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Structuring and texturing gluten-free pasta: egg albumen or whey proteins?

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3 1 **Structuring and texturing gluten-free pasta: egg albumen or whey proteins?**
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3 16 **Abstract**
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5 17 The effects of adding egg albumen or whey proteins to pasta made from parboiled rice flour were
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7 18 investigated. Pasta quality was evaluated in terms of color, of furosine content, and of cooking
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9 19 properties (water absorption, cooking loss, and consistency at the optimal cooking time). The
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11 20 surface heterogeneity of the cooked and uncooked materials was studied, and some starch properties
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13 21 (pasting properties, starch susceptibility to alpha-amylase hydrolysis) were assessed, along with the
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15 22 features of the protein network as determined by conditional solubility studies and with
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17 23 ultrastructural features of the cooked products. Egg albumen improved pasta appearance, and gave a
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19 24 product with low cooking loss, firmer and nutritionally more valuable than the other ones. In
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21 25 albumen enriched pasta, small starch granules appear homogeneously surrounded by a protein
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23 26 network. In the uncooked product, the protein network is stabilized mostly by hydrophobic
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25 27 interactions, but additional disulfide interprotein bonds form upon cooking. Thus, addition of 15%
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27 28 liquid albumen to parboiled rice flour results in significant improvement of the textural and
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29 29 structural features of rice-based gluten-free pasta.
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32 **Keywords:** gluten free pasta, proteins, cooking behaviour, ultrastructure
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34 **Abbreviations:** BU, Brabender units; DTT, dithiothreitol; EA, egg albumen; FV, final viscosity;
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36 HV, hot viscosity; MVAG, micro-viscoamylograph; PaEA, rice pasta with egg albumen; PaPR,
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38 rice pasta; PaWP, rice pasta with whey proteins; PR, parboiled rice; PT, pasting temperature; SB,
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40 setback; WP, whey proteins.
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39 Introduction

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41 The replacement of gluten functionality in gluten-free products represents a major technological
42 challenge. **Gluten-free products are seeing a growing demand, due to the increase in the incidence
43 of pathologies of various type, all linked to some form of intolerance to gluten (1). At a prevalence
44 of the coeliac disease approaching 1/200 in some European countries (2), the potential market for
45 gluten-free products is of interest also for non-specialized industries.**

46 Up-to-now, two main approaches have been proposed and recently reviewed by Marti &
47 Pagani [3]. One is focused on choosing appropriate processing conditions able to create a new and
48 **efficient** arrangement of starch components in the final product [4-6]. The other approach is based
49 on the choice of appropriate ingredients and/or additives (mainly hydrocolloids and emulsifiers)
50 suitable for inducing a cohesive structure that overcomes the absence of gluten. Generally, the
51 additives are obtained through chemical synthesis or are extracted from sources other than cereals.
52 Despite the amply reported positive effects of the addition of emulsifiers and hydrocolloids [7-12],
53 the consumers often associate their presence in gluten-free pasta to a “non-natural” food.

54 Thus, the use of proteins as structuring building ingredients could be an interesting approach
55 for producing gluten-free pasta, also because of their positive role in improving the nutritional value
56 of the product [13]. The effects of whey proteins were recently investigated on rheological and
57 mechanical properties of fresh handmade tagliatelle from pseudocereal flours [14]. **Moreover, the
58 addition of whey proteins to sweet potato gave high quality pasta with strong starch-protein network
59 formation leading to slow starch digestibility (15,16).**

60 The addition of egg white (0.25%) and casein (0.25%) to a rice dough was associated with
61 improved handling and processing [12]. Recent reports deal with the effectiveness of egg white
62 powder (6%) and emulsifiers (1.2%) in improving texture and cooking quality of gluten-free pasta
63 prepared from buckwheat, amaranth, and quinoa flour blends [17].

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3 64 In the present work, we investigated the role of texturing proteins (egg albumen and whey
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5 65 proteins) in defining the overall quality of rice pasta in the absence of other additives. The effects of
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7 66 these proteins on the starch/protein interactions and on the overall pasta structure and cooking
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9 67 performance was also investigated.
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12 13 14 69 **Materials and Methods**

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16 17 18 71 **Materials**

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23 73 Parboiled milled rice (PR; Indica type cultivar of commercial origin; total starch: 85.9% db; protein:
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25 74 7.1% db; lipid: 1.0% db; ash: 0.89% db; amylose: 25% db) used in this study was provided by Riso
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27 75 Viazzo s.r.l. (Crova, Italy). Kernels were ground in an industrial plant (Riso Viazzo s.r.l., Crova,
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29 76 Italy) to produce flour with particles smaller than 250 μm .

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32 77 Liquid egg albumen (EA) (11% protein, 0.8% carbohydrates, 0.03% fat) was purchased
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34 78 from Ovopel s.p.a. (San Giovanni in Croce, Cremona, Italy). Ultrafiltered spray-dried whey proteins
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36 79 (WP) (80% protein, 6% lactose, 6% fat) were provided by Tosi & G. s.r.l. (Vimercate, MB, Italy).
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39 40 41 81 **Pasta production**

