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Supramolecular Allosteric Cofacial Porphyrin Complexes

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Abstract

Nature routinely uses cooperative interactions to regulate cellular activity. For years, chemists have designed synthetic systems that aim toward harnessing the reactivity common to natural biological systems. By learning how to control these interactions *in situ*, one begins to allow for the preparation of man-made biomimetic systems that can efficiently mimic the interactions found in Nature. To this end, we have designed a synthetic protocol for the preparation of flexible metal-directed supramolecular cofacial porphyrin complexes which are readily obtained in greater than 90% yield through the use of new hemilabile porphyrin ligands with bifunctional ether–phosphine or thioether–phosphine substituents at the 5 and 15 positions on the porphyrin ring. The resulting architectures contain two hemilabile ligand–metal domains (Rh^I or Cu^I sites) and two cofacially aligned porphyrins (Zn^{II} sites), offering orthogonal functionalities and allowing these multimetallic complexes to exist in two states, “condensed” or “open”. Combining the ether–phosphine ligand with the appropriate Rh^I or Cu^I transition-metal precursors results in “open” macrocyclic products. In contrast, reacting the thioether–phosphine ligand with Rh^I or Cu^I precursors yields condensed structures that can be converted into their “open” macrocyclic forms via introduction of additional ancillary ligands. The change in cavity size that occurs allows these structures to function as allosteric catalysts for the acyl transfer reaction between *X*-pyridylcarbinol (where *X* = 2, 3, or 4) and 1-acetylimidazole. For 3- and 4-pyridylcarbinol, the “open” macrocycle accelerates the acyl transfer reaction more than the condensed analogue and significantly more than the porphyrin monomer. In contrast, an allosteric effect was not observed for 2-pyridylcarbinol, which is expected to be a weaker binder and is unfavorably constrained inside the macrocyclic cavity.

Introduction

Over the past two decades, chemists have been designing synthetic systems that incorporate cofacial porphyrin entities.^{1–10} These systems are of particular interest owing to their unique photophysical properties,^{11–15} their ability to catalyze small molecule transformations,^{16–20} and their use in molecular recognition.^{21,22} Among these, cofacial porphyrin compounds exhibit unique properties attributed to their spatial arrangement with respect to each other, and therefore, the ability to control that arrangement via incorporation of multiple porphyrins in supramolecular arrays offers a potentially facile route to biomimetic structures with novel physicochemical properties.^{23,24} Taking inspiration from nature, a primary goal of our

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research has been to prepare supramolecular structures that exhibit allosteric behavior in the context of catalysis and small-molecule/ion sensing analogous to that displayed ubiquitously in natural enzymatic regulation.^{25–28} In this context, we reasoned that the ability to manipulate the distance between porphyrins by altering the shape and size of supramolecular cavities in which they reside, via external stimuli, may provide a means for designing abiotic allosteric mimics of biological systems. Indeed, a clear inspiration for this kind of allosteric control over the function of multi-porphyrin assemblies comes from perhaps the most famous example of allosteric control in biology, the cooperative homoallosteric protein hemoglobin, in which O₂ binding at one subunit initiates conformational changes that facilitate subsequent O₂ binding at other porphyrin domains.²⁹

To design synthetic allosteric porphyrin-based structures, an attractive strategy entails the preparation of structures that provide facile control over both the orientation and distance of both porphyrins in the context of a small molecule-mediated reaction. To date, most approaches for the synthesis of cofacial porphyrins rely on the use of covalently attached organic molecules as spacers between the porphyrin units.^{6,9,21,30–32} As cofacial porphyrins have been prepared predominantly through synthetic organic approaches, only a few examples exist in which transition metals have been incorporated to facilitate the formation of cofacial porphyrins.^{24,33–36} Additionally, the backbones used to couple these cofacial porphyrins are often based upon rigid linker molecules, which restrict the overall flexibility of the molecule and make the incorporation of different substrates with varying size and shape challenging. To address this issue, several cofacial porphyrin structures have been designed which incorporate flexible linkages.^{22,37–40} However, few are able to specifically access a particular conformation via the introduction or removal of small-molecule effectors at a distal site (i.e., in an allosteric fashion). To this end, we hypothesized that coordination complexes of type **I** and **II** (Scheme 1), whose cavity size can be modified *in situ* with small molecules to form **II** would be very useful for designing stimulant-responsive porphyrin-based biomimetic systems.

Herein, we present a general, high-yielding synthetic methodology for the synthesis of cofacial porphyrin complexes that utilizes flexible porphyrin-based hemilabile ligands and M^I precursors (where M^I = Rh^I or Cu^I) to form macrocyclic products that can be chemically stimulated to change shape. Our strategy, based upon the Weak-Link Approach (WLA),^{41–43} provides rapid and convergent access to unique cofacial porphyrin systems because it relies on the use of metal-heteroatom interactions to facilitate the formation of the desired macrocyclic structures, thus allowing for the controlled and selective modification of the resulting structures *in situ* via introduction of specific external stimuli. Such stimulation can be used to tune the efficiency of a distance-dependent bimolecular acyl transfer reaction and discriminate geometric isomers of substituted pyridylcarbinol substrates.

General Methods and Instrument Details

All reactions were carried out under an inert atmosphere of nitrogen using standard Schlenk techniques or an inert atmosphere glovebox unless otherwise noted. Tetrahydrofuran (THF), diethyl ether (Et₂O), dichloromethane (CH₂Cl₂), acetonitrile (CH₃CN), and hexanes were purified according to published methods.⁴⁴ All solvents were deoxygenated with nitrogen or argon prior to use. 1-Chloro-2-diphenylphosphinoethane (Organometallics Inc.), deuterated solvents (Cambridge Isotope Laboratories Inc.), [Rh(NBD)Cl]₂ (NBD = norbornadiene, Strem Chemicals), and 4-bromothiophenol (Alfa Aesar) were obtained from commercial sources and used as received. 4-(2-Chloroethoxy)-benzaldehyde,⁴⁵ 2-(4-bromophenylsulfanyl) ethyldiphenylphosphane,⁴⁶ 5-mesityldipyrromethane,⁴⁷ and (2-(mesitylthio)ethyl) diphenylphosphine⁴⁸ were prepared according to literature procedures. The synthesis of [5,10,15,20-tetraphenylporphyrinato]zinc^{II} (Zn(TPP))⁴⁹ was adapted from a literature

synthetic procedure. All other chemicals were used as received from Aldrich Chemical Co. ^1H NMR (300.22 MHz) and $^{13}\text{C}\{^1\text{H}\}$ NMR (75.50 MHz) spectra were recorded on a Varian Mercury 300 MHz FT-NMR spectrometer and referenced relative residual proton resonances. $^{31}\text{P}\{^1\text{H}\}$ NMR (121.53 MHz) spectra were recorded on a Varian Mercury 300 MHz FT-NMR spectrometer and referenced relative to an external 85% H_3PO_4 standard. All chemical shifts are reported in ppm. Electrospray ionization mass spectra (ESIMS) were recorded on a Micromass Quatro II triple quadrupole mass spectrometer or a Micromass Q-ToF Ultima mass spectrometer. Electron impact mass spectra (EIMS) were recorded on a Fisons VG 70-250 SE mass spectrometer. Elemental analyses were performed by Quantitative Technologies Inc., Whitehouse, NJ. Gas chromatography (GC) analyses of reaction mixtures were carried out on a computer-interfaced Agilent Technologies 6890 Network instrument equipped with a flame ionization detector (FID). The column used was a 30-m HP-5 capillary column with a 0.32-mm inner diameter and a 0.25- μm film thickness. GC yields were determined through integration of the product peak against biphenyl (internal standard) using pre-established response factors. GC retention times of products were confirmed with analytically pure samples.

2-[4-(2-Chloroethoxy)-phenyl]-[1,3]dithiane (1)

Under ambient conditions, 4-(2-chloroethoxy)benzaldehyde (3.00 g, 16.3 mmol) and 1,3-propanedithiol (1.95 mL, 19.2 mmol) were combined with $\text{CH}_3\text{-CN}$ (100 mL) in a 250-mL round-bottom flask and allowed to stir at room temperature for 5 min at which point $\text{Y}(\text{OTf})_3$ (437 mg, 5 mol %) was added. The resulting solution was stirred for another 30 min when it gradually became turbid due to product precipitation. The mixture was dried on a rotary evaporator to give an oily residue which was then dissolved in CH_2Cl_2 (100 mL), washed with H_2O (2×25 mL), dried over MgSO_4 , and concentrated *in vacuo*. The crude product was purified via column chromatography (1:1 v/v, CH_2Cl_2 /hexanes as an eluent) to yield **1** as a white solid (3.38 g, 75% yield). ^1H NMR (CD_2Cl_2): δ 1.80 (m, 1H, $\text{CH}_2\text{-CH}_\text{A}\text{H}_\text{B}\text{-CH}_2$), 2.14 (m, 1H, $\text{CH}_2\text{-CH}_\text{A}\text{H}_\text{B}\text{-CH}_2$), 2.86 (m, 2H, SCH_2), 2.92 (m, 2H, SCH_2), 3.81 (t, 2H, CH_2Cl), 4.21 (t, 2H, OCH_2), 5.15 (s, 1H, S-CH-S), 6.87 (d, 2H, $J_{\text{H-H}} = 8.4$ Hz, *ArH*), 7.37 (d, 2H, $J_{\text{H-H}} = 8.7$ Hz, *ArH*). $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3): δ 25.2 ($\text{CH}_2\text{-CH}_2\text{-CH}_2$), 32.4 ($\text{CH}_2\text{-CH}_2\text{-CH}_2$), 42.0 (ClCH_2), 50.8 (S-CH-S), 68.2 (OCH_2), 115.0 (*ArC*), 129.3 (*ArC*), 132.3 (CHC_{ipso}), 158.3 (OC_{ipso}). EIMS (m/z): Calcd. 274.03 [M] $^+$. Found: 274.02. Elemental analysis for $\text{C}_{12}\text{H}_{15}\text{ClOS}_2$: Calcd. C, 52.44; H, 5.50. Found: C, 52.35; H, 5.44.

