Studies On Apple Seed Germination And Survival Of Seedlings As Affected By Gibberellic Acid Under Cold Arid Conditions.

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Abstract: Ladakh, the land between earth and sky, is known for its remoteness, difficult terrain and short agriculture season. Nearly 50% of its total land lies between 16500 and 19500 feet above mean sea level. It is virtually unfit for human life and vegetation. The experiment was undertaken with an objective to investigate the effect of various concentrations of plant growth regulator (GA₃)) under cold arid conditions to standardize the procedure for seed germination of Apple (Malus x domestica Borkh.) so as to increase the dearth of apple seedling plants which can easily acclimatize under these tough conditions with better survival for maximum benefit to nursery growers and orchardists. One of the effects of cold arid conditions is the reduction in seed germination, seedling establishment (survival) and seedling growth but research reveals that pretreatment with growth regulators improves some aspects of apple seedlings as well as to standardize the technique under cold arid conditions, the experiment was conducted in Horticulture research field of High Mountain Agriculture Research Institute Leh, Ladakh India. The study clearly indicated that apple seed treated with GA₃ 500 ppm for 40 hours gave best response regarding seed germination, better growth of seedlings and survival of sapples.

Key words: Apple, Gibberellic acid, Growth, survival, seed germination, seedling establishment, water stress, cold arid conditions.

Introduction

Seed germination is a complex physiological process that it is response to environmental signals such as water potential, light, nitrate and other factors .Poor seed germination is the major limiting factor of Apple (Malus x domestica Borkh.) for large scale production and cultivation under cold arid conditions. Seed germination in general can be controlled by many factors like natural germination (growth) inhibitors (Taiz et al., 1991). These are the derivatives of benzoic acid. cinnamic acid. coumarin. naringenin, jasmonic and abscisic acid (ABA). It has been postulated that seed coat (testa) of many plant species contains considerable amount of germination inhibitor, which prevent their seed germination (EL-Barghathi et al., 2005). The first stage of germination consists of ingesting water and an awakening or activation of the germplasm. Protein components of the cells that were formed as the seed developed became inactive as it matured.

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Control is exercised by four classes of plant hormones: inhibitors such as abscisic acid which block germination; auxins which control root formation and growth; the gibberellins which regulate protein synthesis and stem elongation; and cytokinins that control organ differentiation. Gibberellic acid (actually a group of related substances called gibberellins) was discovered as a metabolic byproduct of the fungus Gibberella fujikuroi. Gibberellic acid (GA3) is known to be concerned in the regulation of plant responses to the external environment (Chakrabarti et al., 2003). Recent studies have revealed that cold stratification has a direct effect on production of gibberellins (GAs) in seeds of Arabidopsis thaliana (Yamaguchi et al., 2000, 2002). Exogenously applied GA₃ overcomes seed dormancy in several species (Baskin and Baskin, 1998; Hartmann et al., 1997) and promotes germination in some species that normally require cold stratification, light, or after-ripening (Bewley et al., 1994). GA promotes the production of enzymes such as endo- β -mannanase, which loosen cell walls in the endosperm, thereby reducing resistance to radicle emergence (Yamaguchi et al., 2002). In some temperate species, dormancy is broken by a period of warm temperatures followed by cold stratification. This response is most often associated with morphophysiological dormancy (et al., 2005); however, seeds with morpho-physiological dormancy have underdeveloped embryos (et al., 2005). In order to accelerate this method, it can be combined with some treatments such as chemical applications or mechanical seed coat removal (Mehanna et al., 1985; Martinez-Gomez et al., 2001). Many investigators have studied the effects of exogenous growth regulators on seed germination. Gibberellins eliminated the chilling requirements of peach and apple seeds and increased their germination (Mehanna et al., 1985). Temperate fruit production in India is based mainly on apple. In India apple 5th in production and roughly occupies 4.5% of fruit ranks area in the country. The apple is mostly cultivated in Jammu and Kashmir followed by Himachal Pradesh and Uttaranchal and Jammu Kashmir alone accounts for 60% of its production in India .Since more than 60% area of Jammu and Kashmir state falls in Ladakh region hence this study