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45 83 Experimental pasta samples were produced using the pilot-plant at DeFENS, University of Milan.
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47 84 Three different pasta samples were prepared starting from the same parboiled rice flour (PR) with
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49 85 or without the addition of texturing proteins: 1) PaPR, pasta from rice; 2) PaEA, rice pasta with egg
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51 86 albumen; 3) PaWP, rice pasta with whey proteins. The amount of EA (15g/100g flour) and WP
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53 87 (3g/100g flour) were chosen to produce pasta samples with a comparable protein content. A
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55 88 conventional pasta-making process was applied according to Marti et al. [4]. Rice flour and water
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57 89 were blended in order to produce a mixture with a final moisture of 40%. When WP were used, a
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3 90 previous suspension in water was prepared and mixed until complete solubilisation of proteins.
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5 91 Liquid albumen was added to the flour-water mixture during mixing.
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7 92 For all the pasta samples, the flour-water mixture was formed into pasta by conventional
8
9 93 extrusion, carried out in a lab scale extruder for semolina pasta (20 kg/h; MAC 30, Italtast, Parma,
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11 94 Italy), keeping the extrusion temperature at 50 °C. Samples were formed into macaroni shape (7
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13 95 mm external diameter) and dried in an experimental drying cell, by using a low-temperature drying
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15 96 cycle (50 °C for 14 hours) [4]. All the samples were stored at room temperature until analysed;
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17 97 when appropriate, pasta samples were ground to 500 µm with a laboratory mill (IKA
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19 98 Universalmühle M20, Staufen, Germany), fitted with a water cooling jacket in order to avoid
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21 99 overheating during grinding.
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27 101 Ultrastructural observations
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31 103 Ultrastructural observations were carried out on cooked and lyophilized pasta. Samples were
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33 104 mounted on aluminum stubs, and sputter-coated with gold. Pasta ultrastructure was imaged in the
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35 105 Scanning Electron Microscope (SEM) LEO438 VP (LEO Electron Microscopy Ltd., Cambridge,
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37 106 UK), under high vacuum conditions (10^{-4} Pa) at an accelerating voltage of 15 kV.
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43 108 Color analysis
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47 110 A reflectance color meter (CR 210, Minolta Co., Osaka, Japan) was used to measure the lightness
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49 111 and saturation of the color intensity of rice flours by utilizing the CIE-LAB uniform color space
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51 112 procedure. CIE-LAB-System color values L^* , a^* , and b^* as measures of lightness, redness-
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53 113 greenness, and yellowness–blueness, respectively, were recorded for each sample. Each
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55 114 measurement was replicated five times and the average value was used.
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3 116 Furosine determination
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7 118 Furosine was determined by HPLC after acid hydrolysis according to Resmini et al. [18], and
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9 119 expressed as mg/100 g of protein. Protein content was determined according to the AACC 46-11.02
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11 120 official method [19].
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15 122 Cooking behaviour
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19 124 Cooking loss was evaluated by determining the amount of solids lost into cooking water according
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21 125 to the AACC 66-50.01 official method [19]. An aliquot of pasta (20 g) was cooked at the optimal
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23 126 cooking time for each sample in boiling natural water (pasta:water ratio = 1:10) with no salt added.
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25 127 The optimum cooking time of rice pasta was evaluated as the time required for disappearance of the
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27 128 central core when gently squeezed between two glass plates, according to the AACC 66-50.01
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29 129 official method [19].
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32 130 After cooking, pasta was drained, water was recovered, and its level brought back to the
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34 131 initial volume. Twenty-five ml of cooking water were then collected and dried to constant weight at
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36 132 105 °C. The residue was weighed and the dry matter reported as percentage of the starting dry
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38 133 material. Results were expressed as grams of matter loss/100 g of dry pasta. Weight increase of
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40 134 pasta during cooking was evaluated by weighing pasta before and after cooking. The results were
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42 135 expressed as the ratio between the weight increase and the weight of uncooked pasta.
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45 136 Textural characteristics of the cooked pasta were determined by using a Texture Analyzer
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47 137 TA.HD-plus (Stable Micro System Ltd., Godalming, United Kingdom), calibrated for a load cell of
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49 138 2.5 kN. The analysis was repeated at least five times: for each replicate, 6 pieces of pasta were
50

51 139 cooked at the optimal cooking time and analyzed using a Kramer cell (test speed of 0.67 mm/s).
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53 140 Firmness (expressed in Newton) was calculated by Texture Exponent TEE32 software (v. 3.0.4.0).
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3 142 Image analysis and surface heterogeneity
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7 144 The lengthwise and cross section images of 20 macaroni for each sample were taken before and
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9 145 after cooking at the optimum cooking time, using a flatbed scanner (Epson Perfection 3170 Photo,
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11 146 Seiko Epson Corp., Japan), at 300 dpi (dots per inch) of resolution and a color depth of 24 bits in
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13 147 standard conditions. During the acquisition, samples were covered with a black box to avoid
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15 148 reflections during acquisitions. The images were saved as TIFF format and then processed using a
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17 149 dedicated software (Image Pro-Plus 4.5.1.29, Media Cybernetics Inc, UK). The assessment of
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19 150 surface texture of uncooked and cooked products was performed on a surface of 50pxl * 30pxl
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21 151 extracted from the images of the macaroni [20]. After conversion in 8-bit grayscale, the surface
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23 152 texture of each image was evaluated and expressed in terms of heterogeneity (HTG). This
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25 153 parameter is defined as the fraction of pixels whose intensity value deviates more than 10%
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27 154 compared to the average intensity of the entire image: a value equal to 0 corresponds to a
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29 155 homogeneous (smooth) surface, whereas a value equal to 1 corresponds to a heterogeneous (rough)
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31 156 surface.
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38 158 Starch properties: α -amylase susceptibility and pasting properties
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43 160 The starch susceptibility to α -amylase hydrolysis was determined by evaluating the damaged starch
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45 161 content (AACC 76-31, [19]; “Starch Damage Assay Kit” by Megazyme International Ireland Ltd.,
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47 162 Bray Business Park, Bray, Co. Wicklow, Ireland). Results are the average of at least four replicates.
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49 163 Pasting properties of rice flours were measured according to Marti et al. [4] using a Brabender
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51 164 Micro-Visco-AmyloGraph (MVAG) (Brabender OHG, Duisburg, Germany). Fifteen grams of
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53 165 sample were dispersed in 100 mL of distilled water, scaling both sample and water weight on a 14%
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55 166 flour moisture basis.
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3 167 The suspensions were subjected to the following temperature profile: heating from 30 up to
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5 168 95 °C, holding at 95 °C for 30 min, cooling from 95 to 50 °C, holding at 50 °C for 30 min and
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7 169 cooling to 30 °C. A heating/cooling rate of 3.0 °C/min was applied. The following indices were
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9 170 considered: pasting temperature (PT, °C; temperature at which an initial increase in viscosity
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11 171 occurs), hot viscosity (HV, Brabender Units, BU; maximum paste viscosity achieved during the
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13 172 heating cycle), final viscosity (FV, BU; paste viscosity achieved at the end of the cooling cycle),
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15 173 and setback (SB, BU; increase in viscosity during cooling and corresponding to the difference
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17 174 between the final viscosity and the viscosity reached after the first holding period). Measurements
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19 175 were performed at least in duplicate and the average value was used.
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177 Properties of the protein network: protein solubility

