Structure of a novel InsP₃ receptor

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Inositol 1,4,5-trisphosphate (InsP₃) constitutes a major intracellular second messenger that transduces many growth factor and neurotransmitter signals. InsP₃ causes the release of Ca²⁺ from intracellular stores by binding to specific receptors that are coupled to Ca²⁺ channels. One such receptor from cerebellum has previously been extensively characterized. We have now determined the full structure of a second, novel InsP₃ receptor which we refer to as type 2 InsP₃ receptor as opposed to the cerebellar type 1 InsP₃ receptor. The type 2 InsP₃ receptor has the same general structural design as the cerebellar type 1 InsP₃ receptor with which it shares 69% sequence identity. Expression of the aminoterminal 1078 amino acids of the type 2 receptor demonstrates high affinity binding of InsP₃ to the type 2 receptor with a similar specificity but higher affinity than observed for the type 1 receptor. These results demonstrate the presence of several types of InsP₃ receptor in brain and raise the possibility that intracellular Ca²⁺ signaling may involve multiple pathways with different regulatory properties dependent on different InsP₃ receptors.

Key words: Ca^{2+} channel/endoplasmic reticulum/ intracellular Ca^{2+} /ryanodine receptor/signal transduction

Introduction

The cellular responses to many growth factors and neurotransmitters is mediated by increases in intracellular Ca^{2+} caused by the release of $InsP_3$ (Berridge and Irvine, 1989). InsP₃ binds to specific intracellular receptors and causes the release of Ca^{2+} from intracellular Ca^{2+} stores that are probably part of the endoplasmic reticulum (Streb *et al.*, 1983). Generation of intracellular InsP₃ also leads to the delayed influx of Ca^{2+} via plasma membrane channels by an unidentified mechanism (Penner *et al.*, 1988). In addition, InsP₃ is instrumental in establishing or maintaining Ca^{2+} oscillations in many cells (Woods *et al.*, 1986; Berridge, 1990; Harootunian *et al.*, 1991; Petersen *et al.*, 1991).

An $InsP_3$ receptor from cerebellum has been well characterized (reviewed in Ross *et al.*, 1990; Shears, 1991) and immunolocalized to all parts of the endoplasmic reticulum in Purkinje cells (Mignery et al., 1989; Ross et al., 1989). The cerebellar InsP₃ receptor consists of a homotetramer of M_r 313 000 subunits that are encoded by a 10 kb mRNA which is subject to at least two different alternative splicing events (Mignery et al., 1989, 1990; Furiuchi et al., 1989; Ferris et al., 1991a). The primary structure of the InsP₃ receptor predicts the presence of eight transmembrane regions (Mignery et al., 1990; De Camilli et al., 1990) although an alternative model with nine transmembrane regions has also been suggested (Furiuchi et al., 1989). Mutagenesis studies demonstrated that the receptor forms homotetramers by virtue of intersubunit interactions localized to the regions of the membrane spanning sequences, and that the ligand binding site is localized to the amino-terminal fourth of the receptor (Mignery and Südhof, 1990). Based on these results, a domain model of the receptor was suggested whereby the receptor contains an amino-terminal binding domain, a carboxy-terminal Ca²⁺ channel domain, and an intervening coupling domain that regulates the relationship between InsP₃ binding and Ca²⁺ channel gating. Consistent with this model, the phosphorylation sites of the InsP₃ receptor were localized to the putative coupling domain (Mignery et al., 1990; Ferris et al., 1991b).

Although the cerebellar InsP₃ receptor is expressed at low levels in virtually all tissues investigated (Mignery et al., 1990), several lines of evidence suggest that there may be more than one type of InsP₃ receptor. Biochemical data demonstrated that InsP₃ binding has different characteristics in different tissues and that Ca²⁺ release by InsP₃ may be subject to different regulatory processes in different tissues (Guillemette et al., 1988; Palmer and Wakelam, 1989; Rossier et al., 1989; Ely et al., 1990; Pietri et al., 1990). These observations raise the possibility that different tissues may express different InsP3 receptors. Furthermore, it has been suggested that the endoplasmic reticulum is subcompartmentalized with respect to its function as a Ca²⁺-storing organelle (Villa et al., 1991; Takei et al., 1992), indicating that different types of InsP₃ receptors could be expressed in different subcompartments. For example, it is conceivable that a novel type of InsP₃ receptor may be localized to peripheral elements of the endoplasmic reticulum and physically coupled to components of the plasma membrane, thereby mediating the observed plasma membrane Ca²⁺ flux (Irvine, 1990). Another possibility is that different InsP3 receptors are localized in different compartments of the endoplasmic reticulum, conferring different Ca²⁺ release properties on these compartments.

All of these possibilities imply the presence of additional types of $InsP_3$ receptors that are distinct from the only currently described $InsP_3$ receptor. These receptors may nevertheless be structurally similar to this receptor although they differ from it in their intracellular targeting and/or

regulation. We now report the presence and full length structure of a novel type of $InsP_3$ receptor that fits these requirements. The presence of different types of $InsP_3$ receptors suggests that the intracellular Ca^{2+} signalling induced by $InsP_3$ may also be a function of the types and distributions of the $InsP_3$ receptors.

Results

In order to search for InsP₃ receptor related messages, a rat brain cDNA library was screened with an oligonucleotide corresponding to the last transmembrane region of the InsP₃ receptor (Mignery et al., 1990). This region was chosen for screening because it constitutes the region of highest homology between the $InsP_3$ and ryanodine receptors (Furiuchi et al., 1989; Mignery et al., 1989; Takeshima et al., 1989). In addition to multiple clones encoding the cerebellar InsP₃ receptor, two overlapping clones were isolated that were different from the cerebellar InsP₃ receptor clones (pI6 and pI15, Figure 1). Sequencing demonstrated that these clones encoded a novel transcript homologous to the InsP₃ receptor. Oligonucleotides corresponding to the 5' sequences of these and subsequent clones were then used to isolate further overlapping cDNA clones covering the entire coding region of the transcript and extending over 10.7 kb (Figure 1), and the sequences of all of these clones were determined.

The complete sequence of the InsP₃ receptor related transcript was assembled from the sequences of the overlapping cDNA clones (Figure 2). Its translated amino acid sequence predicts synthesis of a protein containing 2701 amino acids with a total molecular weight of 307 088 Daltons. The suggested initiation codon conforms well to the consensus sequence for initiator methionine codons (Kozak, 1989) and is preceded by an in-frame stop codon, suggesting that the sequence is full length with respect to the coding region. Clones containing poly(A) tails at two different positions in the 3' untranslated region were isolated (Figure 1). Both poly(A) tails are preceded by AT-rich sequences that may serve as polyadenylation signals (underlined in Figure 2). Northern blots demonstrated the presence of two messages for this cDNA corresponding to approximately 9 and 11 kb in size (data not shown), suggesting that there is differential polyadenylation of the 3' end of the message in vivo.

The amino acid sequence of the new protein was compared with that of the rat cerebellar InsP₃ receptor, revealing a high degree of homology between the two proteins with an overall sequence identity of 69%. Alignment of the two sequences with each other (Figure 3) demonstrated that their homology extends over their entire length but shows a patchy distribution, with regions of identity separated by completely dissimilar sequence stretches. For example, hydrophobicity plots of both sequences suggested the presence of eight transmembrane regions (Mignery et al., 1990a and data not shown) which are underlined in Figure 3 and labeled M1 to M8. Most of the putative transmembrane regions are highly conserved but two transmembrane regions, M2 and M3, show very little sequence similarity. In addition, many but not all loops connecting transmembrane regions are poorly conserved, for example the sequence separating the sixth and seventh transmembrane regions contains no similarity except for two conserved cysteine residues whereas



Fig. 1. Structure of the mRNA of the type 2 $InsP_3$ receptor (top) and distribution of the isolated cDNA clones (bottom). The open bar indicates the localization of the coding region in the mRNA. The scale of the graph is depicted in the lower left corner.

the transmembrane regions themselves are more than 90% identical. Similar patches of identical sequences separated by completely dissimilar regions can also be observed in other parts of the structures. In addition, deletions of one sequence relative to the other are observed, particularly in the coupling domain of the InsP₃ receptor that separates the transmembrane regions from the ligand binding domain. Interestingly, one of these deletions corresponds to an alternatively spliced region in the cerebellar InsP₃ receptor may also be alternatively spliced.

Figure 3 also contains the partial sequence of a third mRNA that is related to the InsP₃ receptor and was isolated by the polymerase chain reaction from a human kidney cDNA library (C.L.Newton, G.A.Mignery and T.C.Südhof, in preparation). This sequence shows the same pattern of similarity and diversity as described above, suggesting that there is a family of related sequences with a similar core of conserved residues.