was undertaken to extend and popularize its cultivation in this vast area with prime objective to increase the seedling rootstocks of apple so that it can establish easily and survival rate is improved for increasing the apple fruit production in this nontraditional area. Sexually raised plants are long lived, have extensive and powerful root system with wide and deep distribution in the soil which accounts for persistent and adequate annual growth of absorbing roots thus helping them acclimatize easily in the environment. Also rootstocks exhibit a great effect on production efficiency yield quality, adaptability, tree vigor, impart resistance to biotic (diseases and insects/pests) and aboitic stresses (salt tolerance and water logging) of scion cultivar with the main function of seedling rootstocks is to provide anchorage by growing deep in to the soil and also regulate uptake of moisture and nutrients. Hence this study was under taken to standardize apple seed germination with regard to different concentrations of gibberellic acid so this large number of apple fruit trees may be established in this nontraditional area of Ladakh India for commercial orcharding and get huge returns of apple fruit production in this remote area of India under cold arid conditions for the benefit of country in general and farmers in particular. Chemically induced germination of unstratified apple (Malus x domestica Borkh.) has two possible uses. First, by eliminating stratification one might obtain seedlings immediately after seed maturation. Second, wide familial variation in germination time might be reduced by chemically treating either partially stratified or unstratified seed. Many investigators have studied the effects of exogenous growth regulators on seed germination. The application of gibberellins increases the seed germination percentage by increase of the amino acid content in embryo and they cause release of hydrolytic enzyme required for digestion of endospermic starch when seeds renew growth at germination. Gibberellic acid acts synergistically with auxins, cytokinins and probably with the other plant hormones; what might be called a system approach, or synergism. The overall development of plant is regulated by the growth hormones, nutrient and environmental factors. They also vary in their germination requirement (Chauhan et al., 2009). It is not known in which concentrations these hormones will cause a response in the cell. This investigation with growth hormones will help in determining which of the hormonal concentration is suitable for seed germination and proper seedling growth. Hence the main objective of this study was carried out to find out the effect of GA₃ on germination of apple seed, their subsequent growth and survival of seedlings. In view of the above back ground, the present investigation was undertaken to study the influence of growth substances like Gibberellic acid (GA3) on seed germination, radical and plumule elongation; to draw the information of timing and control of seed germination and seedling growth of a species in their natural habitat.

Materials and Methods

The experiment was conducted in the Horticulture laboratory of High Mountain Agriculture Research Institute Leh Ladakh in 2009-10 by following completely randomized design with three replications. Three concentrations of GA_3 (450, 500 and 550 ppm) were taken into consideration by long dip method for 30 hrs, 35 hrs and 40 hrs. In this

experiment, seeds of all treatments were placed on filter paper moistened with 3% fungicide solution (Captan) in Petri dishes were placed in an incubator at 21±1 ℃ and 70-80% humidity with darkness. Germination was measured in 3-day intervals during 30 days. All seeds with at least a 5 mm long radicle were considered as germinated. Germination percentage per treatment was calculated as the average of three replicates with 50 seeds. Percentage data of germination were subjected to the angle transformation and the analysis of variance was performed. After long dipping, seeds were sown in the pots in the month of March. Observations on germination of seeds, length of seedlings, diameter of seedling, number of leaves per seedling, length and width of leaves, length and diameter of root and survival of seedling were recorded and compared with control statistically. These were soaked for 30. 35 and 40 h in the above concentrations as per treatment combination and only double distilled water for the control set. Seed germination percentage was determined by the method given by Dahiya et al., (2007), after 14 days of treatment. Mean values were subjected to analysis of variance (ANOVA) to test the significance for germination percent as per the methodology advocated by Panse et al., (1967).

Table 1. The treatments of GA3, with their different
concentrations

Growth hormone	Concentration (ppm)	Seed soaked time (h)	Treatments
GA₃	-	-	T ₀ (Control)
		30	T ₁
	450 ppm	35	T ₂
		40	T ₃
		30	T ₄
	500 ppm	35	T_5
		40	T ₆
		30	T ₇
	550 ppm	35	T ₈
		40	T ₉

The concentrations of the test solutions used for the treatment were 450 ppm, 500 ppm, 550 ppm and 40 ppm for Gibberellic acid with a separate control check using the distilled water treatment (Table 1).