[2-(4-[1,3]Dithian-2-yl-phenoxy)ethyl]diphenylphosphane (2)

In a 100-mL Schlenk round-bottom flask, **1** (1.00 g, 3.64 mmol) was dissolved in THF (50 mL). To this solution, KPPH_2 (7.28 mL of a 0.5 M solution in THF, 3.64 mmol) was added over 10 min and allowed to stir for an additional 30 min. The solvent was removed, and the residue was extracted with degassed $\text{CH}_2\text{Cl}_2/\text{H}_2\text{O}$. The solvent was removed from the organic fraction, yielding **2** as an off-white solid, which was recrystallized from CH_2Cl_2 /hexanes (1.37 g, 86% yield). ^1H NMR (CD_2Cl_2): δ 1.83 (m, 1H, $\text{CH}_2\text{-CH}_\text{A}\text{H}_\text{B}\text{-CH}_2$), 2.11 (m, 1H, $\text{CH}_2\text{-CH}_\text{A}\text{H}_\text{B}\text{-CH}_2$), 2.54 (t, 2H, CH_2P), 2.84 (m, 2H, SCH_2), 3.01 (m, 2H, SCH_2), 4.07 (q, 2H, OCH_2), 5.12 (s, 1H, S-CH-S), 6.73 (d, 2H, $J_{\text{H-H}} = 8.7$ Hz, *ArH*), 7.31–7.50 (br m, 12H, $\text{P}(\text{Ar-H})$). $^{13}\text{C}\{^1\text{H}\}$ NMR (CD_2Cl_2): δ 25.3 ($\text{CH}_2\text{-CH}_2\text{-CH}_2$), 28.3 ($\text{CH}_2\text{-CH}_2\text{-CH}_2$), 28.5 ($\text{CH}_2\text{-CH}_2\text{-CH}_2$), 32.3 (CH_2P), 50.8 (S-CH-S), 65.5, 65.8, 114.8 (*ArC*), 128.7 (*ArC*), 128.8 (*ArC*), 129.0 (*ArC*), 129.0 (*ArC*), 131.9 (*ArC*), 132.7 (*ArC*), 133.0 (*ArC*), 138.3 (*ArC*), 138.4 (CHC_{ipso}), 158.7 (OC_{ipso}). $^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2): δ -21.6 (s). EIMS (m/z): Calcd. 421.11 [M] $^+$. Found: 421.10. Elemental analysis for $\text{C}_{24}\text{H}_{25}\text{OPS}_2$: Calcd. C, 67.90; H, 5.94. Found: C, 67.02; H, 5.86.

2-[4-[2-(Diphenylphosphinothioyl)ethoxy]phenyl]-(1,3)dithiane (3)

In a 100-mL Schlenk round-bottom flask, **2** (1.18 g, 2.76 mmol) and elemental sulfur (88.5 mg, 2.76 mmol) were stirred in THF (50 mL) for 4 h at which point the solvent was removed *in vacuo* and recrystallized from CH₂Cl₂/hexanes to yield **3** as a light-yellow microcrystalline solid (1.23 g, 97% yield). ¹H NMR (CD₂Cl₂): δ 1.83 (m, 1H, CH₂-CH_AH_B-CH₂), 2.11 (m, 1H, CH₂-CH_AH_B-CH₂), 2.84 (m, 6H, (SCH₂)₂ and CH₂P), 4.35 (m, 2H, OCH₂), 5.11 (s, 1H, S-CH-S), 6.62 (d, 2H, *J*_{H-H} = 8.7 Hz, *ArH*), 7.27 (d, 2H, *J*_{H-H} = 8.4 Hz, *ArH*), 7.46–7.91 (br m, 10H, P(*ArH*)). ¹³C{¹H} NMR (CDCl₃): δ 25.2 (CH₂-CH₂-CH₂), 32.4 (CH₂-CH₂-CH₂), 33.0 (CH₂-CH₂-CH₂), 50.9 (CH₂P=S), 53.6 (S-CH-S), 62.5 (OCH₂), 114.8 (ArC), 128.8 (ArC), 129.0 (ArC), 129.1 (ArC), 131.1 (ArC), 131.3 (ArC), 131.8 (ArC), 131.9 (ArC), 132.2 (ArC), 133.3 (ArC), 158.1 (OC_{ipso}). ³¹P-{¹H} NMR (CD₂Cl₂): δ 39.3 (s). EIMS (*m/z*): Calcd. 456.08 [M⁺]. Found. 456.07. Elemental analysis for C₂₄H₂₅OPS₃: Calcd. C, 68.84; H, 5.23. Found: C, 67.43; H, 4.90.

4-[2-(Diphenylphosphinothioyl)ethoxy]benzaldehyde (4)

Under ambient conditions, NaNO₂ (226 mg, 3.27 mmol) and acetyl chloride (0.233 mL, 3.27 mmol) were stirred in CH₂Cl₂ (10 mL) a 100-mL round-bottom flask at 0 °C for 10 min. A solution of **3** (500 mg, 1.09 mmol) in CH₂Cl₂ was added and stirred for an additional 5 min at 0 °C. At this point, H₂O (5 mL) was added and the reaction was brought to room temperature and allowed to stir for an hour. The reaction was neutralized with a saturated aqueous solution of NaHCO₃ and was extracted with CH₂Cl₂ (50 mL). The organic layer was washed with H₂O (2 × 10 mL) and dried over MgSO₄. After removing the solvent *in vacuo*, the crude product was purified via column chromatography (1:1 v/v, ethyl acetate/hexanes as eluent) yielding **4** as a light-yellow microcrystalline solid (352 mg, 88% yield). ¹H NMR (CD₂Cl₂): δ 2.98 (m, 2H, CH₂P), 4.46 (m, 2H, OCH₂), 6.78 (d, 2H, *J*_{H-H} = 9 Hz, *ArH*), 7.48–7.91 (br m, 12H, P(*ArH*)), 9.84 (s, 1H, CHO). ¹³C{¹H} NMR (CD₂Cl₂): δ 32.1 (d, *J*_{C-P} = 56 Hz, CH₂P=S), 63.0 (OCH₂), 114.8 (ArC), 128.8 (C_{ipso}CHO), 128.9 (ArC), 130.4 (ArC), 131.1 (ArC), 131.2 (ArC), 131.9 (ArC), 132.4 (ArC), 133.5 (ArC), 163.2 (C_{ipso}O), 190.7 (CHO). ³¹P{¹H} NMR (CD₂Cl₂): δ 39.2 (s). EIMS (*m/z*): Calcd. 366.08 [M⁺]. Found. 366.08. Elemental analysis for C₂₁H₁₉O₂PS: Calcd. C, 63.13; H, 5.52. Found: C, 62.68; H, 5.22.

5,15-Bis-[4-[2-(diphenylphosphinothioyl)ethoxy]phenyl]-10,20-bis-(mesityl)porphyrin (5)

In an aluminum-foil-wrapped 1000-mL Schlenk round-bottom flask, **4** (1.47 g, 4 mmol), 5-mesityldipyrrromethane (1.06 g, 4 mmol), and activated molecular sieves (4 Å) were stirred in CHCl₃ (600 mL) and degassed under a stream of N₂ for 15 min. BF₃•OEt₂ (0.420 mL) was added dropwise to this solution, and the resulting mixture was allowed to stir for 3 h under N₂. DDQ (1.09 g, 4.8 mmol) was then added as a solid under a stream of N₂, and the reaction was allowed to stir for an additional 30 min at which point NET₃ (4 mL) was added. The reaction mixture was stirred for 1 min before being filtered through a pad of Celite to remove the sieves. The solution was concentrated *in vacuo* and the resulting residue was dissolved in a minimum amount CH₂Cl₂ and poured on top of a silica gel column and purified via flash chromatography (eluent CH₂Cl₂) to yield **5** as a purple microcrystalline solid (1.01 g, 41% yield). ¹H NMR (CD₂Cl₂): δ -2.69 (s, 2H, NH), 1.82 (s, 12H, CH₃), 2.66 (s, 6H, CH₃), 3.16 (m, 4H, CH₂P=S), 4.66 (m, 4H, OCH₂), 7.07 (d, 4H, *J*_{H-H} = 7.8 Hz, *ArH*), 7.30 (s, 4H), 7.59 (br s, 12H, *ArH*), 7.96 (m, 12H, *ArH*), 8.65 (d, 4H, *J*_{H-H} = 5.1 Hz, *ArH*), 8.79 (d, 4H, *J*_{H-H} = 4.5 Hz, *ArH*). ³¹P{¹H} NMR (CD₂Cl₂): δ 39.5 (s). ESIMS (*m/z*): Calcd. 1219.4 [M⁺]. Found: 1219.3. Elemental analysis for C₇₈H₆₈N₄O₂P₂S₂: Calcd. C, 76.82; H, 5.62; N, 4.59. Found: C, 76.04; H, 5.18; N, 4.65.

[5,15-Bis-[4-[2-(diphenylphosphinothioyl)ethoxy]phenyl]-10,20-bis-(mesityl)porphyrinato] zinc (II) (6)

Under ambient conditions, **5** (500 mg, 0.399 mmol) and $\text{Zn}(\text{OAc})_2 \cdot 2\text{H}_2\text{O}$ (700 mg, 3.19 mmol) were combined in a 500-mL round-bottom flask and stirred under reflux for 4 h in a $\text{CHCl}_3/\text{CH}_3\text{OH}$ (4:1 v/v, 350 mL) solution. The solution was then washed with H_2O (100 mL) and extracted with CHCl_3 (2×100 mL). The organic layer was further washed with H_2O (100 mL), dried over Na_2SO_4 , and concentrated to give **6** as a purple microcrystalline solid (507 mg, 96% yield). ^1H NMR ($\text{THF}-d_8$): δ 1.83 (s, 12H, CH_3), 2.60 (m, 6H, CH_3), 2.79 (m, 4H, $\text{CH}_2\text{P}=\text{S}$), 4.36 (m, 4H, OCH_2), 7.16 (br m, 28H, ArH), 8.09 (d, 4H, $J_{\text{H-H}} = 8.4$ Hz, ArH), 8.61 (d, 4H, $J_{\text{H-H}} = 4.5$ Hz, ArH), 8.77 (d, 4H, $J_{\text{H-H}} = 4.5$ Hz, ArH). $^{31}\text{P}\{^1\text{H}\}$ NMR ($\text{THF}-d_8$): δ 39.2 (s). ESIMS (m/z): Calcd. 1282.8 [M^+]. Found: 1282.2. Elemental analysis for $\text{C}_{78}\text{H}_{66}\text{N}_4\text{O}_2\text{P}_2\text{S}_2\text{Zn}$: Calcd. C, 73.03; H, 5.19; N, 4.37. Found: C, 72.89; H, 5.23; N, 4.17.

[5,15-Bis-[4-(2-diphenylphosphanylethoxy)phenyl]-10,20-bis-(mesityl)porphyrinato]zinc(II) (7)

In a 50-mL Schlenk round-bottom flask, **6** (300 mg, 0.234 mmol) and Cp_2ZrHCl (392 mg, 1.52 mmol) were stirred in THF under N_2 (40 mL) at 60 °C for 4 h. The solvent was removed and the reaction was purified via column chromatography (silica gel, THF) in a glove box under an atmosphere of N_2 . The solvent was removed *in vacuo* to yield **7** as a purple microcrystalline solid (254 mg, 89% yield). ^1H NMR ($\text{THF}-d_8$): δ 1.83 (s, 12H, CH_3), 2.60 (s, 6H, CH_3), 2.77 (m, 4H, CH_2P), 4.36 (m, 4H, CH_2O), 7.17 (d, 4H, $J_{\text{H-H}} = 6.6$ Hz, ArH), 7.30–7.61 (bm, 24H, ArH), 8.01 (d, 4H, $J_{\text{H-H}} = 8.7$ Hz, ArH), 8.64 (d, 4H, $J_{\text{H-H}} = 4.5$ Hz, ArH), 8.78 (d, 4H, $J_{\text{H-H}} = 4.5$ Hz, ArH). $^{31}\text{P}\{^1\text{H}\}$ NMR ($\text{THF}-d_8$): δ -21.2 (s). ESIMS (m/z): Calcd. 1218.74 [M^+]. Found: 1217.4. Elemental analysis for $\text{C}_{78}\text{H}_{66}\text{N}_4\text{O}_2\text{P}_2\text{Zn}$: Calcd. C, 76.87; H, 5.46; N, 4.60. Found: C, 76.37; H, 5.18; N, 4.45.

