MedChemComm



RESEARCH ARTICLE

View Article Online
View Journal | View Issue



Cite this: *Med. Chem. Commun.*, 2017, **8**, 2258

Received 28th August 2017, Accepted 14th October 2017

DOI: 10.1039/c7md00442g

rsc.li/medchemcomm

Synthesis and antifungal activity of novel oxazolidin-2-one-linked 1,2,3-triazole derivatives†

Novel oxazolidin-2-one-linked 1,2,3-triazole derivatives (4a-k) were synthesized by straightforward and versatile azide-enolate (3 + 2) cycloaddition. The series of compounds was screened for antifungal activity against four filamentous fungi as well as six yeast species of *Candida* spp. According to their efficiency and breadth of scope, they can be ordered as 4k > 4d > 4h > 4a, especially in relation to the activity displayed against *Candida glabrata* ATCC-34138, *Trichosporon cutaneum* ATCC-28592 and *Mucor hiemalis* ATCC-8690, *i.e.* compounds 4d, 4h and 4k showed excellent activity against *C. glabrata* (MIC 0.12, 0.25 and 0.12 μ g mL⁻¹, respectively), better than that of itraconazole (MIC 1 μ g ml⁻¹). The activity of compound 4d (MIC = 2 μ g mL⁻¹) was higher than that observed for the standard antifungal drug (MIC = 8 μ g mL⁻¹) against *Trichosporon cutaneum*, while compound 4k displayed an excellent antimycotic activity against *Mucor hiemalis* (MIC = 2 μ g mL⁻¹ vs. 4 μ g mL⁻¹ for itraconazole). In addition, we describe herein a novel mild and eco-friendly synthetic protocol for obtaining β -ketosulfones (adducts to afford compounds 4a-k) from α -brominated carbonyls in an aqueous nanomicellar medium at room temperature.

1. Introduction

Both oxazolidin-2-one¹ and 1,2,3-triazole² cores are very well-recognized pharmacophores in the literature. They have become extremely versatile in medicinal chemistry, featuring a number of clinically used drugs (*e.g.* Linezolid 1 or Tazobactam 2, Fig. 1). Consequently, several antimicrobials based on one or the other scaffold have been reported in the literature.³ On the other hand, the study of *both cores in the same molecule* (3) as an antibacterial agent has been reported⁴ and patented (*e.g.* 3a (ref. 5) and 3b (ref. 6)). This year – 2017 – Melinta Therapeutics Inc. announced the acceptance of FDA of the Investigational New Drug (IND) application for topical

Radezolid 3c, a second-generation oxazolidinone/triazole compound discovered by Melinta scientists as a novel antibiotic to treat serious bacterial infections. Curiously, antifungal activity has not been demonstrated for these kinds of compounds to the best of our knowledge.

Even though the mechanism of action of oxazolidinone⁸ and triazole⁹ cores has been described, the inhibition of monoamine oxidase A (MAO-A)^{4a} as well as the RNA-binding process^{4g} has been proposed as the mode of action for oxazolidin-2-one-linked 1,2,3-triazole.

Due to the strong resistance of fungi to current drugs, there is a continuous search for antifungal agents. As part of our ongoing research, we herein describe the synthesis and biological evaluation of novel oxazolidin-2-one-linked 1,2,3-triazole derivatives (4).

2. Chemistry

Our initial study began by obtaining the starting materials, azide/oxazolidin-2-one 8 and ketones 11 (Scheme 1). Firstly, chlorine derivative 7 was synthesized (61%) by coupling between 2-chloroethyl isocyanate 6 and benzoin 5.¹⁰ Subsequently, the nucleophilic substitution of 7 by the azide ion furnished compound 8 (58%).

