

Research Article

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Sulfoquinovosylmonoacylglycerols regulating intestinal inflammation in co-culture system from the brown alga *Turbinaria ornata*

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The inflammatory bowel diseases (IBD) including ulcerative colitis and Crohn disease are characterized by chronic inflammation throughout the gastrointestinal tract. The prevalence of IBD has been increasing worldwide, and has sometimes led to irreversible impairment of gastrointestinal structure and functions. In the present study, we identified a new sulfoquinovosylmonoacylglycerols (SQMG) (1) together with two known SQMGs (2 and 3) regulating intestinal inflammation from the brown alga *Turbinaria ornata*. The anti-inflammatory properties of two bioactive SQMGs, 1 and 2 were evaluated using an *in vitro* co-culture system consisting of human epithelial Caco-2 cells and PMA (phorbol 12-myristate 12-acetate)-differentiated THP-1 macrophages. Treatment with 1 or 2 inhibited the production nitric oxide and prostaglandin E₂ induced by lipopolysaccharide and interferon γ challenge. The expressions of inducible nitric oxide synthase and cyclooxygenase 2 were markedly down-regulated in response to inhibition of nuclear factor κ B translocation to nucleus. These findings suggest the potential use of the brown alga *T. ornata* and its biologically active metabolites SQMGs as pharmaceutical adjuvants in the treatment of inflammation-related diseases, including IBD.

Key Words: co-culture; inflammation; inflammatory bowel disease; sulfoquinovosylmonoacylglycerol; *Turbinaria ornata*

INTRODUCTION

Inflammatory bowel disease (IBD) is a chronic inflammatory disease, including ulcerative colitis (UC) and Crohn disease (CD). Characteristic chronic inflammation of the gastrointestinal tract observed in IBD sometimes causes irreversible damage to the gastrointestinal structure and functions. The two main types of IBD, UC and CD, have quite different clinical and pathological features. Clinical features of CD mainly include abdomi-

nal pain, diarrhea, narrowing of the intestinal lumen, and pathological features lead to intestinal obstruction, including scleritis, granulomas, and fistula (Loftus and Sandborn 2002). On the other hand, UC is characterized by lower abdominal pain, diarrhea, rectal bleeding, frequent and bloody feces (Bouma and Strober 2003). Intestinal homeostasis disorders in IBD are due to inadequate and exaggerated mucosal immune responses to the nor-



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mal components of mucosal microflora, as evidenced by numerous studies involving clinical patients and credited animal models (Bouma and Strober 2003). During the onset and progression of IBD, the mucosal epithelial barrier is damaged, and the intestinal flora causes and induces an abnormal intestinal immune response with a subsequent inflammation (Abraham and Cho 2009).

Marine algae are increasingly targeted for their bioactive metabolites that have the potential to be used in future medicine (Blunt et al. 2013, Wang et al. 2017, Alves et al. 2018, Fernando et al. 2019). The organisms produce extensive bioactive compounds including polyphenols, polysaccharides, sterols, fatty acids, peptides, pigments, and halogenated compounds, which exhibit antioxidant, anti-inflammatory, anticancer, antidiabetic, tyrosinase inhibitory and ultraviolet-protective effects (Faulker 2001, Cardozo et al. 2007). In a recent search for anti-inflammatory secondary metabolites that regulate intestinal inflammation in marine algae, the extract of brown alga *Turbinaria ornata* (Turner) J. Agardh showed attractive activity. To evaluate intestinal anti-inflammatory activity, an advanced *in vitro* co-culture system combining Caco-2 epithelial and THP-1 macrophage cells was established and applied. The differentiated Caco-2 cells are known to provide properties similar to human intestinal tissue, such as transport and permeability of substances (Rubas et al. 1993, Lennernäs et al. 1996). This co-culture system provides characteristic and more representative human intestine-like model (Kämpfer et al. 2017).

T. ornata is known as a kind of brown algae which is distributed throughout the world. It has been reported that a cytotoxic secosqualene carboxylic acid and turbinaric acid was isolated from *T. ornata* collected in southern area of Japan and cytotoxic sterols were identified from the organism collected in the southernmost tip of Taiwan (Asari et al. 1989, Sheu et al. 1997). Besides cytotoxic constituents, the extract of *T. ornata* showed potent antioxidant and antibacterial activity which contributed from phenolic compounds (Zubia et al. 2008, Vijayabaskar and Shiyamala 2012). Recently, anti-inflammatory activity *T. ornata* and its active metabolite fucoidan has been reported (Subash et al. 2016). However, the possibility of *T. ornata* in regulating intestinal inflammation has not been reported, and studies of the metabolites of *T. ornata* remains limited. Hence, in the present study, we evaluated anti-inflammatory potential of *T. ornata* in the treatment of IBD, and attempted to identify bioactive constituents using bioactivity-guided isolation experiments.

MATERIALS AND METHODS

General experimental procedures

Optical rotations were measured on a Jasco P-1010 polarimeter (Jasco Co., Ltd, Tokyo, Japan) with a 5 cm-length cell. IR spectrum was recorded on a Jasco FT/IR 4100 spectrometer. Nuclear magnetic resonance (NMR) spectra were measured on a Varian VNMR5 500 MHz spectrometer (Varian Technologies, Santa Clara, CA, USA) in CD₃OD. High resolution electrospray ionization mass spectra were obtained on an SCIEX X500R instrument and tandem mass spectrometry (MS/MS) data was measured on an SCIEX QTRAP 3200 instrument (Applied Biosystems/Sciex, Foster City, CA, USA). High-performance liquid chromatography (HPLC) experiments were performed on an Agilent 1200 system (Agilent, Waldbronn, Germany) consisting of a binary pump (G1312B), autosampler (G1329B), column oven (G1316B), and 1260 infinity evaporative light scattering detector (ELSD). The LPS (*Escherichia coli* 0127:B8), dimethyl sulfoxide, PMA (phorbol 12-myristate 12-acetate), and monoclonal anti-β-actin antibody were purchased from Sigma-Aldrich (St. Louis, MO, USA). MEM medium, RPMI 1640 medium, fetal bovine serum (FBS), penicillin / streptomycin, and phosphate-buffered saline (PBS) were purchased from Gibco Life Technologies (Grand Island, NY, USA). The DAF-DA assay was purchased from Sigma-Aldrich. Tin protoporphyrin IX (SnPP) was purchased from Porphyrin Products (Logan, UT, USA). Primary antibodies, including inducible nitric oxide synthase (iNOS), cyclooxygenase 2 (COX-2), p65, Lamin B, phospho-p65, IκBα, phospho-ERK, ERK, phospho-p38, p38, phospho-JNK, JNK, HO-1, and Nrf2 were purchased from Cell Signaling Technology, Inc. (Beverly, MA, USA). The secondary antibody was purchased from Jackson ImmunoResearch Laboratories, Inc. (West Grove, PA, USA).

