

STUDY OF THE SOIL–PLANT (CARROT)–ANIMAL CYCLE OF NUTRITIVE AND HAZARDOUS MINERALS IN A RABBIT MODEL

A. BERSÉNYI¹, S. FEKETE^{1*}, I. HULLÁR¹, I. KÁDÁR², M. SZILÁGYI³, R. GLÁVITS⁴,
Margit KULCSÁR¹, M. MÉZES⁵ and L. ZÖLDÁG¹

¹University of Veterinary Science, H-1400 Budapest, P.O. Box 2, Hungary; ²Soil and Agrochemical Research Institute of the Hungarian Academy of Sciences, Budapest; ³Research Institute for Animal Breeding and Nutrition, Herceghalom; ⁴Central Veterinary Institute, Budapest; ⁵University of Agricultural Sciences, Gödöllő, Hungary

(Received September 15, 1998; accepted February 19, 1999)

Carrots were grown on soils polluted by heavy metal salts. Each particular microelement reached a high concentration [molybdenum (Mo) 39.00, cadmium (Cd) 2.30, lead (Pb) 4.01, mercury (Hg) 30.00, and selenium (Se) 36.20 mg/kg dry matter] in the carrot. In a metabolic balance trial conducted with 15 male and 15 female New Zealand White rabbits, the control animals (n = 5) were fed *ad libitum* with concentrate as basal diet, while the other rabbits received the basal diet and carrots containing the particular microelement. Blood samples were taken to determine the activity of serum enzymes. To investigate the metabolism of Mo, Cd, Pb, Hg and Se, samples were taken from the heart, liver, lungs, kidneys, spleen, ovaries/testicles, entire digestive tract, adipose tissue, femur, hair, faeces and urine. Carrot had significantly higher digestibility for all nutrients than the rabbit concentrate. Carrot samples of high Pb content had the lowest digestibility of crude protein. The microelements differed in their rate of accumulation in the organs examined: Mo and Cd accumulated in the kidneys, Pb in the kidneys, liver, bones and lungs, Hg in the kidneys and liver, while Se in the liver, kidneys and heart. The proportions of microelements eliminated from the body either via the faeces and urine (Mo 80.18% and Se 47.41%) or via the faeces (Cd 37.86%, Pb 66.39%, Hg 64.65%) were determined. Pathohistological examination revealed that the rate of spermatogenesis was reduced in the Mo, Cd, Pb and Hg groups compared to the control. Lead, Cd and Hg intake resulted in a considerable decrease in gamma-glutamyltransferase (GGT) and in an increase of alkaline phosphatase (ALP) activity because of damages to the kidneys and bones. All experimental treatments decreased the activity of cholinesterase (CHE) because of lesions in the liver.

Key words: Heavy metals, soil-plant-animal chain, rabbit, carrot, serum enzymes

*Corresponding author; E-mail: Sandor.Fekete@iif.hu; Fax: (36 1) 342 9734

In addition to lead (Pb), cadmium (Cd) has been the subject of numerous studies focusing especially on heavy metal pollution. One of the main reasons why Cd is a hazardous chemical is that it can enter the food chain (Schenkel, 1988): its biological half-life is rather long (17 years for humans). Epidemiological studies have revealed that Cd exposure is correlated with the occurrence of human essential hypertension and Cd concentration of the adrenal gland. Cadmium taken up into the body binds to metallothionein in the liver and kidneys. It can severely damage the kidneys, testicles and ovaries (National Research Council, 1980). Because of the varying metallothionein concentrations and dissimilar metabolic pathways in individual species, values cannot be readily extrapolated from one animal species to another. Data for rabbits are especially scanty. Stowe et al. (1972) carried out a 200-day trial in 3-month-old rabbits, and found that treatment with 160 ppm Cd (administered in the form of CdSO₄) resulted in splenic atrophy, changes in blood parameters, and liver and kidney injury. Fairchild et al. (1977) reported that oral administration of a single dose (43 mg/kg) of cadmium (in the form of CdSO₄) was lethal to rabbits. No data were available in the literature on the effect that Cd naturally taken up from the soil by plants exerted on rabbits.