[(7)RhCl(CO)]₂ Macrocycle (8a)

A small vial was charged with $[\text{Rh}(\text{CO})_2(\text{Cl})]_2$ (4.80 mg, 0.0123 mmol) and CH_2Cl_2 (2 mL). The resulting solution was stirred for 1 min, at which point a solution of **7** (30.0 mg, 0.0246 mmol) in THF (5 mL) was added dropwise over 1 min. The resulting solution was stirred for 3 h. The solvent was removed and the product was recrystallized from $\text{CH}_2\text{Cl}_2/\text{pentane}$ (32.1 mg, 94% yield). ^1H NMR (CD_2Cl_2): δ 1.54 (s, 24H, CH_3), 2.56 (s, 12H, CH_3), 3.34 (br m, 8H, CH_2P), 4.81 (br m, 8H, CH_2O), 7.10 (br m, 22H, ArH), 7.47 (m, 22H, ArH), 7.90 (br m, 20H, ArH), 8.79 (br m, 16H, ArH). $^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2): δ 21.6 (d, $J_{\text{Rh-P}} = 124$ Hz). ESIMS (m/z) for $[\text{C}_{156}\text{H}_{132}\text{N}_8\text{O}_4\text{P}_4\text{Rh}_2\text{Zn}_2(\text{CO})_2]^{2+}$: Calcd. 1349.6. Found: 1349.3. Elemental analysis for $\text{C}_{158}\text{H}_{132}\text{Cl}_2\text{N}_8\text{O}_6\text{P}_4\text{Rh}_2\text{Zn}_2$: Calcd. C, 68.51; H, 4.80; N, 4.04. Found: C, 68.38; H, 4.91; N, 3.52.

[(7)Cu(CH₃CN)₂(PF₆)₂] Macrocycle (8b)

A 50-mL Schlenk flask was charged with $[\text{Cu}(\text{CH}_3\text{CN})_4]\text{PF}_6$ (29.8 mg, 0.0799 mmol) and $\text{CH}_2\text{-Cl}_2$ (5 mL). A THF solution of **7** (100 mg, 0.0799 mmol, 20 mL) was added to the “Cu” solution dropwise over 5 min at room temperature to give a red/purple solution, which was then allowed to stir for 3 h. The solvent was removed to yield a purple microcrystalline solid which was recrystallized from $\text{CH}_2\text{Cl}_2/\text{pentane}$ (107 mg, 92% yield). ^1H NMR (CD_2Cl_2): δ 1.66 (s, 24H, CH_3), 2.07 (s, 12H, CH_3CN), 2.43 (s, 12H, CH_3), 2.95 (br m, 8H, CH_2P), 4.46 (br m, 8H, CH_2O), 7.10 (s, 22H, ArH), 7.50 (s, 22H, ArH), 7.62 (12H, ArH), 8.00 (d, 8H, $J_{\text{H-H}} = 7.8$ Hz, ArH), 8.64 (d, 8H, $J_{\text{H-H}} = 4.2$ Hz, ArH), 8.78 (d, 8H, $J_{\text{H-H}} = 4.2$ Hz, ArH). $^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2): δ -11.5 (s). ESIMS (m/z) for $[\text{C}_{156}\text{H}_{132}\text{N}_8\text{S}_4\text{P}_4\text{Cu}_2\text{Zn}_2]^{2+}$: Calcd. 1282.2. Found: 1282.4. Elemental analysis for $[\text{C}_{164}\text{H}_{144}\text{N}_{12}\text{O}_4\text{P}_6\text{F}_{12}\text{Zn}_2\text{Cu}_2]$: Calcd. C, 65.25; H, 4.81; N, 5.57. Found: C, 65.32; H, 4.56; N, 6.35.

1-Bromo-4-[2-(diphenylphosphinothioyl)ethylsulfanyl]-benzene (9)

In a 100-mL Schlenk round-bottom flask, 2-(4-bromo-phenylsulfanyl)-ethylidiphenylphosphane (3.00 g, 7.48 mmol), and elemental sulfur (264 mg, 8.22 mmol) were stirred in THF (150 mL) under N₂ for 3 h. The reaction mixture was concentrated *in vacuo*, and the product was purified via column chromatography (1:1 v/v, CH₂Cl₂/hexanes).

Compound **9** was isolated as an off-white microcrystalline solid (3.02 g, 92% yield). ¹H NMR (CD₂Cl₂): δ 2.68 (m, 2H, CH₂P=S), 3.10 (m, 2H, SCH₂), 7.12 (d, 2H, J_{H-H} = 8.7 Hz, ArH), 7.39 (d, 2H, J_{H-H} = 8.7 Hz, ArH), 7.48–7.79 (m, 10H, P(ArH)). ¹³C{¹H} NMR (CDCl₃): δ 26.9 (SCH₂), 32.3 (d, CH₂P=S, J_{C-P} = 51.3 Hz), 128.9 (ArC), 129.1 (ArC), 131.0 (ArC), 131.1 (ArC), 131.2 (ArC), 131.6 (ArC), 132.0 (ArC), 132.3 (ArC). ³¹P{¹H} NMR (CD₂Cl₂): δ 41.4 (s). EIMS (*m/z*): Calcd. 431.97 [M⁺]. Found: 431.97. Elemental analysis for C₂₀H₁₈-BrPS₂: Calcd. C, 55.43; H, 4.19. Found: C, 55.59, H, 4.07.

4-[2-(Diphenylphosphinothioyl)ethylsulfanyl]benzaldehyde (10)

Compound **9** (3.00 g, 6.90 mmol) was dissolved in THF (60 mL) in a 100-mL Schlenk round-bottom flask and was cooled to –78 °C. *n*-BuLi (2.76 mL, 6.90 mmol, 2.5 M in hexanes) was added dropwise to the solution over 5 min and the mixture was allowed to stir for 30 min, before DMF (0.802 mL, 10.35 mmol) was added to the flask. This solution was cooled to –78 °C and allowed to stir for an additional 30 min before the temperature was allowed to rise back to room temperature. The mixture was quenched with H₂O followed by extraction with CH₂Cl₂. The organic layer was dried over MgSO₄, filtered, and concentrated *in vacuo* to give a yellow crude product which was then recrystallized (CH₂Cl₂/pentane) to yield **10** as a light-yellow microcrystalline solid (2.35 g, 88% yield). ¹H NMR (CD₂Cl₂): δ 2.76 (m, 2H, CH₂P=S), 3.23 (m, 2H, SCH₂), 7.29 (d, 2H, J_{H-H} = 6.6 Hz, ArH) 7.47 (m, 6H, P(ArH)), 7.73 (d, 2H, J_{H-H} = 8.7 Hz, ArH), 7.79 (m, 4H, P(ArH)), 9.92 (s, 1H, CHO). ¹³C{¹H} NMR (CDCl₃): δ 25.0 (SCH₂), 32.0 (d, CH₂P=S, J_{C-P} = 51.3 Hz), 126.8 (ArC), 129.0 (ArC), 129.1 (ArC), 130.4 (ArC), 131.1 (ArC), 131.3 (ArC), 132.1 (ArC), 132.5 (ArC), 191.4 (CHO). ³¹P{¹H} NMR (CD₂Cl₂): δ 41.5 (s). EIMS (*m/z*): Calcd. 382.01 [M⁺]. Found: 382.06. Elemental analysis for C₂₁H₁₉-OPS₂: Calcd. C, 65.94; H, 5.01. Found: C, 65.23; H, 4.71.

[5,15-Bis-[4-(2-diphenylphosphinothioylethylsulfanyl)phenyl]-10,-20-bis-(mesityl)porphyrin (11)

In an aluminum-foil-wrapped 1000-mL Schlenk flask, compound **10** (1.23 g, 3.20 mmol), 5-mesityldipyrrromethane (848 mg, 3.20 mmol), and activated molecular sieves (4 Å) were stirred in CHCl₃ (600 mL) and degassed under a stream of N₂ for 15 min. BF₃•OEt₂ (0.350 mL) was added dropwise to this solution, and the resulting mixture was allowed to stir for 3 h under N₂. DDQ (862 mg, 3.8 mmol) was then added as a solid under a stream of N₂, and the reaction was allowed to stir for an additional 30 min at which point NET₃ (4 mL) was added. The reaction mixture was stirred for 1 min before being filtered through a pad of Celite to remove the sieves. The solution was concentrated *in vacuo* and the resulting residue was dissolved in CH₂Cl₂ and poured on top of a silica gel column (eluent CH₂Cl₂). Subsequent chromatography yielded **11** as a purple microcrystalline solid (879 mg, 44% yield). ¹H NMR (CD₂Cl₂): δ –2.67 (s, 2H, NH), 1.84 (s, 12H, CH₃), 2.63 (s, 6H, CH₃), 3.03 (m, 4H, CH₂P=S), 3.42 (m, 4H, SCH₂), 7.31 (s, 4H, mesityl H), 7.55 (m, 16H, ArH), 7.89 (m, 8H, ArH), 8.13 (d, 4H, J_{H-H} = 8.1 Hz, ArH), 8.69 (d, 4H, J_{H-H} = 5.1 Hz, ArH), 8.82 (d, 4H, J_{H-H} = 4.8 Hz, ArH). ³¹P{¹H} NMR (CD₂Cl₂): δ 41.6 (s). ESIMS (*m/z*): Calcd. 1251.6 [M⁺]. Found: 1251.3. Elemental analysis for C₇₈H₆₈N₄P₂S₄: Calcd. C, 74.85; H, 5.48; N, 4.48. Found: C, 74.35; H, 4.98; N, 4.43.

[5,15-Bis-[4-(2-diphenylphosphinothioylethylsulfanyl)phenyl]-10,-20-bis(mesityl)porphyrinato] zinc(II) (12)

In a 250-mL round-bottom flask, **11** (400 mg, 0.319 mmol) and Zn(OAc)₂•2H₂O (700 mg, 3.19 mmol) were refluxed for 3 h in a CHCl₃/CH₃OH (4:1 v/v, 200 mL) solution. The solution was washed with H₂O (100 mL) and extracted with CHCl₃ (2 × 50 mL). The organic layer was washed again with H₂O (50 mL), dried over Na₂SO₄, and concentrated to give **12** as a purple microcrystalline solid (412 mg, 98% yield). ¹H NMR (CD₂-Cl₂): δ 1.82 (s, 12H, CH₃), 2.63 (s, 6H, CH₃), 3.03 (m, 4H, CH₂P=S), 3.41 (m, 4H, SCH₂), 7.31 (s, 4H, ArH), 7.54 (m, 12H, ArH), 7.89 (m, 12H, ArH), 8.12 (d, 4H, J_{H-H} = 8.4 Hz, ArH), 8.76 (d, 4H, J_{H-H} = 5.1 Hz, ArH), 8.89 (d, 4H, J_{H-H} = 4.8 Hz, ArH). ³¹P{¹H} NMR (CD₂-Cl₂): δ 41.6 (s). ESIMS (*m/z*): Calcd. 1314.9 [M⁺]. Found: 1314.1. Elemental analysis for C₇₈H₆₆N₄P₂S₄Zn: Calcd. C, 71.24; H, 5.06; N, 4.26. Found: C, 70.69; H, 4.97; N, 3.94.