Isolation of sulfoquinovosylmonoglycerols from the brown alga *Turbinaria ornata*

A specimen of the brown alga *T. ornata* was collected in Fulhadhoo, Maldives, in August 2017. The freeze-dried specimen (1.5 kg) was extracted with 90% aq. EtOH (2 L) twice at room temperature. The extract was concentrated *in vacuo* to give a semi-solid, which was partitioned into four fractions by solvent extraction: hexane, chloroform, ethyl acetate, and butanol. Among them, the butanol fraction (600 mg), obtained from the collaborator (Prof. You-Jin Jeon at Jeju University), was used. For the pur-

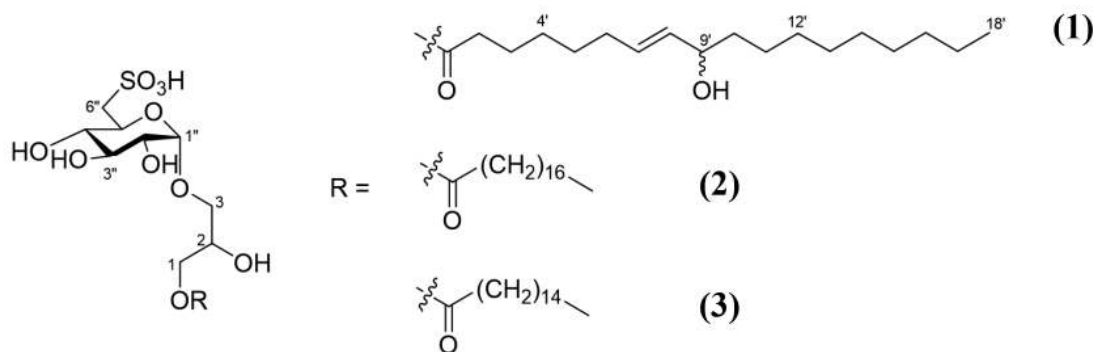


Fig. 1. Structure of compounds 1-3 isolated from *Turbinaria ornata*.

pose of removing any lipids, the fraction was first chromatographed on Sephadex LH-20 eluting 100% MeOH to yield four fractions: M1 (150 mg), M2 (250 mg), M3 (155 mg), and M4 (50 mg). Fractions M1 and M4 were evaluated as inactive effect on NO inhibition, but M2 and M3 showed good activity. Partial fraction of M2 (60 mg) showing the highest NO inhibition effect and interesting ¹H NMR signal was separated again by HPLC using a column Phenomenex C8 (250 × 4.6 mm, 5 μm), gradient solvent system as [A: H₂O with 0.1% formic acid, B: acetonitrile, from 30% B to 50% B for 10 min, then 50% B to 100% B for 10 min], a flow rate of 1.0 mL min⁻¹, and an ELSD detector to yield compounds 1 (2.1 mg), 2 (1.2 mg), and 3 (10.4 mg) (Fig. 1).

Compound 1: a colorless oil; [α]_D²⁵ = + 32.8 (c 0.1, MeOH); ¹H and ¹³C NMR see Table 1; high resolution electrospray ionization mass spectroscopy (HRESIMS) *m/z* 597.2946 [M-H]⁻ (calcd. *m/z* 597.2950 [M-H]⁻ for C₂₇H₅₀O₁₂S).

Cell culture

The human epithelial cell line (Caco-2) and human monocytic cell line (THP-1) were obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA). The Caco-2 cells were cultured in Eagle's minimum essential medium (EMEM; Gibco BRL, Grand Island, NY, USA) containing 20% (v/v) FBS and 1% (v/v) antibiotics (100 U mL⁻¹ penicillin and 100 μg mL⁻¹ streptomycin). THP-1 cells were cultured in RPMI1640 (Gibco BRL) containing 10% (v/v) FBS, 1% (v/v) antibiotics (100 U mL⁻¹ penicillin and 100 μg mL⁻¹ streptomycin) and 0.05 mM 2-mercaptoethanol. The cells were incubated in a humidified atmosphere of 95% air-5% CO₂. After that, THP-1 cells were treated with PMA for macrophage differentiation as below.

Differentiation of THP-1 monocyte to macrophage

THP-1 cells were differentiated in medium containing 50 nM of PMA for three days followed by incubated in fresh medium without PMA for additional two days (Daigneault et al. 2010).

