

Study on methods to extract high contrast image in active dynamic thermography

Saxena, Ashish; Raman, Vignesh; Ng, Eddie Yin Kwee

2019

Saxena, A., Raman, V., & Ng, E. Y. K. (2019). Study on methods to extract high contrast image in active dynamic thermography. *Quantitative InfraRed Thermography Journal*, 16(3-4), 243-259. doi:10.1080/17686733.2019.1586376

<https://hdl.handle.net/10356/144497>

<https://doi.org/10.1080/17686733.2019.1586376>

This is an Accepted Manuscript of an article published by Taylor & Francis in *Quantitative InfraRed Thermography Journal* on 19 Mar 2019, available online:
<http://www.tandfonline.com/10.1080/17686733.2019.1586376>.

Downloaded on 28 Aug 2022 05:19:02 SGT

1 Study on Methods to Extract High Contrast Image in Active 2 Dynamic Thermography

3 Ashish Saxena*, Vignesh Raman, EYK Ng

4 School of Mechanical and Aerospace Engineering, Nanyang Technological University, 50
5 Nanyang Ave, Singapore, 639798

6 *Corresponding Author: ashish008@e.ntu.edu.sg

7 Abstract

8 In the present study, image reconstruction methods are applied in active dynamic thermography
9 (ADT) to visualize the superficial blood vessel with high contrast. ADT is performed on the left
10 forearm of a human subject by applying cooling based external thermal stimulation. Both non-
11 parametrized, viz. sequence image, sequentially subtracted image, Discrete Fourier
12 Transformation (phase and amplitude image), and Thermographic Signal Reconstruction (TSR),
13 and parametrized, viz. Tau time (τ), dT_{norm} , t_{90-10n} , and Tissue Activity Ratio (TAR), types of
14 image reconstruction methods are used. To perform a quantitative comparison, among the image
15 reconstruction methods, the image contrast value is evaluated. While sequentially subtracted
16 image provides a high contrast image in non-parametrized image reconstruction methods, for
17 parametrized image reconstruction methods, it is TAR . Among all the methods considered, TAR
18 provides the best contrast image followed by τ image method.

19 **Keywords:** Active Dynamic Thermography; Thermal Image Reconstruction; Image Contrast;
20 Tissue Activity Ratio; Tau time.

1 **1. Introduction**

2 Active dynamic thermography (ADT), wherein transient sequence of images result upon
3 application of an external thermal or pressure excitation, is one of the most useful quantitative
4 thermography methods in medical diagnosis [1]. Since a diseased tissue responds differently to
5 the external excitation as compared to a normal tissue, ADT proves to be an efficient tool in
6 diagnosing various diseases, such as breast tumor detection, Raynaud's disease, burn wounds,
7 etc. [2]–[4]. While external excitation improves the image contrast that eases the target feature
8 extraction, it also leads to the generation of a large amount of sequence image data to be
9 analyzed. Moreover, the contrast of the image sequence decreases as the recovery phase
10 progresses upon removal of the excitation source [5]. Therefore, reconstruction of a single
11 synthetic image, which is both qualitatively and quantitatively advanced (feature contrast and
12 diagnosis parameter) as compared to the whole image sequence, is needed. To do so, application
13 of various image reconstruction algorithms, to ADT problems, is summarized hereafter.

14
15 It is well known that the original sequence images have a lower signal to noise ratio (SNR). SNR
16 can be improved by deriving phase and amplitude images using Fourier transformation (FT) [6],
17 [7]. In such an attempt, Boue et al. [8] evaluated the diameter of the superficial vein using
18 thermographic amplitude images, of which the contrast is found to be comparatively better than
19 the transient sequence images. Using fast Fourier transform (FFT), Liu et al. [9] showed that the
20 large sub-cutaneous veins can be distinguished from microvasculature and skin tissue without the
21 blood vessel, respectively, in the frequency range of 0.005 to 0.06 Hz. Given that an adequate
22 contrast enhancement cannot be achieved in all the patients, application of thermographic signal

1 reconstruction (TSR) method, which is conventionally used in the field of material
2 characterization and defect detection by ADT [10]–[13], is shown in the work of Liu et al. [14].
3 The authors have applied a 5-minute cuff-based occlusion, followed by transient thermal image
4 acquisition during the reactive hyperaemia. At each pixel, the coefficient values are derived from
5 the first derivative of the logarithmic equation fit to the temperature response over time, and a
6 synthetic image is reconstructed. The authors have shown a substantial improvement in SNR that
7 helps in clear visualization of the angioarchitecture in the synthetic image.

8

9 Since ADT involves a transient process of recovery to normal state post external excitation, to
10 reconstruct the synthetic image, pixel specific parameters pertaining to thermal recovery can also
11 be extracted. One such parameter is the thermal time constant (called Tau time, ' τ ') that
12 quantifies the thermal activity of the tissue. Given the apparent difference in heat transfer rate
13 between the skin tissue with and without vein, it is imperative that the τ time evaluation would
14 differentiate the two; hence, brings out the contrast in the resultant τ image. This image can be
15 used to diagnose skin tissue with pathological conditions as described in the work of Foerster et
16 al. [15], wherein the authors have developed a τ image based tool to diagnose Raynaud's
17 phenomenon (RP). Further, to estimate the τ value more accurately, a non-linear regression-
18 based exponential curve fitting to the experimental data can be done. This method is applied to
19 monitor the cardiosurgery [16]–[18], evaluation of burn wound healing [19]–[21], vascular
20 disease diagnosis [22], etc. Recently, Moderhak et al. [23] described a dynamic thermography
21 method, wherein 60 seconds of thermal excitation and 180 seconds of recovery time is used, to
22 quantitatively determine the success of breast reconstruction procedure by skin flap perfusion
23 assessment. The authors have used a Simplified Magnitude-Temporal Parametrization method

1 (SMTP) to define two unique parameters: dT_{norm} and t_{90-10n} [24], at each pixel, to reconstruct
2 a single image from the ADT sequence, respectively. With the help of the improved contrast
3 images, reconstructed using these two parameters, the authors have shown that the two groups of
4 patients, who will either develop post-surgery necrosis complications or not, can be diagnosed at
5 an early stage. In our recent work [25], a novel method of single image reconstruction,
6 *TAR* image, is introduced. Comparing the new method with the existing τ image method, in three
7 subjects, it is reported that the *TAR* image provides a quantitative and qualitative high contrast
8 image.

9
10 It is evident from the literature review that there exist many single image reconstruction methods
11 that can be applied to the ADT sequence. The reconstructed single thermal image is useful for
12 both qualitative and quantitative analysis of the tissue health monitoring. However, a
13 comprehensive comparison of various image reconstruction methods, applied to a single ADT
14 problem, is not found in the literature. Therefore, in the present study, with the application of
15 cooling excitation, ADT sequence is captured, on the forearm of a human subject, to visualize
16 the superficial vein with high contrast. To do so, various pre-existing image reconstruction
17 methods, viz. FT image (amplitude and phase), TSR image, and parametric image
18 (τ , dT_{norm} , t_{90-10n} , and *TAR*), are used, and a comparative study is done. A contrast factor (W),
19 to quantitatively compare the images obtained from each of the image reconstruction methods
20 studied, is introduced.