Ketones 11a-11g were synthesized since, unlike 11i-11j, they are not commercially available (ketone 11h was kindly donated by Syntex-La Roche). For these purposes, we have developed a novel organic solvent-free synthesis of

^a Departamento de Química Orgánica, Facultad de Química, Universidad Autónoma del Estado de México, Paseo Colón/Paseo Tollocan s/n, Toluca, Estado de México, 50120, Mexico. E-mail: mpagfuentesb@uaemex.mx; Fax: +52 722 217 3890; Tel: +52 722 217 5109x113

b Departamento de Microbiología, Facultad de Química, Universidad Autónoma del Estado de México, Paseo Colón/Paseo Tollocan s/n, Toluca, Estado de México, 50120, Mexico

^c Departamento de Química Orgánica, Escuela Nacional de Ciencias Biológicas, Instituto Politécnico Nacional, Prol. Carpio y Plan de Ayala, Ciudad de México, 11340, Mexico

^d Centro Conjunto de Investigación en Química Sustentable UAEM-UNAM, Carretera Toluca-Atlacomulco Km 14.5, Toluca, 52000, Mexico

[†] Electronic supplementary information (ESI) available: Supplementary data (experimental procedures, characterization data of all compounds and copies of $^{13}\text{C},\,^{1}\text{H},$ and spectra, as well as the study on the synthesis of β-ketosulfones). See DOI: 10.1039/c7md00442g

MedChemComm Research Article

U.S. FDA approved antimicrobial pharmaceuticals:

Oxazolidin-2-ones linked-1,2,3-triazoles:

AstraZeneca patent 3a: $R^2 = R^3 = H$ Pfizer patent 3b: R2= H. R3= C≡CH (antifungal agents)

Fig. 1 The novel series of compounds 4 involves the extremely important oxazolidin-2-one and 1,2,3-triazole pharmacophoric cores. A similar feature is found in antimicrobial oxazolidin-2-one-linked 1,2,3-triazole 3, including lead compounds 1 and 2.

NHCOMe

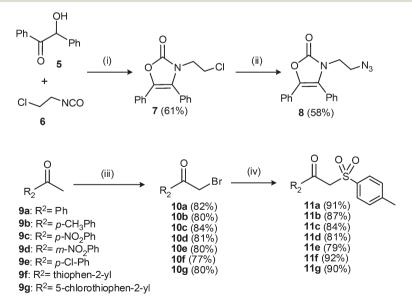
Зс

β-ketosulfones in an aqueous nanomicellar medium by the concomitant use of the surfactant reagent SPGS-550-M or 'Nok' (now available at Sigma-Aldrich¹¹) and sodium p-toluenesulfinate from \alpha-bromoketones. The surfactant nature of NOK (a third generation surfactant) allows it to act as an efficient nanoreactor in catalytic amounts. The study and scope of this novel methodology are described in the ESI† in greater detail. Therefore, we decided to apply such a methodology in the synthesis of compound 11 from 10, which in turn was accomplished by reported protocols12 with N-bromosuccinimide (NBS) as the bromine source.

Although Cu-catalyzed azide-alkyne cvcloaddition (CuAAC) is the conventional method for obtaining 1,2,3triazole moieties, 13 other strategies have emerged as alternatives for such a purpose. We previously reported14 a novel synthetic protocol to achieve the efficient assembly of 1,4,5trisubstituted 1,2,3-triazole cores through azide-enolate cycloaddition. In consequence, oxazolidin-2-one-linked 1,2,3triazole derivatives (4) were synthesized by the efficient 1,3-dipolar cycloaddition of azide/oxazolidin-2-one in the presence of enolates prepared in situ from ketones 11 activated by DBU as the base. Table 1 summarizes these outcomes.