In vitro co-culture model of intestine

For establishment of co-culture system that mimics human intestine, Caco-2 cells were seeded at 3.75 × 10⁵ cells per well on transwell inserts (0.4 μm pore size; Corning CoStar Corp., Cambridge, MA, USA) and maintained

Table 1. The NMR spectral data for compound 1 (500 MHz for ¹H, CD₃OD)

No.	δ _C ,mult	δ _H ,mult (J/Hz)
1	66.5, CH ₂	4.09, dd (13.5, 2.1); 4.19, dd (13.5, 6.1)
2	69.9, CH	4.09, m
3	70.5, CH ₂	3.39, dd (9.5, 3.9); 4.06, dd (9.5, 2.9)
1'	175.2, C	-
2'	34.9, CH ₂	2.36, t (7.6)
3'	25.97 / 25.95, CH ₂	1.61, m
4'	30.30 / 30.27, CH ₂	1.32, m
5'	30.73 / 30.70, CH ₂	1.38, m
6'	33.28 / 33.23, CH ₂	2.03, m
7'	132.46 / 132.40, CH	5.59, dt (15.4, 6.6)
8'	134.53 / 134.46, CH	5.39, dd (15.4, 7.1)
9'	73.79 / 73.75, CH ₂	3.94, dt (7.1, 6.9)
10'	38.47 / 38.45, CH ₂	1.41, m; 1.50, m
11'	26.65 / 26.62, CH ₂	1.31, m
12'-15'	29.9-30.5, CH ₂	1.28-1.31, m
16'	33.1, CH ₂	1.26, m
17'	27.3, CH ₂	1.29, m
18'	14.5, CH ₃	0.89, t (7.1)
1''	100.1, CH	4.77, d (3.7)
2''	73.7, CH	3.40, dd (9.3, 3.7)
3''	75.1, CH	3.64, t (9.3)
4''	74.9, CH	3.06, dd (9.5, 9.3)
5''	69.8, CH	4.09, m
6''	54.2, CH ₂	2.90, dd (14.2, 9.3); 3.34, dd (14.2, 2.0)

NMR, nuclear magnetic resonance.

in growth media EMEM for 14-20 days in an incubator at 37°C with a 5% CO₂ atmosphere. Culture medium was changed every 3 days until the cells were fully differentiated (TEER value > 1,200 Ω·cm²). THP-1 cells were independently seeded at 8.5 × 10⁶ cells into the bottom plate of 6-well transwell plate in growth media RPMI1640. Then, the insert that is polarized with Caco-2 monolayer was added. The upper chamber reflects the apical side and the lower chamber the basolateral side of the intestinal epithelium (Kleiveland 2015). For evaluation of anti-inflammatory activity of the test sample, 1 µg mL⁻¹ of LPS was added on the basolateral compartment of plate and then the test sample was added to the apical compartment of plate at the concentration indicated. After 24 h incubation, the contents of nitrite and inflammatory mediators produced within culture supernatant in the basolateral side were determined.

Measurement of TEER value

TEER value was measured once a week to test monolayer integrity using a Millicell-ERS (Millipore, Burlington, MA, USA). The TEER was calculated as following:

$$\text{TEER } (\Omega \cdot \text{cm}^2) = \text{Resistance} - \text{Blank resistance } (\Omega) \times \text{Membrane surface area } (\text{cm}^2)$$

Cell viability assay

Cell viability was measured in THP-1 cells using CCK-8 assay (Cell Counting Kit-8, #CK04; Dojindo Laboratory, Kumamoto, Japan), according to the manufacturer's instructions. Briefly, THP-1 cells were seeded at 1 × 10⁴ cells per well in 96-well plates and incubated for 24 h. Then, the cells were treated with test sample of various concentrations for 24 h. After incubation, 10 µL of CCK-8 solution was added to each well and incubated for 3 h at 37°C. The absorbance was measured using a microplate reader (Bio-Tek Instruments Inc., Winooski, VT, USA) at 450 nm. The experiments were performed in triplicate.

Measurement of NO and prostaglandin E₂ production

THP-1 cells were seeded at 1 × 10⁴ cells per well in 96-well plates and incubated for 24 h. The cells were treated with test sample for 1 h, followed by treated with LPS (10 µg mL⁻¹) and interferon-gamma (IFNγ; 10 ng mL⁻¹) for 24 h. The content of nitrite produced in culture medium was determined using DAF-DA assay. Briefly, cells were incubated with DAF-DA (3 µmol mL⁻¹) at 37°C for 20 min.

Then washed the cells and replaced with fresh medium incubated for an additional 20 min. The fluorescence was measured at 495 nm of excitation and 515 nm of emission wavelengths with a microplate reader (Bio-Tek Instruments Inc.). The concentration of prostaglandin E₂ (PGE₂) was quantified using an enzyme-linked immunosorbent assay (ELISA) kit (R&D System, Minneapolis, MN, USA) according to the manufacturer's instructions. The culture medium as applied in ELISA assay without dilution.

Western blot

Protein isolated from THP-1 cells incubated in co-culture system was applied to western blot assay. After incubation of cells with test sample and LPS as indicated, THP-1 cells were washed three times with ice-PBS and extracted with RIPA buffer containing a protease inhibitor cocktail (Santa Cruz Biotechnology, Santa Cruz, CA, USA) for 30 min on ice. Protein lysates were centrifuged at 13,000 ×g for 30 min at 4°C. Thirty micrograms of the lysed proteins quantified by Bradford assay were separated on sodium dodecyl sulfate polyacrylamide gel electrophoresis (8-12%) at 100 V and transferred to polyvinylidene difluoride membrane. The membranes were blocked with 5% nonfat milk in PBST buffer for 1 h at room temperature. The membranes were incubated with 1:1,000 diluted primary antibodies (Cell Signaling Technology, Inc., Danvers, MA, USA) at 4°C overnight. The membranes were washed three times with PBST buffer followed by incubated with 1:10,000 diluted secondary antibodies for 1 h at room temperature. Bands were visualized using ECL solution (Thermo Scientific, Rockford, IL, USA) and calibrated by the Chemidoc Imaging System (Bio-Rad, Hercules, CA, USA). The density value of the protein bands was normalized to Lamin B (nuclear) or β-actin (total protein or cytosol).

Statistical analysis

All data were through Shapiro-Wilk normality test and Levene's equality of variance test by SPSS ver. 17.0 program (SPSS Inc., Chicago, IL, USA). Then, the results were analyzed and converted to graph using Prism ver. 5.00 software (GraphPad Software, San Diego, CA, USA); one-way ANOVA were used for parametric data or Kruskal-Wallis for non-parametric data with Dunnett or Dunns *post hoc* tests, respectively. Statistical significance is indicated by p < 0.05. Data were expressed as the mean ± standard deviation of three independent experiments.