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179 Protein solubility in native and denaturing conditions was determined by suspending 0.5 g of finely
180 ground uncooked or cooked sample in 10 mL of 50 mM phosphate, 0.1 M NaCl, pH 7.0, containing
181 6 M urea or 6 M urea and 10 mM dithiothreitol (DTT) when indicated. Suspensions were stirred for
182 60 minutes at 25 °C. After centrifugation (10000 × g for 20 min, 20 °C) the amount of protein in the
183 supernatant was determined by a dye-binding method [21] using bovine serum albumin as a
184 standard. Results were expressed as mg proteins/g pasta.
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186 Statistical analysis

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188 Analysis of variance (ANOVA) was performed on the data adopting the least significant difference
189 (LSD). Data were processed by Statgraphic Plus for Windows v. 5.1. (StatPoint Inc., Warrenton,
190 VA, USA).

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193 Results and Discussion

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195 Color and furosine content

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197 The color is the first quality attribute the consumer takes into consideration to accept or refuse
198 pasta-products. Luminosity and chromatic indices are affected by both processing conditions and
199 formulation, in terms of raw materials characteristics and/or presence of specific ingredients
200 [22]. The color indices of experimental rice pasta are shown in Table 1.

201 PaPR - made solely by rice flour and used as control - showed a similar luminosity as
202 commercial sample from semolina [22]. Adding egg albumen did not affect the colour of rice pasta,
203 whereas a significant ($p < 0.05$) decrease in luminosity and an increase in both redness and
204 yellowness was highlighted by the addition of whey proteins, as a likely consequence of non
205 enzymatic browning phenomena [22]. Indeed, furosine levels were almost twice as high in PaWP
206 with respect to PaEA (Table 1).

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208 Cooking behaviour

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210 The cooking behaviour of pasta samples is shown in Table 2. In GF pasta, because of the lack of
211 gluten, starch polymers are less **efficiently** entrapped in the matrix, giving a product with high
212 cooking loss and low firmness [23]. The use of PR improved the texture of the product, but it
213 seemed quite **efficient** as for limiting the leaching of solid matter into cooking water, that was four
214 times higher than in semolina pasta [23]. Adding soluble proteins to the formulation did not
215 **promote** significant ($p > 0.05$) changes in the amount of absorbed water during cooking. However,
216 the use of proteins decreased the cooking loss, and EA was by far more **efficient** than WP in
217 lowering the cooking loss and in increasing the firmness of pasta (Table 2).

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3 218 Thus, a protein network suitable for retaining starch and the other constituents was formed
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5 219 as a consequence of protein coagulation upon cooking the protein-enriched samples [24]. The high
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7 220 solubility and hydration properties of the proteins used in this study also favoured a homogeneous
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9 221 distribution of albumen and whey proteins inside the matrix during the mixing phase. The proteins
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11 222 used here were hypothesized to contribute emulsifying effects as well [25]. The positive effect of
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13 223 egg addition was observed also in pasta from pseudocereals [17], but only a few studies have been
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15 224 carried out on the use of whey proteins [24].
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21 226 Surface textural characteristics

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25 228 The surface texture of pasta samples before and after cooking, as described by the heterogeneity
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27 229 (HTG) parameter, is shown in Fig. 1. Roughness is relevant here in what it affects the ability to
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29 230 retain condiments. The addition of proteins significantly ($p < 0.05$) decreased roughness of the
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31 231 uncooked product, increasing the surface homogeneity. In particular, whey protein-enriched pasta
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33 232 showed the highest homogeneity (low HTG). In all samples, a decrease in HTG was detected after
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35 233 cooking. A similar phenomenon was observed in durum wheat pasta enriched with buckwheat flour
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37 234 [20].
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40 235 Although cooking lowered the differences in surface heterogeneity among the samples, PaPR
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42 236 still showed the highest HTG, accounting for the high cooking loss (Table 2). Samples with the
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44 237 highest HTG (and therefore with the roughest surface) expose a greater area to water action during
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46 238 cooking and, consequently, a high amount of material can be released into the cooking water [20].
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48 239 This combination of evidences suggest that the high homogeneity in protein-enriched rice pasta
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50 240 may be due to starch-protein interactions that occurred during pasta-making.
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55 242 Ultrastructure of cooked pasta

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SEM images of cross-sections of cooked pasta are shown in Fig. 2. For each sample an image at low magnification is proposed in order to appreciate at the same time the organization of both the external and the central region. Apparently, PaPR sample showed a more compact internal area (Fig. 2a). The addition of soluble proteins was associated with a wider porous structure in the external area. In other words, in PaEA (Fig. 2b) and PaWP (Fig. 2c) macaroni the absorption of boiling water during cooking promoted the formation of a highly hydrated region as shown by the extension and the size of the honeycomb cells that form during the freezing step of highly hydrated gels as required by SEM sample preparation [26].

