



Published in final edited form as:

Nat Rev Cancer. 2015 September ; 15(9): 563–572. doi:10.1038/nrc3978.

Sympathetic nervous system regulation of the tumour microenvironment

Steven W. Cole^{1,*}, Archana S. Nagaraja², Susan K. Lutgendorf³, Paige A. Green⁴, and Anil K. Sood²

¹Department of Medicine, Division of Hematology-Oncology, Geffen School of Medicine, UCLA Molecular Biology Institute, Norman Cousins Center, and Jonsson Comprehensive Cancer Center, University of California, Los Angeles

²Departments of Gynecologic Oncology and Cancer Biology, University of Texas M. D. Anderson Comprehensive Cancer Center

³Departments of Psychology, Obstetrics and Gynecology, Urology, and Holden Comprehensive Cancer Center, University of Iowa

⁴Basic Biobehavioral and Psychological Sciences Branch, Behavioral Research Program, Division of Cancer Control and Population Sciences, United States National Cancer Institute

Abstract

The peripheral autonomic nervous system (ANS) is known to regulate gene expression in primary tumours and their surrounding microenvironment. Activation of the sympathetic division of the ANS in particular modulates gene expression programs that promote metastasis of solid tumours by stimulating macrophage infiltration, inflammation, angiogenesis, epithelial-mesenchymal transition, and tumour invasion, and by inhibiting cellular immune responses and programmed cell death. Haematological cancers are modulated by sympathetic nervous system (SNS) regulation of stem cell biology and hematopoietic differentiation programs. In addition to identifying a molecular basis for physiologic stress effects on cancer, these findings have also identified new pharmacologic strategies to inhibit cancer progression *in vivo*.

Introduction

The sympathetic nervous system (SNS) regulates the function of virtually all human organ systems via localized release of catecholamine neurotransmitters from sympathetic nerve terminals and via systemic circulation of catecholamines from the adrenal gland (Box 1) ^{1–3}. Physiologists have long focused on the effects of acute ‘fight-or-flight’ stress spikes in SNS activity that transiently enhance bodily strength, mobility, perceptual acuity, and tissue defense at the expense of long-term trophic activities such as digestion, reproduction, growth, and exploration ^{1–3}. However, long-term variations in basal levels and circadian cycles of SNS activity can also exert more enduring regulatory effects by altering constitutive gene expression profiles in a wide variety of tissues and organ systems ^{4–11}.

*Correspondence: Steve.Cole@ucla.edu, Department of Medicine/Hematology-Oncology, 11-934 Factor Bldg., UCLA School of Medicine, Los Angeles CA 90095-1678.

These SNS-modulated transcriptional dynamics stem from evolutionarily conserved molecular mobility and defense programs (MMDPs) that adapt a broad range of cellular functions to detect and respond more effectively to challenging, threatening, or novel environments (e.g., mobilizing energy and promoting mobility, strength, perceptual acuity, antimicrobial defenses, and wound healing)^{3,10–12}. In the past decade, it has become apparent that many MMDPs also promote tumour progression and metastasis (reviewed in^{11,13–16}). More recent pharmaco-epidemiologic studies have linked β -adrenergic antagonists to reduced progression of incident tumours^{14,17}, implying that SNS signaling may potentially exert clinically significant effects on tumour biology. This Opinion article surveys some of the key physiological pathways and molecular dynamics involved in SNS regulation of tumour progression, highlights some of the ensuing translational therapeutic opportunities, and outlines critical issues for future research.

Box 1

Sympathetic nervous system signaling

The sympathetic nervous system (SNS) regulates gene expression and cellular function in the nervous, endocrine, cardiovascular, gastrointestinal, respiratory, reproductive, and immune systems by releasing 2 catecholamine neuroeffector molecules:

Norepinephrine is released from SNS nerve fibers that traverse the body to directly innervate target tissues.

Epinephrine is released from the adrenal gland and circulates via blood to target tissues. These adrenergic effector molecules regulate cellular function via 5 adrenergic receptor types that are differentially expressed across target tissues and couple to distinct G protein-mediated signal transduction pathways:

α_1 adrenergic receptors are expressed mainly in smooth muscles and signal through $G_{\alpha q}$ induction of phospholipase C to activate calcium flux and protein kinase C (PKC).

α_2 adrenergic receptors are expressed on smooth muscles and platelets, as well as on neurons where they function as autoreceptors to inhibit norepinephrine release. α_2 receptors signal through $G_{\alpha i}$ inhibition of cyclic 3'-5' adenosine monophosphate (cAMP) activity, which in turn down-regulates the serine-threonine protein kinase A (PKA), the guanine exchange protein activated by adenylyl cyclase (EPAC), and β -arrestin-mediated activation of mitogen-activated protein kinases (MAPK).

β_1 adrenergic receptors enhance cardiac output, mediate neural signaling, and mobilize energy from adipose tissue. They signal through $G_{\alpha s}$ -mediated activation of cAMP, which stimulates PKA, EPAC, and MAPK signaling.

β_2 adrenergic receptors are expressed on smooth muscles of the heart and lung, where they mediate vasodilation, on immune cells, where they mediate cell trafficking and effector activities, and on many tumor cells of epithelial and lymphoid origin. They signal through $G_{\alpha s}$ /cAMP stimulation of PKA, EPAC and MAPK.

β_3 adrenergic receptors are predominately expressed in adipose tissue, where they mobilize energy by signaling through $G_{\alpha s}$ /cAMP stimulation of PKA, EPAC and MAPK.

Each adrenergic pathway stimulates transcription of distinct molecular mobility and defense programs (MMDPs) in characteristic target cells via post-translational activation of transcription factors such as the cAMP response element-binding protein (CREB).

SNS regulation of gene expression

The SNS regulates systemic physiology via two general signaling pathways: one involving direct innervation of target organs throughout the body by SNS nerve fibers that release the sympathetic neurotransmitter norepinephrine, and a second one involving hormonal regulation of organ systems via vascular distribution of epinephrine released from the adrenal gland^{1-3,11} (Box 1 and Figure 1).

Acute SNS activation

Perceptions of acute threat mediated by the central nervous system (CNS) can activate the splanchnic nerves — which stimulate rapid release of pre-synthesized epinephrine (and smaller amounts of norepinephrine) from chromaffin cells of the adrenal medulla¹⁻³. Epinephrine levels in plasma can spike by >10-fold during acute fight-or-flight stress responses, leading to rapid physiological changes in cardiovascular, respiratory, muscular, metabolic, neural, immune, and other functions that typically return to baseline within 20–60 minutes following the abatement of perceived threat^{1,2,18}. These rapid physiological alterations generally involve post-translational modifications of protein function that are mediated by activation of two broad classes of adrenergic receptors, α and β , each of which contains multiple receptor subtypes that are differentially distributed across tissue sites and linked to distinct signal transduction pathways to induce distinct molecular effects¹⁻³ (Box 1). For example, acute fight-or-flight responses increase heart rate and beat strength by activating β_1 -adrenergic receptors in the heart muscle, redistribute blood from superficial tissues to long muscles by activating vascular α_1 - and β_2 -adrenergic receptors, increase respiratory rate and depth by activating bronchial α_1 - and β_2 -adrenergic receptors, mobilize energy by activating β_2 - and β_3 -adrenergic receptors in adipose tissue and the liver, and mobilize leukocytes (especially natural killer (NK) cells) into circulation by activating β_2 -adrenergic receptors on leukocytes¹⁻³.

Activation of sympathetic nerve fibers that directly innervate most of the body's organ systems also has a significant role in acute fight-or-flight stress reactions. This occurs through direct regulation of organ function by μ molar concentrations of norepinephrine released from sympathetic nerve terminals, and by circulation of norepinephrine that spills over from sympathetic innervation of smooth muscles surrounding blood vessels (the primary source of norepinephrine in plasma)¹⁻³ (Figure 1).

Circadian and chronic changes in SNS activity

In addition to the acute fight-or-flight dynamics that are mediated via rapid post-translational modification of protein function, variations in chronic SNS activity can also modulate gene expression and cellular structure^{8,11}. Many of these effects are mediated by transcriptional activation of MMDPs that evolved to prepare the body to respond to physical demands, cognitive challenges, and wounding injury^{1-3,9,10,12}. Unlike other stress-activated neuro-

endocrine systems such as the hypothalamus-pituitary-adrenal axis, the SNS is easily activated by the mere anticipation of threat^{1,18}. For example, first-time parachute jumpers show peak plasma levels of epinephrine, norepinephrine, and target organ cardiovascular responses before they actually commence their first fall (that is, in response to anticipation of the fall, rather than the fall itself)¹⁸. Anticipatory SNS responses can often occur at thresholds below those consciously experienced as fear, stress, or injury. Moreover, unlike the hypothalamus-pituitary-adrenal axis, SNS stress responses do not decay over time with repeated threat exposure^{1,19}. Chronic or repetitive low-grade SNS activation up-regulates norepinephrine levels more strongly than hormonal epinephrine levels (e.g., in post-traumatic stress disorder²⁰), and is commonly observed in people who are chronically exposed to adverse social environments (e.g., poverty, isolation, combat, and demanding or uncontrollable jobs)¹. Experimental studies in animal models have shown that chronic social stress can also increase the growth and branching of sympathetic nerve fibers in peripheral tissues (neo-innervation), and thereby up-regulate basal activity of target tissue adrenergic receptors and downstream MMDPs^{5,21}.

