

## 1 1. Introduction

2 Lutein, a naturally occurring carotenoid, has numerous benefits on human health such as  
3 reducing age-related macular degradation (AMD) (Bian et al. 2012), protecting retinal pigment epithelial  
4 (RPE) cells from photo-oxidative (Domingos et al. 2014), providing antioxidant activity, and preventing  
5 several diseases (Li et al. 2010). Due to its chemical structure, lutein can be easily oxidized and degraded  
6 due to light and heat (Mitri et al. 2011) and also has low water solubility, poor absorption, and low  
7 bioavailability (Kotake-Nara and Nagao 2011). To take full advantage of its potential as an antioxidant,  
8 novel delivery systems have been developed to enhance its ability to be dispersed in water, as well as its  
9 physicochemical stability during processing and storage conditions. Delivery systems developed for  
10 antioxidant delivery include solid lipid nanoparticles, nanocrystals, and nanoliposomes (Mitri et al., 2011;  
11 Mitri et al. 2011; Tan et al. 2013). All these forms were associated with an increase in the stability of the  
12 incorporated drug against physical-chemical degradation. Specially, it has been shown that loss of lutein  
13 was faster when entrapped in single-layer (SL) emulsion, compared to layer-by-layer (LBL) emulsion,  
14 stabilized by gum Arabic (Lim et al. 2014).

15 The addition of stabilizing surfactants to the delivery systems is hypothesized as one of the  
16 simplest and most effective strategies to sustain the release profiles, to improve the physical stability of  
17 the nanodelivery system and chemical stability of entrapped fat-soluble drugs (Podaralla and Perumal  
18 2012). An effective example in this regard is development of a core/shell nanoparticle made with lecithin  
19 as the core and pluronic F127 as a shell layer, for delivery of positively charged proteins engineered to  
20 provide protection, sustained release, and enhanced stability and functionality of entrapped bioactives  
21 (Choi et al. 2010; Oh et al. 2006).

22 A number of recently published studies provided advancing evidence that protein-based  
23 polymeric nanoparticles synthesized from gliadin, soy proteins, lectins, and zein can be successfully  
24 made specifically for food applications (Elzoghby et al. 2012). Among these natural potential  
25 nanocarriers, zein is particularly interesting as a naturally occurring polymer for synthesis of nanodelivery  
26 systems. It is a hydrophobic compound classified as generally recognized as safe (GRAS) as a direct  
27 human food ingredient by the Food and Drug Administration (FDA) (Elzoghby et al., 2012). Several  
28 attempts have been made to synthesize zein nanoparticles with entrapped drugs, antimicrobial agents,

29 and bioactive compounds such as 5-fluorouracil (Lai and Guo 2011), thymol (Zhang et al. 2014),  
30 curcumin (Gomez-Estaca et al. 2012), and essential oils (Parris et al. 2005)(Wu et al. 2012). Solutions  
31 were searched to improve stability of the zein nanoparticles during GI transit. For example, caseinate was  
32 used as an electrosteric stabilizer to prevent aggregation of zein nanoparticles in the neutral pH of the  
33 intestine (Patel et al. 2010). Even though data is available on characteristics of zein nanoparticles loaded  
34 with various antioxidants, little is known about stability of lutein entrapped in zein nanoparticles when  
35 exposed to various processing and storage conditions, and on the effect of surfactants on the release and  
36 stability of lutein under these conditions.

37 The objective of this paper was to assess lutein thermal and photo-stability, and lutein release  
38 from zein nanoparticles in the presence and absence of lecithin and pluronic F127 co-surfactants. Lutein-  
39 loaded zein nanoparticles, made with and without surfactants, were synthesized using a liquid-liquid  
40 dispersion method. A combination of phospholipid soybean lecithin and tri-block copolymer pluronic F127  
41 was used in the formulation as surfactants to promote physicochemical stability of the nanoparticles and  
42 entrapped bioactives. Lutein emulsions were prepared in parallel to be used as a control. Dynamic light  
43 scattering (DLS) and transmission electron microscopy (TEM) were used to characterize particle physical  
44 stability. Lutein release from nanoparticles suspended in phosphate-buffered saline (PBS) was quantified  
45 in the absence and presence of surfactants; the degradation of released lutein was determined under the  
46 same release conditions. In addition, thermal and photo-oxidation of lutein were measured as indicators  
47 of lutein chemical stability. The hypothesis was that lutein entrapped in zein nanoparticles was more  
48 stable under various storage conditions and that the electrostatic affinity between the zein nanoparticles  
49 and surfactants will result in a more sustained release of lutein and improved chemical stability of the  
50 entrapped bioactive.

## 51 2. Material and Methods

### 52 2.1. Material and Reagents

53 Zein (Z3625), pluronic F127, chloroform, and ethanol were purchased from Sigma Aldrich (St.  
54 Louis, MO, USA). Soybean lecithin, hydrochloric acid, and sodium hydroxide were purchased from Fisher  
55 Chemical (Fisher Scientific International, Fairlawn, NJ). Lutein was provided by Kemin Foods, L.C. (Iowa,  
56 USA). Nanopure water obtained using Nanopure Diamond from Barnstead international (IA, USA) was

57 used for all solution preparation. 100kDA Spectra/POR cellulose ester Biotech membrane tubing and  
58 closures was purchased from Spectrum Laboratories Inc. (CA, USA). All other reagents and components  
59 used in this study were of analytical grade.

## 60 **2.2. Synthesis of Zein Nanoparticles with Entrapped Lutein**

61 Nanoparticles were synthesized by a liquid-liquid dispersion method, as follows. Briefly, 10 mg of  
62 zein was dissolved in 1 mL ethanol-aqueous solution (70:30% (v/v)). A lutein solution was prepared at  
63 0.75 mg/mL with 100% ethanol and was added dropwise to the zein solution at a ratio of 1:1 under mild  
64 stirring conditions. The mixture was injected into 7.5 mL of an aqueous phase containing a combination of  
65 lecithin and pluronic F127 0.045:0.09% (w/v) as surfactants. The sample was then processed in a  
66 microfluidizer at 30,000 PSI for 3 cycles (M-110P, Microfluidics, MA, USA). Subsequently, the sample  
67 underwent evaporation to remove ethanol under vacuum (at approximately 500-600 mmHg) and nitrogen  
68 injection (80 mmHg) in a rotovapor (Buchi R-124, Buchi Analytical Inc., DE, USA). The lutein-loaded zein  
69 nanoparticles produced after complete evaporation of ethanol were washed by dialysis using a 100kDa  
70 Spectra/POR CE membrane (Spectrum Rancho, CA, USA). The nanoparticle suspension was placed in  
71 the membrane and suspended in 1.5 L nanopure water for 48 hours; the dialysis medium was changed  
72 every 8 hours to remove free surfactants. The suspension was collected and kept at room temperature for  
73 further analysis. Zein nanoparticles without surfactants were prepared in parallel using the same method,  
74 with the exception that surfactants were not added to the aqueous phase. The lutein emulsion made with  
75 surfactants followed the same protocol was served as a control.