Interesting structures can be distinguished when looking at the core of cooked macaroni at high magnification. PaPR presents an undifferentiated structure (Fig. 2d), in which starch material is organized in a very thin honeycomb network made of numerous small cells. The resulting high surface exposure could contribute to the high cooking loss in PaPR (Table 2). PaEA has a very different structure, as starch material is easily identified as separated by a thin protein layer, and no porous starch organization was distinguishable (Fig. 2d). In PaEA, protein and starch granules appear densely packed, accounting for the high firmness of this sample (Table 2). In PaWP, large agglomerates of gelatinized starch material were separated by long protein fibrils (Fig. 2e). Some discontinuities are present among protein and starch, providing a rationale for the higher cooking loss observed in PaWP with respect to PaEA (Table 2).

Starch properties

Changes in viscosity of pasta samples (ground into flours of uniform size before cooking) was evaluated by the microviscoamylograph test (MVAG), and the results are shown in Fig. 3. Although this approach is conventionally adopted for evaluating the pasting properties of starch and flours, it was also performed on dry pasta, giving information on potential starch behavior during cooking [4]. The MVAG curves of PR and PaPR confirmed previous studies [23]. The low viscosity of PR

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3 270 could be related to the presence of a matrix with a low hydration capacity, as a consequence of both
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5 271 starch gelatinisation and retrogradation phenomena occurring during the parboiling process [27].
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7 272 PaPR showed a lower pasting temperature and a higher hot viscosity compared to PR (Table 3).
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9 273 These results suggested that pasta-making process promoted structural changes, resulting in a
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11 274 product with altered starch properties and accounting for the higher amount of starch quickly
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13 275 accessible to enzymatic hydrolysis, as already discussed by Marti et al. [4]. Starch susceptibility to
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15 276 α -amylase hydrolysis (expressed as damaged starch, see Table 3) as promoted by pasta-processing
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17 277 can complete the information about the starch organisation: the higher the enzymatic susceptibility
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19 278 index, the lower the pasting temperature, and the higher the maximum viscosity [4].
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23 279 PaEA showed a viscosity profile similar to that of PaPR (Fig. 3), and only a slight but not
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25 280 significant ($p>0.05$) increase in the maximum hot viscosity was observed (Table 2). On the
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27 281 contrary, the presence of whey protein significantly ($p<0.05$) decreased the peak viscosity in PaWP,
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29 282 that showed a higher pasting temperature and a lower hot viscosity than PaPR. Results suggest that
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31 283 the whey proteins might have slowed down water uptake by individual starch granule (and,
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33 284 consequently, their gelatinisation) as a consequence of the possible competition of the different
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35 285 biopolymers for available water. A decrease in viscosity was also observed by Marco and Rosell
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37 286 [25] when whey proteins were added to rice flour, likely due to the dilution effect on starch
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39 287 concentration. Indeed, in previous reports, a negative correlation had been established between the
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41 288 protein content and the peak viscosity in rice flour [28,29]. However, the dilution effect could not
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43 289 be **solely** responsible for the decrease in the peak viscosity promoted by whey proteins, because no
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45 290 significant changes were observed when the amount of whey proteins in rice pasta was increased
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47 291 (data not shown). The presence of whey proteins significantly decreased the final viscosity with
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49 292 respect to PaPR, confirming that the increase of viscosity during cooling, usually related to the
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51 293 crystallization of the amylose chains, could be affected also by the reorganization of the denatured
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53 294 proteins [25]. Finally, the presence of large agglomerates of gelatinized starch material in PaWP
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3 295 (see Fig. 2e) could account for the lower susceptibility of PaWP to α -amylase hydrolysis in
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5 296 comparison with PaPR, as also reported in Table 2.
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10 298 Protein network
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14 300 The structural role of covalent and non-covalent interactions among proteins in different pasta
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16 301 samples - before and after cooking - was assessed by detecting the amount of protein solubilized in
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18 302 media with a different ability in dissociating protein-protein complexes (Fig. 4). Albumins and
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20 303 globulins are soluble in plain saline buffer, while proteins involved in aggregates stabilized by non-
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22 304 covalent and/or covalent interactions (that is, interprotein disulfide bonds) are soluble in buffer
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24 305 containing urea or urea/DTT, respectively [30,31]. In the case of uncooked pasta, solubility of PaPR
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26 306 proteins was negligible in saline buffer, and remained very low in the presence of urea unless DTT
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28 307 was added (Fig. 4a). These results indicate that rice proteins were involved in a network stabilized
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30 308 by disulphide bonds.
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34 309 The amount of proteins soluble in saline buffer also was very low in PaEA and PaWP, but it
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36 310 increased significantly after addition of urea, and increased much further when both urea and DTT
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38 311 were present in the extraction buffer. In the presence of urea, about 20 mg proteins per gram of
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40 312 pasta sample can be extracted, suggesting that both albumen and whey proteins form a non-
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42 313 covalently bound protein network in these materials. Indeed, the amount of protein solubilized by
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44 314 the chaotrope is comparable to the quantity of structuring proteins (EA or WP) added to these
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46 315 samples.
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50 316 Presence of a disulfide-reducing agent was necessary to allow solubilization of appreciable
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52 317 amount of proteins from cooked pasta samples (Fig. 4b). Thus, cooking resulted in the formation of
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54 318 a protein network in which the contribution of interprotein disulfide bonds was much greater than in
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56 319 the uncooked product.
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3 320 Information about the compactness of the protein network can be obtained by comparing the
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5 321 amount of protein extracted from each pasta sample before and after cooking. Whereas figures for
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7 322 PaPR and PaWP did not change after cooking, solubility of PaEA protein decreased markedly after
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9 323 cooking. This suggests that aggregates formed by egg albumen proteins in the cooked pasta are so
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11 324 compact to be inaccessible to the disulfide-reducing agents. This compactness provides a rationale
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13 325 for minimizing solid loss and for maintaining good firmness in cooked PaEA (Table 2).
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17 18 327 **Conclusions**