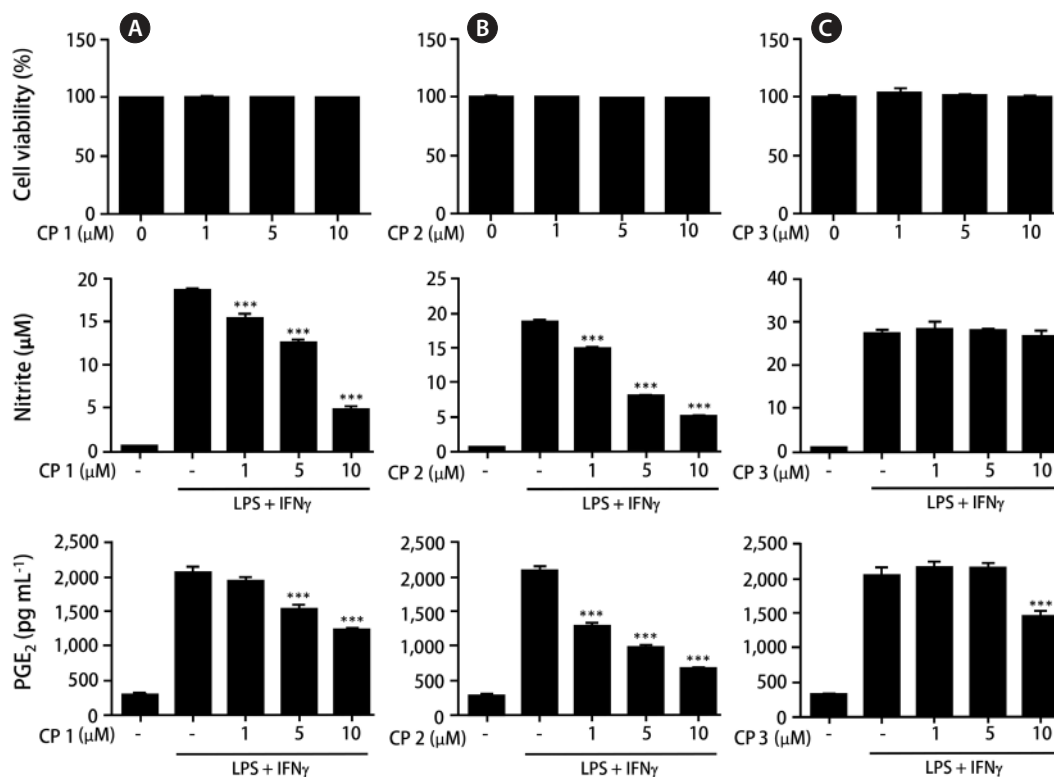


Fig. 2. Cytotoxicity and the effects of compounds 1 (A), 2 (B), and 3 (C) to inhibit the production of nitric oxide (NO) and prostaglandin E₂ (PGE₂) in THP-1 cells. For cytotoxicity assay, THP-1 macrophages differentiated by PMA (phorbol-12-myristate-13-acetate) were treated with various concentration of each compound (1, 5, and 10 µM) for 24 h. For measurement of NO and PGE₂ content, THP-1 macrophages were treated with each compound (1, 5, and 10 µM) followed by co-treated with lipopolysaccharide (LPS) and interferon γ (IFNγ) for 24 h. CP, compound. Results are presented as the mean ± standard deviation of triplicate experiments. ***p < 0.001 compared to LPS-treated cells.

RESULTS

Anti-inflammatory sulfoquinovosylmonoacylglycerols from the brown alga *Turbinaria ornata* using bioactivity-guided separation

Four fractions (M1-M4) of the butanolic layer of *T. ornata* extract separated by the Sephadex LH20 were applied to NO assay. Fraction M2 showed the most potent inhibitory activity on NO production induced by LPS. Hence, fraction M2 was further separated and purified using HPLC to yield several compounds including three sulfoquinovosylmonoacylglycerols (SQMGs). Compound 1 was obtained as a colorless oil and its molecular formula was determined as C₂₇H₅₀O₁₂S by the combination of negative HRESIMS and the ¹³C NMR spectrum. The ¹H NMR spectrum of 1 showed signals distributed in the range of 3.1–4.3 ppm, and a triplet at δ_H 2.36, a multiplet at δ_H 1.61, a broad singlet at δ_H 1.31 and a triplet peak at δ_H 0.89, indicating characteristic signals for a class of glycerolipids. An α-form glycopyranose was assigned by the

COSY correlations from the anomeric proton at δ_H 4.77 (*J* = 3.7 Hz). The coupling constant of each proton and the carbon chemical shifts at C-6'', along with the molecular formula, suggested a 6-deoxy-6-sulfo-α-*D*-glucopyranose moiety. The chemical shifts of the protons and carbons of the unit were consistent with the those of the known ones (Hirao et al. 2012). Further analysis of the COSY and TOCSY spectra led to assign the position of one olefinic group within the long carbon chain to be Δ7'. This was also corroborated by the heteronuclear multiple bond correlation (HMBC) correlations from H-2', H-3', H-5' to C-4' and from H-4', H-6', H-7' to C-5'. The COSY correlation from the olefinic protons (H-8') to the remaining oxymethine proton allowed the hydroxy group to be attached at C-9'. In addition, the position of the functional groups in the carbon chain was determined by MS/MS analysis of the carbon chain cleaved by hydrolysis of 1 with 0.1 N HCl (Supplementary Figs S1-S16). The geometry of the olefinic group was established as *E* by the large proton coupling constant (*J* = 15.4 Hz). The observation of carbons paired from C-3' to C-11' revealed