The strong similarity between the novel sequence described here and the cerebellar InsP₃ receptor suggests that the new protein may represent a new type of InsP₃ receptor. To test this hypothesis, we took advantage of the fact that we had previously localized the ligand binding domain of the cerebellar InsP₃ receptor to the aminoterminal fourth of the receptor (Mignery and Südhof, 1990). Assuming that the ligand binding site of the putative new receptor would have a similar localization, we expressed the first 1078 residues of the new sequence as a soluble protein by transient transfection in COS cells (Figure 4). The ligand binding properties of the amino-terminal fragment of the novel receptor were compared to those of the corresponding homologous fragment from the previously characterized cerebellar InsP₃ receptor. In order to allow recognition of the two different recombinant proteins from the two receptors, the carboxy-termini of both proteins were fused to a 12 residue peptide epitope from the carboxy-terminus of the 116 K subunit of the vacuolar proton pump (Mignery and Südhof, 1990; Perin et al., 1991). After transient transfection, both proteins were expressed at high levels in soluble form in COS cells (Figure 4). Although the calculated molecular weights of the two proteins are very similar, their apparent mobility on SDS-gels differed slightly, possibly reflecting differences in their tertiary structure.

The InsP₃ binding properties of the recombinant proteins were then investigated in the cytosols of COS cells transfected with the expression constructs or control DNAs. Both recombinant proteins bound InsP₃ specifically, with the recombinant protein from the novel receptor having a slightly higher affinity than that of the corresponding fragment of the cerebellar InsP₃ receptor (Figure 5; apparent K_ds were 27 nM and 89.5 nM, respectively). Furthermore, in spite of the considerable sequence differences between the two receptors, InsP₃ binding to the recombinant proteins was displaced by different inositol phosphates to similar extents (Table I). In addition, InsP₃ binding was very sensitive to heparin in both proteins. These results demonstrate that the protein described here represents

a novel $InsP_3$ receptor, from now on referred to as type 2 $InsP_3$ receptor as opposed to the cerebellar type 1 $InsP_3$ receptor. In spite of their sequence differences, both receptors have similar binding specificities although different affinities.

114 GCAGTGTCCCCAGCGGTGGCCTCGCTGCCGCGCCCCGAGCTGAGCCGCCTGGGCCCCCGGGCTCAGAGGAGCCCAAGCTCACCCTTGCGAGCAGCGGAGACAGCGGGGACGGCC TCACGAGCG<u>TGA</u>AGCAGCATGTCTGACAAAATGTCCAGCTTCCTCTACATTGGGGACATCGTGTCCCTGTACGCGGAGGGCCTCGGTCAATGGCTTCATCAGCACCCTGGGGTTG 228 342 32 456 70 CAGTACTGGAAAGCCAAGCAGGCGAAGCAGGGAAACCACACGGAGGCTGCCCTGCTGAAGAAGCTCCAGCACGCCGCAGAGCTGGAACAAAAACAGAATGAGTCAGAGAACAGG 570 108 684 146 798 M R V S L D A A G N E G S W F Y I H P F W K L R S E G D N I V V G D K V V L ATGCCTGTAAATGCTGGGCAGCCCCTGCATGCCAGCAACGTGGAGCTCCTGGACAACCCCGGCTGCAAAGAGGTGAAATGCTGTTAATTGCAACACTAGCTGGAAAATCACTTTA 184 912 NAGOP L H A S N V E L L D N P G C K E V N A V N C N T S W K I T 222 TTCATGAAGTTCAGCTCCTACCGAGAGGATGTATTAAAAGGGGGTGACGTTGTGAGACTGTTTCATGCGGAACAAGAGAAGTTTCTGACCTGTGATGACTATGAGAAAAAACAG 1026 FMKFSSYREDVLKGGDVVRLFHAEQEKFLTCDDYEKKQ CACATTTTCCTGCGGACGACCTTGCGTCAATCAGCAACGTCGGCCACTAGCTCTAAAGCACTCTGGGAGATAGAGGTGGTTCACCATGATCCATGCCGCGGAGGTGCAGGACAG 260 1140 H I F L R T T L R Q S A T S A T S S K A L W E I E V V H H D P C R G G A G Q TGGAACAGCCTGTTCAGGTTTAAGCATCTTGCAACTGGGAACTACTGAGAGGCTTAACCCTGACTATCGAGATGCTCAAAATGAAGGAAAAACTGTGAGAGGAGGGGGAG W N S L F R F K H L A T G N Y L A A E L N P D Y R D A Q N E G K T V R D G E 298 1254 336 CTTCCAACCTCAAAGAAAAAACACCCAGGCAGGGGAGAAGATCATGTACACGCTGGTCTCGGTCCCGCACGGAAATGACATCGCGTCCCTTTTTGAACTTGACGCCACAACTCTG 1368 374 1482 SYVR ERHLCTNTWV STSIP D FFFR I 412 GTCATGCTGAAAATTGGGACCTGCCAGACCAAGGAAGACAAAGAAGCCTTTGCCATCGTGTGCGTCCCGCTGTCTGAGGTCCGAGACCTGGACTTTGCCAACGATGCCAACAAA 1596 V M L K I G T C Q T K E D K E A F A I V C V P L S E V R D L D F A N D A N K GTGTTGGCCACCACGGTGAAGAAGCTGGAGAACGGCAGCATCACCCAGAATGAGAGGAGGAGGTTTGTGGACAAGTTGTTGGAAGACCTTATTTCTTTGTGGCTGATGTGACCAAC 450 1710 A T T V K K L E N G S I T Q N E R R F V T K L L E D L I F F V 488 AACGGACAGGATGTTCTGGATGTGGTCATCACCAAGCCCAAGCCGGGAACGGCAAAAACTAATGAGGGAACAGAATATTCTGGCACAGGTGTTTGGGATCCTTAAAGCCCCTTTC N G Q D V L D V V I T K P N R E R Q K L M R E Q N I L A Q V F G I L K A P F AAGGAGAAGGCTGGGGAAGGCTCGATGCTGAGGCTGGAGGACCTGGGCGACCAGCGCTATGCCCGTACAAGTACGTGCTGCGCTCTGCTGCCGCGGCGTGTGCTGAGGCATCGCAG 1824 526 1938 K E K A G E G S M L R L E D L G D Q R Y A P Y K Y V L R L C Y R V L R H S Q CAGGACTACAGGAAGAACCAGGAGTACATTGCTAAGAACTTCTGCGTCATGCAGTCTCAGATCGGCTATGATATTTTGGCAGAAGAATACGATCACAGCCTTGCTACACAACAAC 564 2052 R K N Q E Y I A K N F C V M Q S Q I G Y D I L A E D T T Т 602 Ail 2166 RKLLEKHITAKEIETFVSLLRRNREPRFLDYLSDLCVS AATAGCACCGCCATCCCTGTGACTCAGGAGCTCATCTGCAAATTCATGCTGAGCCCCGGCAATGCGGACATCCTCATTCAGACGAAGCTGGTGTCCATGCAAGTGGAAAACCCC 640 2280 678 2394 ΜΕΣ ΣΤΙΡΟ Ο ΙΟ Ο ΓΕΥΨΙΥΨΤΟ ΣΝΚΕΡΗ G ΚΑΤΡΗΙΑΟ ΕΑ 716 AGGGAAGGCACCAAGGCTGACCTAGAAGTCCTGACCTATTACAGGTACCAGCTAAACCTCTTTGCAAGGATGTGCTTGGACCGCCAGTACCTGGCCATCAACCAGATTTCAACA 2508 R E G T K A D L E V L T Y Y R Y Q L N L F A R M C L D R Q Y L A I N Q I S T CAGCTGTCTGTAGACCTGATCCTGCGGTGCGTGTCTGACGAGAGCCTGCCCTTTGACCTCGAGCCTCTTCTGCCGACTCATGCTCCACATGCAGCGGGCCGGGACCCCCAG 754 2622 Q L S V D L L R C V S D E S L P F D L R A S F C R L M L H M H V D R D P Q GAGTCTGTGGGGGCCTGGCCCGGCTCTGGACTGAGATCCCCACCAGAGTCACGAGTCTGAGATAGACTCACCAGACTCTTCCAGAAATGACATGAAAAGAGAAA 792 2736 ESVVPVRYARLWTEIPTKITIHEYDSITDSSRNDMKRK TTTGCCCTGACAATGGAATTTGCGAAGAATATTTGAAAGAAGTTGTGAATCAACCGTTCCTTTTGGGGACAAAGAGAAAAATAAACTGACATTTGAGGTGGTCCACCTGGCC 830 2850 FALTMEFVEEYLKEVVNQPFPFGDKEKNKLTFEVVHLA CGGAACCTCATCTACGGATTCTACAGCTTCAGCGAGATGCTGCGGGGTGCGGGGGAGACCTCTGCGGGATCCTGCGAGACCCCCATGTCGTCGTGGTGTTTTGAAAGA 868 2964 IYF GF Y S F S F E I R I T R T E L A T I D T V O A P M SS 906 3078 944 3192 S K Q A S P G E Q E D V T V M D T K L K V I E I L Q F 982 GTCAGGCTGGACTACCGTATCTCTTACATGCTGTCCATATACAAGAAGGAATTTGGGGAGAATGATGGCAACGGTGACCCCTCTGCCAGCGGCACCCCGGAAACATTGCTACCA 3306 1020 3420 S A L V P D I D E I A A Q A E T M F A G R K E K T P V Q L D E G G R T F L AGAGTCCTCATCCATCCATGACTATGCTATGCTCCACTGCTGTCCGGGGCCCTGCAGCTGCTGTTCAAACACTTCAGCCAGAGAGGCAGAAGTGCTTCAGGCCTTCAAACAG R V L I H L I M H D Y A P L L S G A L Q L L F K H F S Q R A E V L Q A F K Q 1058 3534 1096 GTGCAGTTACTGGTGTCGAACCAAGATGTCGATAACTACAAGCAAAATCAAGGCAGACCTGGACCAACTTCGGCTGAAGAGCCGAGAAGTCCGAGCTATGGGTGGAAAAAGAGCGGC 3648 1134 3762 ENGDMGEGQAKGGEEANEESNLLSPVQDGAKT 1172 AGCAACAAGGGCAACAACTACCGGATCGTCAAGGAGATCTTGATTAGGCTGAGTAAGCTCTGTGTGCAGAACAAGAAATGTCGTAATCAACACCAACGGTTGCTGAAAAATATG 3876 1210 3990 LDLLQIP Y E K T D E K M N E V M D L A H T F 1248 AATCCACAGAACCAAGTTCTCCTTCACAAGCATCTCAACCTGTTCCTAACCCCTGGACTCCTGGAGGGGGAAACCATGCGGCACATCTTCATGAACAATTACCATCTGTGCAAT 4104 O N O V L L H K H L N L F L T P G L L E A E T M R H I F M N N Y H L 1286 GAGATCAGCGAGAGGGTGGTCCAGCACTTTGTGCACTGCATTGAGACACACGGCCGCCACGTGGAGTACCTGAGGTTCCTGCAGACCATCGTGAAAGCAGACGGCAAATACGTG 4218 E I S E R V V Q H F V H C I E T H G R H V E Y L R F L Q T I V K A D G K Y V AAGAAATGCCAGGACATGGTGATGACCGAGCTGATCAATGGGGGTGAAGACGTACTGATATTCTACAACGACCGAGCCTCATTTCCCATCCTTCTCAACATGATGTGCTCCCGAG 1324 4332 INGGEDVLIF YNDRASFP 1362 K C O D M V M T F L AGAGCCCGTGGGGATGAGAGCGGCCCCCTGGCCTACCATATCACCCTTGTGGAGTTGCTGGCAGCGTGCACCGGGGAAGAACGTCTACACGGAGATCAAGTGCAATTCTCTC 4446 1400 R G D E S G P L A Y H I T L V E L L A A C T E G K N V Y T E I K C N TTGCCTCTGGACGACATAGTGAGGGTGGTGACCCACGACGACTGCATTCCTGAGGTAAAAATTGCATACGTGAACTTTGTCAACCATTGTTACGTTGACACAGAAGTGGAGATG 4560 L P L D D I V R V V T H D C I P E V K I A Y V N F V N H C Y V D T E V E M AAGGAAATCTATACAAGTAACCACATTTGGAAGTTATTTGGAAGACTTCTTGGTGGATATGGCAAGGGTCTGCAACACCACCACGGAACACCGCAGACACCCTTCTGGAG 1438 4674 L V D M A R V C N T T T D R K H A D T S N H I W K L F E N F 1476 AGGTGTGTGACAGAGTCATGAACATCGTGAGCGGCTTTTCAACTCGCCGTTTTCGGATAACAGTACCAGCCTCCAGACCCATCAGCCGGTCTTTATCCAGCTGCTGCAG 4788 R C V T E S V M N I V S G F F N S P F S D N S T S L Q T H Q P V F I Q L L Q TCCGCCTTCAGAATTTACAACTGCACCTGGCCCAACCCAGCCCAGAAAAGCCTCCGTGGAATCCTGCATCAGAGCCCTGGCTGAAGTGGCCAAAAACCGTGGGATTGCGAT 1514 4902 1552 5016 1590 GCTCTTGGAGGGCCAGCATGGGACTACAGGAATATTATTGAAAAGTTACAGGATGTCGTGGCCTCTCTGGAACAGCAGTTCAGCCCCATGATGCAGGCTGAGTTCTCTGTGCTT 5130