Results and discussion

The analysis of variance (ANOVA) of germination data from growth regulators treated seeds indicated that they were all statistically significant (P<0.05). The significant critical difference (CD) values indicate that apple seeds *were* suitable for the treatments (see table 2). The results obtained showed that application of at all concentrations influenced germination of seeds as well as growth and development of seedlings. Maximum number of germinated seeds (9.66) was found with 500 ppm GA₃ for 40 hrs of seed treatment (table 2 and graph 1). Mehanna *et al.*, (1985) reported that internal GA percentage is at a high level, but the proportion of ABA is at a low level in dormant seeds. While GA in the structure increases the enzymatic

activity, it slows the ABA activity. Gibberellins showed to increase germination in several species (Carrera et al., 1988; Giba et al., 1993; Karam et al., 2001) and to overcome physiological dormancy in seeds with dormant embryos (Hartmann et al., 1997). Seed dormancy may be caused by an inadequate development of embryo and/or an existence of chemical inhibitors (Karam et al., 2001). In addition, physiological dormancy in seeds is closely related to the proportion between inhibitors (especially ABA) and growth regulators (especially gibberellins) (Hartmann et al. 1997). This was found in consistent with the finding of Gercekcioğlu et al., (1999) and Carrera et al., (1988) and Chakrabarti et al., (2003). The application of plant growth regulator could increase the seed germination and other physiological activity by the reason of tolerance to the toxic effects/particles which was found in consonance with the finding of Haroun et al., (1991) and Hogue et al., (2002). Although varied in seed germination and root-shoot elongation by different treatments, the pre-soaking with different treatments was evident that soaked seed could improve in germination and seedling establishment and this observation was found equivalent the observation of Harris et al.. (1999). Same results were obtained by Mohanty et al., (2006) while working on Black gram. Treatment of seed with GA₃ 500 ppm for 40 hrs before sowing also increased the length of seedlings (8.94 cm) see table 2 and graph 2, because additional GA₃ activated α - amylase which digested the available carbohydrate in to simpler sugars, so that energy and nutrition were easily available to faster growing seedlings. Increase in plant growth due to gibberellic acid treatment has also been found by Parera et al., (1994) and Lee et al., (1998). GA₃ could overcome the adverse effects in the seed physiological activity of apple seedling growth, which supports the finding of Chakrabarti et al., (2002), Mikulik et al., (2002) and Roychowdhury et al., (2011). The number, length and width of the leaves also increased in treatment T_6 (500 ppm GA₃ for 40 hrs) as shown in table 2. The diameter of main stem increased from 0.27 (control) to 0.44 cm by the treatment of seed with GA₃ 500 ppm for 40 hrs. With the more effectiveness of low concentration of GA3 (that is ratio of growth hormone and water) could restore retardation in water content; this may be able in tolerance to water stress. This result was considered in parallel to the findings of Shnmungavelu et al., (1970), Powell et al., (1987), Lee et al., (1998), Parera et al., (1994), Hoque et al., (2002) and Ashraf et al., (2005). The length of root also increased from 9.03 cm in control to 15.03cm in the treatment with GA₃ 500 ppm for 40 hrs (see table 2, graph 2). Here again linear increase in length of root was noted for every increase in concentration of GA₃ and duration of treatment of seed before sowing. It is but natural that when the treatment has boosted the growth of seedling above ground, it might have given the same effect below ground *i.e.* in the roots. The diameter of root at the base of its origin was 0.14 cm in control and 0.25 cm in the treatment T_6 (500 ppm GA₃ for 40 hrs). Here again the lower concentration and reduced duration of treatment have linearly decreased the thickness of root which is similar with findings of Copeland et al., (2001), Khan et al., (1992) and Ruan et al., (2002) and Pasalar et al., (2011). It is more important to note that survival of seedling is more important thing. Weaker seedlings are more susceptible to disease and adverse weather conditions than stronger seedlings.