## 76 **2.3. Particle Size, Polydispersity Index (PDI), and Zeta Potential Analyses**

77 Freshly-made zein nanoparticle samples were characterized by measuring average diameter  
78 size, PDI, and zeta potential by dynamic light scattering (DLS), using a Malvern Zetasizer Nano ZS  
79 (Malvern Instruments Ltd., Worcestershire, U.K.). Before the measurements were taken, samples were  
80 prepared at a final concentration of 0.1-0.3 mg/mL, optimum for the instrument. All measurements were  
81 performed in triplicate.

## 82 **2.4. Morphology Analysis**

83 Morphology of freshly-made zein nanoparticle was observed by transmission electron microscopy  
84 (TEM). One droplet of the sample was placed on a copper grid of 400 mesh with a carbon film, and the

85 excess sample was removed with a filter paper. Uranyl acetate was used as a negative stain to improve  
86 the contrast of the sample.

## 87 **2.5. Entrapment Efficiency (EE) Measurement**

88 One milliliter of the freshly-made lutein-loaded zein nanoparticle sample was centrifuged at  
89 64,000 g for 1 hour, from which 95% of particles were recovered (data not shown). The supernatant and  
90 the nanoparticle pellet were collected. Both samples were broken by ethanol and then lutein was  
91 extracted with chloroform (1:1 ratio). The relative solubility of lutein in chloroform (6000 mg/L) is 20 times  
92 higher than that in ethanol (300 mg/L) (Craft and Soares 1992). The concentration of lutein was  
93 measured using a UV/Vis spectrophotometer (GENESYS 6:Thermo Spectronic) with glass cells of 1 cm  
94 path length recorded at 445 nm. The absorbance value was converted to lutein concentration based on  
95 the standard curve for lutein in 1:1 ethanol and chloroform. Entrapment efficiency (%) was estimated as  
96 the ratio of lutein amount in pellet to theoretical lutein entrapped as described by

97 
$$\frac{\text{Lutein amount in pellet}}{\text{Theoretical amount of available lutein}} \times 100 = \% \text{ EE. All measurements were performed in triplicate.}$$

## 98 **2.6. Lutein Release from Zein Nanoparticles in Phosphate-Buffered Saline (PBS)**

99 The release of the entrapped lutein from zein nanoparticles was studied in 0.01 M phosphate-  
100 buffered saline (PBS) solution (pH 7.4) at 37°C; 0.5% of Tween 20 was added to PBS to improve the  
101 solubility of lutein released. Briefly, 10 mL of freshly-prepared nanoparticles were added to 20 mL of  
102 Tween 20 enhanced PBS and mixed thoroughly. The mixture was divided and placed into 1.5 mL  
103 centrifuge tubes, placed into a shaking incubator (C25KC incubator shaker, New Brunswick Scientific, NJ,  
104 USA) at 37°C and 100 rpm. At pre-defined time intervals, a centrifuge tube was sampled and centrifuged  
105 at 64,000 g (Allegra 64R centrifuge, Beckman coulter, Inc., CA, USA) for 1 hour. The supernatant was  
106 removed and extracted with ethanol and chloroform (1:1 ratio) and then vortexed for 10 minutes. The  
107 extracted lutein was determined in the supernatant by measuring the absorbance at 445 nm using a  
108 UV/Vis spectrophotometer as described under entrapment efficiency section. The wavelength was  
109 selected to avoid interference from degraded products of lutein, consisting of low-molecular-weight and  
110 short-chain aldehydes and ketones, with a maximum absorbance ranging from 270 to 345 nm (Landrum  
111 2009). All measurements were performed in triplicate.

## 112 2.7. Degradation of Lutein Entrapped in Zein Nanoparticles

113 The degradation of lutein entrapped in zein nanoparticles (with and without surfactants) and lutein  
114 entrapped in surfactant-stabilized emulsion was determined by measuring and adding the amount of  
115 lutein detected in both pellet and supernatant under the same release condition.

## 116 2.8. Physical-Chemical Stability of Zein Nanoparticles with Entrapped Lutein

117 Freshly-made samples were stored in darkness at three different temperatures: 4°C in a  
118 refrigerator, 25°C at room temperature and 40°C in an incubator over one month. Samples were  
119 monitored for changes in average particle size, surface characteristic, and entrapment efficiency at the  
120 sampling time points of 7, 15, and 30 days of storage. All experiments were performed in triplicate.

## 121 2.9. Photo-Chemical Stability of Lutein Entrapped in Zein Nanoparticles

122 Nanoparticle and emulsion samples were placed in transparent glass vials and stored in a  
123 lightproof cabinet where they were exposed to 365 nm UV lamps (100 W: Blak-Ray model B 100AP) for  
124 up to 10 hours. At exposure time intervals (0.5, 1, 2, 3, 5, 7, and 10 hours), 1 mL was withdrawn from  
125 each sample and then extracted and analyzed by measuring lutein concentration using UV-Vis  
126 spectrophotometer (GENESYS 6: Thermo Spectronic) at 445 nm. The experiment was performed in  
127 triplicate.

## 128 2.10. Degradation Reaction Kinetics

129 A general reaction rate for the lutein degradation and release kinetics can be described by  
130  $-\frac{d[C]}{dt} = k[C]^n$ : where  $C$  is the lutein amount ( $\mu\text{g}$ ),  $k$  is the reaction rate constant, and  $n$  is the order of the  
131 reaction. The correlation coefficient ( $R^2$ ) was used as an indicator of the best fitting of the kinetic models  
132 for lutein release and degradation studies. The degradation of lutein against UV exposure, followed first-  
133 order kinetic as described by  $\ln\left(\frac{[C]}{[C_0]}\right) = -kt$ , similar to the results found in other studies (Abdel-Aal et al.  
134 al. 2010; Aparicio-Ruiz et al. 2011; Dhuique-Mayer et al. 2007; Lim et al., 2014). Lutein degradations  
135 under PBS conditions and storage as a function of time and temperatures followed second-order kinetics  
136 as described by  $\frac{1}{[C]} = \frac{1}{[C_0]} + kt$ : where  $C$  is the lutein amount ( $\mu\text{g}$ ) at time  $t$ ,  $C_0$  is the initial amount of lutein  
137 ( $\mu\text{g}$ ),  $t$  is the time (hours or days) and  $k$  is the reaction rate derived from the slope of linear regressions.

## 138 2.11. Data Statistical Analysis

139 All experiments were performed in triplicate and the results were reported as the mean  $\pm$   
140 standard error. Statistical analysis was performed in SAS (version 9.4, SAS Institute Inc., NC, USA). The  
141 analysis of variance (ANOVA) was used to determine significant differences between the systems. The  
142 significance level ( $P$ ) was set at 0.05.