Low-grade SNS neural activation (and to a lesser extent, adrenal epinephrine responses) also occurs in response to many ‘non-threat’ homeostatic challenges such as temperature change, general bodily movement, sleep disturbance, physical exertion, speaking, and intense concentration or vigilance^{1,3,22–24}. These activity-related dynamics induce a substantial circadian rhythm in SNS activity, which peaks during waking hours and reaches a nadir during periods of extended rest or sleep^{8,22,23,25}. Like chronic low-grade SNS activation, circadian variations in SNS activity exert many of their effects via adrenergic receptor-mediated modulation of gene transcription in target tissues^{8,25}.

Chronic and circadian variations in SNS activity have recently been found to play a major part in regulating constitutive gene expression in a wide range of SNS target tissues. Many of the activated MMDPs involve suppression of long-term growth and maintenance processes and up-regulation of molecular programs that facilitate physical mobility (such as cardiac output, respiration, and glucose mobilization), mental acuity (such as CNS perceptual and mnemonic processes), and response to wounding injuries (such as shock and acute phase responses, inflammation, and wound healing)^{1,2}. In the immune system, for example, tonic SNS activity mobilizes hematopoietic stem cells out of their bone marrow niches and into circulation, where they subsequently transit to peripheral tissues such as the spleen (to facilitate extramedullary hematopoiesis) or sites of injury (to facilitate inflammation and localized tissue remodeling)^{4,8,11,25,26}. Within the bone marrow hematopoietic environment, SNS signaling transcriptionally stimulates the development of monocytes, granulocytes, and other myeloid lineage immune cells at the expense of lymphoid and erythroid lineages, resulting in a pro-inflammatory shift in the overall immunoregulatory set-point of the circulating leukocyte pool^{7,27}. SNS innervation of secondary lymphoid tissues — such as the spleen and lymph nodes — also modulates the evolution of peripheral immune responses, for example by down-regulating lymphocyte trafficking to tissues²⁸, by inhibiting transcription of Type I (α and β) and Type II (γ) interferons, and by stimulating transcription of cytokines expressed in type 2 T helper cells (Th2) and Th17 cells, all of which act to impair innate anti-viral responses and promote humoral immune responses at the expense of cellular immunity^{5,12,29–31}. Adrenergic

and Rad3-related (ATR)/p21 pathway⁶² and β -arrestin-induced activation of the AKT signaling pathway, which stimulates the E3 ubiquitin ligase murine double minute 2 (MDM2) to degrade p53 protein and thereby inhibit p53-mediated responses to chromosomal damage⁶⁰. These effects are sufficient to increase the prevalence of spontaneous chromosomal aberrations in tissues such as the thymus and brain, and such effects can be efficiently blocked by the β -adrenergic antagonist propranolol^{60,61}. Similar effects are observed in neuroblastoma cells, in which propranolol up-regulates p53 levels, promotes apoptosis, and sensitizes tumour cells to the effects of the topoisomerase inhibitor SN-38⁵⁴. However, it is not yet clear whether β -adrenergic inhibition of DNA damage repair is sufficient to increase the rate of spontaneous tumour initiation *in vivo*.

Oncogene activation

β -adrenergic signaling can stimulate several oncogenic signaling pathways including Src⁶³ and HER2 (encoded by the *ERBB2* gene)^{64,65}. In the case of HER2, catecholamine activation of β -adrenergic receptors activates signal transducer and activator of transcription 3 (STAT3), which subsequently activates the *ERBB2* promoter to stimulate gene transcription⁶⁴. In the case of SRC, β -adrenergic signaling stimulates protein kinase A (PKA) to phosphorylate SRC on residue Y419, resulting in SRC-mediated activation of a complex phosphoproteomic network that stimulates tumour growth, migration, and invasion *in vivo*⁶³. In addition to these effects on cellular oncogenes, β -adrenergic signaling can also activate a diverse array of oncogenic viruses¹³. As one example, catecholamine induction of β -adrenergic signaling on B lymphoid cells triggers PKA-mediated activation of the cellular transcription factor cyclic 3'-5' adenosine monophosphate response element binding protein (CREB), which in turn activates a key viral promoter in the episomal Kaposi's Sarcoma-associated Human Herpesvirus 8 (HHV-8) genome⁶⁶. As a result, the viral genome up-regulates expression of the viral transcription factor Rta, which serves as a master regulator of HHV-8 gene expression and stimulates transcription of a diverse array of viral oncogenes and dissemination of the viral genome⁶⁶. HHV-8 contributes to several types of B cell malignancy in addition to Kaposi's Sarcoma of vascular endothelial cells⁶⁷, providing a virally mediated pathway by which SNS activation can contribute to human cancer.

Inflammation and immune response

β -adrenergic signaling also stimulates the transcription of pro-inflammatory cytokines such as IL-6 and IL-8 by tumour cells^{32,68-70} by myeloid lineage immune cells in the tumour microenvironment^{32,49}. Adrenergic stimulation of inflammatory arachidonic acid metabolism can also promote tumour growth⁷¹. Several *in vivo* studies have shown that SNS stimulation of inflammatory signaling can enhance tumour progression and metastasis^{49,72}. However, no studies have yet determined whether SNS effects on inflammation are sufficient to increase rates of tumour initiation.

Macrophages play a key part in mediating inflammation, modulating the tumour microenvironment, and promoting metastasis. β -adrenergic signaling can markedly enhance macrophage recruitment into the tumour parenchyma by stimulating tumour cells' production of chemotactic factors such as macrophage colony stimulating factor (CSF1, also known as M-CSF) and MCP-1^{49,72}. β -adrenergic signaling may also enhance the density of

tumour-associated macrophages by stimulating myelopoietic development of precursor monocytes in the bone marrow^{7,27} and spleen²⁶, which can then be recruited into the tumour microenvironment (Figure 1). Within the tumour microenvironment, β -adrenergic signaling also stimulates macrophage expression of gene programs that promote tumour progression, including transforming growth factor β (*TGFB*), vascular endothelial growth factor (*VEGF*), IL-6, matrix metalloproteinase 9 (*MMP9*), and *PTGS2*. β -adrenergic regulation of macrophage biology has a major role in SNS-induced metastasis as pharmacologic inhibition of CSF1 or MCP-1 signaling can abrogate stress effects on metastasis *in vivo*^{49,72}.

In contrast to its stimulatory effects on inflammation and macrophage biology, SNS signaling can profoundly inhibit the transcription of the Type I and Type II interferons^{49,73,74} that play critical parts in generating cell-mediated immune responses against cancers and tumour-associated viruses. β -adrenergic signaling can also suppress the cytotoxic function of T lymphocytes and NK cells⁵⁹, and these effects contribute to the increased cancer cell dissemination observed during surgery^{56,75,76}. However, several lines of research in severe combined immunodeficiency disease (SCID) and nude mice have found that the SNS can promote tumour metastasis even in the absence of NK and cytotoxic T cells^{35,49,52,77}. As such, neural modulation of cellular immune responses may contribute to SNS influences on tumour progression in some contexts, but it is not an essential mediator of stress effects on cancer progression *in vivo*.

Mesenchymal activation and EMT

Adrenergic signaling can activate a wide range of mesenchymal cell types present in tumour stroma (such as fibroblasts, pericytes, and mesenchymal stem cells)^{33,34,36,78,79}, as well as adipocytes^{80,81} and bone marrow mesenchymal cells that can indirectly regulate cancer biology by altering haematopoiesis of tumour-infiltrating immune cells or modulating the cancer stem cell niche (Figure 2)^{11,82}. Emerging data suggest that β -adrenergic activation of SNAIL family of transcription factors may also promote the expression of mesenchymal gene expression programs in epithelial tumors, promoting EMT, (S. Cole, S. Lutgendorf, and A. Sood, unpublished observations), and thereby modulate pro-metastatic processes such as tumour cell motility^{51,63,83,84} and matrix metalloproteinase-mediated invasion of basement membrane^{63,85–87}. Research is still defining the molecular mediators involved in EMT dynamics and defining their contribution to SNS effects on metastasis *in vivo*.

Angiogenesis

β -adrenergic signaling stimulates the expression of angiogenic growth factors such as VEGF and IL-6^{34,35,68,87–91}, which catalyze the development of vasculature to support tumour growth and metastasis. Studies using pharmacologic and genetic inhibitors of angiogenesis have confirmed that SNS-induced up-regulation of angiogenesis mediates stress effects on tumour growth and metastasis *in vivo*³⁵.