Survival of seedling was 4 out of 10 in control which increased to 9.66 out of 10 in T_6 (500 ppm GA₃ for 40 hrs). Here again the survival of seedlings simultaneously decreased with every decrease in the concentration of GA₃ and duration of treatment of seed before sowing which is in agreement with Singh *et al.*, (1989) and Mueungu *et al.*, (1989). The role of plant growth regulators in overcoming the harmful effects on growth may be due to the change in the endogenous growth regulators (Izumi *et al.*, 1996) and Thomas *et al.*, 2005. It has been confirmed that exogenous application of Gibberellic acid promotes seed germination and growth of the radicle and plumule of many plants (Taiz *et al.*, 2010 and Roychowdhury *et al.*, 2011b, Roychowdhury *et al.*, 2011d).

Conclusion

From the above discussion, it is concluded that different concentrations of Gibberellic acid (GA3) endorsed germination percentage and growth of seedlings in apple seed i.e. radical and plumule elongation. In this study, in order to remove dormancy in apple seeds, exogenous GA₃ application has been most successful in breaking dormancy at 500 ppm concentration(see table 2, graph 1 and 2). This it is recommended from this research that that the concentration of growth regulators GA₃ 500 ppm for 40 hrs favors the increased enzymatic activity that leads to the favorable environment for the seed germination as well as the growth of the radicle and plumule leading to better growth and survival of seedlings. Hence it is inferred that an adaptation and practical application of these findings in nurseries might also have an economic effect.

Literature Cited:

- [1]. A. A. Khan, "Pre- plant physiological seed conditioning," Hort. Rev., 14, 131-181 (1992).
- [2]. A. H. M. A. Mohammed, "Physiological Aspects of Mungbean Plant (Vigna radiata L. wilczek) in Response to Salt Stress and Gibberellic Acid Treatment," Res J Agric Biol. Sci. 3(4): pp. 200-213, 2007.
- [3]. A. Pasalar and B. Rezaee, "Effect of different treatments on dormancy and stimulate germination of henopodium," Fifth national conference on new ideas in agriculture, pp: 4, 2011.
- [4]. C. A. Parera and D. J. Cantliff, "Jannick Journal,"16, pp. 119-141, 1994.
- [5]. C. C. Baskin and J. M. Baskin, "Seeds: Ecology, biogeography, and evolution of dormancy and germination," (Academic Press, Boston, MA) 1998.
- [6]. C. Carrera, M. Reginato and S. E. Alomso, "Seed dormancy and germinations in P. mahaleb L", Seed Abstract: 11–122, 1988.
- [7]. D. Harris, A. Joshi, P. A. Khan, P. Gothkar and P. S. Sodhi, "On-farm seed priming in semi-arid agriculture: development and evaluation in maize, rice and chickpea in India using participatory methods," Exp Agric, 35:pp. 15-29, 1999.

- [8]. F. S. Mueungu, P. Chiduza, L. J. Nyamugafata, W. R. Clark Whalley and W. E. Finch - savage. Field Crops Res, 89, pp. 49 -57, 2004.
- [9]. H. T Hartmann, D. E. Kester, F. Davies Jr. and R.L. Genève, "Plant Propagation Principles and Practices, Sixth Edition," New Jersey, Prentice Hall, 1997.
- [10]. J. D. Bewley and M. Black Seeds, "Physiology of Development and Germination," Plenum Press, New York, USA, (1994).
- [11]. J. Mikulik and V. Vinter, "valuation of factors affecting germination of Dianthus superbus L. sub spp. Superbus," Biologica 40: pp. 13-18, 2002.
- [12]. J. S. Chauhan, Y. K. Tomar., I. N. Singh, S. Ali and A. Debarati, "Effect of growth hormones on seed germination and seedling growth of black gram and horse gram," J American Sci. 5(5):pp. 79-84, 2009.
- [13]. J. Mikulik and V. Vinter, "valuation of factors affecting germination of Dianthus superbus L. sub spp. Superbus," Biologica 40: pp. 13-18, 2002.
- [14]. K. G. Shnmungavelu, "Effect of gibberellic acid on seed germination and development of seedling of some tree species, "Madras Agri- J, 57, pp. 311-314, 1970.
- [15]. L. E. Powell, "Hormonal aspects of bud and seed dormancy in temperate-zone woody plants," Hort. Science, 22: 845–850, 1987.
- [16]. L. O. Copeland and M. B. McDonald, "Principles of Seed Science and Technology 4th edn.," Kluwer Academic Publisher press (2001).
- [17]. L. Taiz and E. Zeiger, Plant Physiology. Sinauer Associates Inc., USA, 2010.
- [18]. M. Ashraf and M.R. Foolad, "Presowing seed treatment –A shotgun approach to improve germination, plant growth, and crop yield under Saline and non saline conditions," Advances in Agronomy, 88, pp. 223-271, (2005).
- [19]. M. F. El-Barghatgi and A. El-Bakkosh, "Effect of some mechanical and chemical pre-treatments on seed germination and seedling growth of Quercus coccifera (Kemes Oaks)," J Jerash Private University, 2005.
- [20]. M. Fenner, and K. Thompson, "The ecology of seeds,"Cambridge University Press, New York, 2005.
- [21]. M. Hoque and S. Haque, "Effects of GA_3 and its mode of application on morphology and yield