L G G P A W D Y R N I I E K L Q D V V A S L E Q Q F S P M M Q A E F 1628 GTGGATGTGCTGTACAGCCCGGAGCTACTGTTCCCAGAAGGAAAGCGATGCCAGGATACGATGTGGTGCCTTCATGTCCAAGCTGATTAATCACACAAAGAAACTGATGGAAAAA V D V L Y S P E L L F P E G S D A R I R C G A F M S K L I N H T K K L M E K GAAGAAAAACTGTGCATTAAAATCCTCCCAGACGTTACGGGAGATGCTGGAGAAAAGACAGCTTCATGGAAGAAAGTAGCAACATTACGGAAAATCCTCCTGAATCGGGAC 5244 1666 5358 EESSTLRKILLNR 1704 E E K L C I K I L Q T L R E M L E K K D S F M AAGGGTGACCACAGCGTTGGTGTGAACGGACCTCTGTCAGGAGCCTACGCCAAGACAGCACCACGGGGGGGCCTTCACTGGACAGGATGCCGATAAGACGGGGATTTCCATG 5472 K G D H S V G V N G P L S G A Y A K T A Q V G G G F T G Q D A D K T G I S M TCCGATATCCAGTGTTTGCTGGACAAGGAAGGGGCCTCCGAACTTGTCATCGATGTCATCGTGAACACCAAAAACGACAGGATTTTTTCCGAGGGGCATCTTGCCTTGGCATTGCC 1742 5586 SELVIDVIVNTKNDRIFSEGILL 1780 G LDKEGA DIOCL CTGCTGGAAGGAAACACACAAAACCCAGAATTCTTTCTACCAGCAGTTGCATGAACAAAAAAAGTCAGAGAAATTCTTCAAAGTCCTTTACGATCGGATGAAGGCTGCTCAG 5700 EGGNTQTQNSFYQQLHEQKKSEKF FKV YDRMK 1818 5814 1856 I D L G S K K R E E D S D L M A L G P R M R D TVN CTGCATTTGAAAAGAGGGAATGAAAGGGCAGTTGACTGAGGCGTCTTCAGCCACGTCCAAAGCATACTGTGTGTACAGAAGGGAAATGGACCCAGACATTGACACGATGTGCCCG 5928 H L K E G M K G Q L T E A S S A T S K A Y C V Y R R E M D P D I D 1894 тмср GGACAGGAAGCAGGAAGTGCGGAGGAAAAGTCTGCAGAGGAAGTTACCATGAGCCCAGCCATCACTATCATGCGGCCCATCCTCAGGTTCCTGCAGTTACTGTGTGAGAATCAC 6042 1932 6156 1970 L R N Q N N K T N Y N L V СЕТ LQFLDCI 6 6 5 RELO NF GGCCTGCTGGGGGCTCTACATCAATGAGAAGAACGTAGCTCTGGTCAACCAGACCCTGGAGAGCTTGACCGAGTACTGCCAGGGCCCGTGTCATGAGAACCAGACCTGTATCGCC 6270 G L L G L Y I N E K N V A L V N Q T L E S L T E Y C Q G P C H E N Q T C I A ACTCACGAATCCAATGGGATTGACATCATCATCGCCTTGATTCTGAGTGACATCAACCCTCTGGGCAAGTACAGAATGGACCTGGTGCTACAGCTGAAGAACAACGCCTCCAAG 2008 6384 2046 6498 MESRHDSENAERILFNMRPKELVDVMKNA 2084 CTGGAATGTAACCATGGGGACGAGGAGGAGGAGGAGGATGATGGTGTTTTCTCCCAAAAGACGTTGGACACAACATCTACATCCTGGCCCACTCAGTTGGCCCGCCACAATAAAACTCCTA 6612 L E C N H G D E E G G D D G V S P K D V G H N I Y I L A H Q L A R H N K L L CAGCAGATGCTCCAAGCCT6GATCCAGATCCAGAGGAAGGGGATGAAGCCTTGAAGTACTATGCCAACCACACCGCACAGATCGAGATTGTGCGGCACCGACCATGGAGCAG Q Q M L K P G S D P E E G D E A L K Y Y A N H T A Q I E I V R H D R T M E Q 2122 6726 2160 6840 TRESKYRVFNTTERDEQGSKVNDF 2198 NICEF CAAACCGGAAGATCTCTACAACGAGATGAAAATGGCAAAAAGAAGATCAGGAACAACCCCGCCCTGTTCTGGTTCTCCAGGCACATCTCCCTGTGGGGGGAGCATCTCCTTCAACCTG 6954 2236 O T E D L Y N E M K W Q K K I R N N P A L F W F S R H ISL WGSIS GCTGTGTTCATCAACCTGGCCGTGGCTCTCTTCTACCCATTCGGGGATGACGGCGATGAAGGTACGCTCTCCCCGCTGTTCTCAGCCCTCCTTTGGGTAGCAGTGGCGATCTGC 7068 A V F I N L A V A L F Y P F G D D G D E G T L S P L F S A L L W V A V A I C ACGTCTATGCTGTTCTTCTTCTCCCAAGCCTGTGGGGCATCCGGGCCATCCGTTGTGTTCTTCATGCTCAGATCAATATACACCATCGGTCTGGGGCCAACGCTAATACTTCTTGGT 2274 7182 V G I R P F L V S I M L R S I Y T I G L G P SKP ті TL 2312 FF SMLF GCTGCCAATCTATGCAATAAAATCGTGTTCCTGGTGAGAATTTTGTGGGAAACCGAGGCACATTCACCCGAGGCAACCGAGCAGTCATTCTGGACATGGCCTTTCTTACCACGTG A A N L C N K I V F L V S F V G N R G F F T R G V R A V I L D M A F L Y H V GCCTATGTCTTGGTTTGCATGCCTGCTGGCCTCTTCGTCCACGAGTTCTTCTACAGGCAGTCATGGCAGAGAAGAGACCCTGCTGAACGTCATCAAAAGCGTCATCAAAAGCGTC 7296 2350 7410 A Y V L V C M L G L F V H E F F Y S F L L F D L V Y R E E T L L N V I K S V ACACGGAATGGCCGCTCCATCATCCTGACTGCGGTCTTGGCTCTTATCCTGGTCTACCTGTTCTCATCAGGCTGCTTTTCTTAAAGGATGACTTCACCATGGAGGTGGAC T R N G R S I I L T A V L A L I L V Y L F S I I G F L F L K D D F T M E V D 2388 7524 2426 AGATTGAAAAAACAGAACTCCAGTCACAGGTAACGACGGGGTTCCCACTATGACCTTAACTTCCATGCTGGGAACCTGCCCTAAGGAAAACTGCTCACCCACGATCCCCTCTTCG 7638 2464 7752 2502 7866 2540 7980 I D T F A D L R S E K Q K K E K I L K T T C F I C G L E R D K F D N K T V S TTTGAGGAGCACATCAAGTCAGAGCACAACATGTGGCATTACTTGTACTTCATCGTCCTGGTGAAGGGACGCAAGGAATACACAGAGGCCTGAGAGCTACGTGGCTCAG 2578 8094 FHTKSEHNMWHYLYFIVLVKVKDPTEYTGPESY 2616 ATGATCACAGAGAAGAATTTAGACTGGTTTCCTCGGATGCGAGCCATGTCACTTGTCAGCAATGAAGGTGACAGTGAGCAAAATGAGATCCGGAACCTGCAGGAGAAGCTGGAG 8208 2654 8322 2692 8436 2701 **N H H M P P H** 8550 ULTITUTIOTION AND A CONTRACT 8664 8778 CGATAGGCCTTGGACTTGTCTCACACACTGACTGCAGTGTCCATCGTGGCTGGTTGAAATTTTTTCTTCAAACTGTGGCACTGGGGTCAGCGAGACAGGAAGCCACACTCTGCT GGCTAAGTCTAAGA<u>AATTTAAAAGATTAAA</u>AAGGAGAATTGÅAAAGGGTGTCGTTAAACTTCCGAACCTTACGTGTTAACTGGACATTTTCTTCTTGGCATGAGACGGGCTCAG 8892 9006 9120 AGAATIGGCIANAGCIGCIGAGAGAAAACTCCGAGGGTAACAGCCCAACACTGAGAAGCGTTACTCGGGTGGCCTTGCCTTTAAAAGCAGGTTAATTTATATTTGAGGTGCTTGACCCAA GTGCTTGCGATCCCCCGAAGCTGAGGGGGGTGGGGGGAACCTAAGAATTGCATTGACTACGTAAGAATATGCAACATTTCAACAGCACATTTAAAGCCGTTAGGAGAACAGCTTT TCAAAATGTAGGCAGATCATAATTCCTTCCCCCCAACTGTTTTCATGTTGACTCATGCAACCAGATTTTGCAAGGGAAAAAATAACTTGGAAGGAGAGATCCTTTTGGAGGA 9234 9348 9462 TGTCTGAGGTGTCAGAGACAGACAGATCATGTAAGTGTGAGACCATCTGGTGGTGAGGGGCCACCAGCCAAGACACTACATTCTGAAACAGAGACCCTCTCTGGCCTCCAGATT 9576 9690 9804 TECACACCTAGGACCACCCAGAGTTACAAAGTGCTCTTGGACAGGGCCCTGCTCCCCTTAGCCTAAAGTGGAAAGATGCTGGTGCCTTGACCTCAGGTGTTCTAGGCTAAAAC 9918 ACTGCACCACTGACCCTCCACTCTTGTAAGAGACCTGTCTTCAGTTACCCTTAATTCTTTGCTAAAGCTTCAAACTATAATCACAAACATTTTTTTGAAGGGTATCACCAAAAT 10032 TAGGCCACGGCAGCCATTTTTAAAGCTCAGGACCATGGATACTTCGATGCGGCATTTGGTAAAAGTAAAATAGGCTGGGCGATCAGAAGGATAGGAGCTGCTTCAGTCTGCTGTAA 10146 TGATATTAAAAATACAGTCACCTCTGACTICTATGCCAGGGAAGGCTTTTTTAAATTTTAGGTTATTAAAATCCATATTGGTTAAAAGTCACAGTTCCCACTGAATAATATTTT 10602 10708