3. Microbiology

Compounds 4a-k were tested for their in vitro activity against four filamentous fungi: Aspergillus fumigatus ATCC-16907, Trichosporon cutaneum ATCC-28592, Rhizopus oryzae ATCC-10329 and Mucor hiemalis ATCC-8690. These compounds were also evaluated, employing standardized microbiological methods developed by the CLSI, against six yeast specimens: Candida albicans ATCC-10231, Candida utilis ATCC-9226, Candida tropicalis ATCC-13803, Candida parapsilopsis ATCC-22019,



Scheme 1 Reagents and conditions: (i) N2, 180 °C, 24 h. (ii) NaN3 (1.1 eq.), DMF anh., 60 °C, 12 h, N2. (iii) NBS (1.1 eq.), TsOH·H2O (1.0 eq.), MeCN, 60 °C, 4 h. (iv) SPGS-550-M aq. (2% w/w), p-Tol-SO₂Na (1.5 eq.), r.t., 12 h.

research Article

Table 1 Synthesis of oxazolidin-2-one-linked 1,2,3-triazoles 4a-k from azide 8 by coupling with active ketones 11

Entry ^a	Ketone	Triazole ^b (yield%) ^c
1	11a: $R^2 = Ph$, $R^1 = SO_2 - p$ -Tol	4a (68)
2	11b: $R^2 = p\text{-CH}_3\text{Ph}, R^1 = SO_2\text{-}p\text{-Tol}$	4b (61)
3	11c: $R^2 = p\text{-NO}_2\text{-Ph}$, $R^1 = SO_2\text{-}p\text{-Tol}$	4c (74)
4	11d: $R^2 = m - NO_2 Ph$, $R^1 = SO_2 - p - Tol$	4d (70)
5	11e: $R^2 = p$ -Cl-Ph, $R^1 = SO_2$ - p -Tol	4e (72)
6	11f: R^2 = thiophen-2-yl, R^1 = SO_2 - p -Tol	4f (68)
7	11g: R^2 = 5-chlorothiophen-2-yl, R^1 = SO_2 - p -Tol	4g (64)
8	11h: R^2 = pentyl, R^1 = SO_2Ph	4h (71)
9	11i: $R^2 = CH_3$, $R^1 = COCH_3$	4i (73)
10	11j: $R^2 = Ph$, $R^1 = COPh$	4j (71)
11	11k: $R^2 = Ph$, $R^1 = CN$	$4\mathbf{k}$ (67)

^a Reaction conditions: To a solution of compound 8 (1.0 eq.) and 11 (1.0 eq.) in DMF anh., DBU (2.0 eq.) was added. The reaction mixture was stirred at 50–60 $^{\circ}$ C for 12–24 h. ^b Confirmed by ¹H-NMR, ¹³C-NMR and MS. ^c Yields refer to chromatographically pure isolated compounds.

Candida glabrata ATCC-34138, and Candida krusei ATCC-14243. Then, the sensitivity of the filamentous microorganisms was determined by the microdilution M38-A method, ¹⁵ and that of the yeast fungi with the M27-A3 method. ¹⁶

Such antifungal activity was compared to that of itraconazole as the standard antifungal drug. The minimum inhibitory concentration (MIC) values of the standard and compounds 4a-k, expressed in micrograms per milliliter, were determined in 96-well plates using MOPS (3-[N-morpholino]propanesulfonic acid buffered RPMI-1640 medium, Sigma-Aldrich).

4. Results and discussion

The antifungal activity of the test compounds is summarized in Table 2. Compounds 4d, 4h and 4k showed excellent activity against *C. glabrata* (MIC 0.12, 0.25 and 0.12 μg mL⁻¹, respectively), better than that of

itraconazole (MIC 1 μ g ml⁻¹). The activity of compound 4d (MIC = 2 μ g mL⁻¹) was higher than that observed for the standard antifungal drug (MIC = 8 μ g mL⁻¹) against *Trichosporon cutaneum*, while compound 4k displayed an excellent antimycotic activity against *Mucor hiemalis* (MIC = 2 μ g mL⁻¹ vs. 4 μ g mL⁻¹ for itraconazole). Compounds 4a, 4c and 4e proved to be moderate antifungal agents against *Aspergillus fumigatus* strains. The current results suggest that the presence of either the *a*-CN or *p*-NO₂Ph group at position 5 of the triazole cores increases the biological activity of these compounds in both yeast and filamentous fungi.