## 143 3. Results and Discussion

### 144 3.1. Physicochemical Characterizations

145 A liquid-liquid dispersion method was successfully used to synthesize lutein-loaded zein  
146 nanoparticles in the presence and absence of surfactants. The combination of lecithin and pluronic F127  
147 was used to stabilize the nanoparticles. Pluronic F127 is a hydrophilic non-ionic surfactant copolymer  
148 consisting of a hydrophobic block of polypropylene located between two hydrophilic blocks of  
149 polyethylene glycol. Gel formation at higher temperatures efficiently overcomes the natural brittleness of  
150 zein, supporting its delivery system application (Li et al. 2013). Lecithin, a phospholipid food emulsifier or  
151 stabilizer, has a hydrophilic head, phosphatidylcholine (PC) and two hydrophobic tails,  
152 phosphatidylethanolamine (PE) and phosphatidylinositol (PI) (Wang and Wang 2008). Because of its  
153 partly mixed structure, lecithin can be used as an effective and stable emulsifier to interact simultaneously  
154 with both hydrophilic and hydrophobic substances (Choi et al., 2010). One or more layers of lecithin cover  
155 the surface of the hydrophobic zein nanoparticles, with lutein entrapped inside the zein matrix by  
156 electrostatic interaction. The hydrophilic head of lecithin connects with hydrophilic polyethylene glycol of  
157 pluronic F127 and the hydrophobic polypropylene possibly connects with zein matrix resulting in a  
158 hydrophilic zein nanoparticle loaded with hydrophobic lutein, which is useful to disperse this bioactive to  
159 the aqueous environment while protecting it from degradation (Fig. 1).

160 [Fig. 1. Near here]

161 Lutein-loaded zein nanoparticles and unloaded nanoparticles, with and without surfactants, were  
162 characterized immediately after purification (Table 1). Average particle size, PDI, and zeta potential of  
163 freshly-made samples were measured after 24 hours dialysis in buffer (pH 7.4) (Podaralla and Perumal  
164 2012). **The statistical analysis of the data revealed that there was significant difference in size, whereas  
165 PDI and zeta potential were not significantly different for zein nanoparticles made with and without**

166 surfactants. The average particle size of lutein loaded in zein nanoparticles with and without surfactants  
167 was  $216.5\pm 29$  nm and  $156.1\pm 18$  nm, respectively. While zein nanoparticles formed in the presence of  
168 surfactants had a relatively small polydispersity (less than 0.3), a higher PDI range of 0.33-0.48 was  
169 observed for nanoparticles made without combined surfactants.

170 [Table 1. Near here]

171 The size results were confirmed by TEM (Fig. 2). Particles with surfactants showed a spherical  
172 shape with a rough surface, with some particles connected in a surfactant mesh (Fig. 2A and 2B).  
173 Nanoparticle without surfactants showed a smaller size, with a more spherical morphology, but were less  
174 uniform in size and more likely to agglomerate (Fig. 2C and 2D) resulting in higher PDI values as  
175 measured by DLS. Similar zein nanoparticle images were reported in other studies (Parris et al., 2005;  
176 Zhang et al. 2014).

177 [Fig. 2. Near here]

178 Zeta potential has long been accepted as a good measure for assessing stability of a  
179 nanoparticles system. A high degree of stability of the nanodelivery system is expected at zeta potential  
180 values higher than +30 mV or lower than -30 mV on the basis of charge repulsion between the  
181 nanoparticles (Murdock et al. 2008). Particles covered by surfactants were found to be more negatively  
182 charged ( $-47.6\pm 1.6$  mV) than particles without surfactants ( $-31.9\pm 4.3$  mV), indicating a good stability of  
183 the surfactant stabilized particles. The particles covered only by lecithin showed a strong anionic surface  
184 charge ( $-51.3\pm 1.2$  mV) beneficial to inducing electrostatic interactions with positively charged compounds;  
185 the negative charge decreased ( $-47.6\pm 1.6$  mV) with the addition of non-ionic pluronic F127, but the  
186 charge was indicative of a stable suspension. Entrapment of lutein resulted in a change in zeta potential  
187 from  $-30.9\pm 3.3$  mV to a less negative value of  $-21.0\pm 8.6$  mV for particles made without surfactants. The  
188 hydrophobic interaction between lutein and zein nanoparticles as nonpolar molecules contributed to the  
189 rearrangement in the zein structure to accommodate the entrapped bioactive, resulting in the observed  
190 zeta potential change.

191 The presence of surfactants not only affected average particle size, PDI, and zeta potential, but  
192 also lutein entrapment inside the zein matrix. Without surfactants, entrapment efficiency was around  
193  $69.1\pm 11.4\%$ ; with the addition of surfactants to the system, which resulted in a thicker and denser zein

194 matrix, the entrapment efficiency increased to  $83\pm 5.8\%$ , revealing a statistically significant difference  
195 between the two systems. This result was expected; Xu and Hanna (Xu and Hanna 2006), suggested that  
196 addition of surfactants to the system can stabilize particles and therefore increase entrapment efficiency.

197 Both lecithin, an anionic phospholipid (Wang and Wang 2008) which forms an ionic complex with  
198 positively charged protein, and the added pluronic F127, which were used simultaneously to stabilize the  
199 nanoparticles, were found to confer a negative charge to the particles suitable for inducing the  
200 electrostatic interactions that allowed for good nanoparticle stability while increasing lipophilic drug  
201 loading (Oh et al. 2006).

### 202 **3.2. Lutein Release from Zein Nanoparticles in PBS**

203 Phosphate buffered saline (PBS) is a water-based salt buffer solution commonly used in  
204 biological research, particularly for testing drug release. The release kinetic of lutein from zein  
205 nanoparticles made with and without surfactants in an aqueous buffer PBS was evaluated (Fig. 3). **The**  
206 **release of the non-entrapped compounds available on the surface of the particles at the beginning stage**  
207 **of the release is referred to an initial-burst release (Fredenberg et al. 2011).** The release profile of zein  
208 nanoparticles can be described as a two-phase pattern, with the initial-burst release within 24 hours  
209 followed by zero-order release profile (Table 3). For particles made without surfactants (LTZN NSF),  
210 lutein released in the initial-burst phase amounted for 43.26%, whereas zein nanoparticles made with  
211 surfactants (LTZN SF) only released 19.83% lutein (Fig. 3). The results are not surprising as surface-  
212 associated lutein was expected to be released quickly from the surface of the particles when surfactants  
213 were not present to inhibit lutein release. Release of lutein after 24 hours followed zero-order kinetics  
214 (Table 3) with 51.51% lutein released at 168 hours from nanoparticles without surfactants versus only  
215 42.67% in the presence of surfactants (Fig. 3). Hydrophobic interaction between lecithin, lutein, and the  
216 polypropylene chains of pluronic F127 inhibited the hydrolytic degradation of zein and slowed the release  
217 of lutein. In the absence of surfactants rapid protein swelling resulted in a faster release of the entrapped  
218 bioactive by diffusion through aqueous channels formed in the hydrated swelled zein matrix (Choi et al.,  
219 2010). The results supported the hypothesis that the electrostatic affinity between the zein nanoparticles  
220 and surfactants; the combined lecithin and pluronic F127, were responsible for a more sustained release



221 of lutein. Statistical results supported that release of lutein from zein nanoparticles with surfactant was  
222 significantly different than release of lutein from zein nanoparticles without surfactant.