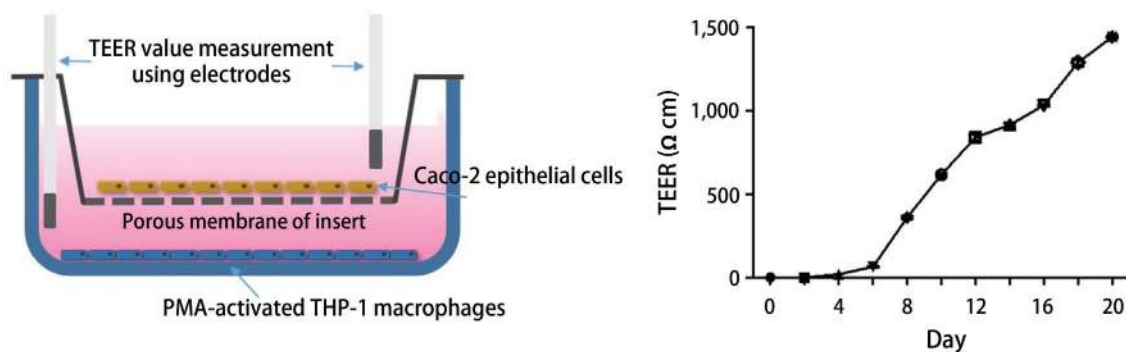


Fig. 3. Co-culture system consists of Caco-2 epithelial cells and PMA (phorbol-12-myristate-13-acetate)-differentiated THP-1 macrophage cells (left), and TEER values for 21 days (right).

that compound **1** is a mixture of two epimers consisting of 9(*R*) and 9(*S*). Finally, HMBC correlations from H-3 to C-1'' and from H-1 to the carbonyl carbon at C-1' established the planar structure of **1**. Thus, compound **1** was determined as 1-*O*-[(7*E*)-9-hydroxyoctadec-7-enoyl]-3-*O*-(6-sulfo- α -*D*-quinovopyranosyl)-*sn*-glycerol. Compared with **1**, compounds **2** and **3** did not display the downfield shifted proton signals corresponding to olefinic groups, but consisted of saturated acyl groups. From the analysis of NMR and HRESIMS, compounds **2** and **3** were identified as 1-stearoyl-3-*O*-(6-sulfo- α -*D*-quinovopyranosyl)-*sn*-glycerol and 1-palmitoyl-3-*O*-(6-sulfo- α -*D*-quinovopyranosyl)-*sn*-glycerol, respectively (Sakamoto et al. 2000, Hanashima et al. 2001).

Anti-inflammatory activities of the isolated compounds **1-3** were screened using THP-1 macrophage cells activated by LPS and IFN γ treatment. Prior to evaluating anti-inflammatory activities of **1-3**, the cytotoxicities of the compounds were measured using CCK-8 assay. All compounds showed no cytotoxicity at the concentration range up to 10 μ M (cell viability > 98% of control). To evaluate the inhibitory activity of **1-3** on NO production, THP-1 cells were treated with each compound (1, 5, and 10 μ M) for 1 h and then challenged with LPS (1 μ g mL⁻¹) and IFN γ (10 ng mL⁻¹). After 24 h of incubation, the content of NO that is released into culture medium was detected using DAF-DA. Among the compounds tested, **1** and **2** showed the significant inhibition on NO production induced by LPS and IFN γ , while **3** was inactive. Inhibitory activities of **1-3** on PGE₂ production were further investigated (Fig. 2). As a result, the pretreatment of cells with **1** or **2** for 24 h led to a significant decrease of LPS and IFN γ -induced PGE₂ production in a concentration-dependent manner, whereas a mild decrease by **3** was observed at the concentration of 10 μ M.

The effects of **1** and **2** on the expression of pro-inflammatory proteins and the phosphorylation of mitogen-activated protein kinase family in intestinal co-culture system

Recently, the advanced and novel co-culture model of Caco-2 and PMA-differentiated THP-1 cells that resembles human intestine has been established (Kämpfer et al. 2017). This *in vitro* model is featured by representing numerous hallmarks of intestinal inflammation. During developing co-culture model, TEER value is extensively used to measure the resistance of tight junctions of cell monolayer. The TEER was measured using an Ohmmeter to assess the tightness of the Caco-2 cell layer on the apical side of the insert as well as the barrier integrity throughout co-culture with THP-1 cells. In our co-culture system, TEER of Caco-2 layer subsequently increased at 4 days after plating and reached the maximum value at day 21 (Fig. 3). In this co-culture system, the effects of **1** and **2** on the expression of iNOS and COX-2 as well as the phosphorylation of mitogen-activated protein kinase (MAPK) family was measured to evaluate the action mechanism of the compounds regulating intestinal inflammation. As shown in Fig. 4, western blot analysis showed that the expression levels of iNOS and COX-2 were upregulated by co-treatment of LPS and IFN γ , whereas the pretreatment of cells with **1** or **2** (1, 5, and 10 μ M) significantly attenuated this upregulation. In addition, the induced phosphorylation of p38, ERK1/2, and JNK induced by LPS and IFN γ was attenuated by pretreatment of **1** or **2**. At the concentration of 1 μ M, the significant decrease on phospho-ERK1/2 and phospho-JNK by **1** and **2**, respectively, were observed (Fig. 5).