1 **2. Method and Material**

2 **2.1. Experiment Setup and Procedure**

3 In-vivo human subject tests are carried out on 3 male subjects (age: 24 ± 1 years), under an
4 approved ethical study, at Nanyang Technological University, Singapore (IRB: SHS-
5 NTU/014/2016). In this experiment, a continuous cooling is applied on the left forearm of each
6 subject for 30 seconds. The cooling application is done with the help of a cooling pad [26]
7 recirculated with ice water at a temperature of 5 ± 0.5 °C. After the removal of cooling, 90
8 seconds of sequential thermal images are captured by an Infrared (IR) thermal camera
9 (VarioCam by InfraTec) at a rate of 2 frames per second (Figure 1). Experiments are performed
10 after allowing the subject to sit for 15 minutes in a controlled temperature environment of $22 \pm$
11 0.5 °C. In each subject, the average value of temperature with standard deviation (\pm SD), over the
12 skin tissue (within the cooling stimulation zone) at the start of the experiment, after the removal
13 of cooling, and end of rewarming, is summarized in Table 1.

14 **2.2. Non-parametric image reconstruction methods**

15 **2.2.1. Discrete Fourier Transformation**

16 For the ADT experiment performed in the present study, Figure 2 shows the variation of skin
17 temperature (T_{skin} in °C) normalized by core body temperature ($T_{core} = 37$ °C) versus time (s).
18 On the removal of the external excitation, to transform the resultant time domain temperature
19 progression into frequency domain, at each pixel within the cooling zone, 1-Dimensional (1-D)
20 Discrete Fourier Transform (DFT) can be applied. The equation for 1-D DFT is given below [7]:

$$21 \quad F_n = \Delta t \sum_{k=0}^{N-1} T(k\Delta t) e^{\frac{-j2\pi nk}{N}} = Re_n + Im_n \quad (1)$$

1 Where, n designates the frequency increment ($n=0,1, 2, \dots,N$), k is the discrete number of
 2 temperature (T in °C) samples, Re_n and Im_n represents the real and imaginary parts of the
 3 transform, respectively, at given n , j is the imaginary number, and Δt represents the time interval
 4 (s). Using the result of Eq. (1), the modulus or amplitude (A in °C) and phase delay (φ in rad) is
 5 computed using the following formulae:

$$6 \quad A = \sqrt{Re_n^2 + Im_n^2} \quad (2)$$

$$7 \quad \varphi = \arctan\left(\frac{Im_n}{Re_n}\right) \quad (3)$$

8 Application of DFT, to the temperature profile obtained after the removal of cooling excitation,
 9 results into a Hermite function [27]. A Hermite function has a symmetrical real and an
 10 asymmetrical imaginary part (at $N/2$). Since both the first and second half of the frequency range
 11 (around 0 Hz) produce alike results, only $N/2$ useful frequencies are considered for further
 12 analysis. For the retained $N/2$ frequencies, the corresponding phase and amplitude images are
 13 extracted.

14 2.2.2. Thermographic Signal Reconstruction (TSR)

15 In this method, a logarithmic polynomial function of degree m (Equation 4) is fit to the \log of
 16 temperature versus time plot, at each pixel, after the removal of cooling.

$$17 \quad \log_{10}(dT) = a_0 + a_1 \log_{10}(t) + a_2 [\log_{10}(t)]^2 + \dots + a_m [\log_{10}(t)]^m \quad (4)$$

18 where, dT is the temperature (°C) increase as a function of time, t (s). a_m is the coefficient of the
 19 polynomial function of degree m . The polynomial function is chosen such that the transient
 20 series of images can be reduced to $(m + 1)$ number of images. The choice of polynomial degree
 21 m is conventionally done with the help of an optimization method, wherein images

1 corresponding to the range of m and its respective coefficient ranks are compared to select the
2 best contrast image revealing maximum thermal expressions [12]. Using this chosen polynomial
3 degree, first derivative images are extracted for further analysis.

4 **2.3. Parametric Image Reconstruction methods**

5 **2.3.1. Tau Image:**

6 The natural rewarming of the skin tissue, to the cooling excitation, can be analyzed using lump
7 capacity analysis model, wherein a uniform rewarming is assumed under the influence of
8 ambient convection thermal boundary condition [28]. This process can be characterized with the
9 help of thermal time constant (called Tau time, ' τ ') which is equal to the time required for a
10 system response to decay to zero [29]. Further, as shown in the work of Dupuis [4], Foerster et
11 al. [15], Jankovic et al. [22], Meffert et al. [30], Merla et al. [31], etc., the rewarming process
12 closely resembles to an exponential response curve described by the transient first order response
13 as given by Equation 5.

$$14 \quad T(t) = T_o + \Delta T(1 - e^{-t/\tau}) \quad (5)$$

15 where, $T(t)$ is the temperature ($^{\circ}\text{C}$) at any time instant, t (s), T_o is the initial temperature ($^{\circ}\text{C}$),
16 ΔT is the total temperature change ($^{\circ}\text{C}$), and τ is the Tau time (s). When the rewarming process
17 reaches the τ time, the temperature of the system attains $(1 - 1/e)$ times of total temperature
18 difference between the initial and post excitation temperature (dT_{excite} in $^{\circ}\text{C}$), which is equal to
19 63% of dT_{excite} . At each pixel, Tau time (τ) is calculated (Figure 2), followed by replacing the
20 original sequence image pixel value with the evaluated τ value, and thereby, a τ image is
21 reconstructed. This τ image is an equivalent representation of the whole thermal image
22 sequence.

1 2.3.2. dT_{norm} and t_{90-10n} Image:

2 As defined in the work of Moderhak et al. [23], dT_{norm} and t_{90-10n} is calculated as follows:

3
$$dT_{norm} = \frac{dT_{recovery}}{dT_{excite}} \quad (6)$$

4
$$t_{90-10n} = \frac{t_{90-10}}{t_{recovery}} \quad (7)$$

5 where, $dT_{recovery}$ is the temperature difference ($^{\circ}\text{C}$) between the start and end of the allowed
6 experimental thermal recovery, dT_{excite} is the temperature difference ($^{\circ}\text{C}$) between the start and
7 end of the thermal excitation, t_{90-10} is the difference in time (s) to reach 90% and 10% of the
8 $dT_{recovery}$, respectively, ($t_{90-10} = t_{90\% \text{ of recovery}} - t_{10\% \text{ of recovery}}$), and $t_{recovery}$ is the total
9 experimental time (s) of the thermal recovery (Figure 2).

10 2.3.3. Tissue Activity Ratio (TAR) Image

11 In this method, a novel approach to reconstruct a unique image, based on the individual pixel's
12 thermal activity, is used [25]. On removal of the external cooling, it is evident that pixel just
13 above the blood vessel would show a higher thermal activity as compared to the other pixels;
14 hence forms the basis for a qualitatively and quantitatively advanced image. Using Equation 8,
15 the thermal activity during the rewarming phase, at each pixel, can be quantified with the help of
16 a ratio called as Tissue Activity Ratio (TAR). TAR accounts for both the rewarming as well as
17 the excitation phase. First, a reference recovery temperature (T_r in $^{\circ}\text{C}$), for all the pixels within
18 the excitation zone, is fixed. This is done by finding the maximum value of temperature reached
19 at each pixel during the recovery phase, and then select the minimum of these values. The
20 selected T_r value ensures that all the pixels will reach at least this temperature within the
21 acquired recovery duration (90s in the present study). Using this T_r value, at each pixel, the time

1 of rewarming (t_r in s), to rewarm from the temperature at the end of cooling (T_c in °C) to T_r , is
2 evaluated and thus, the rate of rewarming (RR in °C/s) is calculated (Equation 9). Further, the
3 rate of cooling (RC in °C/s), at each pixel, is calculated (Equation 10) by subtracting the
4 temperature at the end of the cooling (T_c in °C) from the initial temperature (T_i in °C) before
5 application of cooling, and dividing the resultant temperature difference by the total duration of
6 application of cooling (t_c in s). Unlike the value of t_r in the rewarming phase, the time for
7 cooling (t_c) will be same for all the pixels. The mathematical formulation to calculate TAR , at
8 each pixel, within the cooling zone is given below:

$$9 \quad TAR = \frac{RR}{RC} \quad (8)$$

$$10 \quad \text{where, } RR = \frac{T_r - T_c}{t_r} \quad (9)$$

$$11 \quad RC = \frac{T_i - T_c}{t_c} \quad (10)$$

12 Computing TAR value at each pixel, a unique TAR image is reconstructed.

13 **2.4. Image Contrast Analysis**

14 To compare and evaluate the quality of an image, a contrast factor (W) is evaluated. The contrast
15 factor defines the relative strength of an image target parameter with respect to the background.