[5,15-Bis-[4-(2-diphenylphosphanylethylsulfanyl)phenyl]-10,20-bis(mesityl)porphyrinato] zinc(II) (13)

In a 50-mL Schlenk flask, **12** (300 mg, 0.228 mmol) and Cp₂ZrHCl (382 mg, 1.48 mmol) were stirred in THF (40 mL) at 60 °C for 3 h. The solvent was removed, and the product was purified via column chromatography (THF, silica gel) in a glove box under an atmosphere of N₂. The solvent was removed *in vacuo* to yield **13** as a purple microcrystalline solid (251 mg, 88% yield). ¹H NMR (THF-*d*₈): δ 1.84 (s, 12H, CH₃), 2.47 (m, 4H, CH₂P), 2.61 (s, 6H, CH₃), 3.23 (m, 4H, SCH₂), 7.29 (br m, 28H, ArH), 8.09 (d, 4H, J_{H-H} = 7.8 Hz, ArH), 8.64 (d, 4H, J_{H-H} = 4.8 Hz, ArH), 8.79 (d, 4H, J_{H-H} = 4.2 Hz, ArH). ³¹P{¹H} NMR (THF-*d*₈): δ -15.2 (s). ESIMS (*m/z*): Calcd. 1250.8 [M⁺]. Found: 1249.4. Elemental analysis for C₇₈H₆₆N₄P₂S₂Zn: Calcd. C, 74.90; H, 5.32; N, 4.48. Found: C, 73.15; H, 4.94; N, 4.16.

[(13)Rh(BF₄)₂] Condensed Macrocycle (14a)

A small vial was charged with [Rh(NBD)Cl]₂ (31.0 mg, 0.068 mmol), AgBF₄ (26.0 mg, 0.135 mmol), and CH₂Cl₂ (4 mL). This solution was stirred for 1 h and then filtered dropwise through Celite into a Schlenk flask. The red solution was then diluted with CH₂Cl₂ (20 mL) to give a clear yellow/orange solution. A solution of ligand **13** (170 mg, 0.135 mmol) in THF (20 mL) was added to the “Rh” solution dropwise over 5 min at room temperature to give a dark-purple solution, which was then stirred for an additional 3 h. The solvent was removed to yield **14a** as a purple microcrystalline solid which was recrystallized from CH₂Cl₂/pentane (176 mg, 90% yield). ¹H NMR (CD₂Cl₂): δ 1.48 (br s, 24H, CH₃), 2.44 (br s, 12H, CH₃), 2.76 (br m, 8H, CH₂P), 3.20 (br m, 8H, CH₂S), 6.98 (br m, 8H, ArH), 7.37 – 7.59 (br m, 40H, ArH), 8.02 (br m, 16H, ArH), 8.50 (br m, 16H, ArH). ³¹P{¹H} NMR (CD₂Cl₂): δ 64.1 (d, J_{Rh-P} = 162 Hz). ESIMS (*m/z*) for [C₁₅₆H₁₃₂N₈S₄P₄Rh₂Zn₂]²⁺: Calcd. 1353.7. Found: 1354.1. Elemental analysis for [C₁₅₆H₁₃₂B₂F₈N₈S₄P₄-Rh₂Zn₂]: Calcd. C, 65.03; H, 4.62; N, 3.89. Found: C, 65.56; H, 4.11; N, 3.46.

[(13)Cu(PF₆)₂] Condensed Macrocycle (14b)

A 50-mL Schlenk flask was charged with [Cu(CH₃CN)₄]PF₆ (29.8 g, 0.0799 mmol). The Cu^I precursor was dissolved in CH₂Cl₂ (5 mL), and a THF solution of ligand **13** (100 mg, 0.0799 mmol, 20 mL) was added to the “Cu” solution dropwise over 5 min at room temperature to give a red/purple solution. The solution was then allowed to stir for 3 h. The solvent was removed to yield **14b** as a purple microcrystalline solid which was recrystallized from CH₂Cl₂/pentane (105 mg, 90% yield). ¹H NMR (CD₂Cl₂): δ 1.75 (br s, 24H, CH₃), 2.55 (br s, 12H, CH₃), 3.00 (br m, 8H, CH₂P), 3.65 (br m, 8H, CH₂S), 7.24 – 7.82 (br m, 56H, ArH), 8.16 (br m, 8H, ArH), 8.76 (br s, 16H, ArH). ³¹P{¹H} NMR (CD₂Cl₂): δ 1.0 (s). ESIMS (*m/z*) for [C₁₅₆H₁₃₂N₈S₄P₄Cu₂Zn₂]²⁺: Calcd. 1314.4. Found: 1314.5. Elemental analysis for

$[C_{156}H_{132}N_8S_4P_6F_{12}Cu_2Zn_2]CH_2-Cl_2$: Calcd. C, 62.78; H, 4.50; N, 3.73. Found: C, 61.95; H, 4.18; N, 3.31.

[(13)RhCl(CO)]₂ Macrocycle (15a)

Compound **14a** (20.0 mg, 0.00694 mmol) was dissolved in CD_2Cl_2 (1 g) and placed in an air-free NMR tube. PPNCI (Bis(triphenylphosphoranylidene) ammonium chloride, 2 equiv) was added and the NMR tube was then pressurized with CO (1 atm) for 30 s. Compound **14a** was quantitatively converted to **15a** as determined by ^{31}P NMR spectroscopy. 1H NMR (CD_2Cl_2): δ 1.77 (br s, 24H, CH_3), 2.54 (br s, 12H, CH_3), 3.14 (br m, 8H, CH_2P), 3.48 (br m, 8H, CH_2S), 7.26 – 8.01 (br m, 124H, $(C_6H_5)_3PPN$, $P(C_6H_5)_2$, ArH), 8.65 – 8.80 (br m, 16H, ArH). $^{31}P\{^1H\}$ NMR (CD_2Cl_2): δ 22.1 (s, PPNCI), δ 25.1 (d, $J_{Rh-P} = 123$ Hz). ESIMS (m/z) for $[C_{156}H_{132}N_8S_4P_4-Rh_2Zn_2]^{2+}$ ($M - 2(Cl^-/CO)$): Calcd. 1353.7. Found: 1353.7.

[(13)Cu(pyridine-*d*₅)₂(PF₆)₂] Macrocycle (15b)

Compound **14b** (20.0 mg, 0.00640 mmol) was dissolved in CD_2Cl_2 (1 g) and placed in an air-free NMR tube. C_5D_5N (4 equiv.) was added to this solution and the conversion of **15b** was quantitative as determined by 1H NMR and ^{31}P NMR spectroscopy. 1H NMR (CD_2Cl_2): δ 1.72 (br s, 24H, CH_3), 2.45 (br s, 12H, CH_3), 2.82 (br m, 8H, CH_2P), 3.38 (br m, 8H, CH_2S), 7.17 – 7.55 (br m, 56H, ArH), 8.04 (d, 8H, $J_{H-H} = 7.8$ Hz, ArH), 8.64 (d, 8H, $J_{H-H} = 4.5$ Hz, ArH), 8.70 (d, 8H, $J_{H-H} = 4.8$ Hz, ArH). $^{31}P\{^1H\}$ NMR (CD_2Cl_2): δ –8.0 (s). ESIMS (m/z) for $[C_{156}H_{132}N_8S_4P_4Cu_2Zn_2]^{2+}$: Calcd. 1314.4. Found: 1314.9.

[(Ph₂PCH₂CH₂S–C₆H₂(CH₃)₃)₂Rh]BF₄ Condensed Tweezer-Type Complex (16a)

A 20-mL vial was charged with $[Rh(NBD)Cl]_2$ (63.2 mg, 0.137 mmol), $AgBF_4$ (53.4 mg, 0.274 mmol), and CH_2Cl_2 (4 mL). This solution was stirred for 1 h and then filtered dropwise through Celite into a Schlenk flask. The orange solution was then diluted with CH_2Cl_2 (20 mL) to give a clear yellow solution. A solution of (2-(mesitylthio)ethyl)diphenylphosphine (200 mg, 0.549 mmol) in CH_2-Cl_2 (20 mL) was added to the “Rh” solution dropwise over 5 min at room temperature to give a clear yellow solution, which was then stirred for an additional 3 h. The solvent was removed to yield **16a** as a yellow microcrystalline solid which was recrystallized from CH_2Cl_2 /pentane (113 mg, 90% yield). 1H NMR (CD_2Cl_2): δ 2.22 (d, 12H, CH_3), 2.51 (br s, 10H, CH_3 and SCH_2CH_2P), 2.83 (br m, 4H, CH_2P), 6.71 (s, 4H, ArH), 7.23 – 7.34 (br m, $P(Ar-H)_2$, 20H). $^{31}P\{^1H\}$ NMR (CD_2Cl_2): δ 63.0 (d, $J_{Rh-P} = 163$ Hz). ESIMS (m/z) for $[C_{46}H_{50}N_8S_2P_2Rh]^+$: Calcd. 831.8. Found: 831.6.

(Ph₂PCH₂CH₂S–C₆H₂(CH₃)₃)₂RhCl(CO) Open Tweezer-Type Complex (16b)

Compound **16a** (15.0 mg, 0.0163 mmol) was dissolved in CD_2Cl_2 (1 g) and placed in an air-free NMR tube. PPNCI (bis(triphenylphosphoranylidene)ammonium chloride, 9.38 mg, 0.0163 mmol) was added as a solid and the NMR tube was then pressurized with CO (1 atm) for 30 s. Compound **16a** was quantitatively converted to **16b** as determined by $^{31}P\{^1H\}$ NMR spectroscopy. 1H NMR (CD_2-Cl_2): δ 2.25 (s, 6H, CH_3), 2.37 (s, 12H, CH_3), 2.67 (br m, 4H, CH_2P), 2.82 (br m, 4H, CH_2S), 6.90 (s, 4H, ArH), 7.34 – 7.65 (br m, 50H, $(C_6H_5)_3PPN$, $P(Ar-H)_2$). $^{31}P\{^1H\}$ NMR (CD_2Cl_2): δ 22.1 (s, PPNCI), δ 23.5 (d, $J_{Rh-P} = 123$ Hz). ESIMS (m/z) for $[C_{46}H_{50}N_8S_2P_2Rh]^+$ ($M - (Cl^-/CO)$): Calcd. 831.8. Found: 831.2.

General Procedure for Catalysis Experiments

The formation of 2-, 3-, and 4-acetoxymethylpyridine were monitored by GC relative to an internal standard (biphenyl) and quantified using a previously established calibration curve. Experiments containing **14a**, **15a**, or a mixture of an analogous monomeric Rh^I complex (**16a** or **16b**) and $Zn(TPP)$ (as the control), respectively, were run concurrently in separate vials at room temperature in CH_2Cl_2 under an atmosphere of N_2 inside a glove box.