Fig. 2. Nucleotide and translated amino acid sequences of the type 2 $InsP_3$ receptor from rat. The sequence was assembled from the sequences of the cDNA clones shown in Figure 1. The deduced amino acid sequence is shown in single letter code below the nucleotide sequence, and both sequences are numbered on the right. The in-frame stop codon in the 5' untranslated region preceding the initiator codon is underlined. The position of the poly(A) tail in p115 is shown by an asterisk, and sequences that might serve as polyadenylation signals for this poly(A) tail and for the one at the end of clone p16 are underlined. The following sequence differences were noted between different cDNA clones: The C at position 2289 was a T in p547-17 (silent change); the G at position 2311 was a C in p171, changing D at position 689 to H; the G at position 2397 was a T in p171 (silent change); the T at position 2604 was a C in p547-17 (silent change); the G at position 1013 to C; the G at position 3473 was a C in p170, changing G at position 1065 to A; the T at position 4013 was a C in p170, changing L at position 1256 to P; the C at position 4770 was a T in p165 (silent change); the G at position 5929 was an A in p115 (silent change); the G at position 490 was an A in p165 (silent change); the G at position 5929 was an A in p115 (silent change); the G at position 4890 was a A in p170, changing V at position 4911 was a A in p165 (silent change); the G at position 5929 was a T in p15 (silent change); the G at position 5929 was a T in p15 (silent change); the G at position 5929 was a T in p16, changing E at position 2604 to V. In addition, clone p170 had an out-of-frame deletion from nucleotide 3311 to 3378. These sequence data are available from the EMBL/GenBank/DDBJ databases under accession number X61677 ITPR2.