Materials and methods

Carrot samples (Table 1A) were obtained from the exposure tolerance experiments conducted by the Soil and Agrochemical Research Institute of the Hungarian Academy of Sciences (Budapest). During the experimental period, extremely high doses (810 kg/ha) of Mo [in the form of (NH₄)₆Mo₇O₂₄ × 4 H₂O], Cd [in the form of CdSO₄ × 8 H₂O], Hg [in the form of HgCl₂], Pb [in the form of Pb(NO₃)₂] and Se [in the form of Na₂SeO₃] fertilizers were applied onto the soil and the rate of accumulation of the above heavy metals in carrots was determined. For more technical details see Biacs et al. (1995). Carrot samples with high microelement concentrations (Mo 39.00, Cd 2.30, Pb 4.01, Hg 30.00, and Se 36.20 mg/kg dry matter) were selected.

The animal experiment was carried out in the animal house of the Department of Animal Breeding and Nutrition of the University of Veterinary Science. A total of 15 male and 15 female New Zealand White rabbits (average body weight: 2300 ± 87 g), placed in individual hutches for the metabolic experiment, were used to determine the apparent total tract digestibilities of a breeding rabbit concentrate used as a basal diet (Table 1B). Subsequently the 30 rabbits were divided into six groups (one control and five experimental groups), 2–3 males and females in each. The control group continued to receive the basal diet *ad libitum*, while all five animals in each experimental groups were pre-fed over a period of 10 days. During that period the daily intake of basal diet was re-

stricted to 50 g and a carrot diet containing Mo, Cd, Pb, Hg and Se was offered *ad libitum*. This was followed by a metabolic experiment when a 'feed mixture' containing basal diet and carrot was fed, and the digestibility of nutrients for each of the carrot samples was calculated by the differential experimental formula (Fekete and Gippert, 1982). The mixing ratio, as the actual nutrient ratio in dry matter taken up from carrot, was calculated on the basis of basal diet analysis and the amount of carrot samples fed and remaining. During both experimental periods the total daily urine volume was collected from each animal. Following the metabolic experiment, after the 20-day feeding of carrot samples, blood samples were taken from each animal, and the following blood parameters were determined at the Research Institute for Animal Breeding and Nutrition (Herceghalom) and at the University of Agricultural Sciences (Gödöllő): aspartate aminotransferase (AST), alanine aminotransferase (ALT), gamma-glutamyltransferase (GGT), alkaline phosphatase (ALP), cholinesterase (CHE), creatine kinase (CK), cholesterol (CHOL), triglyceride (TRIG), creatinine (CREA), malondialdehyde, glutathione peroxidase (GSH-Px) and reduced glutathione. The rabbits were weighed, killed painlessly by an overdose of anaesthetic, and subjected to pathological examination. The heart, liver, lungs, kidneys, spleen, ovaries/testicles and entire digestive tract were weighed. Appropriate samples taken from the above organs as well as from the thyroid gland, femoral muscle, jejunum, ileal ampulla and sternum were fixed in buffered formaldehyde solution. In addition to the above-mentioned samples, adipose tissue, femur, hair, faeces and urine samples were taken for chemical analysis including determination of the Mo, Cd, Pb, Hg and Se content of each sample. Samples from animal tissues (kidney, liver, muscle, brain, testicle, ovary, blood, lung and heart) were prepared by lyophilisation, bone, hair and faeces were dried in an oven, and urine was used in natural form. Approximately 1 g samples of animal tissues, hair or faeces, dried at 105 °C, were digested in teflon bombs at 80 °C for 4 h with a mixture of 10 ml of 65 m/m % HNO₃ and 2 ml 30 m/m % H₂O₂. The digest was diluted to 50 ml with deionised water. Having extremely low amounts of animal tissues (e.g. brain, testicle, ovary) the weighed materials (acid, peroxide and deionised water) were reduced proportionally. Approximately 3.0 g of bone was digested 3 times with HNO₃ + H₂O₂ as mentioned above. The final volume of the digest was 150 ml. From the urine samples, 150–200 ml was concentrated to 20 ml, digested with 10 ml HNO₃ + 2 ml H₂O₂ and diluted to 50 ml. Before analysis the samples were diluted 4-fold because of the high concentration of NH₄NO₃ which would have destroyed the plasma without dilution. The analysis of minerals in the HNO₃ + H₂O₂ digest was performed by using a plasma emission spectrometer (ICP), type JY 24 (JOBEN YVON), in a sequential mode at the following wavelengths: Mo 202.03, Cd 228.802, Pb 220.353, Hg 194.227, and Se 196.03 nm.