Survival and programmed cell death

β -adrenergic signaling can modulate a wide variety of growth and survival pathways, including inhibition of anoikis (programmed cell death induced by anchorage-dependent

cells detaching from the surrounding extracellular matrix) mediated by focal adhesion kinase (FAK, also known as *PTK2*)⁹², inhibition of apoptotic responses to chemotherapy mediated by the BCL2-associated agonist of cell death (*BAD*) and p53 pathways^{52–54,93}. β -adrenergic signaling can also modulate the expression of growth and survival factors —such as VEGF, IL-6, and IL-8— that are associated with resistance to tyrosine kinase inhibitors^{91,94}. Several of these studies have documented consequent impacts on tumour growth and metastasis *in vivo*, showing that inhibition of programmed cell death represents a bona fide mediator of SNS effects on cancer progression.

Haematopoiesis

β -adrenergic signaling can promote the growth and dissemination of acute lymphocytic leukaemias *in vivo*^{58,59}, likely via some of the same molecular pathways through which the SNS regulates physiological hematopoiesis^{4,7,11,25–27,95}. Sympathetic innervation of the bone marrow also helps maintain normal stromal cell populations, and may facilitate immunologic recovery following haematopoietic stem cell transplant^{11,96}. However, SNS regulation of the bone marrow hematopoietic environment is complex and can have unpredictable effects that depend greatly on the specific biological interactions between tumour cell biology and the bone marrow microenvironment¹¹. In a model of acute myelogenous leukemia (AML), for example, β -adrenergic antagonists unexpectedly accelerated disease by disrupting the haematopoietic stem cell niche in ways that favoured leukemia stem cell growth^{11,82}. AML colonization of the marrow itself produced similar effects by degrading sympathetic innervation and consequently stimulating leukaemia stem cell growth^{11,82}. Beyond the context of leukaemia, little is known about potential SNS influences on lymphomas and other haematological malignancies.

In summary, a growing body of experimental research has identified specific cellular and molecular mechanisms through which SNS activation can accelerate the progression of diverse tumor types. The general pattern of effects (and non-effects) observed in the experimental literature is broadly consistent with the pattern of relationships observed in epidemiologic studies of stress and cancer^{13,38}, and pharmacologic β -blockers and cancer^{14,17}: SNS activation exerts its most pronounced effects in the early stages of tumour progression as primary tumours interact with the surrounding microenvironment to initiate dissemination and colonization of distant tissues. There is so far little *in vivo* evidence that SNS activation has a significant role in the earlier stage of tumour initiation or that it can significantly affect the subsequent course of already disseminated metastatic disease. However, the experimental literature provides some occasional exceptions to this general pattern. One line of research has shown a paradoxical protective effect of SNS activation in which β -adrenergic signaling altered white adipose tissue production of circulating adipokines such as leptin, which subsequently inhibited the growth of leptin-dependent distant tumors^{80,81}. Another found a protective effect of β -adrenergic signaling on AML progression stemming from SNS maintenance of the bone marrow hematopoietic niche^{11,82}. However, the vast majority of the extant experimental literature indicates that SNS activity generally promotes tumour progression through a pleiotropic array of molecular alterations in the primary tumour microenvironment. This opens up basic questions about the

physiological pathways through which the SNS communicates with the tumor microenvironment.

SNS signaling to the tumour microenvironment

The SNS is a network of distinct neural signaling pathways that share some degree of common regulation by the CNS, but are also subject to independent regulatory input from the target tissues they innervate and from distinct regions of the brain⁸¹. For example, the brain structures that regulate gene expression in adipose tissue are different from those that mediate fight-or-flight control of the cardiovascular system or adrenal medulla⁸¹. SNS innervation of some target tissues — such as the adrenal gland and white adipose tissue — can also stimulate the release of hormones, such as epinephrine or leptin, which are biochemically distinct from the primary norepinephrine signal released by SNS neurons and can circulate more diffusely throughout the body⁸¹. Many SNS nerve fibers also release small amounts of other signaling molecules in tandem with norepinephrine, such as the SNS “co-transmitter” neuropeptide Y¹. Given the diverse array of physiologic mediator pathways that could potentially regulate tumour biology, research begun to map several of the general “nervous system-side” pathways for SNS regulation of tumor progression:

Hormonal catecholamines

Blood supply and tissue perfusion are essential for tumor growth and progression, and also provide a channel through which SNS catecholamines can access tumor tissue^{6,97}. However, clinical studies have so far failed to identify any substantial association between catecholamine levels in plasma and differential gene expression in tumours (even when tumour gene expression profiles are clearly associated with psychological risk factors such as depressive symptoms and low social support⁶). It also remains unclear how readily circulating epinephrine or norepinephrine might penetrate into the parenchyma of solid tumours to exert regulatory effects. Blood-based circulation of catecholamines has played a central role in physiologic conceptions of the fight-or-flight response, but this signaling pathway does not so far seem to constitute a major proximal pathway by which the SNS modulates tumour biology.

Tumour innervation

Although long overlooked by pathologists, many solid tumors receive direct innervation from the SNS. The most common pattern involves sympathetic nerve fibers that enter the tumour in association with the vasculature and occasionally radiate fibers into the tumour parenchyma^{11,32,77,98}. SNS fibers may also infiltrate into the outer perimeter of a growing tumour mass from surrounding healthy tissue^{77,98}, possibly in response to tumour cell expression of neurotrophic factors¹¹. Some experimental data suggest that tumours can actively promote the growth and branching of nerve fibers and may even stimulate development of new neurons (neurogenesis) via the expression of neurotrophic factors such as NGF, BDNF, semaphorins, netrins, and slit molecules^{11,98,99} (A. Sood, unpublished observations). SNS activation may also reciprocally attract tumour cells to neural fibers by up-regulating expression of trophic factors and chemokines such as CXCL12^{11,98–101}. Local sympathetic innervation seems to supply much of the catecholamine content within tumour

tissues because intra-tumor norepinephrine levels are generally higher than (and largely uncorrelated with) blood levels of norepinephrine or epinephrine^{6,97}. Haematological cancers are also subject to regulation by SNS nerve fibers that innervate the bone marrow hematopoietic niche and all other primary and secondary lymphoid organs. Sympathetic innervation of lymphoid tissues and the vasculature regulates cell trafficking and gene expression profiles in both developing progenitor cells and mature leukocytes^{4,7,25,26,27} (Hanoun, 2015 #2226,95).

Indirect hormonal and cellular regulation

SNS activation can also modulate tumour biology via indirect pathways in which SNS innervation of distant tissues triggers secondary hormonal or cellular effects that subsequently affect the tumour microenvironment. For example, SNS signaling to white adipose tissue can suppress circulating leptin levels and thereby inhibit the growth of leptin-sensitive tumours^{80,81}. SNS innervation of bone marrow can also stimulate the production of monocytes, neutrophils, and other myeloid lineage immune cells^{7,26}, which may then transit to the tumour microenvironment and promote metastasis⁴⁹ (Figure 1). Activated macrophages can also synthesize catecholamines under some circumstances¹⁰² and may thus provide a local non-neuronal source of adrenergic signaling within the tumour microenvironment. SNS innervation may also modulate the bone marrow microenvironment to make it a more receptive niche for metastatic colonization¹⁰³ or tumour cell growth and dissemination^{11,82}.

Therapeutic implications

Given the multiple physiologic pathways by which SNS signaling can reach the tumour microenvironment and its pleiotropic effects on tumour biology, pharmacologic antagonism of β -adrenergic signaling might represent a highly leveraged therapeutic opportunity with the potential to favorably impact a wide range of tumor, microenvironmental, and systemic mechanisms of cancer progression. Consistent with that concept, several observational epidemiologic studies have documented associations between β -blocker exposure and reduced progression of prostate^{47,48,104}, breast^{39–41,45}, lung^{44,105}, and ovarian cancer^{106,107}, as well as malignant melanoma^{42,43,46}. However, the epidemiological literature is also inconsistent, and some studies fail to find any evidence of a protective effect (likely due to methodological variations considered below). In experimental animal models of human cancer, β -antagonists can inhibit the progression of prostate^{51,52}, breast^{49,50,103}, ovarian³⁵, lung^{108,109}, pancreatic^{24,57}, and colon cancer¹¹⁰, neuroblastomas^{53,54}, and leukaemia^{58,59}. β -blockade can also inhibit systemic SNS influences on cancer progression such as haematopoietic production of pro-metastatic monocytes^{7,27,49} and bone marrow receptivity to metastatic colonization^{11,103}. β -blockers seem to provide a viable pharmacologic strategy for simultaneously inhibiting many of the pathways through which the SNS can stimulate tumour progression¹⁴.