223 [Fig. 3. Near here]

### 224 **3.3. Entrapped Lutein Degradation in PBS**

225 Lutein is more susceptible to degradation than common carotenes due to conjugated double  
226 bonds and the two hydroxyl groups, considered more heat sensitive (Dhuique-Mayer et al., 2007). The  
227 degradation of lutein was assessed for lutein entrapped in zein nanoparticles with (LTZN SF) and without  
228 surfactants (LTZN NSF) and the result was compared to that of lutein in emulsified form with the same  
229 surfactants (LTEM SF) (Fig. 4). Lutein degradation profiles followed second-order kinetics with no  
230 significant different values (0.00003-0.00004) of the degradation rate constant ( $k$ ) among all systems  
231 studied (Table 3). Emulsified lutein degraded rapidly to approximately 40% under PBS condition after 168  
232 hours, in accordance with findings reported by Shi and Chen (Shi and Chen 1997), who found that 25-  
233 30% pure lutein in distilled water degraded at the same time.

234 [Fig. 4. Near here]

### 235 **3.4. Physical-Chemical Stability as a Function of Time and Temperature**

236 Physical stability of zein nanoparticles was investigated at 4°C, 25°C, and 40°C over 30 days by  
237 measuring size, PDI, and zeta potential. Chemical stability of entrapped lutein was assessed in parallel,  
238 by measuring the absorbance using a UV/Vis spectrophotometer at 445 nm (Table 2). Zein nanoparticles  
239 with and without surfactants were stable at low temperature, measuring between 156.1±18 to 216.5±29  
240 nm when stored at 4°C for 30 days. Nanoparticles increased in size over time when stored at higher  
241 temperature, especially in the absence of surfactants. For example, while size of nanoparticles with  
242 surfactants-increased to 380.5±51 nm over 30 days of storage at 25°C, particles without surfactants  
243 measured up to 3103±332 nm at the same temperature. At 40°C, sizes bigger than 1 µm were detected  
244 after 7 days of storage for the nanoparticles made without surfactants. The PDI generally increased with  
245 temperature and storage time (from 0.27 to 0.80). Zeta potential ranged from -18 mV to -25 mV for  
246 nanoparticles without surfactants, and between -15.2 mV to -38 mV for particles made with surfactants.

247 [Table 2. Near here]

248 The surfactants not only provided long-term storage stability over 30 days for the  
249 nanosuspension, but they also delayed the degradation of lutein (Fig. 5). Only 26% of entrapped lutein  
250 was degraded after 30 days at 25°C when entrapped in LTZN SF, compared to 54% which degraded at  
251 the same time when entrapped in LTZN NSF.

252 [Fig. 5. Near here]

253 Similar trends were found at 40°C in both particles; 13.8% and 7.5% of retained lutein remained  
254 in the lutein-zein nanoparticles with and without surfactants, respectively. Emulsified lutein degraded  
255 faster than lutein entrapped in zein nanoparticles at all temperatures. Lutein degradation under various  
256 temperatures for 30 days followed second-order kinetics, similarly to degradation of lutein in particles  
257 suspended in PBS for 7 days (Table 3). The lowest degradation rate ( $k$ ) values of nanoentrapped lutein  
258 were also found in the presence of zein nanoparticles in the presence of surfactants at all storage  
259 temperatures. Increased temperature resulted in increasing the degradation for all systems. Thus, it is  
260 concluded that while the preferred storage condition of lutein in 4°C (Lai and Guo 2011), stability of lutein  
261 can be improved by loading it in surfactant covered zein nanoparticles even at higher temperatures.

### 262 3.5. Photo-Chemical Stability against UV Exposure

263 Photochemical stability against UV light of lutein loaded in zein nanoparticles with surfactant and  
264 without surfactant was compared to lutein emulsified form made with the same surfactants. There was  
265 statistical significance in the surfactants and time as main effects. Emulsified lutein underwent  
266 photochemical degradation very quickly (Fig. 6). After 10 hours, only 1.42% entrapped lutein remained in  
267 the lutein emulsions whereas 15.91% lutein was protected by entrapment in zein nanoparticles without  
268 surfactants (Fig. 6). The zein nanoparticles combined with surfactants were able to provide the greatest  
269 protection against UV light-induced lutein degradation, with 46.53% of lutein remaining inside the zein  
270 nanoparticles after UV light exposure for 10 hours.

271 [Fig. 6. Near here]

272 The lowest degradation rate constant was found for lutein loaded in zein nanoparticles  
273 combined with surfactants (0.0753) whereas zein nanoparticles without surfactants was higher (0.1869)  
274 indicating that the rate of lutein degradation in the presence of zein nanoparticles plus the effect of  
275 surfactants was noticeably retarded compared with lutein delivered in emulsified form (0.4093) (Table 3).

276 The improved photo-chemical stability of lutein against UV light when entrapped in zein nanoparticles was  
277 mainly based on the competitive absorption of UV photons by zein. Zein has been proven to absorb UV  
278 due to aromatic amino acids such as phenylalanine in its sequence (Stoscheck 1990). Moreover, the  
279 effect of surfactants surrounding zein nanoparticles was another contributor for improved stability of  
280 lutein. Lecithin was successfully used as UV protectant previously (Sundaram and Curry 1996). The  
281 possible mechanism for the ability of lecithin to protect lutein from degradation was due to energy transfer  
282 from the excited lutein species to lecithin. Thus, photo-stability against UV light of the entrapped lutein  
283 was significantly improved by the UV absorption of zein nanoparticles with the support of the combined  
284 lecithin and pluronic F127 as surfactants.

285 [Table 3. Near here]

#### 286 **4. Conclusion**

287 Zein nanoparticles loaded with 7.5% lutein stabilized by the combined lecithin and pluronic F127  
288 surfactants were successfully synthesized using a liquid-liquid dispersion method. The addition of  
289 surfactants increased particle size and improved polydispersity index. Zeta potential slightly changed, and  
290 entrapment efficiency increased significantly. In the presence of the surfactants, the burst release  
291 decreased and the release kinetic was subsequently sustained. Zein nanoparticles showed a great ability  
292 to protect lutein from degradation under various storage conditions as compared to the emulsified lutein.  
293 The preferred storage condition of lutein-loaded zein nanoparticles with surfactants was 4°C for 30 days.  
294 In addition, this complex formulation provided a good protection against UV light for 10 hours. **Further**  
295 **studies are under way on the potential of zein nanoparticles to improve the physicochemical stability and**  
296 **functionality of entrapped antioxidants added to liquid food under simulated gastrointestinal (GI)**  
297 **environments.** Based on characteristics, release, and stability data, it was suggested that with the  
298 addition of surfactants to improve entrapment efficiency of hydrophobic bioactives and to protect lutein  
299 against chemical degradation, zein nanoparticles could provide better protection against bioactive losses  
300 during thermal and neutral conditions than other delivery systems such as emulsions.