parameters of mungbean (Vigna radiate L.)," Pak. J Biol Sci 5: pp. 281-283, 2002.

- [22]. M. Singh, G. N. Sigh, L. N. Singh and B. N. Singh, "Effect of GA3 on seed germination in Mosambi (Citrus sinensis Osbeck)," Haryana J. Horti. Sci., 18, pp. 29-33, 1989.
- [23]. N. Chakrabarti and S. Mukherji, "Effect of phytohormone pretreatment on metabolic changes in Vigna radiate under salt stress," J Environ Biol., 23: pp. 295-300, 2002.
- [24]. N. Chakrabarti and S. Mukherji, "Effect of Phytohormone pretreatment on nitrogen metabolism in Vigna radiate under salt stress," Biol Planta, pp. 46: 63-66, 2003.
- [25]. N. S. Karam and M. M Al-Salem, "Breaking dormancy in Arbutus andrachne L. seeds by stratification and gibberellic acid," Seed Science & Technology, 29: pp. 51–56, 2001.
- [26]. O. S. Dahiya, and P. Kumari, "Seed germination: optimum temperature, moisture and light," In: Plant Physiology research methods (S S Narwal, eds), Scientifi c Publishers, Jodhpur, India, 2007.
- [27]. P. Halmer, "In Seed technology and its biological basis, Commercial seed treatment technology," eds Black M., Bewley J.D. (CRC Press LLC, Boca Raton, FL), pp. 257–28, 2000.
- [28]. P. Martinez-Gomez and F. Dicenta, "Mechanisms of dormancy in seeds of peach (Prunus persica (L.) Batsch) cv. GF 305," Scientia Horticulturae, 91: pp. 51–58, 2001.
- [29]. R. Gercekcioglu and C. Cekic, "Mahlep (Prunus mahaleb L.) tohumlarının cimlenmesi uzerine bazıuygulamaların etkileri," Turk Journal of Agriculture and Forestry, 23: 145–150, 1999.
- [30]. R. Roychowdhury, M. J. F. Alam, S. Bishnu, T. Dalal and J. Tah, "Comparative study for chemical mutagenesis on seed germination, survivability and pollen sterility in M1 and M2 generations of Dianthus," Plant Breeding and Seed Science, 65(1): pp. 29-38, 2012.
- [31]. R. Roychowdhury, J. Tah, "Genetic variability study for yield and associated quantitative characters in mutant genotypes of Dianthus caryophyllus L.," International Journal of Biosciences, 1(5): pp. 38-44, 2011b.
- [32]. R. Roychowdhury, J. Tah, "Mutation breeding in Dianthus caryophyllus L. for economic traits," Electronic Journal of Plant Breeding, 2(2): pp. 282-286, 2011.
- [33]. R. Roychowdhury, J. Tah, T. Dalal and A. Bandyopadhyay, "Selection response and

correlation studies for metrical traits in mutant carnation (Dianthus caryophyllus L.) genotypes," Continental Journal of Agricultural Science, 5: pp. 06-14, 2011d.