Despite the availability of safe, approved, and inexpensive β -antagonists, several practical issues will need to be resolved in order to advance the concept of clinically applying β -blockade in cancer therapy. These issues include the selection of optimal pharmacologic agents (for example, non-selective antagonists that block β_2 receptors — such as propranolol

— are likely to be more effective than the β_1 -selective agents more commonly used in cardiology)^{14,35,40,53,54,107,111}; optimal disease settings (e.g., for reasons that are not yet understood mechanistically, some pharmaco-epidemiologic studies have found that β -antagonists are associated with greater protective effects on triple negative breast tumours than on tumours that express oestrogen receptor (ER), progesterone receptor (PR), or HER2^{41,45}), and optimal intervention timing and duration. For example, experimental models suggest that initiating β -antagonists prior to surgery may reduce SNS-mediated promotion of peri-surgical metastasis^{112,113}. The general pattern of pre-clinical evidence summarized above also implies that β -blockade may have the greatest impact on early stage tumours in which metastatic capacity is physiologically modifiable, and may have much more limited therapeutic impact in the setting of highly disseminated disease.

Beyond general considerations of disease stage, some molecular analyses suggest that it might also be possible to target individual tumours for adjuvant β -blockade based on SNS-related gene expression profiles^{6,16,49}. High expression of adrenergic receptors on tumor cells has not substantially predicted tumor responsiveness to β -antagonists *in vivo*^{54,114}, perhaps because SNS effects on tumour progression can also be mediated by adrenergic receptors on other cells in the surrounding microenvironment, systemic vasculature, and bone marrow haematopoietic and metastatic target tissues. Given the diversity of molecular pathways through which SNS activity might modulate tumour progression, the precise molecular indicators for adjuvant β -blockade will likely need to be empirically defined. However, there are grounds for expecting that the search for such transcriptomic fingerprints of SNS activity might be successful in both emerging patterns of stress- and SNS-related gene expression in tumours¹⁶ and similar precedents in other cell types (e.g., the conserved transcriptional response to adversity observed in circulating immune cells^{10,16}).

These empirical considerations underscore the need for randomized controlled, biomarker-enriched trials to assess initial proof-of-concept evidence that β -blockade can causally influence aspects of tumour biology that are relevant to disease progression. Further observational studies cannot definitively establish a clinical utility for β -blockers in cancer owing to a variety of methodological limitations including: confounding by indication (e.g., the primary historical indication for β -blockers, cardiovascular disease, shares common pathophysiologic drivers with cancer progression such as smoking, adiposity, and systemic inflammation); confounding with other pharmacologic exposures that can affect cancer progression (e.g., angiotensin-converting enzyme inhibitors); poor ascertainment (e.g., archival cardiovascular studies provide poor information on cancer progression and/or mortality, and archival cancer studies provide poor measures of β -blocker exposure); and temporal confounding of trends in cancer survival with trends in β -blocker utilization (particularly for the non-selective β -antagonists which are most likely to be efficacious). Randomized controlled studies will provide the only certain way to overcome these issues and definitively assess effects of β -antagonists on cancer-related outcomes. The availability of pre-clinical data and approved, safe, and inexpensive β -antagonists with well-understood pharmacology and minimal side-effects provide a favorable risk/benefit profile for initial proof-of-concept biomarker trials in clinical oncology.

Some laboratory studies have implicated β_3 - or α -adrenergic receptors in SNS effects on cancer, but the clinical significance of these effects remains to be determined. β_3 -adrenergic receptors are expressed on several types of cancer and stromal cells, and pharmacologic antagonists have been found to inhibit melanoma growth and vascularization in vivo^{34,36}. However, little is known about potential β_3 -adrenergic effects in human cancer, and interpretation of pre-clinical laboratory studies is complicated by the poor specificity of β_3 -adrenergic pharmacologic agents and the potential for non-selective β -antagonists such as propranolol to at least partly modulate β_3 receptors. β_3 -adrenergic receptors also have a key role in SNS regulation of mesenchymal stromal cells in the bone marrow hematopoietic niche^{8,11,115,116}, suggesting potential applications in hematopoietic cell transplantation and anti-tumor immunity. Biological effects of α -adrenergic agents in laboratory models of cancer have been complex and inconsistent^{117,118}. However, human epidemiologic studies have not generally indicated any notable effect of α -adrenergic antagonists on cancer risk or progression (some evidence suggests they may actually weakly promote some cancers)¹¹⁹ and β -adrenergic inhibition has generally been sufficient to block physiological stress effects on cancer in in vivo models (which would not happen if α -adrenergic receptors played a major role). Indeed, some 'α-adrenergic effects' may actually be mediated by downstream stimulation of β -adrenergic signaling that results from blockade of α_2 -adrenergic autoreceptors¹¹⁸. As such, the prospects for α -adrenergic agents in clinical cancer management are more ambiguous than those of β -adrenergic antagonists.

Perspective

Over the past decade it has become evident that malignancy is structured in major ways by interactions between cancer cells and their local tissue microenvironment¹²⁰. We are now beginning to appreciate how the broader physiological macroenvironment of the body can regulate these local tumour microenvironmental dynamics and thereby affect tumour progression and metastasis¹³. SNS regulation of MMDP gene expression programs in tumour cells and their stromal elements represents one of the most clearly defined pathways by which systemic physiology can regulate cancer biology^{15,16}. However, a great deal more research will be required to translate these basic observations into effective therapeutic approaches.

Much remains to be discovered about the cellular and molecular pathways through which SNS activation influences cancer biology. We know little about what effect, if any, the SNS might have on tumor initiation, on the development and conditioning of the metastatic niche, or on responses to therapy. It also remains unclear how much therapeutic leverage might be available from pharmacologic inhibition of SNS activity. If the dominant effect of the SNS in cancer occurs early in progression with the initial development of metastatic potential, the window of opportunity for β -blockade may well have passed once occult tumors become clinically evident. Even so, β -blockade may still have significant clinical utility in disease settings in which tumours are detected relatively early in development and metastatic capacity depends highly on upon physiological conditions. This might be particularly relevant in diseases for which current medical therapies show little efficacy (such as triple-negative breast cancer). Moreover, recent studies showing treatment-sensitizing effects of β -blockade imply some potential for value even in late-stage disease^{24,52–54,62,91,94,109,121,122}.

Much also remains to be clarified regarding regulation of tumour biology by the parasympathetic division of the autonomic nervous system. Recent studies suggest that parasympathetic innervation may contribute to tumour development and progression in certain tissue environments — such as the stomach and prostate gland^{77,123} — via acetylcholine-mediated activation of muscarinic acetylcholine receptors. However, pharmacologic intervention in such effects may be complicated by the fact that parasympathetic activity generally antagonizes the effects of the SNS^{1,3} and anti-cholinergic interventions may have the potential to indirectly stimulate SNS-mediated promotion of cancer. Moreover, the nicotinic acetylcholine receptors that mediate many parasympathetic effects on target tissues also serve as pre-synaptic neurotransmitters in the SNS^{1,3}, so the effects of some anti-cholinergic interventions may act through inhibition of SNS activity rather than (or in parallel with) inhibition of parasympathetic activity¹²⁴. As such, pharmacologic antagonism of cholinergic receptor systems may have more complex and unpredictable effects on tumour progression than the more focal targeting of β -adrenergic receptors. However, the central role of the (surprisingly dispensable) vagus nerve in mediating parasympathetic effects on cancer suggests a novel alternative strategy of surgically denervating selected target organs^{77,123,125}.

The highly pleiotropic effects of SNS activity on tumour biology suggest that even if nervous system-targeted interventions have moderate effects on any single pathway, their integrated effect across many parallel pathways may nevertheless be clinically significant. In an era of highly targeted therapies for the molecular pathogenesis of tumor cell proliferation, an adjuvant therapeutic strategy such as β -blockade that harnesses multiple microenvironmental pathways could provide a highly synergistic approach for controlling cancer progression.

Acknowledgments

Supported by grants from the United States National Institutes of Health (CA083639, CA098258, CA104825, CA109298, CA116778, CA140933, CA151668, CA177909, AG017265, AG033590) and Department of Defense (OC120547, OC093416), the Betty Ann Asche Murray Distinguished Professorship, the Cancer Prevention and Research Institute of Texas (CPRIT RP140106), and the Breast Cancer Research Foundation.