301

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415

**Figure captions**

Fig. 1. Schematic formation of lutein-loaded zein nanoparticle stabilized by lecithin and pluronic F127 surfactants

Fig. 2. Transmission Electron Microscope (TEM) images of zein nanoparticles with surfactants (A and B) and without surfactants (C and D)

Fig. 3. Two-pattern release profiles consisting of an initial burst release within 24 hours and the following zero-order release of lutein from zein nanoparticles made with (LTZN SF) and without surfactants (LTZN NSF) in PBS solution (pH 7.4) at 37°C and 100 rpm for 7 days

Fig. 4. Degradation profiles following second-order reaction of lutein loaded in zein nanoparticles with (LTZN SF) and without (LTZN NSF) surfactants and lutein emulsion made with surfactants (LTEM SF) in PBS solution (pH 7.4) at 37°C and 100 rpm for 7 days

Fig. 5. Degradation profiles following second-order reaction of lutein entrapped in zein nanoparticles at different storage temperatures (A: 4°C, B: 25°C, and C: 40°C) over 30 days

Fig. 6. Photo-chemical stability profiles following first-order reaction of lutein loaded in zein nanoparticles with (LTZN SF) and without surfactants (LTZN NSF) and emulsified lutein with surfactants (LTEM SF) exposed to UV light for 10 hours

**Table captions**

Table 1. Characteristics of unloaded and lutein-loaded zein nanoparticles made with surfactants (SF) or without surfactants (NSF)

Table 2. Characteristics of lutein-loaded in zein nanoparticles at different storage temperatures over 30 days

Table 3. Fitting model for release and degradation of lutein-loaded zein nanoparticles

Fig. 1.

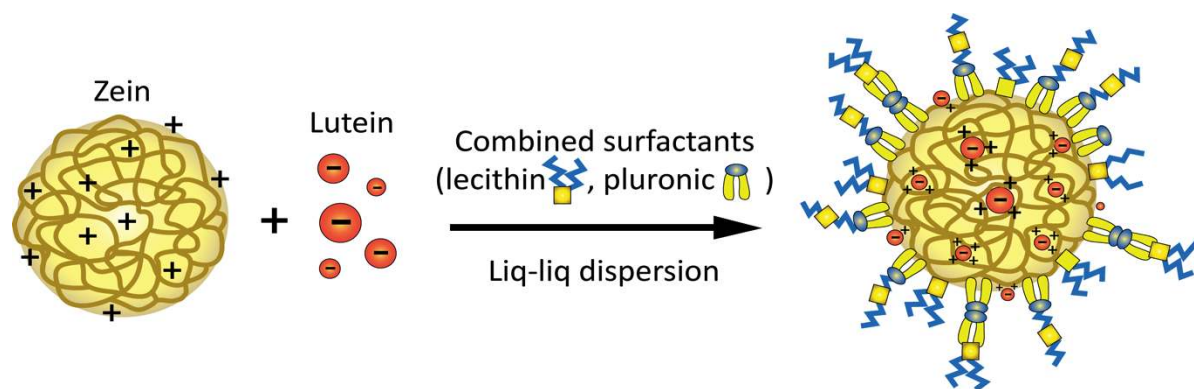
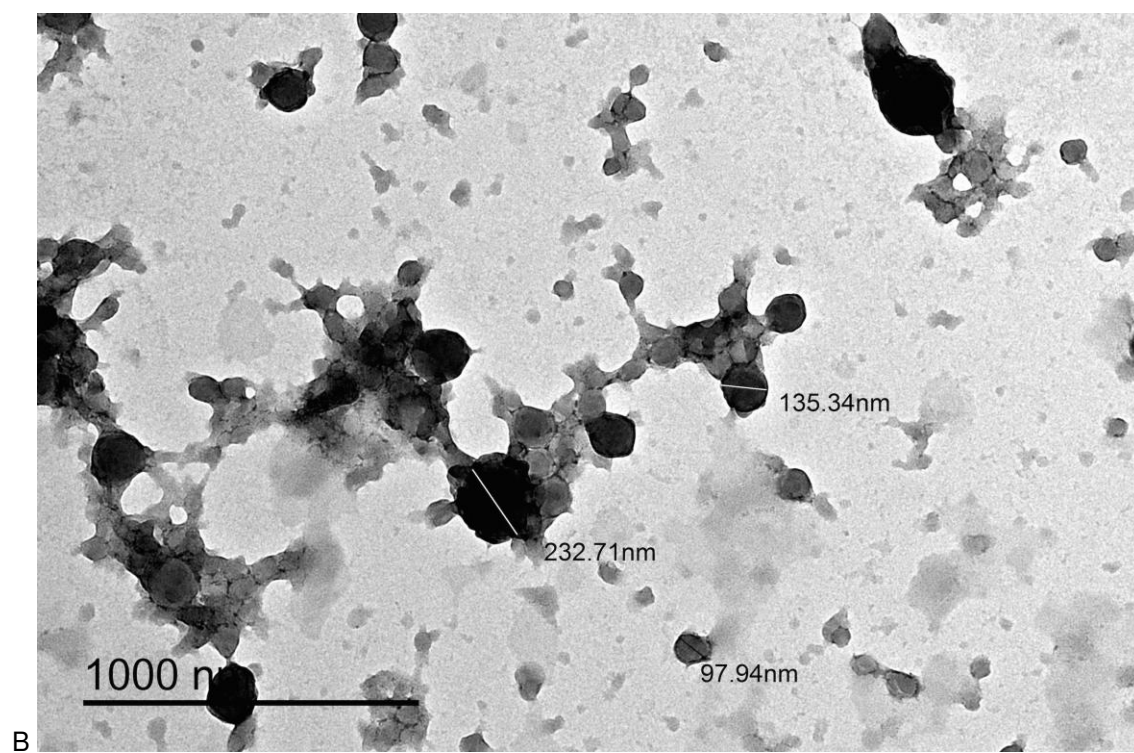
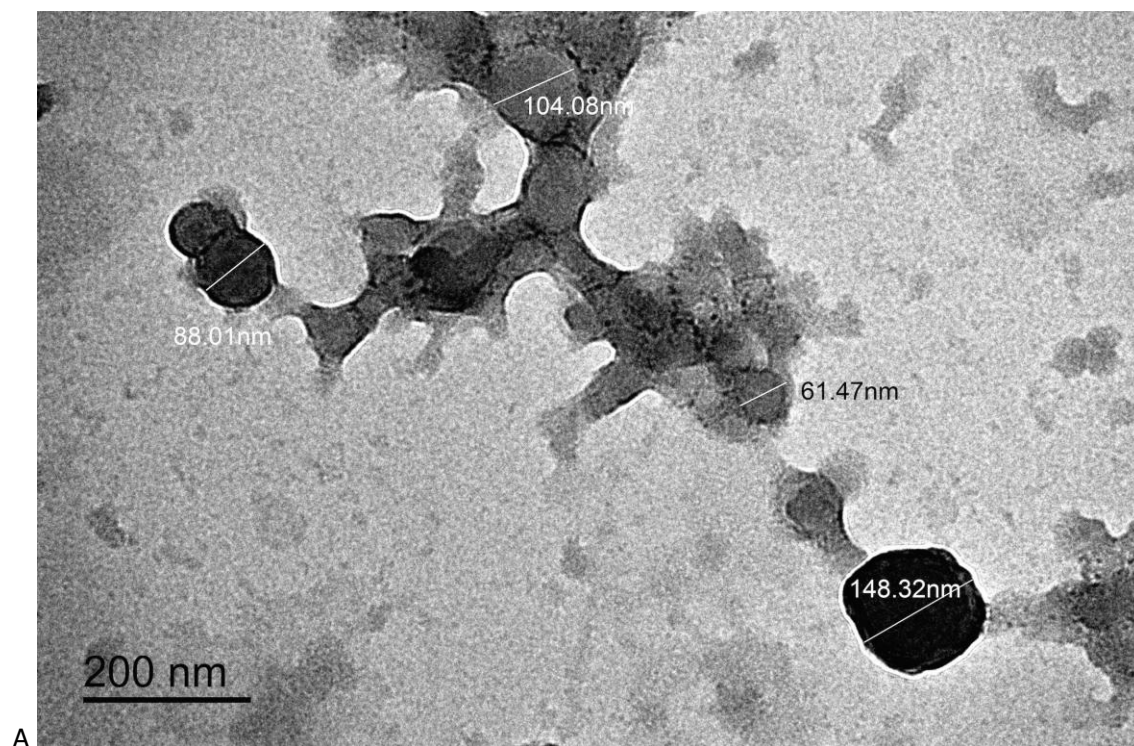




Fig. 2.