- [34]. S. A. Haroun, A. H. .Badawy, W. M. Shukry Auxin induced modifi cation of Zea mays and Lupinus termis seedlings exposed to water stress imposed by polyethylene glycol (PEG 6000). Sci J, 18:pp. 335, 1991.
- [35]. S. G. Thomas, I. Rieu and C. M. Steber, "Gibberellin metabolism and signaling," Vitam Horm, 72, pp. 289-338, 2005.
- [36]. S. K. Mohanty, N. C. Sahoo, "Effect of soaking period, seed size and growth regulators on imbibitions and germination of seeds of some field crops," Orissa J. Agric. Res., 2000.
- [37]. S. Ruan, Q. Xue and K. Tylkowska, "Seed Science technology," 30, pp. 451-458, 2002.
- [38]. S. S. Lee, J. H. Kim, S. B. Honge, S. H. Yuu and E. H. Park, "Korean Journal of crop Science," 43, pp. 194-198, 1998.

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- [39]. S. Yamaguchi, and Y. Kamiya, "Gibberellin biosynthesis: Its regulation by endogenous and environmental signals," Plant Cell Physiol. 41: pp. 251–257, 2000.
- [40]. S. Yamaguchi, and Y. Kamiya, "Gibberellins and light-stimulated germination," J. Plant Growth Regul. 20: pp. 369–376, 2002.
- [41]. T. H. Mehanna, C. M. George and C. Nishijima, "Effects of temperature, chemical treatments and endogenous hormone content on peach seed germination and subsequent seedling growth," Scientia Horticulturae, 27: pp. 63–73, 1985.
- [42]. V. G. Panse and P.V. Sukhatme, "Statistical Methods for Agricultural Workers," 2nd Ed., ICAR publication, New Delhi, India, 1967.
- [43]. Z. Giba, D. Grubisic and R. Konjevic, "The effect of white light, growth regulators and temperature on the germination of blueberry (Vaccinium myrtillus L.) Seeds," Seed Science & Technology, 21: 521– 529, 1993.

Treatment/ Character		Germina tion Percent (%)	Germin ation of seed (N)	Growth of seedlin gs (cm)	No. of leave s/see dling	Len gth of leaf (cm)	Width of leaf (cm)	Diam eter of stem (cm)	Lengt h of root (cm)	Diame ter of root (cm)	survival of seedlings (N)	
	Hour	S										
Control	0	T ₀	54.27	5.33	4.67	8.80	1.09	1.16	0.27	9.03	0.14	4.00
GA₃ 450 ppm	30	T ₁	66.64	6.33	4.95	9.00	1.69	1.58	0.31	10.03	0.15	5.00
	35	T_2	75.75	6.00	4.77	9.00	2.53	1.43	0.31	10.35	0.17	5.00
	40	T_3	62.75	6.66	5.33	8.33	1.39	1.46	0.32	10.62	0.19	6.00
GA₃ 500 ppm	30	T_4	72.08	7.00	4.97	11.00	2.90	1.73	0.34	13.05	0.22	6.00
	35	T_5	87.46	7.00	4.49	11.00	2.78	1.79	0.39	13.35	0.23	7.00
	40	T_6	63.34	9.66	8.94	16.00	3.44	2.25	0.44	15.03	0.25	9.66
GA₃ 550 ppm	30	T ₇	69.54	7.00	5.37	9.66	1.71	1.51	0.34	11.03	0.17	7.00
	35	T ₈	78.92	7.00	4.24	9.00	2.16	1.54	0.33	11.35	0.17	5.00
	40	T ₉	61.28	6.00	5.61	10.00	2.36	1.45	0.34	11.55	0.15	7.00
CD>0.05		0.648	1.450	0.038	1.280	0.03 4	0.120	0.016	0.014	0.017	1.640	

Table 2: Gibberellic acid effect on germination, growth and survival of Apple seeds / seedlings.



Figure 1: Schematic representation showing effect of GA₃ on germination of Apple seeds.