References

1. Weiner, H. *Perturbing the Organism: The Biology of Stressful Experience*. University of Chicago Press; Chicago: 1992.
2. Sapolsky, RM. *Why zebras don't get ulcers: A guide to stress, stress-related diseases, and coping*. Freeman; New York: 1994.
3. Sherwood, L. *Human Physiology: From Cells to Systems*. Cengage Learning; Boston: 2015.
4. Katayama Y, et al. Signals from the sympathetic nervous system regulate hematopoietic stem cell egress from bone marrow. *Cell*. 2006; 124:407–21. [PubMed: 16439213]
5. Sloan EK, et al. Social stress enhances sympathetic innervation of primate lymph nodes: mechanisms and implications for viral pathogenesis. *J Neurosci*. 2007; 27:8857–65. [PubMed: 17699667]
6. Lutgendorf SK, et al. Depression, social support, and beta-adrenergic transcription control in human ovarian cancer. *Brain Behav Immun*. 2009; 23:176–183. [PubMed: 18550328]
7. Powell ND, et al. Social stress up-regulates inflammatory gene expression in the leukocyte transcriptome via beta-adrenergic induction of myelopoiesis. *Proc Natl Acad Sci U S A*. 2013; 110:16574–9. [PubMed: 24062448]

8. Scheiermann C, Kunisaki Y, Frenette PS. Circadian control of the immune system. *Nat Rev Immunol.* 2013; 13:190–8. [PubMed: 23391992]
9. Cole SW. Social regulation of human gene expression: mechanisms and implications for public health. *Am J Public Health.* 2013; 103(Suppl 1):S84–92. [PubMed: 23927506]
10. Cole SW. Human social genomics. *PLoS Genet.* 2014; 10:e1004601. [PubMed: 25166010]
11. Hanoun M, Maryanovich M, Arnal-Estape A, Frenette PS. Neural regulation of hematopoiesis, inflammation, and cancer. *Neuron.* 2015; 86:360–373. [PubMed: 25905810]
12. Irwin MR, Cole SW. Reciprocal regulation of the neural and innate immune systems. *Nat Rev Immunol.* 2011; 11:625–632. [PubMed: 21818124]
13. Antoni MH, et al. The influence of bio-behavioural factors on tumour biology: pathways and mechanisms. *Nat Rev Cancer.* 2006; 6:240–8. [PubMed: 16498446]
14. Cole SW, Sood AK. Molecular pathways: beta-adrenergic signaling in cancer. *Clin Cancer Res.* 2012; 18:1201–6. [PubMed: 22186256]
15. Armaiz-Pena GN, Cole SW, Lutgendorf SK, Sood AK. Neuroendocrine influences on cancer progression. *Brain Behav Immun.* 2013; 30:S19–S25. [PubMed: 22728325]
16. Cole SW. Nervous system regulation of the cancer genome. *Brain Behav Immun.* 2013; 30(Suppl):S10–8. [PubMed: 23207104]
17. Powe DG, Entschladen F. Targeted therapies: Using beta-blockers to inhibit breast cancer progression. *Nat Rev Clin Oncol.* 2011; 8:511–2. [PubMed: 21808268]
18. Richter SD, et al. Time kinetics of the endocrine response to acute psychological stress. *J Clin Endocrinol Metab.* 1996; 81:1956–60. [PubMed: 8626864]
19. Schommer NC, Hellhammer DH, Kirschbaum C. Dissociation between reactivity of the hypothalamus-pituitary-adrenal axis and the sympathetic-adrenal-medullary system to repeated psychosocial stress. *Psychosom Med.* 2003; 65:450–60. [PubMed: 12764219]
20. Wingenfeld K, Whooley MA, Neylan TC, Otte C, Cohen BE. Effect of current and lifetime posttraumatic stress disorder on 24-h urinary catecholamines and cortisol: Results from the Mind Your Heart Study. *Psychoneuroendocrinology.* 2015; 52:83–91. [PubMed: 25459895]
21. Sloan EK, Capitanio JP, Tarara RP, Cole SW. Social temperament and lymph node innervation. *Brain Behav Immun.* 2008; 22:717–726. [PubMed: 18068331]
22. Schoffl C, Becker C, Prank K, von zur Muhlen A, Brabant G. Twenty-four-hour rhythms of plasma catecholamines and their relation to cardiovascular parameters in healthy young men. *Eur J Endocrinol.* 1997; 137:675–83. [PubMed: 9437236]
23. Dimitrov S, et al. Cortisol and epinephrine control opposing circadian rhythms in T cell subsets. *Blood.* 2009; 113:5134–43. [PubMed: 19293427]
24. Eng JW, et al. Housing temperature-induced stress drives therapeutic resistance in murine tumour models through beta2-adrenergic receptor activation. *Nat Commun.* 2015; 6:6426. [PubMed: 25756236]
25. Mendez-Ferrer S, Lucas D, Battista M, Frenette PS. Haematopoietic stem cell release is regulated by circadian oscillations. *Nature.* 2008; 452:442–7. [PubMed: 18256599]
26. Dutta P, et al. Myocardial infarction accelerates atherosclerosis. *Nature.* 2012; 487:325–9. [PubMed: 22763456]
27. Heidt T, et al. Chronic variable stress activates hematopoietic stem cells. *Nat Med.* 2014; 20:754–8. [PubMed: 24952646]
28. Nakai A, Hayano Y, Furuta F, Noda M, Suzuki K. Control of lymphocyte egress from lymph nodes through beta2-adrenergic receptors. *J Exp Med.* 2014; 211:2583–98. [PubMed: 25422496]
29. Sloan EK, Capitanio JP, Cole SW. Stress-induced remodeling of lymphoid innervation. *Brain Behav Immun.* 2008; 22:15–21. Epub 2007 Aug 13. [PubMed: 17697764]
30. Elenkov IJ, Wilder RL, Chrousos GP, Vizi ES. The sympathetic nerve--an integrative interface between two supersystems: the brain and the immune system. *Pharmacol Rev.* 2000; 52:595–638. [PubMed: 11121511]
31. Kohm AP, Sanders VM. Norepinephrine and beta 2-adrenergic receptor stimulation regulate CD4+ T and B lymphocyte function in vitro and in vivo. *Pharmacol Rev.* 2001; 53:487–525. [PubMed: 11734616]

32. Cole S, et al. Computational identification of gene-social environment interaction at the human IL6 locus. *Proc Natl Acad Sci U S A*. 2010; 107:5681–5686. [PubMed: 20176930]
33. Hori Y, et al. Naftopidil, a selective {alpha}1-adrenoceptor antagonist, suppresses human prostate tumor growth by altering interactions between tumor cells and stroma. *Cancer Prev Res (Phila)*. 2011; 4:87–96. [PubMed: 21205739]
34. Calvani M, et al. Norepinephrine promotes tumor microenvironment reactivity through beta3-adrenoreceptors during melanoma progression. *Oncotarget*. 2015; 6:4615–32. [PubMed: 25474135]
35. Thaker PH, et al. Chronic stress promotes tumor growth and angiogenesis in a mouse model of ovarian carcinoma. *Nat Med*. 2006; 12:939–44. Epub 2006 Jul 23. [PubMed: 16862152]
36. Dal Monte M, et al. Functional involvement of beta3-adrenergic receptors in melanoma growth and vascularization. *J Mol Med (Berl)*. 2013; 91:1407–19. [PubMed: 23907236]
37. Sterling, P. Principles of allostasis: Optimal design, predictive regulation, pathophysiology and rational therapeutics. In: Schulkin, J., editor. *Allostasis, homeostasis, and the costs of physiological adaptation*. Cambridge University Press; Cambridge: 2004.
38. Chida Y, Hamer M, Wardle J, Steptoe A. Do stress-related psychosocial factors contribute to cancer incidence and survival? *Nat Clin Pract Oncol*. 2008; 5:466–75. Epub 2008 May 20. [PubMed: 18493231]
39. Powe DG, et al. Beta-blocker drug therapy reduces secondary cancer formation in breast cancer and improves cancer specific survival. *Oncotarget*. 2010; 1:628–38. [PubMed: 21317458]
40. Barron TI, Connolly RM, Sharp L, Bennett K, Visvanathan K. Beta blockers and breast cancer mortality: a population- based study. *J Clin Oncol*. 2011; 29:2635–44. [PubMed: 21632503]
41. Melhem-Bertrandt A, et al. Beta-blocker use is associated with improved relapse-free survival in patients with triple-negative breast cancer. *J Clin Oncol*. 2011; 29:2645–52. [PubMed: 21632501]
42. De Giorgi V, et al. Treatment with beta-blockers and reduced disease progression in patients with thick melanoma. *Arch Intern Med*. 2011; 171:779–81. [PubMed: 21518948]
43. Lemeshow S, et al. Beta-Blockers and Survival among Danish Patients with Malignant Melanoma: A Population-Based Cohort Study. *Cancer Epidemiol Biomarkers Prev*. 2011; 20:2273–9. [PubMed: 21933972]
44. Aydiner A, Ciftci R, Karabulut S, Kilic L. Does beta-blocker therapy improve the survival of patients with metastatic non-small cell lung cancer? *Asian Pac J Cancer Prev*. 2013; 14:6109–14. [PubMed: 24289634]
45. Botteri E, et al. Therapeutic effect of beta-blockers in triple-negative breast cancer postmenopausal women. *Breast Cancer Res Treat*. 2013; 140:567–75. [PubMed: 23912960]
46. De Giorgi V, et al. Effect of beta-blockers and other antihypertensive drugs on the risk of melanoma recurrence and death. *Mayo Clin Proc*. 2013; 88:1196–203. [PubMed: 24182700]
47. Grytli HH, Fagerland MW, Fossa SD, Tasken KA, Haheim LL. Use of beta-blockers is associated with prostate cancer-specific survival in prostate cancer patients on androgen deprivation therapy. *Prostate*. 2013; 73:250–60. [PubMed: 22821802]
48. Grytli HH, Fagerland MW, Fossa SD, Tasken KA. Association between use of beta-blockers and prostate cancer-specific survival: a cohort study of 3561 prostate cancer patients with high-risk or metastatic disease. *Eur Urol*. 2014; 65:635–41. [PubMed: 23351721]
49. Sloan EK, et al. The sympathetic nervous system induces a metastatic switch in primary breast cancer. *Cancer Res*. 2010; 70:7042–52. [PubMed: 20823155]
50. Madden KS, Szpunar MJ, Brown EB. beta-Adrenergic receptors (beta-AR) regulate VEGF and IL-6 production by divergent pathways in high beta-AR-expressing breast cancer cell lines. *Breast Cancer Res Treat*. 2011; 130:747–58. [PubMed: 21234673]
51. Palm D, et al. The norepinephrine-driven metastasis development of PC-3 human prostate cancer cells in BALB/c nude mice is inhibited by beta-blockers. *Int J Cancer*. 2006; 118:2744–9. [PubMed: 16381019]
52. Hassan S, et al. Behavioral stress accelerates prostate cancer development in mice. *J Clin Invest*. 2013; 123:874–86. [PubMed: 23348742]
53. Pasquier E, et al. beta-blockers increase response to chemotherapy via direct antitumour and anti-angiogenic mechanisms in neuroblastoma. *Br J Cancer*. 2013; 108:2485–94. [PubMed: 23695022]