GC Experiments for the Closed Macrocycle (14a)—Inside a glove box, CH₂Cl₂ stock solutions of complex **14a** (0.58 mL of a 2.6 mM solution), biphenyl (0.5 mL of a 25 mM solution), and pyridylcarbinol (0.5 mL of a 90 mM solution) were added to a 20-mL vial. Fresh CH₂-Cl₂ was added to the vial bringing the total reaction volume up to 4.5 mL. After stirring for 5 min, 1-acetylimidazole (0.5 mL of a 60 mM solution) was added to the vial ($t = 0$). At various times, an aliquot (100 μ L) was taken from the solution and added to diethyl ether (2 mL). This was then passed down a plug of Celite (3 cm \times 0.5 cm) to remove the catalyst. The plug was further treated with fresh diethyl ether (5 mL). The combined organics were used for GC analysis.

GC Experiments for the Open Macrocycle (15a)—Inside a glove box, complex **14a** (0.58 mL of a 2.6 mM solution) and benzyltriethylammonium chloride (0.5 mL of a 6.4 mM solution) were added to a 10-mL Schlenk flask. The flask was removed from the glove box and placed on a Schlenk line where CO (1 atm) was bubbled through the solution for 30 s resulting in the formation of **15a**. The flask was brought back into the glove box and the contents were transferred into a vial. Fresh CH₂Cl₂ was added to the flask and transferred into the vial bringing the total reaction volume up to approximately 4 mL. Pyridylcarbinol (0.5 mL of a 90 mM solution) and biphenyl (0.5 mL of a 25 mM solution) were added to the vial and after stirring for 5 min, 1-acetylimidazole (0.5 mL of a 60 mM solution) was added to the flask ($t = 0$). At various times, an aliquot (100 μ L) was taken from the solution and added to diethyl ether (2 mL). This was then passed down a plug of Celite (3 cm \times 0.5 cm) to remove the catalyst. The plug was further treated with fresh diethyl ether (5 mL). The resulting samples were used for GC analysis.

GC Control Experiments for Rh^I-Monomer (16a) and Porphyrin Monomer (Zn(TPP))—Inside a glove box, CH₂Cl₂ stock solutions of complex **16a** (0.892 mL of a 8.0 mM solution), biphenyl (0.5 mL of a 25 mM solution), Zn(TPP) (0.20 mL of a 15 mM solution), and pyridylcarbinol (0.5 mL of a 90 mM solution) were added to a 20-mL vial. Fresh CH₂Cl₂ was added to the vial bringing the total reaction volume up to 4.5 mL. After stirring for 5 min, 1-acetylimidazole (0.5 mL of a 60 mM solution) was added to the vial ($t = 0$). At various times, an aliquot (100 μ L) was taken from the solution and added to diethyl ether (2 mL). This was then passed down a plug of Celite (3 cm \times 0.5 cm) to remove the catalyst. The plug was further treated with fresh diethyl ether (5 mL). The combined organics were used for GC analysis.

GC Control Experiments for Rh^I-Monomer (16b) and Porphyrin Monomer—Inside a glove box, complex **16a** (0.892 mL of a 8.0 mM solution), Zn(TPP) (0.20 mL of a 15 mM solution), and benzyltriethylammonium chloride (1.0 mL of a 6.4 mM solution) were added to a 10-mL Schlenk flask. The flask was removed from the glove box and placed on a Schlenk line where CO (1 atm) was bubbled through the solution for 30 s resulting in the formation of **16b**. The flask was brought back into the glove box and the contents were transferred into a vial. Fresh CH₂Cl₂ was added to the flask and transferred into the vial bringing the total reaction volume up to approximately 4 mL. Pyridylcarbinol (0.5 mL of a 90 mM solution) and biphenyl (0.5 mL of a 25 mM solution) were added to the vial and after stirring for 5 min, 1-acetylimidazole (0.5 mL of a 60 mM solution) was added to the flask ($t = 0$). At various times, an aliquot (100 μ L) was taken from the solution and added to diethyl ether (2 mL). This was then passed down a plug of Celite (3 cm \times 0.5 cm) to remove the catalyst. The plug was further treated with fresh diethyl ether (5 mL). The combined organics were used for GC analysis.

Results and Discussion

The Design of Ether–Phosphine Hemilabile Ligand **7**

To design porphyrin-based supramolecular systems capable of *in situ* allosteric activity, flexibility must be engineered into the backbone of the framework as well as “weak” and “strong” binding structural domains. To this end, we synthesized two new hemilabile porphyrin ligands which incorporate “weak” ether or thioether functionalities in addition to “strong” phosphine binding sites. Ether-based ligand **7** was obtained in eight steps from 4-hydroxybenzaldehyde (Scheme 2). 4-Hydroxy-benzaldehyde was alkylated with excess 1-bromo-2-chloroethane to yield 4-(2-chloroethoxy)benzaldehyde, which was transformed into **3** after a dithiane protection of the aldehyde, conversion of the chloride to the corresponding phosphine, and reaction with elemental sulfur. Deprotection of the dithiane moiety in **3**, followed by condensation of the resulting aldehyde with 5-mesityldipyrromethane affords the freebase porphyrin **5** in gram quantities after column chromatography. Metalation with Zn(OAc)₂•2H₂O gave compound **6**, and desulfurization with Schwartz’s reagent gave ligand **7** in 12% overall yield. The use of a sulfide as the protecting group for the phosphine moiety not only allowed for a convenient nonaqueous protection/deprotection sequence, but also significantly increased the scalability and the yield of the porphyrin synthesis while simplifying the complicated purification and isolation process that often accompanies porphyrin syntheses. Indeed, if one uses an oxide protecting group as opposed to the sulfide, the ligand adheres to the silica column used for chromatography and makes the isolation of the pure product difficult.

Reactivity of Ligand **7** with Rh^I and Cu^I

Initially, our attention was focused on the design, synthesis, and isolation of Rh^I and Cu^I condensed intermediates of type **I** using ligand **7**. Unfortunately, isolation of these products proved impossible under the conditions explored. However, we discovered that the “open” macrocyclic structure **8a** can be obtained directly in quantitative yield upon reaction of ligand **7** with [Rh(CO)₂(Cl)]₂. The ³¹P{¹H} NMR spectrum of **8a** exhibits a single resonance at δ 21.6 (d, $J_{\text{Rh-P}} = 124$ Hz), diagnostic of a highly symmetrical porphyrin complex with *trans*-phosphines and is consistent with the proposed structural formulation for **8a**.⁵⁰ ESIMS analysis of **8a** shows a parent ion at m/z 1349.3, indicating the loss of each Cl[−] ligand bound to the Rh^I centers. Rh^I complexes with analogous coordination environments often lose these ligands during ESIMS.⁵⁰

Similar to the reactivity exhibited with Rh^I, ligand **7** forms the analogous “open” macrocyclic product **8b** when reacted with [Cu(CH₃CN)₄]PF₆. The ³¹P{¹H} NMR spectrum of **8b** exhibits a singlet at δ −11.5 consistent with a tetrahedral Cu^I–P environment.⁵¹ The resonance is substantially shifted downfield from the one observed for free ligand at δ −21.2. Characterization of this product via ESIMS leads to the observation of a parent ion at m/z 1282.4 which corresponds to the loss of two CH₃CN molecules from each Cu^I metal center. This is consistent with the known lability of the bound CH₃CN molecules, which has been observed in analogous systems.⁵¹

X-ray Structure Determination of **8a**⊂DABCO and **8c**⊂DABCO

Initial attempts to grow crystals of **8a** in a variety of solvent mixtures proved unsuccessful, presumably due to the large free volume and flexibility of the cavity. We hypothesized that crystals of **8a** could be obtained in the presence of a ligand that can span and coordinate to the two Zn-centers within the macrocycle, filling the free volume and rigidifying the overall structure. Indeed, X-ray quality crystals were obtained from vapor diffusion of pentane/diethyl ether (1:1 v/v) into a THF/CH₂Cl₂ (1:1 v/v) solution of **8a** containing DABCO which was layered with acetonitrile. The 56-membered macrocyclic product **8a**⊂DABCO contains two porphyrin moieties aligned and locked into a cofacial arrangement in the solid-state by a

DABCO ligand (Figure 1). The Zn–Zn distance of 7.09 Å is significantly longer than the P–Rh–P distance of 4.64 which is made possible by the flexible (CH₂)₂O linkages between the porphyrin moieties and the Rh^I hinges. This large porphyrin–porphyrin distance allows for the mesityl arms of the porphyrin moieties to align in a near superimposable fashion.

Attempts to crystallize **8b** in a similar manner led to an unexpected reaction with the halogenated solvent. Because the solubility of **8b** in pure THF is poor, a mixture of solvents proved to be the most effective for crystallization. Analytically pure samples of **8b** were dissolved in a THF/CH₂Cl₂ solution containing DABCO which was first layered with acetonitrile following by a slow second layer of a solution of pentane/diethyl ether (1:1 v/v). X-ray quality crystals were obtained from this mixture after 1 day and subjected to single-crystal diffraction analysis at the Advanced Photon Source (APS, Argonne National Laboratory). Although **8b** has been shown to contain four molecules of CH₃CN and two PF₆[−] counterions according to elemental analysis, the diffraction experiment yielded a structure with DABCO bridging both Zn-atoms and two Cl[−] anions bound to each tetrahedral Cu metal center, indicating an oxidation of the original Cu^I center to Cu^{II} forming **8cCDABCO** (see Supporting Information). Such a compound could be formed from the dehalogenation of alkyl halide solvents (i.e., CH₂Cl₂ and CHCl₃) by the Cu^I center in **8b**, similar to that observed by Karlin et al. for (TMPA)Cu^I (TMPA= (tris-(2-pyridylmethyl)amine)) in CH₂Cl₂.^{52–54} That **8cCDABCO** possesses the framework expected for **8b** supports our assignment of an open macrocyclic structure for the product isolated from the reaction of **7** and [Cu(CH₃CN)₄]PF₆.

Synthesis of Thioether–Phosphine Hemilabile Ligand 13

Because type **I** structures are difficult to access from the ether–phosphine ligand **7**, we hypothesized that isostructural ligands containing thioether linkages would be more effective at forming condensed structures as has been observed previously with analogous smaller molecules.⁵⁰ To this end, we modified the synthesis of **7** to obtain the thioether-based porphyrin ligand **13** in six steps (Scheme 3). First, 4-bromothiophenol was stoichiometrically alkylated with 1-chloro-2-diphenylphosphinoethane to yield 2-(4-bromophenylsulfanyl) ethyldiphenyl phosphine. To protect the phosphine moiety from oxidation during the porphyrin synthesis, reaction with elemental sulfur yields phosphine sulfide **9**, which was then formylated with *n*-BuLi and DMF. The resulting aldehyde **10** was subsequently condensed with 5-mesityldipyrromethane in the presence of BF₃•OEt₂ to yield porphyrin **11** in 44% yield after a very simple chromatographic separation. Freebase porphyrin **11** was metallated with Zn (OAc)₂•2H₂O to give **12** and followed by deprotection of the phosphine with Schwartz's reagent to yield the phosphine derivative **13** in 33% overall yield.