16 In the present study, the feature of interest (target) is the superficial vein around the skin tissue
17 (background). A high value of W refers to a better visibility of the superficial vein in the image.

18 Using the Weber's formula [32], W is defined as follows:

$$19 \quad W = \left| \frac{I - I_b}{I_b} \right| \quad (11)$$

1 where, I and I_b are the image parameter over the skin tissue with (target) and without superficial
2 vein (background), respectively. In the present study, average values of I and I_b are taken from
3 the reference sampling locations (black and red-dashed rectangular boxes) as shown in Figure 3.
4 While I is taken over the superficial vein (target), I_b is taken from the neighboring skin tissue
5 (background). These reference sampling locations, in each subject, are consistently used for all
6 the images extracted from the various image reconstruction methods studied.

7

8 In Equation 11, the value of I_b is essentially less than the value of I [32], therefore, the value of
9 W provides a measure of difference in the image parameter, between the two regions (target and
10 background), with respect to the minimum of the image parameter over the two regions. By
11 default, it is considered that the background will have the minimum image parameter value as
12 compared to the target ($I > I_b$), however, this is not true in all the cases. For comparison
13 purpose, if all the images, resulting from different image reconstruction methods, provide a trend
14 of either $I > I_b$ or $I < I_b$ consistently, the resultant contrast comparison is balanced. However, if
15 any of the images deviate from the adopted consistent trend, the resultant contrast comparison is
16 imbalanced. In the present study as well, $I > I_b$ is adopted as a consistent trend. To perform a
17 balanced comparison, irrespective of $I > I_b$ or $I < I_b$ trend in the images, the Weber's formula
18 (Equation 11) should be modified (W') such that the minimum of I or I_b should be used in the
19 denominator (as given by Equation 12).

$$20 \quad W' = \left| \frac{I - I_b}{\min(I, I_b)} \right| \quad (12)$$

21 On the other hand, to maintain the standardized usage of the Weber's contrast formula (Equation
22 11) and present a balanced comparison among the adopted ($I > I_b$) and deviating ($I < I_b$) trend

1 images, the deviating trend images can be transformed using the following inversion

2 formulation:

$$3 \quad \phi'_{ij} = \left(\frac{1}{\phi_{ij}}\right) * \phi_{ijmax} \quad (13)$$

4 where, i and j are the number of row and column, respectively, to locate the target pixel, ϕ'_{ij} and

5 ϕ_{ij} are the transformed and original image parameter, respectively, and ϕ_{ijmax} is the maximum

6 image parameter value of all the pixels in the original image. Using Equation 13, the value of I

7 and I_b (where, $I < I_b$) can be transformed as follows:

$$8 \quad I' = \left(\frac{1}{I}\right) * \phi_{ijmax} \quad (14)$$

$$9 \quad I'_b = \left(\frac{1}{I_b}\right) * \phi_{ijmax} \quad (15)$$

10 The proposed transformation results into an equivalent image as the original image but with an

11 inverted scale (inverse proportionality), which provides the desired $I' > I'_b$ trend. Given the

12 multiplication with ϕ_{ijmax} in Equation 14 and 15, the resultant I' and I'_b are dimensionless in

13 nature. Using the standard Weber's contrast formula (Equation 11), the contrast for the

14 transformed image (with I' and I'_b) can be calculated as follows:

$$15 \quad W = \left| \frac{I' - I'_b}{I'_b} \right|, \text{ where } I' > I'_b \quad (16)$$

16 Replacing the value of I' and I'_b from Equations 13 and 14, Equation 15 will result into Equation

17 17.

$$18 \quad W = \left| \frac{I_b - I}{I} \right|, \text{ where } I < I_b \quad (17)$$

1 Since Equation 17 follows the generic nature defined by the modified Weber's contrast formula
2 in Equation 12 (using minimum of I or I_b in the denominator for a balanced comparison among
3 the adopted ($I > I_b$) and deviating ($I < I_b$) trend images), the contrast calculated from the
4 standard Weber's formula (Equation 11), using a transformed value of I and I_b (where, $I < I_b$),
5 will provide a balanced comparison with the images following the adopted $I > I_b$ trend.

6 **3. Results and Discussion**

7 ***3.1. Transient sequence images***

8 After the removal of the external stimulation (cooling), before reaching the original thermal
9 equilibrium with the ambient surroundings, the skin tissue undergoes a transient recovery phase.
10 The thermal activity of the skin tissue, due to blood perfusion and blood flow in the superficial
11 vein, determines the rate of the recovery process. It is evident that the skin tissue over the
12 superficial vein will recover faster as compared to the surrounding skin tissue with no such vein.
13 This brings out the contrast in the visibility of the superficial vein. However, the dynamic
14 sequence images do not hold a consistent clear visibility of the blood vessel as the recovery
15 progresses further. Given the rate of recovery is directly proportional to the temperature
16 difference, due to a continuous decrease in the temperature difference, the rate of recovery slows
17 down as the recovery progresses. Therefore, the visibility of the superficial vein with reference to
18 the nearby skin tissue region first increases and then decreases. To quantify this characteristic,
19 using Equation 11, the contrast of the superficial vein, in all the sequence images, is calculated.
20 Plotting the superficial vein contrast, in all subjects, against time, Figure 4 corroborates with the
21 fact that the visibility of the superficial vein changes dynamically throughout the recovery phase.

1 **3.2. Non-parametric image reconstruction**

2 *3.2.1. Best contrast image from the sequence*

3 Marking the peak of the contrast curve in Figure 4, the maximum contrast point ($W_{max.}$) is
4 determined, which is in the range of 0.2 to 0.6 in the three subjects studied. Using the $W_{max.}$
5 point, a single high contrast image, from the transient sequence of images, can be selected. In
6 each subject, Table 2 shows the image corresponding to $W_{max.}$ point. A contrast value of less
7 than 1 suggests a poor visibility of the superficial vein. It is to be noted that the occurrence of
8 $W_{max.}$ point differs in all the subjects. This is because of the different morphological and
9 hemodynamic characteristics, like thickness of the skin tissue, diameter of the superficial vein
10 and its blood flow velocity, skin tissue blood perfusion rate, etc., which bring varied intensity of
11 thermal activity in each subject and hence, either speed-up or delay the occurrence of
12 $W_{max.}$ point. In Subject-2, the $W_{max.}$ point occurs much later ($W_{max.} = 0.21$ at 40 s) as compared
13 to Subject-1 ($W_{max.} = 0.56$ at 22 s) and Subject-3 ($W_{max.} = 0.61$ at 26.5 s).