Table 1A

Nutrient content of the basal diet and of carrots containing microelements, on air-dry basis (%)

Nutrients	Basal diet	Carrot Mo	Carrot Cd	Carrot Pb	Carrot Hg	Carrot Se
DM	91.88	18.88	20.61	21.26	19.47	22.49
Air DM	91.88	90.12	91.63	90.76	89.09	91.52
Ash	7.03	4.93	5.46	5.07	5.11	5.39
OM	84.85	85.19	86.17	85.69	83.98	86.13
CP	15.20	10.60	10.30	9.80	11.10	11.10
CF	11.99	4.80	4.83	4.73	5.55	4.60
EE	1.80	2.70	1.78	2.28	2.28	2.08
NFE	55.86	67.09	69.26	68.88	65.05	68.35

DM = dry matter; OM = organic matter; CP = crude protein; CF = crude fibre; EE = ether extract; NFE = nitrogen-free extract

Table 1B

Ingredients of the basal diet

Ingredients	%
Barley	38.6
Wheat	10.0
Maize	15.0
Wheat bran	5.7
Extracted sunflower meal I	16.5
Alfalfa meal I	11.7
Lysine	0.2
Limestone	1.5
Salt	0.3
Vitamin-mineral premix	0.5
Total	100.0

Statistical calculations. A nonsequential determination of least squares means and analysis of variance were carried out for the digestibility coefficients as well as for the data on mineral concentrations and enzyme activities. Student's *t*-test (for pair-wise comparison of group means) and one-way analysis of variance giving the least significant difference (LSD) at the level of 5% according to the procedure of SAS (Snedecor and Cochran, 1967; SAS, 1988) were used for the statistical analyses.

Results

The digestibilities of nutrients in the basal diet and in the treated carrots are shown in Table 2. The carrot diet covered 43% of the daily dry matter requirements, which meant the 'feed mixture' in the combined feed utilization experiment. The data presented in this paper provide evidence that the digestibility of carrots for each nutrient tended to be significantly higher than that of the basal diet. The lowest crude protein digestibility was found for samples containing Pb, while the lowest digestibility of ether extract was found for samples containing Hg and Cd. No differences were observed in the digestibility of other nutrients in the carrot samples.

Table 2

Digestibility of the basal diet and of carrots containing microelements, % (mean \pm SD; n = 5)

Nutrients	Basal diet	Carrot Mo	Carrot Cd	Carrot Pb	Carrot Hg	Carrot Se
DM	74.80 ^a	81.64 ^c	79.24 ^b	80.11 ^c	78.78 ^b	79.81 ^c
SD	2.36	2.97	2.47	1.90	1.25	0.88
OM	75.07 ^a	82.25 ^c	80.03 ^b	80.61 ^b	79.56 ^c	81.37 ^d
SD	2.64	2.83	2.41	2.10	1.18	0.63
CP	75.63 ^a	79.81 ^{be}	78.34	75.57 ^f	78.45	77.47
SD	2.32	2.41	2.79	2.22	2.79	2.12
CF	28.63 ^a	42.56 ^d	38.02 ^c	44.91 ^d	39.12 ^c	43.44 ^d
SD	3.11	3.86	3.39	2.86	3.53	2.94
EE	72.17 ^a	81.27 ^{dg}	74.27 ^{he}	78.03 ^{cf}	74.75 ^{he}	75.22 ^b
SD	1.34	1.70	2.66	2.26	2.28	2.04
NFE	81.34 ^a	87.29 ^d	87.08 ^d	87.20 ^d	86.41 ^d	87.88 ^d
SD	1.21	1.96	1.54	1.43	0.90	0.81

DM = dry matter; OM = organic matter; CP = crude protein; CF = crude fibre; EE = ether extract; NFE = N-free extract; a–b: P < 0.05; a–c: P < 0.01; a–d: P < 0.001; e–f: P < 0.05; g–h: P < 0.01

Carrot samples containing Cd resulted in a reduction of heart and liver weight. The heavy metal contents of the diet, urine, faeces and various organs and tissues are presented in Table 3.