54. Wolter JK, et al. Anti-tumor activity of the beta-adrenergic receptor antagonist propranolol in neuroblastoma. *Oncotarget*. 2014; 5:161–72. [PubMed: 24389287]
55. Hasegawa H, Saiki I. Psychosocial stress augments tumor development through beta-adrenergic activation in mice. *Jpn J Cancer Res*. 2002; 93:729–35. [PubMed: 12149137]
56. Goldfarb Y, et al. Improving postoperative immune status and resistance to cancer metastasis: a combined perioperative approach of immunostimulation and prevention of excessive surgical stress responses. *Ann Surg*. 2011; 253:798–810. [PubMed: 21475023]
57. Kim-Fuchs C, et al. Chronic stress accelerates pancreatic cancer growth and invasion: a critical role for beta-adrenergic signaling in the pancreatic microenvironment. *Brain Behav Immun*. 2014; 40:40–7. [PubMed: 24650449]
58. Lamkin DM, et al. Chronic stress enhances progression of acute lymphoblastic leukemia via beta-adrenergic signaling. *Brain Behav Immun*. 2012; 26:635–41. [PubMed: 22306453]
59. Inbar S, et al. Do stress responses promote leukemia progression? An animal study suggesting a role for epinephrine and prostaglandin-E2 through reduced NK activity. *PLoS One*. 2011; 6:e19246. [PubMed: 21559428]
60. Hara MR, et al. A stress response pathway regulates DNA damage through beta2-adrenoreceptors and beta-arrestin-1. *Nature*. 2011; 477:349–53. [PubMed: 21857681]
61. Hara MR, Sachs BD, Caron MG, Lefkowitz RJ. Pharmacological blockade of a beta(2)AR-beta-arrestin-1 signaling cascade prevents the accumulation of DNA damage in a behavioral stress model. *Cell Cycle*. 2013; 12:219–24. [PubMed: 23287463]
62. Reeder A, et al. Stress hormones reduce the efficacy of paclitaxel in triple negative breast cancer through induction of DNA damage. *Br J Cancer*. 2015; 112:1461–70. [PubMed: 25880007]
63. Armaiz-Pena GN, et al. Src activation by beta-adrenoreceptors is a key switch for tumour metastasis. *Nat Commun*. 2013; 4:1403. [PubMed: 23360994]
64. Shi M, et al. The beta2-adrenergic receptor and Her2 comprise a positive feedback loop in human breast cancer cells. *Breast Cancer Res Treat*. 2011; 125:351–62. [PubMed: 20237834]
65. Gu L, Lau SK, Loera S, Somlo G, Kane SE. Protein kinase A activation confers resistance to trastuzumab in human breast cancer cell lines. *Clin Cancer Res*. 2009; 15:7196–206. [PubMed: 19920112]
66. Chang M, et al. Beta-adrenoreceptors reactivate KSHV lytic replication via PKA-dependent control of viral RTA. *Journal of Virology*. 2005
67. zur Hausen, H. *Infections Causing Human Cancer*. Wiley-VCH; Weinheim: 2008.
68. Nilsson MB, et al. Stress hormones regulate interleukin-6 expression by human ovarian carcinoma cells through a Src-dependent mechanism. *J Biol Chem*. 2007; 282:29919–26. Epub 2007 Aug 23. [PubMed: 17716980]
69. Shahzad MM, et al. Stress effects on FosB- and interleukin-8 (IL8)-driven ovarian cancer growth and metastasis. *J Biol Chem*. 2010; 285:35462–70. [PubMed: 20826776]
70. Yang R, Lin Q, Gao HB, Zhang P. Stress-related hormone norepinephrine induces interleukin-6 expression in GES-1 cells. *Braz J Med Biol Res*. 2014; 47:101–9. [PubMed: 24519125]
71. Cakir Y, Plummer HK 3rd, Tithof PK, Schuller HM. Beta-adrenergic and arachidonic acid-mediated growth regulation of human breast cancer cell lines. *Int J Oncol*. 2002; 21:153–7. [PubMed: 12063562]
72. Armaiz-Pena GN, et al. Adrenergic regulation of monocyte chemotactic protein 1 leads to enhanced macrophage recruitment and ovarian carcinoma growth. *Oncotarget*. 2015; 6:4266–4273. [PubMed: 25738355]
73. Collado-Hidalgo A, Sung C, Cole S. Adrenergic inhibition of innate anti-viral response: PKA blockade of Type I interferon gene transcription mediates catecholamine support for HIV-1 replication. *Brain Behav Immun*. 2006; 20:552–63. Epub 2006 Feb 28. [PubMed: 16504464]
74. Cole SW, Korin YD, Fahey JL, Zack JA. Norepinephrine accelerates HIV replication via protein kinase A-dependent effects on cytokine production. *Journal of Immunology*. 1998; 161:610–616.
75. Glasner A, et al. Improving survival rates in two models of spontaneous postoperative metastasis in mice by combined administration of a beta-adrenergic antagonist and a cyclooxygenase-2 inhibitor. *J Immunol*. 2010; 184:2449–57. [PubMed: 20124103]

76. Lee JW, et al. Surgical stress promotes tumor growth in ovarian carcinoma. *Clin Cancer Res.* 2009; 15:2695–702. [PubMed: 19351748]
77. Magnon C, et al. Autonomic nerve development contributes to prostate cancer progression. *Science.* 2013; 341:1236361. [PubMed: 23846904]
78. Bruzzone A, et al. alpha(2)-Adrenoceptors enhance cell proliferation and mammary tumor growth acting through both the stroma and the tumor cells. *Curr Cancer Drug Targets.* 2011; 11:763–74. [PubMed: 21599632]
79. Flint MS, et al. Chronic exposure to stress hormones promotes transformation and tumorigenicity of 3T3 mouse fibroblasts. *Stress.* 2013; 16:114–21. [PubMed: 22506837]
80. Cao L, et al. Environmental and genetic activation of a brain-adipocyte BDNF/leptin axis causes cancer remission and inhibition. *Cell.* 2010; 142:52–64. [PubMed: 20603014]
81. Cao L, During MJ. What is the brain-cancer connection? *Annu Rev Neurosci.* 2012; 35:331–45. [PubMed: 22462541]
82. Hanoun M, et al. Acute myelogenous leukemia-induced sympathetic neuropathy promotes malignancy in an altered hematopoietic stem cell niche. *Cell Stem Cell.* 2014; 15:365–75. [PubMed: 25017722]
83. Lang K, et al. Induction of a metastatogenic tumor cell type by neurotransmitters and its pharmacological inhibition by established drugs. *Int J Cancer.* 2004; 112:231–8. [PubMed: 15352035]
84. Drell, TL, et al. Effects of neurotransmitters on the chemokinesis and chemotaxis of MDA-MB-468 human breast carcinoma cells. *Breast Cancer Res Treat.* 2003; 80:63–70. [PubMed: 12889599]
85. Landen CN Jr, et al. Neuroendocrine modulation of signal transducer and activator of transcription-3 in ovarian cancer. *Cancer Res.* 2007; 67:10389–96. [PubMed: 17974982]
86. Sood AK, et al. Stress hormone-mediated invasion of ovarian cancer cells. *Clin Cancer Res.* 2006; 12:369–75. [PubMed: 16428474]
87. Yang EV, et al. Norepinephrine up-regulates the expression of vascular endothelial growth factor, matrix metalloproteinase (MMP)-2, and MMP-9 in nasopharyngeal carcinoma tumor cells. *Cancer Res.* 2006; 66:10357–64. [PubMed: 17079456]
88. Chakraborty D, Sarkar C, Basu B, Dasgupta PS, Basu S. Catecholamines regulate tumor angiogenesis. *Cancer Res.* 2009; 69:3727–30. [PubMed: 19383906]
89. Yang EV, et al. Norepinephrine upregulates VEGF, IL-8, and IL-6 expression in human melanoma tumor cell lines: implications for stress-related enhancement of tumor progression. *Brain Behav Immun.* 2009; 23:267–75. [PubMed: 18996182]
90. Moretti S, et al. beta-adrenoceptors are upregulated in human melanoma and their activation releases pro-tumorigenic cytokines and metalloproteases in melanoma cell lines. *Lab Invest.* 2013; 93:279–90. [PubMed: 23318885]
91. Liu J, et al. The effect of chronic stress on anti-angiogenesis of sunitinib in colorectal cancer models. *Psychoneuroendocrinology.* 2015; 52:130–42. [PubMed: 25437118]
92. Sood AK, et al. Adrenergic modulation of focal adhesion kinase protects human ovarian cancer cells from anoikis. *J Clin Invest.* 2010; 120:1515–23. [PubMed: 20389021]
93. Sastry KS, et al. Epinephrine protects cancer cells from apoptosis via activation of cAMP-dependent protein kinase and BAD phosphorylation. *J Biol Chem.* 2007; 282:14094–100. [PubMed: 17353197]
94. Deng GH, et al. Exogenous norepinephrine attenuates the efficacy of sunitinib in a mouse cancer model. *J Exp Clin Cancer Res.* 2014; 33:21. [PubMed: 24555849]
95. Dar A, et al. Rapid mobilization of hematopoietic progenitors by AMD3100 and catecholamines is mediated by CXCR4-dependent SDF-1 release from bone marrow stromal cells. *Leukemia.* 2011; 25:1286–96. [PubMed: 21494253]
96. Lucas D, et al. Chemotherapy-induced bone marrow nerve injury impairs hematopoietic regeneration. *Nat Med.* 2013; 19:695–703. [PubMed: 23644514]
97. Lutgendorf SK, et al. Social isolation is associated with elevated tumor norepinephrine in ovarian carcinoma patients. *Brain Behav Immun.* 2011; 25:250–255. [PubMed: 20955777]