These outcomes can also be described by the 'sensitivity' parameters of yeasts, according to the breakpoints described in the M27-A3 document (Table 3). In general, *C. glabrata*, *C. krusei* and *C. parapsilosis* showed some susceptibility to the test compounds, whereas *C. albicans*, *C. tropicalis* and *C. utilis* were resistant to all of them.

Table 2 In vitro antifungal activities of the synthetized compounds (MIC, $\mu g \ mL^{-1}$)

Compound	C. alb	C. trop	C. uti	C. kru	C. gla	C. par	M. hie	A. fum	T. cut	R. ory
4a	8	8	8	0.5	1	8	4	2	8	16
4b	1	8	4	8	2	0.25	16	8	8	16
4c	4	8	4	0.25	4	8	16	2	8	16
4d	8	8	8	8	0.12	0.5	16	16	2	16
4e	8	8	8	8	4	8	16	4	8	16
4f	8	8	8	8	1	8	16	8	8	16
4g	8	8	8	2	2	4	16	16	8	16
4h	8	8	8	8	0.25	8	16	16	8	16
4i	8	8	8	8	2	4	16	16	8	8
4j	8	8	8	8	2	8	16	16	8	16
4k	8	8	8	0.5	0.12	8	2	16	8	16
Standard ^a	0.03	0.06	0.25	0.25	1	0.06	4	1	8	1

Abbreviations: C. alb., Candida albicans; C. trop., Candida tropicalis; C. uti., Candida utilis; C. kru., Candida krusei; C.gla., Candida glabrata, C. par., Candida parapsilosis; M. hie., Mucor hiemalis; A. fum., Aspergillus fumigatus; T. cut., Trichosporon cutaneum; R. ory., Rhizopus oryzae.^a Itraconazole.

Table 3 Determination of the sensitivity of yeast (according to docu-

ment M27-A3): Susceptible (S), dose-dependent sensitive (SDD) and resistant (R)

Compound	C. alb	C. trop	C. uti	C. kru	C. gla	C. par
4a	R	R	R	SDD	R	R
4b	R	R	R	R	R	SDD
4c	R	R	R	SDD	R	R
4d	R	R	R	R	S	SDD
4e	R	R	R	R	R	R
4f	R	R	R	R	R	R
4g	R	R	R	R	R	R
4h	R	R	R	R	SDD	R
4i	R	R	R	R	R	R
4j	R	R	R	R	R	R
4k	R	R	R	SDD	S	R
Standard ^a	S	S	SDD	SDD	R	S

Abbreviations: C. alb., Candida albicans; C. trop., Candida tropicalis; C. uti., Candida utilis; C. kru., Candida krusei; C.gla., Candida glabrata, C. par., Candida parapsilosis. a Itraconazole. Interpretive criteria: breakpoints (MIC, $\mu g \text{ mL}^{-1}$) = 0.12 [S], 0.25-0.5 [SDD], 1 [R].

5. Conclusion

MedChemComm

In summary, eleven oxazolidin-2-one-linked 1,2,3-triazole derivatives (4a-k) were synthesized in good yields based on azide-enolate 1,3-dipolar cycloaddition. In vitro assays demonstrated that compound 4k is the most efficient antimicrobial agent, since it was either better than or comparable to itraconazole against three species (C. glabrata, M. hiemalis and T. cutaneum). The second best antimicrobial activity was exhibited by compound 4d, which was much better than the reference drug against two species (C. glabrata and T. cutaneum). In consequence, these compounds can be considered as drug candidates for future complementary biological studies. In addition, we have developed a novel organic solvent-free synthesis of β-ketosulfones in an aqueous nanomicellar medium. The surfactant nature of SPGS-550-M or 'NOK' (a third generation surfactant) allows it to act as an efficient nanoreactor in catalytic amounts. The notable advantages of this methodology over those previously reported include its simplicity of handling, mild conditions, high yields, cheap reagents and great tolerance of functional groups.

Conflicts of interest

The authors declare no competing interest.