$_{I}^{II}$	ŇŜĎŔŇŜŠĔĹŸĨĠĎĨvŜĹŶĂĔĠŜvŇĠĔĨŜŦĹĠĹŸĎĎŘĊŶŶDĚĔĂĠĦĹŦŇĔĔŔŔŦĔŎĊĹĔŔvĊĔŇŇŔŶŜĂŎŔŎţŶŔŔĸŎĸĸŎĠŇ MSDKMSSFLHIGDICSLYAEGSTNGFISTLGLVDĎŔĊVŶŎĔĔĂĠĦĹŦŇĔĔŔĸĔŔĔŎĊĹĔŔĸĊĔĦŇŔŶŜĂŎŔŎţŶŔŔĸŎĸŔŎĠŇ	82 82
II I	ĦŦĿĂĄĹĨĸŔĹŊŔĂĂĿĹĔŊŔŎŇĔŚĔŇŔŔĹĹĠĿIJĸŶŚĸŶĬŎĹĹĤſŔŚŇŔŶĹĨŶŇŔŔĹĔŶĹĹĔŔŇĂŇŔŨŚĹĎĄĂĠŇĔĠŚŴŦŶĨĦ TTDAVLLNKĹŇHAADLEŘKONETENRKLLGTVIQYGNVIQLLHLKSNKYLTVNKRLPALLEKNAMRVTLDEAGNEGSŴFYIQ	164 164
$_{I}^{II}$	ŶĔŴŔĹŔŜĖĠĎŊŦŮŸĠĎŔŸŶĨMĚŶŇĂĠŎŶĹĬŔĂŜŊŸĿĹĹĎŇĚĠĊŔĔŶŇĂŬŇĊŇŤŠŴŔĬŦĹĔŃŔŗŠSYREĎŸĹŔĠĠĎŶŶŔĹĔĤĂĔ PFYKLRSIGDSŸVIGDKŸŸĹŊPŸŇĂĠŎPĹŀŔĂSSHQĹŸĎŊPĠĊŇĔŸŇSVNĊŊŢŚŴKĬŸĹſĦŔŴSDŇKDDIĹŔĠĠĎŶŶŔĹĔĤĂĔ	246 246
$_{I}^{II}$	ŎĔŔĔĹŤĊĎŊŸĔŔŔŎĤŀĔĹŔŤŤŀŔŎŜĂŤŜĂŤŜĂŤŜĔŔĔĿĔŮŶŀĤĎĔĊŔĠĊĂĊĢŴŇŜĹĔŔĔŔĤĹĂŤĠŊŶĹĂĂĔĿŊĔĎŸŔŊĄŎŊĔ ŎĔĸŦĹĨĊŊĔŀŔĸĸŎŀŀŸFĹĸĨŦĠŖŎSĂŦŜĂŤŜĔŔĬĔŀĔŸĔŸĿŎŀŎŊĊŔĠĠŎġŴŊŜĹŦŔĔŔŀĹĂŤĠŊŶĹĂĂĔĿŊĔŊĿŔĿŎIJŎŢĔĔĔĊĹĔ	328 328
$_{I}^{II}$	GKTVRĎGELPTŠK KKHOÅGĚŘIMÝTĽŮŠŮPHĠŇĎľAŠLPĚĽĎAŤŤĹORAĎCŮŮPŘŇŠÝŮŘĽŘĤĽČŤŇŤŴŤŠŤŠĽPĽĎŤĚĚ FOPSVDPDQDASRSRLRNAQEKMVYSLVSVPEGNDISSIFELDPTTLRGGDSLVPRNSYVRLRHLCTNTWVHSTNIPIDKEE	409 410
$_{I}^{II}$	ĔŖĔŶŇĹŔĨĠŤĊŎŢŔĔĎŔĔĂĔĂĨŶĊŮ₽ĹSĔŶŔĎĹĎĔĂŇĎĂŇŔŶĹAŢŢVĸŔĹĔŇĠSĨŤŎŇĔŔŖĿŶŦŔĹĹĔĎĹŀŢĔŶĂDVŤŇŊĠŎ Ĕĸ₽ŸMĹĸĨġŢŚŖĹĸĔĎĸĔĸĔĸĨŸŶŸŶĔŶĸŔŨĹŊĔĸŇŊŎĸŇŶĸŎĸIJŎĸĬĸĬŎĸĬĬŎŇĔĸŔŖŶĬŔĹĹĔĎĹŀŢĔŶĸŎĿŶĬŇŊĠŎ	491 492
$_{I}^{II}$	ĎŮĨDŮŮ ITŘPŇŘĚŘŎŘĨMŘĚŎŇĨĹAŎVŤGIĨKĂPŤKEKAČEČSMĨŘĹĚDĹĊĎŎŔYĂPYKYVLŘĹČŸŘŮĹŘĤŠŎŎĎŸŘŘŇŎĔŸ DVLEVVFSKPINEROXILMREONILKOIFKLLOAPFTD CGDGPMLRLEELGDORHAPFRHICRLCYRVLRHSOODYRKNOEY	573 573
$_{I}^{II}$	ĨĂŘNĚCVŇŎSŎĨĞŸĎIĹĂĔĎŤĨŤĂĹĹĤŇŇŘŘĹĹĔŔŇĨŤĂĸĔĬEŤĚŸŠĹĹŔŔŇŔĔĚŘĚĹĎŶĹŠĎĹĊŶŠNSTAĨĚŸŤŎĔĹĬĊŔF IAKQFGFMŎKŎĨGYDVLAEDTITALLHNNRKLLEKHITAAEIDTFVSLVRKNREPRFLDYLSDLCVSMNKSIPVTQELICKA	655 655
$_{I}^{II}$	MÎSPGŇĂDÎLÎQÎKÎVSMQVÊNPMEŜS ILPDDIDDÊÊVŴIYŴIDŜŇŘÊPHGŘAIŘHLĂQEÅRĚČTŘADLEVLTŸŸŘŶQĹ VLNPTNADILLÊTKLVLSRFEFEGVSTGENALEAGEDEEEVWLFWRDSNKEIRSKSVRELAQDAKEGQKEDRDVLSYYRYQL	734 737
$_{I}^{II}$	ŇĨĔĂŔŇĊĹĎŔŎŸĨĂĨŇŎĨŠŦŎĨŚŸĎĨĨĹŔĊŸŠĎĔŚĹ₽ĔĎĨŔĂŠĔĊŔĹŇĹŀŇĤŸĎŔĎĔŎĔŚŸŸĔŶŔŶĂŔĹŴŦĔĨĔŦKĬŦĬŀĔŸ NIFARMCLDRŎŸLAINĔISGŎLDVDLILRCMSDENLPYDLRASFCRLMLHHHVDRDPŎĘQVTPVKYARLWSEIPSEIAIDDY	816 819
$_{I}^{II}$	ĎŠITDSŠRNDHŘRKĚALŤŇĚFŮĔĚÝLKEŮŮNŎPĚPĚGĎŘĚKŇŘLŤŤĚŮŮHLĂŘŇLĬÝĚGĚÝSŠĚŠELLŘLŤRTĽLÁŤLĎIŮQA DS SGASKDEIKERFAQTMEFVEEYLRDVVCQRFPFSDKEKNKLTFEVVNLARNLIYFGFYNFSDLLRLTKILLAILDCVHV	898 900
$_{I}^{II}$	PMSSYFERLSKFOD ČŠNŇŮŇŘTÌŇČVČEMŇŤŎNŮĹSŘČSIŤPVSWĎDĂOPSVHPSŘŎASĎCĖQĖĎVTŮŇĎŤŘĹŘVĬĚĬĹŎ TTIFPISKMTKGEENKGS NVMRSIHGVGELMTOVVL RGGGFLPMTPMAĂAPEGNVKOAEP EKEDIMVMDTKLKIIEILO	978 979
$_{I}^{II}$	ŶĨĨŚŶŔĨĎŶŔĨŚŸMĨŚĨŸŔĸĔĔĠĔ ŇŊĠŊĠŊDŶŚĸŜĠŦPEŢĿĹPŜĸĹŶŶĎĬŊĿĔĬĸĸŎĂĔŢĦĔĸĞŖĸĔĸŤ₽ŸŎĹĎĎĿĠĠŔŤ FILNVRĹŊŶŖĨŚĊĹĹĊĨFKREFDEŚŊŚŎŚŚĔŦŚŚĠŊĊŚĠŎŢŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎŎ	1056 1059
$_{I}^{II}$	ŶĹŔŶĹŀŔĹŀŔĤŎŶĄŶĹĿŜĠĂĿŎĹĿĔĸŔĔŜŎŖĄĔŶĿĹŎĂĔŔŎŶŎĹĹŶŚŊŎĎŶĎŇŶŔŎĬŔĄĎĹĎŎĹŔĿŦŶĔŔŜĔĹŀŔŶĔŔ FĹŔŶĹĿŔĹŦĦŔŊŶ₽ĿVSGAĿĄĹĿŖŔĔŜŎŖġĔŸĿŎġŔĸŎŶŎŎĹĿŶŦŚŎŎŶŎŊŶŔŎĬŔĄĎĹĎŎĹŔĿŦŶĔŔŜĔĹŀŔŶĔŔ ŚĊŢ	1137 1141
$_{I}^{II}$	NGÖNGEGQAKGGEEÅNEËSNLLSPVODGAŘTPQIDŠNKGNŇÝŘIŮŘĚILIŘIŠŘLČÚČN KŘCŘNÔHÔŘLLKŇŇGĂĤSŮ PHD GASGENEHKKTEEGTSKPLKHESTSSYNYRVVKEILIRLSKLCVQESASVRKSRKQQQRLLRNMGAHAV	1215 1213
$_{I}^{II}$	ŶĨŊĹĨŎĨŶŶĔŔŢŢĎĔŔŇŊĔŸŇŊĹĂŇŢĔĹŎŇĔĊŖĠŇŖŎŇŎŸĹĨŔŔŇĬŇĬĔĬŢĔŎĿĹĔĂĔŤŴŖŇĬĔŃŇŶŸĤĹĊŊĔĬŚĔŔŶŶŎĤĔ ŸĹĔĹĹŎĨŶŸĔĸĂĔŊŢĸŀŊŎĿĬŊĬĬĊĸŎIJŎŎŎŎŎŎŎŎŎŎŎĬĬĬĬĬĬĬĬĬĬĬĬĬĬĬĬĬĬĬĬĬĬĬ	1296 1295
$_{I}^{II}$	ŶŴĊĨĔĨŴĠŔŀŴĿŶĽŖŀĹŎŢĨŶŔĂŊĊŔŸVŔŔĊŎĎŀŴŀĬĔĹĿŇĠĊĔĎŶĹĿŀŶŴĎŔĂŜŦĿĬĹIJŇŴŀĊŜĔŔĸŔĠĎĔSĠŶĹ AŶĤĬŢ VHCIETHGRNVQYIKFLQŢĬVKAEGKFIKKCQŊĦVMAELVNSGEDVLVFYNDRASFQŢLIQHMRSERDRMDENSPLFMYHIH	1377 1377
$_{I}^{II}$	10£11Aactecknývtěikcnšliþiddivkývtrodciþevki av nárvákovátevénkeivtsná lúklfenflýðma lvellavctecknyvteikcnsliþiddivkyvthedciþevki av inflnhgyvdtevemkeivtsnhmyklfenflydic	1459 1459
