## **Supporting Information**

## Structural and Oxygen Binding Properties of Dimeric Horse Myoglobin

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Data collection				
X-ray source	SPring-8 (BL44XU)			
Wavelength (Å)	0.800			
Space group	P2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>			
Unit cell parameters				
<i>a</i> , <i>b</i> , <i>c</i> (Å)	57.3, 62.5, 83.4			
Resolution (Å)	20-1.05 (1.07-1.05)			
Total reflections	601280			
Number of unique reflections	139129 (6909)			
$R_{\rm merge}^{a}$	0.065 (0.450)			
Completeness (%)	100.0 (99.5)			
< <i>I</i> /σ( <i>I</i> )>	$< I/\sigma(I) >$ 13.2 (3			
Redundancy	4.3 (3.5)			
Refinement				
Resolution (Å)	20-1.05			
Number of reflections	133007			
$R_{\text{cryst}}^{b}$ (%) (F > 0 $\sigma$ )	12.8			
$R_{\text{free}}^{b}$ (%)	16.8			
Number of observations per parameter	4.5			
Number of atoms in an asymmetric				
unit	2398			
Protein	639			
Water	ater 86			
Heme				
Deviations from ideal geometry	RMSD	target $\sigma$		
Bond distance (Å)	0.014	0.020		
Angle distances (Å)	0.030	0.040		
Chiral volumes (Å <sup>3</sup> )				
C sp <sup>3</sup>	0.078	0.100		
C sp <sup>2</sup>	0.088	0.100		

 Table S1
 Statistics of data collection and structure refinement.

Anisotropic displacement parameters DELU ( $Å^2$ ) SIMU ( $Å^2$ )	RMSD 0.005 0.046	target σ 0.010 0.135
ISOR $(Å^2)$	0.098	0.100
Mean isotropic equivalent B-factor (Å <sup>2</sup> ) Main-chain atoms Side-chain atoms Heme atoms Water atoms		11.5 19.8 9.8 28.8
Ramachandran plot (%) Favored Allowed		97.7 2.3

Statistics for the highest-resolution shell are given in parentheses.

<sup>a</sup>  $R_{\text{merge}} = \Sigma_{\text{hkl}} | I - \langle I \rangle | (\Sigma_{\text{hkl}} | I |)^{-1}.$ 

<sup>b</sup>  $R_{\text{cryst}} = \Sigma_{\text{hkl}} ||F_{\text{obs}}| - k|F_{\text{calc}}|| (\Sigma_{\text{hkl}} |F_{\text{obs}}|)^{-1}$ , k: scaling factor.  $R_{\text{free}}$  was computed identically, except where all reflections belong to a test set of 5% of randomly selected data.

**Table S2**Root-mean-square deviation (rmsd) values of the 1–77 amino acid and the 86-153 aminoacid regions between the structures of the monomer and each independent protomer.

	1-77 Amino acid residues	86–153 Amino acid residues
Between the protomers (Å)	0.68	0.38
Between the monomer and each independent protomer (Å)	0.56, 0.77	1.20, 1.22



**Figure S1** Elution curves of horse metMb. a) Elution curve after incubation under 5% (v/v) ethanol, subsequent lyophilization, and dissolution to buffer; b) elution curve of purified monomeric metMb; c) elution curve of purified dimeric metMb; d) elution curve of dimeric metMb after incubation at pH 7.0 for 5 min at 70°C; e) elution curve of dimeric metMb after incubation at pH 7.0 for 3 days at 4°C; f) elution curve of dimeric metMb after incubation at pH 5.0 for 15 min at 4°C. Measurement conditions: column: Superdex 75 10/300 GL; flow rate: 0.2 mL/min; monitoring wavelength: 409 (red) and 280 nm (blue); solvent: 50 mM potassium phosphate buffer; pH: 7.0; temperature: 5 °C.



**Figure S2** Crystal structure of dimeric horse Mb with a 2*F*o-*F*c omit electron density map of the heme. The Heme is shown as a yellow stick model. The 2*F*o-*F*c electron density map is contoured at  $2\sigma$  and shown as a green mesh. Side-chain atoms of His64 and His93 (labeled as H64 and H93) and amino acids in the heme pocket are shown as stick models with labels.



**Figure S3** Hill plots of monomeric and dimeric horse Mbs for oxygen binding.  $pO_2$  and Y represent the partial oxygen pressure (mmHg) and the fractional oxygen saturation of Mb, respectively. Best-fitted linear lines are indicated by broken lines for monomeric (red) and dimeric (blue) Mbs. Measurement conditions are the same as those in Fig. 6.