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23 329 The addition of egg albumen and whey proteins as texturing ingredients is an interesting approach
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25 330 for producing GF pasta with improved cooking quality without using chemical additives. On a
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27 331 similar protein enrichment level, the best results were obtained by using egg albumen. **Egg albumen**
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29 332 **gave pasta of better appearance, with lower cooking loss, and firmer and nutritionally more valuable**
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31 333 **than the one made by using whey proteins.** In pasta made with a 15% addition of liquid albumen to
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33 334 parboiled rice flour, starch molecules are homogeneously surrounded by a protein network
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35 335 stabilized mostly by hydrophobic interactions and by disulfide bonds. Further disulfide interprotein
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37 336 bonds form upon cooking, resulting in significant improvement of the textural and structural
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39 337 features of the product without substantial interference with the pasting behavior of starch or its
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41 338 accessibility to amylolytic enzymes.
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45 46 47 340 **Acknowledgements**

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Table 1 Characteristics of uncooked pasta.

	PaPR	PaEA	PaWP
Luminosity (L*)	89.8 ^a	90.2 ^a	88.6 ^b
Redness (a*)	0.92 ^a	0.82 ^a	1.41 ^b
Yellowness (b*)	20.4 ^a	20.3 ^a	23.7 ^b
Furosine (mg/ 100 g protein)	148 ^a	284 ^b	458 ^c

Means with different superscripts in each line are significantly different (LSD; $p < 0.05$)

PaPR, rice pasta; PaEA, rice pasta with egg albumen; PaWP, rice pasta with whey proteins

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3 459**Table 2** Characteristics of cooked pasta.4
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Means with a different superscript in each line are significantly different (LSD; $p < 0.05$)

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PaPR, rice pasta; PaEA, rice pasta with egg albumen; PaWP, rice pasta with whey proteins

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Table 3 Effect of texturing ingredients on starch properties of rice pasta.

	PR	PaPR	PaEA	PaWP
Starch susceptibility (damaged starch; % db)	8.42 ^a	14.48 ^c	13.92 ^c	10.35 ^b
PT (°C)	76.4 ^c	56.3 ^a	56.8 ^a	60.9 ^b
HV (BU)	114.0 ^a	174.0 ^b	199.0 ^b	130.5 ^a
FV (BU)	272.5 ^a	572.0 ^c	638.5 ^c	375.0 ^b
SB (BU)	158.5 ^a	398.0 ^c	448.0 ^c	245.0 ^b

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478 Means with a different superscript in each line are significantly different (LSD; p<0.05)

479 PT = pasting temperature; HV = hot viscosity; FV = final viscosity; SB = setback.

480 PR = parboiled rice; PaPR, rice pasta; PaEA, rice pasta with egg albumen; PaWP, rice pasta with
481 whey proteins

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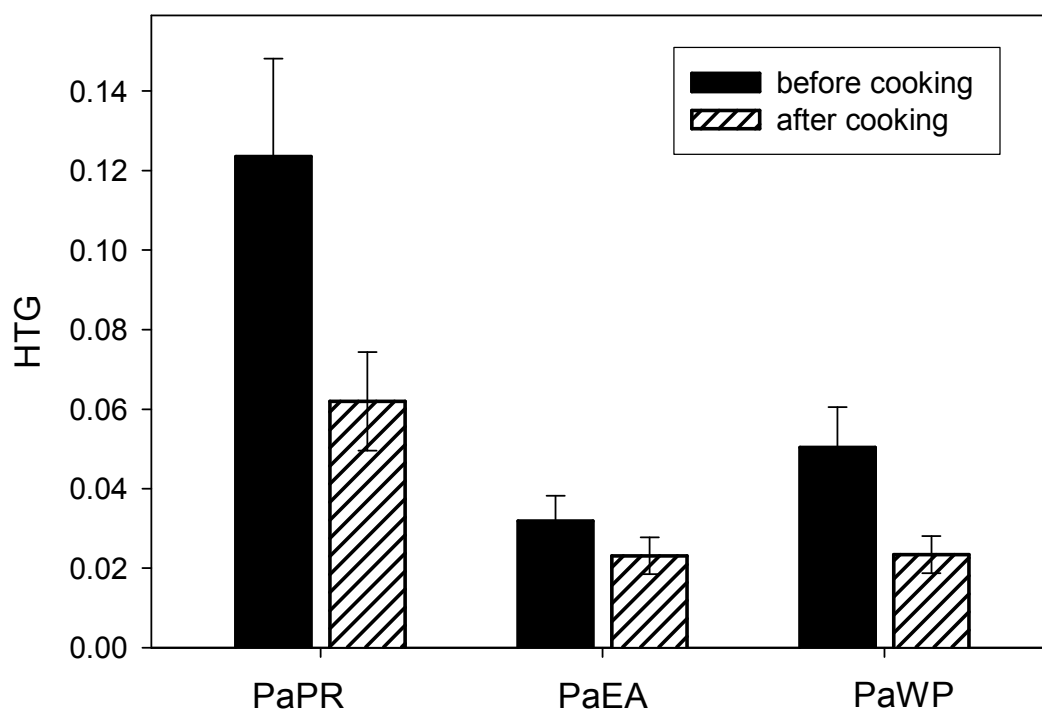
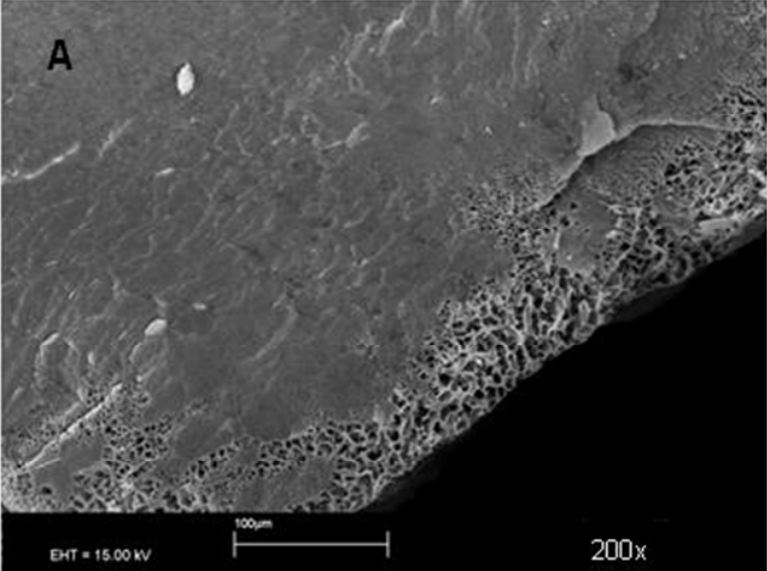