Acknowledgements

We gratefully acknowledge financial support from the Secretaría de Investigación y Estudios Avanzados/UAEMéx (project no. 3804/2014/CID) and CONACYT-Mexico (postgraduate scholarship no. 227581 and no. 273644). The authors would also like to thank the referees for their valuable comments and suggestions, Signa S.A. de C.V. for kindly donating some solvents and reagents, and L. Triana-Cruz (CCIQS UAEMex-UNAM) for technical support.

References

- 1 (a) N. Pandit, R. K. Singla and B. Shrivastava, Int. J. Med. Chem., 2012, 2012, 24; (b) K. J. Shaw and M. R. Barbachyn, Ann. N. Y. Acad. Sci., 2011, 1241, 48-70.
- 2 (a) A. Massarotti, S. Aprile, V. Mercalli, E. Del Grosso, G. Grosa, G. Sorba and G. C. Tron, ChemMedChem, 2014, 9, 2497-2508; (b) S. G. Agalave, S. R. Maujan and V. S. Pore, Chem. - Asian J., 2011, 6, 2696-2718; (c) R. Kharb, P. C. Sharma and M. S. Yar, J. Enzyme Inhib. Med. Chem., 2011, 26, 1-21.
- 3 Oxazolidinone scaffolds: (a) O. A. Phillips and L. H. Sharaf, Expert Opin. Ther. Pat., 2016, 26, 591-605; (b) K. Michalska, I. Karpiuk, M. Król and S. Tyski, Bioorg. Med. Chem., 2013, 21, 577-591; (c) K. Michalska, I. Karpiuk, M. Król and S. Tyski, Bioorg. Med. Chem., 2013, 21, 577-591; (d) 1,2,3-Triazole scaffolds: F. de Carvalho da Silva, M. F. do Carmo Cardoso, P. Garcia-Ferreira and V. F. Ferreira, Biological properties of 1H-1,2,3- and 2H-1,2,3-triazoles, in Chemistry of 1,2,3-triazoles, ed. W. Dehaen and V. A. Bakulev, Springer International Publishing, Switzerland, 2015, pp. 117-165, DOI: 10.1007/7081_2014_124.
- 4 (a) M. V. Nora de Souza, Expert Opin. Ther. Pat., 2008, 18, 1101-1105; (b) F. Reck, F. Zhou, M. Girardot, G. Kern, C. J. Eyermann, N. J. Hales, R. R. Ramsay and M. B. Gravestock, J. Med. Chem., 2005, 48, 499-506; (c) J. A. Demaray, J. E. Thuener, M. N. Dawson and S. J. Sucheck, Bioorg. Med. Chem. Lett., 2008, 18, 4868-4871; (d) O. A. Phillips, E. E. Udo and S. M. Samuel, Eur. J. Med. Chem., 2008, 43, 1095-1104; (e) O. A. Phillips, E. E. Udo, M. E. Abdel-Hamid and R. Varghese, Eur. J. Med. Chem., 2009, 44, 3217-3227; (f) H. Fan, Y. Chen, Z. Jiang, S. Zhang, D. Zhong, R. Ji and Y. Yang, Eur. J. Med. Chem., 2008, 43, 1706-1714; (g) G. Acquaah-Harrison, S. Zhou, J. V. Hines and S. C. Bergmeier, J. Comb. Chem., 2010, 12, 491-496.
- 5 M. B. Gravestock, M. J. Betts, D. A. Griffin and I. R. Matthews, Oxazolidinone derivatives with antibacterial activity, PCT Int. Appl. WO0181350, Astrazeneca U.K. Limited, Astrazeneca AB, Sweden, 2001.
- 6 A. L. Choy and V. P. V. N. Josyula, Benzisoxazole oxazolidinones as antibacterial agents, PCT Int. Appl. WO2007088438A2, Pfizer Products Inc., US, 2007.
- 7 http://melinta.com/.
- 8 (a) B. Bozdogan and P. C. Appelbaum, Int. J. Antimicrob. Agents, 2004, 23, 113-119; (b) A. Chan, J. Cross, Y. He, B. Lippa and D. Ryan, Antibacterial Drugs, in Drug Discovery: Practices, Processes, and Perspectives, ed. J. J. Li and E. J. Corey, John Wiley & Sons, Inc., Hoboken, New Jersey, 2013, ch. 10, pp. 412-413, DOI: 10.1002/9781118354483.ch10.
- 9 (a) F. Schiaffella, A. Macchiarulo, L. Milanese, A. Vecchiarelli, G. Costantino and D. Pietrella, J. Med. Chem., 2005, 48, 7658-7666; (b) J. Y. Choi, L. M. Podust and W. R. Roush, Chem. Rev., 2014, 114, 11242-11271.
- 10 B. M. Santoyo, C. González-Romero, O. Merino, R. Martínez-Palou, A. Fuentes-Benites, H. A. Jiménez-Vázquez, F. Delgado and J. Tamariz, Eur. J. Org. Chem., 2009, 2009, 2505-2518.