14 *3.2.2. Sequential image subtraction*

15 To further enhance the contrast of the sequence images, a primitive noise subtraction technique
16 can be used [26], wherein the very first image after the removal of the cooling (external thermal
17 excitation) is subtracted from the subsequent images in the recovery phase. Given the image to
18 be subtracted is always the same, it is imperative that the contrast of all the subsequent images
19 will increase by a same fraction. Hence, in each subject, to evaluate the highest enhancement in
20 the contrast, the subtracted image corresponding to the $W_{max.}$ point in the original sequence
21 images is selected. From Table 2, it can be observed that the qualitative visibility of the
22 superficial vein in the subtracted images, in all subjects, is better than the corresponding original

1 sequence images. Quantitatively, from Table 5, the contrast of the subtracted images, in all the
2 three subjects, is found to be improved by 52%, 76%, and 28%, respectively, as compared to the
3 corresponding sequence images.

4 3.2.3. *Amplitude and phase image*

5 From the amplitude and phase images, obtained across multiple frequencies through DFT
6 processing, the blind frequency, at which the region of interest (RoI) in the image is clearly
7 visible with high contrast, is determined; RoI corresponds to the superficial vein in the present
8 study. From the literature, it is noticed that there are mainly two methods to find out the blind
9 frequency [27]. In the first method, the phase value over the RoI is plotted against the frequency
10 and the frequency corresponding to the point of inflexion is chosen as the blind frequency. In the
11 second method, the phase contrast between the RoI and the surrounding is plotted against the
12 frequency values, and the frequency corresponding to the maximum contrast value is chosen as
13 the blind frequency. In the present study, the latter method is adopted, wherein using the Weber's
14 contrast formula (Equation 11), the best contrast amplitude and phase images are extracted
15 (Table 2). Given the phase image, for Subject-1 and 2, shows a lower descriptor value over the
16 blood vessel as compared to the background ($I < I_b$), contrast value is calculated after
17 performing the image transformation using Equation 13. For subject-1, 2, and 3, the blind
18 frequencies for phase images are found to be 0.19 Hz, 0.1 Hz, and 0.18 Hz, respectively. Except
19 for Subject-2, the best contrast amplitude images occur at a different frequency than the blind
20 frequency, which is 0.23 Hz and 0.09 Hz for Subject-1 and Subject-3, respectively. From Table
21 5, it can be observed that the contrast value of amplitude images is always higher than phase
22 images. This can be further corroborated by the visual inspection of the images in Table 2, where
23 the phase images, in all subjects, show more non-uniformity in the background skin tissue

1 regions as compared to amplitude images; this degrades the visible quality of the target
2 superficial vein. However, the qualitative clarity of superficial vein in the phase images is found
3 to be better than the amplitude images, viz. the visibility of the branching blood vessel in
4 Subject-1 and the diffusion around the superficial vein in Subject-2 and 3.

5 *3.2.4. TSR coefficient image*

6 In the present study, three degrees of polynomial ($m = 4, 5$ and 6) are tested for the best
7 contrast image. Using these polynomial equations, at each pixel, the first derivative is calculated,
8 and the respective coefficient values (a_m) are used to extract m number of images. To select the
9 best image, in each subject, contrast value (W using Equation 11) is calculated, and the
10 coefficient image with highest contrast value is selected. As observed in Table 3, the best
11 contrast image, for both Subject-1 and 2, is found at $m = 4$ and a_4 , while for Subject-3, it is
12 found at $m = 5$ and a_1 . However, the corresponding images (Table 2) do not comply with the
13 high contrast values in Table 5. This high contrast value is the outcome of the outlier regions in
14 the image; leads to a very low value of the background image parameter (I_b) as compared to the
15 target image parameter (I). This effect is mainly observed in Subject-2 and 3, which results into
16 a qualitatively poor image but with an unrealistically high contrast value.

17 **3.3. Parametric image reconstruction**

18 Reconstruction of a single image, from ADT sequence, not only provides a qualitative
19 representation, but also includes quantitative information. This quantitative information, at each
20 pixel, provides the heat flow characteristics. To do so, quantitative parameters, during the
21 rewarming phase, can be defined. The basis for any of these parameters is the quantification of
22 thermal activity of the tissue, which can be defined as the measure of degree of responsiveness to

1 the external stimulation. This responsiveness, in terms of heat transfer, is the result of the tissue
2 blood perfusion. In the present study, other than the skin tissue perfusion, presence of a
3 superficial vein, which acts as an additional heat source, leads to a higher responsiveness to the
4 external stimulation as compared to the tissue with no such superficial vein. Evaluated using a
5 quantitative parameter, this higher responsiveness brings contrast to the visibility of target
6 superficial vein in the image. The quantitative parameters used to reconstruct the single thermal
7 image are τ , dT_{norm} , t_{90-10n} , and TAR , of which the resultant images, in each subject, are
8 shown in Table 4. As defined in Section 2.2.3 and 2.2.4, of the four quantitative parameters, τ
9 has a unit of seconds (s), while the other three are dimensionless. Given the recovery rate is
10 higher over the superficial vein, the τ value is lower and vice-versa for the tissue with no
11 superficial vein. Therefore, to calculate the contrast value (W), a transformed τ image is used as
12 explained in Section 2.3. A similar image transformation is performed for t_{90-10n} images.
13 Except for Subject-2, the τ image in other two subjects provide a contrast value of higher than 1
14 (Table 5), which can be corroborated from the visual inspection of the images (Table 4).

15

16 For the case of dT_{norm} and t_{90-10n} , the superficial vein part of the reconstructed images, in all
17 subjects, is found distorted and blurred (Table 4) which is reflected in the lower value of contrast
18 as well (Table 5). In a numerical study, on comparison of parametrization methods applied to
19 ADT on a skin flap model with blood vessel perforator, by Moderhak [24], the contrast value, for
20 the first order τ , dT_{norm} , and t_{90-10n} images, is found to be 0.05, 0.11 and 0.06, respectively.
21 For the τ and t_{90-10n} images, it should be noted that the contrast is calculated without any image
22 transformation. Using the proposed transformation in the present study (Equation 13), the new
23 value of contrast, for τ and t_{90-10n} images in [24], is calculated to be 0.06 and 0.08,

1 respectively. The contrast value for dT_{norm} and t_{90-10n} images in both the studies, numerical
2 [24] and present experimental, is found to be less than 1. Given, the ADT procedure and the skin
3 tissue model used, in the numerical study and the present experimental study, are different,
4 comparison among the same kind of parametric images, between the two studies, is not possible.
5 However, the relative difference in the outcome of the different parametric image reconstruction
6 methods, between the two studies, can be compared. From τ to dT_{norm} and t_{90-10n} , the
7 numerical study shows an improvement of 33% and 83% in the image contrast value,
8 respectively, while, among the three subjects, the present experimental study shows a minimum
9 and maximum improvement of 50% and 493% from dT_{norm} to τ and 173% and 786% from
10 t_{90-10n} to τ , respectively (Table 5).