Fig.1 Surface heterogeneity of pasta before and after cooking

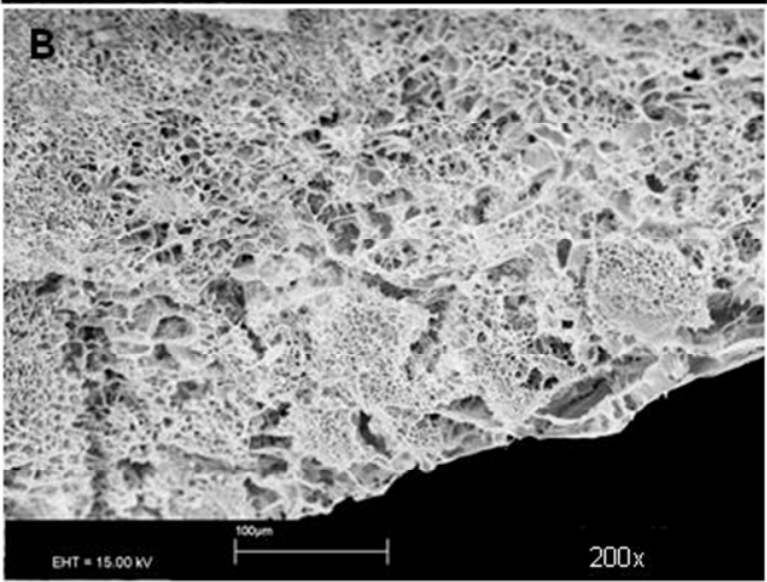
PaPR, rice pasta; PaEA, rice pasta with egg albumen; PaWP, rice pasta with whey proteins

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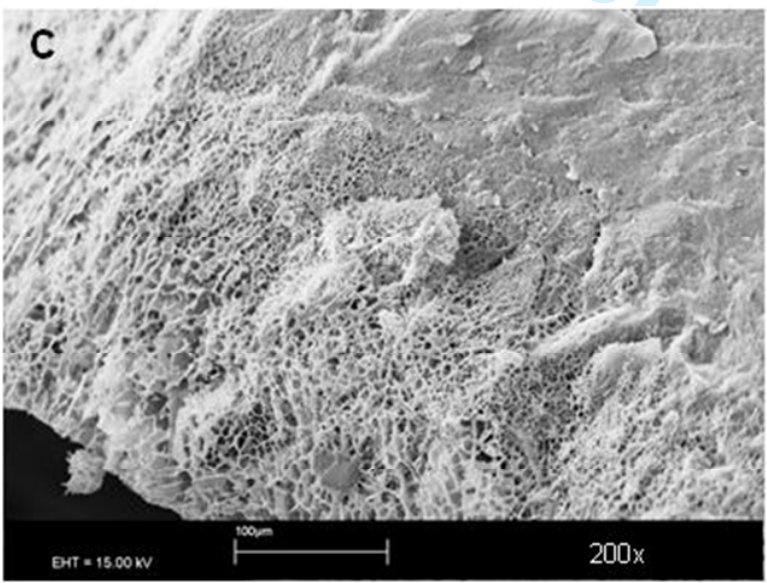
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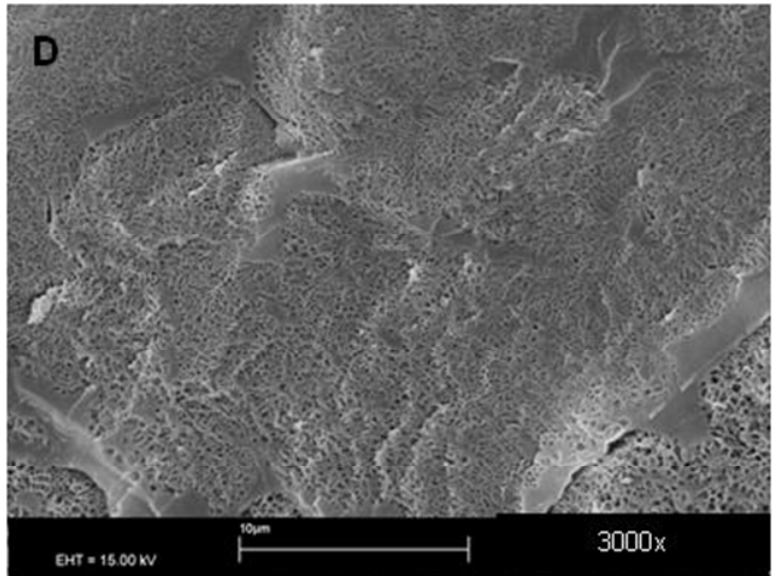