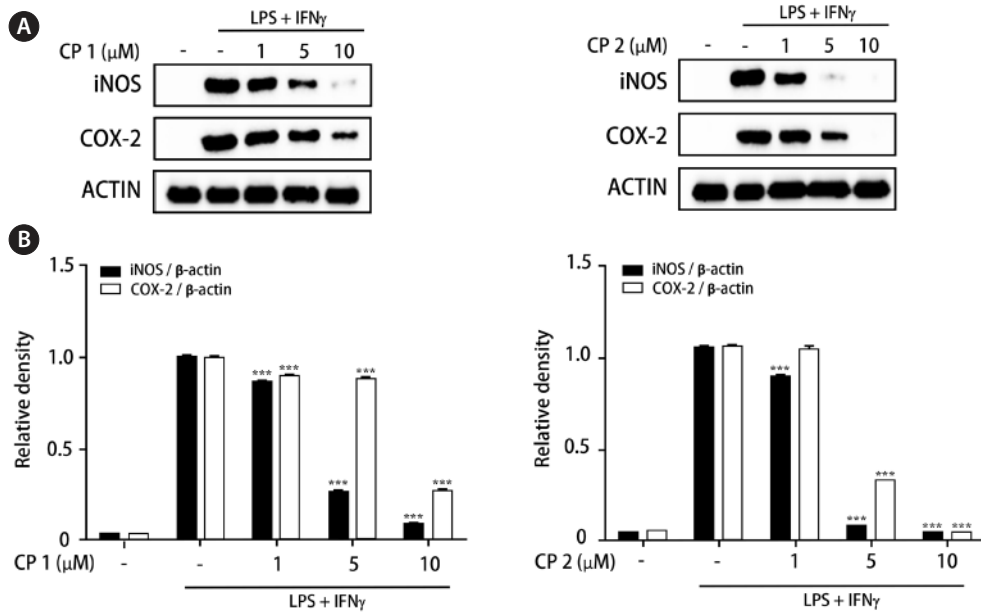


Fig. 4. Compounds 1 and 2 down-regulated the expression of inducible nitric oxide synthase (iNOS) and cyclooxygenase 2 (COX-2) in intestinal co-culture system. Compounds 1 or 2 (1, 5, and 10 μM) were individually treated into the apical compartment of the Caco-2/THP-1 co-culture system. After 1 h, lipopolysaccharide (LPS; 1 $\mu\text{g mL}^{-1}$) and interferon γ (IFN γ ; 10 ng mL^{-1}) were co-treated to the basolateral compartment. The expression levels of iNOS and COX-2 in cells were analyzed by western blot after 24 h of LPS + IFN γ challenge. (A) The representative blots. (B) The calculated intensities after normalization to β -actin. CP, compound. Results are presented as the mean \pm standard deviation of triplicate experiments. *** $p < 0.001$ compared to LPS-treated cells.

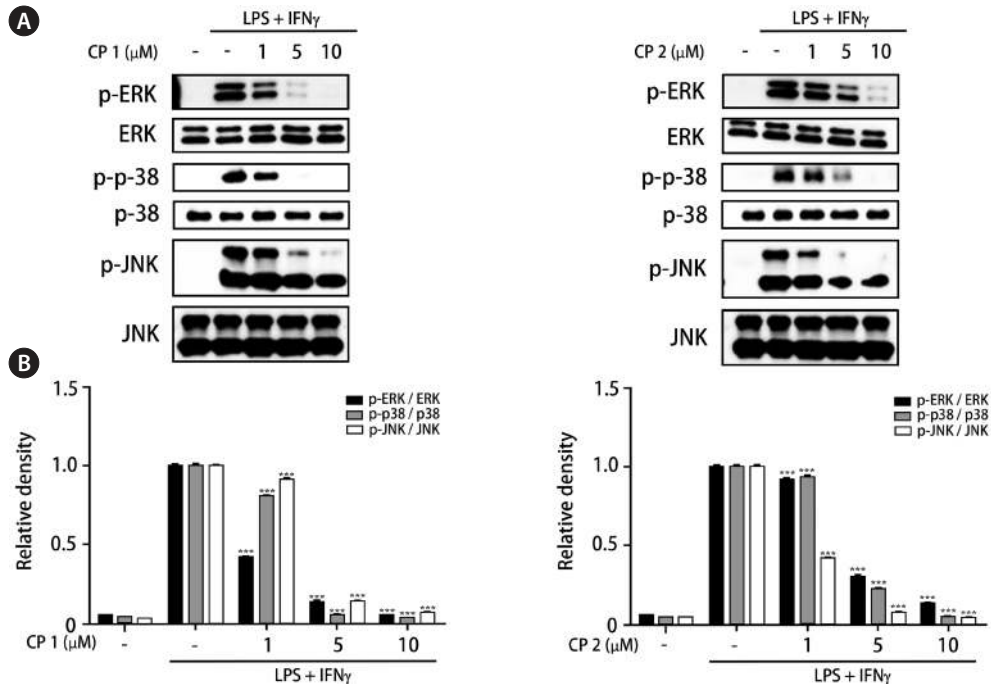


Fig. 5. Compounds 1 and 2 attenuated the phosphorylation of mitogen-activated protein kinase (MAPK) family in intestinal co-culture system. Compounds 1 or 2 (1, 5, and 10 μM) were individually treated into the apical compartment of the Caco-2/THP-1 co-culture system. After 1 h, lipopolysaccharide (LPS; 1 $\mu\text{g mL}^{-1}$) and interferon γ (IFN γ ; 10 ng mL^{-1}) were co-treated to the basolateral compartment. The phosphorylation levels of MAPKs (ERK, p38, and JNK) were analyzed by western blot after 15-30 min of LPS + IFN γ challenge. (A) The representative blots. (B) The calculated intensities after normalization to β -actin. CP, compound. Results are presented as the mean \pm standard deviation of triplicate experiments. *** $p < 0.001$ compared to LPS-treated cells.