11
12 Being one of the parametric methods that depends on the tissue recovery rate, the lower value of
13 contrast in case of dT_{norm} and t_{90-10n} images is possibly because of the inclusion of data until
14 the end of recovery phase as per the definition of these parameters (Equations 6 and 7). It is
15 imperative that as the recovery process approaches towards the end, the temperature difference,
16 between the current skin tissue recovery temperature and the initial temperature (T_i), decreases,
17 which leads to a reduction in the rate of recovery (Figure 2). Hence, the contrast between the
18 tissue with and without vein also decreases, which affects the overall quality of the image;
19 resulting into blurred and distorted images (dT_{norm} and t_{90-10n} images in Table 4). Hence, to
20 satisfy both the conditions, viz. use of rate of recovery as a parameter while considering the data
21 from the initial stages of the recovery process, the TAR parameter is used. This is done by
22 defining a fixed minimax reference recovery temperature value (T_r), within the total recovery
23 phase duration, for all the pixels in the cooling zone. Given the fact that the tissue with

1 superficial vein will rewarm to T_r temperature with a lower value of t_r , while a higher value of
2 t_r for the same T_r over the tissue with no superficial vein, the resulting TAR value will have a
3 higher contrast between the two regions. Visual inspection of the TAR images (Table 4), in all
4 the subjects, reveals a high qualitative visibility of the superficial vein. As compared to the other
5 image reconstruction methods used in the present study, the TAR images are found to be much
6 better in all subjects. Quantitatively, the contrast value of TAR image, in all the subjects, is found
7 to be more than 1, which signifies a higher visibility of the superficial vein with respect to the
8 background (Table 5).

9 **3.4. Comparative analysis**

10 For each subject, comparing among the image reconstruction methods considered, the value of
11 contrast (W) is found to be highest for TAR method along with a high-quality image, except for
12 TSR coefficient method in Subject-2 and 3 (contrast values do not comply with the respective
13 images). Other than the TAR image method, τ image is the only method where a contrast value
14 of more than 1 is achieved (except for Subject-2). For the Subject-1, 2, and 3, the improvement
15 in contrast value from τ image to TAR image is found to be 33%, 243%, and 19%, respectively.
16 For Subject-2, excluding the TSR coefficient method from the analysis, except for TAR image
17 method with a contrast value of 1.03, all other methods give a contrast value in the range of 0.20
18 to 0.37. Among the non-parametric image reconstruction methods, except for TSR coefficient
19 method, subtracted image provides the highest contrast value. The contrast of the subtracted
20 images, in all the subjects, is found even higher than two of the parametric image reconstruction
21 methods, i.e. dT_{norm} and t_{90-10n} . Unlike for Subject-1 and 3, a similar trend is observed for
22 parametric τ image in Subject-2.

1 **4. Conclusions**

2 Active dynamic thermography (ADT) is performed on the forearm of three human subjects,
3 using cold thermal stimulation, to produce high contrast image of the superficial vein with the
4 application of single image reconstruction methods. These methods are categorized into non-
5 parametrized and parametrized groups. While for non-parametrized groups, the best image is
6 selected using maximum contrast values, for non-parametrized group, the output image is itself
7 the best contrast image. Further, a qualitative and quantitative comparative study is performed,
8 wherein it is found that the parameterized based *TAR* image reconstruction method provides the
9 best contrast image, followed by Tau time (τ) image method. In the non-parametrized image
10 reconstruction group, the best method is found to be sequentially subtracted image method.
11 Given the conclusions of the present study is limited to ADT experimentation on three human
12 subjects, subsequent studies on a larger patient cohort shall be performed in future.

13 **Disclosure statement**

14 No potential conflict of interest was reported by the authors.

15 **Funding**

16 The work has been supported by SingHealth-NTU collaborative research grant (Grant number:
17 SHS-NTU/014/2016).

18 **Acknowledgment**

19 The authors would like to acknowledge their heartfelt gratitude to Eetarp Engineering Pte. Ltd.,
20 Singapore for providing the IR thermal camera support, and Mr. Gaurav Singh, Ph.D. Scholar,

- 1 School of Computer Science and Engineering, NTU Singapore, for his consultation on image
- 2 processing.

References

- [1] Y. V. Gulyaev, A. G. Markov, L. G. Koreneva, and P. V. Zakharov, "Dynamical Infrared Thermography in Humans," *IEEE Eng. Med. Biol. Mag.*, vol. 14, no. 6, pp. 766–771, 1995.
- [2] Y. Ohashi and I. Uchida, "Applying dynamic thermography in the diagnosis of breast cancer," *IEEE Eng. Med. Biol. Mag.*, vol. 19, no. 3, pp. 42–51, 2000.
- [3] A. Nowakowski, "Quantitative Active Dynamic Thermal IR-Imaging and Thermal Tomography in Medical Diagnostics," in *Medical Devices and Systems*, 3rd ed., J. D. Bronzino, Ed. Taylor & Francis Group, LLC, 2006, pp. 1–30.
- [4] H. Dupuis, "Thermographic assessment of skin temperature during a cold provocation test," *Scand. J. Work. Environ. Heal.*, vol. 13, no. 4, pp. 352–355, 1987.
- [5] R. B. Barnes, "Barnes : Thermography," *Ann. N. Y. Acad. Sci.*, vol. 121, pp. 34–38, 1964.
- [6] N. Bouzida, A. H. Bendada, J.-M. Piau, M. Akhloufi, X. Maldague, and M. Raymond, "Using lock-in infrared thermography for the visualization of the hand vascular tree," in *Proc. of SPIE*, 2008, vol. 6939, pp. 1–12.
- [7] D. Wu, H. Hamann, A. Salerno, and G. Busse, "Lockin thermography for imaging of modulated flow in blood vessels," *Conf. Quant. InfraRed Thermogr. Pisa*, pp. 1–5, 1997.
- [8] C. Boué, F. Cassagne, C. Massoud, and D. Fournier, "Thermal imaging of a vein of the forearm: Analysis and thermal modelling," *Infrared Phys. Technol.*, vol. 51, no. 1, pp. 13–20, 2007.
- [9] W.-M. Liu, J. Meyer, C. G. Scully, E. Elster, and A. M. Gorbach, "Observing temperature

- fluctuations in humans using infrared imaging,” *Quant. Infrared Thermogr. J.*, vol. 8, no. 1, pp. 21–36, 2011.
- [10] B. Oswald-Tranta, A. Maier, and R. Schledjewski, “Defect depth determination in a CFRP structure using TSR technique,” in *Proceedings of the 2014 International Conference on Quantitative InfraRed Thermography*, 2014, pp. 1–8.
- [11] F. López, V. P. Nicolau, C. Ibarra-Castanedo, S. Sfarra, and X. Maldague, “Comparative study of thermographic signal reconstruction and partial least squares thermography for the detection and evaluation of subsurface defects Flash-lamps,” in *QIRT 2014 Conference*, 2014, no. 1, pp. 1–10.
- [12] J. Roche, F. Leroy, and D. L. Balageas, “Information condensation in defect detection using TSR coefficients images,” in *QIRT2014 Conférence*, 2014, no. 1, pp. 1–10.
- [13] D. L. Balageas, J. M. Roche, F. H. Leroy, W. M. Liu, and A. M. Gorbach, “The thermographic signal reconstruction method: A powerful tool for the enhancement of transient thermographic images,” *Biocybern. Biomed. Eng.*, vol. 35, no. 1, pp. 1–9, 2015.
- [14] Liu, Wei-Min, Jordan Maivelett, Gregory J. Kato, James G. Taylor, Wen-Chin Yang, Yun-Chung Liu, You-Gang Yang, and Alexander M. Gorbach, “Reconstruction of thermographic signals to map perforator vessels in humans,” *Quant. Infrared Thermogr. J.*, vol. 9, no. 2, pp. 123–133, 2012.
- [15] Foerster, J., S. Wittstock, S. Fleischanderl, A. Storch, G. Riemekasten, O. Hochmuth, B. Meffert, H. Meffert, and M. Worm, “Infrared-monitored cold response in the assessment of Raynaud’s phenomenon,” *Clin. Exp. Dermatol.*, vol. 31, no. 1, pp. 6–12, 2005.
- [16] M. Kaczmarek, A. Nowakowski, M. Suchowirski, J. Siebert, and W. Stojek, “Active