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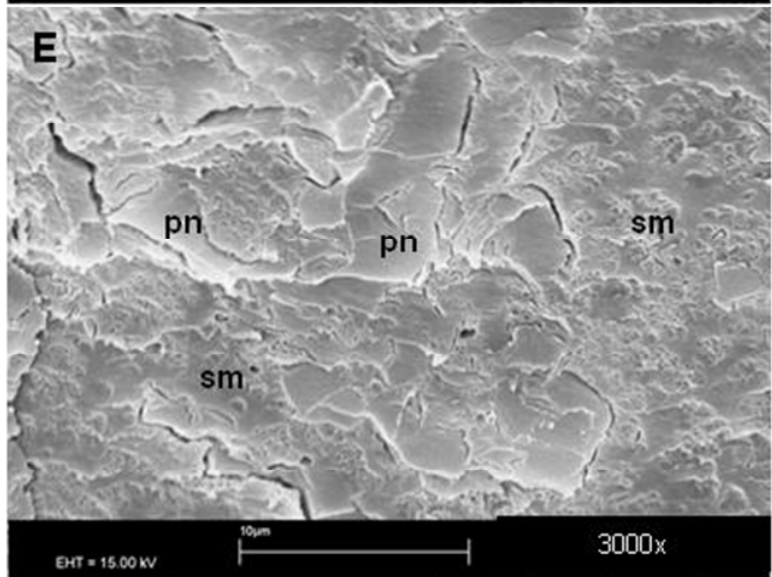


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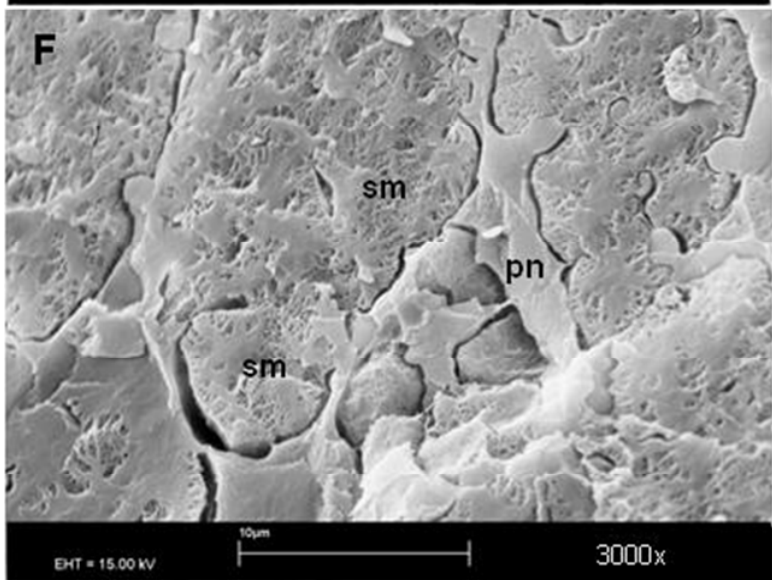
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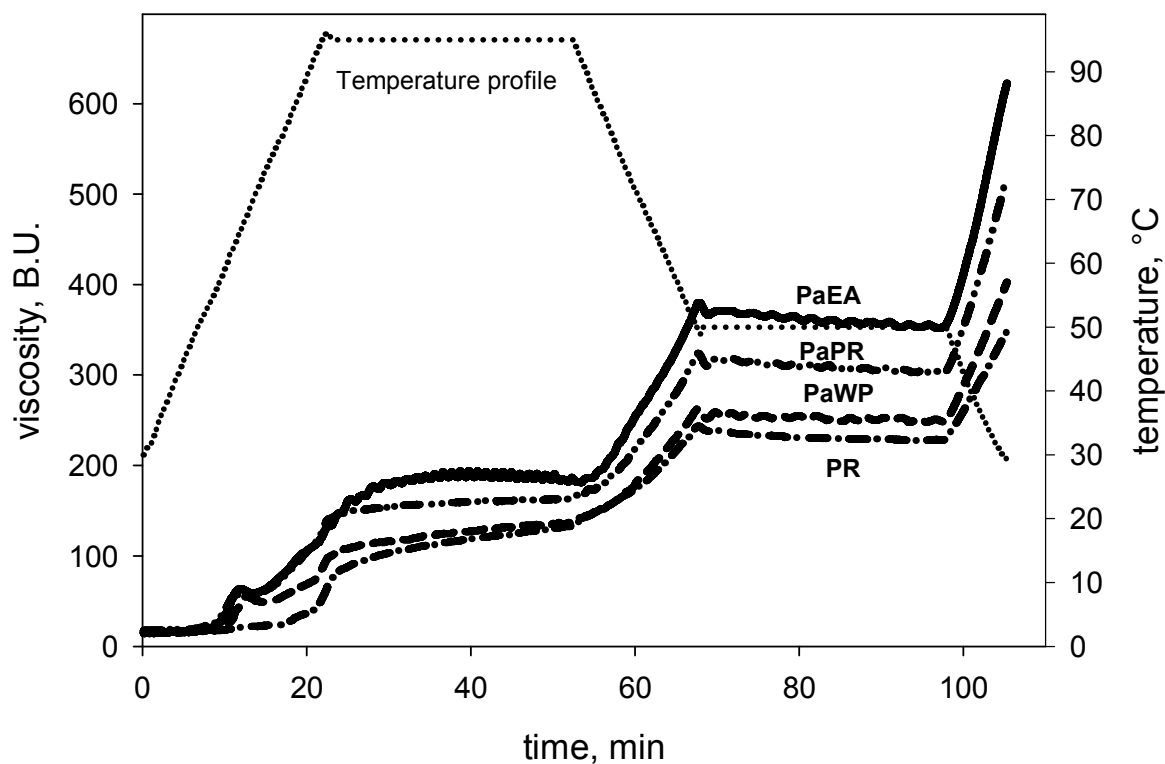
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3 499 **Fig. 2** SEM images of cross-sections of cooked pasta.
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5 500 PaPR at low (A) and high (D) magnification; PaEA at low (B) and high (E) magnification; PaWP at
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7 501 low (C) and high (F) magnification. pn, protein network; sm, starch material.
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9 502 PaPR, rice pasta; PaEA, rice pasta with egg albumen; PaWP, rice pasta with whey proteins
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For Peer Review

