nm)			
		429	555	678	700
Reflectance (R)					
<i>a</i>	0.908**	0.644**	0.903**	0.904**	0.903**
<i>b</i>	0.866**	0.555**	0.904**	0.664**	0.891**
Absorbance (A)					
<i>a</i>	0.978**	0.927**	0.985**	0.960**	0.979**
<i>b</i>	0.888**	0.720**	0.939**	0.745**	0.925**
Transmittance (T)					
<i>a</i>	0.974**	0.937**	0.977**	0.963**	0.981**
<i>b</i>	0.863**	0.708**	0.904**	0.760**	0.904**

** Model significant at the 0.01 probability level (*F*-test).

† The model used was $y = a[1 + b \exp(-kx)]$, where $x = \text{Chl concentration}$, $y = \text{R, A, or T at each wavelength}$, and $a, b,$ and k are constants.

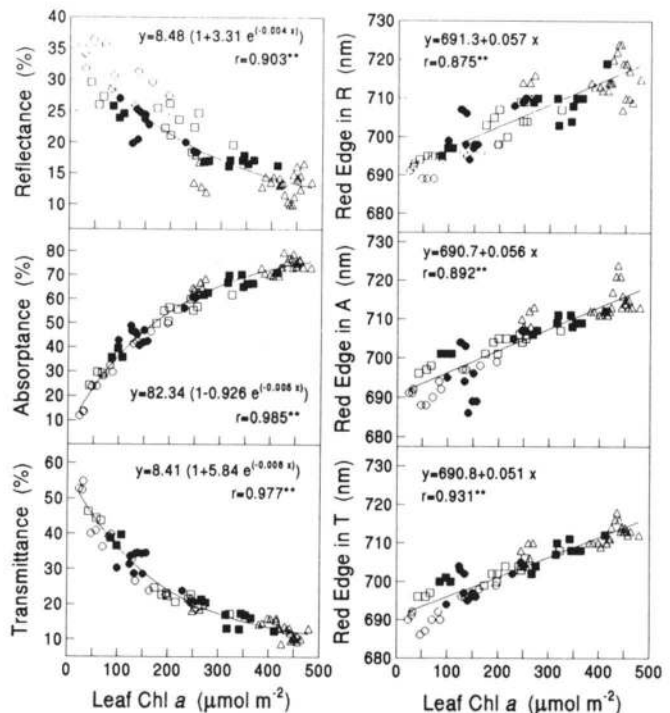


Fig. 5. Relationships between leaf Chl *a* concentration and leaf R, A, and T at 555 nm (values are means of a 10-nm waveband centered on 555 nm) and between leaf Chl *a* concentration and red-edge position in R, A, and T spectra. Legend: open triangles, normal plants; open circles, Fe; solid circles, S; open squares, Mg; solid squares, Mn.

with Fe-, Mg-, and Mn-deficient growth media, for barley and wheat with Fe- and S-deficient media, and for sunflower with the Fe-deficient medium. In wheat and barley, Mn deficiency shifted the red-edge position by only 3 to 4 nm. No correlation was observed between red-edge position and leaf Fe, S, Mg, and Mn concentration. Thus, red-edge position allowed separation by neither species nor mineral deficiency. Red-edge position was linearly correlated with leaf Chl *a* concentration, irrespective of spectral properties, species, or mineral deficiencies. Increasing leaf Chl *a* concentration shifted red-edge position in R spectra to longer wavelengths with an increase of 57 nm per $\text{mmol Chl } a \text{ m}^{-2}$ (Fig. 5).

CONCLUSIONS

Nutritional stress caused first a decrease in leaf chlorophyll concentration and subsequently a decrease in absorbance, and it caused an increase in reflectance and transmittance, and shifted the red-edge position to shorter wavelengths. For the crop species tested, variations in spectral properties and red-edge position were proportional to stress level and leaf Chl concentration, and were observed in the same spectral wavelength.

In barley, wheat, corn, and sunflower, mineral deficiency affected leaf concentration of the deficient element and other elements, with percent variation differing according to species and deficiency. Modifications of leaf spectral properties were not produced by or characteristic of only one deficient element but rather the combined result of several deficient elements.

Many variations in spectral properties can occur, depending also on the interaction between deficiency of a

Table 5. Red-edge position of R, A, and T spectra of the youngest fully expanded leaf of barley, wheat, corn, and sunflower plants grown under normal and Fe-, S-, Mg-, and Mn-deficient conditions. For each species and deficiency, comparison was made between leaves grown under normal and mineral deficient conditions. The position in normal plants differs among elements because of time of sampling.

Crop	Red-edge position (by growth medium treatment)							
	Fe		S		Mg		Mn	
	Normal	Deficient	Normal	Deficient	Normal	Deficient	Normal	Deficient
	nm							
	Reflectance (R)							
Barley	714	696*	715	698*	715	703*	715	712*
Wheat	713	695*	713	696*	713	705*	713	709*
Corn	715	692*	715	707*	723	694*	723	696*
Sunflower	710	689*	714	709*	719	707*	719	710*
	Absorbance (A)							
Barley	713	698*	714	695*	714	702*	714	711*
Wheat	711	694*	712	688*	712	706*	712	709*
Corn	712	692*	712	704*	722	697*	722	701*
Sunflower	708	689*	712	706*	716	705*	716	707*
	Transmittance (T)							
Barley	713	698*	713	696*	713	701*	713	711*
Wheat	710	693*	710	695*	710	705*	710	708*
Corn	709	691*	709	703*	717	697*	717	701*
Sunflower	705	688*	711	704*	713	702*	713	704*

* Significant at the 0.05 probability level (*F*-test).

given mineral and level of deficiency. In wheat, for instance, the same leaf Chl concentration, or leaf R, or red-edge position can be found with a slight Fe deficiency as with a severe Mg deficiency. Therefore, measurements of spectral properties are utilizable for detecting mineral deficiencies in field crops only if both plant species and specific nutrient deficiency are known. In conclusion, measurements of spectral properties of attached leaves may be very interesting for early, easy, and inexpensive determination of mineral deficiencies or other stress conditions (i.e., water stress), only when the species and the deficiency are known a priori.

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