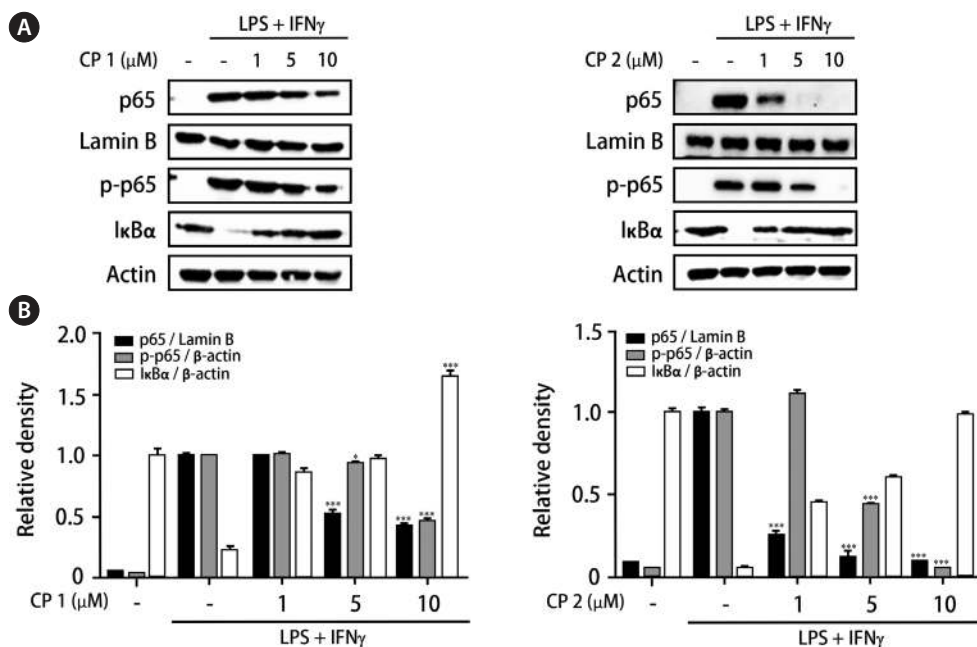


Fig. 6. Compounds 1 and 2 inhibited NF- κ B translocation into the nucleus in intestinal co-culture system. Compounds 1 or 2 (1, 5, and 10 μ M) were individually added into the apical compartment of the Caco-2/THP-1 co-culture system. After 1 h, lipopolysaccharide (LPS; 1 μ g mL⁻¹) and interferon γ (IFN γ ; 10 ng mL⁻¹) were co-treated to the basolateral compartment and the cells were incubated overnight. Cell extracts were biochemically separated into nuclear and cytoplasmic fractions, and the expression of p65, phosphorylated-p65, Lamin B, and I κ B α in cells was analyzed by western blot. (A) The representative blots. (B) The calculated intensities after normalization to Lamin B (nucleus) or β -actin (cytosol and whole cell). CP, compound. Results are presented as the mean \pm standard deviation of triplicate experiments. * p < 0.05 and *** p < 0.001 compared to LPS-treated cells.

The effects of 1 and 2 on the nucleus translocation of nuclear factor- κ B and its inhibitor, I κ B- α in intestinal co-culture system

It has been known that interaction of p65 unit of nuclear factor- κ B (NF- κ B) with specific gene sets promotes subsequent transcription and expression of inflammatory mediators in macrophages (Aggarwal et al. 2004, Sanjeeva et al. 2019). Based on the inhibitory activities of 1 and 2 on the expressions of pro-inflammatory proteins and MAPK family, the effects of these compounds regulating p65 localization in nucleus and cytoplasm have been observed. After treatment of cells with compounds followed by LPS and IFN γ challenge, whole cell lysates were biochemically fractionated into nuclear and cytoplasmic extracts to measure localization of p65 and phosphorylated p65. As shown in Fig. 6, the treatment of cells with 1 or 2 (1, 5, and 10 μ M) markedly inhibited the phosphorylation of p65 in cytoplasm and weakened the translocation of p65 into nucleus. Significantly upregulated total levels of I κ B- α protein by 1 and 2 in the cytoplasm may explain the enhanced binding of I κ B- α to p65, which indicates that NF- κ B access to the nucleus binds

to the promoter can be prevented. The increase of I κ B- α level in cytoplasm was more significant by 1 at the concentration of 10 μ M. These results suggest that interfering with the p65 subunit of NF- κ B and a specific set of target genes by 1 or 2 results in subsequent down-regulation of transcription and expression of inflammatory mediators (Oeckinghaus and Ghosh 2009, Saegusa et al. 2010).

DISCUSSION

T. ornata is a brown alga common in the world. This seaweed is widely consumed as a functional food by Asians due to its antioxidant properties and is also used as a folk medicine for various diseases. *T. ornata* has been reported to exhibit a broad range of biological activities, including antibacterial, anticoagulant, antiviral, antitumor, hepatoprotective activity, and treatment for parasitic infections (Vijayabaskar and Shiyamala 2011, Chia et al. 2015, Abdella et al. 2016, Canoy and Bitacura 2018). Regarding anti-inflammatory properties of *T. ornata*, Subash et al. (2016) reported an anti-inflammatory effect with the bioactive components of *T. ornata*. The granu-

loma weight was reduced and biochemical and inflammatory markers were improved by aqueous extract of *T. ornata* in cotton pellet induced granuloma model in rats. Using gas chromatography–mass spectrometry (GC-MS), fucoidan was characterized as an active constituent in *T. ornata*. Fucoidan referred to the sulfated polysaccharides containing fucose that is contained in cell wall of brown algae. Numerous literatures demonstrated that fucoidan exhibits potential anti-inflammatory activity *in vitro* and *in vivo*. Fucoidan extracted from *T. ornata* from the Maldives inhibited expressions of inflammatory cytokines and protein in RAW264.7 macrophages. Also, the level of reactive oxygen species and NO was reduced by fucoidan treatment in zebrafish embryo model (Jayawardena et al. 2019). Anti-inflammatory action of the sulfated polysaccharide of *T. ornata* was further investigated in Complete Freund's Adjuvant induced arthritis in rats. Ananthi et al. (2017) revealed that the sulfated polysaccharide purified from *T. ornata* inhibited inflammation and bone damage at very low dose itself that was evidenced by the significant reduction in paw volume and arthritic score. Besides the sulfated polysaccharides, a phytochemical analysis to identify the biologically active constituents in *T. ornata* deduced the presence of saponin, alkaloids, amino acids, fixed oil, fat and phenolic compounds by GC-MS and Fourier transform infrared spectroscopy experiments (Deepak et al. 2017). However, characterization of the metabolites in *T. ornata* based on the full assignment by NMR experiments are limited. The aim of this study was to identify anti-inflammatory metabolites of *T. ornata* through isolation and structure elucidation using 1D, 2D-NMR, and MS analysis. Also, we attempted to reveal anti-inflammatory potential for the isolated compounds regulating intestinal inflammation using *in vitro* co-culture system.

Through activity-guided fractionation and isolation, three SQMGs including one new compound were successfully isolated and identified. A new compound **1**, which possesses one hydroxy group and one double bond on the carbon chain, was obtained as a mixture of the two epimers. Compound **3** was isolated in large quantities compared to **1** and **2**, but compounds **1** and **2** showed the significant inhibition against inflammatory mediators in THP-1 macrophage cells. Interestingly, it was found that **1** and **2** with carbon chains of equal length (C18) are active while **3** with C16 chain are inactive. For evaluation of anti-inflammatory property of bioactive compounds, **1** and **2** were further investigated in co-culture system established using human-originated epithelial-like Caco-2 and THP-1 macrophage cells.