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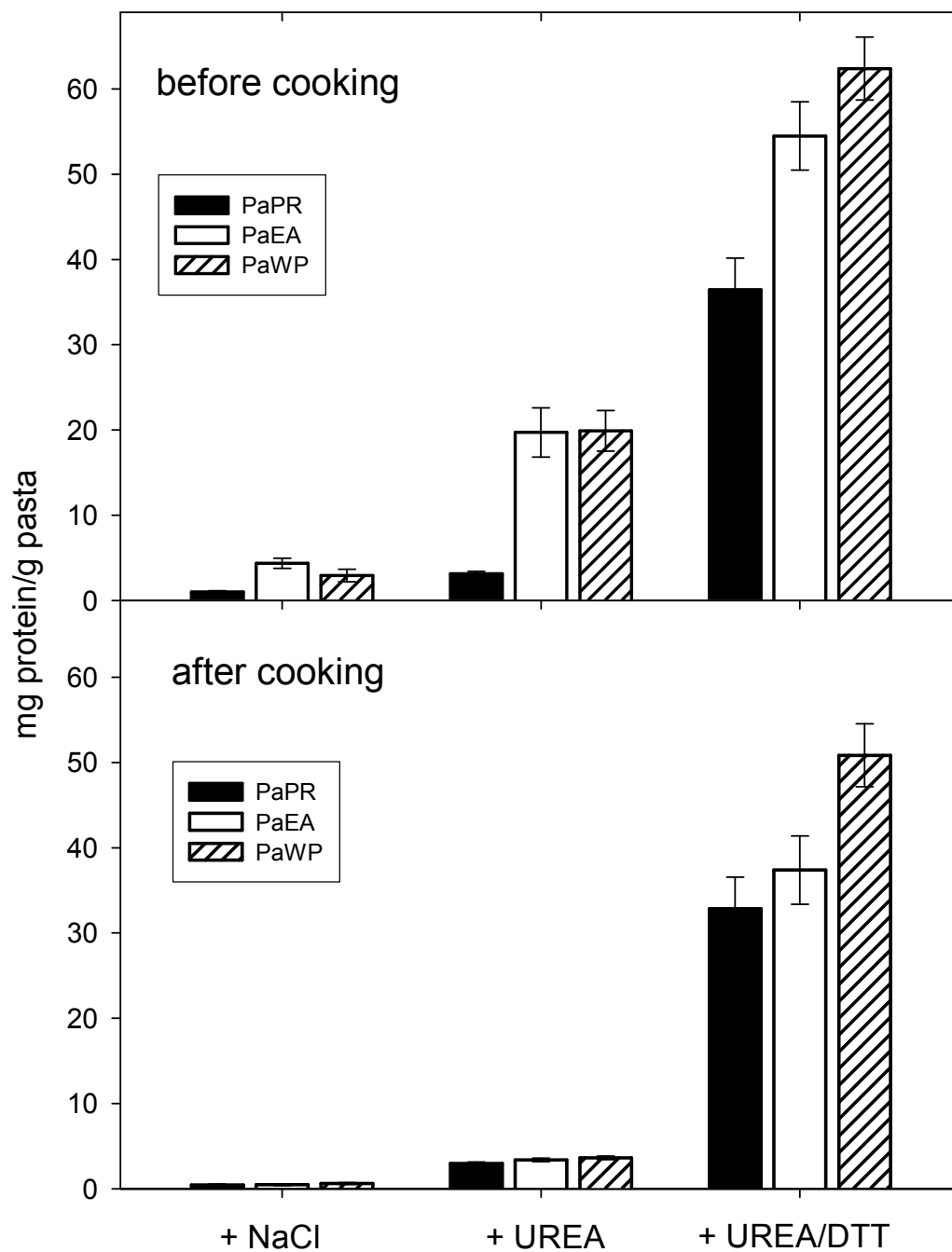
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Fig. 3 Pasting properties of rice pasta.

508 PR = parboiled rice; PaPR, rice pasta; PaEA, rice pasta with egg albumen; PaWP, rice pasta with
509 whey proteins

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Fig 4 Protein solubility in uncooked (upper panel) and cooked (lower panel) rice pasta.

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PaPR, rice pasta; PaEA, rice pasta with egg albumen; PaWP, rice pasta with whey proteins