98. Ayala GE, et al. Cancer-related axonogenesis and neurogenesis in prostate cancer. *Clin Cancer Res.* 2008; 14:7593–603. [PubMed: 19047084]
99. Voss MJ, Entschladen F. Tumor interactions with soluble factors and the nervous system. *Cell Commun Signal.* 2010; 8:21. [PubMed: 20822525]
100. Guo K, et al. Interaction of the sympathetic nerve with pancreatic cancer cells promotes perineural invasion through the activation of STAT3 signaling. *Mol Cancer Ther.* 2013; 12:264–73. [PubMed: 23288783]
101. Xu Q, et al. Stromal-derived factor-1alpha/CXCL12-CXCR4 chemotactic pathway promotes perineural invasion in pancreatic cancer. *Oncotarget.* 2015; 6:4717–32. [PubMed: 25605248]
102. Flierl MA, et al. Phagocyte-derived catecholamines enhance acute inflammatory injury. *Nature.* 2007; 449:721–5. Epub 2007 Sep 30. [PubMed: 17914358]
103. Campbell JP, et al. Stimulation of host bone marrow stromal cells by sympathetic nerves promotes breast cancer bone metastasis in mice. *PLoS Biol.* 2012; 10:e1001363. [PubMed: 22815651]
104. Lu H, et al. Impact of beta-blockers on prostate cancer mortality: a meta-analysis of 16,825 patients. *Onco Targets Ther.* 2015; 8:985–90. [PubMed: 25995645]
105. Wang HM, et al. Improved survival outcomes with the incidental use of beta-blockers among patients with non-small-cell lung cancer treated with definitive radiation therapy. *Ann Oncol.* 2013; 24:1312–9. [PubMed: 23300016]
106. Diaz ES, Karlan BY, Li AJ. Impact of beta blockers on epithelial ovarian cancer survival. *Gynecol Oncol.* 2012; 127:375–8. [PubMed: 22819786]
107. Watkins JL, et al. Clinical impact of selective and non-selective beta-blockers on survival in ovarian cancer patients. *Cancer.* 2015 in press.
108. Schuller HM, Porter B, Riechert A. Beta-adrenergic modulation of NNK-induced lung carcinogenesis in hamsters. *J Cancer Res Clin Oncol.* 2000; 126:624–30. [PubMed: 11079726]
109. Pasquier E, et al. Propranolol potentiates the anti-angiogenic effects and anti-tumor efficacy of chemotherapy agents: implication in breast cancer treatment. *Oncotarget.* 2011; 2:797–809. [PubMed: 22006582]
110. Lin Q, et al. Effect of chronic restraint stress on human colorectal carcinoma growth in mice. *PLoS One.* 2013; 8:e61435. [PubMed: 23585898]
111. Ganz PA, Cole SW. Expanding our therapeutic options: Beta blockers for breast cancer? *J Clin Oncol.* 2011; 29:2612–6. [PubMed: 21632500]
112. Neeman E, Zmora O, Ben-Eliyahu S. A new approach to reducing postsurgical cancer recurrence: perioperative targeting of catecholamines and prostaglandins. *Clin Cancer Res.* 2012; 18:4895–902. [PubMed: 22753587]
113. Horowitz M, Neeman E, Sharon E, Ben-Eliyahu S. Exploiting the critical perioperative period to improve long-term cancer outcomes. *Nat Rev Clin Oncol.* 2015; 12:213–26. [PubMed: 25601442]
114. Boucek RJ Jr, Kirsh AL, Majesky MW, Perkins JA. Propranolol responsiveness in vascular tumors is not determined by qualitative differences in adrenergic receptors. *Otolaryngol Head Neck Surg.* 2013; 149:772–6. [PubMed: 24009211]
115. Mendez-Ferrer S, Battista M, Frenette PS. Cooperation of beta(2)- and beta(3)-adrenergic receptors in hematopoietic progenitor cell mobilization. *Ann N Y Acad Sci.* 2010; 1192:139–44. [PubMed: 20392229]
116. Magnon C, Lucas D, Frenette PS. Trafficking of stem cells. *Methods Mol Biol.* 2011; 750:3–24. [PubMed: 21618080]
117. Szpunar MJ, Burke KA, Dawes RP, Brown EB, Madden KS. The antidepressant desipramine and alpha2-adrenergic receptor activation promote breast tumor progression in association with altered collagen structure. *Cancer Prev Res (Phila).* 2013; 6:1262–72. [PubMed: 24309563]
118. Lamkin DM, et al. alpha2-Adrenergic blockade mimics the enhancing effect of chronic stress on breast cancer progression. *Psychoneuroendocrinology.* 2015; 51:262–70. [PubMed: 25462899]
119. Friedman GD, Udaltsova N, Habel LA. Norepinephrine antagonists and cancer risk. *Int J Cancer.* 2011; 128:737–8. author reply 739. [PubMed: 20333678]

120. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nat Med*. 2013; 19:1423–37. [PubMed: 24202395]
121. Shan T, et al. beta2-adrenoceptor blocker synergizes with gemcitabine to inhibit the proliferation of pancreatic cancer cells via apoptosis induction. *Eur J Pharmacol*. 2011; 665:1–7. [PubMed: 21570961]
122. Obeid EI, Conzen SD. The role of adrenergic signaling in breast cancer biology. *Cancer Biomark*. 2013; 13:161–9. [PubMed: 23912488]
123. Zhao CM, et al. Denervation suppresses gastric tumorigenesis. *Sci Transl Med*. 2014; 6:250ra115.
124. Rosas-Ballina M, et al. Splenic nerve is required for cholinergic antiinflammatory pathway control of TNF in endotoxemia. *Proc Natl Acad Sci U S A*. 2008; 105:11008–13. [PubMed: 18669662]
125. Villanueva MT. Therapeutics: Gastric cancer gets a red carpet treatment. *Nat Rev Cancer*. 2014; 14

Biographies

Steven W. Cole is a Professor of Medicine in the Division of Hematology-Oncology at the David Geffen School of Medicine at UCLA. His research utilizes molecular genetics and computational bioinformatics to analyze the pathways by which social and environmental factors influence the activity of the human genome, as well as viral and tumor genomes. He pioneered the field of human social genomics, and serves as Director of the UCLA Social Genomics Core Laboratory. He is also a member of the Jonsson Comprehensive Cancer Center, the Norman Cousins Center, the UCLA AIDS Institute, and the UCLA Molecular Biology Institute.

Archana S. Nagaraja is a doctoral student in the Department of Cancer Biology and Department of Gynecologic Oncology at the University of Texas at MD Anderson Cancer Center. She is supported by a Research Training Award from the Cancer Prevention and Research Institute of Texas. Her research studies the role of adrenergic signaling in modulating inflammation and metastasis in ovarian cancer.