11 Sigma-Aldrich catalog number: 776033.

Research Article

- 12 J. C. Lee, Y. H. Bae and S. K. Chang, *Bull. Korean Chem. Soc.*, 2003, 24, 407–408.
- 13 (a) N. Jung and S. Bräse, Click Reactions: Azide-Alkyne Cyclo-addition, *Kirk-Othmer Encycl. Chem. Technol.*, 2013, 1–43, DOI: 10.1002/0471238961.clicjung.a01; (b) Click Reactions in Organic Synthesis, ed. S. Chandrasekaran, Wiley-VCH, Weinheim, Germany, 2016, DOI: 10.1002/9783527694174.
- 14 D. González-Calderón, I. Santillán-Iniesta, C. A. González-González, A. Fuentes-Benítes and C. González-Romero, Tetrahedron Lett., 2015, 56, 514–516.
- 15 (a) Clinical and Laboratory Standards Institute (CLSI), Document M38-A2: Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi, Approved Standard, Clinical and Laboratory Standards Institute, Wayne, PA, 2nd edn, 2002; (b) A. L. Barry, An overview of the Clinical and Laboratory Standards Institute (CLSI) and its impact on antimicrobial susceptibility tests, in Antimicrobial Susceptibility Testing Protocols, ed. R. Schwalbe, L. Steele-Moore and A. C. Goodwin, CRC Press Taylor & Francis Group, Florida, 2007, pp. 1–6, ISBN: 9780824741006; (c) A. Espinel-Ingroff and E. Canton, Antifungal susceptibility testing of filamentous fungi, in

- Antimicrobial Susceptibility Testing Protocols, ed. R. Schwalbe, L. Steele-Moore and A. C. Goodwin, CRC Press Taylor & Francis Group, Florida, 2007, pp. 209–241, ISBN: 9780824741006.
- 16 (a) Clinical and Laboratory Standards Institute (CLSI), M27-Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts, Approved Standard, Clinical and Laboratory Standards Institute, Wayne, PA, 3rd edn, 2008, ISBN: 1-56238-666-2; (b) A. Espinel-Ingroff and E. Cantón, Antifungal susceptibility testing of yeasts, in Antimicrobial Susceptibility Testing Protocols, ed. R. Schwalbe, L. Steele-Moore and A. C. Goodwin, CRC Press Taylor & Francis Group, Florida, 2007, pp. 173-208, ISBN: 9780824741006; (c) A. W. Fothergill, Antifungal Susceptibility Testing: Clinical Laboratory and Standards Institute (CLSI) methods, in Interactions of Yeasts, Moulds, and Antifungal Agents, How to Detect Resistance, ed. G. S. Hall, Springer Science-Business Media, 2012, pp. 65-74, DOI: 10.1007/978-1-59745-134-5_2; (d) M. A. Pfaller and D. J. Diekema, J. Clin. Microbiol., 2012, 50, 2846-2856; (e) M. C. Arendrup, G. Garcia-Effron, C. Lass-Flörl, A. G. López, J. L. Rodriguez-Tudela, M. Cuenca-Estrella and D. S. Perlin, Antimicrob. Agents Chemother., 2010, 54, 426-439.