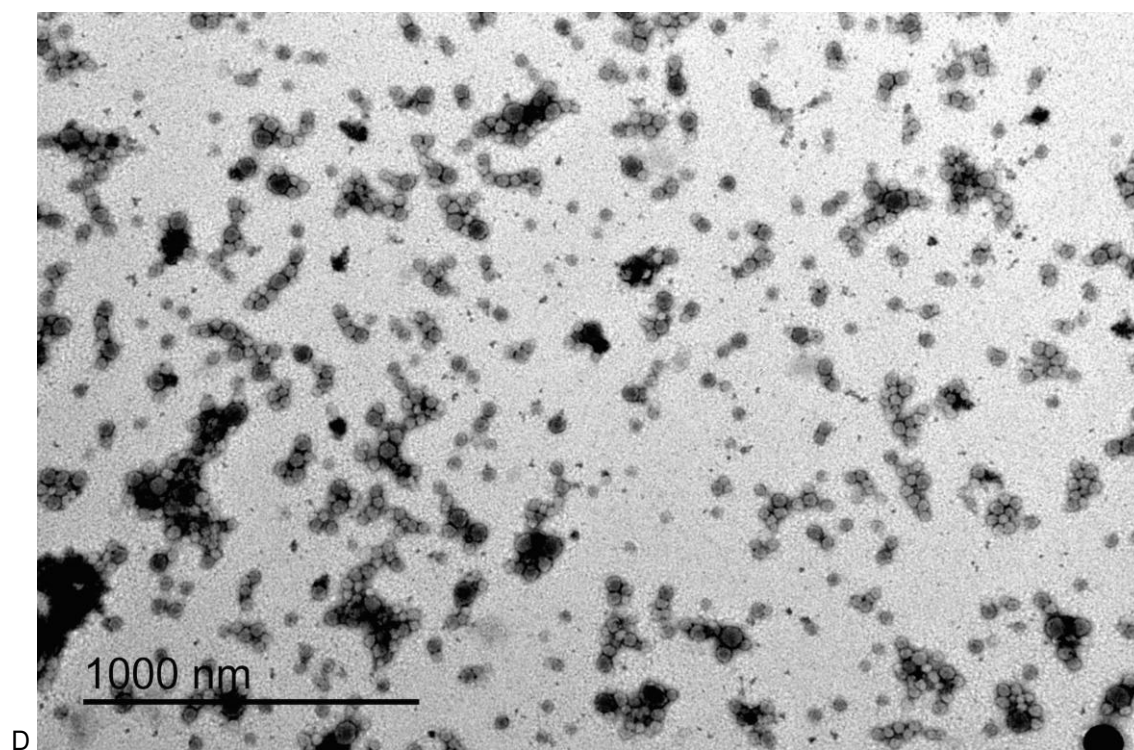
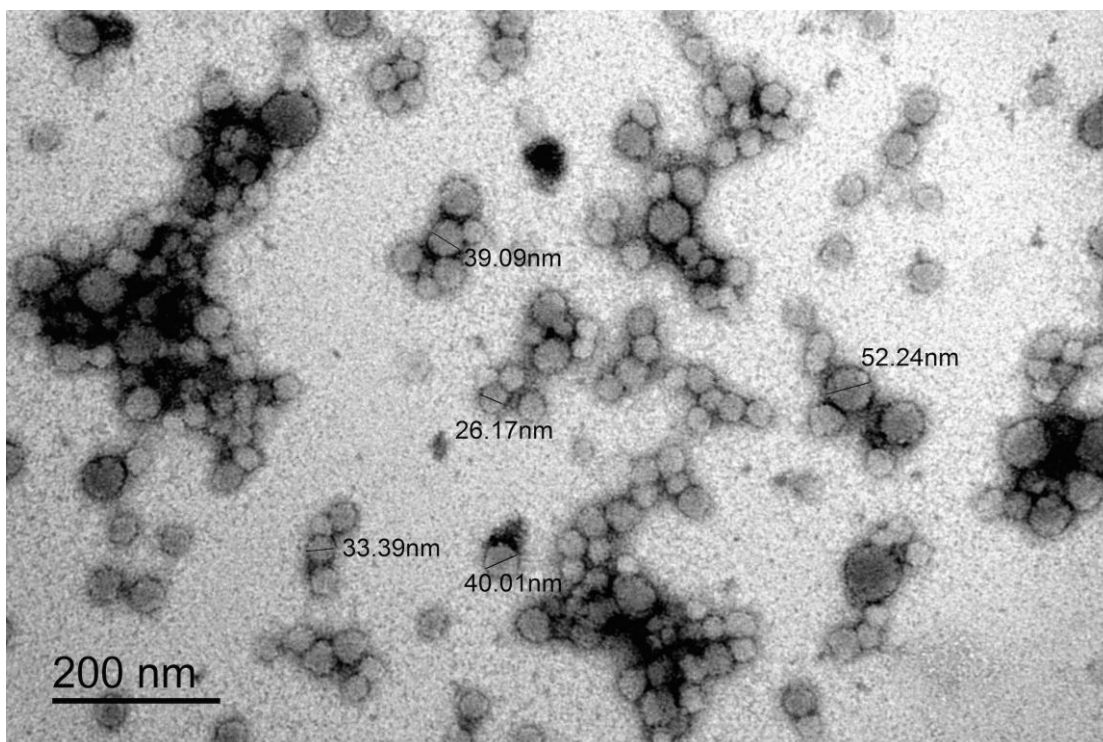


Fig. 3.

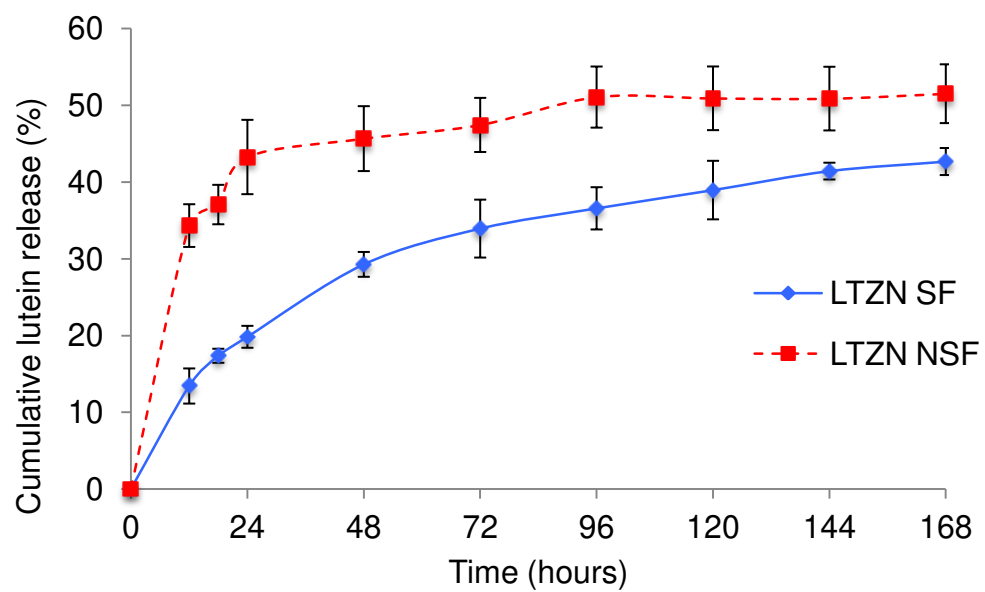


Fig. 4.