The values clearly show that the rate of accumulation of microelements is different in the individual organs: Mo and Cd showed the highest accumulation in the kidneys, Pb in the kidneys, liver, bones and lungs, Hg in the kidneys and liver, while Se in the liver, kidneys and heart.

Table 3

Mo, Cd, Pb, Hg and Se content of basal diet, carrots, faeces, urine and different organs (ppm; n = 5)

	Basal diet Mo	Carrot Mo	Basal diet Cd	Carrot Cd	Basal diet Pb	Carrot Pb	Basal diet Hg	Carrot Hg	Basal diet Se	Carrot Se
Feed	0.527	39.000	0.140	2.300	1.580	4.010	0.000	30.000	0.000	36.200
Faeces	0.416	25.340	0.460	9.072	3.022	9.380	0.000	32.060	0.000	11.703
Urine	0.417	6.602	0.000	0.004	0.040	0.046	0.000	0.016	0.058	3.238
Heart	0.060	1.234	0.000	0.000	0.388	0.000	0.000	0.000	0.584	19.400
Lungs	0.033	1.206	0.007	0.029	0.644	0.507	0.000	0.000	0.727	14.748
Liver	1.262	1.878	0.119	0.722	1.717	1.846	0.000	3.530	1.738	64.950
Kidney	0.754	3.464	1.119	2.590	0.042	4.664	0.000	50.480	4.100	38.550
Spleen	0.000	1.083	0.012	0.000	0.759	0.151	0.000	0.080	1.990	15.350
Adipose tissue	0.000	0.060	0.000	0.000	0.141	0.060	0.000	0.000	0.000	0.634
Muscle	0.000	0.369	0.000	0.000	0.000	0.129	0.000	0.132	1.327	13.490
Bone	0.000	1.202	0.000	0.000	0.000	0.649	0.000	0.000	0.000	3.207
Hair	0.000	0.409	0.004	0.003	0.359	0.000	0.000	0.000	1.371	2.695

Table 4 shows the elimination of heavy metals via the urine and faeces and their accumulation in the tissues, expressed as a proportion of the amount taken up with the feed. It can be seen that Mo and Se are excreted from the body mainly through the faeces and urine, while Cd, Pb and Hg leave the body almost exclusively via the faeces. Cadmium builds up in the tissues in the largest quantity, followed by Se, Hg, Pb and Mo in decreasing order. The proportion of microelements eliminated from the body via the faeces and urine as a percentage of their intake was 80.18% for Mo, 37.86% for Cd, 66.39% for Pb, 64.65% for Hg, and 47.41% for Se.

Table 4

Intake, excretion and deposition of microelements and their proportions (n = 5)

	Intake	Excretion			Deposition
		via faeces	via urine	total	
Mo, mg/day	1.398	0.362	0.759	1.121	0.241
%	100.000	25.890	54.290	80.180	19.820
Cd, mg/day	0.103	0.039	–	0.039	0.064
%	100.000	37.860	0.000	37.860	62.140
Pb, mg/day	0.241	0.157	0.003	0.160	0.081
%	100.000	65.140	1.240	66.390	33.610
Hg, mg/day	0.116	0.055	0.020	0.075	0.041
%	100.000	47.410	17.240	64.650	35.350
Se, mg/day	1.350	0.187	0.453	0.640	0.710
%	100.000	13.850	35.550	47.41	52.59

Immediately before slaughter, the body weight of experimental rabbits was significantly lower ($P < 0.05$) than that of the control animals (Table 5). This can probably be attributed to the fact that the animals' dry matter uptake from the carrot diet was rendered difficult by physical factors rather than by the heavy metal content of the carrot. This is also supported by the finding that the body weight of the experimental groups did not differ. Histological examination revealed that the rate of spermatogenesis in the testis was reduced in the Mo, Cd, Pb and Hg groups as compared to the controls. A large number of syncytial giant cells and degenerated cells indicating abnormal meiotic divisions were found among the spermatogenic cells. Follicles containing healthy ova at different developmental stages were found in the ovaries of some does in the Mo, Se and control groups, but signs indicative of actual ovulation (corpus luteum) were not found.