$_{I}^{II}$	ŔvĊŇŢĨŢĎŔŔĤĂĎŢŦĹĔŖĊŶŢĔŚŶ ŇŊĬŶ ŚĠ ŀŦŊŜŀŦŚĎŊŠŢ ŚĹŎŢĦŎĔŶĔſŎĹĹŎŚĂĔŔIJŶŊĊŢŴ₽ŊĔĄŎŔĂŠŶĔŠĊĬŔĂĹA RACNNTSDRKHADSVLEKYVTEIVĦSIVTT FFSSPFSD QSŢŢĹŎŢŔŎ₽ŸŦŶŎĹĹŎĠVFRVYHĊŊŴĹĦŶSŎĶASVESCIRVLS	1541 1541
$_{I}^{II}$	EVÄKNRGIAIPVÖLÖSÖVÄTLEMKNÄSSTÜÖRAÄÄGÄRÄÄSGPREKEALGGPANÖYRÄIIEKLÖÖVVASLEOOFSEMMÖÄ DVAKSRAIAIPVDLOSOVNNLELKSH NIVOKTAMMIRLSARNAARROSVL AASRDYRNIIERLODIVSALEORLRPLVOA	1623 1621
$_{I}^{II}$	ĚFŠŮĹŮĎŮĹYSĚĽĹĚŘĚGSĎĂŘIRČ ĠAĚMSŘĹĨNŘŤŘKĹM ĚKĚĔŘĹČĬŘIĹŎŤĹŘĚŇLEŘKDSFMĚE ELSVLVDVLHRPELLFPENTDARRKCESGGFICKLIKHTKQLLEENEEKLCIKVLQTLREMMTKDRGYGEKQISIDELENAE	1691 1703
$_{I}^{II}$	SSTLRKILINRYFKGDHS VGVNGPLS GAYAKTAQVGGGFTGQDAD LPQPPEAENSTEELEPSPPLRQLEDHKRGEALRQILVNRYYGNIRPSGRRESLTSFGNGPLSPGGPSK PGGGGGGGGGSGST	1736 1784
$_{I}^{II}$	ĸŢĠŢĨŚŊŊIJŎĊĹĹĎŔĔĠĂŜĔĹŶĨĎŸĬŸŇŢĸŊĎŔŢĔŚĔĠĨĹĹĠĨĂĹĹĔĠĠŇŢŎŢŎŊŠĔŸŎŎĹŀĔŎŔŔŠĔŔĔĔŔŶĹŶĎŔŴŔAĂ ŚRGEMŚLAEVŎĊHĹDKEGAŚŊĹŸĬŊĹĬĦŊAŚŚŊŔŸĔĦĔŚĨĹĹAĨAĹLEGGŊŢŢĬŎŀŊĔŢĔĊŔĹŦĔŎŔĸŚĔĸĔŗĔĸŶŦŶŊŔŴŔŎĂ	1817 1866
$_{I}^{II}$	ŎĸĔĨŖŚĨŶĨŶŇĨĬĎĴĊŚŔĸŖĘĘŊŚIJŀĂLGPŔĦŖŲŖŊSIJĦĿĸĔĢĦĸĊŎĹĬĔĂŜŚĂĨŚŔĂŸĊŲŸŔŔĔMĎŔĎĬĎŢMĊŖĊĢĖĄĠ ŎŎĔĬĸĂŦŲŦŲŇĨŚŊĹĠĸŔĸĸŊDEŲŊŖŊA PŚŖĸĸĸĸĔſŢŢŨĬĔĔŲŔŊŎIJĹĿĔĸŚĸĂŦŖĸĸŦŢŢŖŖĔĂŊĎ ŊĦŶŎŚĠ ĔĠŢ	1899 1945
$_{I}^{II}$	SĂ EEŘSAEEVTŇŠPAĬŤĬŇRĚĬĹŘĚLŎLĹĊĖŇŇŇŘELŎŇĚĹŘNŎŇŇŘŤŇÝŇĹVĊĖŤĹŎĚĹĿĊĊŠŤŤĠĊĹĊĹĽŎĹŸĬŇĚŘ QATTDKAKDDLEMSAVITIMQPILRFLQLLCENHNRDLQNFLRCQNNKTNYNLVCETLQFLDCICGSTTGGLGLLGLYINEK	1980 2027
$_{I}^{II}$	ŇVÄLVNÖTLESLTEVCÖGEPCHENÖTCIÄTHESNÖIDIIIALILSDINPLGKYRMDLVLOLKNNÄSKLLLÄIHESRHDSENÄE NVALINQTLESLTEVCOGPCHENONCIATHESNGIDIITALILNDINPLGKKRHDLVLELKNNASKLLLAIMESRHDSENÄE	2062 2109
$_{I}^{II}$	ŔĨĹĿŇŇŔŸŔĔĹŮDŮŇŔŇĂŶŊŎĠĿĔĊŊŀġĠĿĔĠĠŊĠĊŸŶŔŊĊĬŔŇŶĬŶĬĹĂŀŎĹĂŔŀŇŔĿĹŎĢŇĹŔŶĠSDPEĿĠĎĔĂĹĸŸŶĂ RILYNMRŸKELVEVIKKAYŀŊĠ EVEFEDGENGEDGAASPRNVGŀNIYILAŀŊĹARŀŀNKELŊŤMĹKPĠ GQVDGDEALEFYA	2143 2189
$_{I}^{II}$	ŊŴĨŻŎĨĔĨŮŔĦĎŔĨĤĔŎĨŮŕ₽Ů₽ŊĨĊĔŕĹĨŖĔŠŔŸŔvFŊŤĨĖŔĎĔŎĠŠŔVŇĎŕFŎŎŢĔĎĹŸŇĔŇŔŴŎŔŔĬŔŊŊŶAĬĿŦŴFŠŔĦ KHTAQIEIVRLDRTMEŎIVFPVPSICEFLĨKESKLRIYYTTERDEQGSKINDFFLŔSEDLFNEMNŴŎKKLRAQPVLYWCARŊ	2225 2271
$_{I}^{II}$	ĬŜLŴĠŜĬŜĔŇĹĂŶFIŇĹĂŶĂĿĔŸĔĔGDDGDĔĠŤĹSĚĿĔŠAĹĹŴVĂVAĽĊTSMLFFFSŘĚVĠĨŘPFLVŠIMĹŔSĬŸŢĬĠĹĠĔŤ <u>MSEWSSISENLAVIMNLLVAFFYF</u> KGVRG GTLE <u>PHWSGLLWTAMLISLAIVIAL</u> PKPHGIR <u>ALIASTILRLIE</u> SYGLOPT	2307 2352
II I III	LILLGAANLCNKIVFIVSFVGNRGTFTRGYRAVILDHAFLYHVAYVIVCHLGLFVHEFFYSFLLFDLVYREETLLWVIKSVT LFLLGAFNVCNK <u>IIFLMSFVGNCGTFTRGYRAMVL</u> DVEFLYHLLYLLICAMGLFVHEFFYSLLLFDLVYREETLLWVIKSVT M4	2389 2434 13
II I III	ŔŇĊŔSÌĨĹĽŔAVĹĂĹĨĹŮŶĹĚŜĨIĠFĹĔĹŔĎĎĔŢMĚŸĎŔĽĸŇŔŢ₽VŢĠNDĠVPŢMŢLIŢSMLGŢĊŖĶĘ NČ ŠPŢIPSSNAAG RNGRPIILTAALALILVYLFSIVGYLFFKDDFILEVDRLPNETAGPETGESLANDFLYSDVCRVETGENCTSPAPKEELLPV RNGR <u>SILLTALILVYLFSIVGFLFL</u> KDDFILEVDRLPNNHSTASPLGMPHGAAAFVD TCSGDKMDCVSGLSVPEVLE MG	2468 2516 93
II I III	ÉGGÉDGI ÉRTÉDÍLÍMÉLÍVÍVÍNOGLÍNNGGGVEDVÍRPÍSKDÉPLÍFAÁRÚVÝDÍLÍFFFIVÍLÍVÍNÍLÍFEVÍLÍDÍFÁDÍRS EETEQDK EHTCETLLMCIVTVLSNGLRSGGGVEDVLRKPSKEEPLFAARVIYDLL <u>FFFMVIIIVLNLIFGVII</u> DTFADIRS EDRELDSTERACD <u>TLLMCIVTVMNNGLRNGGGVE</u> DILRKPSKDESLFPARVVYD <u>LL</u> MB	2549 2597 147
$_{I}^{II}$	ĔŔŎŔŔĔĸĬĹŔŦŤĊĔĬĊĞĹĔŔĎŔĔŇŔĬŶĸĔĔĔĔĤĬŔĸĔĤŇŴĤŶĹŸĔĬŬĹŨŔŨŔĎŢĔŶŤĜĔĔŠŶŨĂŎĤĬŢĔĸŇĹĎŴĔĔŔŇŔĂ ĔŔŎŔŔĔĔĬĹŔŦŤĊĔĬĊĞĹĔŔĎŔĔŊŇŔŦŬŸĨĔĔĔĦĬŔĸĔĔĦŇſŴĤŶĹŸĔĬŨĹŨŔŨŔĎŦĔŶŤĜĔĔŠŶŨĂŎĤĬŢĔĸŇĹĎŴĔĔŔŔŔĂ	2631 2679
$_{I}^{II}$	ŘŠLŮŠNEGDSŘÔŇĚ IŘŇLŎĔŘĹĔŠŤŘSLŮK QĽŠĠŎĹAĚLŘEŎŘŤĚŎŘŘNŘŎŘLČFĹČSNTĚŘEŇHHMPPH★ MSLVSSDSEGEQNELRNLQERLESTMKLVTNLSGQLSELKDOMTEORKOKORIGLLGH PPHMNVNPQOPA★	2701 2749