- dynamic thermography in cardiosurgery,” *Quant. Infrared Thermogr. J.*, vol. 4, no. 1, pp. 107–123, 2007.
- [17] Nowakowski A, Kaczmarek M, Stojek W, Beta S, Trzeciak B, Topolewicz J, Rogowski J, Siebert J, “IR-thermal monitoring of cardiosurgery interventions,” in *IFMBE Proceedings* 22, 2008, pp. 1329–1333.
- [18] A. Nowakowski, P. Siondalski, M. Moderhak, and M. Kaczmarek, “A new diagnostic method for evaluation of cardiosurgery wound healing,” *Quant. Infrared Thermogr. J.*, vol. 13, no. 1, pp. 19–34, 2016.
- [19] A. Renkielska *et al.*, “Active dynamic infrared thermal imaging in burn depth evaluation,” *J. Burn Care Res.*, vol. 35, no. 5, pp. e294–e303, 2014.
- [20] A. Renkielska, A. Nowakowski, M. Kaczmarek, and J. Ruminski, “Burn depths evaluation based on active dynamic IR thermal imaging-A preliminary study,” *Burns*, vol. 32, no. 7, pp. 867–875, 2006.
- [21] J. Rumiński, M. Kaczmarek, A. Renkielska, and A. Nowakowski, “Thermal parametric imaging in the evaluation of skin burn depth,” *IEEE Trans. Biomed. Eng.*, vol. 54, no. 2, pp. 303–312, 2007.
- [22] S. Jankovic, S. Stankovic, S. Borjanovic, L. Tenjovic, and M. Bogdanovic, “Cold stress dynamic thermography for evaluation of vascular disorders in hand-arm vibration syndrome,” *J Occup Heal.*, vol. 50, no. 5, pp. 423–425, 2008.
- [23] M. Moderhak, S. Kołacz, J. Jankau, and T. Juchniewicz, “Active dynamic thermography method for TRAM flap blood perfusion mapping in breast reconstruction,” *Quant. Infrared Thermogr. J.*, vol. 14, no. 2, pp. 234–249, 2017.

- [24] M. Moderhak, "Comparison of the exponential thermal transient parameterization methods with the SMTP method in the unipedicled DIEP flap computer modelling and simulation," *Quant. Infrared Thermogr. J.*, vol. 6733, pp. 1–12, 2018.
- [25] A. Saxena, V. Raman, and E. Y. K. Ng, "Single image reconstruction in active dynamic thermography: A novel approach," *Infrared Phys. Technol.*, vol. 93, no. Sept., pp. 53–58, 2018.
- [26] A. Saxena, E. Y. K. Ng, and V. Raman, "Thermographic venous blood flow characterization with external cooling stimulation," *Infrared Phys. Technol.*, vol. 90, pp. 8–19, 2018.
- [27] C. Ibarra-Castanedo and X. Maldague, "Pulsed phase thermography reviewed," *Quant. Infrared Thermogr. J.*, vol. 1, no. 1, pp. 47–70, 2004.
- [28] R. W. Lewis, P. Nithiarasu, and K. N. Seetharamu, *Fundamentals of the Finite Element Method for Heat and Fluid Flow*, vol. 3. John Wiley & Sons, Ltd, 2004.
- [29] G. R. North, "Lessons from energy balance models," in *Physically-Based Modelling and Simulation of Climate and Climatic Change-Part II*, vol. 1, M. E. Schlesinger, Ed. Kluwer Academic Publishers, 1988, pp. 627–651.
- [30] H. Meffert, N. Sonnichsen, and B. Meffert, "Skin-rewarming curves," *Lancet*, vol. 7780, pp. 769–770, 1972.
- [31] A. Merla, L. Di Donato, S. Di Luzio, and G. L. Romani, "Quantifying the relevance and stage of disease with the tau image technique," *IEEE Eng. Med. Biol. Mag.*, vol. 21, no. 6, pp. 86–91, 2002.
- [32] E. Peli, "Contrast in complex images," *J. Opt. Soc. Am. A*, vol. 7, no. 10, p. 2032, 1990.

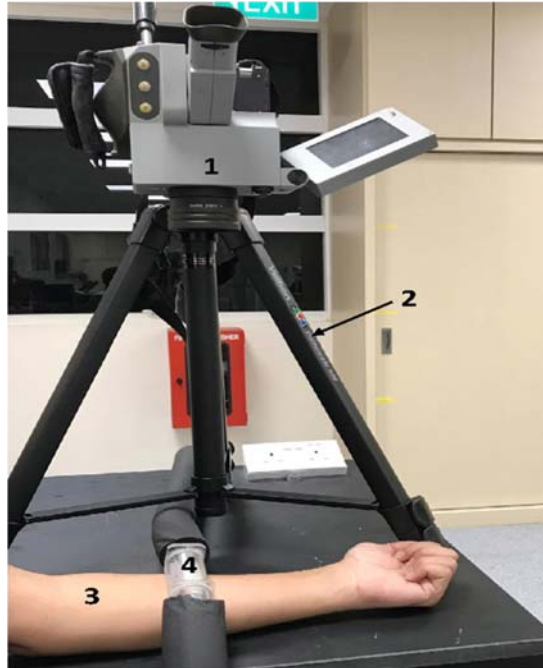


Figure 1: Experimental setup: 1) Infrared thermal camera 2) Tripod to fix the camera 3) Human subject forearm 4) Cooling pad to apply the external stimulation

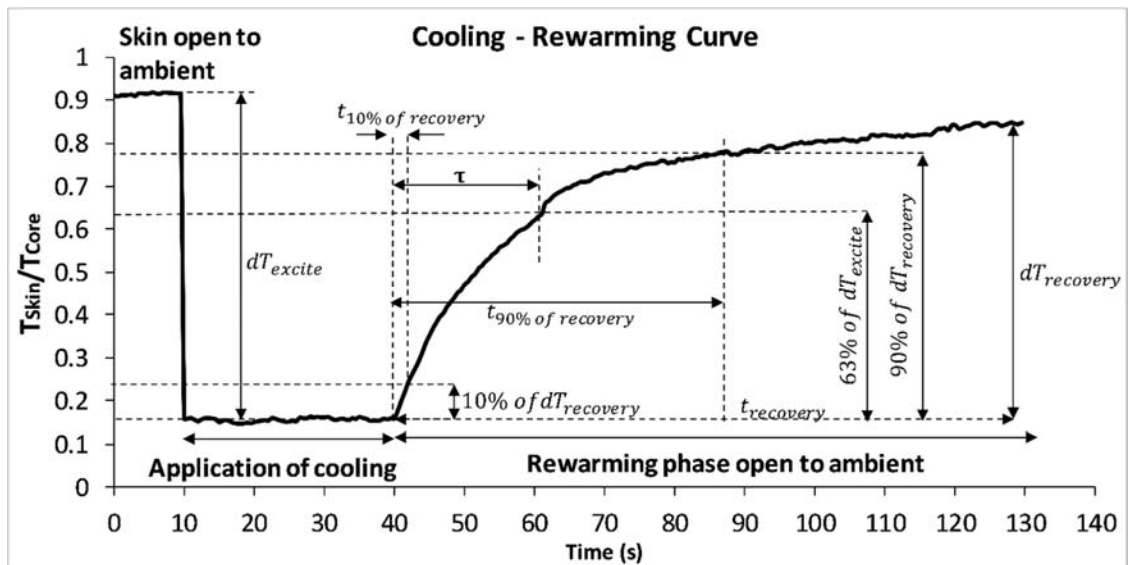


Figure 2: Cooling-rewarming curve in active dynamic thermography depicting curve variables ($T_{\text{core}}=37^{\circ}\text{C}$)