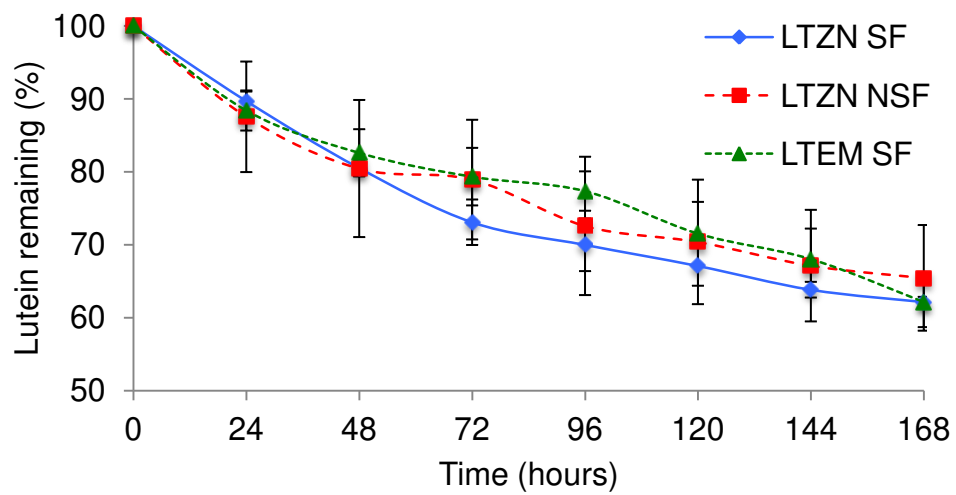


Fig. 5.

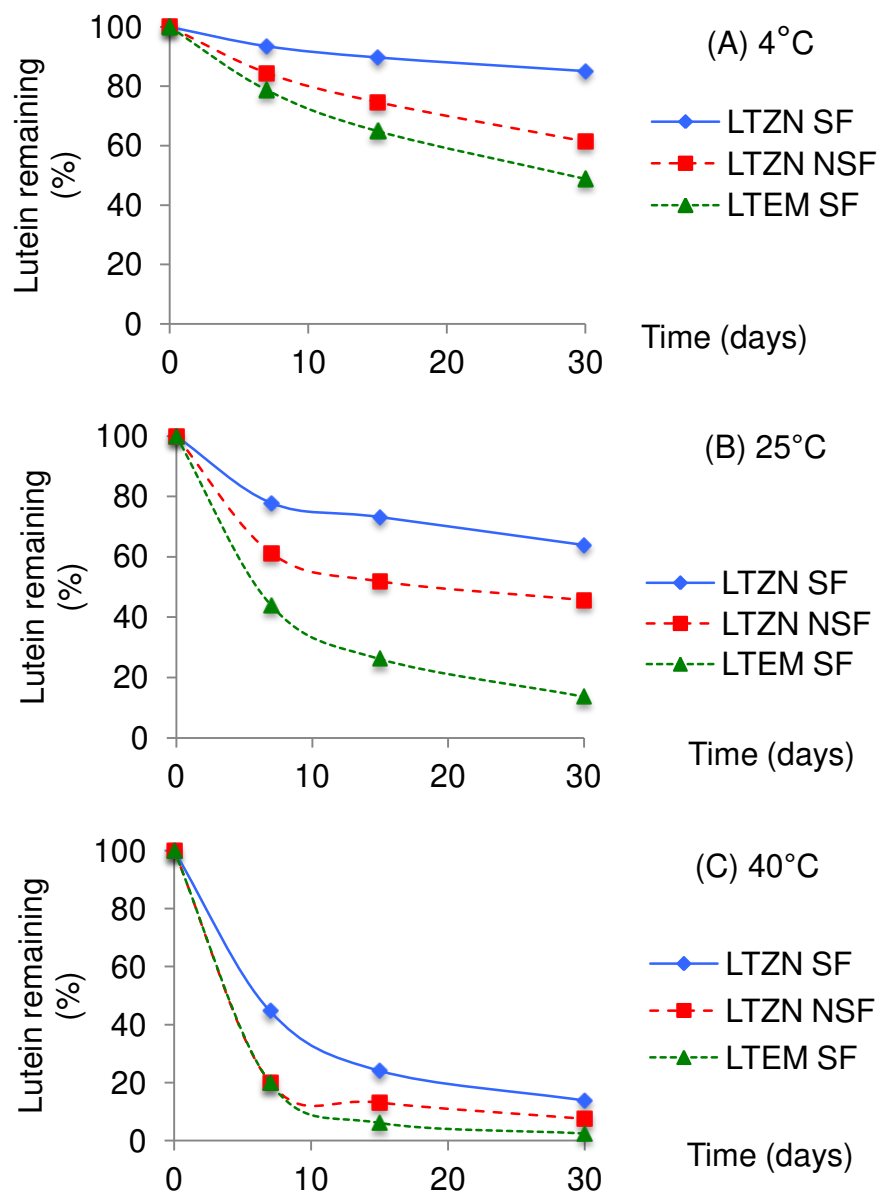
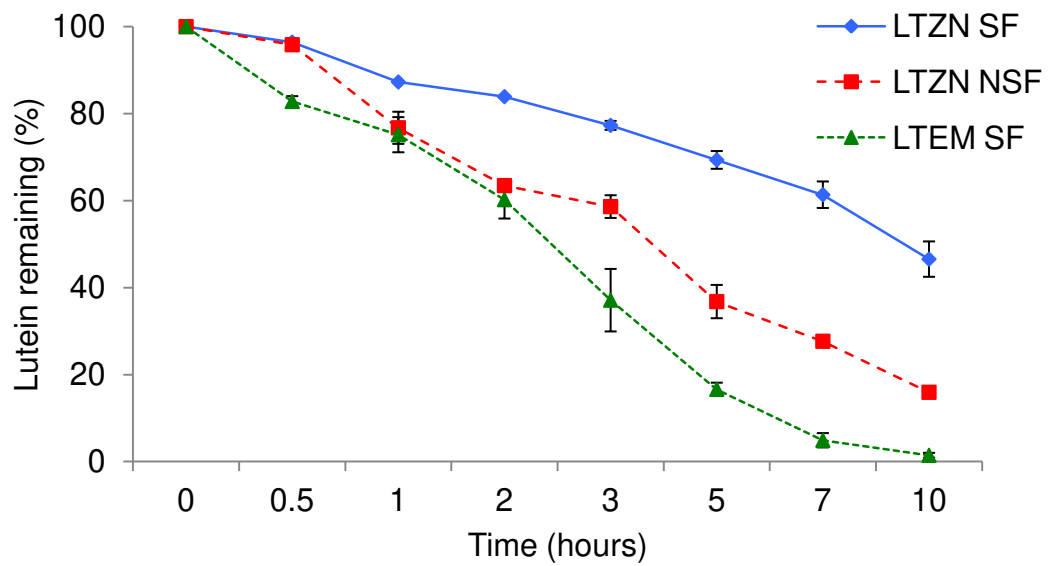


Fig. 6.