Table 5
Weight of organs, g (mean \pm SD; n = 5)

Group	Body weight	Heart	Lungs	Liver	Spleen	Kidney
Control	2790 ^a	7.46 ^a	11.56	77.19	1.58	15.56
SD	193	1.28	1.15	7.13	0.37	1.68
Mo	2484 ^b	6.18	12.45	78.30	1.36	15.30
SD	124	0.85	2.24	6.56	0.10	0.93
Cd	2522 ^b	6.02 ^b	14.09	63.94 ^b	1.42	15.19
SD	99	0.40	2.31	8.66	0.16	2.84
Pb	2496 ^b	6.23	13.22	67.42	1.28	15.24
SD	134	0.75	1.30	9.63	0.17	1.65
Hg	2468 ^b	6.08	13.67	80.24	1.36	14.56
SD	146	0.71	2.04	6.81	0.17	1.50
Se	2515 ^b	6.16	13.56	78.90	1.31	14.46
SD	370	0.87	1.74	13.54	0.15	1.15

a-b: P < 0.05

With regard to changes in blood parameters (Table 6), it should be noted that Cd intake resulted in a considerable decrease in GGT and an increase in ALP activity. Both Cd and Hg had a similar effect on GGT and ALP activity. All experimental treatments decreased the activity of cholinesterase. ALT, AST, CK and creatinine levels did not show appreciable changes.

Table 6
Blood parameters I (means \pm SD; n = 5)

Group	AST U/l	ALT U/l	GGT U/l	ALP U/l	CHE U/l	CK U/l	CHOL U/l	TRIG g/l	GSH-Px U/g pro- tein	CREA μ mol/l
Control	4.40	22.40	8.00	55.20	735	364.0	1.81	1.150	20.00	109.80
SD	1.01	4.49	9.36	31.10	365	90.3	0.31	0.300	8.17	9.32
Mo	5.20	12.80	3.54	96.20	476	623.0	1.51	1.06	13.20	111.60
SD	1.60	3.31	1.84	22.40	391	211.4	0.14	0.38	3.96	7.47
Cd	3.60	17.60	1.88	123.00	429	640.0	1.97	0.714	17.00	119.80
SD	1.49	12.05	1.34	27.90	224	90.6	0.18	0.237	5.83	6.55
Pb	4.20	11.60	3.88	102.60	385	646.0	2.21	0.938	15.00	106.80
SD	1.02	2.65	3.70	21.60	267	260.2	0.93	0.234	6.75	8.05
Hg	5.00	17.80	1.40	96.40	339	523.0	1.97	0.900	13.60	113.20
SD	1.26	7.24	0.38	6.97	585	82.7	0.36	0.208	10.20	15.79
Se	9.75	17.25	2.50	70.50	412	522.0	2.37	0.827	13.20	102.50
SD	5.97	2.86	1.75	12.80	258	224.0	1.39	0.117	0.82	4.15

AST = aspartate aminotransferase; ALT = alanine aminotransferase; GGT = gamma-glutamyl-transferase; ALP = alkaline phosphatase; CHE = cholinesterase; CK = creatine kinase; CHOL = cholesterol; TRIG = triglyceride; GSH-Px = glutathione peroxidase; CREA = creatinine

The carrot diet high in Mo, Cd and Se increased GSH-Px activity by 22–50%, that rich in Pb lowered it by 18%, while Hg caused no change in the activity of that enzyme (Table 7).

Table 7Blood parameters II (means \pm SD; n = 5)

Group	Malondi- aldehyde mmol/l	Glutathione peroxidase		Reduced glutathione	
		nkat/g ml	nkat/g protein	μ mol/l	μ mol/g
Control	2.12	2.20	2.48	109.87	1.41
SD	0.14	0.23	0.52	22.30	0.24
Mo	2.68	2.37	3.15	106.52	1.57
SD	0.30	0.44	0.47	10.52	0.29
Cd	2.65	2.30	3.71	116.74	2.25
SD	0.45	0.59	0.75	8.57	0.29
Pb	2.44	1.46	2.04	118.25	1.78
SD	0.41	0.23	0.44	6.41	0.16
Hg	2.83	2.01	2.73	123.61	1.89
SD	0.64	0.16	0.40	6.20	0.26
Se	2.56	2.31	3.02	122.08	1.73
SD	0.43	0.10	0.38	2.70	0.36