Fig. 3. Alignment of the amino acid sequence of the rat type 2 $InsP_3$ receptor (top line) with that of the type 1 receptor (second line, from Mignery *et al.*, 1990) and the partial sequence of the putative human type 3 $InsP_3$ receptor (third line; C.L.Newton, G.A.Mignery and T.C.Südhof, in preparation). Identical residues are marked by dots above the sequence. Sequences are shown in single letter code and are numbered to the right. Amino acids belonging to putative transmembrane regions are underlined and the transmembrane regions are labeled M1 to M8. The position of the stop codon is indicated by an asterisk.



Fig. 4. Immunoblot analysis of proteins specified by the aminoterminal sequences of the type 1 and type 2 receptors expressed in COS cells. The amino-terminal 1081 and 1078 residues of the type 1 and type 2 InsP3 receptors, respectively, were cloned into an expression vector fused to a sequence encoding the last 12 amino acids of the 116 K proton pump subunit. Cytosol from COS cells transfected with the type 1 InsP₃ receptor expression construct (pIP₃R-Stop1081, lane 1), type 2 InsP₃ receptor construct (pIP₃R2-Stop1078, lane 2) or with control DNA (salmon sperm DNA, lane 3) were analyzed by immunoblotting using an antibody against their common carboxyterminal epitope followed by a peroxidase-labeled secondary antibody. Expression of the constructs results in soluble receptor proteins containing the full-length binding sites of the two InsP3 receptors and ending in the same carboxy-terminal sequence.



Fig. 5. Determination of the binding affinities of the ligand binding domains of the type 1 and type 2 InsP3 receptors. Cytosol from COS cells transfected with constructs encoding the ligand binding domains of the type 1 (closed symbols) and type 2 InsP₃ receptors (open symbols) were used in binding-competition assays whereby the displacement of 3.3 nM [32P]InsP3 by unlabeled InsP3 was determined. The line through the points represents the best fit to the data calculated using GraphPAD version 3.1 software, resulting in the determination of affinities for the type 1 and type 2 receptors of 89.5 nM and of 27.0 nM, respectively. The experiment was repeated twice with similar results. COS cells transfected with control DNA showed no measurable InsP₃ binding (Table I).

Table I. Binding specificities of type 1 and type 2 $Ins(1,4,5)P_3$ receptors

Competitor (10 µM)	$pIP_3R-Stop1081$ (c.p.m./mg × 10 ⁻³)	Percentage of control	pIP_3R2 -Stop1078 (c.p.m./mg × 10 ⁻³)	Percentage of control	Salmon sperm DNA
_	22.86 ± 1.36	100.0	11.33 ± 0.59	100.0	0.00 ± 0.17
Inositol 1,4-bisphosphate	22.86 ± 1.17	100.0	13.61 ± 0.50	120.1	-
Inositol 4,5-bisphosphate	16.03 ± 0.47	70.1	7.71 ± 0.31	68.0	-
Inositol 1,4,5-trisphosphate	0.00 ± 0.18	0.0	0.00 ± 0.15	0.0	0.00 ± 0.96
Inositol 2,4,5-trisphosphate	0.45 ± 1.74	2.0	0.60 ± 0.48	5.3	_
Inositol 1,3,4,5-tetrakisphosphate	8.68 ± 1.20	38.0	5.05 ± 0.93	44.6	_
Inositol 1,4,5,6-tetrakisphosphate	20.13 ± 1.06	88.1	11.59 ± 0.55	102.3	_
Inositol 1,3,4,5,6-pentakisphosphate	12.17 ± 1.29	53.2	7.22 ± 0.61	63.7	-
Inositol hexakisphosphate	17.86 ± 0.49	78.1	9.46 ± 1.11	83.5	-
Heparin (5 µg/ml)	2.62 ± 0.94	11.5	3.58 ± 1.28	31.6	-
Heparin (100 µg/ml)	0.00 ± 0.50	0.0	0.00 ± 0.78	0.0	_

³H-InsP₃-binding (25 nM) was measured with 50 µg protein of the cytosol of COS cells transfected with the indicated DNAs. Quantitative immunoblotting showed that the expression of the pIP₃R-Stop1078 was 77% of that of pIP₃R-Stop1081 (2.95 and 3.81×10^6 c.p.m. ¹²⁵I-labeled antibody binding per mg protein, respectively) with no measurable InsP3 binding or receptor expression in COS cells transfected with control DNA.