Abnormal and persistent inflammatory responses to commensal microbes in genetically sensitive hosts are thought to be the main cause of IBD. In IBD patients, immune cells have been found to release abnormally high levels of pro-inflammatory cytokines compared to normal tissues (MacDermott 1999). In addition, markedly enhanced secretion of these downstream pre-inflammatory cytokines such as tumor necrosis factor α (TNF- α), interleukin (IL)-1 β , and IL-6 is observed in patients with UC and CD. Overexpressed TNF- α , IL-1 β , iNOS, and COX-2 are involved in the development of colitis (Ogata and Hibi 2003, Pettus et al. 2003, Pan et al. 2008). Correspondingly, agents that block the overproduction of these cytokines are proposed to be effective in treating IBD. The first choice of drugs for patients with mild to moderate IBD is anti-inflammatory drugs such as amino salicylate and corticosteroids. Amino salicylates are known to be effective in calming inflammation in the digestive track. However, the side effects including headache, nausea, and diarrhea in high dose limit the use of this medication. In THP-1 macrophage cells, treatment of cells with **1** or **2** inhibited the production of NO and PGE₂ stimulated by LPS and IFN γ . Also, in co-culture system established herein, the expressions of pro-inflammatory proteins, iNOS and COX-2 were markedly attenuated by **1** or **2** in response to inhibition on MAPKs (p38, ERK1/2, and JNK) phosphorylation.

NF- κ B is an essential and ubiquitous transcription factor that controls immune and inflammatory signal transduction by regulating the expression of related genes (Lappas et al. 2002, Aggarwal et al. 2004). Since NF- κ B binds to the proximal promoters of the COX-2 and TNF- α genes, the inhibition of DNA-binding activity of NF- κ B corresponds to the inhibition of the expression of inflammatory mediators, iNOS, COX-2, and TNF- α (Economou et al. 1989). In inactivated cells, NF- κ B complexes containing subunits p50 and p65 are located in the cytoplasm combined with I κ B- α . When cells are activated, I κ B- α is phosphorylated by the I κ B kinase complex and dissociates from NF- κ B to allow translocation of NF- κ B to the nucleus. Translocated NF- κ B promotes subsequent transcription and expression of inflammatory mediators (Oeckinghaus and Ghosh 2009, Saegusa et al. 2010). Based on the inhibitory effects of **1** and **2** on the expression of inflammatory cytokines and proteins, the action mechanisms of these compounds involved in NF- κ B localization in nucleus and cytoplasm of THP-1 macrophages were observed. In THP-1 macrophages co-cultured with Caco-2 cells, the treatment of **1** and **2** attenuated the phosphorylation of p65 and upregulated

the expression level of I κ B- α in cytoplasm, leading to the enhanced interactions of p65 subunit of NF- κ B with I κ B- α . The enhanced binding of I κ B- α with p65 results in cytoplasmic retention of NF- κ B complex, thereby preventing the access of NF- κ B to the nucleus for binding the target gene promoter. Above results suggest that the two SQMGs isolated from *T. ornata* inhibits the expression of pro-inflammatory mediators via regulation of NF- κ B pathway.

A new SQMG together with two known SQMGs were isolated from the brown alga *T. ornata*. The anti-inflammatory potential of the isolated compounds was evaluated in an *in vitro* co-culture system combining epithelial Caco-2 cells and THP-1 macrophages similar to the intestinal environment. The two SQMGs isolated from *T. ornata* showed potent anti-inflammatory activity by down-regulating the expression of iNOS and COX-2 and attenuating nuclear translocation of NF- κ B. Given that NF- κ B is considered a master switch that regulates inflammation in macrophages, the present study supports the potential use of *T. ornata* and its bioactive metabolites as therapeutics for IBD.

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SUPPLEMENTARY MATERIALS

Supplementary Fig. S1. High-performance liquid chromatography (HPLC) chromatogram for the separation of M2 fraction (<https://e-algae.org>).

Supplementary Fig. S2. (A) High resolution negative mass spectrometry (MS) and tandem mass spectrometry (MS/MS) spectra. (B) Optical rotation value for 1 (<https://e-algae.org>).

Supplementary Fig. S3. Infrared spectrum of 1 (<https://e-algae.org>).

Supplementary Fig. S4. ¹H NMR spectrum of 1 (<https://e-algae.org>).

Supplementary Fig. S5. ¹³C NMR spectrum of 1 (<https://e-algae.org>).

Supplementary Fig. S6. COSY NMR spectrum of 1 (<https://e-algae.org>).

Supplementary Fig. S7. TOCSY NMR spectrum of 1 (<https://e-algae.org>).

Supplementary Fig. S8. HSQC NMR spectrum of 1 (blue cross peaks, CH or CH₃; black cross peaks, CH₂) (<https://e-algae.org>).

Supplementary Fig. S9. HMBC NMR spectrum of 1 (<https://e-algae.org>).

Supplementary Fig. S10. ROESY NMR spectrum of 1 (<https://e-algae.org>).

Supplementary Fig. S11. Tandem mass spectrometry (MS/MS) analysis for the cleaved linear carbon chain of 1 (<https://e-algae.org>).

Supplementary Fig. S12. (A) High resolution negative mass spectrometry (MS) and tandem mass spectrometry (MS/MS) spectra. (B) Optical rotation value for 2 (<https://e-algae.org>).

Supplementary Fig. S13. ¹H NMR spectrum of 2 (<https://e-algae.org>).

Supplementary Fig. S14. ¹³C NMR spectrum of 2 (<https://e-algae.org>).

Supplementary Fig. S15. (A) High resolution negative mass spectrometry (MS) and tandem mass spectrometry (MS/MS) spectra. (B) Optical rotation value for 3 (<https://e-algae.org>).

Supplementary Fig. S16. ¹H NMR spectrum of 3 (<https://e-algae.org>).

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