Reactivity of Hemilabile Ligand 13 with Rh^I and Cu^I

In contrast to the results obtained from ether ligand **7**, thioether ligand **13** reacts cleanly with “Rh(NBD)BF₄”⁵⁵ to form the condensed intermediate **14a**. The ³¹P{¹H} NMR spectrum of **14a** exhibits a doublet at δ 64.5 (*J*_{Rh–P} = 162 Hz) which is highly diagnostic of its symmetrical structure and *cis*-Rh–P coordination centers.⁵⁰ Additionally, ESIMS analysis shows a peak corresponding to the M²⁺ ion at 1354.1 *m/z*, indicating the formation of the desired condensed structure. Significantly, compound **14a** can be opened into macrocycle **15a** by introduction of benzyltriethyl ammonium chloride or by PPNCI (where PPNCI = bis (triphenylphosphoranylidene)ammonium chloride) and CO. This transformation can be followed by ³¹P{¹H} NMR through the appearance of a doublet at δ 25.1 (*d*, *J*_{P–Rh} = 123 Hz), significantly upfield from the resonance for **14a**. This resonance is characteristic of a *trans*-phosphine environment about the Rh^I metal center and indicates a conversion from a condensed macrocyclic intermediate to the “open” macrocycle **15a**.⁵⁰ The ESIMS spectrum of **15a** exhibits a peak at 1353.7 *m/z*, corresponding to the M²⁺ ion without the Cl[−] and CO ligands and is consistent with ESIMS data for analogous structures.²⁶

We observed condensed intermediate **14b** as the sole product from the reaction between thioether-based ligand **13** and $[\text{Cu}-(\text{CH}_3\text{CN})_4]\text{PF}_6$. The $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum of **14b** exhibits a singlet at δ 1.0 which is highly diagnostic of a tetrahedral environment at the Cu^{I} metal center.⁵¹ Additionally, ESIMS analysis yields a peak corresponding to the M^{2+} ion at 1314.5 m/z , indicating formation of the desired condensed structure. Notably, compound **14b** can be converted into macrocycle **15b** upon the addition of stoichiometric quantities of pyridine. Once again, the $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum allows us to monitor this transformation through the appearance of a singlet at δ -8.0, which has shifted upfield from the resonance observed for **14b**. The ESIMS analysis of **15b** yields a peak at 1314.9 m/z which corresponds to the pyridine-free molecular ion.

X-ray Structure Determination of **15aCDABCO** and **15cCDABCO**

Similar to the problems encountered in crystallizing **8a**, our attempts to isolate X-ray quality single crystals of **14a** and **15a** proved unfruitful. Based upon the more promising strategy of adding DABCO to pure samples of **8a**, we hypothesized that crystals of **15aCDABCO** could be obtained in a similar fashion. X-ray quality crystals were obtained from vapor diffusion of pentane/diethyl ether (1:1 v/v) into a THF/ CH_2Cl_2 solution of **15a** containing DABCO, which was initially layered with acetonitrile. Similar to the analogous structure **8aCDABCO**, the 56-membered macrocyclic product **15aCDABCO** represents one of the largest macrocycles ever prepared via the Weak-Link Approach. In this structure, the Zn atoms are separated by a distance of distance of 7.02 Å and are bridged by a DABCO ligand (Figure 2). The structure of **15aCDABCO** is slightly puckered toward the Zn(porphyrin) centers, leading to a slightly staggered porphyrin–porphyrin geometry. As expected, the Rh–P and Zn–Zn distance in **8aCDABCO** are comparable to those found for **15aCDABCO** while the Rh–Rh distance differs significantly, presumably due to the presence of the four S atoms which have been incorporated in **15aCDABCO** (Table 2).

Contrary to **8b**, **15b** was quite soluble in pure THF; however, our efforts to crystallize **15b** from THF only yielded microcrystalline powder. Attempts to crystallize **15b** using the same conditions as used for **8b** led to the formation of magenta single crystals of **15cCDABCO**, which were confirmed by X-ray diffraction analysis at APS (see Supporting Information). Macrocyclic **15cCDABCO** encapsulates a bridging DABCO ligand between two Zn porphyrins, flanked by tetrahedral CuCl_2 centers, again indicating the oxidation of Cu^{I} to Cu^{II} via solvent dehalogenation as has been observed previously. As for **8cCDABCO**, isolation of **15cCDABCO** strongly suggests an open macrocyclic structure for **15b** as shown in Scheme 3.

Acyl Transfer Catalytic Experiments

To demonstrate the ability of the Zn^{II} –porphyrin moieties in **14a** and **15a** to act cooperatively in an allosterically controlled fashion, we employed a catalytic acyl transfer reaction that has been shown by Sanders and co-workers to accelerate in the presence of trimetallic Lewis acidic porphyrin assemblies.⁵⁶ We hypothesized that the pyridylcarbinol and acetylimidazole substrates could be brought together within the cavity of **15a** by cofacial Zn^{II} metal centers and converted to the products in a catalytic fashion by virtue of their proximity (Figure 3). Furthermore, reactions involving 1-acetylimidazole and differentially substituted *X*-pyridylcarbinol (where *X* = 2, 3, or 4) can be used to evaluate the ability of **15a** to act allosterically: only the combination of substrates with the right distance can span the cavity and react at an accelerated rate.

The efficiency of both the “closed” (**14a**) and “open” (**15a**) supramolecular catalysts in the acyl transfer reaction were evaluated against a control reaction consisting of the monomer **Zn** (**TPP**) and analogous monomeric Rh^{I} complexes **16a** and **16b** (see Supporting Information).

For 4-pyridylcarbinol (4-PC), both **14a** and **15a** significantly accelerate the reaction rate, with the open macrocycle **15a** being almost 14 times more active than the monomers and twice as fast as the closed macrocycle **14a** (Figure 4a). While **14a** is depicted as a rigid entity in Figure 3, its structure is probably dynamic when in solution and the observed catalytic activity may originate from the conformational flexibility around the S atoms and the low rotational barrier of the porphyrin ligand about the carbon-sulfur bond in the backbone of the ligand. For 3-pyridylcarbinol (3-PC), the background-corrected $\text{rate}_{\text{macrocycle}}/\text{rate}_{\text{monomers}}$ ratio ($\text{rate}_{\text{monomers}}$ = rate of the reaction in the presence of the $[\text{Rh}^{\text{I}} \text{ monomer} + \text{Zn}(\text{TPP})]$ mixture) and the observed allosteric effect ($\text{rate}_{\text{15a}}/\text{rate}_{\text{14a}}$) only drops slightly (Figure 4b), suggesting that the cavities of **14a** and **15a** are still flexible enough to accommodate the change in transition state distance for acyl transfer from acetylimidazole upon binding. For 2-pyridylcarbinol (2-PC), both the $\text{rate}_{\text{macrocycle}}/\text{rate}_{\text{monomers}}$ ratio and the observed allosteric effect ($\text{rate}_{\text{15a}}/\text{rate}_{\text{14a}}$) drop significantly with respect to 3- and 4-pyridylcarbinol and are similar to those observed for the monomers (Figure 4c). This may be explained by a simple geometry argument: if 2-pyridylcarbinol is bound to one of the Zn centers, the carbinol group will be pointed away from the imidazole *N*-acetyl group bound to the other side, resulting in an unfavorable transition state (in comparison to those for 4- and 3-pyridylcarbinol) for productive acyl transfer.

Our catalytic data offers strong evidence that Cl^-/CO can act as positive allosteric effectors in a Rh^{I} -based hemilabile supramolecular catalyst system, enhancing the efficiency of bimolecular acyl transfer reactions when the geometry is optimized. Significantly, these effectors operate cooperatively with each other and are incapable of effecting shape change if added independently to the Rh^{I} species. That 2-pyridylcarbinol can be selectively discriminated against 3- and 4-pyridylcarbinol provides an impetus for employing these supramolecular assemblies for chemical sensing and shape-selective recognition when coupled to catalytic processes that provide signal amplification.

Conclusion

In conclusion, we have developed a coordination chemistry-based synthetic approach for the quantitative preparation of flexible cofacial porphyrin assemblies in which the porphyrins act as functional sites within an allosteric framework that is tunable via modulation of peripheral structure control domains. Importantly, these architectures possess cavities whose sizes can be directly controlled *in situ* via the introduction of simple, cooperative ancillary ligands that bond to the structure control domains. This capability enables the cofacial porphyrin structures to act as allosteric catalysts capable of discriminating different substrate combinations and selectively transforming them into the desired products, two key steps in developing new biomimetic supramolecular systems. Most notably, our synthetic approach should allow for facile access to a range of systems with tunable cofacial porphyrin–porphyrin distances that are useful in studying distance-dependent electron-transfer phenomena, molecular switches, host–guest interactions, and catalysis. Efforts toward elaboration of these areas are currently underway.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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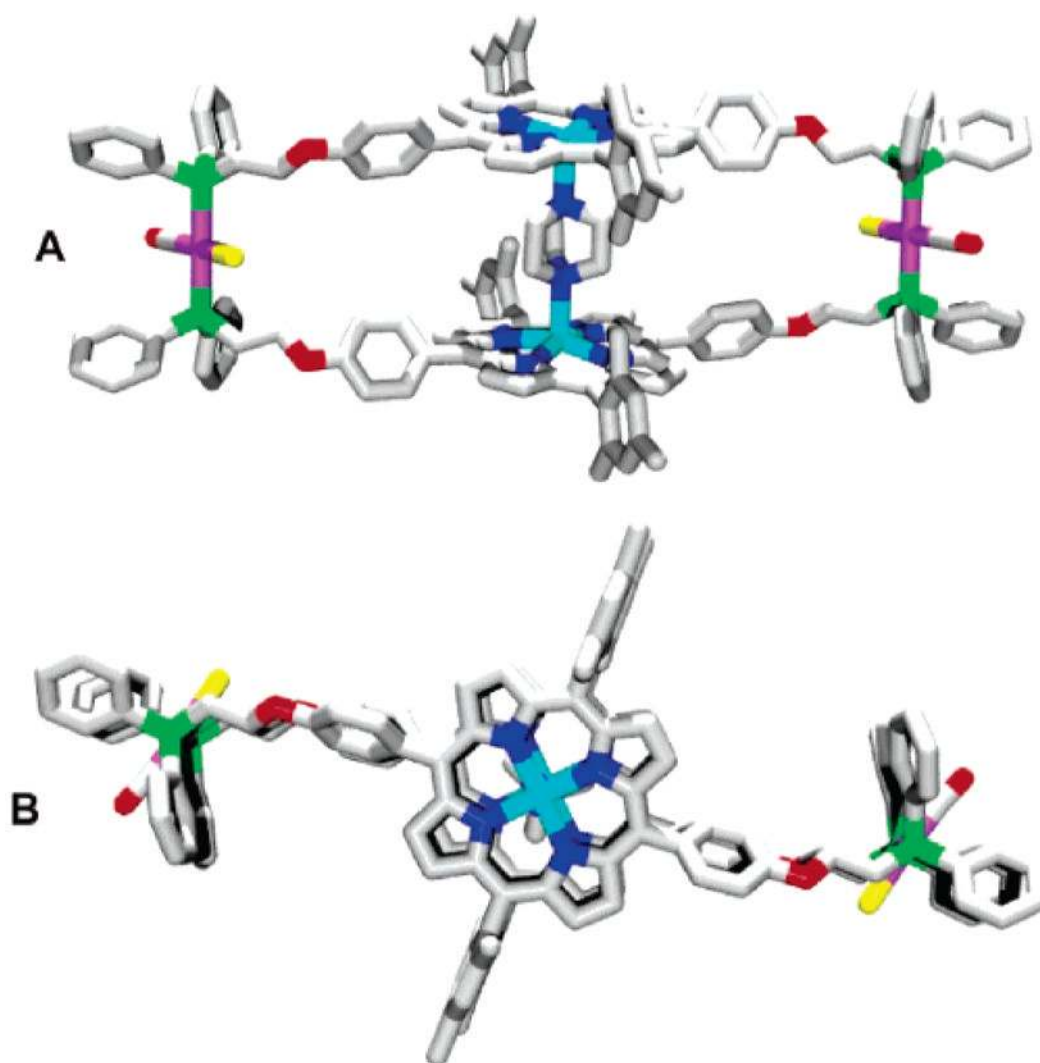


Figure 1.