Discussion

From the results of this experiment it seems that the negative effect exerted by high concentrations of the microelements studied can hardly, if at all, be monitored by determining the usual parameters (body weight gain, digestibility coefficients). Further examinations have revealed that Cd is the element that accumulates in the tissues to the greatest extent, resulting in decreased heart and liver weight. From the fact that Cd, Pb and Hg are excreted almost exclusively in the faeces it can be concluded that the absorbed amount of these elements remains in the body and their further excretion is very slow. In the trial of Hinesly et al. (1985) 60% of the retained Cd was found in the kidneys. The pathohistological examinations have shown that accumulation of these elements in the organism causes two main problems: it can impair spermatogenesis in breeding animals and pose a risk to consumers of products of animal origin.

As a rule, the uptake of Pb significantly reduced the activity of GGT and increased that of ALP. Cadmium and Hg loading also increased ALP activity. The altered activity of GGT and ALP indicates toxicity to the kidneys and the bones, respectively (Goreczky and Sós, 1983). All the treatments resulted in reduced activity of cholinesterase, which suggests damage of the liver parenchyma

(Kaneko, 1989). AST, ALT and CK activity and creatine content of the blood remained practically unchanged. The carrot diet containing Mo, Cd and Se increased the activity of GSH-Px by 20–50%, that containing Pb decreased it, while the dietary uptake of Hg had no appreciable effect on the activity of that enzyme. The changes found in GSH-Px activity show that the experimental treatments altered the organism's immunity to free radicals (Fehér és Verecke, 1985). According to Szilágyi et al. (1995), even the levels of metallothionein were significantly higher in the liver and kidneys of rabbits exposed to Cd.

Acknowledgements

This study was supported by a grant from the Hungarian Scientific Research Fund (OTKA). The authors thank Mr. J. Koncz for the skilful chemical analysis of minerals.

References

- Biacs, P. A., Daood, H. G. and Kadar, I. (1995): Effect of Mo, Se, Zn, and Cr treatments on the yield, element concentration, and carotenoid content of carrot. *J. Agric. Chem.* **43**, 589–591.
- Fairchild, E. J., Lewis, R. J. and Tatkin, R. L. (eds) (1977): Registry of Toxic Effects of Chemical Substances. DHEW Publ. No. (NIOSH) 78-104-B, 2, 526–534.
- Fehér, J. and Verecke, A. (1985): Importance of free residue reactions in medical science (in Hungarian). Biogal Pharmaceutical Factory, Medicina, Budapest.
- Fekete, S. and Gippert, T. (1982): Suggestions for the standardization of metabolic experiments with rabbits (in Hungarian). Research Report, Research Institute of Animal Breeding and Nutrition, Herceghalom.
- Goreczky, L. and Sós, J. (1983): Clinicochemical Laboratory Pocketbook (in Hungarian). Medicina, Budapest.
- Hinesly, D. T., Hansen, L. G., Bray, D. J. and Redborg, K. E. (1985): Transfer of sludge-borne cadmium through plants to chickens. *J. Agric. Food Chem.* **33**, 173–180.
- Kaneko, J. J. (ed.) (1989): Clinical Biochemistry of Domestic Animals. Fourth Edition. Academic Press, Inc., New York–London–Tokyo.
- National Research Council (1980): Mineral Tolerance of Domestic Animals. National Academy of Sciences, Washington, D. C.
- SAS/STAT™ User's Guide, Release 6.03 Edition (1988): SAS Institute, Inc., Cary, NC.
- Schenkel, H. W. (1988): Nutritive Beeinflussung des Stoffwechsel-verhalten von Cadmium bei Rind und Schwein. Habilitationsschrift, Hohenheim.
- Snedecor, G. W. and Cochran, W. G. (1967): Statistical Methods. 6th edition. Iowa State University Press, Ames.
- Stowe, H. D., Wilson, M. and Goyer, R. A. (1972): Clinical and morphologic effects of oral cadmium toxicity in rabbits. *Arch. Pathol.* **94**, 389–397.
- Szilágyi, M., Fekete, S., Sankari, S., Hullár, I., Zsolnainé Harcsi, I. and Kádár, I. (1995): Serum magnesium level and certain biochemical parameters in chicken and rabbits fed normal and heavy metals containing rations. *Proc. Soc. Nutr. Physiol.* **4**, 90–98.