**Table 1.**

<b>Sample <sup>a</sup></b>	<b>Size (nm)</b>	<b>PDI (a.u)</b>	<b>Zeta Potential (mV)</b>	<b>EE (%)</b>
ZN SF	208.8±8.0	0.19±0.04	-47.6±1.6	-
LTZN SF	216.5±29*	0.26±0.09	-30.9±3.3	83.0±5.8*
ZN NSF	149.2±5.5	0.48±0.07	-31.9±4.3	-
LTZN NSF	156.1±18	0.33±0.06	-21.0±8.6	69.1±11.4

Note: Values are expressed as mean ± standard error (n=3). <sup>a</sup> ZN and LTZN represent formulations of zein nanoparticles and lutein-loaded zein nanoparticles, SF and NSF represent formulations with and with no surfactants respectively. Mass ratio of zein:lutein was 1:0.075 (% w/w) and mass ratio of lecithin:pluronic F127 was 0.045:0.09 (% w/v). \* shows statistically significant difference.

Table 2.

Sample	Temperature	Time (days)	Size (nm)	PDI (a.u)	Zeta Potential (mV)
LTZN SF	4°C	0	216.5±50	0.27±0.05	-30.9±3.3
		7	195.7±19	0.29±0.05	-31.1±10.8
		15	183.0±26	0.27±0.06	-32.2±10.7
		30	168.6±2	0.27±0.04	-33.0±10.8
	25°C	0	216.5±50	0.27±0.05	-30.9±3.3
		7	170.8±65	0.38±0.05	-23.3±2.4
		15	221.0±74	0.35±0.06	-21.8±9.5
		30	380.8±51	0.36±0.07	-15.2±0.3
	40°C	0	216.5±50	0.27±0.05	-30.9±3.3
		7	134.5±40	0.54±0.07	-38.0±1.9
		15	203.1±49	0.24±0.06	-31.8±7.4
		30	229.5±27	0.29±0.03	-29.5±2.9
LTZN NSF	4°C	0	156.1±18	0.26±0.06	-21.0±8.6
		7	142.4±32	0.32±0.11	-23.8±1.0
		15	189.2±55	0.26±0.07	-24.6±1.7
		30	198.9±47	0.39±0.13	-25.0±2.6
	25°C	0	156.1±18	0.26±0.06	-21.0±8.6
		7	567.7±203	0.56±0.07	-24.6±1.7
		15	1406.1±279	0.47±0.13	-23.7±1.5
		30	3103±332	0.58±0.15	-23.6±1.4
	40°C	0	156.1±18	0.26±0.06	-21.0±8.6
		7	1096.1±253	0.58±0.05	-18.0±2.7
		15	2434.5±535	0.73±0.19	-21.0±0.6
		30	3599.5±94	0.80±0.10	-22.7±4.2

Note: Values are expressed as mean ± standard error (n=3).



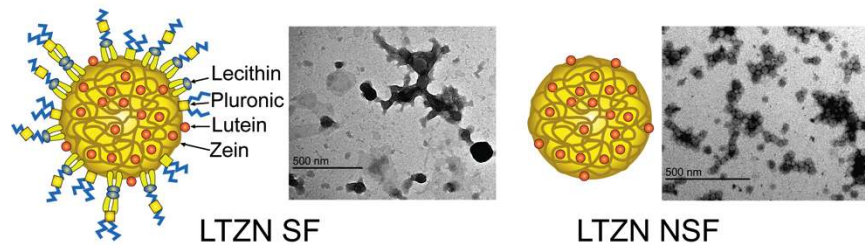
Table 3.

Experiment	Sample	Time	Kinetic model	K	R <sup>2</sup>
Release	LTZN SF	0-24 hours	Zero-order (Initial burst)	0.90930	0.95203
	LTZN NSF			2.02800	0.88626
	LTZN SF	24-168 hours	1 <sup>st</sup> order	0.00120	0.83448
	LTZN NSF			0.01270	0.92693
Degradation	LTZN SF	168 hours	2 <sup>nd</sup> order	0.00004	0.96389
	LTZN NSF			0.00003	0.91575
	LTEM SF			0.00003	0.97666
Physical stability	LTZN SF 4°C	30 days	2 <sup>nd</sup> order	0.00006	0.92430
	LTZN SF 25°C			0.00020	0.84325
	LTZN SF 40°C			0.00210	0.99788
	LTZN NSF 4°C	30 days	2 <sup>nd</sup> order	0.00020	0.99243
	LTZN NSF 25°C			0.00050	0.75685
	LTZN NSF 40°C			0.00420	0.98458
	LTEM SF 4°C	30 days	2 <sup>nd</sup> order	0.00040	0.99878
	LTEM SF 25°C			0.00210	0.99502
	LTEM SF 40°C			0.01210	0.96126
Photo-chemical stability	LTZN SF	10 hours	1 <sup>st</sup> order	0.07530	0.98454
	LTZN NSF			0.18690	0.99256
	LTEM SF			0.40930	0.98301

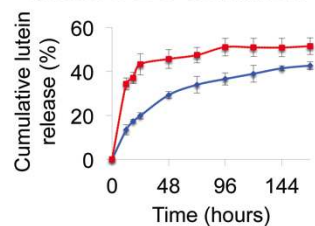
Note: LTZN and LTEM represent formulations of lutein-loaded zein nanoparticles and emulsified lutein, SF and NSF represent formulations with and with no surfactants respectively. K and R<sup>2</sup> represent the degradation rate constant and correlation coefficient, respectively.

## Graphical abstract

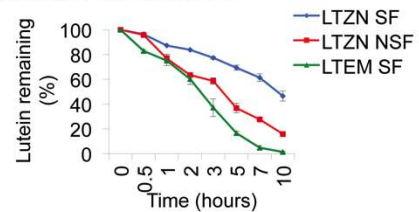
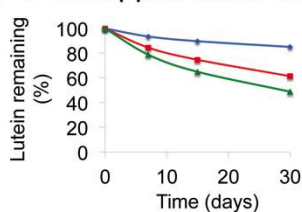
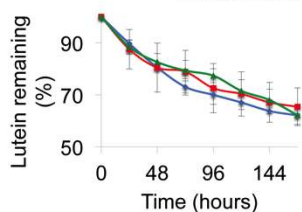
Proposed structures and TEM images of LTZN SF and LTZN NSF



Release profiles under PBS condition



Chemical stability of entrapped lutein under various conditions



PBS solution (pH 7.4, 37°C)

Storage at 4°C for 30 days

365 nm UV lamp for 10 hours