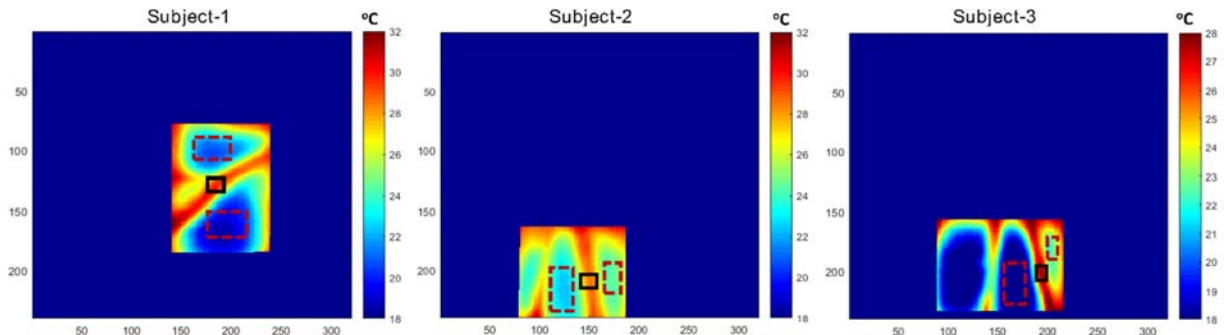


Figure 3: Reference location I and I_b selection in the image: black box is over the blood vessel (I) and red-dashed box is outside the vessel (I_b)

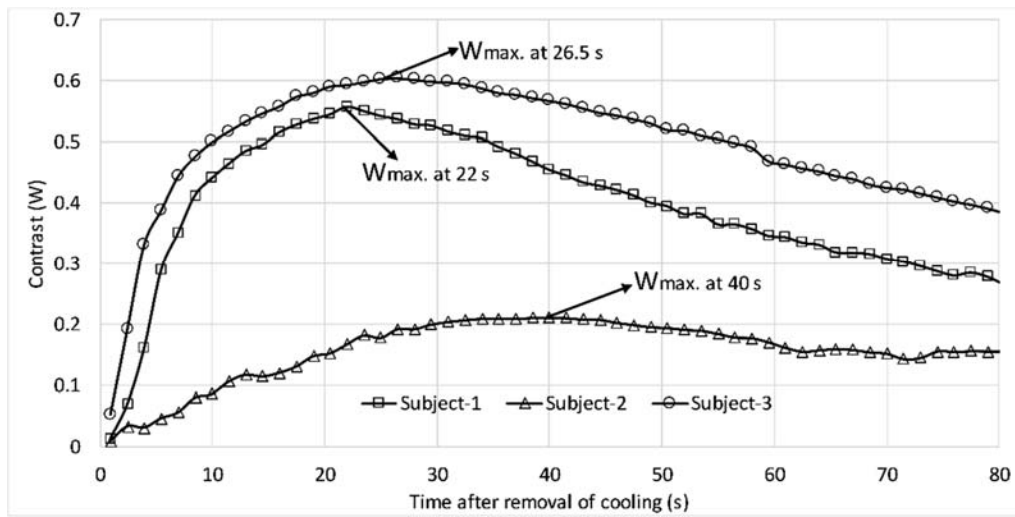


Figure 4: Contrast versus time after removal of cooling for the original sequence of thermal images

Table 1: External cooling stimulation details

Subject Studied	Average temperature within the cooling stimulation zone ($^{\circ}\text{C}$) \pm SD		
	Before cooling ($t=5\text{s}$)	After cooling ($t=40\text{s}$)	End of rewarming ($t=130\text{s}$)
Subject-1	33.53 ± 0.12	8.53 ± 3.5	27.27 ± 1.87
Subject-2	33.52 ± 0.19	9.53 ± 1.98	28.55 ± 1.68
Subject-3	32.82 ± 0.33	9.42 ± 1.03	25.09 ± 2.90

Table 2: Best contrast image in ADT sequence

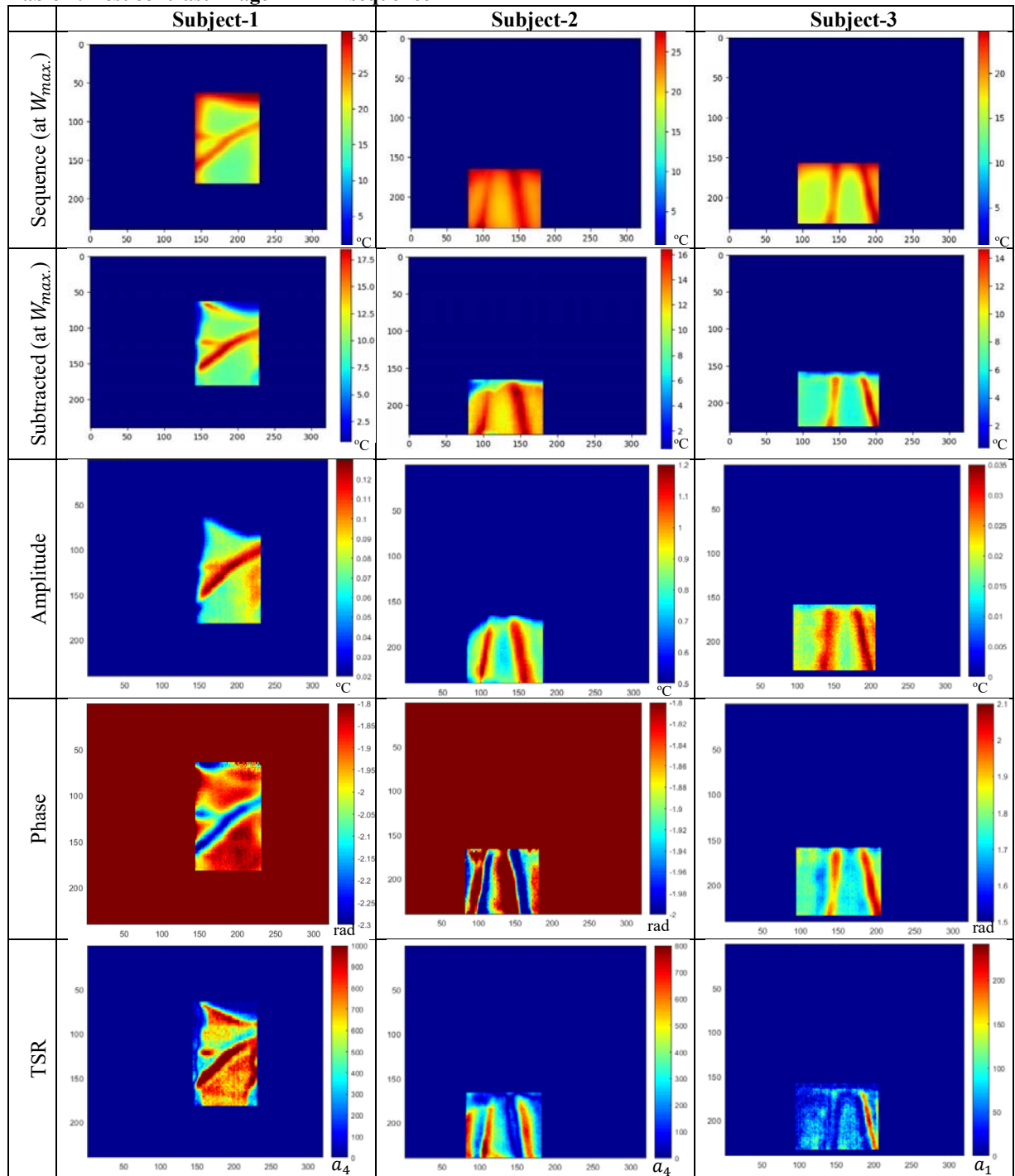


Table 3: Contrast (W) value of first derivative TSR coefficient images at $m = 4, 5, \text{ and } 6$