Graphical representations of the X-ray crystal structure of **8aCDABCO** as viewed (A) from the side and (B) from the top containing a molecule of DABCO bridging both Zn atoms. Hydrogen atoms, disordered DABCO carbon atoms, and solvent molecules have been omitted for clarity. Pink = Rh, Red = O, Yellow = Cl, Green = P, Blue = N, Light Blue = Zn.

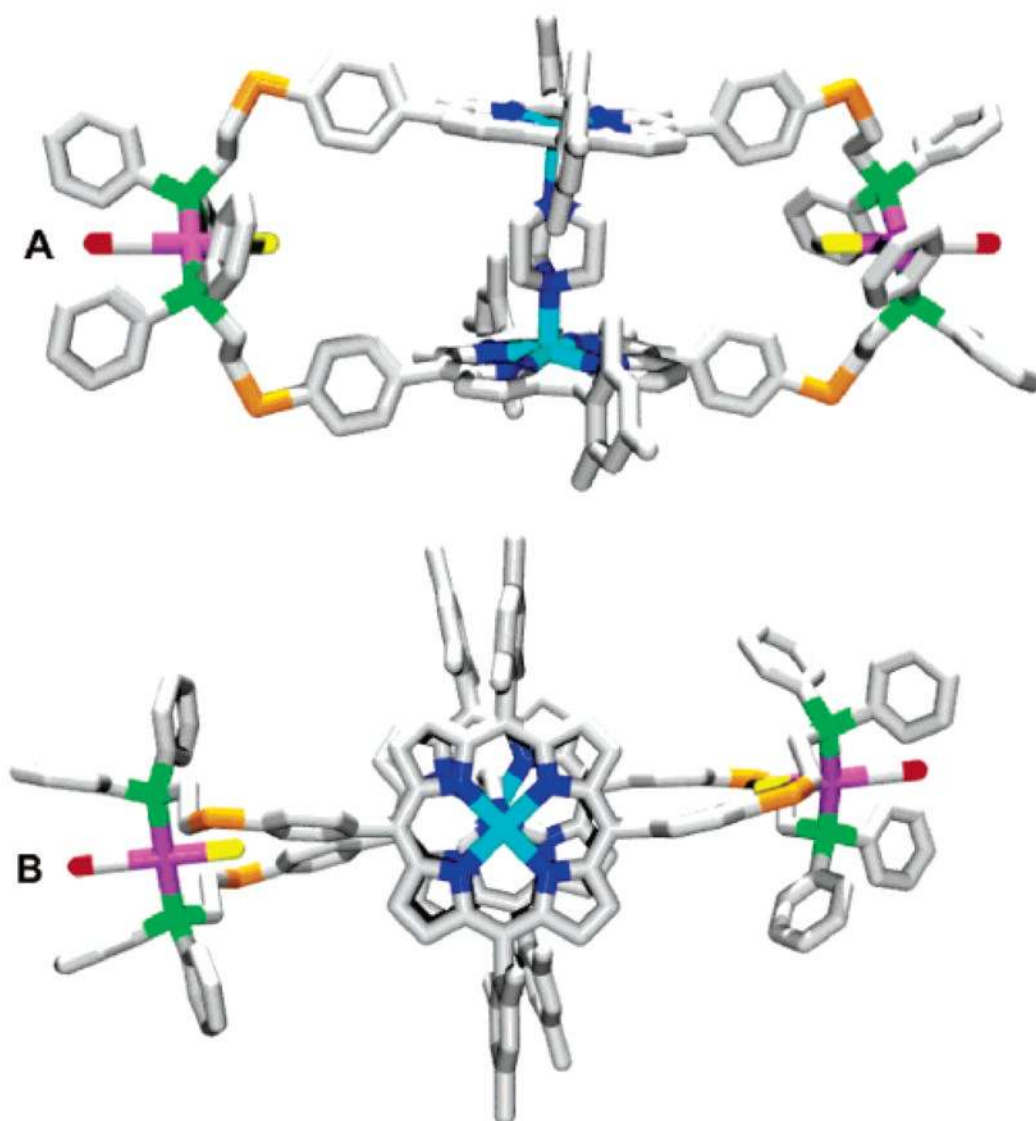


Figure 2. Graphical representations of the X-ray crystal structure of **15aCDABCO** as viewed (A) from the side and (B) from the top containing a molecule of DABCO bridging both Zn atoms. Hydrogen atoms, disordered DABCO carbon atoms and solvent molecules have been omitted for clarity. Gray = Carbon, Pink = Rh, Red = O, Orange = S, Yellow = Cl, Green = P, Dark Blue = N, Light Blue = Zn.

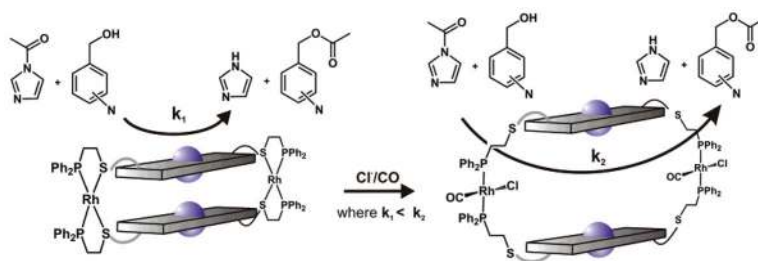


Figure 3. Acyl transfer reactions catalyzed by (left) a closed macrocycle vs (right) the corresponding open macrocycle. The open macrocycle can preorganize the substrates within the cavity, thereby increasing the rate of the reaction (k_2) in comparison to that (k_1) observed in the presence of the closed macrocycle.

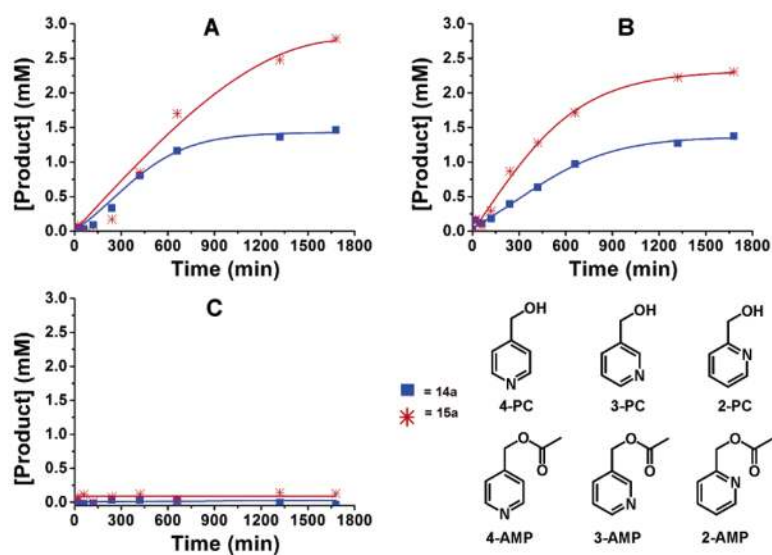
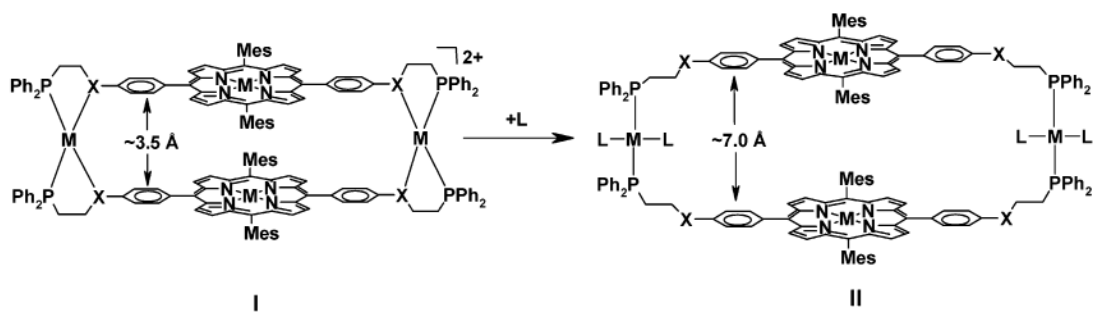
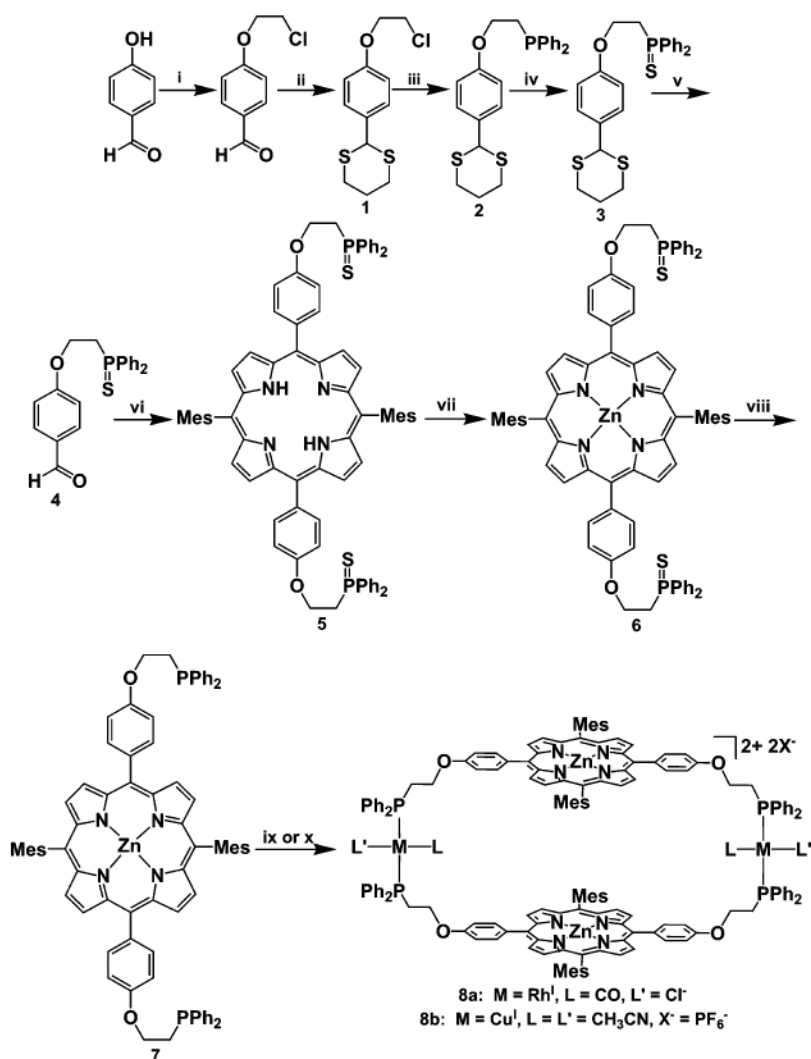