Susan K. Lutgendorf is a Professor in the Departments of Psychology, Obstetrics and Gynecology, and Urology and member of the Holden Comprehensive Cancer Center at the University of Iowa. She directs an NIGMS-funded Training Program entitled Mechanisms of Health and Disease at the Behavioral-Biomedical Interface. Her current work, funded by the National Cancer Institute, examines how stress, depression, and social support are linked to biological processes involved in tumour progression. Her work has been recognized by a New Investigator Award from the Psychoneuroimmunology Research Society, an American Psychological Association Award for Outstanding Contributions to Health Psychology.

Paige A. Green is Chief, Basic Biobehavioral and Psychological Sciences Branch in the Behavioral Research Program, Division of Cancer Control and Population Sciences at the National Cancer Institute (NCI). She serves as Chair for the NCI Network on Biobehavioral Pathways in Cancer, a research consortium that strives to accelerate the discovery, development, and clinical translation of cancer relevant molecular pathways and networks regulated by social, behavioral, and psychological factors through the central nervous system.

Anil K. Sood is Professor and Vice Chairman for Translational Research in the Department of Gynecologic Oncology and has a joint appointment in the Department of Cancer Biology at the University of Texas M.D. Anderson Cancer Center. He is also Co-Director of the Center for RNA Interference (RNAi) and Non-Coding RNA and Director of the Blanton-Davis Ovarian Cancer Research Program. His main research interests include neuroendocrine effects on cancer metastasis, RNAi therapeutics, and development of new strategies for targeting the tumour microenvironment.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

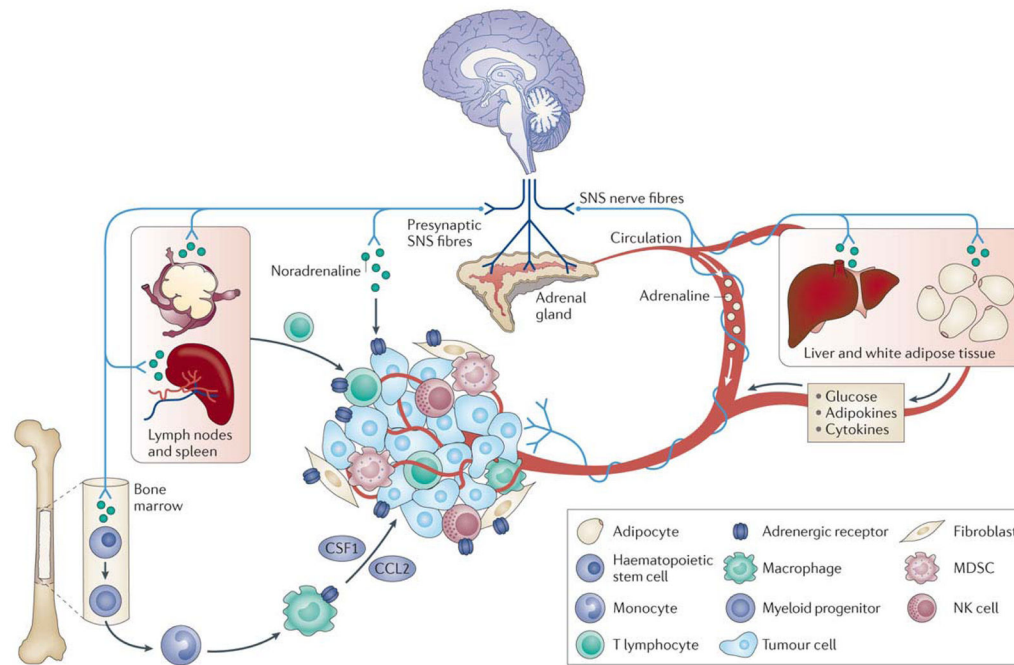


Figure 1. SNS regulation of the tumour microenvironment

SNS activation can regulate gene expression and cellular function in the tumour microenvironment through a variety of pathways. Direct SNS effects on tumour biology are mediated by catecholamine neuroeffector molecules (epinephrine and norepinephrine) that are released into the tumour microenvironment to engage adrenergic receptors that are expressed on many types of tumor cells and their surrounding stromal elements such as tumor-associated macrophages and vascular endothelial cells. Epinephrine is released from the adrenal gland and circulates to the tumour microenvironment through the vasculature, whereas norepinephrine is released from sympathetic nerve fibers within the tumour microenvironment, which generally associate with the vasculature and can sometimes radiate dendritic fibers into the tumour parenchyma. Indirect effects on tumour biology are mediated by release of catecholamine neuroeffector molecules into distal tissue sites that regulate systemic biological processes which subsequently impinge on tumour biology, such as regulation of immune cell development (e.g., myelopoiesis in the bone marrow and spleen, and lymphocyte differentiation in secondary lymphoid organs such as the spleen and lymph nodes) and trafficking (e.g., monocyte/macrophage recruitment via chemokines such as MCP-1/*CCL2* and growth factors such as M-CSF/*CSF1*), or regulation of systemic metabolic and hormonal regulators of tumour growth (e.g., glucose mobilization from the liver and circulating adipokines from white adipose tissue). These multiple regulatory pathways allow the SNS to exert highly pleiotropic effects on tumour progression and metastasis of many solid epithelial tumors (e.g., breast, prostate, ovary, lung, pancreas) as well as hematologic malignancies via innervation of lymphoid organs such as the bone marrow, spleen, and lymph nodes.

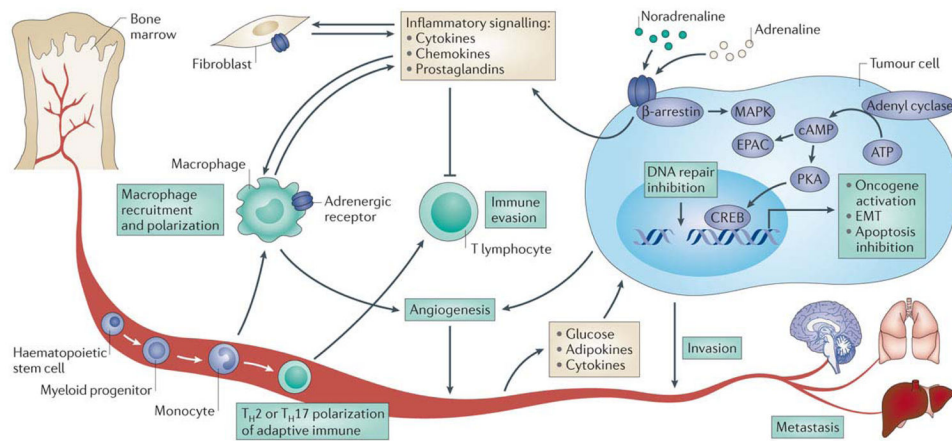


Figure 2. Molecular mechanisms for SNS regulation of tumor progression

SNS signaling through α - and β -adrenergic receptor systems can regulate a wide variety of molecular processes involved in tumour progression and metastasis, including DNA damage repair, signaling by cellular and viral oncogenes, expression of pro-inflammatory mediators (cytokines, chemokines, prostaglandins) by tumour cells and immune cells, recruitment and pro-metastatic transcriptional programming of macrophages, angiogenesis and lymphangiogenesis, epithelial-mesenchymal transition, tumor cell motility and invasive capacity, resistance to apoptosis and chemotherapy-mediated cell death, and inhibition of cytokines and cytotoxic function in adaptive immune responses. SNS activation also exerts immunoregulatory effects through innervation of the bone marrow hematopoietic niche to promote stem cell mobilization and development of myeloid lineage immune cells (monocytes and macrophages and myeloid-derived suppressor cells), through innervation of the spleen to influence extra-medullary myelopoiesis of monocytes, macrophages and myeloid-derived suppressor cells, and through innervation of other primary and secondary lymphoid organs to inhibit cellular immune responses and promote humoral immune responses. SNS activation additionally regulates a wide variety of systemic metabolic and hormonal processes that can impact tumour progression, including mobilization of glucose and fatty acids from the liver and adipokines and pro-inflammatory cytokines from white adipose tissue. Many of these molecular effects have been found to be regulated by β -adrenergic receptors (ADRB), which regulate cellular and viral gene expression via activation of multiple intracellular signal transduction pathways including cyclic-3'-5'-adenosine monophosphate (cAMP)-mediated activation of protein kinase A (PKA), which subsequently phosphorylates transcription factors such as cAMP response element-binding protein (CREB); cAMP-mediated activation of the guanine exchange protein activated by adenylyl cyclase (EPAC); and β -arrestin-mediated activation of MAP kinase signaling pathways. β -adrenergic-induction of multiple intracellular signaling pathways further amplifies the impact of the multiple parallel extracellular signaling pathways (Figure 1) to generate a highly pleiotropic network of molecular effects that generally stimulate tumor progression and metastasis.