Coefficient Image	Image Contrast (W) in TSR images								
	Subject 1			Subject 2			Subject 3		
	$m = 4$	$m = 5$	$m = 6$	$m = 4$	$m = 5$	$m = 6$	$m = 4$	$m = 5$	$m = 6$
a_1	0.817	0.079	0.061	1.734	1.613	0.088	1.140	1.904	1.036
a_2	0.830	0.091	0.061	1.920	1.558	0.433	1.146	1.885	1.042
a_3	0.842	0.105	0.062	2.240	1.498	1.899	1.152	1.866	1.048
a_4	0.852	0.119	0.063	2.844	1.432	0.165	1.158	1.848	1.053
a_5	NA	0.135	0.064	NA	1.358	0.751	NA	1.830	1.059
a_6	NA	NA	0.066	NA	NA	0.519	NA	NA	1.065

Table 4: Image output from different single image reconstruction methods in ADT

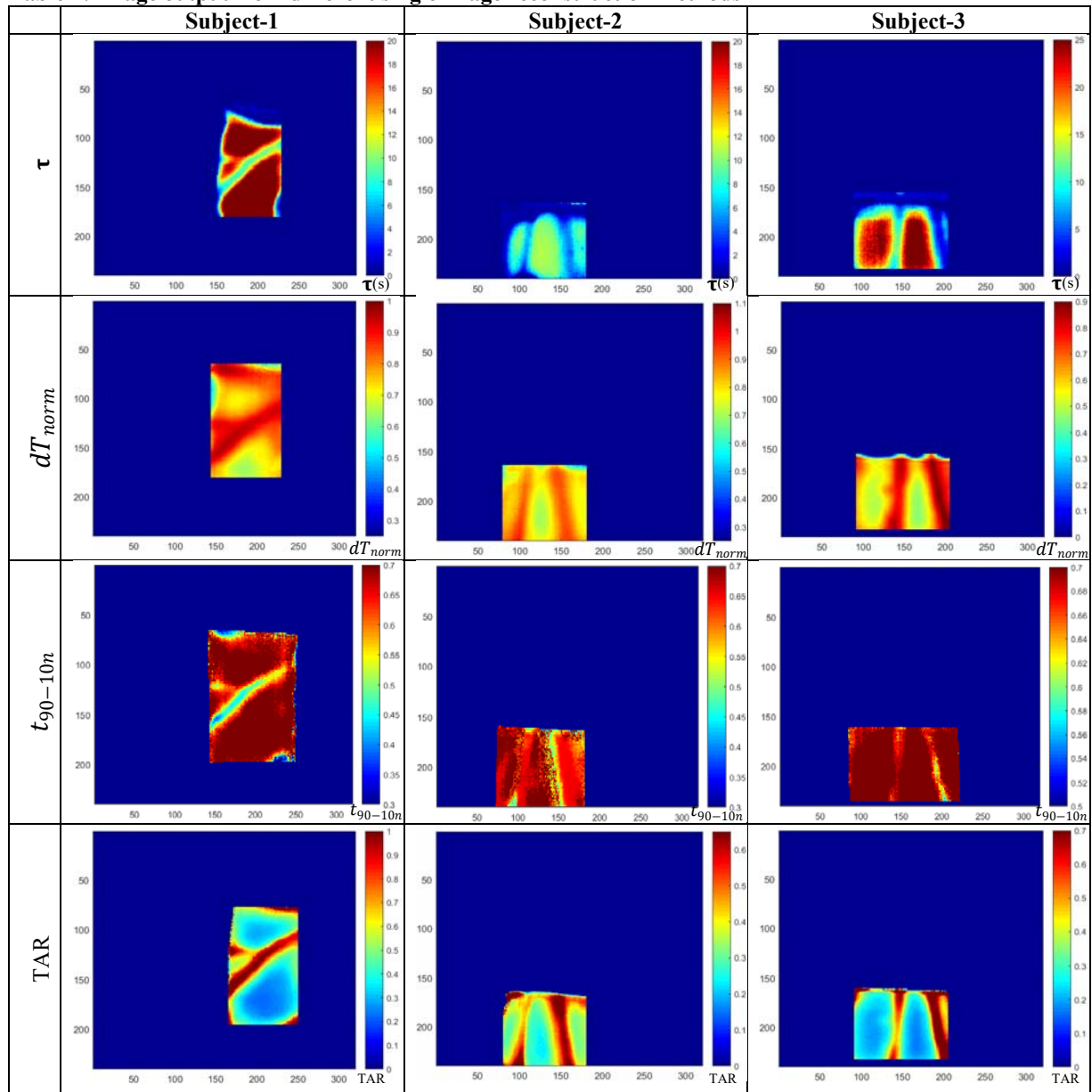


Table 5: Contrast and image parameter value over the skin tissue with (target, I) and without (background, I_b) the superficial vein in the reconstructed image from different image reconstruction methods

Image Reconstruction Methods		Contrast (W)		
		Subject 1	Subject 2	Subject 3
Sequence Image (W_{max})		0.56 $I=23.87$ °C, $I_b=15.30$ °C	0.21 $I=26.34$ °C, $I_b=21.79$ °C	0.61 $I=23.83$ °C, $I_b=14.80$ °C
Subtracted Image (at sequence W_{max})		0.85 $I=18.14$ °C, $I_b=9.80$ °C	0.37 $I=16.36$ °C, $I_b=11.96$ °C	0.78 $I=11.58$ °C, $I_b=6.50$ °C
Fourier Transform	Amplitude	0.53 $I=0.12$ °C, $I_b=0.07$ °C	0.35 $I=1.11$ °C, $I_b=0.82$ °C	0.50 $I=0.03$ °C, $I_b=0.02$ °C
	Phase	0.20* $I'=1.46$, $I'_b=1.22$	0.04* $I'=1.22$, $I'_b=1.17$	0.18 $I=2.04$ rad, $I_b=1.72$ rad
Thermographic Signal Reconstruction (TSR)		0.85 $I=1230$, $I_b=664$	2.84 $I=615$, $I_b=160$	1.90 $I=205$, $I_b=70.70$
Parametric Image Reconstruction	τ	1.78* $I'=3.99$, $I'_b=1.43$	0.30* $I'=1.87$, $I'_b=1.44$	1.24* $I'=3.09$, $I'_b=1.38$
	dT_{norm}	0.30 $I=0.94$, $I_b=0.73$	0.20 $I=0.91$, $I_b=0.76$	0.54 $I=0.81$, $I_b=0.53$
	t_{90-10n}	0.60* $I' = 1.93$, $I'_b=1.21$	0.11* $I'=1.41$, $I'_b=1.28$	0.14* $I'=1.26$, $I'_b=1.10$
	Tissue Activity Ratio (TAR)	2.37 $I=1.01$, $I_b=0.30$	1.03 $I=0.61$, $I_b=0.30$	1.47 $I=0.52$, $I_b=0.21$
<p>I: Image parameter over the skin tissue with superficial vein (target) I_b: Image parameter over the skin tissue without superficial vein (background) *Contrast (W) is calculated after the transformation of the images (I' and I'_b of I and I_b, respectively) using Equation 13 (I' and I'_b are dimensionless)</p>				