Figure 4. Formation of the three *X*-(acetoxymethyl)pyridine (*X* = 2, 3, or 4) isomers by an acyl transfer reaction between 1-acetylimidazole and *X*-pyridylcarbinol, as catalyzed by Zn-porphyrin complexes **14a** and **15a**. Concentration vs time plots are shown for the formation of 4-(acetoxymethyl)pyridine (**A**, 4-AMP), 3-(acetoxymethyl)pyridine (**B**, 3-AMP), and 2-(acetoxymethyl)pyridine (**C**, 2-AMP). All data were corrected for background reactions (see Supporting Information). Conditions: CH₂Cl₂, rt, 9 mM *X*-pyridylcarbinol, 6 mM 1-acetylimidazole, 2.5 mM biphenyl (GC reference standard), and 0.3 mM supramolecular catalyst (**14a** and **15a**). CO (1 atm) and appropriate amounts of benzyltriethylammonium chloride when indicated.



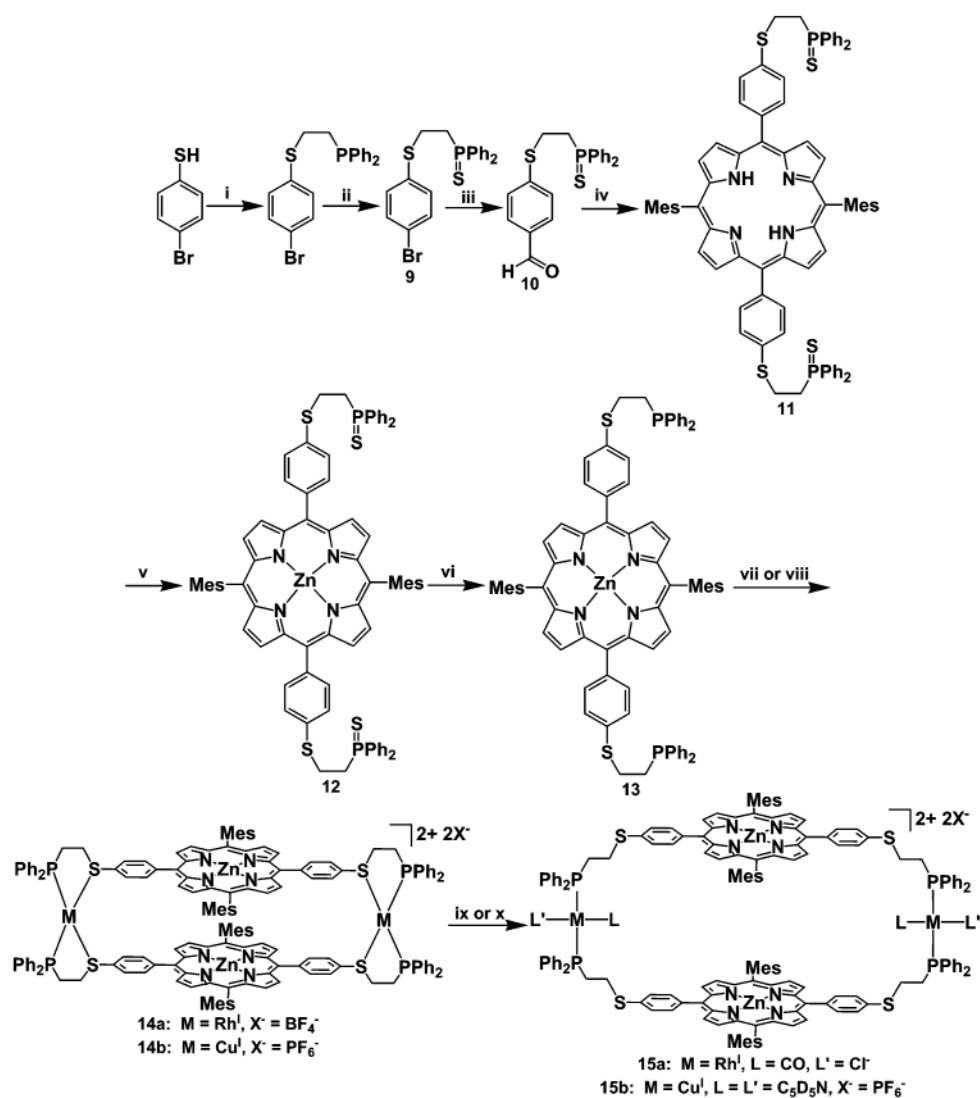
Scheme 1. Design of Allosteric Porphyrin-Based Supramolecules Whose Cavity Sizes Can Be Modified by the Binding of a Ligand L^a

^a **I**: Condensed Macrocycle, **II**: Open Macrocycle. PPh₂ = diphenylphosphine and MES = 1,3,5-trimethylbenzene.



Scheme 2. Synthesis of Ether-Based Ligand 7 and Macrocycles 8a and 8b^a

^a (i) 1-bromo-2-chloroethane, K_2CO_3 , Acetone, Reflux; (ii) 1,3-propanedithiol, $\text{Y}(\text{OTf})_3$ (5 mol %), CH_3CN ; (iii) KPPH_2 , THF; (iv) S_8 , THF; (v) NaNO_2 , $\text{AcCl}/\text{H}_2\text{O}$, CH_2Cl_2 , $0^\circ\text{C} \rightarrow \text{rt}$; (vi) 5-mesityldipyrrromethane, $\text{BF}_3 \cdot \text{OEt}_2$, DDQ, NEt_3 , CHCl_3 , 4 Å Molecular Sieves; (vii) $\text{Zn}(\text{OAc})_2 \cdot 2\text{H}_2\text{O}$, 4:1 $\text{CHCl}_3/\text{MeOH}$, Reflux; (viii) Cp_2ZrHCl , THF, 60°C ; (ix) $[\text{Rh}(\text{CO})_2(\text{Cl})]_2$, $\text{CH}_2\text{Cl}_2/\text{THF}$; (x) $[\text{Cu}(\text{CH}_3\text{CN})_4]\text{PF}_6$, $\text{CH}_2\text{Cl}_2/\text{THF}$.



Scheme 3. Synthesis of Thioether-Based Ligand 13 and Macrocycles 14a–b, 15a–b^a

^a (i) $\text{ClCH}_2\text{CH}_2\text{PPh}_2$, Cs_2CO_3 , CH_3CN , Reflux; (ii) S_8 , THF; (iii) $n\text{-BuLi}$, DMF, THF, -78°C ; (iv) 5-mesityldipyrrromethane, $\text{BF}_3 \cdot \text{OEt}_2$, DDQ, NEt_3 , CHCl_3 , 4 Å Molecular Sieves; (v) $\text{Zn}(\text{OAc})_2 \cdot 2\text{H}_2\text{O}$, 4:1 $\text{CHCl}_3/\text{MeOH}$, Reflux; (vi) Cp_2ZrHCl , THF, 60°C ; (vii) for **14a**: $[\text{Rh}(\text{NBD})\text{Cl}]_2$, AgBF_4 , $\text{CH}_2\text{Cl}_2/\text{THF}$; (viii) for **14b**: $[\text{Cu}(\text{CH}_3\text{CN})_4]\text{PF}_6$, $\text{CH}_2\text{Cl}_2/\text{THF}$; (ix) for **15a**: PPNCl/CO (1 atm); (x) for **15b**: $\text{C}_5\text{D}_5\text{N}$.

Table 1
X-ray Crystallographic Data for **8aCDABCO** and **15aCDABCO**

	8aCDABCO	15aCDABCO
empirical formula	C ₁₈₁ H ₁₆₆ Cl ₄ N ₁₂ O ₈ P ₄ Rh ₂ Zn ₂	C ₁₇₂ H ₁₄₂ Cl ₂ N ₁₀ O ₄ P ₄ Rh ₂ S ₄ Zn ₂
formula weight	3239.5	3072.54
Temperature	153(2) K	100(2) K
Wavelength	0.71073 Å	0.48595 Å
crystal system, space group	triclinic, <i>P1</i>	monoclinic, <i>C2/c</i>
unit cell dimensions	<i>a</i> = 13.9395(13) Å <i>α</i> = 83.044(2)° <i>b</i> = 17.1623(17) Å <i>β</i> = 73.368(2)° <i>c</i> = 20.894(2) Å <i>γ</i> = 74.250(2)°	<i>a</i> = 17.0086(10) Å <i>b</i> = 44.661(3) Å <i>β</i> = 104.252(2)° <i>c</i> = 23.0053(13) Å
volume	4604.5(8) Å ³	16937.3(17) Å ³
Z, calculated density	1, 1.168 Mg/m ³	4, 1.205 Mg/m ³
absorption coefficient	0.581 mm ⁻¹	0.342 mm ⁻¹
<i>F</i> (000)	1678	6336
crystal size	0.170 × 0.170 × 0.50 mm	0.150 × 0.025 × 0.010 mm
theta range for data collection	1.02 to 28.83°	0.90 to 15.70°
reflections collected/unique	42335/21291 [<i>R</i> (int) = 0.0712]	123297/12229 [<i>R</i> (int) = 0.0780]
absorption correction	integration	empirical
max. and min. transmission	0.9695 and 0.8265	1.000000 and 0.649500
refinement method	full-matrix least-squares on <i>F</i> ²	full-matrix least-squares on <i>F</i> ²
data/restraints/parameters	21291/0/915	12229/0/921
goodness-of-fit on <i>F</i> ²	0.912	1.063
final <i>R</i> indices [<i>I</i> > 2σ(<i>I</i>)]	<i>R</i> 1 = 0.0910, <i>wR</i> 2 = 0.2301	<i>R</i> 1 = 0.0937, <i>wR</i> 2 = 0.2816
<i>R</i> indices (all data)	<i>R</i> 1 = 0.1924, <i>wR</i> 2 = 0.2591	<i>R</i> 1 = 0.1199, <i>wR</i> 2 = 0.3081
largest diff. peak and hole	1.378 and -0.747 e ⁻ /Å ⁻³	2.236 and -0.800 e ⁻ /Å ⁻³

Table 2
Selected Distances for **8aCDABCO** and **15aCDABCO**

selected distance	8aCDABCO (Å)	15aCDABCO (Å)
Rh-Rh	24.8	22.5
Rh-P	2.32, 2.31	2.30
Zn-Zn	7.09	7.02