Discussion

In this study we have isolated and sequenced a set of overlapping cDNA clones encoding a novel InsP3 receptor. The complete primary structure of the new receptor was determined and consisted of 2701 amino acids and was found to be homologous over its entire length to the cerebellar InsP₃ receptor—the only InsP₃ receptor previously characterized. Expression of the amino-terminal domains of the novel receptor (referred to as type 2 InsP₃ receptor) and of the cerebellar receptor (referred to as type 1 receptor) in COS cells demonstrates that both bind InsP3 with high affinity and similar specificities, although the type 2 receptor has a significantly higher affinity than the cerebellar type 1 receptor. Together our results demonstrate the presence of different types of InsP₃ receptors in brain tissue, whose sequences and properties suggest that they may have different InsP₃ binding affinities and regulatory characteristics.

Alignment of the sequences of the type 1 and type 2 InsP₃ receptors reveals a scattered distribution of identical and diverse sequences with an overall sequence homology of 69%. The structural design of the two InsP₃ receptors is similar, suggesting that they are comprised of similar 3204

functional domains. We have previously proposed a domain model for the cerebellar InsP₃ receptor that divides its sequence into a ligand binding domain, a coupling domain transducing the ligand binding signal, and a Ca²⁺ channel domain (Mignery and Südhof, 1990). Analysis of the sequence similarity between the receptors as a function of these domains suggests the ligand binding site is the most conserved region between the two receptors (Figure 6). This agrees well with the similar binding characteristics of the two receptors and suggests that their primary functional differences may be localized to the coupling domain and the Ca²⁺ channel domain.

The coupling domain separating the ligand binding domain from the putative channel domain is the least similar domain between the two types of receptors. The coupling domain contains the cAMP-dependent phosphorylation sites of the cerebellar InsP₃ receptor, suggesting that it is the principal target of regulatory signals in the InsP₃ receptor (Mignery et al., 1990; Ferris et al., 1991b). The lack of conservation between the two types of InsP₃ receptors in this region suggests that the two receptors may be subject to different types of regulation. In addition, significant sequence



Fig. 6. Domain model of the $InsP_3$ receptors and sequence identities between the type 1 and type 2 $InsP_3$ receptors in the different domains. The three principal domains of the $InsP_3$ receptor are described on top (Mignery and Südhof, 1990). The eight putative transmembrane regions in the carboxy-terminal fourth of the receptors are numbered and their position indicated by vertical lines. The sequence identity between the two types of $InsP_3$ receptors in each domain is shown on the bottom.

differences are observed in the putative Ca^{2+} channel domain, particularly in the loops separating transmembrane regions. These differences suggest that the Ca^{2+} gating characteristics of the two receptors may also be different.

Our results demonstrate the presence of multiple types of $InsP_3$ receptors co-expressed in brain. Many of the proteins involved in signal transduction at the cell surface have been shown to be present in multiple isoforms with different regulatory properties but this is the first such demonstration for a protein functioning downstream of the generation of $InsP_3$. What is the biological relevance of the presence of different types of $InsP_3$ receptors? We would like to suggest three major hypotheses that are not mutually exclusive and are based on examples of differentially regulated isoforms of proteins involved in signal transduction cascades.

- 1. Different types of $InsP_3$ receptors may be functionally similar but have different regulatory properties. This would result in differences of the properties of intracellular Ca^{2+} stores dependent on which $InsP_3$ receptors are expressed. This hypothesis is supported by the fact that the putative coupling domains of the $InsP_3$ receptors that connect their ligand binding sites to the transmembrane regions is the least conserved between the receptor forms, suggesting that they may indeed be subject to differential regulation.
- 2. Different InsP₃ receptors could have different intracellular functions specified by different intracellular localizations. Currently it seems unlikely, although not excluded, that an InsP₃ receptor might be present in a subcellular membrane other than the endoplasmic reticulum, such as the plasma membrane (Penner *et al.*, 1988). It is more likely that there are specialized subcompartments of the endoplasmic reticulum which may contain differentially regulated Ca²⁺ stores (Lechleiter *et al.*, 1991; Villa *et al.*, 1991).
- 3. Different types of $InsP_3$ receptors could have different intracellular functions analogous to the two forms of the ryanodine receptor. Ryanodine receptors, similar to $InsP_3$ receptors, release Ca^{2+} from intracellular stores. The ryanodine receptors from cardiac and skeletal muscle differ from each other in the coupling between membrane depolarization to Ca^{2+} release, and their sequences are 65% identical (Takeshima *et al.*, 1989; Otsu *et al.*, 1990; Zorzato *et al.*, 1990). It is possible that of the different types of $InsP_3$ receptors, one could be autonomous in the cell interior whereas the other similar to the skeletal muscle ryanodine receptor could be coupled to the plasma membrane. The low abundance of the type 2 $InsP_3$ receptor would support such a model.

All these hypotheses (independent of which will be proved to be correct) imply that Ca^{2+} signaling induced by InsP₃ is much more complex than that envisioned by a single type of receptor. Clearly Ca^{2+} -release from intracellular stores is not a uniform process but dependent on receptor types as well as secondary regulatory events.

Materials and methods

cDNA cloning and DNA sequencing

A rat cDNA library was screened as described (Südhof et al., 1987; Mignery *et al.*, 1990) with an oligonucleotide complementary to the sequence of the last putative transmembrane region of both the ryanodine and the InsP₃ receptors (oligonucleotide sequence: CAGCTGCAGGACGATGATGAT-GACCATGAAGAAGAA). Of the 15 clones isolated, most encoded the cerebellar InsP₃ receptor but two clones upon sequencing were found to be distinct from but homologous to the cerebellar InsP₃ receptor. Although both of these clones terminated in poly(A) tails, one clone had a 1.9 kb longer 3' untranslated region than the other, suggesting differential use of polyadenylation signals. The two clones were fully sequenced and further 5' clones were isolated using oligonucleotides based on the sequences of the 5' ends of these clones. The complete receptor was cloned in this manner on nine overlapping cDNA clones, several of which were isolated more than one time.

Polymerase chain reaction cloning of $InsP_3$ receptor related transcripts from a human kidney library was performed as described (Perin *et al.*, 1991) using the oligonucleotide described above as the specific primer and primers from the flanking sequences of the vector as the second primer. Only two transcripts with homology to the cerebellar $InsP_3$ receptor were isolated, one of which was the human homologue of the cerebellar receptor, whereas the second encoded a novel sequence. DNA sequencing was performed by the chain termination method (Sanger *et al.*, 1977) either manually using ³²P- and ³⁵S-labeled nucleotides or automatically on an ABI 370A sequencer using single-stranded M13 subclones of the cDNA clones. Sequences were analyzed on an IBM-AT computer using Microgenie software and searched against GenBank release 64 and NBRF release 25, with no significant homology observed with any sequences in the databanks except for the $InsP_3$ receptor and ryanodine receptor.

Expression of the ligand binding sites of type 1 and type 2 $InsP_3$ receptors by transfection

pIP₃R2-Stop1078 is a mammalian expression vector in which the cytomegalovirus promoter drives the expression of the first 1078 amino acids of the type-2 InsP3 receptor. This sequence is followed by the 12 carboxy-terminal acids of the 116 K subunit of the proton pump (Perin et al., 1991) against which we obtained an antipeptide antibody that was used both to visualize and to quantify expression. pIP₃2-Stop1078 was constructed by cloning the 2.45 kb EcoRI-KpnI fragment from p567-13 into pCMV2 (kind gift of Dr D.W.Russell, University of Texas Southwestern Medical Center, Dallas), followed by the 1.04 kb KpnI-PstI fragment from pI71 and by an oligonucleotide encoding the carboxy-terminal epitope. The corresponding type-1 InsP3 receptor expression vector pIP3R-Stop1081 was described previously (Mignery and Südhof, 1990). Purified DNA from both vectors was transiently transfected into COS cells and expression was analyzed by immunoblotting using peroxidase-labeled secondary antibodies and quantified using iodinated secondary antibodies and an Ambis radioanalytic imaging system. The cytosol from transfected cells was prepared as described (Mignery and Südhof, 1990) and used for binding measurements. All binding measurements were performed using the PEG precipitation assay (Chadwick et al., 1990) and tritiated InsP₃ (17 Ci/mmole) (NEN-Du Pont) except for the assays used for the determination of the binding affinities in which 32 P-labeled InsP₃ (155 Ci/mmole) was used because of the required higher sensitivity. COS cells transfected with salmon sperm DNA were used as negative controls in all experiments. Binding data were evaluated and affinities calculated using GraphPAD InPlot version 3.1 software.

RNA blotting experiments

Total RNA was isolated from rat tissues and used for RNA-blots as described (Perin *et al.*, 1986). All blots were probed with uniformly labeled single-stranded DNA probes generated on M13 templates, and washed at high stringencies.

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